



Universiteit
Leiden
The Netherlands

Reagent controlled glycosylations for the assembly of well-defined Pel oligosaccharides

Wang, L.; Zhang, Y.; Overkleeft, H.S.; Marel, G.A. van der; Codée, J.D.C.

Citation

Wang, L., Zhang, Y., Overkleeft, H. S., Marel, G. A. van der, & Codée, J. D. C. (2020). Reagent controlled glycosylations for the assembly of well-defined Pel oligosaccharides. *Journal Of Organic Chemistry*, 85(24), 15872-15884. doi:10.1021/acs.joc.0c00703

Version: Publisher's Version

License: [Creative Commons CC BY-NC-ND 4.0 license](#)

Downloaded from: <https://hdl.handle.net/1887/3133291>

Note: To cite this publication please use the final published version (if applicable).

Reagent Controlled Glycosylations for the Assembly of Well-Defined Pel Oligosaccharides

Liming Wang, Yongzhen Zhang, Herman S. Overkleeft, Gijsbert A. van der Marel, and Jeroen D. C. Codée*

Cite This: *J. Org. Chem.* 2020, 85, 15872–15884

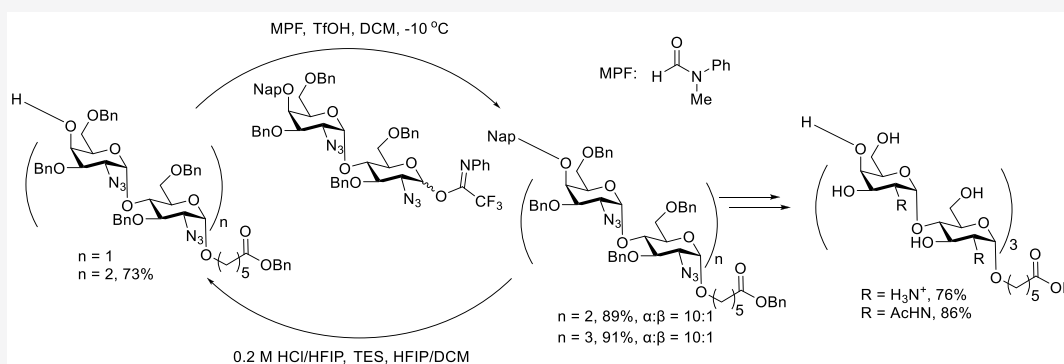
Read Online

ACCESS |

Metrics & More

Article Recommendations

Supporting Information



ABSTRACT: A new additive, methyl(phenyl)formamide (MPF), is introduced for the glycosylation of 2-azido-2-deoxyglucose building blocks. A linear α -(1,4)-glucosamine tetrasaccharide was assembled to prove the utility of MPF. Next, a hexasaccharide fragment of the *Pseudomonas aeruginosa* exopolysaccharide Pel was assembled using a [2 + 2 + 2] strategy modulated by MPF. The used [galactosazide- α -(1,4)-glucosazide] disaccharide building blocks were synthesized using a 4,6-*O*-DTBS protected galactosyl azide donor.

INTRODUCTION

Pel is one of the exopolysaccharides that is involved in the biofilm formation of *Pseudomonas aeruginosa*, an opportunistic Gram-negative pathogen that is the major cause of morbidity and mortality in cystic fibrosis patients.¹ Pel is a linear polysaccharide composed of 1,4-linked α -*N*-acetyl galactosamine (GalNAc) and α -*N*-acetyl glucosamine (GlcNAc) residues, present in a $\pm 6:1$ ratio, of which some of the residues have been deacetylated to generate positively charged galactosamine (GalN) and glucosamine (GlcN) moieties (Figure 1).^{1b} Well-defined Pel fragments can be used to unravel their role in biofilm formation to study their biosynthesis and possibly as synthetic antigens in the development of a (semi)-synthetic vaccine against *P. aeruginosa*. Because of the random distribution of the monosaccharides in Pel, it is impossible to isolate well-defined oligosaccharides from natural sources, and therefore, organic synthesis is necessary to provide these structures.

The key challenge in the generation of these oligosaccharides is the stereoselective construction of the 1,2-*cis*-glycosidic linkages. Four kinds of *cis*-glycosidic linkages, namely α -D-GlcN-(1 \rightarrow 4)-D-GlcN, α -D-GlcN-(1 \rightarrow 4)-D-GalN, α -D-GalN-(1 \rightarrow 4)-D-GlcN, and α -D-GalN-(1 \rightarrow 4)-D-GalN have to be constructed. Zhang et al. recently reported an effective synthetic strategy to assemble galactosaminogalactans

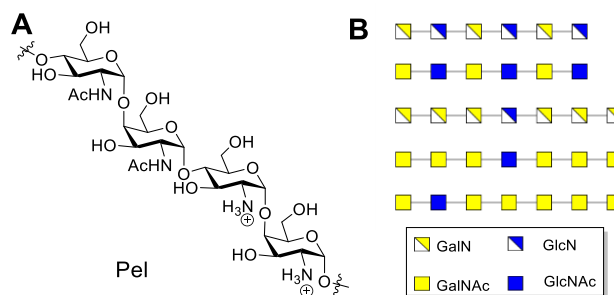


Figure 1. Repeating structures of Pel.

(GAGs), fungal polysaccharides composed of 1,4-linked α -D-Gal, α -D-GalN, and α -D-GalNAc moieties.² For the formation of the 1,2-*cis* linkages in these structures, 4,6-di-*tert*-butylsilylene (4,6-*O*-DTBS) protected GalN donors were

Special Issue: A New Era of Discovery in Carbohydrate Chemistry

Received: March 18, 2020

Published: May 7, 2020

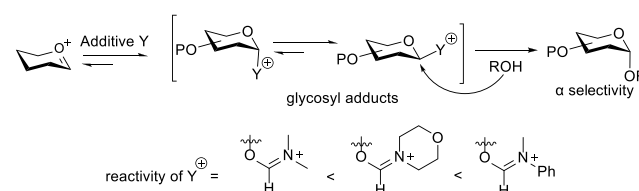


used to control the selectivity.³ This strategy allowed the use of galactosamine donors bearing differently masked amine functionalities. Galactosazide and trichloroacetyl protected GalN donors were used to combine GalN and GalNAc at predetermined sites in the target GAG oligosaccharides. Of note, the stereodirecting capacity of the DTBS group in GalN donors effectively overrides the neighboring group participation by C2-participating functionalities such as the trichloroacetamide. Thus, DTBS-GalN donors also represent attractive building blocks for Pel assembly. For the stereoselective introduction of α -D-GlcN linkages, no general solution exists, even though the construction of this type of glycosidic linkage has attracted significant attention,^{4,5} as it is present in many important natural polysaccharides and glycoconjugates such as heparin, heparan sulfate,⁶ GPI anchors, and various bacterial polysaccharides.⁷

Additive controlled glycosylations are gaining increasing interest for the stereoselective construction of glycosidic linkages.⁸ In these approaches, the nature of the additive determines the reactivity of *in situ* formed glycosylating species, and the influence of the additive can be tuned to match the reactivity of the glycosyl donor⁹ and acceptor¹⁰ building blocks. We have recently reported on the fully stereoselective assembly of a branched α -glucan with an α -(1 \rightarrow 4)-linked backbone from *Mycobacterium tuberculosis*, α -(1,3)-glucans from the *Aspergillus fumigatus* fungal cell wall as well as the assembly of α -(1,3)-glucans found attached to lipoteichoic acids of *Enterococcus faecalis*.¹¹ The synthetic strategy used in these approaches hinged on the use of additive controlled glycosylation reactions in combination with the use of a single benzyl-type protecting group (Bn, PMB, Nap). For glycosylations with relatively reactive primary alcohol acceptors, the trimethylsilyl iodide (TMSI)-triphenylphosphine oxide (Ph₃P=O) activator couple was used, while the condensations with less reactive secondary alcohols required the use of the trifluoromethanesulfonic acid (TfOH)-dimethylformamide (DMF) pair. The successful construction of multiple 1,4- α -glucosidic linkages was an incentive to explore this strategy for the assembly of the Pel oligosaccharides. Mong and coworkers previously described how formamide additives can be used for the construction of 1,2-*cis*-GalN₃ and GlcN₃ linkages. They introduced *N*-formyl-morpholine (NFM) to modulate the reactivity of tri-*O*-benzyl GlcN₃ and 4,6-benzylidene-GalN₃ donors and showed that glycosylations mediated by NFM proceeded with higher stereoselectivity than the corresponding DMF-modulated condensations.^{7c} Because of the stronger electron withdrawing effect of the azide group with respect to a benzyl ether, 2-azido donors are generally less reactive than their 2-*O*-benzyl counterparts. This lower reactivity can be counterbalanced by the use of a somewhat less nucleophilic additive, resulting in a better leaving group Y, thereby explaining why NFM outperforms DMF in these glycosylations (see Scheme 1).

We here describe a strategy to synthesize Pel oligosaccharides using additive-controlled glycosylations to match the reactivities of the GlcN₃ donor and the Pel acceptors. Because of the relatively low nucleophilicity of the GlcN₃-C4-OH and especially the GalN₃-C4-OH, a new additive is introduced that generates intermediates that are more reactive than the previously introduced DMF and NFM-imidinium ions.

Scheme 1. Relative Reactivity of DMF and NFM Glycosyl Adducts

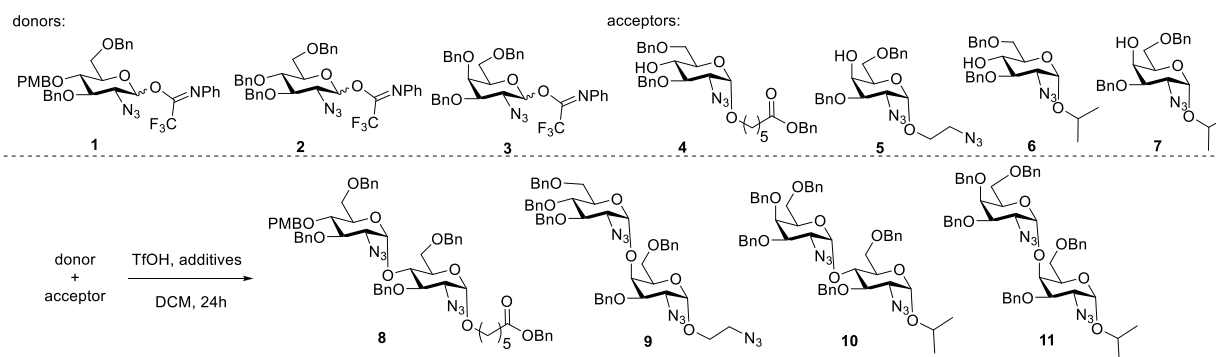


RESULTS AND DISCUSSION

First, we paid attention to the formation of α -D-GlcN-(1 \rightarrow 4)-D-GlcN linkages. In line with previous work, solely benzyl type protecting groups (PMB, Nap, Bn) were used (besides the azide at C2) to generate orthogonally protected building blocks of uniform reactivity. With donor **1** and acceptor **4** (see SI for the syntheses of these building blocks), DMF was investigated as an additive to control the selectivity according to previous successful experiments. Thus, donor α -D-GlcN **1**, acceptor **4**, and the additive were mixed in DCM with molecular sieves and cooled to -78 °C. Next, TfOH was added, and after stirring for 0.5 h, the mixture was placed at 0 °C and allowed to stir for 24 h. As shown in Table 1, this produced the desired disaccharide product **8** with complete α -selectivity, but the yield was only 32% (entry 1). Performing the reaction at room temperature did not lead to erosion of stereoselectivity but only marginally improved the yield (entry 2). Likely, the low reactivity of the donor and acceptor led to the observed poor yield, and NFM was therefore probed as additive.^{6c} Use of this additive provided complete α -selectivity and raised the yield of the condensation to 55% yield. To further improve the reaction, a slightly less nucleophilic additive was sought, and *N*-methyl-*N*-phenylformamide (MPF) was explored. It was expected that the imidinium ion formed from this additive would be more reactive because the aniline-type nitrogen would be less capable of supporting the (partial) positive charge in the ion (see Scheme 1). The reaction of donor **1** and acceptor **4** proceeded with excellent yield (91%) when performed at 0 °C, and the disaccharide **8** was obtained with 15:1 α : β -selectivity (entry 4). Although the stereoselectivity of this condensation is somewhat less than the DMF or NFM mediated glycosylations, the improved yield allows for an overall more productive reaction.¹²

Next, our attention was turned to the formation of the α -GlcN-(1 \rightarrow 4)-GalN linkage exploring the additives as described above. First, donor **2** was coupled with acceptor **5** using DMF to provide product **9** in low yield and poor selectivity (Table 1, entry 5). The use of NFM instead of DMF did not improve the outcome of this glycosylation (entry 6). Likely, the poor reactivity of the GalN₃-C4-OH hampers the union of the two carbohydrate building blocks. Next, the use of MPF was explored. At 0 °C, disaccharide **9** was obtained in high yield (83%) but with moderate α : β -selectivity (5:1). Performing the same reaction at -10 °C increased the α -selectivity (α : β = 10:1) but led to a relatively low yield (43%, entry 8). To increase the yield of the reaction, the concentration was raised from 0.1 to 0.2 M (entry 9). This led to the formation of the desired compound **9** with a yield of 88% and a 10:1 α : β ratio. Having defined adequate conditions for the construction of α -GlcN-(1 \rightarrow 4)-GlcN and α -GlcN-(1 \rightarrow 4)-GalN linkages, the use of MPF in combination with galactosazide donor **3** was explored for the construction of the

Table 1. Glycosylation between 2-Azido Glu/Gal Donors and 4-OH-2-azido Glu/Gal Acceptors



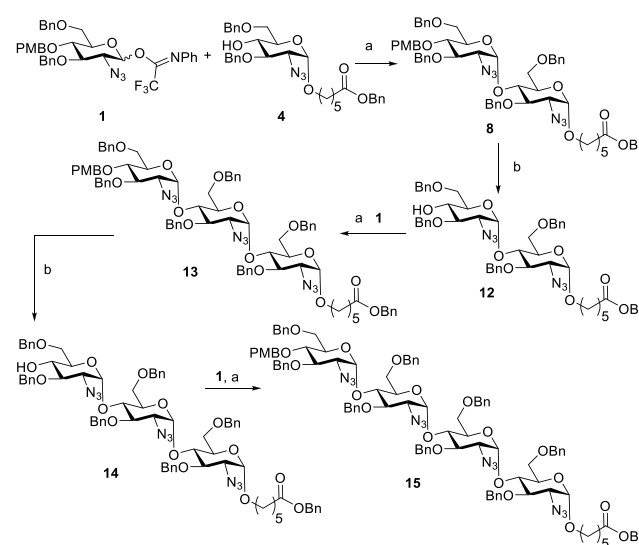
entry	donor	acceptor	conc (mmol/mL)	additive	equiv	T (°C)	product	yield (%) ^a	$\alpha:\beta$ ^b
1	1	4	0.1	DMF	16	0	8	32	>20:1
2	1	4	0.1	DMF	16	rt	8	38	>20:1
3	1	4	0.1	NFM	16	rt	8	55	>20:1
4	1	4	0.1	MPF	16	0	8	91	~15:1
5	2	5	0.1	DMF	16	0	8	23	6:1
6	2	5	0.1	NFM	16	0	9	24	6:1
7	2	5	0.1	MPF	16	0	9	83	5:1
8	2	5	0.1	MPF	16	-10	9	43	10:1
9	2	5	0.2	MPF	16	-10	9	88	10:1
10	3	6	0.1	MPF	16	-10	10	88	8:1
11	3	7	0.1	MPF	16	-10	11	80	4:1

^aIsolated yield. ^bThe $\alpha:\beta$ ratio was determined by ¹H NMR.

target α -GalN-(1 \rightarrow 4)-GlcN and α -GalN-(1 \rightarrow 4)-GalN linkages. Under the conditions established above, donor 3 was coupled with glucosyl acceptor 6 to give the disaccharide 10 in excellent yield and 8:1 α/β -stereo selectivity (entry 10). Contrary, disaccharide 11, formed from donor 3 and galactosyl acceptor 7, was obtained with relatively poor selectivity ($\alpha:\beta = 4:1$, entry 11). From these model reactions, it can be concluded that three out of four Pel-type linkages can effectively be installed using the MPF-mediated glycosylations. For the α -GalN-(1 \rightarrow 4)-GalN linkages, the previously reported approach using 4,6-O-DTBS galactosamine donors is clearly superior.

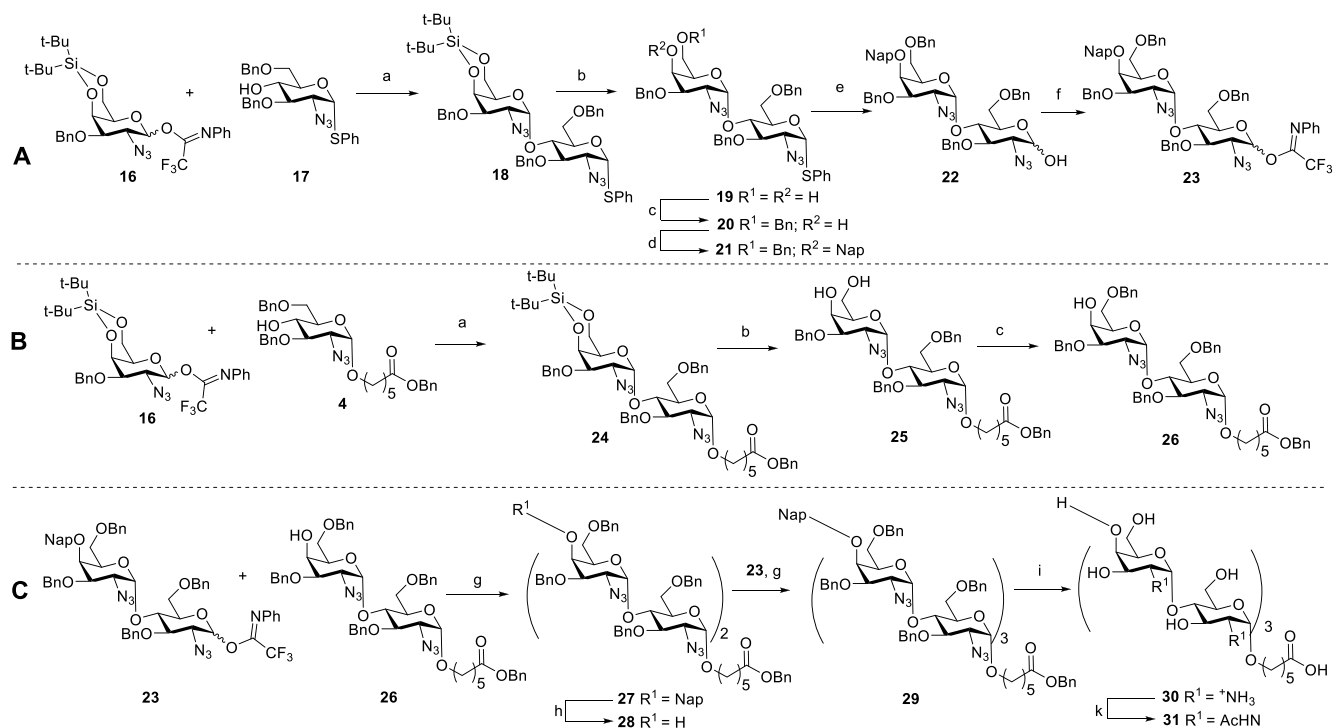
Next, we probed the robustness of the MPF-mediated protocol in the synthesis of Pel-type oligosaccharides. First, the assembly of an all-1,2-*cis* linked tetraglucosamine was explored, as depicted in Scheme 2. Thus, donor 1 and acceptor 4 were coupled under the above identified reaction conditions to provide the desired disaccharide 8. The PMB was removed using a catalytic amount of HCl to give disaccharide acceptor 12 in 88% yield.¹³ Next, compound 12 was glycosylated with donor 1 under the MPF conditions to form the desired trisaccharide 13 in 83% yield and excellent stereoselectivity ($\alpha:\beta > 19:1$). Repetition of the deprotection and glycosylation reactions then uneventfully provided tetrasaccharide 15. The successful assembly of this tetrasaccharide indicates that the yield and stereoselectivity do not decrease with the growing of the sugar chain.

Next, the synthesis of a Pel hexasaccharide featuring both GalN and GlcN residues was undertaken. A [2 + 2 + 2] strategy was designed to streamline the assembly of the structures, building on MPF-mediated glycosylations of the GalN₃-GlcN₃ donor 23. The procedure for the synthesis of the required building blocks 23 and 26 is depicted in Scheme 3A and B. First, donor 16 was coupled with glucoazide 17 in a

Scheme 2. Assembly of an α -Glucosazide Tetrasaccharide Using MPF Mediated Glycosylations^a

^a(a) MPF, TfOH, DCM, -78 to 0 °C, 8: 91%, $\alpha:\beta = 15:1$; 13: 83%, $\alpha:\beta > 19:1$; 15: 90%, $\alpha:\beta > 20:1$. (b) 0.2 M HCl/HFIP, TES, HFIP/DCM, 12: 88%; 14: 78%.

chemoselective glycosylation to form disaccharide 18 as a single anomer. Next, the silylidene ketal was cleaved with HF-pyridine, after which a benzyl ether was regioselectively introduced under the aegis of Taylor's borinic acid catalyst.¹⁴ Protection of the remaining C4'-OH with a naphthyl group delivered compound 21. Next, the anomeric thiophenol group was removed using *N*-iodosuccinimide in acetone/water, and the resulting hydroxyl group turned into the desired *N*-phenyltrifluoroimidate functionality to provide donor 23.

Scheme 3. (A) Synthesis of Donor 23, (B) Synthesis of Acceptor 26, and (C) Assembly of Pel Fragment 31^a

^a(a) TfOH , DCM, **18**: 70%; **24**: 92%. (b) HF -pyridine, THF, **19**: 98%, **25**: 91%. (c) BnBr , borinic acid-catalyzed, K_2CO_3 , KI, CH_3CN , 60 °C, **20**: 96%; **26**: 95%. (d) NapBr , NaH, DMF, **21**: 93%. (e) NIS, acetone, H_2O . (f) 2,2,2-Trifluoro-*N*-phenylacetimidoyl chloride, Cs_2CO_3 , acetone, **23**: 83% over two steps. (g) MPF, TfOH , DCM, -10 °C, 48 h, **27**: 89%, $\alpha:\beta = 10:1$; **29**: 91%, $\alpha:\beta = 10:1$. (h) 0.2 M HCl /HFIP, TES, HFIP/DCM, **28**: 73%. (i) H_2 , $\text{Pd}(\text{OH})_2/\text{C}$, CH_3COOH , THF/ $\text{H}_2\text{O}/t\text{-BuOH}$, **30**: 76%. (k) Ac_2O , NaHCO_3 , H_2O , **31**: 86%.

Acceptor **26** was obtained from donor **16** and acceptor **4**. These two building blocks were united to stereoselectively provide disaccharide **24**. Removal of the silylidene ketal and introduction of the C6'-*O*-benzyl ether as described above provided **26**. With building blocks **23** and **26** in hand, the assembly of the target hexasaccharides was undertaken (Scheme 3C). First, donor **23** was glycosylated with acceptor **26** using MPF as additive at -10 °C at a 0.2 M concentration to form tetrasaccharide **27** in 89% yield as a 10:1 α/β -mixture. Then, the Nap ether was removed using HCl and triethylsilane in DCM/HFIP to give the tetrasaccharide acceptor **28**. Compound **28** was treated with donor **23** under the optimal MPF-mediated glycosylation conditions to deliver hexasaccharide **29** in high yield and stereoselectivity. Reduction of the six azides and removal of the benzyl ester and ethers were accomplished in a one-step reduction to give the compound **30**, of which the amino groups were acetylated with acetic anhydride to afford the Pel structure **31**.

CONCLUSION

In conclusion, MPF is here reported for the first time as a moderator to enable the stereoselective construction of α -GlcN₃ linkages. This additive complements previously introduced glycosylation additives such as DMF and NFM and expands the "nucleophilic additive toolbox" that can be used to match the reactivity of glycosyl donor-acceptor pairs. The applicability of the MPF-mediated glycosylations in oligosaccharide synthesis has been demonstrated by the hand of the assembly of Pel-type oligosaccharides. A linear glucosazide tetrasaccharide was assembled through highly stereoselective glycosylation reactions, using building blocks

solely equipped with benzyl type (Bn and PMB) hydroxyl protecting groups. A [2 + 2 + 2] strategy was developed for the assembly of a (GalN-GlcN)₃ hexasaccharide in which the α -GlcN linkages were constructed in glycosylation reactions using MPF as an additive.

EXPERIMENT SECTION

General Experimental Procedures. All reagents were of commercial grade and used as received. All moisture sensitive reactions were performed under an argon atmosphere. DCM used in the glycosylation reactions was dried with flamed 4 Å molecular sieves before being used. Reactions were monitored by TLC analysis with detection by UV (254 nm) and where applicable by spraying with 20% sulfuric acid in EtOH or with a solution of $(\text{NH}_4)_6\text{Mo}_7\text{O}_{24}\cdot 4\text{H}_2\text{O}$ (25 g/L) and $(\text{NH}_4)_4\text{Ce}(\text{SO}_4)_4\cdot 2\text{H}_2\text{O}$ (10 g/L) in 10% sulfuric acid (aq.) followed by charring at ~150 °C. Column chromatography was carried out using silica gel (0.040–0.063 mm). Size-exclusion chromatography was carried out using Sephadex LH-20. ¹H and ¹³C spectra were recorded on a Bruker AV 400 and Bruker AV 500 in CDCl_3 or D_2O . Chemical shifts (δ) are given in ppm relative to tetramethylsilane as internal standard (¹H NMR in CDCl_3) or the residual signal of the deuterated solvent. Coupling constants (*J*) are given in Hz. All ¹³C spectra are proton decoupled. NMR peak assignments were made using COSY and HSQC experiments; where applicable, Clean TOCSY, HMBC and GATED experiments were used to further elucidate the structure. The anomeric product ratios were analyzed through integration of proton NMR signals.

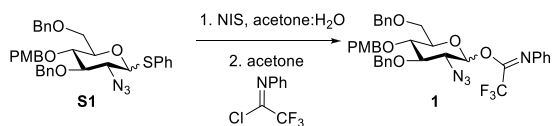
Procedure A for the Glycosylation of Secondary Alcohols. A mixture of donor (1.0 equiv), acceptor (0.7 equiv) (donors and acceptors coevaporated with toluene three times), and MPF (16 equiv) in dry DCM was stirred over fresh flame-dried 3 Å molecular sieves under nitrogen. The solution was cooled to -78 °C, after which TfOH (1.0 equiv) was added. After 30 min, the reaction was stirred at 0 or -10 °C until TLC analysis showed complete conversion of the

acceptor. The reaction was quenched with Et₃N, filtered, and concentrated *in vacuo*. The products were purified by size exclusion and silica gel column chromatography.

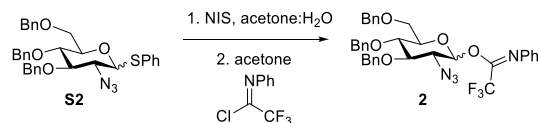
Procedure B for the Glycosylation of Primary Alcohols. A mixture of donor (1.0 equiv), acceptor (0.7 equiv) (donors and acceptors coevaporated with toluene three times), Ph₃P=O (6 equiv) in dry DCM was stirred over fresh flame-dried 3 Å molecular sieves under nitrogen. Then, TMSI (1.0 equiv) was added slowly into the mixture. The reaction was stirred at room temperature until TLC analysis indicated the reaction to be complete. The solution was diluted, and the reaction was quenched with saturated Na₂S₂O₃. The organic phase was washed with water and brine, dried with anhydrous MgSO₄, filtered, and concentrated *in vacuo*. The products were purified by size exclusion and silica gel column chromatography.

Procedure C for Deprotection of the PMB and Nap Protecting Group.¹³ The starting material (1 equiv) was dissolved in DCM:HFIP (1:1, 0.1 M). TES (2.0 equiv) and 0.2 M HCl/HFIP (0.1–1 equiv) were added to the mixture. The reaction mixture was stirred until TLC analysis indicated full consumption of the starting material (15 min to 2 h). Then, the mixture was diluted with DCM, and the reaction was quenched with saturated NaHCO₃. The organic phase was washed with water and brine, dried with anhydrous MgSO₄, filtered, and concentrated *in vacuo*. The product was purified by silica gel column chromatography.

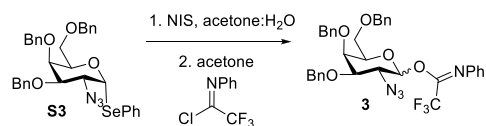
Experimental Procedures and Characterization Data of Products. For the synthesis procedure and data of known compounds **9**,^{15a} **S1**,^{15a} **S2**,^{15b} **S3**,^{15c} and **S10**,^{5e} see references. We used “a”, “b”, “c”, “d”, “e”, “f”, “g”, “h”, and “i” to specify the ¹H and ¹³C NMR signals of sugar rings from the “reducing” to the “non-reducing” end and “o” to specify the ¹H and ¹³C NMR signals of the spacer.



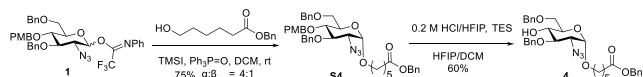
N-Phenyl Trifluoroacetimidate 2-N₃-glucose Donor 1. Compound **S1** (9.1 g, 15.2 mmol) was dissolved in acetone:H₂O (10:1, 150 mL). *N*-Iodosuccinimide (NIS) (6.9 g, 30.5 mmol) was added in one portion, and the reaction mixture was stirred at room temperature for 2 h. The solution was diluted with DCM, and the reaction was quenched with saturated aqueous Na₂S₂O₃. Then, the organic layer was washed with water and brine. The organic layer was dried with anhydrous MgSO₄, filtered, and concentrated *in vacuo*, and the product was purified by column chromatography (pentane:ethyl acetate (EA) = 3:1). The lactol (**7.2** g, 14.3 mmol) was obtained as colorless syrup. Next, the lactol was dissolved in acetone (150 mL). Cs₂CO₃ (7.0 g, 21.3 mmol) and 2,2,2-trifluoro-*N*-phenylacetimidoyl chloride (3.4 mL, 21.3 mmol) were added to the solution, respectively. The reaction was stirred overnight, then quenched with Et₃N, filtered, and concentrated *in vacuo*. The product was purified by column chromatography (pentane:EA = 40:1–20:1). Compound **1** (8.5 g, 79% over two steps, pentane:EA = 10:1, R_f = 0.45–0.55) was obtained as yellow syrup. IR (neat, cm⁻¹) ν 697, 737, 1029, 1082, 1119, 1210, 1251, 1312, 1514, 1720, 2112 (N₃), 2872, 2912. ¹H NMR (CDCl₃, 500 MHz, 60 °C) δ 7.38–7.20 (m, aromatic H), 7.11–7.06 (m, aromatic H), 6.82–6.78 (m, aromatic H), 6.37 (bs, 1 H), 5.41 (bs, 1 H), 4.92–4.80 (m), 4.74–4.69 (m), 4.60–4.48 (m), 3.96 (t, J = 10.0 Hz, 1 H), 3.90 (bd, 1 H), 3.77–3.58 (m), 3.43 (t), 3.33 (bs, 1 H). ¹³C-APT (CDCl₃, 125 MHz, 60 °C) δ 159.8, 159.8, 143.6, 143.5, 138.3, 138.2, 138.1, 130.3 (aromatic C), 129.7, 128.9, 128.6, 128.6, 128.5, 128.1, 128.0, 127.8, 127.9, 127.9, 127.8, 124.7, 124.6, 119.6, 114.2, 114.2 (aromatic CH), 96.2 (C-1), 94.4 (C-1), 83.3, 80.5, 77.7, 77.3, 76.4, 75.7, 75.0, 74.8, 73.9, 73.8, 73.7, 68.5, 65.8, 63.5, 55.4. HRMS (ESI) *m/z*: Calculated for [M - [O(C=NPh)CF₃] + OH + Na]⁺ C₂₈H₃₁O₆N₃Na: 582.21051, found: 582.20943.



Synthesis of N-Phenyl Trifluoroacetimidate 2-N₃-glucose Donor 2. Compound **S2** (8.5 g, 15 mmol) was dissolved in acetone:H₂O (10:1, 150 mL). NIS (6.7 g, 30 mmol) was added in one portion, and the reaction mixture was stirred at room temperature for 2 h. The solution was diluted with DCM, and the reaction was quenched with saturated aqueous Na₂S₂O₃. Then, the organic layer was washed with water and brine. The organic layer was dried with anhydrous MgSO₄, filtered, and concentrated *in vacuo*, and the product was purified by column chromatography (pentane:ethyl acetate (EA) = 3:1). The lactol (**6.1** g, 13 mmol) was obtained as colorless syrup. Next, the lactol was dissolved in acetone (150 mL). Cs₂CO₃ (6.4 g, 19.6 mmol) and 2,2,2-trifluoro-*N*-phenylacetimidoyl chloride (3.4 mL, 21.3 mmol) were added to the solution, respectively. The reaction was stirred overnight, then quenched with Et₃N, filtered, and concentrated *in vacuo*. The product was purified by column chromatography (pentane:EA = 40:1–20:1). Compound **2** (7.3 g, 87%) was obtained as yellow syrup. IR (neat, cm⁻¹) ν 694, 734, 1027, 1073, 1116, 1150, 1208, 1312, 1361, 1452, 1490, 1497, 1598, 1717, 2110 (N₃), 2869, 3032. ¹H NMR (CDCl₃, 500 MHz, 60 °C) δ 7.52–6.81 (m, aromatic H), 6.37 (bs, 1 H, H-1α), 5.43 (bs, 1 H, H-1β), 4.89–4.76 (m, CHH), 4.60–4.48 (m, CHH), 3.98 (t, J = 9.5 Hz, 1 H), 3.91 (bd, 1 H), 3.80–3.59 (m), 3.46 (t), 3.36 (bs, 1 H). ¹³C-APT (CDCl₃, 125 MHz, 60 °C) δ 143.6, 143.5, 138.2, 138.2, 138.1, 138.1, 138.1 (aromatic C), 129.5, 128.9, 128.8, 128.6, 128.6, 128.5, 128.2, 128.1, 128.1, 128.0, 128.0, 127.97, 127.95, 127.91, 127.9, 126.5, 124.7, 124.6, 120.8, 119.6 (aromatic CH), 96.2 (C-1), 94.4 (C-1), 83.3, 80.5, 78.0, 77.6, 76.3, 75.7, 75.7, 75.4, 75.2, 73.9, 73.8, 73.7, 68.5, 65.8, 63.5. HRMS (ESI) *m/z*: Calculated for [M - [O(C=NPh)CF₃] + OH + Na]⁺ C₂₇H₂₉O₅N₃Na: 498.19994, found: 498.19848.

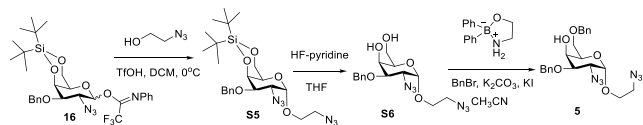


Synthesis of N-Phenyl Trifluoroacetimidate 2-N₃-galactose Donor 3. Compound **S3** (3.7 g, 6.0 mmol) was dissolved in acetone:H₂O (10:1, 150 mL). NIS (2.7 g, 12 mmol) was added in one portion, and the reaction mixture was stirred at room temperature for 2 h. The solution was diluted with DCM, and the reaction was quenched with saturated aqueous Na₂S₂O₃. Then, the organic layer was washed with water and brine. The organic layer was dried with anhydrous MgSO₄, filtered, and concentrated *in vacuo*, and the product was purified by column chromatography (pentane:EA = 3:1). The lactol was obtained as colorless syrup. Next, the lactol was dissolved in acetone. Cs₂CO₃ (3.0 g, 9 mmol) and 2,2,2-trifluoro-*N*-phenylacetimidoyl chloride (1.5 mL, 9 mmol) were added to the solution, respectively. The reaction was stirred overnight, then quenched with Et₃N, filtered, and concentrated *in vacuo*. The product was purified by column chromatography (pentane:EA = 40:1–20:1). Compound **3** (3.3 g, 86%) was obtained as yellow syrup. IR (neat, cm⁻¹) ν 695, 734, 751, 986, 1027, 1153, 1316, 1364, 1454, 1490, 1497, 1590, 1717, 2114 (N₃), 2870, 2915. ¹H NMR (CDCl₃, 400 MHz) δ 7.56–6.79 (m, aromatic H), 6.35 (bs, 1 H, H-1), 5.49 (bs, 1 H, H-1), 5.28 (d), 4.90–4.84 (m, CHH), 4.78–4.31 (m), 4.15–3.83 (m), 3.76 (dd), 3.65–3.31 (m). ¹³C-APT (CDCl₃, 100 MHz) δ 143.5, 143.4, 138.5, 138.3, 138.3, 138.2, 138.1, 137.7, 137.7, 137.6, 137.6, 137.4, 137.3, 135.2 (aromatic C), 129.5, 128.8, 128.7, 128.6, 128.6, 128.6, 128.5, 128.5, 128.43, 128.37, 128.3, 128.19, 128.17, 128.15, 128.14, 128.07, 128.03, 128.00, 127.95, 127.9, 126.48, 124.46, 120.6, 119.4 (aromatic CH), 96.5 (C-1), 92.5 (C-1), 80.9, 80.7, 77.4, 75.1, 74.9, 74.8, 74.7, 74.6, 73.8, 73.67, 73.65, 73.62, 73.5, 72.9, 72.7, 72.6, 72.5, 72.4, 72.3, 72.2, 71.9, 69.7, 69.3, 68.7, 68.3, 68.1, 64.7, 62.2, 60.4, 59.2. HRMS (ESI) *m/z*: [M + Na]⁺ Calculated for C₂₇H₂₉O₅N₃Na: 669.22953, found: 669.22913.



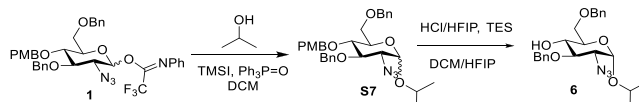
Synthesis of Monosaccharide 4. The reaction was carried out according to the standard procedure B. A mixture of donor **1** (1.0 g, 1.5 mmol), benzyl 6-hydroxyhexanoate (520 mg) (donors and acceptors coevaporated with toluene three times), and $\text{Ph}_3\text{P}=\text{O}$ (2.6 g, 9.3 mmol) in dry DCM (15 mL) was stirred over fresh flame-dried 3 Å molecular sieves under nitrogen. Then, TMSI (222 μL , 1.5 mmol) was added slowly into the mixture. The reaction was stirred at room temperature until TLC analysis indicated the reaction to be complete. The solution was diluted, and the reaction was quenched with saturated $\text{Na}_2\text{S}_2\text{O}_3$. The organic phase was washed with water and brine, dried with anhydrous MgSO_4 , filtered, and concentrated *in vacuo*. The products were purified by silica gel column chromatography (pentane:EA = 8:1, R_f = 0.63). Compound **S4** (800 mg, 75% yield, $\alpha:\beta$ = 5:1) was obtained as a colorless syrup. IR (neat, cm^{-1}) ν 697, 736, 1002, 1029, 1037, 1075, 1150, 1248, 1358, 1454, 1611, 1733 (C=O), 2105 (N_3), 2866, 2933. ^1H NMR (CDCl_3 , 400 MHz) δ 7.40–7.21 (m, 15 H, aromatic H), 7.00 (bd, 2 H, aromatic H), 6.79 (bd, 2 H, aromatic H), 5.09 (s, 2 H, PhCH_2), 4.90 (d, J = 3.6 Hz, 1 H, H-1a), 4.88 (s, 2 H, PhCH_2), 4.71 (d, J = 10.4 Hz, 1 H, CHH), 4.63 (d, J = 12.4 Hz, 1 H, CHH), 4.49 (d, J = 12.4 Hz, 1 H, CHH), 4.43 (d, J = 10.4 Hz, 1 H, CHH), 3.975 (t, J = 9.6 Hz, 1 H, H-3a), 3.79–3.63 (m, 5 H, H-2a, H-4a, H-5a, H-6a, H-1 $^\circ$ _a), 3.47–3.37 (m, 1 H, H-1 $^\circ$ _b), 3.33 (dd, 1 H, J_1 = 10.0 Hz, J_2 = 2.0 Hz, H-2a), 2.36 (t, J = 7.6 Hz, 2H, H-5 $^\circ$), 1.70–1.58 (m, 4 H, H-2 $^\circ$, H-4 $^\circ$), 1.43–1.36 (m, 2 H, H-3 $^\circ$). ^{13}C -APT (CDCl_3 , 100 MHz) δ 173.4 (C=O), 159.4, 138.1, 137.9, 130.1 (aromatic C), 129.6, 128.6, 128.5, 128.5, 128.2, 127.99, 127.96, 127.9, 127.8, 113.9 (aromatic CH), 97.9 (C-1a), 80.2 (C-3a), 78.0 (C-4a), 75.3, 74.8, 73.6 (CH_2), 70.7 (C-5a), 68.3 (C-6a), 68.0 (C-1 $^\circ$), 66.1 (PhCH_2), 63.4 (C-2a), 55.3 (OCH_3), 34.2 (C-5 $^\circ$), 29.1 (C-2 $^\circ$), 25.7 (C-3 $^\circ$), 24.7 (C-4 $^\circ$). HRMS (ESI) m/z : $[\text{M} + \text{NH}_4]^+$ Calculated for $\text{C}_{41}\text{H}_{51}\text{N}_4\text{O}_8$: 727.37014, found: 727.37015.

Then, the reaction was carried out according to the standard procedure C. The starting material **S4** (700 mg, 0.99 mmol) was dissolved in DCM:HFIP (1:1, 0.1 M). TES (314 mL) and 0.2 M HCl/HFIP (0.5 mL) were added to the mixture. The reaction mixture was stirred until TLC analysis indicated full consumption of the starting material (15 min). Then, the mixture was diluted with DCM, and the reaction was quenched with saturated NaHCO_3 . The organic phase was washed with water and brine, dried with anhydrous MgSO_4 , filtered, and concentrated *in vacuo*. The product was purified by silica gel column chromatography (pentane:EA = 4:1, R_f = 0.34). Compound **4** (350 mg, 60% yield) was obtained as a colorless syrup. $[\alpha]_{\text{D}}^{20}$ +59.3 (c = 1, CHCl_3). IR (neat, cm^{-1}) ν 697, 737, 1050, 1147, 1455, 1734 (C=O), 2105 (N_3), 2866, 2926, 3478. ^1H NMR (CDCl_3 , 400 MHz) δ 7.41–7.23 (m, 15 H, aromatic H), 5.10 (s, 2 H, PhCH_2), 4.90 (d, J = 11.2 Hz, 1 H, CHH), 4.87 (d, J = 3.6 Hz, 1 H, H-1a), 4.81 (d, J = 11.2 Hz, 1 H, CHH), 4.59 (d, J = 12.0 Hz, 1 H, CHH), 4.53 (d, J = 12.0 Hz, 1 H, CHH), 3.86–3.64 (m, 6 H, H-2a, H-3a, H-4a, H-5a, H-6a, H-1 $^\circ$ _a), 3.47–3.41 (m, 1 H, H-1 $^\circ$ _b), 3.25 (dd, 1 H, J_1 = 10.0 Hz, J_2 = 2.0 Hz, H-2a), 2.37 (t, J = 7.6 Hz, 2H, H-5 $^\circ$), 1.72–1.61 (m, 4 H, H-2 $^\circ$, H-4 $^\circ$), 1.47–1.37 (m, 2 H, H-3 $^\circ$). ^{13}C -APT (CDCl_3 , 100 MHz) δ 173.6 (C=O), 138.2, 137.9, 136.1 (aromatic C), 128.7, 128.6, 128.5, 128.3, 128.3, 128.1, 128.05, 127.9, 127.7, 127.5 (aromatic CH), 98.0 (C-1a), 79.8 (C-3a), 75.0 (C-6a), 73.7 (CH_2), 72.2 (c-4a), 70.2 (c-5a), 69.8 (PhCH_2), 68.1 (C-1 $^\circ$), 66.2 (PhCH_2), 62.8 (C-2a), 34.2 (C-5 $^\circ$), 29.1 (C-2 $^\circ$), 25.7 (C-3 $^\circ$), 24.7 (C-4 $^\circ$). HRMS (ESI) m/z : $[\text{M} + \text{NH}_4]^+$ Calculated for $\text{C}_{33}\text{H}_{43}\text{O}_7\text{N}_4$: 607.31263, found: 607.31238.

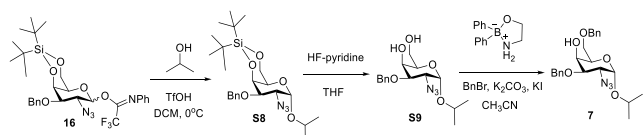


Synthesis of Acceptor 5. Donor **16** (620 mg, 1.0 mmol) and 2-azidoethanol (178 mg, 2.0 mmol) were dissolved in DCM and cooled to 0 °C, and TfOH (15 μL , 0.1 mmol) was added. The reaction was

stirred at 0 °C until TLC analysis showed complete conversion of the donor. The reaction was quenched with Et_3N after completion, checked by TLC, filtered, and concentrated *in vacuo*. Compound **S5** (370 mg, 73%) was obtained with full α -selectivity. Then, compound **S5** was dissolved in THF. HF-pyridine was added to the solution. After TLC analysis showed complete consumption of the starting material, the reaction was quenched with saturated NaHCO_3 . The mixture was diluted with ethyl acetate, washed with H_2O and brine, dried with anhydrous MgSO_4 , filtered, concentrated *in vacuo*. Crude compound **S6**, K_2CO_3 , KI, and borinic acid-catalyzed were mixed in CH_3CN , and then BnBr was added in the solution. The reaction was stirred at 60 °C until TLC analysis showed complete conversion of the starting material. The reaction was quenched with H_2O after completion, checked by TLC, filtered, and concentrated *in vacuo*, purified by column chromatography (pentane:EA = 5:1). Compound **5** (280 mg, 84% yield over two steps) was obtained as colorless syrup. $[\alpha]_{\text{D}}^{20}$ +89.9 (c = 1, CHCl_3). IR (neat, cm^{-1}) ν 698, 738, 1052, 1096, 1146, 1454, 2108 (N_3), 2873, 2923, 3483. ^1H NMR (CDCl_3 , 500 MHz) δ 7.40–7.28 (m, 10 H, aromatic H), 4.95 (d, J = 3.5 Hz, 1 H, H-1a), 4.71 (d, J = 11.5 Hz, 1 H, CHH), 4.68 (d, J = 11.5 Hz, 1 H, CHH), 4.60 (d, J = 12.0 Hz, 1 H, CHH), 4.57 (d, J = 12.0 Hz, 1 H, CHH), 4.12 (t, J = 1.5 Hz, 1 H, H-4a), 3.98 (t, J = 6.0 Hz, 1 H, H-5a), 3.93 (dd, 1 H, J_1 = 10.5 Hz, J_2 = 3.0 Hz, H-3a), 3.90–3.86 (m, 1 H, H-1 $^\circ$ _a), 3.77–3.63 (m, 4 H, H-2a, H-6a, H-1 $^\circ$ _b), 3.57–3.52 (m, 1 H, H-2 $^\circ$ _a), 3.37–3.33 (m, 1 H, H-2 $^\circ$ _b), 2.61 (bt, 1 H, OH), 1.21–1.18 (bt, 6 H, 2 CH_3). ^{13}C -APT (CDCl_3 , 125 MHz) δ 137.9, 137.3 (aromatic C), 128.8, 128.6, 128.4, 128.2, 127.9, 127.8 (aromatic CH), 98.5 (C-1a), 76.0 (C-3a), 73.8, 72.1 (CH_2), 69.6 (C-6a), 69.2 (C-5a), 67.2 (C-1 $^\circ$), 66.8 (C-4a), 59.0 (C-2a), 50.8 (C-2 $^\circ$). HRMS (ESI) m/z : $[\text{M} + \text{NH}_4]^+$ Calculated for $\text{C}_{22}\text{H}_{30}\text{O}_5\text{N}_7$: 472.23029, found: 472.23003.



Synthesis of Acceptor 6. Donor **1** (820 mg, 1.2 mmol), isopropanol (200 μL , 2.6 mmol), and $\text{Ph}_3\text{P}=\text{O}$ (2 g) were dissolved in DCM (12 mL), and TMSI (173 μL) was added at room temperature. The reaction was stirred at rt until TLC analysis showed complete conversion of the donor. The reaction was quenched with Et_3N after completion, checked by TLC, filtered, and concentrated *in vacuo*, purified by column chromatography. Compound **S7** was obtained with $\alpha:\beta$ = 5:1. Then, compound **S7** was dissolved in DCM/HFIP (1.5 mL: 1.5 mL). TES (380 μL) and 0.2 M HCl/HFIP (600 μL) were added to the mixture. The reaction mixture was stirred until TLC analysis indicated full consumption of the starting material (30 min). Then, the mixture was diluted with DCM, and the reaction was quenched with saturated NaHCO_3 . The organic phase was washed with water and brine, dried with anhydrous MgSO_4 , filtered, and concentrated *in vacuo*. The product was purified by silica gel column chromatography (pentane:EA = 5:1). Compound **6** (240 mg, 47% yield over two steps) was obtained as colorless syrup. $[\alpha]_{\text{D}}^{20}$ +83.4 (c = 1, CHCl_3). IR (neat, cm^{-1}) ν 697, 735, 1029, 1047, 1120, 1454, 2105 (N_3), 2920, 2974, 3476. ^1H NMR (CDCl_3 , 400 MHz) δ 7.40–7.20 (m, 10 H, aromatic H), 4.97 (d, J = 3.6 Hz, 1 H, H-1a), 4.88 (d, J = 11.2 Hz, 1 H, CHH), 4.78 (d, J = 11.2 Hz, 1 H, CHH), 4.57 (d, J = 12.0 Hz, 1 H, CHH), 4.50 (d, J = 12.0 Hz, 1 H, CHH), 3.93–3.82 (m, 3 H, H-3a, H-5a, H-1 $^\circ$), 3.73–3.61 (m, 3 H, H-4a, H-6a), 3.18 (dd, 1 H, J_1 = 10.0 Hz, J_2 = 3.6 Hz, H-2a), 2.76 (bs, 1 H, OH), 1.23, (d, J = 8.4 Hz, 3 H, CH_3), 1.21 (d, J = 8.4 Hz, 3 H, CH_3). ^{13}C -APT (CDCl_3 , 100 MHz) δ 138.2, 137.8 (aromatic C), 128.5, 128.4, 128.0, 127.9, 127.7, 127.6 (aromatic CH), 96.4 (C-1a), 79.6 (C-3a), 74.9, 73.6 (CH_2), 72.1 (C-4a), 70.8 (C-1 $^\circ$), 70.1 (C-5a), 69.7 (C-6a), 62.5 (C-2a), 23.2 (CH_3), 21.5 (CH_3). HRMS (ESI) m/z : $[\text{M} + \text{NH}_4]^+$ Calculated for $\text{C}_{23}\text{H}_{33}\text{O}_5\text{N}_4$: 445.24455, found: 445.24441.



Synthesis of Acceptor 7. Donor **16** (2.77 g, 4.6 mmol) and isopropanol were dissolved in DCM (40 mL), cooled to 0 °C and TfOH (40 μ L) was added. The reaction was stirred at 0 °C until TLC analysis showed complete conversion of the donor. The reaction was quenched with Et₃N after completion, checked by TLC, filtered, and concentrated *in vacuo*. Compound **8** was obtained with full α -selectivity. Then, compound **8** was dissolved in THF (20 mL). HF-pyridine (1 mL) was added to the solution. After TLC analysis showed complete consumption of the starting material, the reaction was quenched with saturated NaHCO₃. The mixture was diluted with ethyl acetate, washed with H₂O and brine, dried with anhydrous MgSO₄, filtered, concentrated *in vacuo*, purified by column chromatography (pentane:EA = 3:1). Compound **9** (1.45 g) was obtained with 94% yield over two steps. Then, compound **9** (665 mg, 1.97 mmol), K₂CO₃ (293 mg), KI (327 mg), and borinic acid-catalyzed (44 mg) were mixed in CH₃CN (20 mL), and then BnBr was added in the solution. The reaction was stirred at 60 °C in oil bath until TLC analysis showed complete conversion of the starting material. The reaction was quenched with H₂O after completion, checked by TLC, filtered, and concentrated *in vacuo*, purified by column chromatography (pentane:EA = 5:1). Compound **7** (745 mg, 80% yield) was obtained as colorless syrup. [α]_D²⁰ +102.7 (c = 1, CHCl₃). IR (neat, cm⁻¹) ν 698, 737, 1052, 1454, 2108 (N₃), 2892, 2926, 2972. ¹H NMR (CDCl₃, 400 MHz) δ 7.42–7.27 (m, 10 H, aromatic H), 5.02 (d, J = 3.6 Hz, 1 H, H-1a), 4.71 (bs, 2 H, PhCH₂), 4.58 (bs, 2 H, PhCH₂), 4.15 (t, J = 1.6 Hz, 1 H, H-4a), 4.01 (bt, 1 H, H-5a), 3.95–3.89 (m, 2 H, H-3a, H-1°), 3.76 (dd, 1 H, J₁ = 10.0 Hz, J₂ = 6.0 Hz, H-6a), 3.70–3.62 (m, 2 H, H-6a_b, H-2a), 2.60 (bs, 1 H, OH), 1.23 (d, 3 H, J = 10.4 Hz, CH₃), 1.21 (d, 3 H, J = 10.4 Hz, CH₃). ¹³C-APT (CDCl₃, 100 MHz) δ 138.0, 137.5 (aromatic C), 128.8, 128.6, 128.3, 128.1, 127.9, 127.8 (aromatic CH), 96.7 (C-1a), 76.1 (C-3a), 73.8, 72.0 (CH₂), 70.9 (C-1°), 69.6 (C-6a), 68.7 (C-5a), 66.8 (C-4a), 59.0 (C-2a), 23.3 (CH₃), 21.6 (CH₃). HRMS (ESI) *m/z*: [M + NH₄]⁺ Calculated for C₂₃H₃₃O₅N₄: 445.24455, found: 445.24455.

Synthesis of Disaccharide 8. The reaction was carried out according to the standard procedure A. A mixture of donor **1** (320 mg, 0.47 mmol), acceptor **4** (185 mg, 0.31 mmol) (donors and acceptors coevaporated with toluene three times), and MPF (610 μ L) in dry DCM (3 mL) was stirred over fresh flame-dried 3 Å molecular sieves under nitrogen. The solution was cooled to –78 °C, after which TfOH (42 μ L) was added. After 30 min, the reaction was stirred at –10 °C until TLC analysis showed complete conversion of the acceptor. The reaction was quenched with Et₃N, filtered, and concentrated *in vacuo*. The product was purified by size exclusion (DCM:MeOH = 1:1). Compound **8** (304 mg, 88% yield, α : β = 15:1, PE:EA = 4:1, R_f = 0.51) was obtained as a colorless syrup. IR (neat, cm⁻¹) ν 697, 736, 1027, 1147, 1249, 1358, 1454, 1514, 1734 (C=O), 2103 (N₃), 2866, 2928. ¹H NMR (CDCl₃, 400 MHz) δ 7.39–7.21 (m, 25 H, aromatic H), 7.00 (bd, 2 H, aromatic H), 6.79 (bd, 2 H, aromatic H), 5.66 (d, J = 4.0 Hz, 1 H, H-1b), 5.11 (s, 2 H, PhCH₂), 4.98 (d, J = 10.4 Hz, 1 H, CHH), 4.93 (d, J = 4.0 Hz, 1 H, H-1a), 4.89–4.82 (m, 3 H, 3 CHH), 4.66 (d, J = 10.0 Hz, 1 H, CHH), 4.54–4.47 (m, 3 H, 3 CHH), 4.37 (d, J = 10.4 Hz, 1 H, CHH), 4.23 (d, J = 10.4 Hz, 1 H, CHH), 4.07 (t, J = 9.2 Hz, 1 H, H-3a), 3.98 (t, J = 9.2 Hz, 1 H, H-4a), 3.87–3.61 (m, 10 H, H-3b, H-4b, H-5a, H-5b, H-6a, H-6b, OCH₃), 3.54–3.44 (m, 2 H, H-6b, H-1°), 3.35–3.29 (m, 3 H, H-2a, H-2b, H-1°), 2.38 (t, J = 7.6 Hz, 2H, H-5°), 1.73–1.63 (m, 4 H, H-2°, H-4°), 1.46–1.38 (m, 2 H, H-3°). ¹³C-APT (CDCl₃, 100 MHz) δ 173.5 (C=O), 159.4, 138.2, 138.0, 137.84, 137.82, 136.2, 130.2 (aromatic C), 129.7, 128.7, 128.5, 128.4, 128.2, 128.1, 128.0, 127.9, 127.84, 127.78, 127.6, 127.4, 113.9 (aromatic CH), 97.8 (C-1b), 97.7 (C-1a), 80.9 (C-3a), 80.3 (C-3b), 77.8 (C-4b), 75.5, 74.7, 74.5, 73.6, 73.5 (PhCH₂), 73.4 (c-4a), 71.6 (c-5b), 70.2 (C-5a), 69.1 (C-6a), 68.2 (C-6b), 67.9 (C-1°), 66.2 (PhCH₂), 63.8 (C-2), 63.4

(C-2), 55.4 (OCH₃), 34.2 (C-5°), 29.2 (C-2°), 25.8 (C-3°), 24.8 (C-4°). HRMS (ESI) *m/z*: [M + NH₄]⁺ Calculated for C₆₁H₇₂N₇O₁₂: 1094.52335, found: 1094.52388.

Synthesis of Disaccharide 9. The reaction was carried out according to the standard procedure A. A mixture of donor **2** (146 mg, 0.22 mmol), acceptor **5** (50 mg, 0.11 mmol) (donors and acceptors coevaporated with toluene three times), and MPF (216 μ L) in dry DCM was stirred over fresh flame-dried 3 Å molecular sieves under nitrogen. The solution was cooled to –78 °C, after which TfOH (19 μ L) was added. After 30 min, the reaction was stirred at –10 °C until TLC analysis showed complete conversion of the acceptor. The reaction was quenched with Et₃N, filtered, and concentrated *in vacuo*. The product was purified by size exclusion (DCM:MeOH = 1:1). Compound **9** (86 mg, 87%, α : β = 10:1) was obtained as a colorless syrup. IR (neat, cm⁻¹) ν 697, 736, 1027, 1046, 1093, 1127, 1150, 1259, 1359, 1454, 2105 (N₃), 2869, 2923. ¹H NMR (CDCl₃, 400 MHz) δ 7.40–7.05 (m, 25 H, aromatic H), 4.99 (bt, 2 H, H-1a and H-1b), 4.90–4.76 (m, 3 H, 3 CHH), 4.69 (d, J = 10.8 Hz, 1 H, CHH), 4.63 (d, J = 10.8 Hz, 1 H, CHH), 4.59–4.53 (m, 2 H, 2 CHH), 4.39 (bt, 2 H, 2 CHH), 4.31 (d, J = 2.4 Hz, 1 H), 4.13–3.49 (m, 13 H), 3.39–3.29 (m, 2 H), 3.22 (dd, J₁ = 12.4 Hz, J₂ = 2.0 Hz, 1 H), 2.96 (dd, J₁ = 10.8 Hz, J₂ = 2.0 Hz, 1 H), 4.48 (d, J₁ = 10.8 Hz, J₂ = 1.6 Hz, 1 H). ¹³C-APT (CDCl₃, 100 MHz) δ 138.1, 137.8, 137.7, 137.5 (aromatic C), 128.6, 128.5, 128.4, 128.4, 128.2, 128.07, 128.06, 127.9, 127.82, 127.78, 127.75, 127.7, 127.2 (aromatic CH), 98.9 (C-1), 98.5 (C-1), 80.2, 78.1, 75.6, 75.4, 74.9, 73.7, 73.3, 73.3, 72.0, 70.9, 69.6, 67.3, 67.3, 67.0, 64.0, 59.4, 50.7. HRMS (ESI) *m/z*: [M + NH₄]⁺ Calculated for C₄₉H₅₇O₉N₁₀: 929.43045, found: 929.43039.

Synthesis of Disaccharide 10. The reaction was carried out according to the standard procedure A. A mixture of donor **3** (77 mg, 0.12 mmol), acceptor **6** (34 mg, 0.08 mmol) (donors and acceptors coevaporated with toluene three times), and MPF (156 μ L) in dry DCM was stirred over fresh flame-dried 3 Å molecular sieves under nitrogen. The solution was cooled to –78 °C, after which TfOH (8 μ L) was added. After 30 min, the reaction was stirred at –10 °C until TLC analysis showed complete conversion of the acceptor. The reaction was quenched with Et₃N, filtered, and concentrated *in vacuo*. The product was purified by size exclusion (DCM:MeOH = 1:1). Compound **10** (56 mg, 88% yield, α : β = 8:1) was obtained as a colorless syrup. IR (neat, cm⁻¹) ν 697, 737, 1050, 1097, 1122, 1258, 1454, 2108 (N₃), 2869, 2928. ¹H NMR (CDCl₃, 400 MHz) δ 7.41–7.19 (m, 25 H, aromatic H), 5.64 (d, J = 3.6 Hz, 1 H, H-1a), 5.03 (d, J = 3.6 Hz, 1 H, H-1b), 4.96 (d, J = 10.0 Hz, 1 H, CHH), 4.91 (d, J = 10.0 Hz, 1 H, CHH), 4.81 (d, J = 11.2 Hz, 1 H, CHH), 4.67 (d, J = 11.2 Hz, 1 H, CHH), 4.61 (d, J = 11.2 Hz, 1 H, CHH), 4.56 (d, J = 12.4 Hz, 1 H, CHH), 4.48 (d, J = 11.2 Hz, 1 H, CHH), 4.44 (d, J = 12.4 Hz, 1 H, CHH), 4.29 (d, J = 11.6 Hz, 1 H, CHH), 4.22 (d, J = 11.6 Hz, 1 H, CHH), 4.07 (dd, J = 8.0, 10.0 Hz, 1 H, H-3b), 3.98–3.78 (m, 7 H), 3.72–3.63 (m, 2 H, H-6), 3.48–3.37 (m, 2 H, H-6), 3.29 (dd, J = 3.6, 10.0 Hz, 1H, H-2b), 1.28 (d, J = 6.4 Hz, 1 H, CH₃), 1.24 (d, J = 6.4 Hz, 1 H, CH₃). ¹³C-APT (CDCl₃, 100 MHz) δ 138.4, 138.2, 137.9, 137.6 (aromatic C), 128.62, 128.57, 128.5, 128.42, 128.37, 128.3, 128.0, 127.92, 127.89, 127.85, 127.8, 127.5, 127.4 (aromatic CH), 98.0 (C-1a), 96.2 (C-1b), 80.8 (C-3b), 77.6 (C-3a), 74.9, 74.5 (PhCH₂), 74.0 (C-2a), 73.6, 73.2 (PhCH₂), 72.9 (C-4b), 72.2 (PhCH₂), 71.1 (C-4a), 70.2 (C-5b), 70.1 (C-5a), 69.5 (C-6), 68.5 (C-6), 63.6 (C-2b), 59.8 (C-1°), 23.4 (CH₃), 21.7 (CH₃). HRMS (ESI) *m/z*: [M + NH₄]⁺ Calculated for C₅₀H₆₀O₉N₇: 902.44470, found: 902.44467.

Synthesis of Disaccharide 11. The reaction was carried out according to the standard procedure A. A mixture of donor **3** (77 mg, 0.12 mmol), acceptor **7** (34 mg, 0.08 mmol) (donors and acceptors coevaporated with toluene three times), and MPF (156 μ L) in dry DCM was stirred over fresh flame-dried 3 Å molecular sieves under nitrogen. The solution was cooled to –78 °C, after which TfOH (8 μ L) was added. After 30 min, the reaction was stirred at –10 °C until TLC analysis showed complete conversion of the acceptor. The reaction was quenched with Et₃N, filtered, and concentrated *in vacuo*. The product was purified by size exclusion (DCM:MeOH = 1:1).

Compound **11** (62 mg, 80% yield, $\alpha:\beta = 4:1$) was obtained as a colorless syrup. $[\alpha]_D^{20} +85.8$ ($c = 1$, CHCl_3). IR (neat, cm^{-1}) ν 697, 736, 986, 1037, 1117, 1209, 1261, 1454, 2106 (N_3), 2870, 2925. ^1H NMR (CDCl_3 , 400 MHz) δ 7.43–7.12 (m, 25 H, aromatic H), 5.05 (d, $J = 3.6$ Hz, 1 H, H-1a), 4.98 (d, $J = 3.6$ Hz, 1 H, H-1b), 4.88 (d, $J = 12.0$ Hz, 1 H, CHH), 4.80 (d, $J = 10.8$ Hz, 1 H, CHH), 4.72 (d, $J = 11.2$ Hz, 1 H, CHH), 4.63 (d, $J = 11.2$ Hz, 1 H, CHH), 4.54 (bd, 3 H, 3 CHH), 4.47 (d, $J = 10.8$ Hz, 1 H, CHH), 4.36 (dd, $J = 5.2, 9.2$ Hz, 1 H, H-5a), 4.28 (d, $J = 2.8$ Hz, 1 H, H-4a), 4.10 (s, 1 H, H-4b), 4.03–3.85 (m, 8 H, H-6b_a, H-5b, H-3b, H-3a, H-2b, H-2a, H-1 $^\circ$), 3.60 (dd, $J = 3.6, 11.2$ Hz, 1H, H-2a), 3.56–3.49 (m, 2 H, H-6b_b, H-6a_a), 3.14–3.09 (m, 2 H, H-6a_b), 1.20 (d, $J = 6.0$ Hz, 1 H, CH_3), 1.19 (d, $J = 6.0$ Hz, 1 H, CH_3). ^{13}C -APT (CDCl_3 , 100 MHz) δ 138.7, 138.0, 137.7, 137.6 (aromatic C), 128.64, 128.58, 128.55, 128.4, 128.3, 128.2, 128.12, 128.09, 128.0, 127.90, 127.87, 127.75, 127.74, 127.6, 127.3 (aromatic CH), 98.2 (C-1b), 96.8 (C-1a), 77.4 (C-3b), 75.9 (C-3a), 75.0, 73.7, 73.2 (PhCH_2), 73.0 (C-4b), 72.9 (C-4a), 71.9, 71.9 (PhCH_2), 71.0 (C-1 $^\circ$), 69.4 (C-5b), 69.2 (C-5a), 67.7 (C-6a), 67.2 (C-6b), 60.4 (C-2b), 59.5 (C-2a), 23.4 (CH_3), 21.7 (CH_3). HRMS (ESI) m/z : $[\text{M} + \text{NH}_4]^+$ Calculated for $\text{C}_{50}\text{H}_{60}\text{O}_9\text{N}_7$: 902.44470, found: 902.44482.

Synthesis of Disaccharide 12. The reaction was carried out according to the standard procedure C. Compound **8** (200 mg, 0.18 mmol) was dissolved in DCM:HFIP (1:1, 0.1 M). TES (60 μL) and 0.2 M HCl/HFIP (100 μL) were added to the mixture. The reaction mixture was stirred until TLC analysis indicated full consumption of the starting material (30 min). Then, the mixture was diluted with DCM, and the reaction was quenched with saturated NaHCO_3 . The organic phase was washed with water and brine, dried with anhydrous MgSO_4 , filtered, and concentrated *in vacuo*. The product was purified by silica gel column chromatography (pentane:EA = 5:1, $R_f = 0.22$). Compound **12** (152 mg, 88% yield) was obtained as a colorless syrup. $[\alpha]_D^{20} +62.9$ ($c = 1$, CHCl_3). IR (neat, cm^{-1}) ν 697, 736, 1029, 1043, 1146, 1261, 1454, 1734 (C=O), 2105 (N_3), 2868, 2926, 3491. ^1H NMR (CDCl_3 , 400 MHz) δ 7.42–7.20 (m, 25 H, aromatic H), 5.64 (d, $J = 3.6$ Hz, 1 H, H-1b), 5.11 (s, 2 H, PhCH_2), 4.98 (d, $J = 10.4$ Hz, 1 H, CHH), 4.93 (d, $J = 3.6$ Hz, 1 H, H-1a), 4.89–4.82 (m, 3 H, 3 CHH), 4.55 (d, $J = 12.0$ Hz, 1 H, CHH), 4.51 (d, $J = 12.0$ Hz, 1 H, CHH), 4.08 (dd, $J_1 = 8.8$ Hz, $J_2 = 10.0$ Hz, 1 H, H-3a), 3.99 (t, $J = 8.8$ Hz, 1 H, H-4a), 3.86–3.65 (m, 7 H, H-3b, H-4b, H-5a, H-5b, H-6b, H-6a_a), 3.53–3.44 (m, 2 H, H-6a_b, H-1 $^\circ$), 3.40–3.33 (m, 2 H, H-2a, H-1 $^\circ$), 3.24 (dd, $J_1 = 3.6$ Hz, $J_2 = 10.0$ Hz, 1 H, H-2b), 2.68 (bs, 1 H, OH), 2.38 (t, $J = 7.6$ Hz, 2H, H-5 $^\circ$), 1.73–1.64 (m, 4 H, H-2 $^\circ$, H-4 $^\circ$), 1.47–1.39 (m, 2 H, H-3 $^\circ$). ^{13}C -APT (CDCl_3 , 100 MHz) δ 173.56 (C=O), 138.23, 138.17, 137.8, 137.7, 136.2 (aromatic C), 128.7, 128.6, 128.5, 128.4, 128.3, 128.2, 128.1, 128.0, 127.9, 127.83, 127.79, 127.6, 127.4 (aromatic CH), 97.7 (C-1a), 97.6 (C-1b), 80.9 (C-3a), 79.7 (C-3b), 75.2, 74.5, 73.7, 73.4 (PhCH_2), 73.1 (C-4b), 72.8 (C-4a), 70.6 (C-5b), 70.2 (C-5a), 69.9 (C-6a), 69.0 (C-6b), 68.2 (C-1 $^\circ$), 66.3 (PhCH_2), 63.8 (C-2a), 62.8 (C-2b), 34.3 (C-5 $^\circ$), 29.2 (C-2 $^\circ$), 25.8 (C-3 $^\circ$), 24.8 (C-4 $^\circ$). HRMS (ESI) m/z : $[\text{M} + \text{NH}_4]^+$ Calculated for $\text{C}_{53}\text{H}_{64}\text{N}_7\text{O}_{11}$: 974.46583, found: 974.46576.

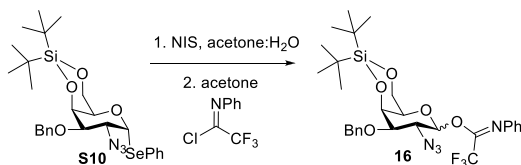
Synthesis of Trisaccharide 13. The reaction was carried out according to the standard procedure A. A mixture of donor **1** (160 mg, 0.24 mmol), acceptor **12** (150 mg, 0.16 mmol) (donors and acceptors coevaporated with toluene three times), and MPF (307 mL) in dry DCM (1.5 mL) was stirred over fresh flame-dried 3 Å molecular sieves under nitrogen. The solution was cooled to -78 °C, after which TfOH (300 μL) was added. After 30 min, the reaction was stirred at -10 °C until TLC analysis showed complete conversion of the acceptor. The reaction was quenched with Et_3N , filtered, and concentrated *in vacuo*. The product was purified by size exclusion (DCM:MeOH = 1:1). Compound **13** (186 mg, 81% yield, $\alpha:\beta > 19:1$, PE:EA = 4:1, $R_f = 0.40$) was obtained as a colorless syrup. $[\alpha]_D^{20} +75.8$ ($c = 1$, CHCl_3). IR (neat, cm^{-1}) ν 697, 736, 1029, 1147, 1249, 1359, 1454, 1514, 1734 (C=O), 2106 (N_3), 2866, 2932. ^1H NMR (CDCl_3 , 400 MHz) δ 7.39–7.21 (m, 35 H, aromatic H), 7.00 (bd, 2 H, aromatic H), 6.79 (bd, 2 H, aromatic H), 5.69 (d, $J = 3.6$ Hz, 1 H, H-1), 5.67 (d, $J = 3.6$ Hz, 1 H, H-1), 5.11 (s, 2 H, PhCH_2), 5.02–4.82 (m, 7 H, 6 CHH, H-1a), 4.66 (d, $J = 10.0$ Hz, 1 H, CHH),

4.56–4.46 (m, 3 H, 3 CHH), 4.39–4.33 (m, 2 H, 2 CHH), 4.26 (d, $J = 12.0$ Hz, 1 H, CHH), 4.18 (d, $J = 12.0$ Hz, 1 H, CHH), 4.14–3.98 (m, 4 H), 3.90–3.59 (m, 11 H), 3.56–3.44 (m, 3 H), 3.37–3.24 (m, 3 H), 2.38 (t, $J = 7.6$ Hz, 2H, H-5 $^\circ$), 1.73–1.63 (m, 4 H, H-2 $^\circ$, H-4 $^\circ$), 1.47–1.39 (m, 2 H, H-3 $^\circ$). ^{13}C -APT (CDCl_3 , 100 MHz) δ 173.5 (C=O), 159.4, 138.3, 138.2, 138.0, 137.8, 137.7, 137.6, 136.1, 130.3 (aromatic C), 129.6, 128.6, 128.57, 128.55, 128.4, 128.3, 128.26, 128.1, 127.9, 127.86, 127.8, 127.7, 127.6, 127.5, 127.4, 127.3, 113.8 (aromatic CH), 97.8, 97.7, 97.4 (C-1a, 1b and 1c), 81.0, 80.7, 79.9 (C-3a, 3b and 3c), 77.7 (C-4c), 75.3, 74.7, 74.6, 74.2, 73.6, 73.5 (PhCH_2), 73.0, 72.5 (C-4a and 4b), 71.5, 71.1, 70.2 (C-5a, 5b and 5c), 68.9, 68.7 (2 C-6), 68.2 (C-1 $^\circ$), 67.7 (C-6), 66.2 (PhCH_2), 63.9, 63.6, 63.1 (C-2a, 2b and 2c), 55.3 (OCH₃), 34.2 (C-5 $^\circ$), 29.1 (C-2 $^\circ$), 25.7 (C-3 $^\circ$), 24.7 (C-4 $^\circ$). HRMS (ESI) m/z : $[\text{M} + \text{NH}_4]^+$ Calculated for $\text{C}_{81}\text{H}_{93}\text{N}_{10}\text{O}_{16}$: 1461.67655, found: 1461.67594.

Synthesis of Trisaccharide Acceptor 14. The reaction was carried out according to the standard procedure C. The starting material **13** (320 mg, 0.22 mmol) was dissolved in DCM:HFIP (1:1, 0.1 M). TES (71 μL) and 0.2 M HCl/HFIP (110 μL) were added to the mixture. The reaction mixture was stirred until TLC analysis indicated full consumption of the starting material (15 min). Then, the mixture was diluted with DCM, and the reaction was quenched with saturated NaHCO_3 . The organic phase was washed with water and brine, dried with anhydrous MgSO_4 , filtered, and concentrated *in vacuo*. The product was purified by silica gel column chromatography (pentane:EA = 4:1). Compound **14** (230 mg, 78% yield) was obtained as a colorless syrup. $[\alpha]_D^{20} +51.0$ ($c = 3$ mg/mL, CHCl_3). IR (neat, cm^{-1}) ν 697, 737, 1028, 1148, 1454, 1736 (C=O), 2106 (N_3), 2866, 2926. ^1H NMR (CDCl_3 , 400 MHz) δ 7.42–7.17 (m, 35 H, aromatic H), 5.67–5.65 (m, 2 H, H-1b and H-1c), 5.12 (s, 2 H, PhCH_2), 5.01–4.85 (m, 7 H, 6 CHH, H-1a), 4.56 (d, $J = 12.0$ Hz, 1 H, CHH), 4.50 (d, $J = 12.0$ Hz, 1 H, CHH), 4.37–4.32 (m, 3 H, 3 CHH), 4.22 (d, $J = 12.0$ Hz, 1 H, CHH), 4.14–3.99 (m, 4 H), 3.87–3.62 (m, 8 H), 3.56–3.43 (m, 3 H), 3.37–3.30 (m, 4 H), 3.18 (dd, $J_1 = 3.6$ Hz, $J_2 = 10.0$ Hz, 1 H, H-2c), 2.76 (bs, 1 H, OH), 2.39 (t, $J = 7.6$ Hz, 2H, H-5 $^\circ$), 1.74–1.64 (m, 4 H, H-2 $^\circ$, H-4 $^\circ$), 1.47–1.41 (m, 2 H, H-3 $^\circ$). ^{13}C -APT (CDCl_3 , 100 MHz) δ 173.6 (C=O), 138.3, 138.20, 138.17, 137.8, 137.6, 137.5, 136.2 (aromatic C), 138.7, 128.7, 128.6, 128.55, 128.5, 128.4, 128.3, 128.13, 128.09, 128.0, 127.95, 127.9, 127.8, 127.7, 127.5, 127.48, 127.3 (aromatic CH), 97.8, 97.7, 97.4 (C-1a, 1b and 1c), 81.1, 80.8, 79.1 (C-3a, 3b and 3c), 75.0, 74.6, 74.3, 73.7, 73.5, 73.4 (PhCH_2), 73.0, 72.9, 72.3 (C-4a, 4b and 4c), 71.1, 70.3, 70.2 (C-5a, 5b and 5c), 70.0, 68.9, 68.6 (C-6a, 6b and 6c), 68.3 (C-1 $^\circ$), 66.26 (PhCH_2), 63.9, 63.7, 62.5 (C-2a, 2b and 2c), 34.3 (C-5 $^\circ$), 29.2 (C-2 $^\circ$), 25.8 (C-3 $^\circ$), 24.8 (C-4 $^\circ$). HRMS (ESI) m/z : Calculated for $\text{C}_{73}\text{H}_{85}\text{N}_{10}\text{O}_{15}$: 1341.61904, found: 1341.61923.

Synthesis of Tetrasaccharide 15. The reaction was carried out according to the standard procedure A. A mixture of donor **1** (40 mg, 0.06 mmol), acceptor **14** (35 mg, 0.03 mmol) (donors and acceptors coevaporated with toluene three times), and MPF (52 μL) in dry DCM (0.3 mL) was stirred over fresh flame-dried 3 Å molecular sieves under nitrogen. The solution was cooled to -78 °C, after which TfOH (5 μL) was added. After 30 min, the reaction was stirred at -10 °C until TLC analysis showed complete conversion of the acceptor. The reaction was quenched with Et_3N , filtered, and concentrated *in vacuo*. The product was purified by size exclusion (DCM:MeOH = 1:1). Compound **15** (43 mg, 87% yield, $\alpha:\beta > 20:1$) was obtained as a colorless syrup. $[\alpha]_D^{20} +94.8$ ($c = 1$, CHCl_3). IR (neat, cm^{-1}) ν 697, 737, 1029, 1148, 1251, 1359, 1454, 1514, 1735 (C=O), 2106 (N_3), 2868, 2928. ^1H NMR (CDCl_3 , 400 MHz) δ 7.42–7.15 (m, 45 H, aromatic H), 7.00 (bd, 2 H, aromatic H), 6.79 (bd, 2 H, aromatic H), 5.70–5.67 (m, 3 H, H-1b, 1c, 1d), 5.11 (s, 2 H, PhCH_2), 5.02–4.87 (m, 8 H, 7 CHH, H-1a), 4.81 (d, $J = 10.4$ Hz, 1 H, CHH), 4.66 (d, $J = 10.4$ Hz, 1 H, CHH), 4.54 (s, 2 H, PhCH_2), 4.65 (d, $J = 12.0$ Hz, 1 H, CHH), 4.38–4.28 (m, 4 H, 4 CHH), 4.22–4.00 (m, 8 H), 3.90–3.59 (m, 15 H), 3.52–3.44 (m, 3 H), 3.39–3.34 (m, 7 H), 2.38 (t, $J = 7.6$ Hz, 2H, H-5 $^\circ$), 1.73–1.64 (m, 4 H, H-2 $^\circ$, H-4 $^\circ$), 1.47–1.38 (m, 2 H, H-3 $^\circ$). ^{13}C -APT (CDCl_3 , 100 MHz) δ 173.5 (C=O), 159.4, 138.3, 138.2, 138.0, 137.84, 137.78, 137.6, 136.2, 130.3 (aromatic C), 129.7, 128.7, 128.65, 128.61, 128.59, 128.48,

128.45, 128.4, 128.33, 128.30, 128.2, 128.02, 127.98, 127.93, 127.87, 127.8, 127.74, 127.71, 127.6, 127.5, 127.4, 127.3, 113.9 (aromatic CH), 97.9, 97.8, 97.5, 97.48 (C-1a, 1b, 1c and 1d), 81.0, 80.9, 80.8, 80.0 (C-3a, 3b, 3c and 3d), 77.8 (C-4), 75.3, 74.7, 74.4, 74.3, 73.6, 73.6, 73.54, 73.51 (PhCH₂), 73.1 (C-4), 72.4 (C-4), 72.1 (C-4), 71.5, 71.2, 71.1, 70.2 (c-5a, 5b, 5c and 5d), 68.9, 68.6, 68.3 (3 C-6), 68.29 (C-1°), 67.8 (C-6), 66.3 (PhCH₂), 63.8, 63.7, 63.6, 63.2 (C-2a, 2b, 2c and 2d), 55.4 (OCH₃), 34.3 (C-5°), 29.2 (C-2°), 25.8 (C-3°), 24.8 (C-4°).



Synthesis of *N*-Phenyl Trifluoroacetimidate 2-*N*₃-galactose Donor **16.** NIS (9.15 g, 40.68 mmol) was added to the solution of compound **S3** (18 g, 31.3 mmol) in Acetone/H₂O (210 mL/72 mL) at 0 °C. The reaction was slowly warmed to room temperature and stirred until TLC analysis indicated full consumption of the starting material (\pm 1h). Then, the mixture was diluted with DCM and washed with saturated Na₂S₂O₃ and brine, dried with anhydrous MgSO₄, filtered, and concentrated *in vacuo*. The lactol was purified by silica gel column chromatography (pentane:EA = 4:1). Cs₂CO₃ was added to the solution of The lactol (10.59g, 24.33 mmol) in 140 mL acetone. The mixture was stirred at 0 °C for 15 min. Then, CF₃C(=NPh)Cl (6.06 g, 29.2 mmol) was added to the solution. which was slowly warmed to room temperature and stirred overnight. The reaction was quenched with Et₃N and concentrated *in vacuo*. The product **16** was purified by silica gel column chromatography (pentane:Et₂O = 30:1–10:1). Compound **16** (13.3 g, a/b = 2:1, 90% yield, PE: Et₂O = 10:1, Rf = 0.45–0.55) was obtained as white solid. α isomer: ¹H NMR (CDCl₃, 400 MHz) δ 7.50–7.24 (m, 7H, aromatic H), 7.15–7.05 (m, 1H, aromatic H), 6.84 (d, *J* = 7.7 Hz, 2H, aromatic H), 6.47 (bs, 1H, H-1), 4.78 (d, *J* = 11.4 Hz, 1H, CH₂Ph), 4.69 (d, *J* = 11.4 Hz, 1H, CH₂Ph), 4.63 (s, 1H, H-4), 4.22 (q, *J* = 12.8 Hz, 2H, H-6), 4.10 (t, *J* = 6.3 Hz, 1H, H-2), 3.89 (d, *J* = 9.5 Hz, 1H, H-3), 3.76 (s, 1H, H-5), 1.09–1.02 (m, 18H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ 143.29, 137.45, 128.74, 128.56, 128.01, 127.91, 124.40, 119.35 (aromatic C/CH), 94.73 (C-1), 76.04 (C-3), 70.71 (CH₂Ph), 69.89 (C-5), 69.16 (C-4), 66.76 (C-6), 57.71 (C-2), 27.59 (CH₃), 27.23 (CH₃), 23.38 (C-Si), 20.73 (C-Si). β isomer: ¹H NMR (CDCl₃, 400 MHz) δ 7.48–7.25 (m, 7H, aromatic H), 7.14–7.04 (m, 1H, aromatic H), 6.85 (d, *J* = 7.7 Hz, 2H, aromatic H), 5.50 (bs, 1H, H-1), 4.77 (d, *J* = 11.9 Hz, 1H, CH₂Ph), 4.66 (d, *J* = 11.9 Hz, 1H, CH₂Ph), 4.43 (s, 1H, H-5), 4.19 (s, 2H, H-6), 4.02 (s, 1H, H-4), 3.30 (s, 2H, H-2, 3), 1.15–1.00 (m, 18H, CH₃). ¹³C NMR (100 MHz, CDCl₃) δ 143.5, 137.5, 128.8, 128.7, 128.2, 128.0, 124.5, 119.4 (aromatic C/CH), 95.8 (C-1), 79.6 (C-3), 72.2 (C-2), 71.0 (CH₂Ph), 68.6 (C-5), 66.8 (C-6), 60.8 (C-4), 27.7 (CH₃), 27.4 (CH₃), 23.6 (C-Si), 20.9 (C-Si). HRMS (ESI) *m/z*: [M + NH₄]⁺ Calculated for C₂₁H₃₇N₃O₅Si: 629.2383, found: 629.2376.

Synthesis of Disaccharide **18.** Donor **16** (5 g, 8.2 mmol) and acceptor **17** (3.32 g, 6.95 mmol) (donors and acceptors coevaporated with toluene three times) were dissolved in DCM (65 mL) and cooled to 0 °C, and TfOH (60 μ L) was added. The reaction was stirred at 0 °C until TLC analysis showed complete conversion of the donor. The reaction was quenched with Et₃N after completion, checked by TLC, filtered, and concentrated *in vacuo*. The product **16** was purified by silica gel column chromatography (pentane:Et₂O = 10:1). Compound **18** (4.36g, 70% yield) was obtained with full α -selectivity as a colorless syrup. [α]_D²⁰ +153.3 (c = 1, CHCl₃). IR (neat, cm⁻¹) ν 651, 698, 738, 797, 826, 984, 1043, 1066, 1100, 1171, 1364, 1473, 2107 (N₃), 2859, 2933. ¹H NMR (CDCl₃, 400 MHz) δ 7.53–7.51 (m, 2 H, aromatic H), 7.44–7.22 (m, 18 H, aromatic H), 5.64 (d, *J* = 3.6 Hz, 1 H, H-1b), 5.59 (d, *J* = 5.2 Hz, 1 H, H-1a), 5.00 (d, *J* = 10.4 Hz, 1 H, CHH), 4.91 (d, *J* = 10.4 Hz, 1 H, CHH), 4.73 (d, *J* = 11.6 Hz, 1 H, CHH), 4.63 (d, *J* = 11.6 Hz, 1 H, CHH), 4.43–4.37 (m, 4 H, 2 CHH, H-4b, H-5a), 3.96–3.77 (m, 6 H, H-2a, H-2b,

H-3a, H-4a, H-6), 3.72–3.64 (m, 2 H, H-3b, H-6a), 3.53 (dd, *J*₁ = 2.0 Hz, *J*₂ = 10.8 Hz, 1 H, H-6_b), 3.42 (s, 1 H, H-5b), 1.03 (s, 9 H, 3 CH₃), 0.97 (s, 9 H, 3 CH₃). ¹³C-APT (CDCl₃, 100 MHz) δ 137.9, 137.8, 137.4, 133.5 (aromatic C), 132.1, 129.2, 128.62, 128.58, 128.51, 128.48, 128.00, 127.98, 127.8, 127.5 (aromatic CH), 97.7 (C-1b), 87.1 (C-1a), 82.3 (C-3a), 75.5 (C-3b), 75.0, 73.3 (PhCH₂), 72.8 (c-4a), 71.3 (c-5a), 70.5 (PhCH₂), 69.6 (C-4b), 68.9 (C-6), 68.0 (C-5b), 66.9 (C-6), 64.6 (C-2a), 58.1 (C-2b), 27.7 (3 CH₃), 27.3 (3 CH₃), 23.4, 20.7. HRMS (ESI) *m/z*: [M + NH₄]⁺ Calculated for C₄₇H₆₂N₇O₈Si: 912.41444, found: 912.41409.

Synthesis of Disaccharide **20.** Compound **18** (4.1 g, 4.6 mmol) was dissolved in THF (40 mL) in a round flask. Then, HF-pyridine (1.2 mL) was added in the solution. The reaction mixture was stirred until TLC analysis indicated full consumption of the starting material (30 min). Then, the mixture was diluted with DCM, and the reaction was quenched with saturated NaHCO₃. The organic phase was washed with water and brine, dried with anhydrous MgSO₄, filtered, and concentrated *in vacuo*. The crude compound **19** was dissolved in CH₃CN (47 mL). Then, BnBr (880 μ L), borinic acid-catalyzed (110 mg), K₂CO₃ (710 mg), KI (800 mg) were added into the mixture. The reaction mixture was stirred at 60 °C in oil bath until TLC analysis indicated full consumption of the starting material (24 h). Then, the mixture was diluted with ethyl acetate and the reaction quenched with saturated NaHCO₃. The organic phase was washed with water and brine, dried with anhydrous MgSO₄, filtered, and concentrated *in vacuo*. The product was purified by silica gel column chromatography (pentane:Et₂O = 5:1). Compound **20** (3.6 g, 94% yield with two steps) was obtained as a colorless syrup. [α]_D²⁰ +11.7 (c = 1, CHCl₃). IR (neat, cm⁻¹) ν 697, 737, 1029, 1046, 1077, 1266, 2106 (N₃), 2870, 2919, 3493. ¹H NMR (CDCl₃, 400 MHz) δ 7.54–7.51 (m, 2 H, aromatic H), 7.41–7.23 (m, 23 H, aromatic H), 5.61 (d, *J* = 3.6 Hz, 1 H, H-1b), 5.60 (d, *J* = 5.2 Hz, 1 H, H-1a), 5.00 (d, *J* = 10.4 Hz, 1 H, CHH), 4.94 (d, *J* = 10.4 Hz, 1 H, CHH), 4.66 (s, 2 H, PhCH₂), 4.50–4.37 (m, 3 H, 2 CHH, 5a), 4.08 (s, 1 H, H-4b), 3.97–3.91 (m, 2 H, H-2b, H-4a), 3.85–3.37 (m, 5 H), 3.66 (dd, *J*₁ = 2.4 Hz, *J*₂ = 10.8 Hz, 1 H, H-6_b), 3.59 (dd, *J*₁ = 5.6 Hz, *J*₂ = 9.6 Hz, 1 H), 3.51 (dd, *J*₁ = 5.6 Hz, *J*₂ = 9.6 Hz, 1 H), 2.63 (s, 1 H, OH). ¹³C-APT (CDCl₃, 100 MHz) δ 138.3, 137.8, 137.6, 137.2, 133.5 (aromatic C), 132.3, 128.8, 128.6, 128.5, 128.4, 128.3, 128.1, 128.0, 127.9, 127.86, 127.85, 127.6, 127.55 (aromatic CH), 98.2 (C-1b), 87.1 (C-1a), 82.1 (C-3a), 76.3 (C-3b), 75.1 (PhCH₂), 74.4 (c-4a), 73.8, 73.2, 71.8 (PhCH₂), 71.3 (c-5a), 69.5 (C-6), 69.4 (C-6), 69.3 (C-5b), 66.5 (C-4b), 64.8 (C-2b), 59.0 (C-2a). HRMS (ESI) *m/z*: [M + NH₄]⁺ Calculated for C₄₆H₅₂N₇O₈S: 862.35926, found: 862.35895.

Synthesis of Thio-disaccharide **21.** The compound **20** (3.83 g, 4.53 mmol) was dissolved in DMF (10 mL). Then, NaH (544 mg) and NapBr (1.5 g) were added into the mixture. The reaction mixture was stirred until TLC analysis indicated full consumption of the starting material (2 h). Then, the mixture was diluted with ethyl acetate and the reaction quenched with ice water. The organic phase was washed with water and brine, dried with anhydrous MgSO₄, filtered, and concentrated *in vacuo*. The product was purified by silica gel column chromatography (pentane:Et₂O = 10:1). Compound **21** (4.15 g, 93% yield) was obtained as a colorless syrup. [α]_D²⁰ +208.6 (c = 1, CHCl₃). IR (neat, cm⁻¹) ν 697, 737, 1028, 1049, 1093, 1123, 1362, 1454, 2106 (N₃), 2870, 2914. ¹H NMR (CDCl₃, 400 MHz) δ 7.80–7.11 (m, 32 H, aromatic H), 5.64 (d, *J* = 3.6 Hz, 1 H, H-1b), 5.59 (d, *J* = 5.2 Hz, 1 H, H-1a), 5.01–4.89 (m, 3 H, 3 CHH), 4.70–4.62 (m, 3 H, 3 CHH), 4.50 (d, *J* = 12.0 Hz, 1 H, CHH), 4.44–4.40 (m, 2 H, CHH, 5b), 4.27 (d, *J* = 11.6 Hz, 1 H, CHH), 4.17 (d, *J* = 11.6 Hz, 1 H, CHH), 4.04 (s, 1 H, H-4b), 3.97–3.81 (m, 6 H, H-2a, H-2b, H-3a, H-3b, H-4a, H-5a), 3.77–3.65 (m, 2 H, H-6), 3.51–3.40 (m, 2 H, H-6). ¹³C-APT (CDCl₃, 100 MHz) δ 138.3, 137.8, 137.60, 137.58, 135.6, 133.6, 133.2, 133.1 (aromatic C), 133.08, 129.2, 128.7, 128.6, 128.5, 128.4, 128.3, 128.1, 127.99, 127.97, 127.93, 127.80, 127.6, 127.4, 127.2, 126.5, 126.2, 126.1 (aromatic CH), 98.3 (C-1b), 87.1 (C-1a), 82.2 (C-3a), 77.5 (C-3b), 75.2, 74.9 (PhCH₂), 74.1 (c-4a), 73.6, 73.1 (PhCH₂), 72.8 (C-4b), 72.3 (PhCH₂), 71.3 (C-5b), 70.4 (C-5a), 69.4 (C-6), 68.6 (C-6), 64.8 (C-2a), 69.8 (C-2b).

HRMS (ESI) m/z : $[M + NH_4]^+$ Calculated for $C_{57}H_{60}N_7O_8S$: 1002.42186, found: 1002.42125.

N-Phenyl Trifluoroacetimidate Disaccharide Donor 23. Compound **21** (4.15 g, 4.21 mmol) was dissolved in acetone:H₂O (10:1, 44 mL). NIS (2.0 g, 8.8 mmol) was added in one portion, and the reaction mixture was stirred at room temperature for 2 h. The solution was diluted with DCM, and the reaction was quenched with saturated aqueous Na₂S₂O₃. Then, the organic layer was washed with water and brine. The organic layer was dried with anhydrous MgSO₄, filtered, and concentrated *in vacuo*, and the product was purified by column chromatography (pentane:EA = 3:1). The lactol **22** was obtained as colorless syrup. Next, the lactol was dissolved in acetone (40 mL). Cs₂CO₃ (1.9 g) and 2,2,2-trifluoro-N-phenylacetimidoyl chloride (960 μ L) were added to the solution, respectively. The reaction was stirred overnight, then quenched with Et₃N, filtered, and concentrated *in vacuo*. The product was purified by column chromatography (pentane:EA = 40:1–20:1). Compound **23** (3.7 g, 81% over two steps) was obtained as yellow syrup. IR (neat, cm⁻¹) ν 695, 734, 818, 1027, 1116, 1209, 1312, 1454, 1489, 1497, 1717, 2107, 2870, 2918. ¹H NMR (CDCl₃, 500 MHz) δ 7.80–7.09 (m, aromatic H), 6.81 (bt, 1 H), 5.65 (dd, 1 H), 5.01–4.87 (m), 4.68–4.54 (m), 4.45–4.42 (m), 4.33–4.18 (m), 4.03–3.41 (m). ¹³C-APT (CDCl₃, 125 MHz) δ 143.4, 143.2, 138.20, 138.18, 137.8, 137.6, 137.55, 137.5, 135.6, 133.3, 133.2 (aromatic C), 128.9, 128.7, 128.53, 128.52, 128.44, 128.43, 128.3, 128.08, 128.05, 128.0, 127.97, 127.9, 127.84, 127.81, 127.69, 127.66, 127.65, 127.6, 127.2, 127.17, 124.7, 124.6 (aromatic CH), 119.4 (C-1), 98.3 (C-1), 98.2 (C-1), 83.6, 81.0, 77.6, 77.3, 75.5, 75.18, 75.16, 75.0, 74.9, 73.64, 73.60, 73.30, 73.2, 73.1, 73.0, 72.8, 72.7, 72.3, 72.0, 70.4, 70.3, 69.0, 68.5, 65.8, 63.7, 59.7, 59.6. HRMS (ESI) m/z : $[M - [O(C=NPh)CF_3] + OH + Na]^+$ Calculated for C₆₉H₅₆F₃N₇O₈Na: 910.41340, found: 910.41374.

Synthesis of Disaccharide 24. Donor **16** (1.09 g) and acceptor **4** (790 mg) (donors and acceptors coevaporated with toluene three times) were dissolved in DCM (12 mL) and cooled to 0 °C, and TfOH (12 μ L) was added. The reaction was stirred at 0 °C until TLC analysis showed complete conversion of the donor. The reaction was quenched with Et₃N after completion, checked by TLC, filtered, and concentrated *in vacuo*. The product was purified by size exclusion (DCM:MeOH = 1:1). Compound **24** (1.24 g, 92% yield) was obtained with full α -selectivity as a colorless syrup. $[\alpha]_D^{20} +95.9$ (c = 1, CHCl₃). IR (neat, cm⁻¹) ν 651, 698, 737, 765, 797, 826, 984, 1004, 1040, 1130, 1144, 1171, 1455, 1474, 1735 (C=O), 2106 (N₃), 2860, 2933. ¹H NMR (CDCl₃, 400 MHz) δ 7.44–7.24 (m, 20 H, aromatic H), 5.67 (d, J = 3.6 Hz, 1 H, H-1b), 5.12 (s, 2 H, PhCH₂), 4.96 (d, J = 10.4 Hz, 1 H, CHH), 4.92 (d, J = 3.6 Hz, 1 H, H-1a), 4.86 (d, J = 10.4 Hz, 1 H, CHH), 4.71 (d, J = 11.6 Hz, 1 H, CHH), 4.61 (d, J = 11.6 Hz, 1 H, CHH), 4.48 (s, 2 H, PhCH₂), 4.36 (d, J = 2.0 Hz, 1 H, H-4b), 4.06 (dd, J₁ = 10 Hz, J₂ = 8.4 Hz, 1 H, H-3a), 3.91–3.78 (m, 4 H), 3.74–3.45 (m, 6 H), 3.34–3.30 (m, 2 H, H-2a, H-1^b), 2.39 (t, J = 7.6 Hz, 2H, H-5^o), 1.74–1.64 (m, 4 H, H-2^o, H-4^o), 1.48–1.42 (m, 2 H, H-3^o), 1.03 (s, 9 H, 3 CH₃), 0.95 (s, 9 H, 3 CH₃). ¹³C-APT (CDCl₃, 100 MHz) δ 173.5 (C=O), 138.0, 137.9, 137.7, 136.2 (aromatic C), 128.7, 128.6, 128.3, 128.0, 127.96, 127.9, 127.86, 127.6 (aromatic CH), 97.9 (C-1b), 97.5 (C-1a), 81.0 (C-3a), 75.5 (C-3b), 74.3, 73.5 (PhCH₂), 72.4 (c-4a), 70.5 (PhCH₂), 70.1 (c-5a), 69.6 (C-4b), 69.1 (C-6), 68.3 (C-1^o), 67.9 (C-5b), 66.9 (C-6), 66.3 (PhCH₂), 63.6 (C-2a), 58.1 (C-2b), 34.3 (C-5^o), 29.2 (C-2^o), 27.7 (3 CH₃), 27.3 (3 CH₃), 25.8 (C-3^o), 24.8 (C-4^o), 23.4, 20.7. HRMS (ESI) m/z : $[M + NH_4]^+$ Calculated for C₅₄H₇₄N₇O₁₁Si: 1024.52101, found: 1024.52157.

Synthesis of Disaccharide 25. Compound **24** (1.16 g, 1.15 mmol) was dissolved in THF (11 mL) in a round flask. Then, HF-pyridine (300 μ L) was added in the solution. The reaction mixture was stirred until TLC analysis indicated full consumption of the starting material (30 min). Then, the mixture was diluted with DCM, and the reaction was quenched with saturated NaHCO₃. The organic phase was washed with water and brine, dried with anhydrous MgSO₄, filtered, and concentrated *in vacuo*. The product was purified by silica gel column chromatography (pentane:Et₂O = 3:1). Compound **25** (910 mg, 91% yield) was obtained as a colorless syrup. $[\alpha]_D^{20} +80.6$ (c = 1,

CHCl₃). IR (neat, cm⁻¹) ν 698, 738, 1040, 1145, 1262, 1354, 1455, 1733 (C=O), 2106 (N₃), 2872, 2932, 3461. ¹H NMR (CDCl₃, 400 MHz) δ 7.38–7.25 (m, 20 H, aromatic H), 5.65 (d, J = 3.6 Hz, 1 H, H-1b), 5.12 (s, 1 H, PhCH₂), 4.97 (d, J = 10.4 Hz, 1 H, CHH), 4.92 (d, J = 3.6 Hz, 1 H, H-1a), 4.88 (d, J = 10.4 Hz, 1 H, CHH), 4.67–4.54 (m, 4 H, 4 CHH), 4.05 (dd, J₁ = 10.0 Hz, J₂ = 8.4 Hz, 1 H, H-3a), 3.91–3.80 (m, 2 H, H-4a, H-5), 3.74–3.57 (m, 8 H), 3.51–3.44 (m, 1 H, H-1^b), 3.31 (dd, J₁ = 10.0 Hz, J₂ = 3.6 Hz, 1 H, H-2a), 2.65 (s, 1 H, OH), 2.39 (t, J = 7.6 Hz, 2H, H-5^o), 2.29 (s, 1 H, OH), 1.74–1.64 (m, 4 H, H-2^o, H-4^o), 1.47–1.40 (m, 2 H, H-3^o). ¹³C-APT (CDCl₃, 100 MHz) δ 173.60 (C=O), 138.1, 137.80, 137.1, 136.1 (aromatic C), 128.8, 128.7, 128.6, 128.5, 128.4, 128.31, 128.29, 128.1, 127.9, 127.8, 127.7 (aromatic CH), 97.9 (C-1b), 97.7 (C-1a), 80.8 (C-3a), 76.1 (C-3b), 74.5 (PhCH₂), 73.8 (c-4a), 73.7 (PhCH₂), 71.9 (PhCH₂), 70.3 (c-5b), 70.2 (C-5a), 69.7 (C-6), 68.3 (C-1^o), 67.2 (C-4b), 66.3 (PhCH₂), 63.8 (C-2a), 62.9 (C-6), 58.8 (C-2b), 34.3 (C-5^o), 29.2 (C-2^o), 25.8 (C-3^o), 24.8 (C-4^o). HRMS (ESI) m/z : $[M + NH_4]^+$ Calculated for C₄₆H₆₀N₇O₁₁: 884.41888, found: 884.41942.

Synthesis of Disaccharide Acceptor 26. The compound **25** (865 mg, 1.0 mmol) was dissolved in CH₃CN (10 mL). Then, BnBr (182 μ L), borinic acid catalyst (22 mg), K₂CO₃ (148 mg), and KI (166 mg) were added into the mixture. The reaction mixture was stirred at 60 °C in oil bath until TLC analysis indicated full consumption of the starting material (24 h). Then, the mixture was diluted with ethyl acetate, and the reaction was quenched with saturated NaHCO₃. The organic phase was washed with water and brine, dried with anhydrous MgSO₄, filtered, and concentrated *in vacuo*. The product was purified by silica gel column chromatography (pentane:Et₂O = 5:1). Compound **26** (910 mg, 95%) was obtained as a colorless syrup. $[\alpha]_D^{20} +66.6$ (c = 1, CHCl₃). IR (neat, cm⁻¹) ν 697, 736, 1040, 1096, 1259, 1455, 1734 (C=O), 2106 (N₃), 2869, 2928. ¹H NMR (CDCl₃, 400 MHz) δ 7.40–7.21 (m, 25 H, aromatic H), 5.64 (d, J = 3.6 Hz, 1 H, H-1b), 5.11 (s, 1 H, PhCH₂), 4.96 (d, J = 10.4 Hz, 1 H, CHH), 4.91 (d, J = 3.6 Hz, 1 H, H-1a), 4.88 (d, J = 10.4 Hz, 1 H, CHH), 4.63 (bs, 2 H, 2 CHH), 4.55 (d, J = 12.0 Hz, 1 H, CHH), 4.45 (d, J = 12.0 Hz, 1 H, CHH), 4.47–4.36 (m, 3 H, 3 CHH), 4.07–4.03 (m, 2 H, H-3a, H-5a), 3.91–3.44 (m, 12 H), 3.32 (dd, J₁ = 11.2 Hz, J₂ = 3.6 Hz, 1 H, H-2a), 2.65 (s, 1 H, OH), 2.38 (t, J = 7.6 Hz, 2H, H-5^o), 1.73–1.63 (m, 4 H, H-2^o, H-4^o), 1.47–1.39 (m, 2 H, H-3^o). ¹³C-APT (CDCl₃, 100 MHz) δ 173.5 (C=O), 138.3, 137.8, 137.3, 136.1 (aromatic C), 129.1, 128.7, 128.65, 128.54, 128.50, 128.4, 128.3, 128.0, 127.9, 127.8, 127.6, 127.6 (aromatic CH), 97.9 (C-1b), 97.7 (C-1a), 80.8 (C-3a), 76.3 (C-3b), 74.5 (PhCH₂), 73.9 (C-4a), 73.8, 73.3, 71.7 (PhCH₂), 70.1 (C-4b), 69.5, 69.4 (C-6), 69.2 (C-5b), 68.2 (C-1^o), 66.4 (C-5a), 66.2 (PhCH₂), 63.7 (C-2a), 58.9 (C-2b), 34.2 (C-5^o), 29.1 (C-2^o), 25.7 (C-3^o), 24.8 (C-4^o). HRMS (ESI) m/z : $[M + NH_4]^+$ Calculated for C₅₃H₆₄N₇O₁₁: 974.466583, found: 974.46660.

Synthesis of Tetrasaccharide 27. The reaction was carried out according to the standard procedure A. A mixture of donor **23** (520 mg, 0.49 mmol), acceptor **26** (238 mg, 0.25 mmol) (donors and acceptors coevaporated with toluene three times), and MPF (490 μ L) in dry DCM (1 mL) was stirred over fresh flame-dried 3 Å molecular sieves under nitrogen. The solution was cooled to –78 °C, after which TfOH (40 μ L) was added. After 30 min, the reaction was stirred at –10 °C until TLC analysis showed complete conversion of the acceptor. The reaction was quenched with Et₃N, filtered, and concentrated *in vacuo*. The product was purified by size exclusion (DCM:MeOH = 1:1). Compound **27** (280 mg, 89%, α : β = 10:1) was obtained as a colorless syrup. IR (neat, cm⁻¹) ν 697, 736, 1028, 1096, 1258, 1319, 1356, 1454, 1497, 1731 (C=O), 2105 (N₃), 2868, 2925. ¹H NMR (CDCl₃, 400 MHz) δ 7.77–7.69 (m, 3 H, aromatic H), 7.60 (bs, 1 H, aromatic H), 7.42–7.17 (m, 48 H, aromatic H), 5.73 (d, J = 3.6 Hz, 1 H, H-1d), 5.65 (d, J = 3.6 Hz, 1 H, H-1b), 5.10 (s, 1 H, PhCH₂), 4.96–4.90 (m, 7 H), 4.74–4.48 (m, 7 H), 4.36–3.61 (m, 22 H), 3.51–3.26 (m, 6 H), 3.20 (d, J = 10.0 Hz, 1 H), 3.03 (d, J = 10.0 Hz, 1 H), 2.36 (t, J = 7.6 Hz, 2H, H-5^o), 1.71–1.61 (m, 4 H, H-2^o, H-4^o), 1.45–1.37 (m, 2 H, H-3^o). ¹³C-APT (CDCl₃, 100 MHz) δ 173.4 (C=O), 138.4, 138.2, 137.7, 137.68, 137.6, 137.54, 137.47, 136.1, 135.6, 133.1, 133.0 (aromatic C), 128.6, 128.5, 128.4, 128.38, 128.35,

128.27, 128.25, 128.2, 128.1, 128.07, 128.0, 127.9, 127.88, 127.86, 127.78, 127.76, 127.66, 127.6, 127.4, 127.3, 127.1, 127.0, 126.4, 126.0, 125.9 (aromatic CH), 98.7 (C-1), 98.1 (C-1), 97.8 (C-1), 97.7 (C-1), 81.0 (C-3), 80.7 (C-3), 76.8 (C-3), 75.1 (C-3), 74.9, 74.5, 74.4 (CH₂), 73.5 (C-4), 73.5, 73.3 (CH₂), 73.2 (C-4), 72.9 (C-4), 72.8 (C-4), 72.8, 72.2, 71.7 (CH₂), 70.6 (C-5), 70.1 (C-5), 69.9 (C-5), 69.8 (C-5), 69.2, 68.4, 68.1, 67.9, 66.8, 66.1 (CH₂), 64.7 (C-2), 63.7 (C-2), 59.5 (C-2), 59.4 (C-2), 34.1 (C-5°), 29.0 (C-2°), 25.6 (C-3°), 24.7 (C-4°). HRMS (ESI) *m/z*: [M + NH₄]⁺ Calculated for C₁₀₄H₁₁₄N₁₃O₁₉: 1848.83484, found: 1848.83541.

Synthesis of Tetrasaccharide Acceptor 28. The reaction was carried out according to the standard procedure C. Compound 27 (700 mg, 0.38 mmol) was dissolved in DCM:HFIP (1:1, 0.1 M). TES (304 μL, 1.91 mmol) and 0.2 M HCl/HFIP (1.9 mL) were added to the mixture. The reaction mixture was stirred until TLC analysis indicated full consumption of the starting material (15 min). Then, the mixture was diluted with DCM, and the reaction was quenched with saturated NaHCO₃. The organic phase was washed with water and brine, dried with anhydrous MgSO₄, filtered, and concentrated *in vacuo*. The product was purified by silica gel column chromatography (pentane:Et₂O = 5:1). Compound 28 (297 mg, 73% yield) was obtained as a colorless syrup. [α]_D²⁰ +106.7 (c = 1, CHCl₃). IR (neat, cm⁻¹) ν 696, 737, 1040, 1100, 1261, 1454, 1735 (C=O), 2106 (N₃), 2869, 2926. ¹H NMR (CDCl₃, 500 MHz) δ 7.42–7.17 (m, 45 H, aromatic H), 5.71 (d, *J* = 3.5 Hz, 1 H, H-1d), 5.65 (d, *J* = 3.5 Hz, 1 H, H-1b), 5.10 (s, 1 H, PhCH₂), 4.95–4.89 (m, 6 H, H-1a, H-1c, 4 CHH), 4.72 (d, *J* = 12.0 Hz, 1 H, CHH), 4.68 (s, 2 H, 2 CHH), 4.57–4.48 (m, 3 H), 4.36–4.17 (m, 7 H), 4.09–3.58 (m, 17 H), 3.51–3.30 (m, 3 H), 3.36–3.30 (m, 3 H), 3.91–3.44 (m, 12 H), 3.17 (dd, *J*₁ = 11.5 Hz, *J*₂ = 2.5 Hz, 1 H), 3.02 (dd, *J*₁ = 11.5 Hz, *J*₂ = 2.5 Hz, 1 H), 2.66 (s, 1 H, OH), 2.37 (t, *J* = 7.5 Hz, 2H, H-5°), 1.71–1.63 (m, 4 H, H-2°, H-4°), 1.45–1.39 (m, 2 H, H-3°). ¹³C-APT (CDCl₃, 125 MHz) δ 173.5 (C=O), 138.5, 138.3, 137.8, 137.7, 137.68, 137.6, 137.4, 136.2 (aromatic C), 128.8, 128.7, 128.6, 128.6, 128.5, 128.3, 128.2, 128.18, 128.0, 127.9, 127.9, 127.86, 127.8, 127.7, 127.69, 127.5, 127.5, 127.3, 127.2 (aromatic CH), 98.8 (C-1), 98.1 (C-1), 97.9 (C-1), 97.8 (C-1), 80.9 (C-3), 80.8 (C-3), 76.1 (C-3), 75.8 (C-3), 74.54, 74.46, 73.7 (CH₂), 73.6 (C-4), 73.56, 73.4 (CH₂), 73.1 (2 C-4), 73.07, 71.8, 71.75 (CH₂), 70.7 (C-4), 70.2 (C-5), 69.9 (C-5), 69.3, 69.1 (CH₂), 68.8 (C-5), 68.5, 68.3, 68.9 (CH₂), 66.5 (C-5), 66.3 (PhCH₂), 64.7 (C-2), 63.8 (C-2), 59.6 (C-2), 58.8 (C-2), 34.3 (C-5°), 29.2 (C-2°), 25.8 (C-3°), 24.8 (C-4°). HRMS (ESI) *m/z*: [M + NH₄]⁺ Calculated for C₉₃H₁₀₆N₁₃O₁₉: 1708.77224, found: 1708.77299.

Synthesis of Hexasaccharide 29. The reaction was carried out according to the standard procedure A. A mixture of donor 23 (540 mg, 0.5 mmol), acceptor 28 (360 mg, 0.21 mmol) (donors and acceptors coevaporated with toluene three times), and MPF (400 μL) in dry DCM (1 mL) was stirred over fresh flame-dried 3 Å molecular sieves under nitrogen. The solution was cooled to –78 °C, after which TfOH (44 μL) was added. After 30 min, the reaction was stirred at –10 °C until TLC analysis showed complete conversion of the acceptor (48 h). The reaction was quenched with Et₃N, filtered, and concentrated *in vacuo*. The product was purified by size exclusion (DCM:MeOH = 1:1). Compound 29 (500 mg, 91%, α:β = 10:1) was obtained as a colorless syrup. [α]_D²⁰ +123.0 (c = 1, CHCl₃). IR (neat, cm⁻¹) ν 697, 736, 1039, 1099, 1261, 1319, 1359, 1454, 1734 (C=O), 2106 (N₃), 2870, 2926. ¹H NMR (CDCl₃, 400 MHz) δ 7.77–7.69 (m, 3 H, aromatic H), 7.60 (bs, 1 H, aromatic H), 7.44–7.07 (m, 68 H, aromatic H), 5.73 (d, *J* = 3.6 Hz, 1 H, H-1), 5.71 (d, *J* = 3.2 Hz, 1 H, H-1), 5.65 (d, *J* = 3.6 Hz, 1 H, H-1), 5.09 (s, 1 H, PhCH₂), 4.96–4.91 (m, 9 H), 4.77–4.65 (m, 5 H), 4.58–4.48 (m, 4 H), 4.33–3.62 (m, 37 H), 3.52–3.16 (m, 10 H), 3.05 (d, *J* = 10.0 Hz, 1 H), 2.99 (d, *J* = 10.0 Hz, 1 H), 2.36 (t, *J* = 7.6 Hz, 2H, H-5°), 1.71–1.61 (m, 4 H, H-2°, H-4°), 1.45–1.37 (m, 2 H, H-3°). ¹³C-APT (CDCl₃, 100 MHz) δ 173.4 (C=O), 138.5, 138.4, 138.2, 137.7, 137.7, 137.6, 137.59, 137.57, 137.5, 137.4, 136.1, 135.7, 133.2, 133.0 (aromatic C), 128.6, 128.5, 128.49, 128.48, 128.45, 128.39, 128.36, 128.3, 128.2, 128.1, 128.05, 128.1, 128.0, 127.94, 127.90, 127.86, 127.82, 127.78, 127.77, 127.72, 127.67, 127.6, 127.42, 127.35, 127.3, 127.12, 127.11,

127.03, 127.0, 126.4, 126.0, 125.9 (aromatic CH), 98.8 (C-1), 98.7 (C-1), 98.1 (C-1), 97.9 (C-1), 97.8 (C-1), 97.7 (C-1), 80.9 (C-3), 80.8 (C-3), 80.7 (C-3), 76.9 (C-3), 76.1 (C-3), 75.7 (C-3), 74.9, 74.5, 74.4, 73.5, 73.3 (CH₂), 73.3 (C-4), 73.2 (C-4), 73.0 (CH₂), 72.83 (C-4), 72.80 (CH₂), 72.7 (C-4), 72.2, 71.9, 71.7 (CH₂), 70.6 (C-5), 70.5 (C-5), 70.1 (C-5), 69.9 (C-5), 69.8 (C-5), 69.6 (C-5), 69.2, 68.3, 68.1, 67.9, 66.8, 66.5, 66.1 (CH₂), 64.7 (2 C-2), 63.7 (C-2), 59.5 (C-2), 59.47 (C-2), 59.4 (C-2), 34.2 (C-5°), 29.1 (C-2°), 25.75 (C-3°), 24.7 (C-4°).

Synthesis of Hexasaccharide 30. Compound 29 (20 mg, 0.0078 mmol) was dissolved in THF/H₂O/*tert*-BuOH (2 mL/2 mL/1 mL) before a catalytic amount of Pd(OH)₂/C was added. The reaction mixture was stirred for 3 days under a H₂ atmosphere, filtered, and concentrated *in vacuo*. A white powder 30 (6.7 mg, 76%) was obtained after purification by gel filtration (HW-40, 0.15 M NH₄OAc in H₂O). ¹H NMR (D₂O, 500 MHz) δ 5.40–5.35 (m, 3 H, 3 H-1), 4.85–4.81 (m, 3 H, 3 H-1), 4.13–4.06 (m, 2 H), 3.97–3.50 (m, 35 H), 3.41–3.37 (m, 1 H), 3.15–3.08 (m, 2 H), 2.81 (dd, 2 H), 2.72–2.70 (m, 2 H), 2.06 (t, 2 H), 1.55–1.43 (m, 5 H), 1.29–1.23 (m, 2 H). ¹³C-APT (CDCl₃, 125 MHz) δ 99.6 (C-1), 99.5 (C-1), 99.5 (C-1), 99.2 (2 C-1), 97.4 (C-1), 76.9, 76.7, 76.6, 76.7, 73.7, 73.3, 72.4, 72.3, 71.9, 71.1, 71.0, 70.5, 69.2, 68.8, 68.4, 68.3, 61.2, 60.6, 60.4, 55.3, 55.3, 54.6, 51.1, 51.1, 51.0, 37.5, 28.3, 25.7, 25.4. HRMS (ESI) *m/z*: [M + 2H]⁺/2 Calculated for C₄₂H₈₀N₆O₂₇: 550.25302; found: 550.25247.

Synthesis of Hexasaccharide 31. Compound 30 (5 mg) was dissolved in H₂O. Then, Ac₂O and NaHCO₃ were added in the solution. The reaction mixture was stirred for 3 days until TLC analysis showed complete conversion of the starting materials. The product was purified by gel filtration (HW-40, 0.15 M NH₄OAc in H₂O). Compound 31 (5.5 mg, 86%) was obtained as a white solid. ¹H NMR (D₂O, 500 MHz) δ 5.36–5.34 (m, 4 H, 4 H-1), 5.27 (d, *J* = 4.0 Hz, 1 H, H-1), 4.79 (bt, 2 H, 2 H-1), 4.73 (d, *J* = 3.0 Hz, 1 H, H-1), 4.21–4.06 (m, 5 H), 3.97–3.57 (m, 37 H), 3.40–3.35 (m, 1 H), 2.24 (bt, 2 H), 1.97–1.91 (m, 18 H, 6 CH₃), 1.80–1.74 (m, 2 H), 1.54–1.46 (m, 4 H), 1.32–1.26 (m, 2 H). ¹³C-APT (CDCl₃, 125 MHz) δ 180.2, 174.7, 174.69, 174.6, 174.5, 174.46 (6 C=O), 98.2, 98.15, 96.6 (6 C-1), 77.4, 77.2, 76.1, 75.7, 75.4, 72.5, 72.4, 71.8, 71.7, 71.3, 71.2, 70.7, 70.7, 70.4, 68.5, 68.1, 67.9, 67.6, 66.9, 61.6, 60.7, 60.3, 60.0, 54.4, 54.2, 49.9, 34.6, 28.2, 25.0, 24.4, 22.1 (CH₃), 22.0 (CH₃), 22.0 (CH₃), 21.9 (CH₃), 21.9 (2 CH₃). HRMS (ESI) *m/z*: [M + 2H]⁺/2 Calculated for C₅₄H₉₂O₃₃N₆: 676.28472; found: 676.28489.

■ ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acs.joc.0c00703>.

Full experimental details and characterization and NMR spectra of all new compounds (PDF)

■ AUTHOR INFORMATION

Corresponding Author

Jeroen D. C. Codée – Leiden Institute of Chemistry, Leiden University, 2333 CC Leiden, The Netherlands; orcid.org/0000-0003-3531-2138; Email: jcodee@chem.leidenuniv.nl

Authors

Liming Wang – Leiden Institute of Chemistry, Leiden University, 2333 CC Leiden, The Netherlands

Yongzhen Zhang – Leiden Institute of Chemistry, Leiden University, 2333 CC Leiden, The Netherlands

Herman S. Overkleeft – Leiden Institute of Chemistry, Leiden University, 2333 CC Leiden, The Netherlands; orcid.org/0000-0001-6976-7005

Gijsbert A. van der Marel – Leiden Institute of Chemistry, Leiden University, 2333 CC Leiden, The Netherlands

Complete contact information is available at:
<https://pubs.acs.org/10.1021/acs.joc.0c00703>

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

This work was supported by the Chinese Scholarship Council (CSC grant to L.W.) and the European Research Council (ERC-CoG-726072-“GLYCONTROL”, to J.D.C.C.).

REFERENCES

- (1) (a) Ramsey, D. M.; Wozniak, D. J. Understanding the control of *Pseudomonas aeruginosa* alginate synthesis and the prospects for management of chronic infections in cystic fibrosis. *Mol. Microbiol.* **2005**, *56*, 309–322. (b) Jennings, L. K.; Storek, K. M.; Ledvina, H. E.; Coulon, C.; Marmont, L. S.; Sadvovskaya, I.; Secor, P. R.; Tseng, B. S.; Scian, M.; Filloux, A.; Wozniak, D. J.; Howell, P. L.; Parsek, M. R. Pel is a cationic exopolysaccharide that cross-links extracellular DNA in the *Pseudomonas aeruginosa* biofilm matrix. *Proc. Natl. Acad. Sci. U. S. A.* **2015**, *112*, 11353–11358.
- (2) (a) Bamford, N. C.; Le Mauff, F.; Subramanian, A. S.; Yip, P.; Millán, C.; Zhang, Y.; Zacharias, C.; Forman, A.; Nitz, M.; Codée, J. D. C.; Usón, I.; Sheppard, D. C.; Howell, P. L. Ega3 from the fungal pathogen *Aspergillus fumigatus* is an endo- α -1,4-galactosaminidase that disrupts microbial biofilms. *J. Biol. Chem.* **2019**, *294*, 13833–13849. (b) Le Mauff, F.; Bamford, N. C.; Alnabelseya, N.; Zhang, Y.; Baker, P.; Robinson, H.; Codée, J. D. C.; Howell, P. L.; Sheppard, D. C. Molecular mechanism of *Aspergillus fumigatus* biofilm disruption by fungal and bacterial glycoside hydrolases. *J. Biol. Chem.* **2019**, *294*, 10760–10772. (c) Kazakova, E. D.; Yashunsky, D. V.; Krylov, V. B.; Bouchara, J.-P.; Cornet, M.; Valsecchi, I.; Fontaine, T.; Latgé, J.-P.; Nifantiev, N. E. Biotinylated Oligo- α -(1 \rightarrow 4)-d-galactosamines and Their N-Acetylated Derivatives: α -Stereoselective Synthesis and Immunology Application. *J. Am. Chem. Soc.* **2020**, *142*, 1175–1179.
- (3) (a) Imamura, A.; Matsuzawa, N.; Sakai, S.; Udagawa, T.; Nakashima, S.; Ando, H.; Ishida, H.; Kiso, M. The Origin of High Stereoselectivity in Di-tert-butylsilylene-Directed α -Galactosylation. *J. Org. Chem.* **2016**, *81*, 9086–9104. (b) Imamura, A.; Ando, H.; Korogi, S.; Tanabe, G.; Muraoka, O.; Ishida, H.; Kiso, M. Di-tert-butylsilylene (DTBS) group-directed α -selective galactosylation unaffected by C-2 participating functionalities. *Tetrahedron Lett.* **2003**, *44*, 6725–6728.
- (4) (a) Polat, T.; Wong, C.-H. Anomeric Reactivity-Based One-Pot Synthesis of Heparin-Like Oligosaccharides. *J. Am. Chem. Soc.* **2007**, *129*, 12795–12800. (b) Hu, Y.-P.; Lin, S.-Y.; Huang, C.-Y.; Zulueta, M. M. L.; Liu, J.-Y.; Chang, W.; Hung, S.-C. Synthesis of 3-O-sulfonated heparan sulfate octasaccharides that inhibit the herpes simplex virus type 1 host–cell interaction. *Nat. Chem.* **2011**, *3*, 557–563. (c) Zulueta, M. M. L.; Lin, S.-Y.; Lin, Y.-T.; Huang, C.-J.; Wang, C.-C.; Ku, C.-C.; Shi, Z.; Chyan, C.-L.; Irene, D.; Lim, L.-H.; Tsai, T.-I.; Hu, Y.-P.; Arco, S. D.; Wong, C.-H.; Hung, S.-C. α -Glycosylation by D-Glucosamine-Derived Donors: Synthesis of Heparosan and Heparin Analogues That Interact with Mycobacterial Heparin-Binding Hemagglutinin. *J. Am. Chem. Soc.* **2012**, *134*, 8988–8995. (d) Yoshida, K.; Yang, B.; Yang, W.; Zhang, Z.; Zhang, J.; Huang, X. Chemical Synthesis of Syndecan-3 Glycopeptides Bearing Two Heparan Sulfate Glycan Chains. *Angew. Chem., Int. Ed.* **2014**, *53*, 9051–9058. (e) Chang, C.-H.; Lico, L. S.; Huang, T.-Y.; Lin, S.-Y.; Chang, C.-L.; Arco, S. D.; Hung, S.-C. Synthesis of the Heparin-Based Anticoagulant Drug Fondaparinux. *Angew. Chem., Int. Ed.* **2014**, *53*, 9876–9879.
- (5) (a) Codée, J. D. C.; Stubba, B.; Schiattarella, M.; Overkleeft, H. S.; van Boeckel, C. A. A.; van Boom, J. H.; van der Marel, G. A. A Modular Strategy Toward the Synthesis of Heparin-like Oligosaccharides Using Monomeric Building Blocks in a Sequential Glycosylation Strategy. *J. Am. Chem. Soc.* **2005**, *127*, 3767–3773. (b) Arungundram, S.; Al-Mafraji, K.; Asong, J.; Leach, F. E.; Amster, I. J.; Venot, A.; Turnbull, J. E.; Boons, G.-J. Modular Synthesis of Heparan Sulfate Oligosaccharides for Structure-Activity Relationship Studies. *J. Am. Chem. Soc.* **2009**, *131*, 17394–17405. (c) Orgueira, H. A.; Bartolozzi, A.; Schell, P.; Seeberger, P. H. Conformational Locking of the Glycosyl Acceptor for Stereocontrol in the Key Step in the Synthesis of Heparin. *Angew. Chem., Int. Ed.* **2002**, *41*, 2128–2131. (d) Park, J.; Kawatkar, S.; Kim, J.-H.; Boons, G.-J. Stereoselective Glycosylations of 2-Azido-2-deoxy-glucosides Using Intermediate Sulfonium Ions. *Org. Lett.* **2007**, *9*, 1959–1962. (e) Hagen, B.; van Dijk, J. H. M.; Zhang, Q.; Overkleeft, H. S.; van der Marel, G. A.; Codée, J. D. C. Synthesis of the *Staphylococcus aureus* Strain M Capsular Polysaccharide Repeating Unit. *Org. Lett.* **2017**, *19*, 2514–2517. (f) Zhang, Y.; Zhang, H.; Zhao, Y.; Guo, Z.; Gao, J. Efficient Strategy for α -Selective Glycosidation of D-Glucosamine and Its Application to the Synthesis of a Bacterial Capsular Polysaccharide Repeating Unit Containing Multiple α -Linked GlcNAc Residues. *Org. Lett.* **2020**, *22*, 1520–1524.
- (6) (a) Casu, B. In *Structure and Biological Activity of Heparin*, Vol. 43; Tipson, R. S., Horton, D., Eds.; Academic Press, 1985; pp 51–134. (b) Turnbull, J.; Powell, A.; Guimond, S. Heparan sulfate: decoding a dynamic multifunctional cell regulator. *Trends Cell Biol.* **2001**, *11*, 75–82. (c) Gandhi, N. S.; Mancera, R. L. The structure of glycosaminoglycans and their interactions with proteins. *Chem. Biol. Drug Des.* **2008**, *72*, 455–482.
- (7) (a) Wei, P.; Kerns, R. J. Factors Affecting Stereocontrol during Glycosidation of 2,3-Oxazolidinone-Protected 1-Tolylthio-N-acetyl-D-glucosamine. *J. Org. Chem.* **2005**, *70*, 4195–4198. (b) Li, J.; Dai, Y.; Li, W.; Laval, S.; Xu, P.; Yu, B. Effective Synthesis of α -D-GlcN-(1 \rightarrow 4)-D-GlcA/L-IdoA Glycosidic Linkage under Gold(I) Catalysis. *Asian J. Org. Chem.* **2015**, *4*, 756–762. (c) Ingle, A. B.; Chao, C.-S.; Hung, W.-C.; Mong, K.-K. T. Tuning Reactivity of Glycosyl Iminium Intermediate for 2-Azido-2-deoxyglycosyl Donors in α -Glycosidic Bond Formation. *Org. Lett.* **2013**, *15*, 5290–5293. (d) Manabe, S.; Ishii, K.; Ito, Y. N-Benzyl-2,3-oxazolidinone as a Glycosyl Donor for Selective α -Glycosylation and One-Pot Oligosaccharide Synthesis Involving 1,2-cis-Glycosylation. *J. Am. Chem. Soc.* **2006**, *128*, 10666–10667. (e) Mensah, E. A.; Nguyen, H. M. Nickel-Catalyzed Stereoselective Formation of α -2-Deoxy-2-Amino Glycosides. *J. Am. Chem. Soc.* **2009**, *131*, 8778–8780. (f) Amin, M. N.; Ishiwata, A.; Ito, Y. Synthesis of N-linked glycan derived from Gram-negative bacterium, *Campylobacter jejuni*. *Tetrahedron* **2007**, *63*, 8181–8198.
- (8) (a) Oka, N.; Kajino, R.; Takeuchi, K.; Nagakawa, H.; Ando, K. α -Selective Ribofuranosylation of Alcohols with Ribofuranosyl Iodides and Triphenylphosphine Oxide. *J. Org. Chem.* **2014**, *79*, 7656–7664. (b) Kobashi, Y.; Mukaiyama, T. Glycosyl Phosphonium Halide as a Reactive Intermediate in Highly α -Selective Glycosylation. *Bull. Chem. Soc. Jpn.* **2005**, *78*, 910–916. (c) Mukaiyama, T.; Kobashi, Y. Highly α -Selective Synthesis of Disaccharide Using Glycosyl Bromide by the Promotion of Phosphine Oxide. *Chem. Lett.* **2004**, *33*, 10–11. (d) Kobashi, Y.; Mukaiyama, T. Highly α -Selective Glycosylation with Glycosyl Acetate via Glycosyl Phosphonium Iodide. *Chem. Lett.* **2004**, *33*, 874–875. (e) Lu, S. R.; Lai, Y. H.; Chen, J. H.; Liu, C. Y.; Mong, K. K. Dimethylformamide: an unusual glycosylation modulator. *Angew. Chem., Int. Ed.* **2011**, *50*, 7315–7320. (f) Dourtoglou, V.; Gross, B. Activation de la Ponction Hydroxyle Anomere via les Sels d’Iminium. O-Glycosylation des Tri-O-benzyl-2,3,5-D-arabino et -ribotorannoses. *J. Carbohydr. Chem.* **1983**, *2*, 57–73. (g) Zeng, J.; Wang, R.; Zhang, S.; Fang, J.; Liu, S.; Sun, G.; Xu, B.; Xiao, Y.; Fu, D.; Zhang, W.; Hu, Y.; Wan, Q. Hydrogen-Bonding-Assisted Exogenous Nucleophilic Reagent Effect for β -Selective Glycosylation of Rare 3-Amino Sugars. *J. Am. Chem. Soc.* **2019**, *141*, 8509–8515.
- (9) (a) Ngoje, G.; Li, Z. Study of the stereoselectivity of 2-azido-2-deoxyglucosyl donors: protecting group effects. *Org. Biomol. Chem.* **2013**, *11*, 1879–1886. (b) van der Vorm, S.; Overkleeft, H. S.; van der Marel, G. A.; Codee, J. D. C. Stereoselectivity of Conformationally Restricted Glucosazide Donors. *J. Org. Chem.* **2017**, *82*, 4793–4811. (c) Lourenco, E. C.; Ventura, M. R. Improvement of the stereoselectivity of the glycosylation reaction with 2-azido-2-deoxy-1-

thioglycoside donors. *Carbohydr. Res.* **2016**, *426*, 33–39. (d) van der Vorm, S.; Hansen, T.; van Hengst, J. M. A.; Overkleeft, H. S.; van der Marel, G. A.; Codée, J. D. C. Acceptor reactivity in glycosylation reactions. *Chem. Soc. Rev.* **2019**, *48*, 4688–4706. (e) Codée, J. D. C.; Litjens, R. E. J. N.; van den Bos, L. J.; Overkleeft, H. S.; van der Marel, G. A. Thioglycosides in sequential glycosylation strategies. *Chem. Soc. Rev.* **2005**, *34*, 769–782. (f) Zhang, Z.; Ollmann, I. R.; Ye, X.-S.; Wischnat, R.; Baasov, T.; Wong, C.-H. Programmable One-Pot Oligosaccharide Synthesis. *J. Am. Chem. Soc.* **1999**, *121*, 734–753. (g) Douglas, N. L.; Ley, S. V.; Lücking, U.; Warriner, S. L. Tuning glycoside reactivity: New tool for efficient oligosaccharide synthesis. *J. Chem. Soc., Perkin Trans. 1* **1998**, 51–66. (h) Andrews, C. W.; Rodebaugh, R.; Fraser-Reid, B. A Solvation-Assisted Model for Estimating Anomeric Reactivity. Predicted versus Observed Trends in Hydrolysis of n-Pentenyl Glycosides. *J. Org. Chem.* **1996**, *61*, 5280–5289.

(10) (a) Hagen, B.; Ali, S.; Overkleeft, H. S.; van der Marel, G. A.; Codée, J. D. C. Mapping the Reactivity and Selectivity of 2-Azidofucosyl Donors for the Assembly of N-Acetylglucosamine-Containing Bacterial Oligosaccharides. *J. Org. Chem.* **2017**, *82*, 848–868. (b) van der Vorm, S.; Hansen, T.; Overkleeft, H. S.; van der Marel, G. A.; Codée, J. D. C. The influence of acceptor nucleophilicity on the glycosylation reaction mechanism. *Chem. Sci.* **2017**, *8*, 1867–1875. (c) van der Vorm, S.; Hansen, T.; van Hengst, J. M. A.; Overkleeft, H. S.; van der Marel, G. A.; Codée, J. D. C. Acceptor reactivity in glycosylation reactions. *Chem. Soc. Rev.* **2019**, *48*, 4688–4706.

(11) (a) Wang, L.; Overkleeft, H. S.; van der Marel, G. A.; Codée, J. D. C. Reagent Controlled Stereoselective Synthesis of α -Glucans. *J. Am. Chem. Soc.* **2018**, *140*, 4632–4638. (b) Wang, L.; Overkleeft, H. S.; van der Marel, G. A.; Codée, J. D. C. Reagent Controlled Stereoselective Assembly of α -(1,3)-Glucans. *Eur. J. Org. Chem.* **2019**, *10*, 1994–2003.

(12) To shed light on the mode of action of MPF, a low temperature NMR experiment was conducted to study the formed reactive intermediates upon activation of donor **13**. Diagnostic signals were observed for the anomeric protons corresponding to the α/β -imidinium ions.

(13) Volbeda, A. G.; Kistemaker, H. A. V.; Overkleeft, H. S.; van der Marel, G. A.; Filippov, D. V.; Codée, J. D. C. Chemoselective Cleavage of p-Methoxybenzyl and 2-Naphthylmethyl Ethers Using a Catalytic Amount of HCl in Hexafluoro-2-propanol. *J. Org. Chem.* **2015**, *80*, 8796–8806.

(14) (a) Chan, L.; Taylor, M. S. Regioselective Alkylation of Carbohydrate Derivatives Catalyzed by a Diarylborinic Acid Derivative. *Org. Lett.* **2011**, *13*, 3090–3093. (b) Lee, D.; Taylor, M. S. Borinic Acid-Catalyzed Regioselective Acylation of Carbohydrate Derivatives. *J. Am. Chem. Soc.* **2011**, *133*, 3724–3727. (c) Lee, D.; Williamson, C. L.; Chan, L.; Taylor, M. S. Regioselective, Borinic Acid-Catalyzed Monoacylation, Sulfonylation and Alkylation of Diols and Carbohydrates: Expansion of Substrate Scope and Mechanistic Studies. *J. Am. Chem. Soc.* **2012**, *134*, 8260–8267.

(15) (a) Hansen, S. U.; Miller, G. J.; Barath, M.; Broberg, K. R.; Avizienyte, E.; Helliwell, M.; Raftery, J.; Jayson, G. C.; Gardiner, J. M. Synthesis and Scalable Conversion of L-Iduronamides to Heparin-Related Di- and Tetrasaccharides. *J. Org. Chem.* **2012**, *77*, 7823–7843. (b) Dietrich, H.; Espinosa, J. F.; Chiara, J. L.; Jimenez-Barbero, J.; Leon, Y.; Varela-Nieto, I.; Mato, J.-M.; Cano, F. H.; Foces-Foces, C.; Martin-Lomas, M. Glycosyl Inositol Derivatives Related to Inositolphosphoglycan Mediators: Synthesis, Structure, and Biological Activity. *Chem. - Eur. J.* **1999**, *5*, 320–336. (c) Czernecki, S.; Ayadi, E. Preparation of diversely protected 2-azido-2-deoxyglycopyranoses from glycals. *Can. J. Chem.* **1995**, *73* (3), 343–350.