

Parkinson's disease age at onset genome-wide association study: Defining heritability, genetic loci, and alpha-synuclein mechanisms Blauwendraat, C.; Heilbron, K.; Vallerga, C.L.; Bandres-Ciga, S.; Coelln, R. von; Pihlstrom, L.; ...; IPDGC

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RESEARCH ARTICLE

Parkinson's Disease Age at Onset Genome-Wide Association Study: Defining Heritability, Genetic Loci, and α -Synuclein Mechanisms

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ABSTRACT: Background: Increasing evidence supports an extensive and complex genetic contribution to PD. Previous genome-wide association studies (GWAS) have shed light on the genetic basis of risk for this disease. However, the genetic determinants of PD age at onset are largely unknown. Objectives: To identify the genetic determinants of PD age at onset.

Methods: Using genetic data of 28,568 PD cases, we performed a genome-wide association study based on PD age at onset.

Results: We estimated that the heritability of PD age at onset attributed to common genetic variation was \sim 0.11, lower than the overall heritability of risk for PD (\sim 0.27), likely, in part, because of the subjective nature of this measure. We found two genome-wide significant association signals, one at *SNCA* and the other a protein-coding variant in *TMEM175*, both of which are known PD risk loci and a Bonferronicorrected significant effect at other known PD risk loci, *GBA*,

INPP5F/BAG3, *FAM47E/SCARB2*, and *MCCC1*. Notably, *SNCA*, *TMEM175*, *SCARB2*, *BAG3*, and *GBA* have all been shown to be implicated in α-synuclein aggregation pathways. Remarkably, other well-established PD risk loci, such as *GCH1* and *MAPT*, did not show a significant effect on age at onset of PD.

Conclusions: Overall, we have performed the largest age at onset of PD genome-wide association studies to date, and our results show that not all PD risk loci influence age at onset with significant differences between risk alleles for age at onset. This provides a compelling picture, both within the context of functional characterization of disease-linked genetic variability and in defining differences between risk alleles for age at onset, or frank risk for disease. © 2019 International Parkinson and Movement Disorder Society

Key Words: age at onset; *GBA*; Parkinson's disease; *SNCA*; *TMEM175*

Parkinson's disease (PD) is the most common neurodegenerative movement disorder. PD is pathologically characterized by the loss of dopaminergic neurons in the SN and α -synuclein (encoded by *SNCA*) protein aggregates. Current estimates are that in 2015 there were 6.9 million PD patients worldwide, and this number is predicted to be 14.2 million in 2040. PD has a strong age-dependent prevalence and male-female differences, where males are \sim 1.5 times more likely to develop PD.^{2,3}

The exact cause of PD is unknown; however, there is clear evidence that genetic variability plays a role in both disease development and progression. In the past two decades, mutations in several genes have been identified that cause monogenic forms of PD, accounting for around 1% to 5% of all PD cases. The majority of PD cases are therefore referred to as sporadic PD. Genome-wide association studies (GWAS) have successfully identified over 40 loci robustly associated with PD. Interestingly, variability in several genes, including SNCA, LRRK2, VPS13C, GCH1, and GBA, appear to play a role in both monogenic and sporadic disease.

Substantial evidence suggests that genetic variation plays a role in age at onset (AAO), where, for example, monogenic forms of PD often present as early-onset cases, but the identity of the exact genetic component in sporadic PD remains unclear. Additionally, there is some evidence that AAO may be different between males and females. Multiple studies have nominated variants and genes of interest, but replication and large cohort studies are lacking. Additionally, rare coding variants in PD risk genes like *GBA* and *LRRK2* have been associated with an earlier

AAO.¹²⁻¹⁷ Currently, the best predictor of PD AAO is a genetic risk score (GRS) based on cumulative genetic PD risk,^{7,18} implying broad genetic overlap between PD susceptibility and PD AAO.

In this study, we performed the largest PD AAO GWAS to date including 28,568 PD cases. Using this large cohort, we have estimated the heritability of PD AAO and have identified several associated variants. Additionally, we have shown that the latest PD GRS remains highly correlated with of AAO.

Materials and Methods

Processing of International Parkinson's Disease Genomics Consortium Data Sets

Genotyping data (all Illumina platform based) were obtained from International Parkinson's Disease Genomics Consortium (IPDGC) members, collaborators, and public resources (Supporting Information Table S1). All data sets underwent quality control separately, both on individual-level data and variant-level data, before imputation. See Supporting Information Methods for a detailed description of data processing. Where possible, AAO was defined based on patient report of initial manifestation of parkinsonian motor signs (tremor, bradykinesia, rigidity, or gait impairment). Where this information was not available, age of diagnosis was used as a proxy for onset age. Note that the correlation was high between age of diagnosis and AAO.

The resulting quality-controlled and imputed data sets of PD cases (total n = 17,415) were analyzed with the formula AGE_AT_ONSET ~ SNP + SEX + PC1-PC5. Analyses were performed per data set with rvtests linear regression using imputed dosages. ¹⁹ System Genomics of Parkinson's Disease (SGPD) data (n = 581) was processed as above with the following minor changes: A linear regression model in PLINK was used to run a GWAS for AAO, and relatedness cutoff of 0.05 was used. The association analysis was adjusted for sex and the first 10 eigenvectors from principal component analysis.

Processing of the 23andMe Data Set

As an independent second cohort, we used the 23 and Me PD data set (n = 10,572), which consisted of customers of the personal genetics company, 23andMe, Inc., who had consented to participate in research. PD patients were recruited through a targeted e-mail campaign in conjunction with the Michael J. Fox Foundation or other partners, patient groups, and clinics. PD cases were individuals who self-reported having been diagnosed with PD and who selfreported their age of PD diagnosis. We used age of diagnosis rather than age of symptom onset because symptom onset is often gradual and may be more difficult to selfreport accurately. Note that the correlation was high between age of diagnosis and AAO ($\rho = 0.917$; P < 1E-300), and the average difference between age of diagnosis and AAO was 2.58 years. We have previously shown that selfreported PD case status is accurate with 50 of 50 cases confirmed by telemedicine interview.²⁰ See Supporting Information Methods for a detailed description of data processing. After quality control, which included $R^2 > 0.5$ variant removal, we analyzed 11,956,580 single-nucleotide polymorphisms (SNPs). Remaining PD cases (n = 10.572)were analyzed using the formula AGE AT DIAGNOSIS ~ SNP + SEX + PC1-PC5 + genotyping platform. The genomic inflation factor was $\lambda = 1.015$.

Meta-Analyzing Data Set

The 17 IPDGC data sets were meta-analyzed using METAL (v.2011-03-25) fixed effects using default settings. We excluded SNPs with an I² statistic >50% and variants that were present in fewer than 66.7% of the data sets. One genome-wide significant variant was excluded because of this filter criteria (rs2737029, I² = 58.1%). This resulted in a total of 6,850,647 variants and a genomic inflation factor of λ = 1.001. For the combined GWAS, we meta-analyzed all 17 IPDGC data sets with the 23andMe data set using the same quality-control steps.

Additional Analyses and Figures

PD GWAS loci were obtained from using Table 1 and Table 2 of Chang et al 2017⁵ and using P.META from the provided summary stats (see Supporting Information Table S3). Additionally, to assess the influence of a GRS for

PD case-control status on PD AAO, we obtained the GRS from the most recent PD GWAS Supplementary. The GRS was calculated and processed using PLINK for each individual as described previously. Associations were performed using the formula AGE_AT_ONSET ~ GRS + SEX + PC1-PC5. Quantile-quantile (QQ)-plots and Manhattan plots were generated using FUMA.²² Forest plots were generated in R using the package rmeta (https://cran.r-project. org/web/packages/rmeta/index.html). Locus plots²³ were generated for the genome-wide significant loci and were compared to the latest published PD GWAS.⁵ GCTA-coio analyses were performed to identify whether there were multiple independent signals in loci of interest using all IPDGC data sets (excluding SGPD) described in Supporting Infromation Table S1 as a reference panel. 24,25 This reference panel was created using variants passing the following criteria: R² < 0.8, minor allele frequency >0.001, Hardy-Weinberg equilibrium P < 1E-6, and a maximum variant missingness of 5%. The genetic correlation between the PD AAO GWAS and the PD GWAS⁵ was calculated using linkage disequilibrium score regression (LDSC)²⁶ using default settings for European populations. Heritability was estimated using LDSC and GCTA based on GWAS summary statistics and individual-level genotypes, respectively.

Power Calculations

We performed power calculations using the method of Brion and colleagues:²⁷

Power =
$$1 - P(\chi_{NCP}^2 > \chi_{0.95}^2)$$

where χ^2_{NCP} is a random variable from a noncentral χ^2 with one degree of freedom, and $\chi^2_{0.95}$ is the threshold of a central χ^2 distribution with one degree of freedom and a type 1 error rate of 0.05. The noncentrality parameter was calculated using the method of Sham and Purcell:²⁸

$$\lambda = N \times \frac{2p(1-p) \times \beta^2}{variance(Y)}$$

where λ is the noncentrality parameter, p is the minor allele frequency, β is the effect size in years, and variance(Y) is the variance in PD AAO. We estimated variance(Y) by taking the mean variance across each cohort, weighted by their sample size. We assessed effect sizes between 0.01 and 1 years and reported the minimum effect size that yielded >80% power.

RESULTS

Initial Data Overview

In total, we included 18 data sets: the IPDGC data contain 17 independent cohorts (n = 17,996) and the 23andMe PD

cohort (n = 10,572; see Supporting Information Table S1 for more details). The average AAO in the IPDGC data set was 62.14 (range, 20–96; standard deviation [SD] = 12.08), whereas in the 23andMe dataset the average age of diagnosis was 60.71 (range, 40–97; SD = 9.98). We found minor differences in AAO in females and males in both the IPDGC data set (females = 62.15, SD = 11.71; males = 62.03, SD = 11.95) and the 23andMe data set (females = 59.95, SD = 9.79; males = 61.20, SD = 10.07).

Heritability of PD Age at Onset

Using LDSC, the heritability of PD AAO was $h^2 = 0.076$ (standard error [SE] = 0.0277) in the IPDGC cohort, $h^2 = 0.0805$ (SE = 0.0403) in the 23andMe cohort, and 0.109 (SE = 0.0255) in the complete meta-analysis. These heritability estimates for PD AAO were similar to estimates derived using GCTA in the largest IPDGC data set (IPDGC_NeuroX, $h^2 = 0.0798$; SE = 0.0391; P = 0.0184; N = 5,428) and in the 23andMe dataset ($h^2 = 0.1235$; SE = 0.0341; P = 1.031E-4; N = 10,697). Heritability estimates in the other IPDGC data sets were considered less reliable because of the low number of included cases.

SNCA as Top Associated Loci With PD Age at Onset

Two loci reached genome-wide significance, SNCA and TMEM175, both of which are established PD risk loci based on PD case/control GWASes (Fig. 1). Common variation at the SNCA locus has clearly been established as a risk factor PD, and both rare mutations and whole-gene multiplications have been identified to cause monogenic PD. ^{29,30} Several independent signals have been reported in this locus, where initially a variant in intron 4 was identified in PD;³¹ subsequently a second independent signal at the 5' end was identified, ³² which is also associated with dementia with Lewy bodies; ³³ and currently at least three independent signals are present in this locus.³⁴ Here, we identified both signals to be associated with AAO (Fig. 2A). The 3' end signal is the strongest with rs356203 as the most associated SNP, resulting in a reduction of ~ 0.6 year in AAO (P meta = 1.90E-12; beta = -0.626; SE = 0.0890). Using conditional analysis, we also identified the 5' end signal (Fig. 2B; rs983361, P conditional = 6.82E-6; beta = -0.484; SE = 0.108).

A Coding Variant in *TMEM175* Is Associated With PD Age at Onset

The *TMEM175/GAK* locus is another known PD locus where two independent signals have been identified.³² Whereas it is more parsimonious to assume that a single GWAS peak is the product of a single causal gene in the locus, there is evidence that both *TMEM175* and *GAK* may modulate PD risk.^{35,36} *TMEM175* and *GAK* share a promoter, suggesting that they may be coregulated and

may perform related cellular functions. Interestingly, the TMEM175/GAK locus is one of the few PD GWAS loci where a coding variant (TMEM175 p.M393 T, exon 11, rs34311866) is among the highest associated variants (Fig. 2A). This coding variant was also the most associated variant in this locus in our PD AAO analysis (P_meta = 9.62E-9; beta = -0.613; SE = 0.107), resulting in an average reduction of \sim 0.6 years (Figs. 3B and 5). Using conditional analysis based on rs34311866, we did not identify a second independent signal in this locus (P = 0.879; beta = -0.0240; SE = 0.158; Supporting Information Fig. S9).

PD High-Risk Variants in GBA and LRRK2 and Age at Onset

LRRK2 p.G2019S and GBA variants are the most commonly identified genetic risk factors for PD. LRRK2 p.G2019S is identified in ~1% of the general European PD population, but higher frequencies have been reported in other populations.³⁷ GBA variants are more commonly identified in Ashkenazi Jewish populations, but are also present in the European population.³⁸

LRRK2 p.G2019S (rs34637584) has already been described to have a large range of AAO and reduced penetrance. 12,14 Because of the low frequency and low imputation quality of this variant, it did not pass the variant-level quality control and was only tested in 9 of the 18 data sets. Similar to previous studies, ³⁷ we identified 140 carriers in the IPDGC data sets (\sim 0.8% of all PD cases). The average AAO for p.G2019S carriers was 66.58 (range = 36-95; SD = 11.66), which is \sim 4 years later compared to the average AAO of noncarriers in the IPDGC data sets (62.14; range, 20-96; SD = 12.08). However, this is based on a relatively small amount of cases, and some cohorts excluded LRRK2 p.G2019S carriers pregenotyping or excluded familial PD cases. Besides, the noncoding variant at the 5' end of LRRK2 (rs76904798), which is also identified as a risk factor PD did not show an association with AAO (P = 0.1267; beta = -0.184; SE = 0.121).

For GBA, there have been several reports that variants in GBA affect AAO in PD.³⁹⁻⁴¹ When solely looking at the GBA locus, we identified three significant hits for the GBA region including p.N370S variant (rs76763715, P = 2.628E-6; beta = -2.600; SE = 0.553), p.E326K (rs2230288, P = 0.00123; beta = -0.929; SE = 0.287 and p.T369M (rs75548401: P = 0.01153; beta = -1.281; SE = 0.507). All resulted in relatively large reduction AAO ranging from of 2.6 to 0.9 year. Note that the p.N370S variant was only tested in 13 of the 18 data sets because of the low frequency or low imputation quality. Using conditional analysis, we did not identify other clear significant signals in the GBA gene (Supporting Information Fig. S11).

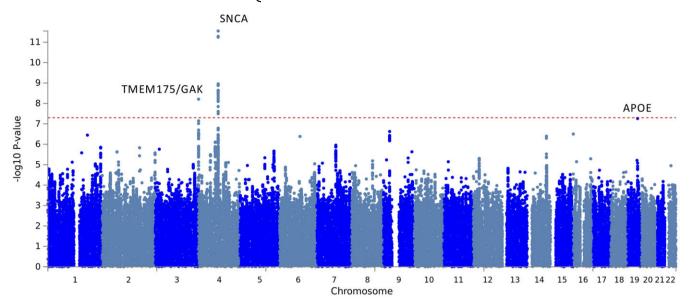


FIG. 1. Manhattan plot of Parkinson's disease age at onset GWAS. Based on meta-analyses of data sets (n = 28,568) using 7,426,111 SNPs. Two genome-wide significant loci were identified: *SNCA* and the *TMEM175/GAK*. Additionally, one borderline significant locus was found, *APOE*. [Color figure can be viewed at wileyonlinelibrary.com]

PD Genetic Risk Loci Are Associated With Age at Onset

Previously, a GRS based on PD GWAS loci has been identified as the main genetic predictor for AAO. Here, we tested the association between the updated 47-SNP GRS used in the latest PD GWAS study. For each IPDGC data set, the GRS was calculated and we identified a consistent association between the Chang and colleagues GRS and PD AAO. After meta-analyzing results from the individual cohorts, we found that each 1-SD increase in GRS led to an earlier AAO by \sim 0.8 years (summary effect = -0.801; 95% confidence interval CI: -0.959, -0.643; $I^2 = 11.6\%$; Supporting Information Fig. S12). Although this effect was highly significant, the genetic proportion explained by the

GRS was $\sim 0.56\%$, and the *SNCA* and *TMEM175* loci combined explained $\sim 0.3\%$, most likely attributed to the lower heritability. Furthermore, there was a significantly negative genetic correlation between the AAO GWAS summary statistics and the most recent PD GWAS⁵ using LDSC (rg = -0.5511; se = 0.103; P = 9.027E-8).

When solely looking at the SNPs that were genome-wide significant in the most recent PD GWAS,⁵ we identified a significant effect in six loci after Bonferroni correction: *SNCA*, *TMEM175*, *BST1*, *INPP5F/BAG3*, *FAM47E/SCARB2*, and *MCCC1* (Fig. 4; Supporting Information Table S3). Interestingly, for some well-established loci, no significant *P* values were identified in any of the data sets, including *GCH1* (P_meta = 0.9769; 80% power to detect

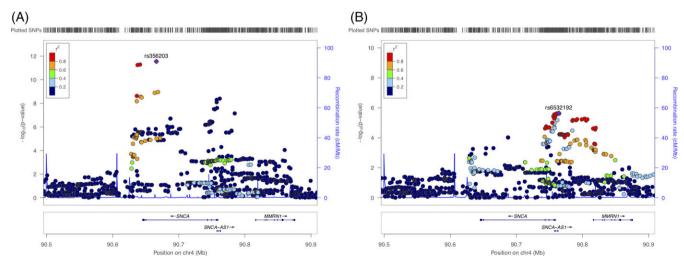


FIG. 2. SNCA association with Parkinson's disease age at onset. (A) Locus zoom plot of the association signal of the SNCA locus. The age at onset association signal is primarily based at the 3' end of the SNCA gene, highly similar as the PD GWAS signal of Chang and colleagues⁵ (Supporting Information Fig. S8). (B) Conditional analysis based on the most associated SNP (rs356203) shows that there is a secondary signal at the 5' end of the SNCA gene rs6532192, as well similar as previously reported for the PD GWAS.⁵ [Color figure can be viewed at wileyonlinelibrary.com]

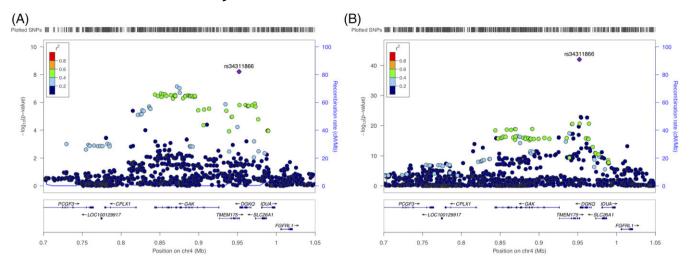


FIG. 3. *TMEM175/GAK* locus association with Parkinson's disease age at onset and Parkinson's disease. (A) Locus zoom plot of the association signal of the *TMEM175/GAK* locus with PD age at onset. The association signal is primarily based on the coding variant *TMEM175* p.M393T, rs34311866. (B) Locus zoom plot of the association signal of the *TMEM175/GAK* locus with PD versus controls. Similarly, as in (A), the main signal is based on the coding variant *TMEM175* p.M393 T, rs34311866. [Color figure can be viewed at wileyonlinelibrary.com]

a change in AOO >0.28 years), *RAB7L1*/NUCKS1 (PARK16; P_meta = 0.124; 80% power to detect a change in AOO >0.26 years), and *MAPT* (P_meta = 0.745; 80% power to detect a change in AOO >0.32 years; Fig. 5; see Supporting Information Table S3 for all power calculations).

APOE E4 Allele as Positive Control Longevity Marker

Interestingly, the variant representing the *APOE* E4 allele (rs429358) was among the borderline significant loci with a P_meta value of 5.696E-8 resulting in a 0.7 year earlier of AAO (beta = -0.707; SE = 0.130). The *APOE* E4 allele

is the most common genetic risk factor for Alzheimer's disease, ⁴² but has no effect on PD (*P* value in latest GWAS = 0.625).^{32,43} Interestingly, it has already been previously identified as an aging marker.^{44,45} To investigate this in our data, we performed a GWAS on the reported age of controls from the IPDGC data sets and this demonstrated a result consistent with that observed in the PD cases: *P* value of 1.49E-5 (effect = -0.644; SE = 0.149), suggesting that associations observed at this locus are general age-related effects (Fig. 5). Conditional tests based on rs429358 showed that the whole signal is coming from the *APOE* E4 allele (Supporting Information Fig. S10).

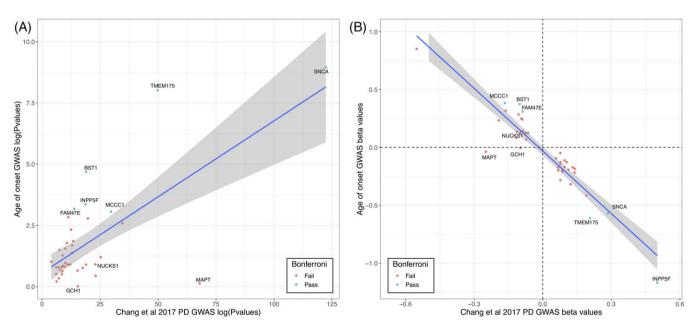


FIG. 4. *P*-value and beta-value correlation plot between Parkinson's disease GWAS and age at onset GWAS. (A) Log-transformed *P* values were plotted of the Chang and collegaues⁵ PD GWAS (x-axis) and the current age at onset GWAS (y-axis). SNPs are annotated by their closest gene. Green dots are loci that pass Bonferroni correction and red are loci that did not pass Bonferroni correction. (B) Beta values were plotted of the Chang and colleagues⁵ PD GWAS (x-axis) and the current age at onset GWAS (y-axis). SNPs are annotated by their closest gene. Green dots are loci that pass Bonferroni correction, and red are loci that did not pass Bonferroni correction. [Color figure can be viewed at wileyonlinelibrary.com]

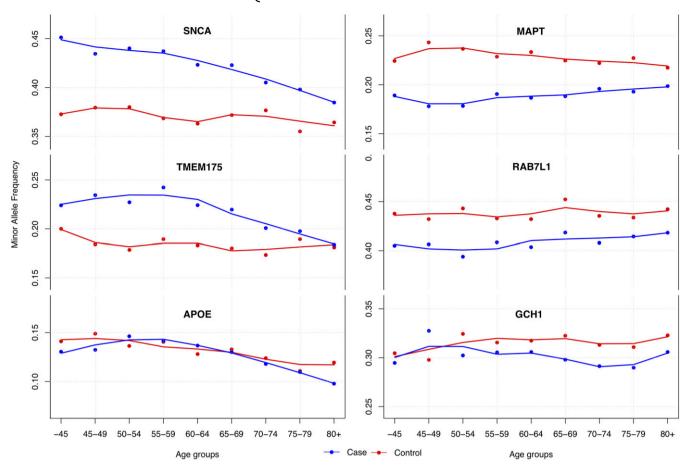


FIG. 5. Minor allele frequency differences correlated with age groups separated by case-control status. Genotype data were merged and minor allele frequency was calculated per age group and separated by case-control status (IPDGC data only: N_cases = 17,996 and N_controls = 16,502; each age group contains >850 individuals). Lines were using LOESS regressions based on the minor allele frequency per age group. On the left, the three most significant loci associated with PD age at onset are shown; *SNCA* = rs356203, *TMEM175/GAK* = rs34311866, and *APOE* = rs429358. On the right, three significant PD case-control GWAS loci are shown to have no clear effect on PD age at onset; *MAPT* = rs17649553, *RAB7L1* = rs823118, and *GCH1* = rs11158026. Standard-error bars are not shown in the figure, but are neglectable (~0.015) for each age group. LOESS, locally estimated scatterplot smoothing. [Color figure can be viewed at wileyonlinelibrary.com]

Additionally, none of the PD loci in the control dataset had a significant P value after correction for multiple testing (Supporting Information Table S5).

Replication of Previous Associated Loci and Potential Sex Effects

In the past decade, several studies have performed PD AAO analyses and nominated many genes and variants of interest (Supporting Information Table S2). Many of these variants and genes were identified in smaller data sets and were reported with nominal *P* values or could not be replicated in other data sets. In our current GWAS, we replicate the findings in *SNCA*, *GBA*, and *TMEM175* as described above (Supporting Information Table S2).

Additionally, some evidence exists there that there might be sex differences in AAO. To investigate this, we divided the IPDGC data sets (excluding the SGPD data set) into a males-only data set (N=11,411) and a females-only data set (N=6,621) and performed sex-specific AAO GWASes and meta-analyses. No genome wide-significant hits were identified in either the male or the female meta-analysis, and

the effect sizes for the most recent PD GWAS variants Chang and colleagues were similar (Pearson correlation = 0.532; P = 0.00034; Supporting Information Table S4). Interestingly, we did identify a similar sex-specific effect of the COMT coding variant rs4680 (p.V158M) as reported previously: P = 0.0133 in males (effect = 0.370; SE = 0.150) and P = 0.922 in females (effect = -0.0188; SE = 0.193).

Discussion

Here, we performed a genome-wide AAO analysis using 28,568 PD cases and discovered several PD loci associated with AAO. We identified two genome-wide significantly associated loci—SNCA (P_meta = 1.90E-12) and TMEM175 (P_meta = 9.62E-9)—as well as several subsignificant loci including GBA, SCARB2, and BAG3. Notably, this is the first study reporting genome-wide significance for PD GWAS loci for PD AAO. The heritability estimate of PD AAO was \sim 0.11, which is much less than the heritability of PD case-control status \sim 0.27. 5,47 Our heritability estimate was lower than an

estimate from a previous study in familial PD, although this was based on a relatively small data set (n = 504 families).⁶ This is likely, in large part, attributed to the subjective nature of this phenotype.

It has been shown that PD case-control GWAS loci are associated with PD AAO.7 We found that a GRS based on the latest PD GWAS⁵ was also highly associated with an earlier age of PD onset (Supplementary Figure 12). Furthermore, there was a significant negative genetic correlation between the PD case-control GWAS and the PD AAO GWAS. Six individual loci remained significant after Bonferroni correction, including two well-established PD loci—SNCA and TMEM175—and four less-studied PD loci—BST1, INPP5F/BAG3, FAM47E/SCARB2, and MCCC1. Interestingly, no effect was identified for several well-established loci, including GCH1.(PARK16), and MAPT (Supporting Information Table S3). For these three loci, our study had 80% power (at P = 0.05) to detect changes greater than 0.28, 0.26, and 0.32 years, respectively. For comparison, the two genomewide significant hits in SNCA and TMEM175 modified AAO by ~0.6 year. We believe this provides compelling evidence that only a subset of PD case-control risk SNPs also modulates PD AAO, and notably that variability at MAPT, a major GWAS identified risk factor for PD, does not influence AAO. We believe these data may suggest that different PD loci modulate risk through different pathways and that this likely has therapeutic consequences.

Interestingly, three of our top hits (SNCA, TMEM175, and GBA) are strongly implicated in α-synuclein mechanisms of PD pathogenesis pathology. SNCA encodes for the α-synuclein protein, which is the major constituent of Lewy bodies, the defining pathology of PD. Both common and rare variants at SNCA are associated with increased PD risk, including duplications or triplications of the genomic locus.²⁹ One of the distinguishing features of autosomal-dominant PD caused by SNCA gene multiplications is an early AAO. 48,49 The common variants at SNCA associated with PD risk and AAO have also been demonstrated to enhance SNCA gene expression. 34,50 Thus, based on the genetics of both Mendelian and sporadic PD, SNCA dosage appears to be a major driver of disease onset age. TMEM175 has been shown to impair lysosomal and mitochondrial function and increases α-synuclein aggregation.³⁶ GBA has been shown to be involved in several α -synuclein aggregation mechanisms, including autophagy and enhancing α -synuclein cell-to-cell transmission. ^{51–53} Other loci identified by our PD AAO analyses have also been linked to α-synuclein clearance, for example for SCARB2, encoding the endoplasmic reticulum-to-lysosome transporter of GBA, and BAG3.^{54,55} Taken together, it is tempting to speculate about the direct link between PD AAO loci influencing α-synuclein accumulation and clearance pathways and the other nonsignificant PD loci acting in another pathway by a different mechanism, but more experiments are needed to confirm this.

Several previous studies have performed PD AAO analyses in smaller data sets. We were unable to replicate the majority of these previous reported variants, likely because of the lack of power to detect a true signal in the smaller studies (<2,000 samples; see Supporting Information Table S2). We may have been unable to replicate previous work by Hill-Burns and colleagues, given that much of their analysis was restricted to familial PD cases. ¹¹ Their analysis of sporadic PD included almost 4,000 cases, but unlike our study and several others, ^{7,40,56} they did not find significant associations with SNCA (rs356203, P = 0.355) or TMEM175 (rs34311866, P = 0.525).

Although we included a very large amount of data, there were still several limitations to our study. First, we had limited power to detect rare variants that modulate PD AAO because of the use of genotyping arrays and large number of cohorts with a relatively small size. Several reports have shown that rare variants may influence AAO of PD. For example, a recent report showed that rare variants in *LRRK2* lowered AAO. ¹⁶ This likely also explains the relatively moderate *P* values for rare coding variants in *GBA* and *LRRK2* on age onset.

Second, because of the lack of reliable genotype data, we excluded chromosomes X and Y from our analysis. Both of these limitations could be addressed by using genome sequencing data, and large genome sequencing projects are currently underway. Third, the heritability of PD AAO (~ 0.11) was much lower than the heritability of PD casecontrol status, ⁴⁷ which, in part, explains the lack of genomewide significantly associated loci in our study. Fourth, there was a reasonable amount of heterogeneity in the mean AAO our different cohorts. Age of PD diagnosis was self-reported in some of our cohorts and assessed by physicians in other cohorts. Some cohorts were specifically designed to include younger-onset cases. Other cohorts provided age of diagnosis whereas others used AAO, although these are likely to be highly correlated (r > 0.9 in our data). By analyzing each cohort separately, we were able to mitigate many of these interstudy sources of variation. Indeed, there was relatively little heterogeneity of SNP effect-size estimates between studies (Supporting Information Table S3). However, future data collection and studies would probably benefit from more specific and more predefined structured and symptomspecific AAO diagnostic criteria. Implementation of such criteria in large studies and cohorts or even in health care systems could significantly improve the understanding of the genetics of PD AAO.

Overall, we have performed the largest AAO of PD GWAS to date, and our results reveal an interesting and significant genetic component. Our results show that not all PD risk loci influence AAO with significant differences between risk alleles for AAO, which implies different mechanisms for risk for developing PD and PD AAO. This provides a compelling picture, both within the context of functional characterization of disease-linked genetic variability and in defining differences between risk alleles for AAO, or frank risk for

disease. The significantly associated variability is centered on the gene encoding α -synuclein and on variability in several other lysosomal proteins that have been shown to directly influence α -synuclein aggregation or clearance. Thus, these data support the notion that α -synuclein is an important target for future disease-modifying or -preventive therapies and that drugs targeting α -synuclein production and clearance may be most valuable.

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Data Availability

IPDGC GWAS summary statistics are available on the IPDGC website (http://pdgenetics.org/resources), and 23andMe GWAS summary statistics will be made available to qualified researchers under an agreement with 23andMe that protects the privacy of the 23andMe participants. Please visit http://research.23andme.com/collaborate/#publication for more information and to apply to access the data.

References

- Dorsey ER, Bloem BR. The Parkinson pandemic—a call to action. JAMA Neurol 2018;75:9–10.
- Reeve A, Simcox E, Turnbull D. Ageing and Parkinson's disease: why is advancing age the biggest risk factor? Ageing Res Rev 2014;14:19–30.
- Moisan F, Kab S, Mohamed F, et al. Parkinson disease male-to-female ratios increase with age: French nationwide study and meta-analysis. J Neurol Neurosurg Psychiatry 2016;87:952–957.
- Singleton A, Hardy J. The evolution of genetics: Alzheimer's and Parkinson's diseases. Neuron 2016;90:1154–1163.
- Chang D, Nalls MA, Hallgrímsdóttir IB, et al. A meta-analysis of genome-wide association studies identifies 17 new Parkinson's disease risk loci. Nat Genet 2017;49:1511–1516.
- Hamza TH, Payami H. The heritability of risk and age at onset of Parkinson' disease after accounting for known genetic risk factors. J Hum Genet 2010;55:241–243.
- Nalls MA, Escott-Price V, Williams NM, et al. Genetic risk and age in Parkinson's disease: continuum not stratum. Mov Disord 2015; 30:850-854
- Alves G, Müller B, Herlofson K, et al. Incidence of Parkinson's disease in Norway: the Norwegian ParkWest study. J Neurol Neurosurg Psychiatry 2009;80:851–857.
- Haaxma CA, Bloem BR, Borm GF, et al. Gender differences in Parkinson's disease. J Neurol Neurosurg Psychiatry 2007;78:819–824.
- Latourelle JC, Pankratz N, Dumitriu A, et al. Genomewide association study for onset age in Parkinson disease. BMC Med Genet 2009;10:98.
- Hill-Burns EM, Ross OA, Wissemann WT, et al. Identification of genetic modifiers of age-at-onset for familial Parkinson's disease. Hum Mol Genet 2016;25:3849–3862.
- Lee AJ, Wang Y, Alcalay RN, et al. Penetrance estimate of LRRK2 p.G2019S mutation in individuals of non-Ashkenazi Jewish ancestry. Mov Disord 2017;32:1432–1438.
- Malek N, Weil RS, Bresner C, et al. Features of -associated Parkinson's disease at presentation in the UKstudy. J Neurol Neurosurg Psychiatry 2018;89:702–709

- Marder K, Wang Y, Alcalay RN, et al. Age-specific penetrance of LRRK2G2019S in the Michael J. Fox Ashkenazi Jewish LRRK2 Consortium. Neurology 2015;85:89–95.
- Nichols WC, Pankratz N, Marek DK, et al. Mutations in GBA are associated with familial Parkinson disease susceptibility and age at onset. Neurology 2009;72:310–316.
- Xiao B, Deng X, Ng EY, et al. Association of LRRK2 haplotype with age at onset in Parkinson disease. JAMA Neurol 2018;75:127–128.
- Gan-Or Z, Amshalom I, Kilarski LL, et al. Differential effects of severe vs mild GBA mutations on Parkinson disease. Neurology 2015;84:880–887.
- Escott-Price V, International Parkinson's Disease Genomics Consortium, Nalls MA, et al. Polygenic risk of Parkinson disease is correlated with disease age at onset. Ann Neurol 2015;77:582–591.
- Zhan X, Hu Y, Li B, Abecasis GR, Liu DJ. RVTESTS: an efficient and comprehensive tool for rare variant association analysis using sequence data. Bioinformatics 2016;32:1423–1426.
- Dorsey ER, Ray Dorsey E, Darwin KC, et al. Virtual research visits and direct-to-consumer genetic testing in Parkinson's disease. DIGI-TAL HEALTH 2015;1:205520761559299.
- Willer CJ, Li Y, Abecasis GR. METAL: fast and efficient metaanalysis of genomewide association scans. Bioinformatics 2010;26: 2190–2191.
- Watanabe K, Taskesen E, van Bochoven A, Posthuma D. Functional mapping and annotation of genetic associations with FUMA. Nat Commun 2017;8:1826.
- Pruim RJ, Welch RP, Sanna S, et al. LocusZoom: regional visualization of genome-wide association scan results. Bioinformatics 2010; 26:2336–2337.
- Yang J, Ferreira T, Morris AP, et al. Conditional and joint multiple-SNP analysis of GWAS summary statistics identifies additional variants influencing complex traits. Nat Genet 2012;44:369–375, S1–S3.
- Yang J, Lee SH, Goddard ME, Visscher PM. GCTA: a tool for genomewide complex trait analysis. Am J Hum Genet 2011;88:76–82.
- Bulik-Sullivan BK, Loh PR, Finucane HK, et al. LD Score regression distinguishes confounding from polygenicity in genome-wide association studies. Nat Genet 2015;47:291–295.
- Brion MJ, Shakhbazov K, Visscher PM. Calculating statistical power in Mendelian randomization studies. Int J Epidemiol 2013;42:1497–1501.
- Sham PC, Purcell SM. Statistical power and significance testing in large-scale genetic studies. Nat Rev Genet 2014;15:335–346.
- Singleton AB, Farrer M, Johnson J, et al. alpha-Synuclein locus triplication causes Parkinson's disease. Science 2003;302:841.
- Polymeropoulos MH, Lavedan C, Leroy E, et al. Mutation in the alpha-synuclein gene identified in families with Parkinson's disease. Science 1997;276:2045–2047.
- Simón-Sánchez J, Schulte C, Bras JM, et al. Genome-wide association study reveals genetic risk underlying Parkinson's disease. Nat Genet 2009:41:1308–1312.
- Nalls MA, Pankratz N, Lill CM, et al. Large-scale meta-analysis of genome-wide association data identifies six new risk loci for Parkinson's disease. Nat Genet 2014;46:989–993.
- 33. Guerreiro R, Ross OA, Kun-Rodrigues C, et al. Investigating the genetic architecture of dementia with Lewy bodies: a two-stage genome-wide association study. Lancet Neurol 2018;17:64–74.
- Pihlstrom L, Blauwendraat C, Cappelletti C, et al. A comprehensive analysis of SNCA-related genetic risk in sporadic Parkinson disease. Ann Neurol 2018;84:117–129.
- Dumitriu A, Pacheco CD, Wilk JB, et al. Cyclin-G-associated kinase modifies α-synuclein expression levels and toxicity in Parkinson's disease: results from the GenePD Study. Hum Mol Genet 2011;20:1478–1487.
- Jinn S, Drolet RE, Cramer PE, et al. TMEM175 deficiency impairs lysosomal and mitochondrial function and increases α-synuclein aggregation. Proc Natl Acad Sci U S A 2017;114:2389–2394.
- Correia Guedes L, Ferreira JJ, Rosa MM, Coelho M, Bonifati V, Sampaio C. Worldwide frequency of G2019S LRRK2 mutation in Parkinson's disease: a systematic review. Parkinsonism Relat Disord 2010;16:237–242.

- Sidransky E, Nalls MA, Aasly JO, et al. Multicenter analysis of glucocerebrosidase mutations in Parkinson's disease. N Engl J Med 2009;361:1651–1661.
- Gan-Or Z, Giladi N, Rozovski U, et al. Genotype-phenotype correlations between GBA mutations and Parkinson disease risk and onset. Neurology 2008;70:2277–2283.
- Lill CM, Hansen J, Olsen JH, Binder H, Ritz B, Bertram L. Impact of Parkinson's disease risk loci on age at onset. Mov Disord 2015; 30:847–850.
- Davis AA, Andruska KM, Benitez BA, Racette BA, Perlmutter JS, Cruchaga C. Variants in GBA, SNCA, and MAPT influence Parkinson disease risk, age at onset, and progression. Neurobiol Aging 2016;37:209. e1–209.e7.
- 42. Liu CC, Liu CC, Kanekiyo T, Xu H, Bu G. Apolipoprotein E and Alzheimer disease: risk, mechanisms and therapy. Nat Rev Neurol 2013;9:106–118.
- Federoff M, Jimenez-Rolando B, Nalls MA, Singleton AB. A large study reveals no association between APOE and Parkinson's disease. Neurobiol Dis 2012;46:389–392.
- Garatachea N, Emanuele E, Calero M, et al. ApoE gene and exceptional longevity: Insights from three independent cohorts. Exp. Gerontol 2014;53:16–23.
- Broer L, Buchman AS, Deelen J, et al. GWAS of longevity in CHARGE consortium confirms APOE and FOXO3 candidacy. J Gerontol A Biol Sci Med Sci 2015;70:110–118.
- Klebe S, Golmard JL, Nalls MA, et al. The Val158Met COMT polymorphism is a modifier of the age at onset in Parkinson's disease with a sexual dimorphism. J Neurol Neurosurg Psychiatry 2013;84: 666–673.
- Keller MF, Saad M, Bras J, et al. Using genome-wide complex trait analysis to quantify 'missing heritability' in Parkinson's disease. Hum Mol Genet 2012;21:4996–5009.
- Konno T, Ross OA, Puschmann A, Dickson DW, Wszolek ZK. Autosomal dominant Parkinson's disease caused by SNCA duplications. Parkinsonism Relat Disord 2016;229(Suppl 1):S1–S6.
- Ross OA, Braithwaite AT, Skipper LM, et al. Genomic investigation of α-synuclein multiplication and parkinsonism. Ann Neurol 2008; 63:743–750.
- Soldner F, Stelzer Y, Shivalila CS, et al. Parkinson-associated risk variant in distal enhancer of α-synuclein modulates target gene expression. Nature 2016;533:95–99.
- Du TT, Wang L, Duan CL, et al. GBA deficiency promotes SNCA/αsynuclein accumulation through autophagic inhibition by inactivated PPP2A. Autophagy 2015;11:1803–1820.
- Mazzulli JR, Xu YH, Sun Y, et al. Gaucher disease glucocerebrosidase and α-synuclein form a bidirectional pathogenic loop in synucleinopathies. Cell 2011;146:37–52.
- Bae EJ, Yang NY, Song M, et al. Glucocerebrosidase depletion enhances cell-to-cell transmission of α-synuclein. Nat Commun 2014;5:4755.
- Cao YL, Yang YP, Mao CJ, et al. A role of BAG3 in regulating SNCA/α-synuclein clearance via selective macroautophagy. Neurobiol Aging 2017;60:104–115.
- Rothaug M, Zunke F, Mazzulli JR, et al. LIMP-2 expression is critical for β-glucocerebrosidase activity and α-synuclein clearance. Proc Natl Acad Sci U S A 2014;111:15573–15578.
- Brockmann K, Schulte C, Hauser AK, et al. SNCA: major genetic modifier of age at onset of Parkinson's disease. Mov Disord 2013; 28:1217–1221.

Supporting Data

Additional Supporting Information may be found in the online version of this article at the publisher's web-site.