



FlbA-Regulated Gene *rpnR* Is Involved in Stress Resistance and Impacts Protein Secretion when *Aspergillus niger* Is Grown on Xylose

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ABSTRACT Proteins are secreted throughout the mycelium of *Aspergillus niger* except for the sporulating zone. A link between sporulation and repression of protein secretion was underlined by the finding that inactivation of the sporulation gene *flbA* results in mycelial colonies that secrete proteins throughout the colony. However, $\Delta flbA$ strain hyphae also lyse and have thinner cell walls. This pleiotropic phenotype is associated with differential expression of 36 predicted transcription factor genes, one of which, *rpnR*, was inactivated in this study. Sporulation, biomass, and secretome complexity were not affected in the $\Delta rpnR$ deletion strain of the fungus. In contrast, ribosomal subunit expression and protein secretion into the medium were reduced when *A. niger* was grown on xylose. Moreover, the $\Delta rpnR$ strain showed decreased resistance to H₂O₂ and the proteotoxic stress-inducing agent di-thiothreitol. Taking the data together, RpnR is involved in proteotoxic stress resistance and impacts protein secretion when *A. niger* is grown on xylose.

IMPORTANCE *Aspergillus niger* secretes a large amount and diversity of industrially relevant enzymes into the culture medium. This makes the fungus a widely used industrial cell factory. For instance, carbohydrate-active enzymes of *A. niger* are used in biofuel production from lignocellulosic feedstock. These enzymes represent a major cost factor in this process. Higher production yields could substantially reduce these costs and therefore contribute to a more sustainable economy and less dependence on fossil fuels. Enzyme secretion is inhibited in *A. niger* by asexual reproduction. The sporulation protein FlbA is involved in this process by impacting the expression of 36 predicted transcription factor genes. Here, we show that one of these predicted transcriptional regulators, RpnR, regulates protein secretion and proteotoxic stress resistance. The gene is thus an interesting target to improve enzyme production in *A. niger*.

KEYWORDS asexual development, aspergillus, *flbA*, fungus, protein secretion, proteotoxic stress

Fungi in the genus *Aspergillus* feed on organic waste and can be opportunistic pathogens of plants, animals, and humans (1). They secrete a large amount and diversity of enzymes that are instrumental in these lifestyles and that also make *Aspergillus* species, such as *Aspergillus niger*, important cell factories (2, 3). However, the costs of these enzymes can still be high. For instance, they represent up to 28% of the costs of biofuel production from lignocellulosic biomass (4). The biofuel industry, therefore, is in need of reduced production costs.

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Asexual development in *A. niger* is initiated as a response to changing environmental conditions and as a result of an environment-independent differentiation program (1). It results in conidiophores that form conidia. These spores are heterogeneous in cellular composition and germination rate (5); are dispersed by wind, water, and insects; and give rise to new mycelia. The mycelia, also known as colonies, consist of zones that show heterogeneity in function and composition (3). For instance, the sporulation zone within a colony does not secrete proteins, while the neighboring nonsporulating zones do release proteins into the medium (6). Covering the mycelium with a porous membrane prevents sporulation but does not affect spatial secretion within the mycelium. Apparently, the capacity to sporulate, but not the sporulation process itself, represses secretion.

F1bA was shown to control asexual reproduction, to repress protein secretion in the sporulation zone, to activate mycotoxin production, to inhibit vegetative growth and autolysis, and to impact cell wall architecture (6–9). It does so by activating the $G\alpha$ subunit FadA, which is at the start of a signaling pathway. A total of 36 predicted transcription factor genes are differentially expressed when the $\Delta flbA$ strain is grown on xylose medium, 18 of which are downregulated and 18 upregulated (10, 11). The most downregulated predicted transcription factor gene, *fum21*, is involved in production of the mycotoxins fumonisin and pyranonigrin A, but it does not affect sporulation, secretion, vegetative growth, autolysis, and cell wall architecture (9). Here, we show that the predicted transcription factor *rpnR*, which is upregulated in the $\Delta flbA$ strain (10), is involved in proteotoxic stress resistance and impacts protein secretion when *A. niger* is grown on xylose.

RESULTS

Inactivation of *rpnR*. The predicted transcription factor gene An08g06850 (12) is upregulated 9-, 11-, and 6-fold in the central, middle, and outer concentric zones, respectively, of $\Delta flbA$ strain colonies, making it the most upregulated transcription factor in the strain (10). The protein encoded by An08g06850 showed a bidirectional hit with Rpn4p of *Saccharomyces cerevisiae* (13) and Rpn4 of *Neurospora crassa* (14). The former protein was shown to be an activator of 26S proteasome subunit expression. The function of the *N. crassa* orthologue is not known, but it binds to the same DNA sequences as Rpn4p of *S. cerevisiae* (14). The proteins of *A. niger*, *S. cerevisiae*, and *N. crassa* are 701, 531, and 714 amino acids long. The zinc finger C2H2 domains share 44% and 77% identity, respectively, with that of An08g06850, while the overall identities are 16% and 45%. Based on these data, An08g06850 is now called *rpnR*. The gene was inactivated in the wild-type strain MA234.1, resulting in the $\Delta rpnR$ strain. Deletion of the gene was verified by PCR (see Fig. S1 in the supplemental material). Two independent complemented strains showed restoration of the wild-type phenotype compared to the $\Delta rpnR$ strain phenotypes described below.

Phenotypic analysis of the $\Delta rpnR$ strain. Growth of the $\Delta rpnR$ strain was assessed using minimal medium with xylose as a carbon source (MM-X). The sugar was chosen because differential expression of *rpnR* in the $\Delta flbA$ strain was found on this carbon source (10). On MM-X agar (MM-XA) plates, the $\Delta rpnR$ strain formed colonies with a smaller diameter, but the total biomass was greater than that of the wild type. The diameters of 7-day-old wild-type and $\Delta rpnR$ strain colonies were 6.08 ± 0.20 cm and 5.11 ± 0.34 cm ($n = 6$; $P < 0.05$), whereas the biomasses were 27.4 ± 7.5 mg and 37.4 ± 4.5 mg ($n = 8$; $P < 0.05$), respectively. When xylose was replaced with 1% beechwood xylan, no difference in biomass was found, but the $\Delta rpnR$ strain diameter was again smaller (3.96 ± 0.05 versus 3.66 ± 0.11 cm [$n = 5$; $P < 0.05$]). Together, these data show that the $\Delta rpnR$ strain grows more compactly than the wild type. Both the wild type and the $\Delta rpnR$ strain sporulated in the subperipheral zone and the centers of the colonies (Fig. 1A and B), with similar production of spores per square centimeter (Fig. 1C).

Since Rpn4 of *S. cerevisiae* activates expression of 26S proteasome subunits, it was hypothesized that the $\Delta rpnR$ strain would cope less well with unfolded proteins, i.e., its

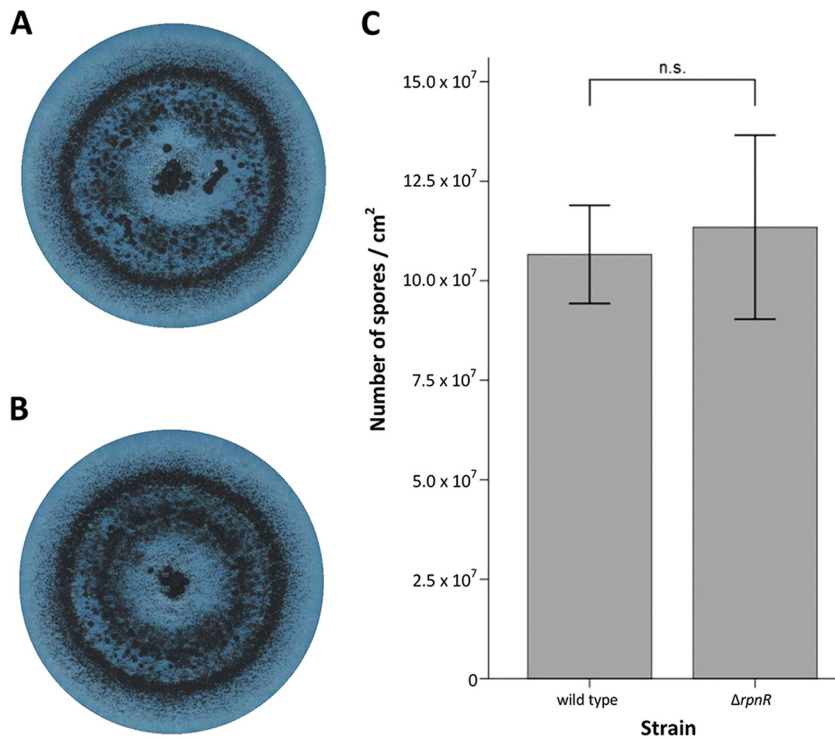


FIG 1 Spatial distribution of sporulation of wild-type strain MA234.1 (A) and the $\Delta rpnR$ strain (B) and numbers of spores per square centimeter produced by MA234.1 and the $\Delta rpnR$ strain (C). Localization and quantification of sporulation were monitored 48 h after removal of the upper membrane of sandwiched colonies. n.s., not significant. The error bars indicate standard deviations.

proteotoxic-stress resistance would be affected. Indeed, germination of the $\Delta rpnR$ strain in MM-X containing 1 mM the unfolded protein response-inducing agent dithiothreitol (DTT) was delayed compared to the wild type, especially at a high concentration of spores (Fig. 2A, D, and G). The growth rate of the $\Delta rpnR$ strain was also reduced at high spore concentrations but did not differ from that of the wild type at lower spore concentrations (Fig. 2A, D, and G). Germination of the $\Delta rpnR$ strain in MM-X containing 0.02% H_2O_2 was also delayed at all spore concentrations and was even absent at low spore concentrations. In addition, the growth rate was decreased at intermediate spore concentrations, while the growth rate was not affected at high spore concentrations (Fig. 2B, E, and H). Interestingly, germination of the $\Delta rpnR$ strain was also delayed when no stressor was present. In fact, the differences between the $\Delta rpnR$ strain and the wild type in germination timing and growth speed were equal in medium containing DTT or H_2O_2 and in medium without these stressors when a low or high spore concentration, respectively, was examined (Fig. 2C, F, and I).

SDS-PAGE revealed that protein secretion was not affected in shaken liquid cultures of the $\Delta rpnR$ strain when maltose was used as a carbon source (data not shown). However, it was reduced in the presence of xylose (Fig. 3A). Proteomics was performed to examine whether secretome complexity was also affected. To this end, similar amounts of total secreted protein from MA234.1 and the $\Delta rpnR$ strain were analyzed by mass spectrometry (MS). A total of 316 proteins were identified in the medium of the $\Delta rpnR$ strain, 223 of which were detected in at least 3 out of 4 replicates. These numbers were 341 and 257, respectively, for the wild type. A signal peptide for secretion was found in 194 and 222 of the secreted $\Delta rpnR$ and wild-type proteins, respectively, while 11 and 10 of the proteins that did not have a signal peptide are predicted to have an intracellular localization. The wild-type secretome was relatively enriched for 28 proteins compared to the $\Delta rpnR$ strain secretome (Table 1; see Table S1 in the supplemental material). Conversely, the $\Delta rpnR$ secretome was not enriched for proteins

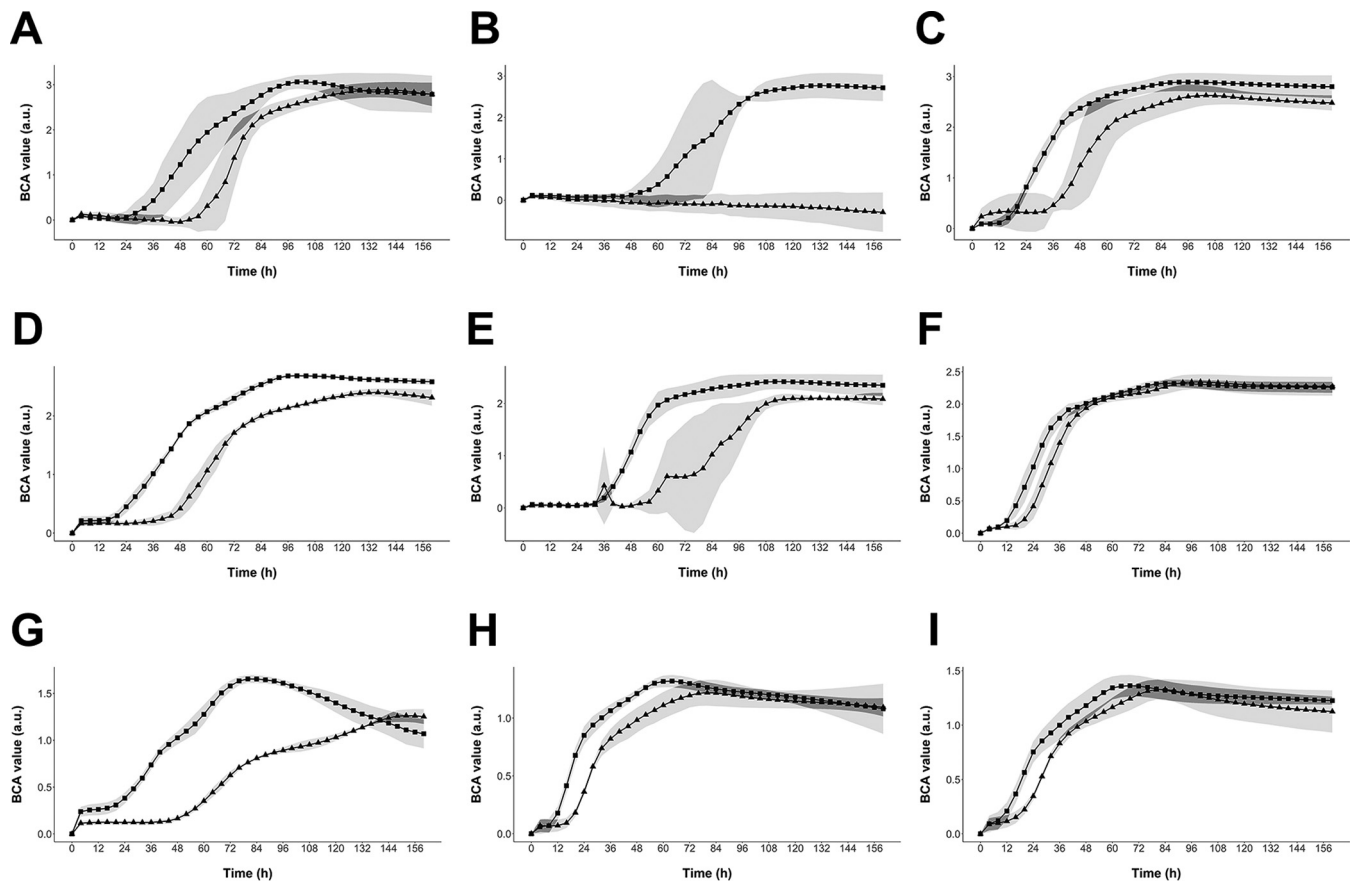


FIG 2 Growth curves of wild-type strain MA234.1 (squares) and the $\Delta rpnR$ strain (triangles) in wells with MM-X containing 1 mM DTT (A, D, and G), 0.02% H_2O_2 (B, E, and H), or no stressor (C, F, and I) with 10^3 (A, B, and C), 10^4 (D, E, and F), and 10^5 (G, H, and I) conidia per well. The data points and lines for each strain show the average of 3 measurements, while the light-gray shading shows the 95% confidence intervals. Dark-gray shading indicates overlap between the confidence intervals of the two strains. Absence of overlap implies significant differences between the strains.

compared to the wild type. Of the proteins enriched in the wild type (i.e., relatively reduced in the $\Delta rpnR$ strain), 11 could not be assigned to a Gene Ontology (GO) subset (GO slim) term (process component), and 13 were associated with carbohydrate metabolic process, 2 with developmental process, and 2 with response to stress. The GO slim terms protein catabolic process, cellular amino acid metabolic process, cellular protein modification process, asexual sporulation, and pathogenesis could each be assigned to one protein (Fig. 3B; see Table S1). Closer inspection of the set of proteins without a GO term showed that it contains the β -1,3-glucanase BgxB (An02g13180) (15), the orthologue of an XlnR-regulated lipase of *Aspergillus oryzae* (An10g00790), a putative muconate cycloisomerase (An01g14730), a glutamyl-tRNA^{Gln} amidotransferase (An05g01860), and a putative α -1,6-mannanase (An07g07700). From the 16 proteins involved in carbohydrate metabolism (the 13 associated with carbohydrate metabolic process plus BgxB, An07g07700, and Msd5), 9 are involved in carbon source modification, while 7 are (putatively) involved in remodeling of the cell wall (Table 1).

Gene expression analysis. RNAs of the $\Delta rpnR$ strain and its progenitor, MA234.1, that had been transferred to MM-X for 4 h after pregrowth in nutrient-rich transformation medium (TM) were sequenced (see Materials and Methods). A total of 505 genes were differentially expressed, 253 and 252 of which were down- and upregulated, respectively, in the $\Delta rpnR$ strain (see Table S2 in the supplemental material). Expression of *rpnR* was 0, thus confirming its proper deletion. Of the downregulated genes in the $\Delta rpnR$ strain, 0 and 14 genes were also downregulated in the $\Delta flbA$ and $\Delta fum21$ strains, respectively (Table 2). Similarly, 2 and 4 of the upregulated genes in the $\Delta rpnR$ strain were also upregulated in the $\Delta flbA$ and $\Delta fum21$ strains, respectively (Table 2). GO term

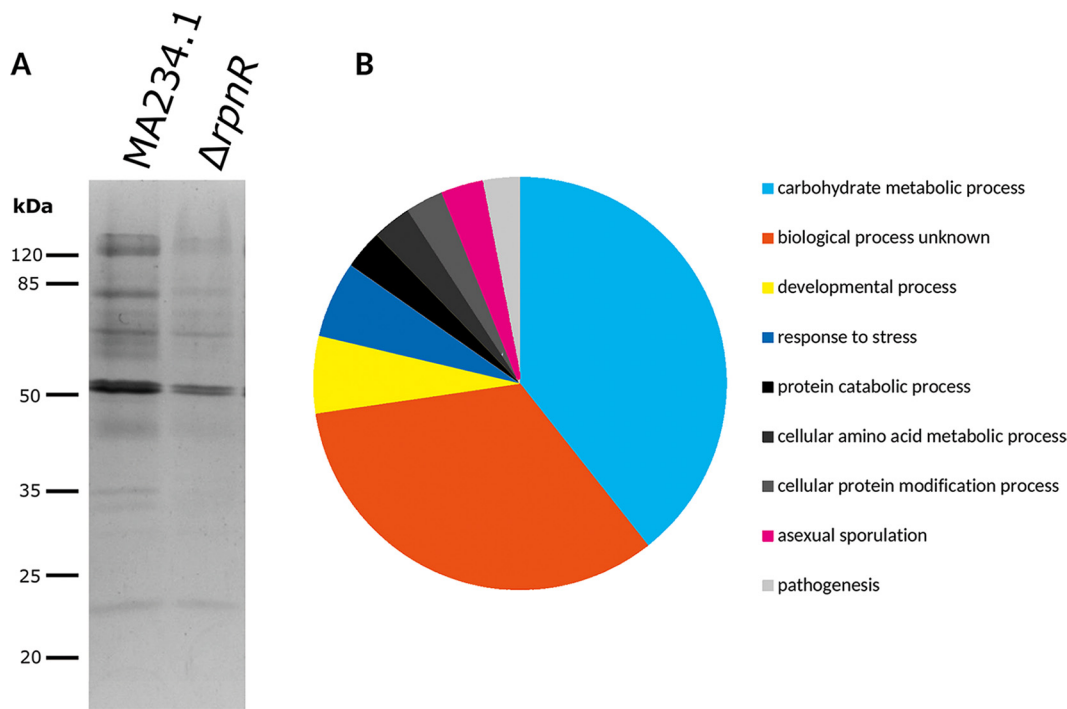


FIG 3 Differences in the secretomes of the $\Delta rpnR$ strain and the wild type. (A) SDS-PAGE of medium proteins of shaken liquid cultures of the wild type and the $\Delta rpnR$ strain pregrown in TM-G for 16 h and transferred to MM-X for 24 h. The gel was stained with Coomassie brilliant blue R-250. (B) Pie chart of GO slim terms of the 28 relatively underrepresented secreted proteins of the $\Delta rpnR$ strain. Proteins PepE, Bgt1, and An01g14960 occur in 2 categories and MsdS in 3.

enrichment analysis of the downregulated genes showed that GO terms related to translation were overrepresented (see Fig. S2 in the supplemental material). In fact, about 20% of the downregulated genes were 60S or 40S ribosomal subunits, while 7 out of 13 subunits of translation initiation factor 3 (eIF3) (16) were also downregulated (see Table S2). This was reflected in the overrepresentation of the KEGG annotation terms ribosome, translation, genetic information processing, nucleotide metabolism, and RNA transport (Table 3). In addition, the GO term alpha amino acid metabolic process was overrepresented (see Fig. S2). Furthermore, small secreted proteins were

TABLE 1 Underrepresented secreted carbohydrate metabolism proteins in the $\Delta rpnR$ strain medium compared to the wild type

Process	Name or protein ID	Function
Carbon source utilization	AxeA	Acetyl xylan esterase
	Suc1	Invertase
	AguA	α -Glucuronidase
	An11g00390	Rhamnogalacturonan lyase
	An04g03170	β -Glucosidase
	An11g02100	β -Glucosidase
	CbhB	Cellobiohydrolase
	An08g01900	Xylan β -xylosidase
	An08g05230	Endoglucanase 4
	Cell wall remodeling	Bgt1
AgnB		α -1,3-Glucanase (53)
BgxB		β -1,3-Glucanase
An08g08370 ^a		α -1,2-Mannosidase
An08g03060 ^a		α -Mannosidase (54)
An07g07700 ^a		α -1,6-Mannanase
MsdS ^a		α -1,2-Mannosidase

^aThe protein is likely active on the fungal cell wall (55) and might play a role in glycosylation of proteins like those present in the cell wall.

TABLE 2 Differentially expressed genes in the $\Delta rpnR$ strain that are also down- and upregulated in the $\Delta flbA$ or $\Delta fum21$ strain and fold changes in the strains compared to the wild type^a

Expression in the $\Delta rpnR$ strain	Protein ID	Functional annotation ^b	Strain	Fold change	
				$\Delta rpnR$ strain	$\Delta flbA$ or $\Delta fum21$ strain ^c
Downregulated	1142051	No annotation	$\Delta fum21$	131.0	192.7
	1142053	Zinc finger transcription factor, C2H2 type	$\Delta fum21$	106.7	1312.2
	1166044	No annotation	$\Delta fum21$	37.2	126.6
	1087288	Taurine catabolism dioxygenase TauD	$\Delta fum21$	16.1	16.5
	1082505	Major facilitator superfamily transporter	$\Delta fum21$	15.7	22.3
	1107461	Chitinase	$\Delta fum21$	7.9	4.8
	1089440	Major facilitator superfamily transporter	$\Delta fum21$	4.9	11.6
	1157348	UDP-glucose 4-epimerase	$\Delta fum21$	3.5	5.6
	1150465	No annotation	$\Delta fum21$	3.2	4.2
	45784	No annotation	$\Delta fum21$	2.8	14.8
	1186369	No annotation	$\Delta fum21$	2.8	33.6
	1109756	No annotation	$\Delta fum21$	2.7	8.5
	1124492	Phosphoglycerate mutase	$\Delta fum21$	2.3	4.4
	1184525	NRPS-like protein	$\Delta fum21$	2.1	8.0
	Upregulated	1145520	Triacylglycerol lipase	$\Delta flbA$	20.5
1152605		Acyl-coenzyme A dehydrogenase	$\Delta flbA$	2.5	3.5
1183897		AnAFP	$\Delta fum21$	9.6	6.6
1184369		Carbohydrate esterase family 16 protein	$\Delta fum21$	3.4	4.5
1156756		No annotation	$\Delta fum21$	2.6	5.4
1184413		No annotation	$\Delta fum21$	2.4	4.0

^aAll the strains were grown in TM-G for 16 h, followed by 4 h in MM-X. Differentially expressed genes in the $\Delta flbA$ and $\Delta fum21$ strains can be found in references 9 and 10.

^bWhen no function of the gene could be inferred from MycoCosm, GO/KEGG terms, and a blastp search, the functional annotation is given as "no annotation." AnAFP, *Aspergillus niger* antifungal protein.

^cAs indicated in the "Strain" column.

overrepresented, as well as genes of secondary-metabolism cluster 49 (4 out of 17 genes) (Table 3). Terms related to regulation of translation and regulation of the ribosome were underrepresented (see Fig. S3 in the supplemental material). This implies that RpnR specifically regulates constituents of the ribosome but not their regulators. Other underrepresented terms were related to the nucleus, DNA binding, and RNA synthesis (see Fig. S3), indicating that RpnR specifically affects proteins that act outside the nucleus and does not regulate the ribosome at the RNA level. Terms related to zinc ion binding were also underrepresented (see Fig. S3).

Among the upregulated genes in the $\Delta rpnR$ strain, GO terms related to arabinose metabolism, oxidoreductive activity, and coenzyme binding were overrepresented (see Fig. S4 in the supplemental material). More precisely, genes encoding the arabinofuranosidases AbfB, AbfC, and AbfE were upregulated in the $\Delta rpnR$ strain (see Table S2). However, since no proteins were overrepresented in the proteome of the $\Delta rpnR$ strain (see above), these arabinofuranosidases are apparently not upregulated at the protein level. Many KEGG annotation terms were overrepresented (Table 3). Of these, the 2nd-level term amino acid metabolism represented 12% of the upregulated genes, carbohydrate metabolism 11%, xenobiotics biodegradation and metabolism 10%, lipid metabolism 10%, and metabolism of cofactors and vitamins 7%. Furthermore, 5 Pfam domains were overrepresented (Table 3), including domains associated with lipid metabolism (PF13561.1) and oxidoreductive activity (PF00106.20 and PF00107.21). Also, genes encoding proteins with a transmembrane domain and genes belonging to secondary-metabolism cluster 75 (5 out of 18) were overrepresented (Table 3). Together, these data indicate that RpnR deficiency reduces expression of ribosomal subunits, whereas it activates arabinose metabolism, lipid metabolism, coenzyme metabolism, and oxidoreductase activity.

DISCUSSION

The transcription factor Rpn4 of *S. cerevisiae* is involved in regulation of DNA repair (17, 18) and functions as a transcriptional activator of 26S proteasomal subunits (13, 19)

TABLE 3 Overrepresented KEGG pathways, Pfam domains, and secondary-metabolism-related annotation terms among the down- and upregulated genes of the $\Delta rpnR$ strain compared to the wild type^a

Gene regulation	Location	Overrepresented term in $\Delta rpnR$ strain	
Downregulated	KEGG pathways	KEGG third level: ribosome KEGG second level: translation KEGG first level: genetic information processing KEGG second level: nucleotide metabolism KEGG third level: RNA transport	
	Secondary-metabolism clusters	Part of secondary-metabolism cluster 49	
Upregulated	KEGG pathways	KEGG first level: metabolism KEGG second level: overview KEGG third level: valine, leucine and isoleucine degradation KEGG second level: xenobiotics biodegradation and metabolism KEGG third level: fatty acid metabolism KEGG second level: amino acid metabolism KEGG third level: fatty acid biosynthesis KEGG fourth level: K00059:fabG; 3-oxoacyl-[acyl-carrier protein] reductase (EC 1.1.1.100) KEGG EC: acyl-carrier protein KEGG third level: biosynthesis of unsaturated fatty acids KEGG second level: lipid metabolism KEGG second level: carbohydrate metabolism KEGG third level: biotin metabolism KEGG third level: propanoate metabolism KEGG third level: chlorocyclohexane and chlorobenzene degradation KEGG third level: benzoate degradation KEGG third level: toluene degradation KEGG EC: EC 1.1.1.65 KEGG fourth level: K05275:E1.1.1.65; pyridoxine 4-dehydrogenase (EC 1.1.1.65) KEGG fourth level: K01714:dapA; 4-hydroxy-tetrahydrodipicolinate synthase (EC 4.3.3.7) KEGG EC: EC 4.3.3.7 KEGG second level: metabolism of cofactors and vitamins KEGG third level: metabolism of xenobiotics by cytochrome P450 KEGG third level: aminobenzoate degradation KEGG third level: caprolactam degradation KEGG third level: fatty acid degradation KEGG third level: carbon metabolism KEGG third level: vitamin B ₆ metabolism KEGG third level: PPAR signaling pathway KEGG third level: butanoate metabolism KEGG third level: chemical carcinogenesis	
		Pfam domains	PF13561.1 enoyl-(acyl carrier protein) reductase PF08659.5 KR domain PF00106.20 short-chain dehydrogenase PF00107.21 zinc-binding dehydrogenase PF00701.17 dihydrodipicolinate synthetase family
			Secondary-metabolism clusters
		Other terms	Transmembrane domain

^aAll the strains were grown in TM-G for 16 h, followed by 4 h in MM-X.

by binding to PACE promoter sequences (13, 17, 20). Here, it is shown that the *A. niger* orthologue of Rpn4, RpnR, functions differently than its *S. cerevisiae* counterpart. Deletion of its encoding gene resulted in reduced proteotoxic- and oxidative-stress resistance, which may well explain why expression of the ribosomal subunits and the total amount of secreted proteins were also reduced when *A. niger* was grown on xylose. Expression of genes encoding 26S proteasome subunits remained unchanged (data not shown). This contrasts with the *S. cerevisiae rpn4* deletion strain, in which 26S proteasome subunit expression was decreased while ribosomal subunit genes were unaffected (17, 21). It is, however, in line with results in *N. crassa*, where the majority of proteasome genes do not contain the DNA sequences to which *N. crassa* Rpn4 binds (14). It should be noted that reduced proteotoxic- and oxidative-stress resistance was not found at low and high spore concentrations, respectively. How can this be explained? With increasing spore concentrations, the amounts of stressor and nutrient

per spore decrease. These amounts may impact the phenotype. For instance, the effect of H₂O₂ is expected to be lower at high spore concentrations because its effective amount per spore is less.

There is a link between unfolded-protein levels in the cell and ribosomal gene expression and RNA translation. Proteotoxic stress is counteracted by the unfolded-protein response (UPR), which not only increases the abundance of chaperones and heat shock proteins in the endoplasmic reticulum (ER), but also, importantly, reduces the influx of new proteins into the ER through downregulating translation. Translation attenuation in animals (22) and *S. cerevisiae* (21) is achieved by inhibition of translation initiation factor eIF2 α activity by phosphorylation. In addition, *S. cerevisiae* downregulates genes encoding ribosomal subunits (23, 24), which increases resistance to proteotoxic stress (21, 25). Whether these processes also occur in filamentous fungi, such as *A. niger*, is not known. It is known that activation of genes involved in the UPR occurs via transcription factor Hac1 in *S. cerevisiae* (26, 27) and its homologue HacA in *A. niger* (28). Hac1/HacA is activated by Ire1/IreA, a molecule that senses proteotoxic stress in the ER. As a result, it removes an intron from Hac1 pre-mRNA, so that an active Hac1 protein is produced (29). On the other hand, downregulation of *S. cerevisiae* genes encoding ribosomal subunits is regulated by deactivation of protein kinase A (30), which in turn activates the general stress response regulator Msn4.

Colonies of *A. niger* that are transferred from a nutrient-rich medium to a defined medium (i.e., TM to MM) with xylose as a carbon source induce genes encoding secreted enzymes. This induction has been shown to induce a response reminiscent of the UPR in *A. niger* (31), likely due to increased influx of proteins in the ER. The fact that the $\Delta rpnR$ strain is more sensitive to proteotoxic stress agrees with the finding that genes encoding ribosomal proteins are downregulated in this deletion strain. In addition, 7 out of the 15 subunits of eIF3 (16) were found to be downregulated in the $\Delta rpnR$ strain. This might represent another way in which translation is inhibited, analogous but not similar to eIF2 α activity inhibition in yeast and animals (see above). Decreased translation would reduce the amount of unfolded proteins. This, apparently, is effective, since protein secretion, but not biomass formation, is affected in the $\Delta rpnR$ strain. The fact that protein secretion was not affected when the $\Delta rpnR$ strain was grown in maltose-containing medium may be explained by the less complex secretome that is found on this carbon source (32).

The biomass of the $\Delta rpnR$ strain was higher or not reduced in colonies grown on agar medium. In all cases, the diameter was reduced, implying that the biomass per surface area was increased. This was probably caused by increased hyphal branching, but the underlying mechanism is not known. What also remains unexplained is why the $\Delta rpnR$ strain is less tolerant of proteotoxic stress. UPR malfunction could be an explanation. However, the fact that chaperones or heat shock proteins are not differentially expressed in the $\Delta rpnR$ strain suggests that the rest of the UPR is induced equally strongly as in the wild type. This is also reflected by the fact that both HacA and IreA are not differentially expressed. Therefore, other factors make the $\Delta rpnR$ strain more sensitive to proteotoxic stress. It is also unclear why 28 specific proteins have lower abundance in the secretome of the $\Delta rpnR$ strain than in the wild type, in addition to the overall secretion reduction. Their corresponding genes are not differentially expressed, so the difference in the secreted protein level might arise from posttranscriptional events, such as high sensitivity to ribosome depletion.

What do these data mean for the function of RpnR in FlbA-mediated sporulation-inhibited protein secretion? In wild-type colonies, FlbA is active in sporulating regions of the colony, where it downregulates *rpnR*. Our results indicate that this leads to less proteotoxic-stress resistance and lower ribosome abundance and thus to lower protein secretion in sporulating regions than in the nonsporulating zones of the colony. Indeed, this has been observed in wild-type colonies. Downregulation of ribosomal protein genes has been shown to occur during sporulation in *Aspergillus fumigatus* (33), also supporting this theory. The facts that this ribosomal protein downregulation is dependent on BrIA (33) and that *rpnR* is also upregulated in zones of maltose-grown colonies

TABLE 4 Primers used in this study

Primer name	5'–3' sequence
1	AGTATTCCAAGCTCTGGACTAACCTCTTTAATGGGTCAGTCAATGCCTGGACTATC
2	TGATGTGTGTGACGTGATGTGTATATCCTAGCAGGTATAGACACGGAATTTGCCAC
3	TGATGATGGATATATGGAAGCTGGAGGATGTCCAATTGAGAGGTAAGCCAGGG
4	TAGCAGGTGAGAACTCACTGTACTCGACTCCTGCAGAAACCACAAAGGTACCCTACGTC
5	TAGGATATACACATCACGTC
6	CATCCTCCAGCTTCCATATATC
7	GGTTTAGCGGGTGCAGAAG
8	GGCGTCGGTTTCCACTATC
9	AAAGTTCGACAGCGTCTCC
10	GTTTGGGCTTCAATGTCTTAC
11	ATAGGGAGAGCGGCCCGGAGATGCAGATCGAGTG
12	GAAGATCTGGCGGCCACCCGTGACCCAATGAGAA
13	ATCCAAGTGCACCTCAGAGCC
14	CATGCATGGTTGCCTAGTGAA

lacking *fluG* (34) imply that downregulation of ribosomal proteins can be activated by several genes in the sporulation pathway. Taken together, the data show that the putative transcription factor RpnR of *A. niger* regulates proteotoxic-stress resistance and, possibly as a consequence, protein secretion and may therefore be an important target for improving *A. niger* as a cell factory for enzyme production.

MATERIALS AND METHODS

Strains and culture conditions. *A. niger* strain MA234.1 (transient *kusA::amdS*; *pyrG*⁺) (35) and its derivative Δ *rpnR* strain (see below) were grown at 30°C on MM (36) with 25 mM xylose as a carbon source and either containing (MM-XA) or not containing (MM-X) 1.5% agar. Alternatively, 1% (wt/vol) beechwood xylan was used instead of xylose. Liquid cultures were pregrown in TM (MM containing 0.5% yeast extract and 0.2% Casamino Acids) (37) containing 25 mM glucose (TM-G). To this end, 100 ml medium was inoculated with 5×10^8 freshly harvested conidia from 3-day-old MM-XA cultures and shaken in 250-ml Erlenmeyer flasks at 200 rpm. After 16 h, the mycelium was washed with 0.9% NaCl, and 10 g (wet weight) was transferred to a 1-liter Erlenmeyer flask containing 150 ml MM-X. Growth was prolonged for 4 h (RNA sequencing) or 24 h (SDS-PAGE and proteomics) at 200 rpm. Biomass formation and sporulation were assessed by growing colonies in a 0.45-mm-thick layer of 1.25% agarose between perforated polycarbonate membranes (0.1- μ m pores; diameter, 76 mm; Profiflora, Almere, The Netherlands) (38). To this end, the sandwiched cultures were inoculated in the center of the agarose layer with 10^4 conidia contained in 2 μ l H₂O. The upper polycarbonate membrane was placed 24 h postinoculation to prevent spreading of conidia. After 7 days, the diameter of the colony was measured and the dry-weight biomass was determined by subtracting the average weight of a noninoculated agarose layer. Alternatively, the upper membrane was taken off to enable the colony to sporulate during a 48-h period. Differences in biomass, colony diameter, or spore production were calculated using an independent-samples *t* test.

Inactivation of *rpnR* and complementation of the deletion strain. The *rpnR* deletion construct was made by amplifying the 5' and 3' flanks of *rpnR* (protein ID, An08g06850) (12) from genomic DNA by PCR using primer pairs 1/2 and 3/4, respectively (Table 4). The hygromycin resistance gene *hph* was amplified from the vector pAN7.1 (39) using primer pair 5/6 (Table 4). Split marker fragments of this selection marker fused to the 3' and 5' flanks were created by fusion PCR (40) using primer pairs 7/8 and 9/10 (Table 4) for the 5' and 3' fragments, respectively.

The *rpnR* complementation construct was made by amplifying the genomic coding sequence of *rpnR* together with its 1,017-bp promoter and 559-bp terminator sequences. To this end, MA234.1 genomic DNA was used in combination with Phusion polymerase (Thermo Scientific, Wilmington, DE, USA) and primer pair 11/12 (Table 4). The resulting fragment was inserted in the NotI site of vector pMA357 (9) using an InFusion HD cloning kit (Clontech, Mountain View, CA, USA). This resulted in vector pDA2, containing the *amdS* selection cassette and the gene *rpnR* under the control of its own promoter and terminator.

The split marker fragments and the complementation construct were transformed to strain MA234.1 and the Δ *rpnR* strain, respectively, as described previously (41). Transformants were purified twice on MM-XA containing 100 μ g ml⁻¹ hygromycin or 15 mM CsCl and 10 mM acetamide as a nitrogen source for selection of deletion and complemented strains, respectively.

RNA sequencing and analysis. Mycelium from biological triplicates was taken up in TRIzol reagent (Invitrogen, Bleiswijk, The Netherlands) after homogenization in a Tissue Lyzer II (Qiagen, Venlo, The Netherlands). After extraction with chloroform, total RNA was precipitated with isopropanol, washed with ethanol, and resuspended in RNase-free water. RNA was purified using a NucleoSpin RNA kit (Macherey-Nagel, Düren, Germany), and its concentration and purity were checked using a Nanodrop ND-1000 (Thermo Scientific).

Library construction, cluster generation, and sequencing of cDNA were performed by ServiceXS (Leiden, The Netherlands). Library construction and cluster generation were performed with the NEBNext Ultra Directional RNA Library Prep kit for Illumina (New England Biolabs, Ipswich, MA, USA). Sequencing of cDNA was performed using an Illumina NextSeq 500 with NextSeq control software 2.0.2 (Illumina, San Diego, CA, USA), generating single reads of 75 bp. For image analysis, base calling, and quality checking, the Illumina data analysis pipelines RTA v2.4.11 and Bcl2fastq v2.17 were used. The sequences were then aligned to the *A. niger* ATCC 1015 genome assembly (version Aspni7) (42) obtained from MycoCosm (43) using HISAT2 version 2.0.5 (44). Functional annotation of the predicted genes was obtained from MycoCosm as described previously (5), which, in the case of the differentially expressed genes, was manually approved by examining the GO and KEGG terms and by blastp search. However, ribosomal subunits were named by homology with *S. cerevisiae*, as proposed by Nakamura et al. (45).

The expression levels of genes were normalized to fragments per kilobase of exon model per million fragments (FPKM). Cuffdiff version 2.2.1 (46) was used to identify reads mapping to predicted genes and to identify differentially expressed genes. The bias correction method was used while running Cuffdiff (47). In addition to Cuffdiff's requirements for differential expression (including a false-discovery rate [FDR]-adjusted *P* value of <0.05), we selected for a >2-fold change and a minimal expression level of 10 FPKM in at least one of the samples. The quality of the results was analyzed using CummeRbund (48).

Custom scripts were developed in Python and R to analyze over- and underrepresentation of functional annotation terms in sets of differentially regulated genes using the Fisher exact test. The Benjamini-Hochberg correction was used to correct for multiple tests using a *P* value of <0.05. Over- and underrepresented GO terms were visualized using RamiGO (49). GO terms, KEGG pathways, potential protein-degrading activity, presence of a secretion signal or a transmembrane domain, requirements for terming a protein a small secreted protein, and a function in secondary metabolism were predicted as described previously (5).

SDS-PAGE. Proteins were precipitated with acetone and taken up in a 5% volume of sample buffer compared to the original volume of culture medium. Proteins were separated in 12.5% SDS-PAGE gels using a Pierce prestained protein molecular weight marker (Thermo Scientific). Gels were fixed with 50% methanol and 10% acetic acid; stained with 0.1% Coomassie brilliant blue R-250; and destained with 25% methanol, 10% acetic acid. Protein bands were imaged using a Universal Hood III with Image Lab software (Bio-Rad Laboratories BV, Veenendaal, The Netherlands).

Proteomics. Culture medium (4-ml samples of biological quadruplicates) was concentrated 100-fold for 30 min at $4\,000 \times g$ and 4°C using an Amicon Ultra-4 centrifugal filter unit with a cutoff of 10 kDa (Millipore, Billerica, MA, USA). The concentrated medium was washed twice by adding 2.5 ml phosphate-buffered saline (PBS), followed by centrifugation steps. The protein concentrations of the samples were estimated by SDS-PAGE analysis. Based on this, sample buffer was added to correct for protein concentrations in the different samples. Samples were separated on a 12% Bis-Tris SDS-PAGE gel (Bio-Rad). The gel was run for 2 to 3 cm and stained with colloidal Coomassie dye G-250 (Gel Code blue stain reagent; Thermo Scientific), after which each lane was cut into two pieces to reduce complexity. The gel pieces were reduced, alkylated, and digested overnight with trypsin at 37°C. The peptides were extracted with 100% acetonitrile, dried in a vacuum concentrator, and resuspended in 10% (vol/vol) formic acid. An ultrahigh-performance liquid chromatography (UHPLC) 1290 system (Agilent) coupled to an Orbitrap Q Exactive HF Biopharma mass spectrometer (Thermo Scientific) was used for MS. Peptides were trapped for 5 min in solvent A (0.1% formic acid in water) on a Dr Maisch Reprosil C₁₈ column (3 μm by 2 cm by 100 μm) before being separated on an analytical column (Agilent Poroshell EC-C₁₈; 2.7 μm by 50 cm by 75 μm) using a gradient of solvent A consisting of 13 to 44% in 95 min, 44 to 100% in 3 min, 100% for 1 min, 100 to 0% in 1 min, and finally 0% for 10 min. The flow was passively split to 300 nl min⁻¹. The mass spectrometer was operated in data-dependent mode. Full-scan MS spectra from *m/z* 375 to 1,600 were acquired at a resolution of 60,000 at *m/z* 200 after accumulation to a target value of 3E6. Up to 15 most intense precursor ions were selected for fragmentation. High-energy collisional dissociation fragmentation was performed at a normalized collision energy of 27% after accumulation to a target value of 1E5. Tandem MS (MS-MS) was acquired at a resolution of 15,000. Raw data files were processed using Proteome Discover (version 1.4.1.14). A database search was performed against the *A. niger* CBS 513.88 database (12) using Mascot (version 2.5.1; Matrix Science, London, UK) as a search engine. Cysteine carbamidomethylation was set as a fixed modification and methionine oxidation as a variable modification. Trypsin was specified as the enzyme, and up to two missed cleavages were allowed. Filtering was done at a 1% false-discovery rate and an ion score of >20. For CRAPome analysis (50), proteins with a SAINTexpress (51) probability score of ≥0.75 were considered differentially expressed. Differentially expressed proteins were mapped to GO slim terms using the GO slim mapper on AspGD (12). SignalP 4.1 (52) was used to identify signal peptides for secretion.

Stress resistance assays. Resistance to H₂O₂ and DTT was determined in biological triplicates by inoculating 10³ to 10⁵ spores in 96-well plates filled with 200 μl MM-X per well and either supplemented or not with 0.02% H₂O₂ or 1 mM DTT. The spores were incubated at 30°C for 7 days. Growth was monitored using oCelloScope (BioSense Solutions, Farum, Denmark) and UniExplorer (BioSense) software. The bicinchoninic acid (BCA) normalized algorithm was used to determine the total surface area of the fungal mycelium.

Accession number(s). The RNA sequencing data have been deposited in NCBI GEO with accession number GSE102899 (<https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi>).

SUPPLEMENTAL MATERIAL

Supplemental material for this article may be found at <https://doi.org/10.1128/AEM.02282-18>.

SUPPLEMENTAL FILE 1, PDF file, 1.3 MB.

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We declare no conflict of interest.

D.A., S.G.V.D.B., H.P., M.A., and R.A.O. designed and performed experiments; D.A., H.P., M.A.F.A., M.A., A.F.J.R., R.A.O. and H.A.B.W. interpreted data; D.A. and H.A.B.W. wrote the manuscript.

REFERENCES

- Krijgsheld P, Bleichrodt R, van Veluw GJ, Wang F, Müller WH, Dijksterhuis J, Wösten HAB. 2013. Development in *Aspergillus*. *Stud Mycol* 74:1–29. <https://doi.org/10.3114/sim0006>.
- Meyer V, Wu B, Ram AFJ. 2011. *Aspergillus* as a multi-purpose cell factory: current status and perspectives. *Biotechnol Lett* 33:469–476. <https://doi.org/10.1007/s10529-010-0473-8>.
- Wösten HAB, van Veluw GJ, de Bekker C, Krijgsheld P. 2013. Heterogeneity in the mycelium: implications for the use of fungi as cell factories. *Biotechnol Lett* 35:1155–1164. <https://doi.org/10.1007/s10529-013-1210-x>.
- Johnson E. 2016. Integrated enzyme production lowers the cost of cellulosic ethanol. *Biofuels Bioprod Bioref* 10:164–174. <https://doi.org/10.1002/bbb.1634>.
- Teertstra WR, Tegelaar M, Dijksterhuis J, Golovina EA, Ohm RA, Wösten HAB. 2017. Maturation of conidia on conidiophores of *Aspergillus niger*. *Fungal Genet Biol* 98:61–70. <https://doi.org/10.1016/j.fgb.2016.12.005>.
- Krijgsheld P, Nitsche BM, Post H, Levin AM, Müller WH, Heck AJR, Ram AFJ, Altelaar AFM, Wösten HAB. 2013. Deletion of *flbA* results in increased secretome complexity and reduced secretion heterogeneity in colonies of *Aspergillus niger*. *J Proteome Res* 12:1808–1819. <https://doi.org/10.1021/pr301154w>.
- Wieser J, Lee BN, Fondon IIIJW, Adams TH. 1994. Genetic requirements for initiating asexual development in *Aspergillus nidulans*. *Curr Genet* 27:62–69. <https://doi.org/10.1007/BF00326580>.
- Yu J-H, Wieser J, Adams TH. 1996. The *Aspergillus* FlbA RGS domain protein antagonizes G protein signaling to block proliferation and allow development. *EMBO J* 15:5184–5190. <https://doi.org/10.1002/j.1460-2075.1996.tb00903.x>.
- Aerts D, Hauer EE, Ohm RA, Arentshorst M, Teertstra WR, Phippen C, Ram AFJ, Frisvad JC, Wösten HAB. 2018. The FlbA-regulated predicted transcription factor Fum21 of *Aspergillus niger* is involved in fumonisin production. *Antonie Van Leeuwenhoek* 111:311–322. <https://doi.org/10.1007/s10482-017-0952-1>.
- Krijgsheld P, Wösten HAB. 2013. Transcriptome analysis of zones of colonies of the $\Delta flbA$ strain of *Aspergillus niger*. *Fungal Genomics Biol* 3:109.
- Aerts D. 2018. Regulators controlled by the sporulation gene *flbA* of *Aspergillus niger*. Ph.D. thesis. Universiteit Utrecht, Utrecht, The Netherlands.
- Cerqueira GC, Arnaud MB, Inglis DO, Skrzypek MS, Binkley G, Simison M, Miyasato SR, Binkley J, Orvis J, Shah P, Wymore F, Sherlock G, Wortman JR. 2014. The *Aspergillus* Genome Database: multispecies curation and incorporation of RNA-Seq data to improve structural gene annotations. *Nucleic Acids Res* 42:D705–D710. <https://doi.org/10.1093/nar/gkt1029>.
- Mannhaupt G, Schnall R, Karpov V, Vetter I, Feldmann H. 1999. Rpn4p acts as a transcription factor by binding to PACE, a nonamer box found upstream of 26S proteasomal and other genes in yeast. *FEBS Lett* 450:27–34. [https://doi.org/10.1016/S0014-5793\(99\)00467-6](https://doi.org/10.1016/S0014-5793(99)00467-6).
- Gasch AP, Moses AM, Chiang DY, Fraser HB, Berardini M, Eisen MB. 2004. Conservation and evolution of cis-regulatory systems in ascomycete fungi. *PLoS Biol* 2:e398. <https://doi.org/10.1371/journal.pbio.0020398>.
- van Munster JM, Nitsche BM, Krijgsheld P, van Wijk A, Dijkhuizen L, Wösten HAB, Ram AFJ, van der Maarel MJE. 2013. Chitinases CtcB and Cfcl modify the cell wall in sporulating aerial mycelium of *Aspergillus niger*. *Microbiology* 159:1853–1867. <https://doi.org/10.1099/mic.0.067967-0>.
- Smith MD, Gu Y, Querol-Audí J, Vogan JM, Nitido A, Cate JH. 2013. Human-like eukaryotic translation initiation factor 3 from *Neurospora crassa*. *PLoS One* 8:e78715. <https://doi.org/10.1371/journal.pone.0078715>.
- Jelinsky SA, Estep P, Church GM, Samson LD. 2000. Regulatory networks revealed by transcriptional profiling of damaged *Saccharomyces cerevisiae* cells: Rpn4 links base excision repair with proteasomes. *Mol Cell Biol* 20:8157–8167. <https://doi.org/10.1128/MCB.20.21.8157-8167.2000>.
- Karpov DS, Spasskaya DS, Tutyayeva VV, Mironov AS, Karpov VL. 2013. Proteasome inhibition enhances resistance to DNA damage via upregulation of Rpn4-dependent DNA repair genes. *FEBS Lett* 587:3108–3114. <https://doi.org/10.1016/j.febslet.2013.08.007>.
- Xie Y, Varshavsky A. 2001. RPN4 is a ligand, substrate, and transcriptional regulator of the 26S proteasome: a negative feedback circuit. *Proc Natl Acad Sci U S A* 98:3056–3061. <https://doi.org/10.1073/pnas.071022298>.
- Shirozu R, Yashiroda H, Murata S. 2015. Identification of minimum Rpn4-responsive elements in genes related to proteasome functions. *FEBS Lett* 589:933–940. <https://doi.org/10.1016/j.febslet.2015.02.025>.
- Guerra-Moreno A, Isasa M, Bhanu MK, Waterman DP, Eapen VV, Gygi SP, Hanna J. 2015. Proteomic analysis identifies ribosome reduction as an effective proteotoxic stress response. *J Biol Chem* 290:29695–29706. <https://doi.org/10.1074/jbc.M115.684969>.
- Bernales S, Papa FR, Walter P. 2006. Intracellular signaling by the unfolded protein response. *Annu Rev Cell Dev Biol* 22:487–508. <https://doi.org/10.1146/annurev.cellbio.21.122303.120200>.
- Haugen AC, Kelley R, Collins JB, Tucker CJ, Deng C, Afshari CA, Brown JM, Ideker T, van Houten B. 2004. Integrating phenotypic and expression profiles to map arsenic-response networks. *Genome Biol* 5:R95. <https://doi.org/10.1186/gb-2004-5-12-r95>.
- Hosiner D, Lempiäinen H, Reiter W, Urban J, Loewith R, Ammerer G, Schweyen R, Shore D, Schüller C. 2009. Arsenic toxicity to *Saccharomyces cerevisiae* is a consequence of inhibition of the TORC1 kinase combined with a chronic stress response. *Mol Biol Cell* 20:1048–1057. <https://doi.org/10.1091/mbc.e08-04-0438>.
- Steffen KK, McCormick MA, Pham KM, MacKay VL, Delaney JR, Murakami CJ, Kaeblerlein M, Kennedy BK. 2012. Ribosome deficiency protects against ER stress in *Saccharomyces cerevisiae*. *Genetics* 191:107–118. <https://doi.org/10.1534/genetics.111.136549>.
- Mori K, Kawahara T, Yoshida H, Yanagi H, Yura T. 1996. Signalling from endoplasmic reticulum to nucleus: transcription factor with a basic-leucine zipper motif is required for the unfolded protein-response pathway. *Genes Cells* 1:803–817. <https://doi.org/10.1046/j.1365-2443.1996.d01-274.x>.
- Cox JS, Walter P. 1996. A novel mechanism for regulating activity of a transcription factor that controls the unfolded protein response. *Cell* 87:391–404. [https://doi.org/10.1016/S0092-8674\(00\)81360-4](https://doi.org/10.1016/S0092-8674(00)81360-4).
- Mulder HJ, Saloheimo M, Penttilä M, Madrid SM. 2004. The transcription factor HACA mediates the unfolded protein response in *Aspergillus niger*, and up-regulates its own transcription. *Mol Genet Genomics* 271:130–140. <https://doi.org/10.1007/s00438-003-0965-5>.
- Mori K, Ogawa N, Kawahara T, Yanagi H, Yura T. 2000. mRNA splicing-mediated C-terminal replacement of transcription factor Hac1p is required for efficient activation of the unfolded protein response. *Proc Natl Acad Sci U S A* 97:4660–4665. <https://doi.org/10.1073/pnas.050010197>.
- Pincus D, Aranda-Díaz A, Zuleta IA, Walter P, El-Samad H. 2014.

- Delayed Ras/PKA signaling augments the unfolded protein response. *Proc Natl Acad Sci U S A* 111:14800–14805. <https://doi.org/10.1073/pnas.1409588111>.
31. Jørgensen TR, Goosen T, van den Hondel CAMJJ, Ram AFJ, Iversen JLL. 2009. Transcriptomic comparison of *Aspergillus niger* growing on two different sugars reveals coordinated regulation of the secretory pathway. *BMC Genomics* 10:44. <https://doi.org/10.1186/1471-2164-10-44>.
 32. Lu X, Sun J, Nimtz M, Wissing J, Zeng AP, Rinas U. 2010. The intra- and extracellular proteome of *Aspergillus niger* growing on defined medium with xylose or maltose as carbon substrate. *Microb Cell Fact* 9:23. <https://doi.org/10.1186/1475-2859-9-23>.
 33. Twumasi-Boateng K, Yu Y, Chen D, Gravelat FN, Nierman WC, Sheppard DC. 2009. Transcriptional profiling identifies a role for *BrlA* in the response to nitrogen depletion and for *StuA* in the regulation of secondary metabolite clusters in *Aspergillus fumigatus*. *Eukaryot Cell* 8:104–115. <https://doi.org/10.1128/EC.00265-08>.
 34. Wang F, Krijgheld P, Hulsman M, de Bekker C, Müller WH, Reinders M, de Vries RP, Wösten HAB. 2015. *FluG* affects secretion in colonies of *Aspergillus niger*. *Antonie Van Leeuwenhoek* 107:225–240. <https://doi.org/10.1007/s10482-014-0321-2>.
 35. Park J, Hulsman M, Arentshorst M, Breeman M, Alazi E, Legendijk EL, Rocha MC, Malavazi I, Nitsche BM, van den Hondel CAMJJ, Meyer V, Ram AFJ. 2016. Transcriptomic and molecular genetic analysis of the cell wall salvage response of *Aspergillus niger* to the absence of galactofuranose synthesis. *Cell Microbiol* 18:1268–1284. <https://doi.org/10.1111/cmi.12624>.
 36. de Vries RP, Burgers K, van de Vondervoort PJJ, Frisvad JC, Samson RA, Visser J. 2004. A new black *Aspergillus* species, *A. vadensis*, is a promising host for homologous and heterologous protein production. *Appl Environ Microbiol* 70:3954–3959. <https://doi.org/10.1128/AEM.70.7.3954-3959.2004>.
 37. Kusters-van Someren MA, Harmsen JA, Kester HC, Visser J. 1991. Structure of the *Aspergillus niger pelA* gene and its expression in *Aspergillus niger* and *Aspergillus nidulans*. *Curr Genet* 20:293–299. <https://doi.org/10.1007/BF00318518>.
 38. Wösten HAB, Moukha SM, Sietsma JH, Wessels JGH. 1991. Localization of growth and secretion of proteins in *Aspergillus niger*. *J Gen Microbiol* 137:2017–2023. <https://doi.org/10.1099/00221287-137-8-2017>.
 39. Punt PJ, Oliver RP, Dingemans MA, Pouwels PH, van den Hondel CAMJJ. 1987. Transformation of *Aspergillus* based on the hygromycin B resistance marker from *Escherichia coli*. *Gene* 56:117–124. [https://doi.org/10.1016/0378-1119\(87\)90164-8](https://doi.org/10.1016/0378-1119(87)90164-8).
 40. Arentshorst M, Jing N, Ram AFJ. 2015. Efficient generation of *Aspergillus niger* knock out strains by combining NHEJ mutants and a split marker approach, p 263–272. *In* van den Berg MA, Maruthachalam K (ed), Genetic transformation systems in fungi, vol 1. Springer, Basel, Switzerland.
 41. de Bekker C, Wiebenga A, Aguilar G, Wösten HAB. 2009. An enzyme cocktail for efficient protoplast formation in *Aspergillus niger*. *J Microbiol Methods* 76:305–306. <https://doi.org/10.1016/j.mimet.2008.11.001>.
 42. Andersen MR, Salazar MP, Schaap PJ, van de Vondervoort PJ, Culley D, Thykaer J, Frisvad JC, Nielsen KF, Albang R, Albermann K, Berka RM, Braus GH, Braus-Stromeyer SA, Corrochano LM, Dai Z, van Dijk PW, Hofmann G, Lasure LL, Magnuson JK, Menke H, Meijer M, Meijer SL, Nielsen JB, Nielsen ML, van Ooyen AJ, Pel HJ, Poulsen L, Samson RA, Stam H, Tsang A, van den Brink JM, Atkins A, Aerts A, Shapiro H, Pangilinan J, Salamov A, Lou Y, Lindquist E, Lucas S, Grimwood J, Grigoriev IV, Kubicek CP, Martinez D, van Peij NN, Roubos JA, Nielsen J, Baker SE. 2011. Comparative genomics of citric-acid-producing *Aspergillus niger* ATCC 1015 versus enzyme-producing CBS 513.88. *Genome Res* 21:885–897. <https://doi.org/10.1101/gr.112169.110>.
 43. Grigoriev IV, Nikitin R, Haridas S, Kuo A, Ohm R, Otilar R, Riley R, Salamov A, Zhao X, Korzeniewski F, Smirnova T, Nordberg H, Dubchak I, Shabalov I. 2014. MycoCosm portal: gearing up for 1000 fungal genomes. *Nucleic Acids Res* 42:D699–D704. <https://doi.org/10.1093/nar/gkt1183>.
 44. Kim D, Langmead B, Salzberg SL. 2015. HISAT: a fast spliced aligner with low memory requirements. *Nat Methods* 12:357–360. <https://doi.org/10.1038/nmeth.3317>.
 45. Nakamura S, Sato H, Tanaka R, Yaguchi T. 2016. Verification of ribosomal proteins of *Aspergillus fumigatus* for use as biomarkers in MALDI-TOF MS identification. *Mass Spectrom (Tokyo)* 5:A0049. <https://doi.org/10.5702/massspectrometry.A0049>.
 46. Trapnell C, Williams BA, Pertea G, Mortazavi A, Kwan G, van Baren MJ, Salzberg SL, Wold BJ, Pachter L. 2010. Transcript assembly and quantification by RNA-Seq reveals unannotated transcripts and isoform switching during cell differentiation. *Nat Biotechnol* 28:511–515. <https://doi.org/10.1038/nbt.1621>.
 47. Roberts A, Trapnell C, Donaghey J, Rinn JL, Pachter L. 2011. Improving RNA-Seq expression estimates by correcting for fragment bias. *Genome Biol* 12:R22. <https://doi.org/10.1186/gb-2011-12-3-r22>.
 48. Goff L, Trapnell C, Kelley D. 2013. CummeRbund: analysis, exploration, manipulation, and visualization of Cufflinks high-throughput sequencing data. R package version 2.16.0.
 49. Schröder MS, Gusenleitner D, Quackenbush J, Culhane AC, Haibe-Kains B. 2013. RamiGO: an R/Bioconductor package providing an AmiGO visualize interface. *Bioinformatics* 29:666–668. <https://doi.org/10.1093/bioinformatics/bts708>.
 50. Mellacheruvu D, Wright Z, Couzens AL, Lambert JP, St-Denis NA, Li T, Miteva YV, Hauri S, Sardi ME, Low TY, Halim VA, Bagshaw RD, Hubner NC, Al-Hakim A, Bouchard A, Faubert D, Fermin D, Dunham WH, Goudreault M, Lin ZY, Badillo BG, Pawson T, Durocher D, Coulombe B, Aebersold R, Superti-Furga G, Colinge J, Heck AJ, Choi H, Gstaiger M, Mohammed S, Cristea IM, Bennett KL, Washburn MP, Raught B, Ewing RM, Gingras AC, Nesvizhskii AI. 2013. The CRAPome: a contaminant repository for affinity purification-mass spectrometry data. *Nat Methods* 10:730–736. <https://doi.org/10.1038/nmeth.2557>.
 51. Teo G, Liu G, Zhang J, Nesvizhskii AI, Gingras AC, Choi H. 2014. SAINTexpress: improvements and additional features in Significance Analysis of INTERactome software. *J Proteomics* 100:37–43. <https://doi.org/10.1016/j.jprot.2013.10.023>.
 52. Petersen TN, Brunak S, von Heijne G, Nielsen H. 2011. SignalP 4.0: discriminating signal peptides from transmembrane regions. *Nat Methods* 8:785–786. <https://doi.org/10.1038/nmeth.1701>.
 53. van Munster JM, Dobruchowska JM, Veloo R, Dijkhuizen L, van der Maarel MJEC. 2015. Characterization of the starvation-induced chitinase CfcA and α -1,3-glucanase AgnB of *Aspergillus niger*. *Appl Microbiol Biotechnol* 99:2209–2223. <https://doi.org/10.1007/s00253-014-6062-3>.
 54. van Munster JM, Nitsche BM, Akeroyd M, Dijkhuizen L, van der Maarel MJEC, Ram AFJ. 2015. Systems approaches to predict the functions of glycoside hydrolases during the life cycle of *Aspergillus niger* using developmental mutants $\Delta brlA$ and $\Delta flbA$. *PLoS One* 10:e0116269. <https://doi.org/10.1371/journal.pone.0116269>.
 55. Henry C, Fontaine T, Heddergott C, Robinet P, Aïmanianda V, Beau R, Beauvais A, Mouyna I, Prevost MC, Fekkar A, Zhao Y, Perlin D, Latgé JP. 2016. Biosynthesis of cell wall mannan in the conidium and the mycelium of *Aspergillus fumigatus*. *Cell Microbiol* 18:1881–1891. <https://doi.org/10.1111/cmi.12665>.