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# High-affinity antigen association to cationic liposomes via coiled coil-forming peptides induces a strong antigen-specific CD4<sup>+</sup> T-cell response

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## ABSTRACT

Liposomes are widely investigated as vaccine delivery systems, but antigen loading efficiency can be low. Moreover, adsorbed antigen may rapidly desorb under physiological conditions. Encapsulation of antigens overcomes the latter problem but results in significant antigen loss during preparation and purification of the liposomes. Here, we propose an alternative attachment method, based on a complementary heterodimeric coiled coil peptide pair pepK and pepE.

PepK was conjugated to cholesterol (yielding CPK) and pepE was covalently linked to model antigen OVA323 (yielding pepE-OVA323). CPK was incorporated in the lipid bilayer of cationic liposomes (180 nm in size). Antigen was associated more efficiently to functionalized liposomes (Kd 166 nM) than to cationic liposomes (Kd not detectable). *In vivo* co-localization of antigen and liposomes was strongly increased upon CPK-functionalization (35% → 80%). CPK-functionalized liposomes induced 5-fold stronger CD4<sup>+</sup> T-cell proliferation than non-functionalized liposomes *in vitro*. Both formulations were able to induce strong CD4<sup>+</sup> T-cell expansion in mice, but more IFN- $\gamma$  and IL-10 production was observed after immunization with functionalized liposomes. In conclusion, antigen association via coiled coil peptide pair increased co-localization of antigen and liposomes, increased CD4<sup>+</sup> T-cell proliferation *in vitro* and induced a stronger CD4<sup>+</sup> T-cell response *in vivo*.

## 1. Introduction

The latest generation of vaccines moves away from whole pathogens and instead uses pathogen-derived proteins or peptides (i.e., subunits) as antigen. Subunit based vaccines are safer than whole pathogen-based vaccines but require adjuvants because these antigens alone are poorly immunogenic. Liposomes, vesicles composed of a phospholipid bilayer, are a widely investigated adjuvant because of their versatility and

proven success [1–4]. The lipid composition directs the physico-chemical properties of the liposomes, such as size, zeta potential, membrane fluidity and rigidity [1,3]. These properties greatly influence their behavior upon injection and the immune response they may induce [3,5–8]. Cationic liposomes have been used extensively as vaccine adjuvants. They are known to induce strong T-cell expansion and pro-inflammatory T helper (Th) 1-skewed immune response [9–14].

For a maximum adjuvant effect, the adjuvant and antigen need to be

**Abbreviations:** ACK, ammonium-chloride-potassium; ACN, acetonitrile; APC, antigen-presenting cell; BMDC, Bone marrow-derived dendritic cells; CC, coiled coil; CD, far UV circular dichroism spectroscopy; Chol, cholesterol; DC, dendritic cell; DCM, dichloromethane; DiD, 1,1'-dioctadecyl-3,3,3',3'-tetramethylindodicarbocyanine 4-chlorobenzenesulfonate; DIPEA, N,N-diisopropylethylamine; DLS, dynamic light scattering; DOTAP, 1,2-dioleoyl-3-trimethylammonium-propane; dpf, days post fertilization; DSPC, 1,2-distearoyl-sn-glycero-3-phosphocholine; EDTA, ethylenediaminetetraacetic acid; FCS, fetal calf serum; FOXP3, forkhead box P3; GM-CSF, granulocyte-macrophage colony-stimulating factor; hpi, hours post injection; IFN- $\gamma$ , interferon gamma; IL, interleukin; IMDM, Iscove's Modified Dulbecco's Medium; ITC, isothermal titration calorimetry; LN, lymph nodes; MFI, mean fluorescence intensity; MHC, major histocompatibility complex; MW, molecular weight; Oxyma, ethyl cyanohydroxyiminoacetate; PBS, phosphate-buffered saline; PDI, polydispersity index; PenStrep, penicillin-streptomycin; PMA, phorbol 12-myristate 13-acetate; ROR $\gamma$ T, retinoic acid receptor-related orphan receptor gamma; RPMI 1640, Roswell Park Memorial Institute Medium; TFA, trifluoroacetic acid; Th, T helper.

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taken up by the same antigen presenting cell. This is most efficiently achieved when the antigen and the adjuvant are physically or chemically associated [15,16]. Antigens can be associated with liposomes in various ways. For instance, hydrophilic molecules can be encapsulated in the aqueous core, whereas lipophilic molecules can be incorporated in the lipid bilayer. Although encapsulation usually ensures sustained colocalization, the encapsulation of an antigen in liposomes can be a challenging and costly process. Very few examples have shown 100% encapsulation efficiency, which oftentimes means that a large amount (up to 99%) of a valuable antigen is lost during liposome preparation and purification [10,17–19]. Moreover, the encapsulation process has to be optimized for each new antigen, and might require a change of composition of the liposomes, thereby potentially sacrificing adjuvant potential [19,20]. A more straightforward method is association of antigens on the outside of the liposome, which could be achieved by simple mixing of the antigen with preformed liposomes. Administration of antigens adsorbed to liposomes, via electrostatic interaction, has been shown to enhance the induced immune response compared to administration of plain antigen [12,21]. However, the degree of association may vary between antigens and formulations [22–25]. Moreover, after administration of liposomes with electrostatically adsorbed antigen, the antigen may rapidly diffuse away from the liposomes because of competition between the antigen and extracellular biomolecules. Another disadvantage of electrostatic adsorption of antigens is that it requires an opposite charges on the liposome and antigen [26]. This complicates the development of a general vaccine adjuvant that can be used for a diverse range of antigens.

Here we provide an attractive alternative method of antigen association to functionalized liposomes based on designer coiled coil (CC) motifs. In this method, complementary peptides interact resulting in the formation of noncovalent intermolecular helices [27,28]. The formation of these helices is based on a complementary peptide pair, peptide E (pepE) and peptide K (pepK), which form a parallel heterodimer CC [27–29]. The pepE-pepK peptide pair has been shown to remain intact under physiological conditions. Moreover, liposomes functionalized with pepK have shown to be able to target cells functionalized with pepE *in vitro* and *in vivo* [30–34]. We investigated the possibility to use this CC forming peptide pair as an attachment platform for association of antigens to liposomes. We hypothesized that the binding affinity of these CC-forming peptides results in a higher association efficiency of the antigen and a more stable association *in vivo* as compared to electrostatic interaction. Here we report that indeed the association of antigen (pepE-OVA323) with pepK-functionalized liposomes remained intact under physiological conditions. Moreover, we show this way of association significantly enhanced the immunogenicity of the antigen, resulting in a stronger CD4<sup>+</sup> T-cell activation.

## 2. Material and methods

### 2.1. Chemicals

Cholesterol (Chol), 1,2-distearoyl-*sn*-glycero-3-phosphocoline (DSPC) and 1,2-dioleoyl-3-trimethylammonium-propane (DOTAP) were purchased from Avanti Lipids (AL, USA). OVA323-339 and granulocyte-macrophage colony-stimulating factor (GM-CSF) were supplied by BioConnect (Huissen, Netherlands), 1,1'-Dioctadecyl-3,3',3'-Tetramethylindodicarbocyanine, 4-Chlorobenzenesulfonate (DiD), phorbol 12-myristate 13-acetate (PMA), ionomycin, Brefeldin A, Penicillin-Streptomycin (PenStrep), GlutaMAX<sup>™</sup> were supplied by Thermo Fisher (Bleiswijk, Netherlands). Iscove's Modified Dulbecco's Medium (IMDM) and Roswell Park Memorial Institute 1640 (RPMI) were supplied by Lonza (Basel, Switzerland). Fetal Calf Serum (FCS) was purchased from PAA Laboratories (Ontario, Canada). Dimethylformamide (DMF), piperidine, acetic anhydride, pyridine, trifluoroacetic acid (TFA) and acetonitrile (ACN) were purchased from Biosolve (Valkenswaard, Netherlands). *N,N*-diisopropylethylamine (DIPEA), and Ethyl

cyanohydroxyiminoacetate (Oxyma) were obtained from Carl Roth (Karlsruhe, Germany). Dichloromethane (DCM) and diethyl ether were supplied by Honeywell (Landsmeer, Netherlands). Tentagel HL-RAM was obtained from Rapp Polymere (Tübingen, Germany). All amino acids were supplied by NovaBioChem (Darmstadt, Germany). Fmoc-NH-PEG<sub>4</sub>-COOH was purchased from Iris Biotech GmbH (Marktredwitz, Germany). All fluorescent antibodies for flow cytometry were purchased from eBioscience (MA, USA) and are displayed in [Supplementary table 1](#). CD4 and CD8 T-cell enrichment kit was purchased from Miltenyi (Leiden, Netherlands).

All other chemicals were purchased from Sigma Aldrich (Zwijndrecht, Netherlands).

### 2.2. Mouse experiments

C57Bl/6, OT-I and OT-II transgenic mice on a C57Bl/6 background were purchased from Jackson Laboratory (CA, USA), bred in-house under standard laboratory conditions, and provided with food and water *ad libitum*. All animal work was performed in compliance with the Dutch government guidelines and the Directive 2010/63/EU of the European Parliament. Experiments were approved by the Ethics Committee for Animal Experiments of Leiden University.

### 2.3. Peptide synthesis

Peptides were synthesized on a microwave-assisted, automated peptide synthesizer (Liberty Blue). An overview of all the synthesized compounds can be found in [Supplementary table 2](#). Synthesis was performed at a 0.1 mmol scale on the solid-phase Tentagel HL-RAM resin with a loading of 0.39 mmol/g. Fmoc-deprotection was achieved with 20% piperidine in DMF at 90 °C for 60 s. Amide coupling was achieved using 5 equiv. of Fmoc-protected amino acid with 5 equiv. of DIC as activator and 5 equiv. of Oxyma as the activator base heated at 95 °C for 240 s. Upon completion of synthesis, peptides were acetylated with an excess of acetic anhydride and pyridine in DMF. Cholesterol-coupled pepK (CPK) was synthesized as described elsewhere [35]. In short, resin-bound peptides were PEGylated with 2.5 equiv. of Fmoc-NH-PEG<sub>4</sub>-COOH in the presence of 5 equiv. of DIPEA and 2.5 equiv. of HATU for 4 h at room temperature. Subsequently, the protecting Fmoc was removed with 20% piperidine in DMF and the reactive amine was coupled to 1.05 equiv. amino-cholestene hemisuccinate in the presence of 5 equiv. DIPEA and 2.5 equiv. HATU for 4 h at room temperature, before cleavage from the resin was performed by using a mixture of TFA:TIPS:water, 95:2.5:2.5. The peptide was precipitated in ice-cold diethyl ether; the precipitate was subsequently collected by centrifugation and dissolved in a water and ACN mixture. The ACN was removed using a rotary evaporator and water was removed by lyophilization, resulting in crude peptide as an off-white powder.

All peptides and conjugates were purified by reversed-phase HPLC (RP-HPLC) on a Kinetic Evo C18 column with a Shimadzu system comprising two LC-8A pumps and an SPD-10AVP UV-Vis detector. Peptides were purified using a gradient of 20–80% B, (where B is ACN containing 1% TFA, and A is water with 1% TFA) over 20 min with a flow rate of 12 mL/min. The collected fractions were analyzed on a LC-MS system (Thermo Scientific TSQ quantum access MAX mass detector connected to a Ultimate 3000 liquid chromatography system fitted with a 50 × 4.6 mm Phenomenex Gemini 3 μm C18 column) and those deemed to be pure were pooled. Organic solvent was removed under reduced pressure (150 mbar) before lyophilization to obtain a dry purified peptide powder.

AlexaFluor488-labeled pepE-OVA323 was synthesized starting from pepE-OVA323 with an additional glycine and cysteine at the C-terminus. The peptide was incubated with 1.1 equiv. AlexaFluor 488 C<sub>5</sub> maleimide (Thermo Fisher, Netherlands) in 100 mM HEPES buffer (pH 7.4) in the dark at room temperature. After 2 h, free dye was removed by dialysis under constant stirring in a 2 k MWCO Slide-A-Lyzer<sup>™</sup> (Thermo Fisher,

Netherlands) at 4 degrees to Milli-Q (18,2 MOhm/cm) overnight. Finally, the pure peptide was obtained by centrifugation at 1000g for 30 min.

#### 2.4. Liposome preparation and characterization

Liposomes were prepared by the gold standard dehydration-rehydration method as described elsewhere [17]. In brief: 15 μmol total lipids with or without CPK were dissolved in 1:2 methanol:chloroform in the desired ratio (2:1:1 DSPC:DOTAP:cholesterol molar ratio with 1 mol% CPK). In case of fluorescent liposomes, 0.1 mol% of total lipid DiD was added in this step. Subsequently, the organic solvent was evaporated at 150 mbar and 50 °C in a rotary evaporator, yielding a lipid film. This film was hydrated in the presence of glass beads with a 10 mM HEPES, 280 mM sucrose, pH 7.4 (H/S buffer), with antigen in the case of encapsulation, frozen in liquid nitrogen and lyophilized overnight. The resulting lipid cake was rehydrated with Milli-Q to a final volume of 1 mL (resulting in a 15 mM lipid suspension) and homogenized with a LIPEX extruder (Evonik, Canada) by repeated passage over stacked filters of 400 nm and 200 nm (Nuclepore Track-Etch membrane from Whatman, Netherlands).

Hydrodynamic diameter and polydispersity were measured by dynamic light scattering (DLS) using a Zetasizer Nano ZS (Malvern Instruments Ltd., Worcestershire, UK). The zeta potential was measured using laser Doppler electrophoresis on the same machine with a zeta dip cell (Malvern Instruments Ltd.). Each sample was diluted 100-fold in 10 mM HEPES buffer (pH 7.4) before measurement.

#### 2.5. Isothermal titration calorimetry

Isothermal titration calorimetry (ITC) was performed with a Micro-Cal PEAQ-ITC Automated machine (Malvern Instruments Ltd., Worcestershire, UK). Free pepK or liposomes containing CPK (300 μl, 50 μM pepK or CPK in H/S buffer) were used in the receptor compartment. 3 μl of pepE, pepE-OVA323 or pepE-OVA257 (500 μM) was added every 180 s.

For data analysis, the first injection was removed from the raw data. Subsequently a One-sided fitting was performed, which was optimized by 100 iterations until the values for binding constant, stoichiometry and dH did not change anymore.

#### 2.6. Far-UV circular dichroism spectroscopy

Far-UV circular dichroism (CD) measurements were performed on a Jasco J815 CD spectrometer equipped with a Jasco PTC 123 Peltier temperature controller in a 1 mm quartz cuvette. Far-UV CD spectra between 190 and 260 nm were also collected at T = 25 °C. The molar ellipticity [θ] was calculated from the measured ellipticity θ, the path length l in centimeter, the molar monomer concentration cM, and the number of amino acids per peptide N as  $[\theta] = \frac{100 \cdot \theta}{l \cdot (cM1 \cdot N1) + (cM2 \cdot N2)}$  (Equation 1) as described elsewhere [36].

#### 2.7. Association efficiency

To determine the association efficiency of pepE-OVA323 to liposomes, liposomes with and without CPK (800 μg lipids/mL) were incubated with antigen (30 μg/mL) for 30 min. Subsequently, unbound antigen was separated from liposomes with a centrifuge membrane concentrator (Vivaspin2, 300.000 MWCO, Sartorius) by spinning down for 10 min at 500g. The unbound fraction (flow through) was measured by RP-UPLC (Waters ACQUITY UPLC, Waters, MA, USA).

Liposomes with and without CPK (800 μg lipids/mL) were mixed with pepE-OVA257 (30 μg/ml) and dialyzed in a Float-A-Lyzer® G2 (Spectrum labs, CA, United States) of 1 mL and MWCO of 100 kD for 5 days against 10 mM HEPES buffer pH 7.4. Remaining antigen in the

dialysis membrane was measured by RP-UPLC (Waters ACQUITY UPLC, Waters, MA, USA).

#### 2.8. Ex vivo T-cell stimulation

Bone marrow-derived dendritic cells (BMDCs) were prepared as described elsewhere [37]. In short, bone marrow was isolated from murine tibia and femurs of C57BL/6 mice. Bone marrow cells were stimulated for 10 days with 20 ng/mL GM-CSF in complete IMDM (cIMDM, IMDM supplemented with 100 U/mL PenStrep, 2 mM glutamax and 8% FCS). After 10 days, the BMDCs (50,000 cells per well) were exposed to the different formulations for 4 h. Subsequently the BMDCs were washed twice with cIMDM and incubated with either 100,000 CD4<sup>+</sup> T-cells derived from OT-II mice or 100,000 CD8<sup>+</sup> T-cells derived from OT-I mice, which were purified according to manufacturer's protocol [38], for 72 h in complete RPMI (cRPMI, supplemented with 100 U/mL PenStrep, 2 mM glutamax, 50 μM β-mercaptoethanol and 10% FCS). After 72 h, the cell suspension was harvested and prepared for flow cytometry.

#### 2.9. Flow cytometry measurements and analysis

For flow cytometry measurements, the cell suspension was washed with FACS buffer (PBS with 1% FCS and 2 mM EDTA). Subsequently, the suspension was stained for 30 min in the dark at 4 °C with FACS buffer containing fluorescent antibodies against the surface markers of interest (an overview of the used antibodies is found in Supplementary table 1). After the staining, the cells were washed with PBS and measured in the flow cytometer (CytoFLEX S, BeckmanCoulter, CA, US). For intracellular markers, the cells were subsequently fixed and stained according to manufacturer's protocol with the Transcription Factor Staining Buffer Set (eBioscience, catalogue number 00-5523-00) for transcription factors, or the Intracellular Fixation & Permeabilization Buffer Set (eBioscience, catalogue number 88-8824-00). Before measurement, the cells were resuspended in PBS. All flow cytometry data were analyzed using FlowJo vX.

#### 2.10. Peptide uptake in BMDCs

BMDCs were prepared as described above and 50,000 BMDCs per well were plated in a flat bottomed 96 wells plate. Subsequently, different liposome formulations containing DiD (ex/em wavelength: 644/665 nm) were added to the culture medium of these BMDCs. After 4 h, cell medium was refreshed twice to remove any unbound formulation. Subsequently, cells were prepared for flow cytometry.

#### 2.11. Biodistribution studies in zebrafish embryos

Zebrafish (Danio rerio, strain AB/TL or Tg(kdrl:RFP-CAAX)s916 [39]) were maintained and handled according to the guidelines from the Zebrafish Model Organism Database (<http://zfin.org>) and in compliance with the directives of the local animal welfare committee of Leiden University. Fertilization was performed by natural spawning at the beginning of the light period, and eggs were raised at 28.5 °C in egg water (60 g/mL Instant Ocean sea salts). Prior to injection, zebrafish embryos were embedded and anesthetized in 0.4% agarose containing 0.01% tricaine. Liposomal formulations (containing 5 mM lipids and/or 200 μg/ml pepE-OVA323-AF488) were injected with 1 nL volume in the duct of Cuvier, at 2.5 days post fertilization (dpf) as described previously [40]. For each treatment, two independently formulated liposome preparations were imaged using confocal microscopy. Embryos were randomly picked from a dish of 10–20 successfully injected embryos (exclusion criteria were: no backward translocation of erythrocytes after injection and/or damage to the yolk ball).

Confocal z-stacks were captured on a Leica TCS SPE confocal microscope, using a 10 × air objective (HCX PL FLUOTAR) or a 40 × water-

immersion objective (HCX APO L). For whole-embryo views, 3–5 overlapping z-stacks were captured to cover the complete embryo. Laser intensity, gain, and offset settings were identical between stacks and sessions. Images were processed using the Fiji distribution of ImageJ [41,42]. The greyscale threshold for both liposomal and peptide signal was determined and set identical for an entire experiment. All pixels with intensity above the threshold were set at a maximum value of 255, whereas negative pixels were set at 0. Subsequently, the “3D-multi coloc” plugin of 3d Image suite was used to determine co-localization [43].

### 2.12. Adoptive transfer and vaccination study

C57Bl/6 mice (10–15 week-old) were randomized into groups. On day 0, all mice received 500,000 CD4<sup>+</sup> T-cells, that were purified from sex-matched OT-II transgenic mice with a CD4 + T-cell enrichment kit (Miltenyi, Netherlands) according to manufacturer's protocol [38], via the tail vein. On day 1, mice were immunized subcutaneously with a single injection of formulation in a total volume of 200 µl in H/S buffer. Seven days after immunization, mice were sacrificed and blood, spleen and axillary and brachial lymph nodes (LNs) were removed. Organs were processed and measured by flow cytometry.

Splenocytes (10<sup>6</sup> per well) of each mouse were stimulated in cRPMI for 1 h with either medium, pepE-OVA323, or PMA and ionomycin, after which brefeldin A was added and the cells were incubated for another 5 h and subsequently prepared for flow cytometry.

### 2.13. Statistical analysis

Data was processed and analyzed in GraphPad v8 (Prism) for Windows. Statistical analysis was performed with the same program and the method of analysis is indicated in the figure legends.

## 3. Results

### 3.1. Peptide and liposome characterization

The MHC-II (I-A<sup>b</sup>) restricted epitope of ovalbumin (OVA323-339, ISQAVHAAHAEINEAGR) was synthesized at the C-terminus of pepE resulting in the pepE-OVA323 conjugate. The same was done for the MHC-I (H2-K<sup>b</sup>) restricted epitope (OVA257-264, SIINFELK), resulting in pepE-OVA257. All peptides were of >95% purity according to RP-HPLC (Supplementary figures 1–4). Subsequently, positively charged (DOTAP-containing) liposomes were prepared with and without CPK to assess the effect of its incorporation on the physico-chemical properties of liposomes composed of DOTAP, DSPC and cholesterol. The size (179 vs 177 nm for liposomes without and with CPK respectively), polydispersity index (PDI, 0.073 vs 0.071 for liposomes without and with CPK respectively) and zeta potential (between 48 vs 45 mV for liposomes without and with CPK respectively) of liposomes were unaffected by the addition of 1 mol% CPK (Table 1).

To confirm the presence of CPK in the functionalized liposomes, and the ability of CPK to interact with pepE-conjugates, we performed far-UV CD spectroscopy and ITC, respectively. Far-UV CD spectra showed

minima at 222 and 208 nm in CPK-functionalized liposomes which indicates presence of  $\alpha$ -helices. Upon mixing with pepE, the peak at 208 nm and overall signal increased, pointing to interaction and the formation of a CC structure (Supplementary figure 5). ITC was used to determine the binding energy and dissociation constant (Kd) between peptide and liposome. No binding energy was measured in non-functionalized liposomes (Fig. 1), whereas the binding energy of pepE and pepE-conjugates titrated into a suspension of CPK-functionalized liposomes decreased slowly during the first 6 injections after which a plateau was reached (Fig. 1). Kd values were determined for both pepE-conjugates onto CPK-functionalized liposomes and all were in the order of 10<sup>-7</sup> M, but could not be measured with non-functionalized liposomes. Moreover, antigen was mixed with both formulations and unbound antigen was removed by centrifugal filtration or dialysis to measure association efficiency. We observed an association efficiency of 78% after mixing pepE-OVA323 with non-functionalized liposomes, which substantially increased after mixing with functionalized liposomes (Table 1), where the unbound antigen concentration was below the limit of detection. Thus, CC interaction provided a simple and highly efficient method to associate antigen to liposomes, which appeared stronger than adsorption to non-functionalized liposomes.

### 3.2. Peptide uptake in BMDCs

We assessed whether the increased affinity provided by CC-mediated association would result in an increased antigen and liposome uptake by BMDCs. Fluorescently labeled pepE-OVA323 and the DiD-labeled fluorescent liposomes were used to facilitate cell uptake studies using flow cytometry. The incorporation of a small amount of fluorescent dye resulted in a slightly, but not substantially smaller size (160 vs 180 nm) of the liposomes (Supplementary Figure 6). BMDCs were incubated for 4 h with fluorescent formulations containing pepE-OVA323. Over 90% of all BMDCs had taken up liposomes, irrespective of CPK-functionalization or presence of antigen (Fig. 2). Uptake of peptides in the absence of liposomes was limited (39%). Association with non-functionalized liposomes increased the uptake from 40% to 75%, but was this increased to >99% after association to CPK-functionalized liposomes (Fig. 2A). The fluorescent signal in cells (Mean Fluorescence Intensity, MFI) after exposure to pepE-OVA323 was significantly increased with functionalized liposomes (20-fold), compared to a 4-fold increase by association to non-functionalized liposomes (Fig. 2B), which indicates more uptake of pepE-OVA323 per cell. These data show that under physiological conditions (including serum), the strong association of antigen to CPK functionalized liposomes via coiled coil formation results in an improved uptake of the antigen.

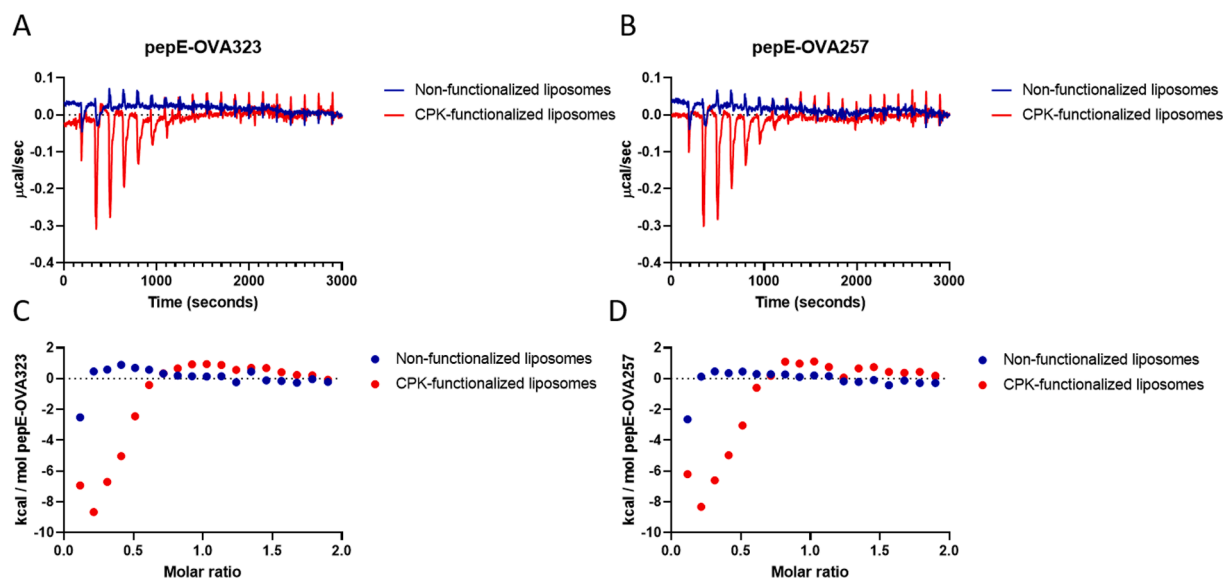
### 3.3. In vivo distribution in zebrafish embryos

The *in vitro* studies revealed that CPK-functionalized liposomes increased antigen and liposome co-localization compared to non-functionalized liposomes. To evaluate how these *in vitro* findings translate to *in vivo* administration in complex tissue, we investigated the biodistribution of pepE-OVA323 and cationic liposomes after intravenous injection in zebrafish embryos. Free peptide, free liposomes, non-

**Table 1**

Overview of the particle size (z-average), polydispersity index (PDI) and zeta potential of non-functionalized liposomes (DOTAP: DSPC: cholesterol) and CPK-functionalized liposomes before and after mixing with pepE-OVA323 or pepE-OVA257. Moreover, the dissociation constant (Kd) as measured by ITC is shown. \* = significantly different from the liposomes without antigen. # = beyond detection limit. Values represent average values  $\pm$  SD (n  $\geq$  2). N.a. = not applicable.

Liposome formulation	Z-average (nm)	PDI	Zeta potential (mV)	Kd (nM)	Association efficiency (%)
Non-functionalized liposomes	179.3 $\pm$ 13.8	0.073 $\pm$ 0.039	48.1 $\pm$ 3.3	n.a.	n.a.
CPK-functionalized liposomes	176.7 $\pm$ 14.4	0.071 $\pm$ 0.047	44.9 $\pm$ 6.2	n.a.	n.a.
Non-functionalized liposomes + pepE-OVA323	188.6 $\pm$ 6.6	0.116 $\pm$ 0.028 *	35.0 $\pm$ 5.2 *	> 10 <sup>5</sup> #	78.1 $\pm$ 4.0
CPK-functionalized liposomes + pepE-OVA323	185.8 $\pm$ 4.2	0.080 $\pm$ 0.014	28.6 $\pm$ 7.0 *	166 $\pm$ 68	>95% #
Non-functionalized liposomes + pepE-OVA257	182.5 $\pm$ 2.3	0.092 $\pm$ 0.004	45.0 $\pm$ 2.4	> 10 <sup>5</sup> #	1.69 $\pm$ 2.4
CPK-functionalized liposomes + pepE-OVA257	184.6 $\pm$ 2.5	0.089 $\pm$ 0.004	38.8 $\pm$ 1.3 *	392 $\pm$ 264	47.5 $\pm$ 7.1



**Fig. 1.** Antigen binding to liposomes. Buffer corrected heat plots of isothermal titration calorimetry measurements in which non-functionalized and functionalized liposomes were titrated with pepE-OVA323 (A) and pepE-OVA257 (B). The calculated energy per mol of pepE-OVA323 (C) or pepE-OVA257 (D) was used to derive the dissociation constant  $K_d$  (B).

functionalized and CPK-functionalized liposomes incubated with pepE-OVA323 were injected in zebrafish embryos. Confocal fluorescence microscopy was used to assess the biodistribution of both liposomes and peptides after injection. Co-localization throughout the embryo was observed after injection of functionalized liposomes with pepE-OVA323, and was especially prominent in the caudal vein (Fig. 3 A, B). In contrast, for cationic liposomes with pepE-OVA323, co-localization was only observed in the caudal aspect of the tail and at the injection site for non-functionalized liposomes with pepE-OVA323 (Fig. 3A). Non-functionalized liposomes that were associated with pepE-OVA323 displayed similar distribution to the individual components alone (Supplementary figure 7). This difference of pepE-OVA323 distribution after association with functionalized liposomes and non-functionalized liposomes was more profound when zoomed in on the tail vein (Fig. 3B). We observed 75% of the functionalized liposome signal co-localized with pepE-OVA323, in contrast to pepE-OVA323 associated to non-functionalized liposomes (34%) (Fig. 3C). In 3D representation, the co-localization can be observed at the subcellular level as well (Supplementary video 1 and 2). Thus, pepE-OVA323 remained strongly associated with functionalized liposomes *in vivo*, but only in part associated with non-functionalized liposomes.

### 3.4. Ex vivo T-cell stimulation and proliferation

Next, we investigated whether the strong association and co-localization of antigen and liposome results in improved immunogenicity. We pulsed BMDCs with plain pepE-OVA323 or pepE-OVA323 adsorbed to liposomes with and without CPK for 4 h, after which  $CD4^+$  T-cells derived from OT-II transgenic mice were co-cultured with these BMDCs. We observed that BMDCs pulsed with soluble pepE-OVA323 successfully induced concentration-dependent T-cell proliferation (Fig. 4A). Compared to plain peptide, BMDCs pulsed with pepE-OVA323 associated with non-functionalized liposomes resulted in an approximately 5-fold lower EC50. The same antigen associated to CPK-functionalized liposomes, however, showed an 18-fold lower EC50 than soluble peptide.

A similar experiment was performed with pepE-OVA257, an MHC-I restricted epitope of ovalbumin and  $CD8^+$  T-cells derived from OT-I transgenic mice. There we also found a concentration-dependent T-cell proliferation profile. Interestingly, in contrast to the  $CD4^+$  T-cell

proliferation, there were no differences in proliferation induced by plain peptide, or antigen associated with either liposome formulation (Fig. 4B).

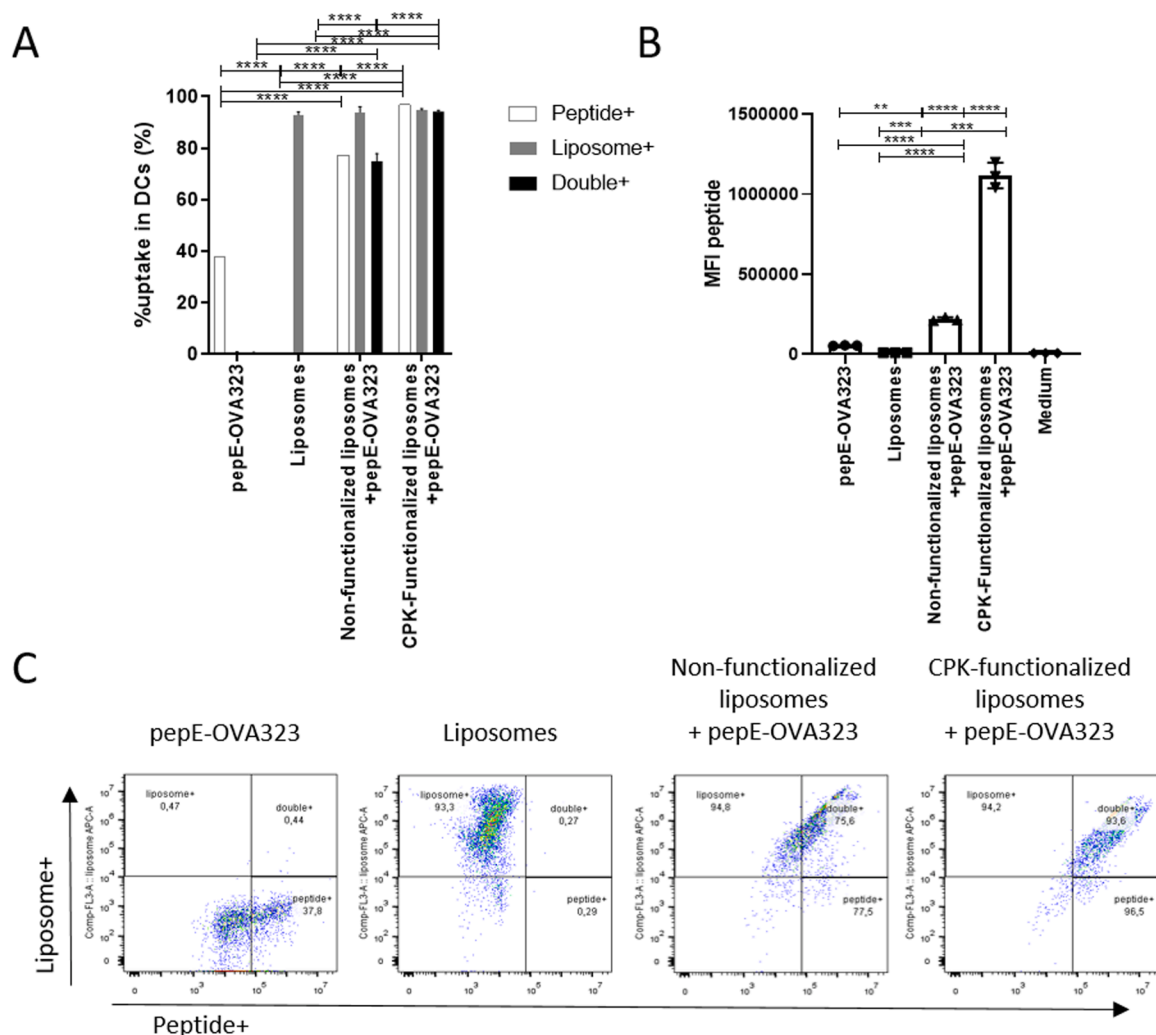
### 3.5. In vivo immune response in mice

As both non-functionalized and CPK-functionalized liposomes increased the  $CD4^+$  T-cell responses *in vitro*, we vaccinated mice that had received an adoptive transfer of ovalbumin-specific  $CD4^+$  T-cells derived from OT-II mice to compare the effect of non-functionalized and CPK-functionalized liposomes on the *in vivo* immunogenicity. One day after the adoptive transfer, mice received pepE-OVA323 in buffer or associated to either non-functionalized or CPK-functionalized liposomes. The expansion of the transferred  $CD4^+$  T-cells was successfully induced by both liposome formulations (Fig. 5A & B), whereas few OT-II cells were present in mice that were vaccinated with peptide alone. Two different doses were used for vaccination. There were no differences in T-cell expansion in the high dose, but in the low dose treatment we observed increased expansion in the group which received the antigen with functionalized liposomes (Fig. 5B).

The majority of the OT-II cells that were found in all organs were positive for the transcription factor T-bet. Few OT-II cells expressed Gata3, ROR $\gamma$ t or FoxP3 (Supplementary figure 8), suggesting a Th1 profile. Indeed, upon *ex vivo* stimulation with PMA and ionomycin, IFN- $\gamma$  was the most abundantly expressed cytokine (Fig. 6A) while almost no IL-4 or IL-17 was expressed (Supplementary figure 9). We observed a significant increase in IFN- $\gamma$  production after immunization with CPK-functionalized liposomes compared to non-functionalized liposomes. Interestingly, the expanded cells also produced IL-10 (Fig. 6B) despite the absence of FoxP3 expression. The IL-10 production was significantly higher in mice that received functionalized liposomes. Further inspection of these cytokine-producing T-cells revealed an increase of double-producing OT-II cells after vaccination with functionalized liposomes (Fig. 6C, D).

## 4. Discussion

In this study, we demonstrated a novel antigen association method to liposomes based on a complementary peptide pair that forms a CC upon interaction. PepK was coupled to cholesterol to yield CPK, which was



**Fig. 2.** Uptake of pepE-OVA323 and liposomes in MHCII<sup>+</sup>CD11c<sup>+</sup> BMDCs. BMDCs were incubated in cIMDM for 4 h with different formulations containing fluorescent liposomes and fluorescent pepE-OVA323. Subsequently, the BMDCs were analyzed by flow cytometry for peptide and liposome uptake. C shows representative FACS plots of BMDC population for each formulation. A Two-way ANOVA with a Tukey's multiple comparison test was performed to determine statistically significant differences in antigen and liposome uptake. MFI was compared with a One-way ANOVA and Tukey's multiple comparison test. \*\* =  $p < 0.01$ , \*\*\* =  $p < 0.001$ , \*\*\*\* =  $p < 0.0001$ .

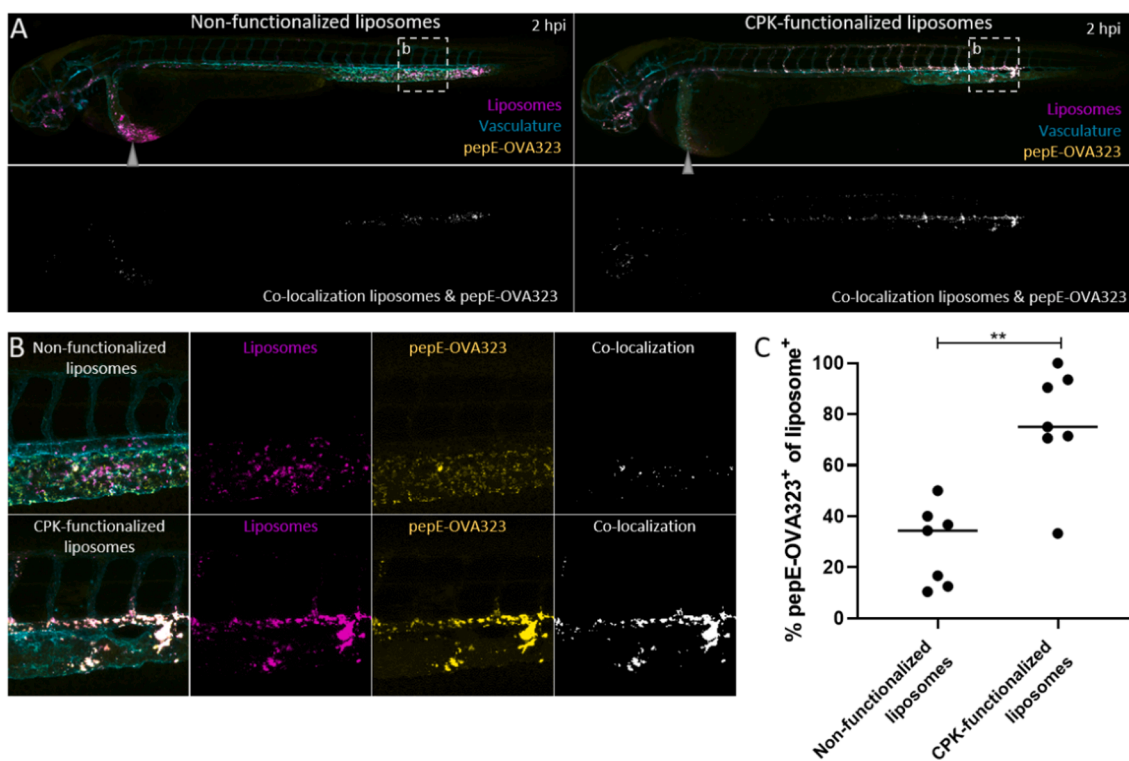
encapsulated in the lipid bilayer, whereas pepE was synthesized with two different antigenic epitopes. The interaction between pepE and pepK was already known to be stable under physiological conditions [33,44] and we now demonstrate its use as a method of antigen association. We established that incorporation of CPK in the liposomal bilayers did not affect the size, polydispersity or zeta potential of liposomes composed of DSPC, DOTAP and cholesterol.

The binding affinity between pepK and pepE is in line with expected values reported before [45]. We observed Kds around  $10^{-7}$  M when mixing both soluble peptides. This did not change when CPK-functionalized liposomes were used instead of soluble peptide or pepE-antigen was used instead of pepE. Moreover, we observed an increase in ellipticity in the CD spectrum, which strongly suggests that the association indeed occurs via the formation of a CC [36,46]. By simple mixing of the antigen and liposomes, high efficiency adsorption was achieved. This adsorption is very stable and resulted in high co-localization of antigen and liposome both *in vitro* and *in vivo*.

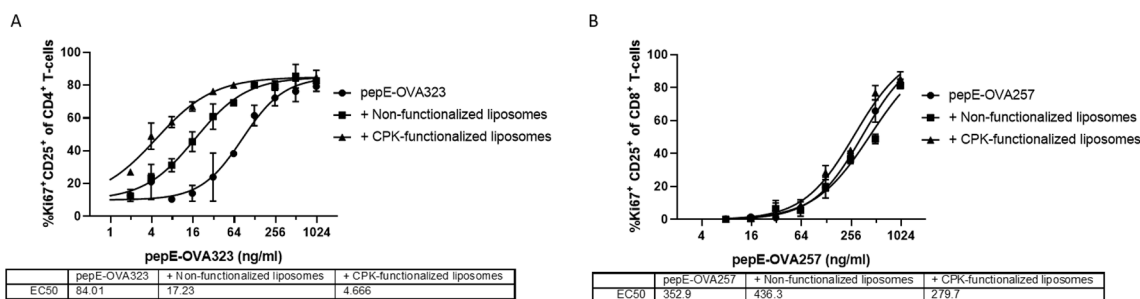
We showed in both BMDCs and zebrafish embryos that peptides associated to non-functionalized liposomes, but not to functionalized liposomes, can rapidly dissociate from the liposomes. While we saw a degree of co-localization of antigen with non-functionalized liposomes, this was significantly increased when antigen was associated with

functionalized liposomes. PepE-OVA323 was taken up by BMDCs and showed accumulation in zebrafish endothelial cells even without liposomes. This is consistent with the net anionic charge of this peptide, which leads to rapid clearance by scavenger endothelial cells which are similar to liver sinusoidal endothelial cells in mammals [40]. CC association, as demonstrated, results in prolonged co-localization and therefore potentially increases exposure to antigen-coated liposomes and could result in more antigen presentation and TCR stimulation.

This prolonged TCR stimulation is in line with our *in vitro* observations, where we confirmed the adjuvant effect of cationic liposomes [1,5]. Antigen association with non-functionalized liposomes resulted in a stronger T-cell proliferation, reducing the EC50 approximately 5-fold. Stronger antigen association to CPK-functionalized liposomes resulted in an 18-fold EC50 reduction compared to plain antigen in CD4<sup>+</sup> T-cells. Although the presence of the coiled-coil construct have increased the immunogenicity independent of the strong increase in antigen-liposome association; the absence of any immune potentiating effect for CD8<sup>+</sup> T-cells argues against that. The proliferation of CD8<sup>+</sup> T-cells was barely affected by the addition of either non-functionalized or functionalized liposomes to the formulation. This suggests that the T-cell proliferation is only induced by unbound antigen, but not liposome-associated antigen. Possibly the antigens associated to these liposomes are less capable



**Fig. 3.** Liposome and antigen distribution in zebrafish embryos. Representative images of zebrafish embryos 2 h post injection (hpi) of pepE-OVA323 adsorbed to non-functionalized liposomes and CPK-functionalized liposomes. Gray arrows indicate the site of injection. Images were taken by confocal microscopy at 10× magnification (A) and 40× magnification (B), compressed and processed by ImageJ. PepE-OVA323 (yellow), liposomes (magenta) and the vascular system (cyan; kdrl:mCherry) of the zebrafish are shown in the upper image; co-localization of pepE-OVA323 and liposomes (i.e., the pixels in which the signals of fluorescence were above the background signal) is white in the lower image of whole fish (A). The percentage of all pixels that had liposome signal which also had pepE-OVA323 signal (C) was compared with a Mann Whitney test, \*\* =  $p < 0.01$ .



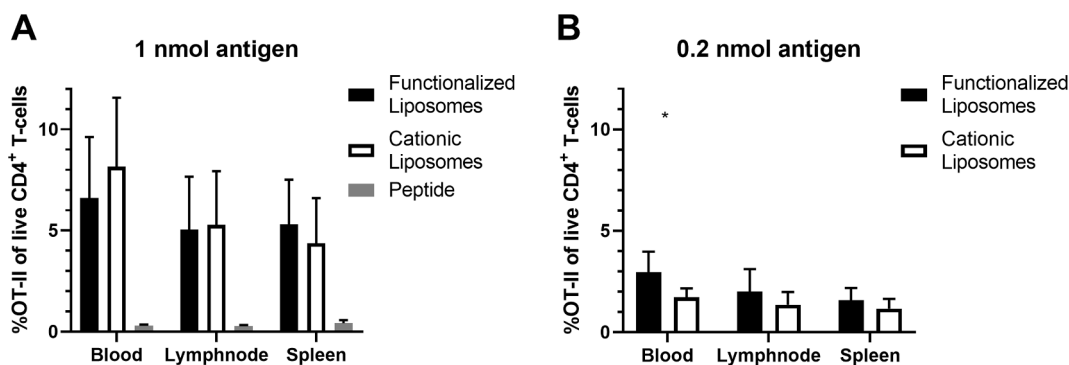
**Fig. 4.** *In vitro* T-cell proliferation. A) CD8<sup>+</sup> T-cell proliferation induced by BMDCs pulsed with pepE-OVA257 in different formulations. B) CD4<sup>+</sup> T-cell proliferation induced BMDCs pulsed with pepE-OVA323 in different formulations. EC50 values were calculated based on a non-linear dose-response model fit with variable slope (four parameters) in which top and bottom were shared between all groups. Graph shows average of 3 points for each concentration and is a representative of 3 separate experiments.

of endosomal escape and therefore do not enter the cytosol after uptake by BMDCs. MHC-II restricted epitopes are loaded into MHC-II molecules in the late endosome, and therefore not affected by the absence of endosomal escape [47–50]. MHC-I restricted epitopes however require this escape to be efficiently cross-presented to MHC-I molecules in the ER [47,50–52]. The potential lack of endosomal escape would be surprising, as cationic liposomes are often used to induce CD8<sup>+</sup> T-cell responses *in vitro* and *in vivo* [17–19], and are even found to promote cross presentation [22,53,54]. Apparently, this is not the case for the liposomes we have investigated.

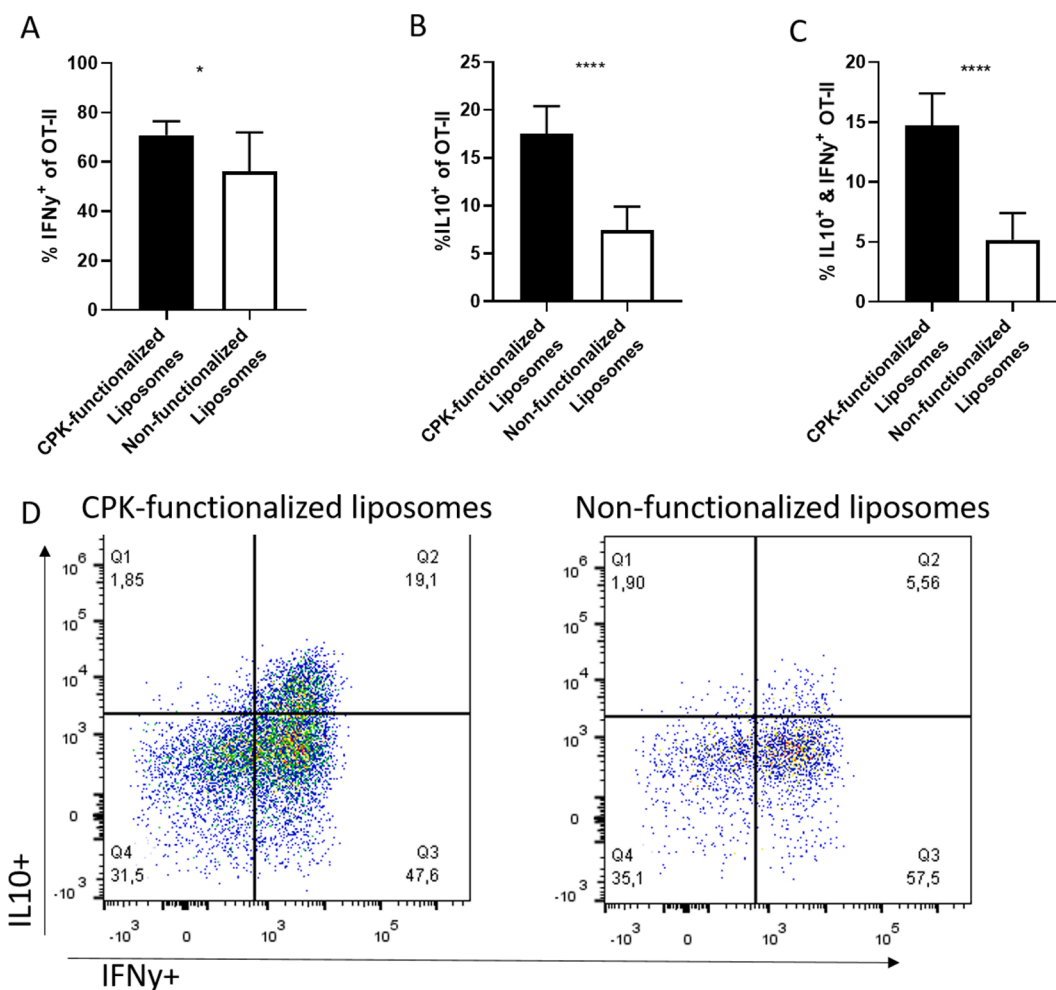
Strong T-cell expansion of transferred OT-II derived CD4<sup>+</sup> T-cells was observed 1 week after vaccination with antigen associated to both non-functionalized and functionalized liposomes. When the vaccination was performed with 1 nmol of antigen, we found that approximately 5–6% of

all CD4<sup>+</sup> T-cells were expanded OT-II cells, which is in line with previous work [10]. Both liposomes directed the induced immune response towards a Th1 phenotype, as most of our OT-II cells expressed T-bet, which is in agreement with previous findings [9,10,21,26]. Hardly any cells were expressing transcription factors associated with Th2 (Gata3), Th17 (RORγT) or Treg (FoxP3) phenotypes. This was confirmed by the cytokine production upon *ex vivo* stimulation of spleen-derived lymphocytes. Approximately 55% of all OT-II cells in the spleen were producing IFN-γ, a typical Th1 cytokine, in mice which received antigen associated with non-functionalized liposomes. In mice that received functionalized liposomes significantly more OT-II cells produced IFN-γ, suggesting a stronger immune response.

A striking difference was the production of IL-10 after immunization. In mice that received non-functionalized liposomes, approximately 7%



**Fig. 5.** OT-II T-cell expansion after vaccination. OT-II CD4<sup>+</sup> T-cell expansion *in vivo* induced by vaccination with 1 nmol antigen (A, mean ± SD, n = 8 for liposomes, n = 4 for peptide) and 0.2 nmol antigen (B, mean ± SD, n = 7) as measured by flow cytometry. Effect of the formulation in each organ was compared by multiple student's t tests. \* = p < 0.05.



**Fig. 6.** Cytokine production by expanded OT-II T-cells after *ex vivo* stimulation. OT-II cells that produced IFN- $\gamma$  (A), IL-10 (B), and both IL-10 and IFN- $\gamma$  (C) after stimulation for 6 h with PMA and ionomycin from the spleen after vaccination with 1 nmol antigen associated with either CPK-functionalized or non-functionalized liposomes (mean ± SD, n ≥ 6). A representative FACS plot of OT-II cells for each formulation is shown in D. Mice where less than 200 OT-II cells were detected, were excluded from the analysis. Outliers were detected with Grubb's outliers test and groups were compared with an unpaired student's t test. \* = p < 0.05, \*\*\*\* = p < 0.0001.

of all OT-II cells produced IL-10, but in mice that were vaccinated with CPK-functionalized liposomes, over 15% produced IL-10. Practically all of these IL-10-producing cells were also producing IFN- $\gamma$ . The double-producing CD4<sup>+</sup> T-cells were previously observed in parasitic infections and have proven critical for host-survival. They are considered to dampen the ongoing immune response more effectively than ordinary

regulatory T-cells [55–58]. They could be an interesting phenotype to induce in the treatment of auto-immune diseases and allergy, as both IFN- $\gamma$  and IL-10 have an inhibitory effect on the Th2 immune response [56,59,60]. It is thought that this subset of IL-10- and IFN- $\gamma$ -producing Th1 cells is the result of continuous antigen presentation and T-cell receptor (TCR) overstimulation [55,58,61–64]. This suggests that

immunization with antigen associated to CPK-functionalized liposomes resulted in more stimulation of the TCRs by antigen presenting cells than antigen associated to non-functionalized liposomes, which could be explained by the enhanced affinity of pepE-OVA323 to functionalized liposomes.

We are not the first to show surface coupling of antigen results in a stronger immune response [65–70]. Most approaches consist of proteins or peptides which are covalently attached to reactive groups on the vesicle surface [66,67,69] or covalent attachment of the antigen to a lipid tail [68,70]. The CC based association presented in this paper is non covalent, but strong enough to remain intact in circulation. Moreover, simple mixing liposomes and antigen is sufficient to yield high association efficiency and formulations can be administered within a few minutes after mixing. The association mechanism is based on biodegradable peptides, which are readily synthesized and could serve as a platform technique for other antigenic epitopes.

In conclusion, we have demonstrated the use of CC-forming peptides as an antigen-attachment tool for peptide-based antigens. The antigens have a high affinity for the liposomes and have a high association efficiency. Moreover, the association remains intact *in vitro* and *in vivo* and could be used for peptide-based MHC-II restricted epitopes. The induced immune response by DCs pulsed with CC-adsorbed antigen to liposomes was much higher than for electrostatically adsorbed antigen. Moreover, *in vivo* we observed a strong increase in IL-10- and IFN- $\gamma$ -producing antigen-specific CD4<sup>+</sup> T-cells, which suggests a stronger TCR stimulation after CC-mediated association of antigen.

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## Declaration of Competing Interest

None.

## Appendix A. Supplementary material

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.ejpb.2020.11.005>.

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