

Cholesterol metabolism and hematopoiesis interaction in atherothrombosis

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General introduction and thesis outline

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PREFACE

Atherosclerosis and its complications are the underlying cause for most cardiovascular diseases. It is a chronic, multifactorial disease in which lipid accumulates in the arterial wall, leading to a local inflammatory reaction and atherosclerotic plaque formation. Atherosclerotic disease develops largely asymptomatic over a lifetime. However, plaque rupture or erosion can cause the formation of a superimposed thrombus blocking the flow of blood. Upon rupture or erosion of an atherosclerotic plaque, platelets, the body's first responders upon endothelial disruption, initiate the formation of a platelet plug, which activates the coagulation cascade to form an arterial thrombus. Although in principle a protective response, this thrombus formation can cause acute cardiovascular events such as myocardial infarction or ischemic stroke $1,2$.

Bone marrow derived cells from both the innate and adaptive immune system, especially monocytes/macrophages and T cells, and platelets have been implicated in the pathophysiology of atherosclerosis¹, The primary site of immune cell and platelet production is the bone marrow. During bone marrow hematopoiesis, hematopoietic stem cells (HSCs) mature and differentiate until the formed immune cells and platelets enter the circulation or home towards their site of action or organ for final maturation 3 . During homeostasis, hematopoiesis is tightly balanced between the different blood cell lineages. In that way, by controlling the output of immune cells and platelets, hematopoiesis plays a major role in the resolution or persistence of inflammatory diseases, such as atherosclerosis. An increased number of peripheral blood leukocytes, especially monocytes and granulocytes, as well as increased platelet counts and reactivity is associated with occurrence and outcome of cardiovascular disease⁴⁻¹⁰. Interestingly, defects in cholesterol metabolism and hypercholesterolemia have been shown to affect hematopoiesis, immune cell production and platelet counts and reactivity 11,12. Therefore, bone marrow cholesterol handling is an interesting target in the battle against cardiovascular disease.

This chapter summarizes relevant information to understand the role of cholesterol metabolism in the production and reactivity of immune cells and platelets, and its effects on atherosclerosis and atherothrombosis development.

HEMATOPOIESIS

All circulating blood cells, i.e. erythrocytes, leukocytes and platelets, originate from bone marrow, in which they are formed from HSCs through hematopoiesis. Hematopoiesis is divided into three main branches: the myeloid, lymphoid and megakaryocyte-erythroid arms. HSCs constitute the top of the hematopoietic tree and give rise to committed

progenitor cells, which in turn mature into differentiated cells that home towards their site of action or organ for final maturation, Figure 1.

HSCs are self-renewing and either multi- or uni-potent. They remain quiescent to maintain their undifferentiated state. The HSC pool is divided into long-term HSCs (LT-HSCs) and short-term HSCs (ST-HSCs). LT-HSCs can provide long term engraftment and can serially engraft irradiated mice, whereas ST-HSCs have more limited self-renewal capacity in a serial transplantation assay ³. ST-HSCs are believed to give rise to multipotent progenitors (MPPs), which are capable of producing all blood cell linages, but lack the capacity for serial transplantation. These MPPs then differentiate further into either myeloid or lymphoid lineages. In the classical model of hematopoiesis, the next steps are the formation of common myeloid progenitors (CMPs), which give rise to all myeloid cells, including the megakaryo-erythroid lineage, and the common lymphoid progenitors (CLPs)³. However, this classical tree has been subject of debate. Recent evidence has suggested that possibly the first step from the HSC pool may not be a lymphoidmyeloid differentiation decision step, but rather one that allows the megakaryocyteerythrocyte lineage to split off, leaving a progenitor with both lymphoid and myeloid potential, but without the capacity to form megakaryocytes or erythrocytes. Named a lymphoid-primed multipotent progenitor (LMPP), this cell then differentiates into the granulocyte-macrophage progenitor (GMP) and CLP³. The GMP can differentiate into a neutrophil, monocyte (which in turn can differentiate into a macrophage or dendritic cell in the periphery), or an eosinophil 13 . The CLP differentiates into a B cell, a natural killer cell (NK cell) or becomes an early thymic progenitor (ETP) that migrates from the bone marrow to the thymus to become a $CD4+T$ cell or a $CD8+T$ cell¹³. Megakaryocyteerythroid progenitors (MEPs) differentiate further into erythrocytes (through stimulation with erythropoietin) or megakaryocytes (stimulated by thrombopoietin). Cells in the erythroid lineage are released into the bloodstream as reticulocytes that mature to erythrocytes in the circulation. Megakaryocytes mature fully in the bone marrow, after which they release platelets into the bloodstream and perish¹⁴.

CHOLESTEROL METABOLISM

Cholesterol is an important precursor for steroid hormones and an essential component of cell membranes, in which it modulates membrane fluidity. Cholesterol can exist in eukaryotic cells in two forms: an unbound, free from in cell membranes, or stored as cholesteryl esters. Esterification for storage is important, as excess free cholesterol is cytotoxic. See Figure 2 for a schematic overview.

Figure 1: Schematic overview of hematopoiesis. For explanation see text. LT-HSC: long-term hematopoietic stem cell. ST-HSC: short-term hematopoietic stem cell. MPP: multipotent progenitor. LMPP: lymphoidprimed multipotent progenitor. CMP: common myeloid progenitor. CLP: common lymphoid progenitor. GMP: granulocyte-macrophage progenitor. MEP: megakaryocyte-erythroid progenitor. B: B cell. T: T cell. NK: natural killer cell. GR: granulocyte. MO: monocyte. E: erythrocyte. MK: megakaryocyte.

Lipoproteins

General structure and function

Cholesterol is a hydrophobic molecule and is insoluble in aqueous solutions. Therefore, it is transported thought the blood circulation by lipoproteins. Lipoproteins are watersoluble macromolecular complexes, comprised of a hydrophobic core containing cholesteryl esters, neutral lipids, and triglycerides, and a hydrophilic monolayered shell of phospholipids, unesterified free cholesterol and specific apolipoproteins. Lipoproteins are classified according to their size and density: chylomicrons, very-low-density lipoproteins (VLDL), low-density lipoproteins (LDL), and high-density lipoproteins (HDL) 15. Chylomicrons and VLDL are the largest and least dense lipoproteins, as they consist primarily of a triglyceride rich core. LDL, a remnant of VLDL, and HDL contain a core which is comprised primarily of cholesteryl esters 15 . Moreover, the different lipoproteins can be distinguished by their apolipoprotein composition. Chylomicrons, VLDL and LDL have apolipoprotein B (ApoB) as their primary apolipoprotein, whereas apolipoprotein A1 (ApoA1) is the principle apolipoprotein of HDL.

The liver plays a central role in cholesterol metabolism, which can be divided in an exogenous, endogenous and reverse cholesterol transport pathway 6 .

Figure 2: Schematic overview of lipid metabolism. For explanation, see text.

Exogenous lipid and cholesterol metabolism

The amount of cholesterol and triglycerides absorbed from the diet is a major contributor the lipid and cholesterol levels in the circulation. Upon absorption by the enterocytes dietary triglycerides and cholesteryl esters are, together with intestinal ApoB molecules incorporated in nascent chylomicrons. These newly synthesized chylomicrons are then exocytosed into the lymph and subsequently enter the blood circulation, where they acquire Apolipoprotein E (ApoE) and apolipoprotein C (ApoC) apolipoproteins to form mature chylomicrons ^{16,17}. Mature chylomicrons circulate in the blood, where their triglycerides molecules are lipolyzed by lipoprotein lipase (LPL) and taken up for energy production and storage 18,19. The formed chylomicron remnants are then removed from the circulation through interaction of ApoE with hepatic LDL receptor related protein (LRP), remnant receptors, heparin sulphate proteoglycans, and scavenger receptor BI $(SR-BI)$ ²⁰⁻²². In line with the importance of ApoE for removal of lipoproteins and chylomicron remnants from the circulation, targeted deletion of ApoE is associated with severe hypercholesterolemia. Hypercholesterolemia in these mice already occurs on a standard chow diet which does not contain cholesterol, and is strongly aggravated on a high-cholesterol diet $23,24$.

Endogenous lipid and cholesterol metabolism

Lipids derived from hepatic uptake of lipoprotein remnants and de novo synthesized cholesterol are packaged and (re)secreted into the circulation by the liver as ApoB100 containing nascent VLDL particles 25 . In the circulation, VLDL further acquires ApoE and ApoC¹⁵. Hydrolysis of triglycerides transforms VLDL to intermediate-density lipoproteins (IDL), which are partly taken up by the liver via the LDL receptor (LDLr) and LRP 26 . Further hydrolysis of IDL particles results in the formation of smaller cholesteryl ester-rich LDL, which has ApoB100 as the sole apolipoprotein 26 . Finally, LDL is recognized by the LDLr mostly on the liver as well as peripheral tissues and removed from the circulation for intracellular metabolism. In line with its important role in the removal of VLDL remnants and its ultimate remnant LDL from the circulation, mice which are genetically deficient for the LDLr have a prolonged half-life of plasma VLDL and LDL 27 . These mice display a modestly elevated plasma cholesterol level when fed a normal chow diet. However, when placed on a cholesterol-rich diet they show strongly elevated plasma cholesterol levels (hypercholesterolemia)^{28,29}.

Reverse cholesterol transport

Reverse cholesterol transport is the pathway through which excess cholesterol from peripheral tissues is transported back to the liver by HDL for recycling or biliary excretion.

HDL metabolism

HDL is primarily produced by the liver, and to a lesser extent by the intestine, though interaction of lipid-poor ApoA1 with the ATP-binding cassette transporter A1 (ABCA1)^{30,31}. This is an essential part of HDL formation, as both genetic deficiency of ApoA1 as well as lack of ABCA1 diminishes circulating HDL levels ^{32,33}. Lipidation of ApoA1 results in the formation of nascent pre-β HDL particles 34 . These nascent HDL particles are potent acceptors of excess cholesterol from peripheral tissues. Upon association of, primarily, free cholesterol and phospholipids from peripheral tissues, discoidal HDL particles mature to spherical HDL₃ particles upon conversion of its free cholesterol into cholesteryl esters by lecithin:cholesteryl acyltransferase (LCAT)³⁵ and storage in the core of the particle. HDL₃ is subsequently converted into large α -migrating mature HDL₂ by further acquirement of phospholipids by transfer through phospholipid transfer protein (PLTP) and apolipoproteins that are released during lipolysis of triglycerides in chylomicrons or VLDL. Mature HDL circulates in the blood where it takes up excess cholesterol from tissues, excreted through cholesterol efflux transporters. The cholesteryl esters are delivered to the liver via SR-BI for recycling and biliary excretion or transferred to ApoB containing lipoproteins by cholesteryl ester transfer protein (CETP) for redistribution through the endogenous lipid and cholesterol metabolism pathway.

Cholesterol efflux towards ApoA1 and HDL

Cholesterol efflux is an active mechanism, which is regulated by various transporters, the most important of which are members of the ATP binding cassette (ABC) transporter family. Several members of this family of transmembrane proteins mediate the transport of cholesterol between various cellular domains and eventual efflux of cholesterol to ApoA1 or HDL on the plasma membrane.

ABCA1 is present on the cellular plasma membrane, and mediates cholesterol efflux to lipid poor apolipoproteins, such as ApoA1 and ApoE, but not large HDL particles 36 . ABCA1 is highly expressed in hepatocytes and enterocytes, the two cell types that together generate the majority of the plasma HDL 37 . As a consequence, ABCA1 knockout mice are unable to lipidate circulating ApoA1, resulting in HDL deficiency 32 . In line, mutations in the ABCA1 gene in humans are the causal factor in Tangier disease, of which HDL deficiency is a major clinical manifestation ³⁸⁻⁴⁰. However, ABCA1 is most known for its crucial function in lipid homeostasis and cholesterol efflux from macrophages, as the deletion of ABCA1 in bone marrow leads to a dramatic increase in foam cell formation and subsequent atherosclerosis in hypercholesterolemic mice ⁴¹.

Another important cholesterol transporter is ABCG1. ABCG1 is highly expressed in macrophages, endothelial cells, epithelial cells and brain. However, in contrast to ABCA1, not in hepatocytes or intestinal enterocytes 42 . Moreover, contrary to ABCA1, which is a full transporter, ABCG1 needs to dimerize with another ABC transporter to become functional 43. ABCG1 mediates in cholesterol efflux to mature HDL particles, but not lipid-free apolipoproteins^{42,44,45}. In line, ABCG1 does not facilitate HDL generation⁴². Interestingly, whereas ABCA1 is present on the plasma membrane and directly binds to circulating ApoA1⁴⁶, ABCG1 is involved in intracellular sterol movement. Although the precise cellular localization of ABCG1 remains elusive, it is involved in trafficking of cholesterol over the endoplasmic reticulum membrane and endosomes. In that way it does not only increase availability of cholesterol for efflux to lipoprotein acceptors, but is also important for the regulation of endoplasmic reticulum-localized cholesterol biosynthesis genes 46,47.

Besides ABCA1 and ABCG1, ABCG4 has also been implicated as a cholesterol efflux transporter ⁴⁴. ABCG4 shows high levels of homology to ABCG1, and effluxes to HDL particles. Interestingly, it is not expressed in macrophages, but is highly expressed in megakaryocytes, the precursor cells of platelets⁴⁸.

Besides ABC transporter family members, SR-BI has also been implicated as a cholesterol efflux transporter, as it induces free cholesterol flux down a cholesterol-concentration gradient ^{49,50}. The exact mechanism by which SR-BI mediates cholesterol efflux is largely unknown. Possibly, SR-BI simply mediates tethering of acceptor HDL particles closely to the cell surface 51 . Another possibility is that SR-BI reorganizes the lipids in cholesterol-rich domains in the plasma membrane, thereby facilitating the movement of cholesterol towards HDL 51.

ATHEROSCLEROSIS

Atherosclerosis is a chronic, multifactorial disease induced by lipid accumulation in the vessel wall and subsequent arterial wall inflammation, leading to atherosclerotic plaque formation. Atherosclerosis develops asymptomatic over a lifetime. However, when a plaque ruptures or erodes, a superimposed thrombus is formed causing acute cardiovascular events such as myocardial infarction or ischemic stroke.

Dyslipidemia is a major risk factor for atherosclerosis

A major risk factor for the development of premature atherosclerosis is dyslipidemia. Dyslipidemia is a broad term, which refers to a number of lipid disorders, including elevated LDL cholesterol, low HDL cholesterol, excess lipoprotein(a), hypertriglyceridemia, and mixed lipid disorders. Although familial causes account for ~20% of these disorders, most (~80%) are related to diet and lifestyle 52 . It is well established that there is a direct relationship between dyslipidemia and cardiovascular risk 53 . In healthy individuals, cholesterol levels are below 5 mmol/L. A rise of 2 mmol/L cholesterol increases the risk of death by cardiovascular disease by 50% 54. This is most likely attributable to high LDL cholesterol, the main carrier of cholesterol in human plasma. In contrast, large population studies have consistently shown that low HDL cholesterol, as well as ApoA1 levels are independent, inverse predictors of cardiovascular disease risk ⁵⁵⁻⁵⁹. Normalization of dyslipidemia is therefore a major target in the prevention of atherosclerosis and its clinical manifestations.

Pathophysiology of atherosclerosis

Atherosclerotic plaques form at specific arterial sites with differences in blood flow dynamics, such as arterial tree branches and bifurcations. Low shear stress, oscillatory or turbulent flow, together with dyslipidemia, which can cause lipoprotein retention in the subendothelial matrix, cause damage and endothelial dysfunction. This is generally regarded as the primary step in atherosclerosis development. Damage to the endothelium increases arterial wall permeability and increases expression of adhesion molecules, and the production of cytokines and chemokines, thereby leading to the migration of inflammatory cells such as monocytes, lymphocytes and neutrophils from the circulation into the arterial wall intima. Macrophages in the intimal wall then rapidly

engulf the accumulated lipids and the first foam cells in the nascent lesion are formed. Macrophage foam cell formation is the hallmark of early atherosclerotic lesions, also called fatty streaks. These fatty streaks can then progress towards intermediate fibrofatty lesions, which consist of several layers of foam cells and vascular smooth muscle cells with T cells, surrounded by a relatively poor developed matrix of connective tissue. These intermediate plaques can then progress further into advanced fibrous lesions, which are characterized by a fibrous cap that covers a core of extracellular lipid and necrotic material, together with macrophages, smooth muscle cells and T cells $^{\text{1}}$. In line with the importance of immune cell influx and inflammation in atherosclerosis development, studies in mice have indicated a causal relationship between the levels of blood monocytes and neutrophils and the extent of atherosclerosis $60-62$. Moreover, the levels of blood monocytes and neutrophils are predictive of acute cardiovascular events ^{4,5}. Importantly, monocytosis in the absence of hyperlipidemia is insufficient to induce atherosclerosis ^{11,63}. Only when the endothelium becomes inflamed in response to hypercholesterolemia, monocytosis and neutrophilia contribute to lesion development 11,64-66

Atherothrombosis

Vulnerable plaques

Atherosclerotic plaques develop slowly, and remain largely asymptomatic as long as they remain intact. However, advanced plaques can cause complications through atherothrombosis formation. Atherothrombosis can occur in two ways: by rupture of an unstable lesion, or by erosion of the overlying endothelium. Plaques prone to rupture, so-called vulnerable plaques, are characterized by a thin fibrous cap, and a large lipid core with a relative abundance of inflammatory leukocytes 57 . In contrast, vulnerable plaques underlying areas of superficial erosion do not have thin fibrous caps. Moreover, they harbor fewer inflammatory cells, and lack large lipid pools⁶⁸. From autopsy studies done several decades ago, it was suggested that plaque rupture most commonly led to fatal coronary atherothrombosis $67,69$, whereas a minority of fatal events was caused by superficial plaque erosion. However, more recently, questions have been raised about the dominant mechanisms implicated in atherothrombosis. Recent evidence suggests that plaques with thin fibrous caps and large lipid pools seldomly rupture and cause clinical events $70,71$. Moreover, recent studies have shown that there is a shift towards plagues with significantly more fibrous, non-inflammatory characteristics ⁷²⁻⁷⁴. Possibly, this shift in plaque characteristics could lead to a subsequent shift in plaque rupture versus erosion occurrence 75,76.

Pathophysiology of atherothrombosis

When a plaque ruptures or erodes, subendothelial collagen and thrombogenic plaque material, such as macrophage tissue factor, are exposed to the arterial circulation. This leads to thrombus formation on top of the ruptured or eroded plaque². Upon the breach of the arterial endothelial lining, platelets are the first responders. Within one minute after rupture, platelets adhere and aggregate on collagenous plaque components. In line with their importance in atherothrombotic development, platelet characteristics such as density and volume are associated with the risk of acute coronary syndromes $6-9$, and implies mean platelet volume, a determinant of platelet reactivity, as both a causal and prognostic factor 10 . Following the platelet response, approximately three minutes after breaching of the endothelial lining, the thrombus is characterized by thrombin and fibrin formation, and by tissue factor-triggered coagulation cascade activation 77 . This thrombus formation can lead to rapid occlusion of the vessel, a cause myocardial infarction, ischemic stroke and sudden death. This deadly nature of atherothrombosis has made it a critical target for investigation.

Interplay between cholesterol and bone marrow cells in atherosclerosis

Atherosclerosis is generally characterized as an inflammatory response to hypercholesterolemia. However, there is increasing evidence that the inflammatory process driving the atherogenic response to hypercholesterolemia may be initiated in the bone marrow. A large body of evidence indicates that bone marrow functioning and hematopoiesis are affected by changes in HSC cholesterol homeostasis, i.e. hypercholesterolemia and cholesterol efflux.

Hypercholesterolemia and HSCs

Several studies on the effects of hypercholesterolemia on bone marrow function have been done in high-fat, high-cholesterol (HFHC) diet -fed LDLr and ApoE knockout mice. Both HFHC diet-fed ApoE knockout, and to a lesser extent HFHC diet-fed LDLr knockout mice display and augmented expansion and proliferation of HSCs 64,78,79 . This phenotype is paralleled by an increase in myeloid cell production and increased circulating counts of monocytes and neutrophils ^{64,78,79}.

Interestingly, hypercholesterolemia-induced hyperproliferation of HSCs, as well as the consequent leukocytosis, are also evident in recipients of transplanted bone marrow from hypercholesterolemic mice $79,80$. Thus, long-term intrinsic changes in stem cells are induced by the hypercholesterolemia.

Besides HSC proliferation and leukocytosis, hypercholesterolemia also alters platelet production and reactivity. Rabbits and guinea pigs fed a high-cholesterol diet develop megakaryocytes with increased ploidy and size $81,82$. Hypercholesterolemia in humans is also associated with higher megakaryocyte ploidy, indicative of altered megakaryocyte

maturation ⁸³. Notably, the change in megakaryocyte maturation in hypercholesterolemic human subjects coincided with a higher mean platelet volume ⁸³.

Cholesterol efflux and HDL and HSCs

Most of the atherosclerosis-protective properties of APOA1 and HDL are attributed to their role in cholesterol efflux and subsequent reverse cholesterol transport from macrophage foam cells, as well as direct anti-inflammatory properties ^{84–86}. HSCs utilize cholesterol efflux pathways to oppose the excessive production of myeloid cells ^{11,64,78,87}. Genetic deletion of ABCA1 and ABCG1 leads to dramatic expansion and proliferation of HSCs and an increase in committed myeloid progenitor populations ¹¹. This stimulates the development of severe monocytosis and neutrophilia and infiltration of multiple organs with myeloid cells and accumulation of macrophage foam cells 11. A similar phenotype is observed when ABCA1/ABCG1 double knockout bone marrow is transplanted into LDLr homozygous or heterozygous knockout mice $11,88$. Interestingly, increased HSC numbers and the myeloproliferative phenotype caused by combined deletion of ABCA1/ABCG1 in bone marrow could be rescued by transgenic overexpression of ApoA1 in recipient mice ¹¹. Infusion of ApoA1 into LDLr/ApoA1 double knockout mice also reduces the number of LSK cells in bone marrow⁸⁷, and reduces HSC proliferation in hypercholesterolemic LDLr knockout mice, and ApoE knockout mice ^{64,78}.

Transplantation of ABCA1 knockout bone marrow into LDLr knockout mice is associated with lymphocytosis, although no such phenotype is found in total body ABCA1 knockout mice 41. Similarly, chow-fed ApoA1 knockout mice do not display monocytosis or proliferation of HSCs 64 . As both ABCA1 and ApoA1 knockout mice are hypocholesterolemic through the inability to produce HDL, this indicates that the importance of cholesterol efflux may depend on the levels of circulating cholesterol levels. Genetic deletion of only ABCG1 has thus far not been shown to affect bone marrow hematopoiesis, neither as a total body knockout nor after transplantation to hypercholesterolemic recipient mice.

ABCG4 is specifically expressed in MEP cell populations and megakaryocyte progenitors. In line, ABCG4 knockout megakaryocyte progenitors show defective cholesterol efflux to HDL and increased MEP and megakaryocyte progenitor cell populations, which are relatively more responsive to thrombopoietin. As a consequence, ABCG4 knockout transplanted LDLr knockout mice have increased platelet counts⁴⁸.

OUTLINE OF THE THESIS

Defects in cholesterol metabolism and hypercholesterolemia have been shown to affect hematopoiesis, immune cell production and platelet counts and reactivity. Therefore, bone marrow cholesterol handling is an interesting target in the battle against cardiovascular diseases, and acute cardiovascular events in particular.

This thesis will describe novel interactions between cholesterol metabolism and the production immune cells and platelets, and its effects on atherosclerosis and atherothrombosis development.

The first part of this thesis focusses on cholesterol metabolism in hematopoiesis.

It is well established that dyslipidemia is an important risk factor for cardiovascular disease. Circulating lipid levels are highly correlated with atherosclerotic plaque development. In addition, experimental evidence suggests that lipids also directly influence thrombosis and influence the risk and the outcome of acute cardiovascular events. In **chapter 2**, the effects of lipoproteins on three important components of atherothrombosis pathophysiology, i.e. endothelial function, platelets and secondary coagulation, are reviewed.

A substantial body of clinical evidence indicates that platelet characteristics such as density and volume are associated with increased risk of acute coronary syndromes $6-9$, and implies mean platelet volume, a determinant of platelet reactivity, as both a causal and prognostic factor 10 . Therefore, modification of platelet functionality is a promising therapeutic target in the prevention and treatment of cardiovascular disease. In **chapter 3**, we show that elevated plasma unesterified cholesterol levels impairs megakaryopoiesis and platelet production in SR-BI knockout mice. Our findings suggest that impaired platelet production, in addition to the previously reported augmented platelet clearance, may explain part of the thrombocytopenic phenotype associated with SR-BI deficiency. In **chapter 4**, we describe novel and opposing roles of the cholesterol efflux transporters ABCA1 and ABCG1 in megakaryopoiesis, platelet production and effects on platelet characteristics.

In addition to effects on hematopoietic stem cell functionality and hematopoiesis, ApoA1 and HDL also directly influence inflammatory cell responses ⁸⁶. In **chapter 5**, we dissociated the direct effects of ApoA1 deficiency on immune cells in the context of atherosclerosis from effects that are generated through changes in bone marrow immune cell production.

The second part of this thesis focusses on the development of experimental animal models of atherothrombosis.

Although atherosclerosis is widely studied in experimental mouse models, experimental studies regarding atherothrombosis are hampered due to lack of suitable mouse models. In **chapter 6** recent advances in the development of mouse models for spontaneous atherothrombosis are reviewed. In **chapter 7**, we describe a novel RNA interference-based mouse model for spontaneous atherothrombosis in ApoE knockout mice. Further characterization and attempts to optimize this model through administration of phenylephrine to ApoE knockout mice are described in **chapter 8**. Murine SR-BI deficiency is associated with increased platelet reactivity and a higher susceptibility for FeCl₃ damage-induced arterial thrombosis in the carotid artery ¹². In **chapter 9**, we apply the developed atherothrombosis model to hypercholesterolemic SR-BI knockout mice.

Finally, all the results obtained in this thesis and future perspectives are summarized and discussed in **chapter 10**.

REFERENCES

- 1. Ross R. Atherosclerosis An inflammatory disease. *N Engl J Med*. 1999;340(2):115-126. doi:10.1056/nejm199901143400207.
- 2. Furie B, Furie BC. Mechanisms of thrombus formation. *N Engl J Med*. 2008;359(9):938- 949. doi:10.1056/NEJMra0801082.
- 3. Orkin SH, Zon LI. Hematopoiesis: An Evolving Paradigm for Stem Cell Biology. *Cell*. 2008;132(4):631-644. doi:10.1016/j. cell.2008.01.025.
- 4. Lee CD, Folsom AR, Nieto FJ, Chambless LE, Shahar E, Wolfe DA. White blood cell count and incidence of coronary heart disease and ischemic stroke and mortality from cardiovascular disease in African-American and White men and women: atherosclerosis risk in communities study. *Am J Epidemiol*. 2001;154(8):758-764.
- 5. Sweetnam PM, Thomas HF, Yarnell JW, Baker IA, Elwood PC. Total and differential leukocyte counts as predictors of ischemic heart disease: the Caerphilly and Speedwell studies. *Am J Epidemiol*. 1997;145(5):416-421.
- 6. Martin JF, Plumb J, Kilbey RS, Kishk YT. Changes in volume and density of platelets in myocardial infarction. *Br Med J (Clin Res Ed)*. 1983;287(6390):456-459. doi:10.1136/ bmj.287.6390.456.
- 7. Cameron HA, Phillips R, Ibbotson RM, Carson PH. Platelet size in myocardial infarction. *Br Med J (Clin Res Ed)*. 1983;287(6390):449-451. doi:10.1136/ bmj.287.6390.449.
- 8. Pizzulli L, Yang A, Martin JF, Lüderitz B. Changes in platelet size and count in unstable angina compared to stable angina or non-cardiac chest pain. *Eur Heart J*. 1998;19(1):80-84. doi:10.1053/ euhj.1997.0747.
- 9. Endler G, Klimesch A, Sunder-Plassmann H, Schillinger M, Exner M, Mannhalter C, Jordanova N, Christ G, Thalhammer R, Huber K, Sunder-Plassmann R. Mean

platelet volume is an independent risk factor for myocardial infarction but not for coronary artery disease. *Br J Haematol*. 2002;117(2):399-404.

- 10. Martin J, Bath P, Burr ML. Influence of Platelet Size on Outcome after Myocardial Infarction. *Lancet*. 1991;338(8780):1409- 1411. doi:0140-6736(91)92719-I [pii].
- 11. Yvan-Charvet L, Pagler T, Gautier EL, Avagyan S, Siry RL, Han S, Welch CL, Wang N, Randolph GJ, Snoeck HW, Tall AR. ATPbinding cassette transporters and HDL suppress hematopoietic stem cell proliferation. *Science*. 2010;328(5986):1689-1693. doi:10.1126/science.1189731.
- 12. Korporaal SJA, Meurs I, Hauer AD, Hildebrand RB, Hoekstra M, Cate HT, Pratico D, Akkerman J-WN, Van Berkel TJC, Kuiper J, Van Eck M. Deletion of the High-Density Lipoprotein Receptor Scavenger Receptor BI in Mice Modulates Thrombosis Susceptibility and Indirectly Affects Platelet Function by Elevation of Plasma Free Cholesterol. *Arterioscler Thromb Vasc Biol*. 2011;31(1):34-42. doi:10.1161/ATVBAHA.110.210252.
- 13. Nakajima H. Role of transcription factors in differentiation and reprogramming of hematopoietic cells. *Keio J Med*. 2011;60(2):47-55. doi:10.2302/kjm.60.47.
- 14. Machlus KR, Italiano JE. The incredible journey: From megakaryocyte development to platelet formation. *J Cell Biol*. 2013;201(6):785-796. doi:10.1083/ jcb.201304054.
- 15. Ginsberg HN. Lipoprotein physiology. *Endocrinol Metab Clin North Am*. 1998;27(3):503-519.
- 16. Mansbach CM 2nd, Gorelick F. Development and physiological regulation of intestinal lipid absorption. II. Dietary lipid absorption, complex lipid synthesis, and the intracellular packaging and secretion of chylomicrons. *Am J Physiol Gastroin-*

test Liver Physiol. 2007;293(4):G645-50. doi:10.1152/ajpgi.00299.2007.

- 17. Hussain MM. A proposed model for the assembly of chylomicrons. *Atherosclerosis*. 2000;148(1):1-15.
- 18. Goldberg IJ. Lipoprotein lipase and lipolysis: central roles in lipoprotein metabolism and atherogenesis. *J Lipid Res*. 1996;37(4):693-707.
- 19. Zechner R. The tissue-specific expression of lipoprotein lipase: implications for energy and lipoprotein metabolism. *Curr Opin Lipidol*. 1997;8(2):77-88.
- 20. Cooper AD. Hepatic uptake of chylomicron remnants. *J Lipid Res*. 1997;38(11):2173- 2192.
- 21. Ji ZS, Fazio S, Lee YL, Mahley RW. Secretioncapture role for apolipoprotein E in remnant lipoprotein metabolism involving cell surface heparan sulfate proteoglycans. *J Biol Chem*. 1994;269(4):2764-2772.
- 22. Out R, Kruijt JK, Rensen PCN, Hildebrand RB, de Vos P, Van Eck M, Van Berkel TJC. Scavenger receptor BI plays a role in facilitating chylomicron metabolism. *J Biol Chem*. 2004;279(18):18401-18406. doi:10.1074/jbc.M401170200.
- 23. Zhang SH, Reddick RL, Piedrahita J a, Maeda N. Spontaneous hypercholesterolemia and arterial lesions in mice lacking apolipoprotein E. *Science*. 1992;258(5081):468- 471. http://www.ncbi.nlm.nih.gov/ pubmed/1411543.
- 24. Plump a S, Smith JD, Hayek T, Aalto-Setälä K, Walsh a, Verstuyft JG, Rubin EM, Breslow JL. Severe hypercholesterolemia and atherosclerosis in apolipoprotein E-deficient mice created by homologous recombination in ES cells. *Cell*. 1992;71(2):343-353. doi:10.1016/0092-8674(92)90362-G.
- 25. Olofsson SO, Stillemark-Billton P, Asp L. Intracellular assembly of VLDL: two major steps in separate cell compartments. *Trends Cardiovasc Med*. 2000;10(8):338-345.
- 26. Brown MS, Kovanen PT, Goldstein JL. Regulation of plasma cholesterol by lipoprotein

receptors. *Science*. 1981;212(4495):628- 635.

- 27. Kowala MC, Recce R, Beyer S, Gu C, Valentine M. Characterization of atherosclerosis in LDL receptor knockout mice: macrophage accumulation correlates with rapid and sustained expression of aortic MCP-1/ JE. *Atherosclerosis*. 2000;149(2):323-330.
- 28. Ishibashi S, Brown MS, Goldstein JL, Gerard RD, Hammer RE, Herz J. Hypercholesterolemia in low density lipoprotein receptor knockout mice and its reversal by adenovirus-mediated gene delivery. *J Clin Invest*. 1993;92(2):883-893. doi:10.1172/ JCI116663.
- 29. Knowles JW, Maeda N. Genetic modifiers of atherosclerosis in mice. *Arterioscler Thromb Vasc Biol*. 2000;20(11):2336-2345.
- 30. Brunham LR, Kruit JK, Iqbal J, Fievet C, Timmins JM, Pape TD, Coburn BA, Bissada N, Staels B, Groen AK, Hussain MM, Parks JS, Kuipers F, Hayden MR. Intestinal ABCA1 directly contributes to HDL biogenesis in vivo. *J Clin Invest*. 2006;116(4):1052-1062. doi:10.1172/JCI27352.
- 31. Timmins JM, Lee J-Y, Boudyguina E, Kluckman KD, Brunham LR, Mulya A, Gebre AK, Coutinho JM, Colvin PL, Smith TL, Hayden MR, Maeda N, Parks JS. Targeted inactivation of hepatic Abca1 causes profound hypoalphalipoproteinemia and kidney hypercatabolism of apoA-I. *J Clin Invest*. 2005;115(5):1333-1342. doi:10.1172/ JCI23915.
- 32. McNeish J, Aiello RJ, Guyot D, Turi T, Gabel C, Aldinger C, Hoppe KL, Roach ML, Royer LJ, de Wet J, Broccardo C, Chimini G, Francone OL. High density lipoprotein deficiency and foam cell accumulation in mice with targeted disruption of ATP-binding cassette transporter-1. *Proc Natl Acad Sci U S A*. 2000;97(8):4245-4250.
- 33. Plump AS, Azrolan N, Odaka H, Wu L, Jiang X, Tall A, Eisenberg S, Breslow JL. ApoA-I knockout mice: characterization of HDL metabolism in homozygotes and identifi-

cation of a post-RNA mechanism of apoA-I up-regulation in heterozygotes. *J Lipid Res*. 1997;38(5):1033-1047.

- 34. Schmitz G, Langmann T. Structure, function and regulation of the ABC1 gene product. *Curr Opin Lipidol*. 2001;12(2):129-140.
- 35. Kostner GM, Knipping G, Groener JE, Zechner R, Dieplinger H. The role of LCAT and cholesteryl ester transfer proteins for the HDL and LDL structure and metabolism. *Adv Exp Med Biol*. 1987;210:79-86.
- 36. Wang N, Silver DL, Costet P, Tall AR. Specific binding of ApoA-I, enhanced cholesterol efflux, and altered plasma membrane morphology in cells expressing ABC1. *J Biol Chem*. 2000;275(42):33053-33058. doi:10.1074/jbc.M005438200.
- 37. Wellington CL, Walker EKY, Suarez A, Kwok A, Bissada N, Singaraja R, Yang Y-Z, Zhang L-H, James E, Wilson JE, Francone O, McManus BM, Hayden MR. ABCA1 mRNA and protein distribution patterns predict multiple different roles and levels of regulation. *Lab Invest*. 2002;82(3):273-283.
- 38. Brooks-Wilson A, Marcil M, Clee SM, Zhang LH, Roomp K, van Dam M, Yu L, Brewer C, Collins JA, Molhuizen HO, Loubser O, Ouelette BF, Fichter K, Ashbourne-Excoffon KJ, Sensen CW, Scherer S, Mott S, Denis M, Martindale D, Frohlich J, Morgan K, Koop B, Pimstone S, Kastelein JJ, Genest JJ, Hayden MR. Mutations in ABC1 in Tangier disease and familial high-density lipoprotein deficiency. *Nat Genet*. 1999;22(4):336-345. doi:10.1038/11905.
- 39. Bodzioch M, Orso E, Klucken J, Langmann T, Bottcher A, Diederich W, Drobnik W, Barlage S, Buchler C, Porsch-Ozcurumez M, Kaminski WE, Hahmann HW, Oette K, Rothe G, Aslanidis C, Lackner KJ, Schmitz G. The gene encoding ATP-binding cassette transporter 1 is mutated in Tangier disease. *Nat Genet*. 1999;22(4):347-351. doi:10.1038/11914.
- 40. Rust S, Rosier M, Funke H, Real J, Amoura Z, Piette JC, Deleuze JF, Brewer HB, Du-

verger N, Denefle P, Assmann G. Tangier disease is caused by mutations in the gene encoding ATP-binding cassette transporter 1. *Nat Genet*. 1999;22(4):352-355. doi:10.1038/11921.

- 41. van Eck M, Bos IST, Kaminski WE, Orso E, Rothe G, Twisk J, Bottcher A, Van Amersfoort ES, Christiansen-Weber TA, Fung-Leung W-P, Van Berkel TJC, Schmitz G. Leukocyte ABCA1 controls susceptibility to atherosclerosis and macrophage recruitment into tissues. *Proc Natl Acad Sci U S A*. 2002;99(9):6298-6303. doi:10.1073/ pnas.092327399.
- 42. Kennedy MA, Barrera GC, Nakamura K, Baldan A, Tarr P, Fishbein MC, Frank J, Francone OL, Edwards PA. ABCG1 has a critical role in mediating cholesterol efflux to HDL and preventing cellular lipid accumulation. *Cell Metab*. 2005;1(2):121-131. doi:10.1016/j.cmet.2005.01.002.
- 43. Dean M, Hamon Y, Chimini G. The human ATP-binding cassette (ABC) transporter superfamily. *J Lipid Res*. 2001;42(7):1007- 1017.
- 44. Wang N, Lan D, Chen W, Matsuura F, Tall AR. ATP-binding cassette transporters G1 and G4 mediate cellular cholesterol efflux to high-density lipoproteins. *Proc Natl Acad Sci U S A*. 2004;101(26):9774-9779. doi:10.1073/pnas.0403506101.
- 45. Nakamura K, Kennedy MA, Baldan A, Bojanic DD, Lyons K, Edwards PA. Expression and regulation of multiple murine ATP-binding cassette transporter G1 mRNAs/isoforms that stimulate cellular cholesterol efflux to high density lipoprotein. *J Biol Chem*. 2004;279(44):45980-45989. doi:10.1074/ jbc.M408652200.
- 46. Wang N, Ranalletta M, Matsuura F, Peng F, Tall AR. LXR-induced redistribution of ABCG1 to plasma membrane in macrophages enhances cholesterol mass efflux to HDL. *Arterioscler Thromb Vasc Biol*. 2006;26(6):1310-1316. doi:10.1161/01. ATV.0000218998.75963.02.
- 47. Tarling EJ, Edwards PA. ATP binding cassette transporter G1 (ABCG1) is an intracellular sterol transporter. *Proc Natl Acad Sci U S A*. 2011;108(49):19719-19724. doi:10.1073/pnas.1113021108.
- 48. Murphy AJ, Bijl N, Yvan-Charvet L, Welch CB, Bhagwat N, Reheman A, Wang Y, Shaw JA, Levine RL, Ni H, Tall AR, Wang N. Cholesterol efflux in megakaryocyte progenitors suppresses platelet production and thrombocytosis. *Nat Med*. 2013;19(5):586-594. doi:10.1038/nm.3150.
- 49. Ji Y, Jian B, Wang N, Sun Y, Moya ML, Phillips MC, Rothblat GH, Swaney JB, Tall AR. Scavenger receptor BI promotes high density lipoprotein-mediated cellular cholesterol efflux. *J Biol Chem*. 1997;272(34):20982- 20985.
- 50. Jian B, de la Llera-Moya M, Ji Y, Wang N, Phillips MC, Swaney JB, Tall AR, Rothblat GH. Scavenger receptor class B type I as a mediator of cellular cholesterol efflux to lipoproteins and phospholipid acceptors. *J Biol Chem*. 1998;273(10):5599-5606.
- 51. de la Llera-Moya M, Rothblat GH, Connelly MA, Kellner-Weibel G, Sakr SW, Phillips MC, Williams DL. Scavenger receptor BI (SR-BI) mediates free cholesterol flux independently of HDL tethering to the cell surface. *J Lipid Res*. 1999;40(3):575-580.
- 52. Fiorucci S, Cipriani S, Baldelli F, Mencarelli A. Bile acid-activated receptors in the treatment of dyslipidemia and related disorders. *Prog Lipid Res*. 2010;49(2):171-185. doi:10.1016/j.plipres.2009.11.001.
- 53. Lusis AJ. Atherosclerosis. *Nature*. 2000;407(6801):233-241. doi:10.1038/35025203.
- 54. Libby P, Aikawa M. Stabilization of atherosclerotic plaques: new mechanisms and clinical targets. *Nat Med*. 2002;8(11):1257- 1262. doi:10.1038/nm1102-1257.
- 55. Gordon DJ, Knoke J, Probstfield JL, Superko R, Tyroler HA. High-density lipoprotein cholesterol and coronary heart disease in hypercholesterolemic men: the Lipid Re-

search Clinics Coronary Primary Prevention Trial. *Circulation*. 1986;74(6):1217-1225.

- 56. Miller NE, Thelle DS, Forde OH, Mjos OD. The Tromso heart-study. High-density lipoprotein and coronary heart-disease: a prospective case-control study. *Lancet (London, England)*. 1977;1(8019):965-968.
- 57. Gordon T, Castelli WP, Hjortland MC, Kannel WB, Dawber TR. High density lipoprotein as a protective factor against coronary heart disease. The Framingham Study. *Am J Med*. 1977;62(5):707-714.
- 58. Miller M, Seidler A, Kwiterovich PO, Pearson TA. Long-term predictors of subsequent cardiovascular events with coronary artery disease and "desirable" levels of plasma total cholesterol. *Circulation*. 1992;86(4):1165-1170.
- 59. Di Angelantonio E, Sarwar N, Perry P, Kaptoge S, Ray KK, Thompson A, Wood AM, Lewington S, Sattar N, Packard CJ, Collins R, Thompson SG, Danesh J. Major lipids, apolipoproteins, and risk of vascular disease. *JAMA*. 2009;302(18):1993-2000. doi:10.1001/jama.2009.1619.
- 60. Rajavashisth T, Qiao JH, Tripathi S, Tripathi J, Mishra N, Hua M, Wang XP, Loussararian A, Clinton S, Libby P, Lusis A. Heterozygous osteopetrotic (op) mutation reduces atherosclerosis in LDL receptor- deficient mice. *J Clin Invest*. 1998;101(12):2702-2710. doi:10.1172/JCI119891.
- 61. Qiao JH, Tripathi J, Mishra NK, Cai Y, Tripathi S, Wang XP, Imes S, Fishbein MC, Clinton SK, Libby P, Lusis AJ, Rajavashisth TB. Role of macrophage colony-stimulating factor in atherosclerosis: studies of osteopetrotic mice. *Am J Pathol*. 1997;150(5):1687-1699.
- 62. Smith JD, Trogan E, Ginsberg M, Grigaux C, Tian J, Miyata M. Decreased atherosclerosis in mice deficient in both macrophage colony-stimulating factor (op) and apolipoprotein E. *Proc Natl Acad Sci U S A*. 1995;92(18):8264-8268.
- 63. Out R, Hoekstra M, Meurs I, de Vos P, Kuiper J, Van Eck M, Van Berkel TJC. Total

body ABCG1 expression protects against early atherosclerotic lesion development in mice. *Arterioscler Thromb Vasc Biol*. 2007;27(3):594-599. doi:10.1161/01. ATV.0000257136.24308.0c.

- 64. Murphy AJ, Akhtari M, Tolani S, Pagler T, Bijl N, Kuo C-L, Wang M, Sanson M, Abramowicz S, Welch C, Bochem AE, Kuivenhoven JA, Yvan-Charvet L, Tall AR. ApoE regulates hematopoietic stem cell proliferation, monocytosis, and monocyte accumulation in atherosclerotic lesions in mice. *J Clin Invest*. 2011;121(10):4138-4149. doi:10.1172/ JCI57559.
- 65. Swirski FK, Libby P, Aikawa E, Alcaide P, Luscinskas FW, Weissleder R, Pittet MJ. Ly-6Chi monocytes dominate hypercholesterolemia-associated monocytosis and give rise to macrophages in atheromata. *J Clin Invest*. 2007;117(1):195-205. doi:10.1172/ JCI29950.
- 66. Drechsler M, Megens RTA, van Zandvoort M, Weber C, Soehnlein O. Hyperlipidemia-triggered neutrophilia promotes early atherosclerosis. *Circulation*. 2010;122(18):1837-1845. doi:10.1161/ CIRCULATIONAHA.110.961714.
- 67. Davies MJ. Stability and Instability: Two Faces of Coronary Atherosclerosis The Paul Dudley White Lecture 1995. *Circulation*. 1996;94:2013-2020. http://circ.ahajournals.org/content/94/8/2013.long.
- 68. Libby P, Theroux P. Pathophysiology of coronary artery disease. *Circulation*. 2005;111(25):3481-3488. doi:10.1161/ CIRCULATIONAHA.105.537878.
- 69. Falk E, Shah PK, Fuster V. Coronary Plaque Disruption. *Circulation*. 1995;92:657- 671. http://circ.ahajournals.org/content/92/3/657.long.
- 70. Buffon A, Biasucci LM, Liuzzo G, D'Onofrio G, Crea F, Maseri A. Widespread coronary inflammation in unstable angina. *N Engl J Med*. 2002;346(24):1845-1853.
- 71. Crea F, Liuzzo G. Pathogenesis of Acute Coronary Syndromes. *J Am Coll*

Cardiol. 2013;61(1):1-11. doi:10.1016/j. jacc.2012.07.064.

- 72. Libby P, Pasterkamp G. Requiem for the 'vulnerable plaque.' *Eur Heart J*. 2015:ehv349. doi:10.1093/eurheartj/ehv349.
- 73. Underhill HR, Yuan C, Zhao X-Q, Kraiss LW, Parker DL, Saam T, Chu B, Takaya N, Liu F, Polissar NL, Neradilek B, Raichlen JS, Cain V a., Waterton JC, Hamar W, Hatsukami TS. Effect of rosuvastatin therapy on carotid plaque morphology and composition in moderately hypercholesterolemic patients: A high-resolution magnetic resonance imaging trial. *Am Heart J*. 2008;155(3):584. e1-584.e8. doi:10.1016/j.ahj.2007.11.018.
- 74. Libby P. How does lipid lowering prevent coronary events? New insights from human imaging trials. *Eur Heart J*. 2015;36(8):472- 474. doi:10.1093/eurheartj/ehu510.
- 75. Hu S, Jia H, Vergallo R, Abtahian F, Tian J, Soeda T, Rosenfield K, Jang I-K. Plaque Erosion : In Vivo Diagnosis and Treatment Guided by Optical Coherence Tomography. *JACC Cardiovasc Interv*. 2014;7(6):e63-e64. doi:10.1016/j.jcin.2013.10.024.
- 76. Braunwald E. Coronary Plaque Erosion : Recognition and Management. *JACC Cardiovasc Imaging*. 2013;6(3):288-289. doi:10.1016/j.jcmg.2013.01.003.
- 77. Lippi G, Franchini M, Targher G. Arterial thrombus formation in cardiovascular disease. *Nat Rev Cardiol*. 2011;8(9):502-512. doi:10.1038/nrcardio.2011.91.
- 78. Feng Y, Schouteden S, Geenens R, Van Duppen V, Herijgers P, Holvoet P, Van Veldhoven PP, Verfaillie CM. Hematopoietic stem/progenitor cell proliferation and differentiation is differentially regulated by high-density and low-density lipoproteins in mice. *PLoS One*. 2012;7(11):e47286. doi:10.1371/journal.pone.0047286.
- 79. Seijkens T, Hoeksema MA, Beckers L, Smeets E, Meiler S, Levels J, Tjwa M, de Winther MPJ, Lutgens E. Hypercholesterolemia-induced priming of hematopoietic stem and progenitor cells aggravates ath-

erosclerosis. *FASEB J Off Publ Fed Am Soc Exp Biol*. 2014;28(5):2202-2213. doi:10.1096/ fj.13-243105.

- 80. van Kampen E, Jaminon A, van Berkel TJC, Van Eck M. Diet-induced (epigenetic) changes in bone marrow augment atherosclerosis. *J Leukoc Biol*. 2014;96(5):833-841. doi:10.1189/jlb.1A0114-017R.
- 81. Martin JF, Slater DN, Kishk YT, Trowbridge EA. Platelet and megakaryocyte changes in cholesterol-induced experimental atherosclerosis. *Arteriosclerosis*. 1985;5(6):604- 612.
- 82. Schick BP, Schick PK. The effect of hypercholesterolemia on guinea pig platelets, erythrocytes and megakaryocytes. *Biochim Biophys Acta*. 1985;833(2):291-302. http:// www.ncbi.nlm.nih.gov/pubmed/3970955.
- 83. Pathansali R, Smith N, Bath P. Altered megakaryocyte–platelet haemostatic axis in hypercholesterolaemia. *Platelets*. 2001;12(5):292-297. doi:10.1080/09537100120058810.
- 84. Zhang Y, Zanotti I, Reilly MP, Glick JM, Rothblat GH, Rader DJ. Overexpression of apolipoprotein A-I promotes reverse transport of cholesterol from macrophages to feces in vivo. *Circulation*. 2003;108(6):661-663. doi:10.1161/01.CIR.0000086981.09834.E0.
- 85. Rader DJ. Molecular regulation of HDL metabolism and function: implications for novel therapies. *J Clin Invest*. 2006;116(12):3090-3100. doi:10.1172/ JCI30163.
- 86. Navab M, Berliner JA, Subbanagounder G, Hama S, Lusis AJ, Castellani LW, Reddy S, Shih D, Shi W, Watson AD, Van Lenten BJ, Vora D, Fogelman AM. HDL and the inflammatory response induced by LDL-derived oxidized phospholipids. *Arterioscler Thromb Vasc Biol*. 2001;21(4):481-488.
- 87. Gao M, Zhao D, Schouteden S, Sorci-Thomas MG, Van Veldhoven PP, Eggermont K, Liu G, Verfaillie CM, Feng Y. Regulation of high-density lipoprotein on hematopoietic stem/progenitor cells in atherosclerosis requires scavenger receptor type BI expression. *Arterioscler Thromb Vasc Biol*. 2014;34(9):1900-1909. doi:10.1161/ ATVBAHA.114.304006.
- 88. Out R, Hoekstra M, Habets K, Meurs I, de Waard V, Hildebrand RB, Wang Y, Chimini G, Kuiper J, Van Berkel TJC, Van Eck M. Combined deletion of macrophage ABCA1 and ABCG1 leads to massive lipid accumulation in tissue macrophages and distinct atherosclerosis at relatively low plasma cholesterol levels. *Arterioscler Thromb Vasc Biol*. 2008;28(2):258-264. doi:10.1161/ ATVBAHA.107.156935.