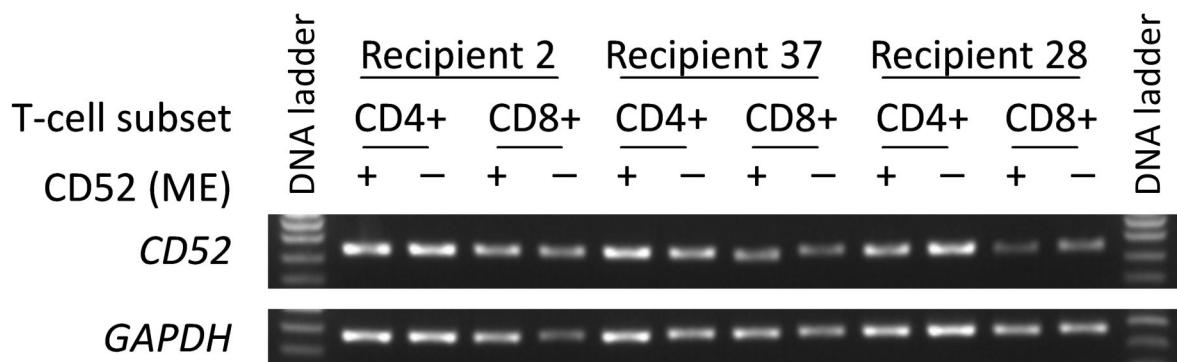
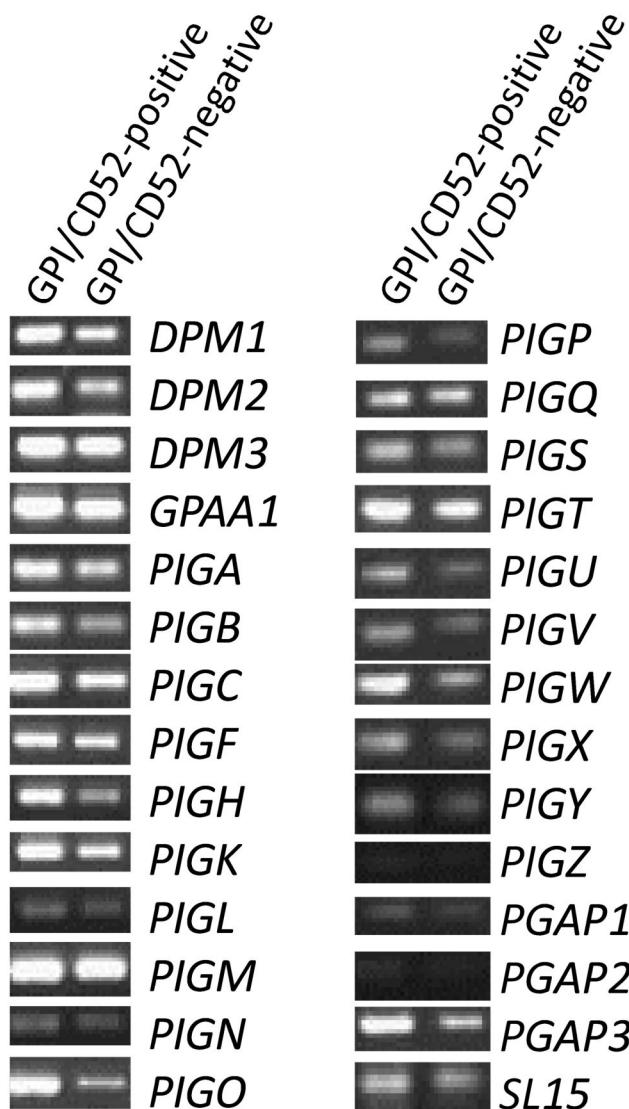


Supplemental Figure S1.

A



B



1 **Supplemental Figure Legends**

2 **Supplemental Fig S1. No loss of mRNA expression of CD52 or any of the GPI-anchor synthesis
3 pathway genes in GPI/CD52-negative T cells**

4 **(A)** Agarose gel electrophoresis results from a representative CD52 mRNA expression analysis of CD4
5 and CD8 T cells derived from recipients 2, 37, and 28 which were positive (+) or negative (-) for CD52
6 membrane expression (ME). CD52-negative and CD52-positive CD4 and CD8 T cells were purified by
7 flow cytometric cell sorting, followed by mRNA isolation, cDNA synthesis, and a PCR amplification
8 using specific primers (Supplemental Table 2). Equimolar amounts of cDNA of the CD52-negative and
9 CD52-positive samples were used for PCR amplification. As a control *GAPDH* was amplified. **(B)**
10 Representative example of mRNA expression analysis for the GPI-anchor synthesis pathway genes
11 performed on FACS purified GPI/CD52-negative and GPI/CD52-positive CD4 T cells from recipient 37.
12 GPI/CD52-negative and GPI/CD52-positive CD4 T cells were purified by FACS, followed by mRNA
13 isolation, and cDNA synthesis. PCR amplification was performed using primer sets specifically
14 designed to individually amplify all 28 proteins involved in the GPI-anchor biosynthesis pathway.
15 Equimolar amounts of cDNA were used for each amplification. Depicted is the agarose gel
16 electrophoresis analysis from the resulting amplicons.

Supplemental Table II. Primer sequences used for mRNA expression analysis and mutation analysis

Set 1	GCCATGGAACTCACCGGTAATA	AAAACGTTGGCCCTTCAG
Set 2a	CTCACCGGTAAATAGAGGACAC	GCTTGTGTTGTAAGCACCGAGC
Set 2b	GTTGGGAGAGAGTCACGATA	TTCAGCACCAAAGCTCTC
<i>CD52</i>	AGACAGCCCTGAGATCACCTA	GCCCCTACATCATTACCCCC
<i>GAPDH</i>	ATGTTCGTCATGGGTGTGAACCA	TGGCAGGTTTCTAGACGGCAG
<i>PIGA</i>	GTTGGGAGAGAGTCACGATA	GCTTGTGTTGTAAGCACCGAGC
<i>PIGB</i>	CTTACCCCTCTTATTCATGGCTG	GGTTAATGAGTATCCACAGAACAC
<i>PIGC</i>	CTGCGCAACCCTAGGAACCT	GAAGAAGCCAGACCAAGTCCC
<i>PIGF</i>	GTAGTTCCCGCTCCCTTC	CTCCAAGCCATGCTCTACA
<i>PIGH</i>	GCCATTACATGCAGAAGGT	GAAGTGTCACCTGATGGT
<i>PIGK</i>	GGGAAGTCTGAAGCGGTAA	CTAGGTGGATCCTCCAGT
<i>PIGL</i>	TACCTAAAGGGTGTCTGTG	CCGGGAGAAGATAATGTAGAGG
<i>PIGM</i>	TCACCGCTTCCTTATACC	TGGGAAGGATGTAAGTCACTG
<i>PIGN</i>	AGAAGTGAAGAACCAAGCC	TCAACACTGATACAACAAGGTC
<i>PIGO</i>	TCGTTGCCCTGAAGAGACAC	ATGCCAATGGATGGCTGGAA
<i>PIGP</i>	GGTGGAAAATTCAACGTCGC	TGGATGGAGTCGAGTGGAGA
<i>PIQQ</i>	CTGTGGATCAGCTACATCCA	CCAGGTATAGGAACAGGAG
<i>PIGS</i>	GCGGCTACACACCTAGAGG	CTGGGAGTAAGGCAACGAGG
<i>PIGT</i>	GCGGGAGGAATTGTCATCA	CAAGAGCTTCTCCAGGGGG
<i>PIGU</i>	TTCATTTCCGAGCGGGTGG	TGCGGGGATGAAATCCCAAG
<i>PIGV</i>	CATGTTCAAGTTCTACCAAG	GCCTAGAATGTATCGTGTGAC
<i>PIGW</i>	AGCCATCTCCTGTTCCGTG	TGCACACCAGCCATGTGTAT
<i>PIGX</i>	CATAACAGAGGCAGTGATGG	CATTCTCAAAGCACAAGGGG
<i>PIGY</i>	TGTTCTACTCAGCCTGTG	CCCATCAAAGTCAAAGGTG
<i>PIGZ</i>	GCCCTGGAGTTTACCCC	GAAGGTGGAACCAGAGATCAGC
<i>DPM1</i>	ACAGAATTCTCTAACAGACCACG	CTCCATTCTTGTAGCGA
<i>DPM2</i>	TTAGCCTGATCATCTCACCT	ATGAACAGTCCCACAAACAG
<i>DPM3</i>	ATGACGAAATTAGCGCAGTGG	TTAGGCTGTCAGAACGCGAG
<i>GPAA1</i>	CCACGAGCGCTATATGGTGT	ATTGATGCCACGCAGGGTTA
<i>PGAP1</i>	TTCTATGTGCCTGCAAGGGG	ACCTCGTACCGACAGTCTGA
<i>PGAP2</i>	TCCCACCACTGGATCGG	GTGTGCTTCTGGTCAACCG
<i>PGAP3</i>	ACCTGTCGGGACGACTGTAA	AAGAGCGGTGGGAAGTCAAG
<i>SL15</i>	CGTTCACTGGGACTTGCTTC	GTGTGCCGTTGTGGTAGTT

Supplemental Table III. Tetramers for detection of virus specific T cells

	HLA	Virus	Protein	Peptide
Recipient 2	A*02:01	CMV	IE-1	VLEETSVML
Recipient 28	B*35:01	CMV	pp65	IPSINVHHY
Recipient 37	B*0801	CMV	IE-1	QIKVRVDMV
Recipient 37	B*0801	EBV	BZLF1	RAKFKQLL