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Dendrimer-encapsulated nanoparticle-core micelles as a modular strategy for particle-in-a-box-in-a-box nanostructures†

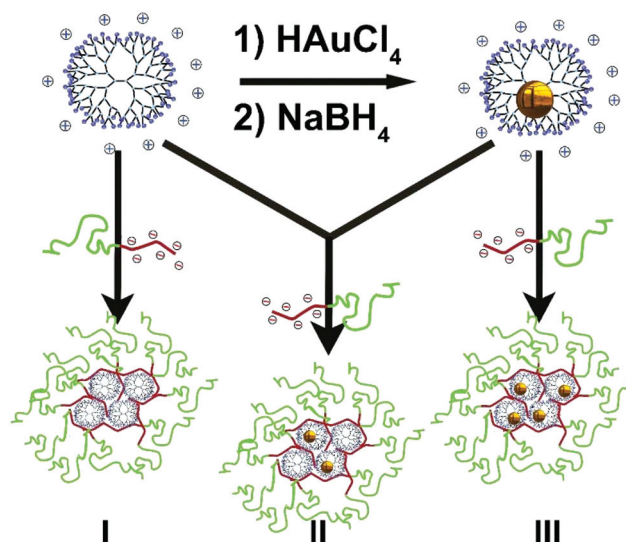
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The hierarchically controlled synthesis and characterization of self-assembling macromolecules and particles are key to explore and exploit new nanomaterials. Here we present a versatile strategy for constructing particle-in-a-box-in-a-box systems by assembling dendrimer-encapsulated gold nanoparticles (DENS) into dendrimicelles. This is realized by combining positively charged PAMAM dendrimers with a negative-neutral block copolymer. The number of particles per dendrimicelle can be controlled by mixing DENS with empty PAMAM dendrimers. The dendrimicelles are stable in solution for months and provide improved resistance for the nanoparticles against degradation. The dendrimicelle strategy provides a flexible platform with a plethora of options for variation in the type of nanoparticles, dendrimers and block copolymers used, and hence is tunable for applications ranging from nanomedicine to catalysis.

Introduction

The hierarchical organization of molecules, macromolecules, nanoparticles and self-assembled systems is at the heart of supramolecular and nanoscale chemistry, with nanocontainers being among the most appealing ones for applications ranging from drug delivery to catalysis.^{1–4} Within the vast interest in obtaining well-defined nanoparticles in confined systems, micelles and dendrimers have proved to be excellent building blocks. Dendrimers are hyperbranched molecules and are archetypical soft nanoparticles in the 1–10 nanometer range

with the potential to encapsulate molecules,⁵ or nanoparticles.^{6–8} Amis and Crooks and collaborators extensively studied the encapsulation of bare nanoparticles with polyamidoamino (PAMAM) dendrimers,^{8–10} and *via* various NMR techniques we unambiguously proved that (palladium) nanoparticles are formed and reside inside these dendrimers, thus forming so-called dendrimer-encapsulated nanoparticles (DENS).^{11,12} Micelles provide a method to self-assemble multiple, *e.g.*, polymeric, molecular building blocks into well-defined structures typically in the 10–100 nanometer range.^{13,14} In our investigations of coacervate core micellar systems for *e.g.* medical applications,^{15,16} we recently showed



Scheme 1 The formation of (particle-in)-a-box-in-a-box. After synthesis of dendrimer-encapsulated Au₂₅₆ nanoparticles inside 7th generation, amine-terminated, PAMAM dendrimers, the charge-stoichiometric addition of an anionic-neutral pMAA₆₄PEO₈₈₅ block copolymer results in the formation of dendrimicelles. The number of nanoparticles per dendrimer can be tuned by mixing empty with nanoparticle-filled dendrimers.

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the excellent control over the assembly of negatively charged, carboxylic acid-terminated, PAMAM dendrimers with a positive-neutral poly(vinylpyridine)-poly(ethylene oxide) diblock copolymer into well-defined micelles. These micelles contain 1–100 dendrimers per micelle, depending on the dendrimer generation used.¹⁷ We here present a methodology in which gold nanoparticles in amine-terminated dendrimers, AuDENS, can form micelles by assembling with negative-neutral block copolymers to form a nanoparticle-in-a-box-in-a-box system (Scheme 1).

Results and discussion

PAMAM dendrimers exist with a variety of terminating groups.^{18,19} For the incorporation of AuNPs, the use of (amine-terminated) PAMAMs is preferred, as PAMAM dendrimers with carboxylic acid end groups tend to form dendrimer stabilized nanoparticles rather than DENS,^{20,21} and hydroxyl-terminated dendrimers lack the required surface charge for self-assembly into coacervate core micelles. We synthesized Au₂₅₆DENS following the original work of Amis for the formation of well-defined single particles, focusing on amine-terminated 7th generation PAMAM.¹⁰ To prove that the AuNPs reside inside the PAMAM dendrimers, *e.g.* AuDENS as opposed to dendrimer-stabilized nanoparticles (Au-DSNPs), we used ¹H-NMR spectroscopy and DOSY (Fig. S1†) to verify the complexation of metal ions in the interior of PAMAM and the consecutive formation of dendrimer-encapsulated nanoparticles.^{11,12} In addition, Transmission Electron Microscopy (TEM) showed that the AuDENS are ~1.7 nm in diameter, in agreement with the expected size of Au₂₅₆ NPs (Fig. S2†).

Next, we assembled dendrimicelles by mixing the Au₂₅₆DENS at pH 7 with an anionic–neutral block copolymer having a 64-subunit polymethacrylic acid block and a 885-subunit polyethyleneoxide block, pMAA₆₄pEO₈₈₅. pH 7 was chosen since both the pMAApEO block copolymer as well as the PAMAM dendrimers are charged at this pH.^{22,23} This allows electrostatic interactions of the positively charged amine-terminal groups of PAMAM to associate with the negatively charged methacrylic acid block of the pMAApEO, forming a coacervate core. The neutral PEO block forms the corona of the micelle and stabilizes the resulting nanostructure. Dynamic light scattering, DLS, reveals this self-assembly as an increase in the scattered light intensity and, concurrently, as an increase in size (Fig. 1, S3†).

For the optimal building block ratios, *i.e.* the preferred micelle composition (PMC), we measured DLS at different mixing ratios of block copolymer to PAMAM, keeping the amount of positive charge (PAMAM) constant, and varying the amount of negative pMAA₆₄pEO₈₈₅ (Fig. S4†). From this experiment the PMC was determined to be at a mixing ratio of 0.92, corresponding to a slight excess of PAMAM to block copolymer. This deviation from the theoretical molar charge neutrality might be attributed to the known presence of defects in higher generation PAMAM dendrimers, resulting in fewer

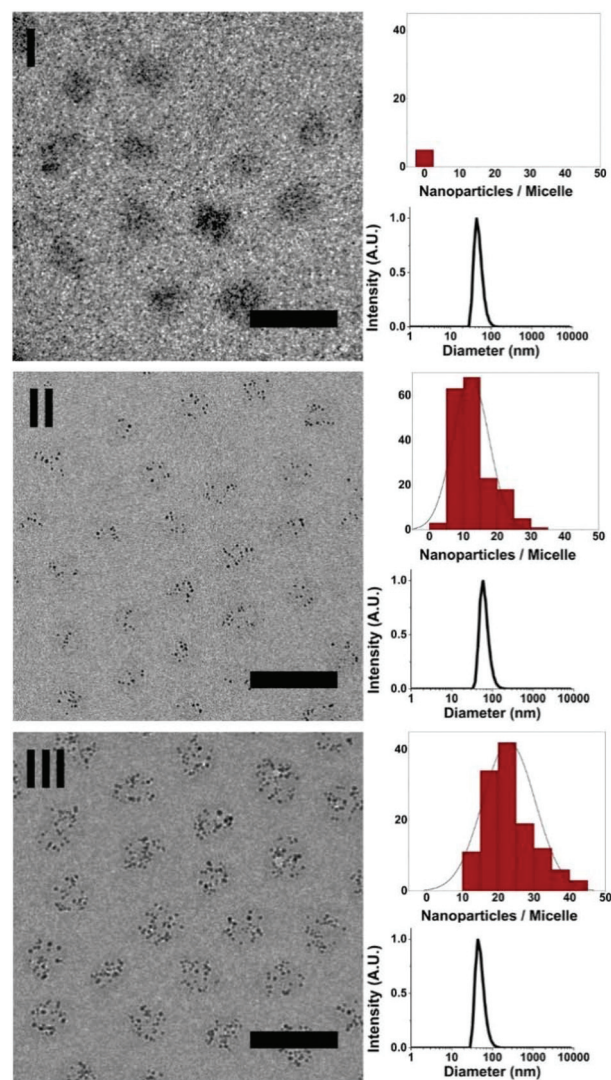


Fig. 1 Dendrimicelle characterization. In each panel, the figure on the left shows a cryoTEM image, and the figures on the right show the counted nanoparticles per micelle (upper), and number-averaged DLS size plot (lower). Dendrimicelles made from: top (I) empty dendrimers. The contrast observed originates from the dense dendrimicelle cores. Middle (II), from 50% Au₂₅₆-filled and 50% empty dendrimers; they contain 12 ± 5 AuNPs per micelle, and an equal amount of empty dendrimers which are not individually observable. Bottom (III), from Au₂₅₆-filled dendrimers. The dendrimicelles contain 23 ± 7 AuNPs per micelle. The AuNPs encapsulated inside the dendrimers greatly improve the contrast of the TEM images (of II and III) and allow for determining aggregation numbers. Scale bars are 50 nm.

charged end-groups and hence observed charge-stoichiometry at lower charge fraction.^{17,24,25}

To investigate whether or not the presence of a nanoparticle inside a dendrimer affects the formation of dendrimicelles, we mixed Au₂₅₆DENS with block copolymer at the previously determined PMC. DLS analysis showed that dendrimicelles formed and exhibited a similar size in solution as empty dendrimicelles. Additionally, we also prepared dendrimicelles by mixing a block copolymer with a dendrimer solution contain-



ing 50% Au₂₅₆DENs and 50% empty PAMAM dendrimers at the PMC. Here, similarly sized dendrimicelles were again formed. DLS only indicates the formation of the dendrimicelles revealing the hydrodynamic diameter; however, it cannot discriminate whether dendrimicelles with only PAMAM and dendrimicelles with DENs were formed. Therefore, we characterized the obtained nanostructures in more detail and determined their stability under various conditions using both cryoTEM and DLS. CryoTEM allows for visualization of the obtained nanostructures in near-native state from which micelle properties such as shape, micelle core and corona size, and aggregation number can be derived. Representative CryoTEM micrographs and DLS data of the dendrimicelles discussed above are shown in Fig. 1 and S5–7,† revealing the dendrimicelle structures for the PAMAM and DEN systems, and that mixing PAMAM and DENs result in mixed-core dendrimicelles.

The dendrimicelle sample with empty PAMAM in the core can be visualized by cryoTEM (I, Fig. 1) since the dense coacervate core provides a higher electron-density, and hence contrast, in the cryoTEM images.²⁶ However, the presence of electron dense AuNPs greatly improves the contrast and facilitates image analysis. In fact, Fig. 1 shows dendrimicelles made using 50% empty PAMAM mixed with 50% Au₂₅₆DENs (II), and shows the dendrimicelles made of 100% Au₂₅₆DENs (III). CryoTEM corroborates that the ~50 nm-sized structures observed with DLS are indeed well-defined nano-assemblies with a core-shell structure.

Cryo-TEM micrographs of the micelles made from 100% Au₂₅₆DENs (III, Fig. 1, S8†) show that the average core diameter is 27 ± 5 nm. DLS data show the hydrodynamic diameter of these micelles to be 52 nm. The expected size for a PEO₈₈₅ corona is ~14 nm,²⁷ hence showing the commensurability for the core size between the DLS (hydrodynamic radius minus corona) and cryoTEM (visible core) results. On the other hand, the micelle size (core + corona) obtained from the micelle core-core distance in the cryoTEM images is 39 ± 5 nm (Fig. S9†), which indicates a corona thickness of ~6 nm. This difference is attributed to the compression of the PEG shell as a result of cryoTEM sample preparation and packing.²⁸

The formation of dendrimicelles straightforwardly allows for incorporation and tuning of the number of NPs per micelle by mixing DENs with empty, amine-terminated PAMAM. Moreover, the presence of only one AuNP per PAMAM allows for the direct determination of the aggregation number of the dendrimicelles by simply counting the number of nanoparticles per micelle. The number of Au₂₅₆DENs per micelle is 23 ± 7 for the dendrimicelle sample made using 100% Au₂₅₆DENs and 12 ± 5 for the sample made using 50% empty PAMAM and 50% Au₂₅₆DENs. Assuming a 1 : 1 association of positive and negative charges in the dendrimicelle core, this corresponds to an average micelle molecular weight of ~12 MDa, roughly three times as heavy as the biggest proteins (Titins) found in nature.²⁹

Using DLS, we further studied the stability of dendrimicelles formed at the PMC against different environmental

conditions. Fig. S10† shows the observed scattering intensity and size of dendrimicelles *versus* pH. The dendrimicelles are stable between pH 6 and 8, in correspondence with the pK_a values of the charged building blocks (~6 for methacrylic acid,²³ and ~9 for amine-terminated PAMAM²²), supporting the hypothesis that the dendrimicelles are made by coacervation of the positively and negatively charged building blocks.

The critical salt concentration (CSC), a measure of the salt stability of (dendri)micelles formed from the self-assembly of oppositely charged building blocks, is 1.5 M for NaCl (Fig. S11†), and the critical micelle concentration (CMC) of these dendrimicelles is ~0.6 mg L⁻¹ total polymer concentration (Fig. S12†). Compared to dendrimicelles made from

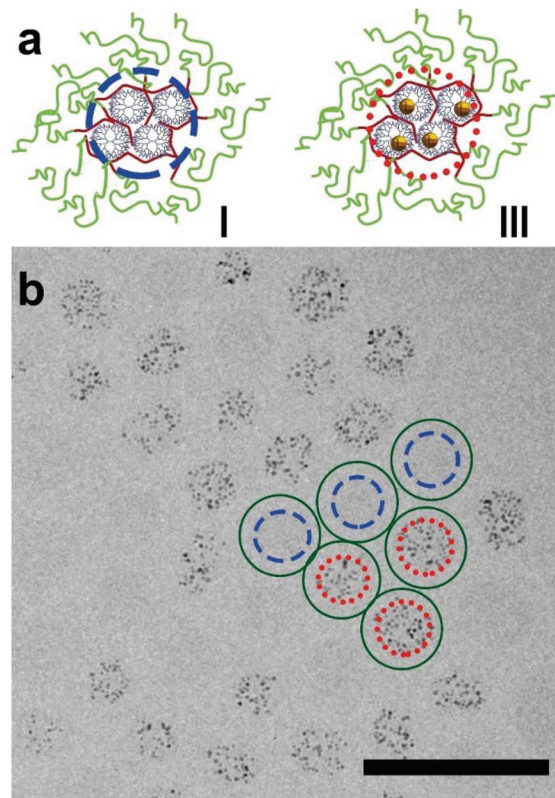


Fig. 2 The effect of mixing dendrimicelles formed with empty dendrimers (I) and dendrimicelles formed with Au-DENs, *i.e.* Au-nanoparticle filled dendrimers (III). (a) Schematic representation of the two dendrimicelles that were mixed: the blue, dashed circle is drawn around the core of a selection of three empty dendrimicelles (I) and the red, dotted, circle is drawn around the nanoparticle-filled dendrimer core of another selection of three micelles (III). (b) Cryo-TEM micrograph of a mix of dendrimicelles I and III, containing empty, and Au₂₅₆-filled dendrimers, respectively; these were separately prepared and consecutively mixed, and measured after 30 days. As can be seen, mixing empty and filled micelles does not result in half-filled micelles, showing the high stability of the dendrimicelles. For visualization purposes, the blue circles are drawn around the micelle-core of three empty dendrimicelles, whereas red-dotted lines are drawn around the cores of three dendrimicelles containing Au-DENs. Additional images can be found in the ESI.† The scale bar is 100 nm.



acid-terminated PAMAM dendrimers,¹⁷ both CSC and CMC are slightly improved, and in line with other micelle systems.¹⁴ DLS results obtained over time (Fig. S13†) show that both the scattered light intensity as well as the obtained size remained virtually constant over a period of seven days.

To investigate whether the stability of the dendrimicelle as a whole also is representative of the stability of the micelle core, *i.e.* to see whether the dendrimicelle core is dynamic, we mixed dendrimicelles made from empty PAMAM with dendrimicelles containing Au₂₅₆DENS. Here cryo-TEM micrographs recorded one day (Fig. S14†) or after 3 months at RT (Fig. 2 and S15/16†) both show dendrimicelles either without any AuDENSs, or completely filled with AuDENSs. Analysis of the dendrimicelles containing AuDENSs shows that these contain 26 ± 8 NPs per dendrimicelle, in agreement with the number of AuNPs per dendrimicelle found before (Fig. 1). Clearly, even at these timescales, no exchange between filled and empty dendrimicelles occurs. This implies that the dendrimer core is (kinetically) trapped and no exchange of dendrimers between the dendrimicelles takes place.

Besides the conceptual value, the advantage of a nano-container lies in the fact that properties of the contained particle/molecule are altered, *e.g.* protected, activated, and selectively accessible. The presented AuDENSs in dendrimicelles indeed have properties that are different from AuDENSs in solution; we used thiol-induced etching of AuNPs to study the differences.³⁰ Upon adding two molar equivalents of mercaptoethanol (ME) to Au atoms the AuDENSs are slowly degraded, the appearance of an absorption band at 395 nm (Fig. 3) being indicative of the formation of Au(I)-thiol complexes.^{31,32} In the case of the dendrimicelle sample, it takes about six times as long before this peak appears. Additionally, the rate at which the peak appears, *i.e.* the slope in the intensity *vs.* time plots, is also higher for the sample with just the Au₂₅₆NPs than for the dendrimicelle sample. These observations suggest that the dendrimicelle forms a barrier that delays degradation.

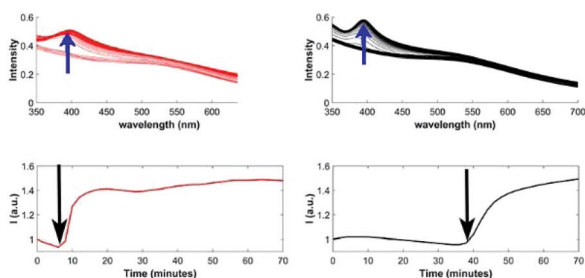


Fig. 3 Stability against degradation of Au₂₅₆DENS in solution or encapsulated inside dendrimicelles. Top: UV-Visible absorption spectra of Au₂₅₆DENS (red) and Au₂₅₆ dendrimicelles (black) after the addition of two equivalents of mercaptoethanol. Bottom: normalized absorption plots, showing absorption at 395 nm *versus* time after the addition of mercaptoethanol. For the Au₂₅₆ dendrimicelles, a significant delay in the first appearance of the Au(I)-thiol complex peak (indicated by an arrow), as well as smaller slope, is observed.

Conclusions

Nanoparticles, dendrimers and micelles find wide applications in fields like nanomedicine, responsive materials and catalysis,³³ and the controlled assembly strategy presented here opens up a vast horizon of applications. It offers a great variety for generating box-in-a-box structures and nanoparticle/molecule-in-a-box-in-a-box structures. There is, first, the possibility of changing the AuNP size by tuning the metal-dendrimer ratio during the synthesis of the DENSs. Second, the use of different dendrimer generations can be exploited. Third, the system allows for making dendrimicelles with different metal NPs or even mixed-metal NPs. Likewise, combinations can be made by mixing different generations, using different types of dendrimers, by encapsulating molecules inside the dendrimer, or even using other macromolecular systems. Finally, coacervate-core micelles form upon charge interactions, but other micellar strategies can be explored as well, *e.g.* based on hydrophobic or supramolecular interactions.

Conflicts of interest

There are no conflicts to declare.

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Notes and references

- 1 T. S. Koblenz, J. Wassenaar and J. N. H. Reek, *Chem. Soc. Rev.*, 2008, **37**, 247–262.
- 2 P.-C. Chen, G. Liu, Y. Zhou, K. A. Brown, N. Chernyak, J. L. Hedrick, S. He, Z. Xie, Q.-Y. Lin, V. P. Dravid, S. A. O'Neill-Slawecki and C. A. Mirkin, *J. Am. Chem. Soc.*, 2015, **137**, 9167–9173.
- 3 S. Mirtschin, A. Slabon-Turski, R. Scopelliti, A. H. Velders and K. Severin, *J. Am. Chem. Soc.*, 2010, **132**, 14004–14005.
- 4 B. M. Rosen, C. J. Wilson, D. A. Wilson, M. Peterca, M. R. Imam and V. Percec, *Chem. Rev.*, 2009, **109**, 6275–6540.
- 5 J. Jansen, E. M. M. de Brabander-Van den Berg and E. W. Meijer, *Science*, 1994, **266**, 1226–1229.
- 6 Z. Zhou, Y. Wang, Y. Yan, Q. Zhang and Y. Cheng, *ACS Nano*, 2016, **10**, 4863–4872.



- 7 L. Luo, L. Zhang, Z. Y. Duan, A. S. Lapp, G. Henkelman and R. M. Crooks, *ACS Nano*, 2016, **10**, 8760–8769.
- 8 S. V. Myers, M. G. Weir, E. V. Carino, D. F. Yancey, S. Pande and R. M. Crooks, *Chem. Sci.*, 2011, **2**, 1632–1646.
- 9 R. M. Crooks, M. Zhao, L. Sun, V. Chechik and L. K. Yeung, *Acc. Chem. Res.*, 2001, **34**, 181–190.
- 10 F. Gröhn, B. J. Bauer, Y. A. Akpalu, C. L. Jackson and E. J. Amis, *Macromolecules*, 2000, **33**, 6042–6050.
- 11 V. M. Gomez, J. Guerra, A. H. Velders and R. M. Crooks, *J. Am. Chem. Soc.*, 2009, **131**, 341–350.
- 12 V. M. Gomez, J. Guerra, S. V. Myers, R. M. Crooks and A. H. Velders, *J. Am. Chem. Soc.*, 2009, **131**, 14634–14635.
- 13 G. Riess, *Prog. Polym. Sci.*, 2003, **28**, 1107–1170.
- 14 I. K. Voets, A. de Keizer and M. A. C. Stuart, *Adv. Colloid Interface Sci.*, 2009, **147–148**, 300–318.
- 15 J. Y. Wang, A. H. Velders, E. Gianolio, S. Aime, F. J. Vergeldt, H. Van As, Y. Yan, M. Drechsler, A. de Keizer, M. A. C. Stuart and J. van der Gucht, *Chem. Commun.*, 2013, **49**, 3736–3738.
- 16 J. Y. Wang, A. Groeneveld, M. Oikonomou, A. Prusova, H. Van As, J. W. M. van Lent and A. H. Velders, *Soft Matter*, 2016, **12**, 99–105.
- 17 J. Wang, I. K. Voets, R. Fokkink, J. van der Gucht and A. H. Velders, *Soft Matter*, 2014, **10**, 7337–7345.
- 18 L. Balogh, R. Valluzzi, K. S. Laverdure, S. P. Gido, G. L. Hagnauer and D. A. Tomalia, *J. Nanopart. Res.*, 1999, **1**, 353–368.
- 19 R. Esfand and D. A. Tomalia, *Drug Discovery Today*, 2001, **6**, 427–436.
- 20 R. W. J. Scott, O. M. Wilson and R. M. Crooks, *J. Phys. Chem. B*, 2005, **109**, 692–704.
- 21 O. M. Wilson, R. W. J. Scott, J. C. Garcia-Martinez and R. M. Crooks, *J. Am. Chem. Soc.*, 2005, **127**, 1015–1024.
- 22 Y. Niu, L. Sun and R. M. Crooks, *Macromolecules*, 2003, **36**, 5725–5731.
- 23 L. Ruiz-Pérez, A. Pryke, M. Sommer, G. Battaglia, I. Soutar, L. Swanson and M. Geoghegan, *Macromolecules*, 2008, **41**, 2203–2211.
- 24 D. G. Mullen, A. Desai, M. A. van Dongen, M. Barash, J. R. Baker and M. M. Banaszak Holl, *Macromolecules*, 2012, **45**, 5316–5320.
- 25 M. A. van Dongen, A. Desai, B. G. Orr, J. R. Baker and M. M. B. Holl, *Polymer*, 2013, **54**, 4126–4133.
- 26 Y. Zheng, Y.-Y. Won, F. S. Bates, H. T. Davis, L. E. Scriven and Y. Talmon, *J. Phys. Chem. B*, 1999, **103**, 10331–10334.
- 27 C. J. Fee and J. M. Van Alstine, *Bioconjugate Chem.*, 2004, **15**, 1304–1313.
- 28 Y. Zheng, Z. Lin, J. L. Zakin, Y. Talmon, H. T. Davis and L. E. Scriven, *J. Phys. Chem. B*, 2000, **104**, 5263–5271.
- 29 S. Labeit and B. Kolmerer, *Science*, 1995, **270**, 293–296.
- 30 C.-Y. Ke, T.-H. Chen, L.-C. Lu and W.-L. Tseng, *RSC Adv.*, 2014, **4**, 26050–26056.
- 31 Y. Shichibu, Y. Negishi, H. Tsunoyama, M. Kanehara, T. Teranishi and T. Tsukuda, *Small*, 2007, **3**, 835–839.
- 32 T. G. Schaaff and R. L. Whetten, *J. Phys. Chem. B*, 1999, **103**, 9394–9396.
- 33 H. Ye and R. M. Crooks, *J. Am. Chem. Soc.*, 2005, **127**, 4930–4934.

