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## **A novel formulation for skin barrier repair : from ex vivo assessment towards clinical studies**

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The background of the page is an abstract composition of numerous overlapping, wavy lines. These lines originate from the bottom left and flow towards the top right, creating a sense of movement and depth. The color palette is primarily composed of various shades of blue, ranging from light sky blue to deep navy blue, with some lines in a muted purple or lavender hue. The lines vary in thickness and opacity, with some appearing as thin, delicate strokes and others as thick, bold bands. The overall effect is a dynamic and textured visual field.

# Part 3

# Chapter 7

## **Applying a vernix caseosa based formulation accelerates skin barrier repair by modulating lipid biosynthesis**

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## Abbreviations

AD	Atopic dermatitis
ATR-FTIR	Attenuated total reflection Fourier transform infrared spectroscopy
AUC	Area under the barrier recovery curve
C34	Ceramide with 34 carbon atoms
FWHM	Full width at half maximum
LMMs	Linear mixed models
PCA	Principal component analysis
PC	Principal component
SAXD	Small angle X-ray diffraction
SC	Stratum corneum
SQ	SquameScan value
TEWL	Transepidermal water loss
VC	Vernix caseosa

## Keywords

Epidermis, clinical study, mass spectrometry, lipidomics, sphingolipids, lipid organization, vernix caseosa, barrier recovery, stratum corneum, treatment

## Abstract

Restoring the lipid homeostasis of the stratum corneum (SC) is a common strategy to enhance the skin barrier function. Here, we used a ceramide containing vernix caseosa (VC) based formulation and were able to accelerate barrier recovery in healthy volunteers. The recovery was examined over 16 days by monitoring trans-epidermal water loss (TEWL) after barrier disruption by tape-stripping. Four skin sites were used to examine the effects of both treatment and barrier recovery. After 16 days, samples were harvested at these sites to examine the SC ceramide composition and the lipid organization. Changes in ceramide profiles were identified using principal component analysis (PCA). After barrier recovery, the untreated sites showed increased levels of ceramide subclass AS and ceramides with a 34 total carbon atom chain length, while the mean ceramide chain length was reduced. These changes were diminished by treatment with the studied formulation, which concurrently increased the formulated ceramides. Correlations were observed between SC lipid composition, lipid organization, and TEWL, and changes in the ceramide subclass composition suggesting changes in the ceramide biosynthesis. These results suggest that VC based formulations enhance skin barrier recovery and are attractive candidates to treat skin disorders with impaired barrier properties.

## Introduction

Vital functions of the stratum corneum (SC) are preventing excessive water loss from the body and protecting the body from the hostile outside environment by acting as an inside-out and an outside-in barrier, respectively. The SC is generated by a dynamic process in which keratinocytes in the lower layer of the epidermis undergo terminal differentiation and transform into corneocytes.<sup>1</sup> During this differentiation process, lipids are synthesized, stored in lamellar bodies and, at the interface between the viable epidermis and SC, these lamellar bodies are secreted in the intercellular regions.<sup>2</sup> The secreted lipids form an extracellular lipid matrix crucial for a proper skin barrier.<sup>3,4</sup> Three main lipid classes have been identified: cholesterol, fatty acids, and ceramides. The latter constitutes the group of SC lipids that have the most diverse chemical structure. Their building blocks are an array of sphingoid bases and acyl chains chemically linked together by an amide bond, resulting in 16 ceramide subclasses.<sup>5</sup> Both chains (the sphingoid base and acyl chain) can vary substantially in carbon chain length. The composition of the SC ceramide fraction (the ceramide profile) was correlated with and shown to be important for lipid organization and skin barrier function.<sup>6-8</sup> In the present study, the effect of a ceramide containing formulation on skin barrier recovery, the ceramide composition, and lipid organization were studied.

Through the processes of cornification and desquamation, the SC is continuously rejuvenated<sup>9,10</sup> and can thereby quickly recover the barrier function after disruption.<sup>11</sup> An impaired skin barrier function is encountered in several inflammatory skin diseases; one of which is atopic dermatitis (AD). In AD, an inadequate recovery of the impaired skin barrier perpetuates the condition. Normalization of the barrier function in AD is crucial to reduce the penetration of irritants and allergens into the skin. Application of lipophilic formulations that act as an additional barrier is a common treatment.<sup>12-15</sup> Another approach aims at normalization of the ceramide composition.<sup>4</sup> Changes in this composition were correlated to the reduced skin barrier function in AD.<sup>8,16,17</sup> Previously, ceramide containing formulations have been used to study skin barrier repair<sup>18-21</sup> and it was reported that the ratios between the three main SC lipids in a formulation was important for short term recovery.<sup>22</sup> Furthermore, it was suggested that long term application could enhance barrier recovery.<sup>21</sup> Nonetheless, it is disputed whether ceramides in a formulation actually normalize the SC lipid composition or remain on the skin surface.<sup>23</sup> It has been shown that the orally administered essential fatty acids were incorporated in the epidermis.<sup>24</sup>

How these formulations accelerate barrier recovery is not clearly understood. Therefore, the primary aim of this study was to examine how treatment with a formulation during barrier recovery affects the SC lipid composition and subsequently lipid organization. This was done by examining the normal physiological responses in healthy volunteers. In this study, we applied a ceramide containing formulation based on the composition of the vernix caseosa (VC). This is a natural ceramide containing formulation that serves as a protective lipid film covering the fetus in the last trimester of pregnancy and during delivery.<sup>25</sup> Previously, it was shown that VC like formulations were able to significantly accelerate barrier recovery in mice.<sup>26,27</sup> This vernix formulation has the advantage that ceramides can be formulated. The formulation is semi-occlusive.<sup>28</sup> To examine the effect of the formulation on barrier recovery, we used barrier disruption

by tape-stripping, as this is an effective way to remove the barrier and induce barrier recovery in healthy skin.<sup>27,29-32</sup>

Here, it is shown that treatment with the formulation resulted in accelerated barrier recovery of tape-stripped skin compared to the untreated tape-stripped skin. In regenerated untreated SC, an altered ceramide composition was observed. These alterations correlated to a decrease in skin barrier function. When applying the ceramide containing formulation, the ceramide composition was modulated towards a ceramide composition more closely mimicking that of control skin. Combined, these results indicate that barrier disruption induces changes in lipid processing during recovery, related to a decreased barrier function and that these changes in lipid processing can be modulated by application of the VC formulation.

## Materials and methods

### Chemicals

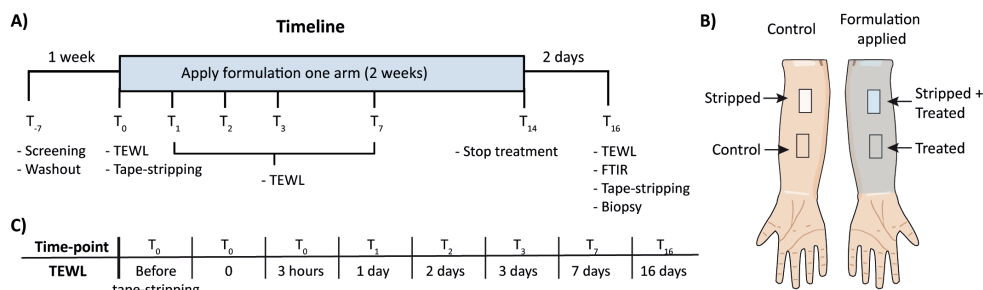
The following solvents were used for lipid extraction and analysis: Millipore water (18.2 mΩ), HPLC grade methanol and chloroform from Lab-Scan (Gliwice, Poland), UPLC grade isopropanol and ethanol from Biosolve (Valkenswaard, the Netherlands), and HPLC grade heptane from Actu-All (Oss, the Netherlands). The synthetic ceramides listed in Table S1 were used as calibrators for the LC/MS analysis. Trypsin and trypsin inhibitor for SC isolation were purchased from Sigma-Aldrich (Zwijndrecht, the Netherlands).

### Clinical study design

The clinical study was approved by the Leiden University Medical Center ethical committee and performed according to the declaration of Helsinki. All volunteers signed an informed consent. 8 female and 7 male Caucasian volunteers, age 18 to 29 years (mean age 23 years) were recruited and checked by a dermatologist. Volunteers were excluded if they had dermatological disorders or a history of dermatological disorders, suffered from chronically inflammatory diseases, used systemic drug therapies, had abundant hair or unnatural abnormalities on the ventral forearms, had a history of drug abuse, or if they were pregnant. All enrolled volunteers completed the clinical study. Figure 1A depicts the timeline of the study. Before the study commenced, volunteers had a one week washout period in which no soaps or cosmetics were applied on the ventral forearms. At  $T_0$  a site of 3.5 by 2.5 cm of SC was removed at both ventral forearms by consecutive tape-stripping using DSquame tape (Cuderm, Dallas, USA). The tape-stripping continued until the site appeared shiny<sup>33</sup> and a transepidermal water loss (TEWL) of  $> 60$  g/m<sup>2</sup>/h was reached (these sites are further referred to as stripped). TEWL was measured using AquaFlux AF200 (BIOX, London, UK). Thereafter, two fingertip units of a VC like formulation (composition is shown in Table S2) were applied twice daily for 14 days on one ventral forearm (referred to as treated), followed by a two day washout period. Treatment allocation to either left or right arm was alternated between volunteers. Figure 1B depicts the 4 sites investigated in this study: control



(no treatment, no tape-stripping), treated (treated with formulation), stripped (tape-stripped at  $T_0$ ), and stripped+treated (tape-stripped at  $T_0$  and treated with formulation). At the final time point ( $T_{16}$ ) 21 tape-strips, using polyphenylene sulfide tape (Nichiban, Tokyo, Japan), were harvested at all 4 sites. The amount of SC on each tape-strip was determined by measuring the tape-strips with a SquameScan (Heiland Electronic, Wetzlar, Germany). Tape-strip zero was discarded. After every other tape-strip an infrared spectrum of the tape-stripped skin region was made (see below). After tape-stripping, a 4 mm biopsy of both recovered sites was collected, near the site where Nichiban tape-strips were harvested, to perform small angle X-ray diffraction (SAXD), see below. During the study, TEWL of all sites was monitored at distinct time points, these measurements were performed before formulation application.



**Figure 1. Study design.** **A)** Timeline of the clinical study (Time points are indicated as T with days as subscripts). Volunteers were first screened by a dermatologist ( $T_{-7}$ ), followed by a one week washout period. At  $T_0$  the SC was removed by tape-stripping. The formulation was applied 2x daily for 2 weeks followed by two days washout. The activities and measurements are indicated at each time point ( $T_{-7}$  till  $T_{16}$ ). At all indicated time-points TEWL was measured, except for  $T_{-7}$ . **B)** The studied sites on the ventral forearms. On both arms one site was tape-stripped; on one arm formulation was applied. **C)** The time-points at which TEWL was measured to monitor barrier recovery of the stripped sites.

## TEWL measurements to determine barrier recovery

To determine barrier recovery of the stripped sites TEWL was monitored. At each time point depicted in Figure 1C, TEWL was measured at three different sections of each site (Figure S1). The barrier recovery percentage was calculated by defining the average of three TEWL measurements at the non-stripped sites as 100% barrier and TEWL at the stripped sites directly after tape-stripping as 0% barrier. The following equation was used:<sup>26</sup>

Recovery%  $T_n = (\text{TEWL } T_0 - \text{TEWL } T_n) / (\text{TEWL } T_0 - \text{Average TEWL non-stripped site } T_n) * 100$

For each volunteer several values were calculated for both the treated and untreated stripped sites: I) the recovery percentage of the site at three sections at each time-point, II) the mean of the three previous values, III) the area under the barrier recovery curve (AUC) of the means over time, using a point-to-point linear fit, IV) using the same fit, the time it took to reach designated recovery percentages between 5-100%, with increment of 5%, V) the difference in time between the stripped and stripped+treated sites at each of the previously mentioned recovery %. Recovery percentages >90% were excluded, as only 5 volunteers reached these values at both stripped sites, biasing these data.

## Quantification of ceramides by LC/MS

Tape-stripped SC can be used to examine the lipid content by mass spectrometry.<sup>34-36</sup> Samples for ceramide analysis by LC/MS were prepared from tape-strips 5-8 and 17-20, harvested at the 4 sites on T<sub>16</sub>. Samples prepared from tape-strips at a depth of 5-8 and 17-20 tapes were analyzed separately to determine if the SC ceramide composition generated in the initial stage of the recovery process (tapes 5-8) was different from that generated in later stages of the recovery process (tapes 17-20). Tape-strips were punched to an area of 2 cm<sup>2</sup>, extracted, analyzed and quantified as described in Boiten et al.<sup>34</sup> Shortly, a modified 4 step Bligh and Dyer extraction at 40°C was performed. Extracts were analyzed using an Acquity UPLC H-class (Waters, Milford, MA, USA) connected to an XEVO TQ-S mass spectrometer (Waters, Milford, MA, USA). Separation was performed on a pva-silica column (5 µm particles, 100 × 2.1 mm i.d.) (YMC, Kyoto, Japan). Using the calibrators, a response model was build and used to quantify the ceramides in the analyzed samples. All data were corrected for the cumulative SquameScan value (SQ) of the four tape-strips comprising the corresponding sample. The data resulted in quantitative SC ceramide profiles for each volunteer, at all 4 sites, and at two depths per site. Ceramides are named according to hydrophilic head group (subclass) and chain length (e.g. NS C42, ceramide subclass NS with a total chain length of 42 carbon atoms).<sup>37</sup> For an overview of subclasses and ceramide structures see Figure S2.

Quantitative data were used to calculate several parameters. Ceramides EOS C66 and NS C40 (supplied in the formulation) were excluded from these calculations. The following parameters were calculated:

- I) Total amount of the individual subclasses (ng/SQ);
- II) Total amount of ceramides with a total chain length 34 carbons (ng/SQ);
- III) Mean carbon chain length: ( $\Sigma$  each ceramide (molar amount ceramide × carbon chain length))/total molar amount all ceramides;
- IV) Percentage of EO ceramides, as ng EO / total ng ceramides.

## Lateral organization and conformational ordering of SC lipids

The lateral organization and conformational ordering of the SC lipids were examined by attenuated total reflection Fourier transform infrared spectroscopy (ATR-FTIR). FTIR spectra were collected using a Varian 670-IR spectrometer (Agilent Technologies, Inc., Santa Clara, CA) equipped with a mercury-cadmium-telluride detector and an external sample compartment containing an ATR accessory (GladiATR, PIKE Technologies, Maddison, WI) with a single reflection diamond. The sample compartment was constantly purged with dry air. Each spectrum was an average of 150 scans, with a spectral resolution of 2 cm<sup>-1</sup>. Conformational ordering of the lipids was determined using the center of gravity of the peak of the CH<sub>2</sub> symmetric stretching vibrations at 90% of the peak height. The bandwidth from the second derivative of the CH<sub>2</sub> scissoring region was selected as a measure for lateral packing of the lipids and determined as described before.<sup>38,39</sup> CH<sub>2</sub> stretching vibration peak position values <2850 cm<sup>-1</sup> indicate an ordered phase, whereas values above 2852 cm<sup>-1</sup> indicate a high degree of disordering being a liquid phase. The second derivative of the CH<sub>2</sub> scissoring region

was baseline-corrected between the endpoints of the scissoring region (~1460-1480  $\text{cm}^{-1}$ ) and the full width at half maximum (FWHM) was calculated.<sup>39</sup> Values of 5 spectra (tape 2-10 and tape 12-20) per site per volunteer were used for the analyses.

## Small angle X-ray diffraction

SC isolated from biopsies harvested at stripped and stripped+treated sites were examined by SAXD. SAXD was used to determine the SC lipids lamellar organization. To isolate SC for SAXD analysis, biopsies were placed overnight in a 0.1% trypsin solution in PBS at 4°C, followed by one hour incubation at 37°C. The SC was peeled of and washed in 0.1 % trypsin inhibitor solution in PBS and twice in Millipore water. Dried SC sheets were stored under argon and over silica gel until use. All SAXD measurements were performed at the European Synchrotron Radiation Facility (Grenoble, France) at station BM26B. SAXD patterns were collected on a Pilatus 1M detector at room temperature for 5 minutes. The sample-to-detector distance was 2 m. Prior to data collection, the SC samples were hydrated over a 27% (w/v) NaBr solution for 24 hours at room temperature. The scattering intensity  $I$  (arbitrary units) was measured as a function of the scattering vector  $q$  (in  $\text{nm}^{-1}$ ), defined as:  $q = 4\pi \sin\theta/\lambda$ , in which  $\theta$  is the scattering angle and  $\lambda$  is the wavelength. From the position of the main peak (referred to as peak 2 with position  $q_2$ ), the spacing ( $d$ ) of the lipid lamellae was calculated using  $d = 2\pi/q_2$ . As in SC this peak position is determined by the structure of the two lamellar phases, a change in the spacing indicates a change in these lamellar phases.<sup>8</sup>

## Software and statistical analysis

All LC/MS data were processed using MassLynx and TargetLynx software (V4.1 SCN 843 Waters Inc.). FTIR data were processed and analyzed using Resolutions Pro 4.1 (Agilent Technologies, Inc.). Principal component analysis (PCA) is a commonly used analysis of lipidomics data sets.<sup>40,41</sup> It is a mathematical dimensional reduction that can effectively visualize variation and the difference between samples in large data sets by reducing all variables to principal components (PCs). PCA was performed using Multibase add-in for excel (NumericalDynamics.Com). For PCA, the quantitative data obtained from the ceramide analyses were normalized to the total amount of ceramides in the samples. The four individual sites (control, treated, stripped, and stripped+treated) of each volunteer were defined as PCA samples. All quantified ceramides at both depths were defined as variables of that "PCA sample" (total of 584 variables), thereafter the mean of each ceramide/variable in all "PCA samples" was centered at zero (subtracting the mean).

Correlation analysis and point-to-points fits were performed using GraphPad Prism (V6.05, Graphpad). Group-wise comparisons were performed using linear mixed models (LLMs) in SPSS (V24 IBM). All measurements within the same subject were treated as repeated measurement. LMMs were used because these models have the advantage that I) the effect size of individual variables and their interactions can be examined, and II) they can handle missing data.<sup>42</sup> Figure S3 explains how the LMMs were used here. All interactions between fixed variables were included in the first run of a model and when non-significant ( $p>0.05$ ) taken out one by one. Significance levels

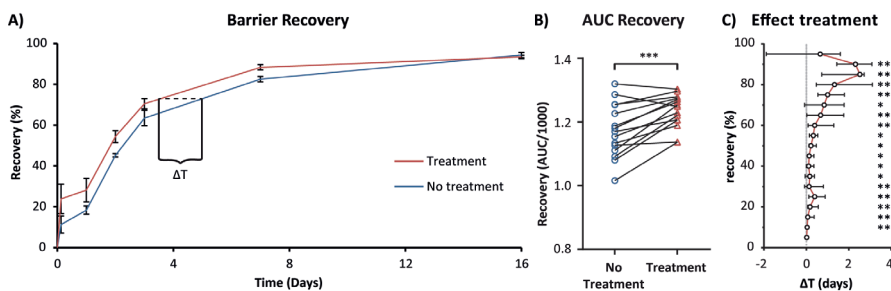
are indicated as follows: \* ( $p < 0.05$ ), \*\* ( $p < 0.01$ ), \*\*\* ( $p < 0.005$ ) and \*\*\*\* ( $p < 0.001$ ).

## Results

### Barrier recovery in healthy volunteers

To determine the effect of the formulation on barrier recovery, changes in skin barrier over time (measured as TEWL and expressed as recovery %) were examined. Figure 2A shows a typical example of barrier recovery % as function of time for untreated and treated stripped sites of one volunteer. After 16 days, all volunteers had reached a recovery %  $> 85$  at both sites. For each volunteer the AUC was calculated of both the untreated and treated stripped sites (Figure 2B). A larger AUC implies less barrier dysfunction during the 16 days. Treatment with the formulation significantly increased the AUC, indicating increased recovery.

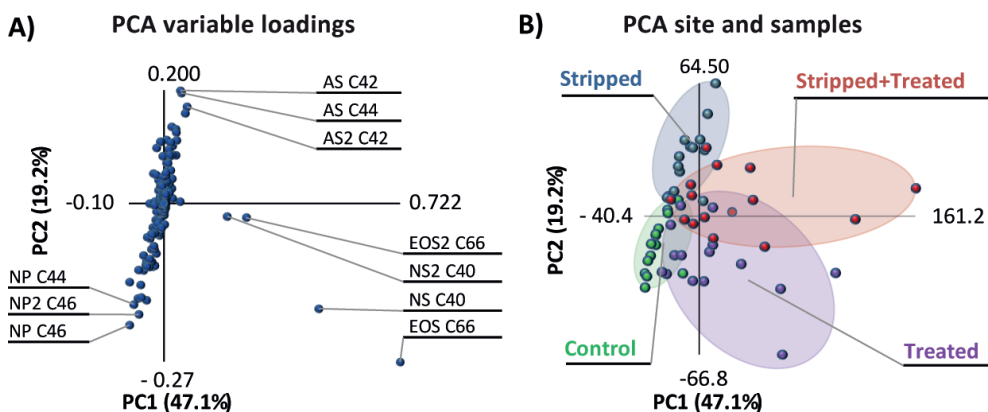
Yet, the AUC does not take into account the kinetics of the recovery process. To determine if treatment accelerated barrier recovery, differences in time between untreated and treated stripped sites were examined at multiple designated recovery percentages ( $\Delta T$  in Figure 2A). Figure 2C depicts the median and interquartile range of  $\Delta T$  at different recovery percentages for all volunteers. A median of zero would indicate no change in recovery time due to treatment. A positive deviation from zero was observed at almost all recovery percentages with a spike at 25%, and a steady increase above 50%. Treatment significantly accelerated barrier recovery ending with a more than two days faster median recovery. As control both the AUC and the  $\Delta T$  were tested for correlation to the TEWL directly after tape-stripping (Table S3). No correlations were observed, indicating that these parameters were not dependent on the amount of barrier disruption.



**Figure 2. The effect of treatment on barrier recovery.** **A)** The typical elapse of barrier recovery over time for untreated and treated stripped sites, for a single volunteer. Points are means ( $\pm$ SD) of three measurements within one site. Lines are point-to-point connections. The dotted line indicated the difference in time ( $\Delta T$ ) between the untreated and treated curve at a designated recovery %. **B)** The AUCs/1000 for all volunteers. Lines connect data points of the same volunteer. A paired t-test showed significant increase at the treated sites. **C)** The  $\Delta T$  for all volunteers at recovery percentages between 5 and 90 %. The data were non-normally distributed and are shown as median and interquartile range. Deviations from zero were tested with a Wilcoxon signed-rank test.

## Principle component analysis of ceramide profiles

To gain insight into differences between the ceramide profiles of the four different sites PCA was performed on the quantitative ceramide data. Figure 3A depicts the variable loadings, indicating how strongly each variable (ceramides detected in the samples) correlated to PC1 and PC2. If the value is higher (deviates from center of the graph in x or y position) it has contributed more to the variation explained by PC1 or PC2, respectively. The 10 variables with the highest correlation coefficients are indicated in the plot. PC1 and PC2 explained a total inertia (variance) of 66.3%. Figure 3B shows the plot of the “PCA samples”, depicting the PC1 and PC2 scores of each “PCA sample”. Samples were assigned to clusters corresponding to the 4 sites indicated by different colors. Separation between clusters was observed demonstrating that variation between the ceramide profiles corresponded to variation between samples of different sites. Therefore, changes in ceramide profile due to treatment and tape-stripping can be explained by the variable loadings of the PCs.



**Figure 3. PCA of all SC ceramides and the amount of ceramides NS C40 and EOS C66.** **A)** The PCA loading plot. Inertia per PC is indicated as % variation. Variables are all ceramides quantified in samples obtained from tape-strips 5-8 and samples obtained from tape-strips 17-20, the latter are indicated with a 2 after the subclass name. **B)** A plot of the PCA samples (the 4 sites of all volunteers). The samples were clustered per site.

Clusters of treated sites were separated from untreated sites as depicted in Figure 3B. By examining plot 3A it shows that this was due to 4 variables primarily. These variables corresponded to the two formulated ceramides (NS C40 and EOS C66), detected in samples of both depths. The PCA demonstrated that a large part of the variation in the ceramide profiles was due to the two ceramides in the formulation. The increase, due to formulation application, of these naturally occurring ceramides was examined. Figure S4 shows that the amount at treated sites was increased at both tape-stripping depths. An LMM of the logarithm of these amounts confirmed these observations (Table S4). Next, separation of clusters corresponding to stripped and non-stripped sites was investigated. Variables correlating strongly to PC2 were primarily responsible for the observed separation. (Figure S5 shows a complete graphic overview of the PC2 loadings). When examining the loadings of PC2, highly positive correlations were

observed for ceramides of subclass AS and, to a lesser extent, ceramides for subclass NS. Highly negative correlations were observed for ceramides from subclass NP. Changes in ceramide profile were observed between the different sites. In the next section, the results of the PCA were used as leads to investigate if these changes correlated to changes in barrier function and the recovery process.

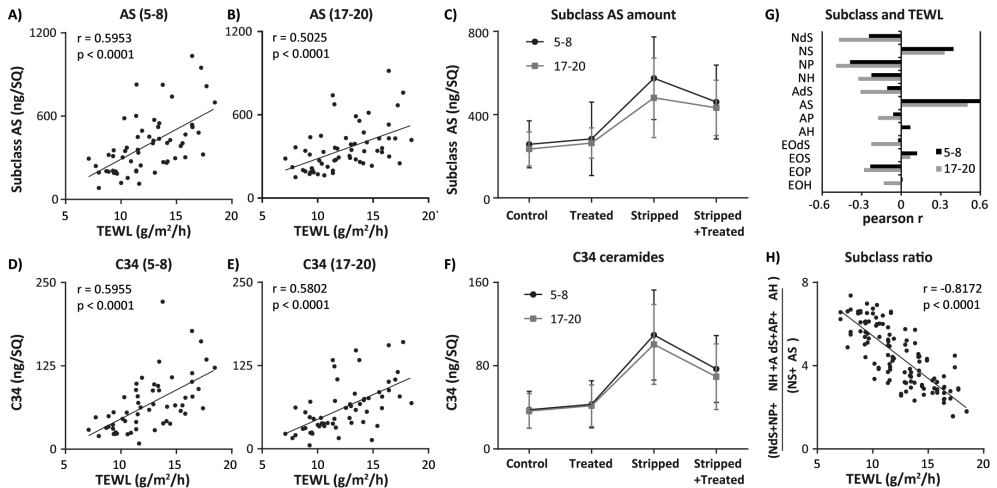
### Correlations between ceramide profile and barrier function

To investigate if barrier function correlated to changes in SC ceramide profile, changes in ceramides AS and NS, identified by PCA, were correlated to the TEWL at T<sub>16</sub> (Figure S6 shows the TEWL at T<sub>16</sub>). Figure 4A and B depict these correlations for the total amount of subclass AS. At both depths, a significant positive correlation between the amount of subclass AS and TEWL was observed. This illustrates that an increase in subclass AS correlated with a reduced skin barrier function. Next, it was examined whether stripping, treatment, SC depth, or the interaction between these conditions had an effect on the amount of subclass AS using a LMM. The model used stripped, treated, and depth as fixed factors, and subject as random factor. Table 1 depicts the effect sizes obtained with the model. Figure 4C depicts the mean ( $\pm$ SD) of the amount of subclass AS measured at all 4 sites at both depths. Using the model it was determined that subclass AS increased significantly at stripped sites. Treatment or depth alone showed no significant effects compared to the control. However, at the stripped site treatment did have a significant effect. The interaction stripped\*treated in Table 1 illustrates this effect. When subjected to stripping and treatment the amount of AS changed by the effect sizes of stripped and treatment, in addition to both effects the interaction term stripped\*treated is added. At stripped+treated sites treatment significantly reduced the increase in AS observed due to stripping with 81.2 ng/SQ (treatment + stripped\*treated). Similar results were obtained when examining NS C44 (Figure S7 and Table 1). It was decided not to use the total amount of subclass NS, as this value was influenced by ceramide NS C40 of the formulation. NS C44 is most abundantly present of the NS ceramide subclass members and chosen as a proper representative of subclass NS.

Besides subclasses, ceramide chain lengths, especially ceramides with a total carbon chain length of 34 carbons (C34), are known to correlate to barrier dysfunction.<sup>17</sup> Here, a significant positive correlation was observed between the total amount of C34 ceramides (summation of amounts of NdS, NS, NP, NH, AdS, AS, AP, and AH with a total of 34 carbons) and TEWL, at both depths (Figure 4D and E), demonstrating that higher levels of C34 ceramides correlate with reduced barrier function at day 16. A similar LMM as that of subclass AS and NS C44 was built for C34 ceramides (Table 1). The total amount of C34 ceramides at the 4 sites at two depths was plotted as the mean  $\pm$ SD in Figure 4F. By using the model a significant increase at the stripped sites was observed. Comparable to subclass AS, treatment only significantly affected the stripped+treated sites. Treatment reduced the increase in C34 ceramides with 31.6 ng/SQ compared to stripped sites.

Next, the correlation of each subclass to TEWL was examined (Figure 4G: Pearson r for each subclass). This correlation underlined the observations with PCA, where subclass NS and AS had a positive correlation and subclass NP a negative correlation to PC2. Both subclasses NS and AS have a distinct synthesis route different from

subclasses NP and AP, this suggested a ratio dividing the subclasses as follows:  $(NdS+NP+NH+AdS+AP+AH)/(NS+AS)$ . This ratio correlated to TEWL (Figure 4G) even for the samples of each site independently. Together, these data showed that I) barrier function correlated to the ratio of ceramide subclasses from different synthesis routes, II) certain ceramide subgroups are increased in stripped SC, III) the amount of these subgroups correlated to TEWL, and IV) treatment is able to reduce the increase of these ceramide subgroups that correlate positively to an increased TEWL (AS, NS C44 and C34 ceramides).



**Figure 4. The effect of treatment on ceramide groups correlating to barrier function.** **A,B)** Correlations between TEWL and the amount of subclass AS (ng/SQ). The Pearson  $r$  and  $p$ -value are indicated (A: Samples of tape-strips 5-8 B: Samples of tape-strips 17-20). **C)** The amount subclass AS (ng/SQ) at all sites at depths of both 5-8 and 17-20 tape-strips (mean  $\pm$ SD). Effect size was determined with a linear mixed model. **D,E)** Correlations between TEWL and the amount of ceramides with a chain length of 34 carbon atoms (ng/SQ). The Pearson  $r$  and  $p$ -value are indicated (D: Samples of tape-strips 5-8 E: Samples of tape-strips 17-20). **F)** The amount of ceramides with a chain length of 34 carbon atoms (ng/SQ) at all sites at depths of both 5-8 and 17-20 tape-strips (mean  $\pm$ SD). Effect size was determined with a linear mixed model. **G)** Pearson  $r$  of the correlation between TEWL and the total amount of each subclass at all sites at depths of both 5-8 and 17-20 tape-strips. **H)** The correlation between TEWL and ratio of subclasses  $(NdS+NP+NH+AdS+AP+AH)/(NS+AS)$  of both depths, excluding NS C40. Pearson  $r$  per site were: control: -0.648 Stripped: -0.808 Treated: -0.729 Stripped+Treated: -0.694. All statistical outputs of the LMM are found in Table 1.

**Table 1. The output of the LMMs per ceramide subgroup.** The intercept is the estimated amount at the control site at a depth of 5-8 tape-strips. Estimates are in ng/SQ. The effect size is the amount in ng/SQ with which the variable changes the intercept. Significance levels of all effect sizes and interaction are given.

		Subclass AS		Subclass NS C44		C34 ceramides	
		Estimate	p-value	Estimate	p-value	Estimate	p-value
Effect size	Intercept (control)	267.1	<0.001	56.6	<0.001	39.4	<0.001
	Stripped	+281.5	<0.001	+56.4	<0.001	+68.0	<0.001
	Treated	+27.3	0.402	+7.2	0.194	+5.6	0.438
	Depth 17-20	-41.3	0.076	-2.1	0.597	-5.2	0.297
Interactions	Stripped*Treated	-108.5	0.020	-19.5	0.013	-37.2	<0.001

## Mean carbon chain length and spacing related to the lamellar phases

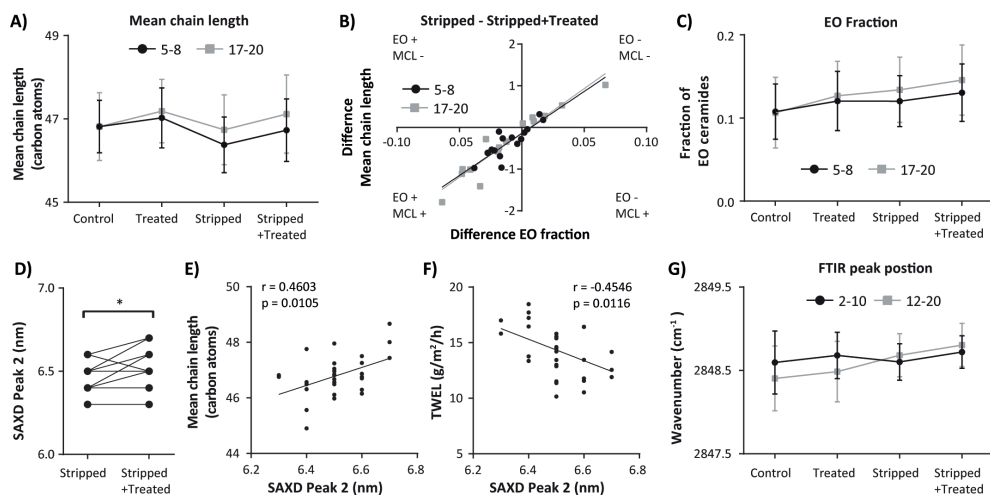
Previous studies have shown that the main ceramide chain length is an important parameter of the SC ceramide fraction.<sup>43</sup> For this reason it was decided to study whether a correlation existed between mean chain length and both the EO ceramide fraction and lamellar spacing. Figure 5A shows the mean chain length of the ceramides in samples prepared from tapes 5-8 and 17-20. A similar LMM as for subclass AS was made (Table 2). This showed that stripping decreased the mean chain length significantly. Treatment was able to increase the mean chain length restoring the mean chain length at the stripped+treated sites. The increase in mean chain length of the stripped+treated sites could be due to an increase in the fraction of esterified omega-acyl-ceramide (EO fraction) (Figure S8 shows this correlation per site). Changes in EO fraction due to treatment of the stripped site (stripped - stripped+treated) correlated to changes in the mean chain length due to treatment of the stripped site (Figure 5B). Treatment was able to increase the EO fraction significantly (Figure 5C and Table 2 show the EO fraction and the output of LMM, respectively).

The carbon chain length of the ceramides could influence the spacing of the strong diffraction peak in the diffraction pattern as measured by SAXD. From the position of the strong diffraction peak in the X-ray diffraction curve (peak 2), the corresponding spacing was calculated (referred to as peak 2 in Figure 5D). A significant increase in spacing due to treatment was observed and a positive correlation was observed between the mean chain length and the spacing, (Figure 5E). When comparing the spacing to TEWL, a correlation was observed between an increased TEWL and a decrease in spacing (Figure 5F). Stripping and treatment thus affected the mean chain length, which correlated to EO fraction and the lamellar spacing.

## Effects of stripping and treatment on lateral packing

Next to the lamellar spacing, the lateral packing is important for a proper barrier function. For this reason the effect of treatment and stripping on the lateral packing was also examined. All skin sites were studied using ATR-FTIR, where both the  $\text{CH}_2$  symmetric stretching vibrations and the  $\text{CH}_2$  scissoring vibrations were examined. A LMM was built for the  $\text{CH}_2$  symmetric stretching peaks. Stripped, treated, and depth were used as fixed factors and subject as random factor (Table 2). A significant increase in peak position was observed due to barrier disruption and treatment suggesting a slight increase in disordering of the lipids. The effects were stronger deeper in the SC. However, the peak position at all sites is located at wavenumbers between  $2848.46 \text{ cm}^{-1}$  and  $2848.80 \text{ cm}^{-1}$ , indicating an ordered lateral lipid organization at all sites. A similar linear mixed model was built for the  $\text{CH}_2$  scissoring peak. We determined the FWHM of the scissoring vibrations. Table S5 depicts the result of this model. At the control site the FWHM was significantly decreased at deeper layers (tape-strips 12-20), yet, the FWHM was  $11.3 \text{ cm}^{-1}$  or higher, indicative for a large fraction of lipids forming an orthorhombic lateral packing. No deviation from an ordered structure by treatment or stripping was observed.





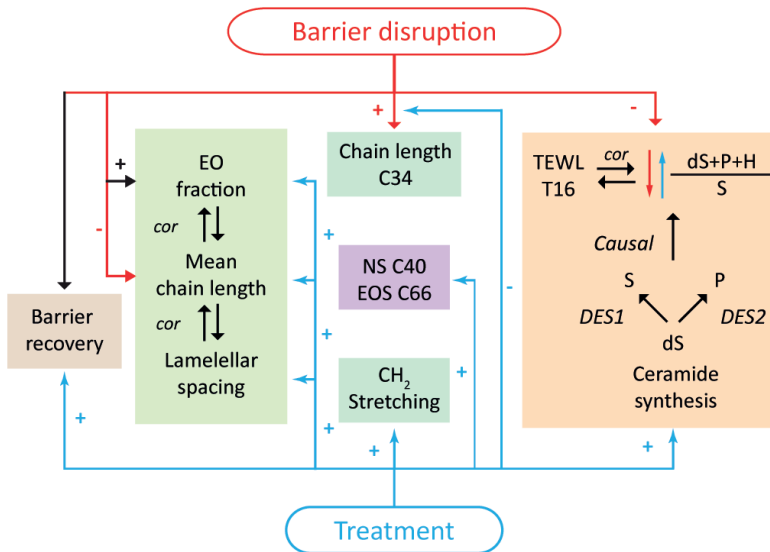
**Figure 5. Mean chain length and lamellar spacing.** **A)** The mean total chain length (carbon atoms) for the 4 different sites at two depths excluding the ceramides of the formulation in the calculation. Data were compared using a LMM. **B)** The correlation between changes in fraction EO ceramides and changes in the mean chain lengths (carbon atoms) at the stripped site due to treatment (amount at the stripped site subtracting the stripped+treated). **C)** The EO fraction of the samples at the 4 different sites at two depths. Data were compared using a LMM. **D)** The difference between the SAXD peak 2 position in the stripped and stripped+treated sites. The differences were tested with a Wilcoxon matched pairs signed rank test. **E)** The correlation between the SAXD peak 2 (nm) and TEWL (g/m<sup>2</sup>/h) at the end of the study. Pearson r and p-value are provided. **F)** The correlation between the SAXD peak 2 (nm) and the mean chain length (carbon atoms) of all ceramides. Pearson r and p-value are provided. **G)** The wavenumber in cm<sup>-1</sup> of the stretching peak position. Data were compared using a LMM.

**Table 2. The output of the LLMs for mean chain length, EO fraction, and stretching peak position.** The mean chain length and EO fraction were calculated excluding ceramides NS C44 and EOS C66. The wavenumber in cm<sup>-1</sup> of the stretching peak position was calculated using the center of gravity at 90% peak height. As fixed factors stripped, treated and depth were used. The intercept was the control site at a depth of 5-8 tape-strips or for spectra of tapes 2-10.

		Mean chain length		EO fraction		Wavenumber cm <sup>-1</sup>	
		Estimate	p-value	Estimate	p-value	Estimate	p-value
Effect size	Intercept (control)	46.76	<0.001	0.107	<0.001	2848.60	<0.001
	Stripped	-0.37	<0.001	+0.011	0.007	+0.02	0.622
	Treated	+0.33	<0.001	+0.014	<0.001	+0.09	0.013
	Depth	+0.08	0.366	0.003	0.537	-0.17	<0.001
Interactions	Stripped*Depth	+0.29	0.021	+0.012	0.043	+0.26	<0.001

## Discussion

In this study we showed that a VC based formulation is able to accelerate the barrier recovery of tape-strip disrupted healthy human skin and, for the first time, demonstrate that formulations can act by normalizing the SC lipid composition. Multiple effects of barrier disruption and treatment on barrier recovery, ceramide composition, and lipid organization were examined (Figure 6 provides an overview). Using PCA, samples were clustered into groups of treatment and stripped, based on changes in ceramide profile. Moreover, correlations of ceramide levels to PC2 had great similarities to correlations between ceramide levels and barrier function after barrier recovery. Thus, the loading variables for PC2 were a powerful tool to identify ceramides responsible for profile changes related to barrier disruption. At the stripped sites after recovery, multiple changes in the ceramide profiles were observed. Treatment partially normalized these changes in SC ceramide composition caused by barrier disruption. These ceramide profile changes correlated to the barrier function, indicating a relation of the changes in these lipids and an accelerated barrier recovery process.



**Figure 6. An overview of the effects of barrier disruption and treatment.** Negative effects are indicated with an – sign and positive effects with a + sign. Barrier disruption induces barrier recovery and treatment accelerates this process. Changes in the EO fraction, mean chain length, and lamellar spacing showed correlations (*cor*) indicated by the arrows. The changes in lipid composition, organization and subsequent effects are indicated. The ratio  $(dS+P+H)/S$ , is mainly determined by the enzymes human-dihydro- $\Delta^4$ -desaturase (DES1) and human-sphingolipid-C4-hydroxylase (DES2), responsible for the production of sphingosine and phytosphingosine ceramides from dihydrosphingosine ceramides, respectively. The arrows indicate that barrier disruption changed the lipid synthesis decreasing this ratio and treatment had the opposite effect. The correlation of the TEWL at day 16 (T16) to the subclass ratio is indicated, yet, the amount of C34 and the SAXD Peak 2 position correlated to TEWL as well.

When examining barrier recovery, two phases could be distinguished; a first steep increase in barrier recovery % after three hours, followed by second gradually diminishing barrier recovery rate. Treatment increased the level of the first phase, where after three hours a recovery of ~25% was reached compared to ~10% without treatment. However, early effects of formulation on barrier recovery measured by TEWL can be caused by leftover formulation on the skin<sup>27</sup>, even though leftover formulation was removed prior to TEWL measurement. The second recovery phase was not affected by residual formulation on the skin surface, because the formulation was applied after TEWL measurement. During the second phase, treatment had a pronounced effect on the barrier recovery rate.

One of the aims of the tested formulation was to replenish SC ceramides. Treatment caused significant increase in the formulated ceramides level; even at deeper SC layers. Although, a 2-day washout period would have removed almost all superficial formulation, it cannot be stated with 100% certainty that the applied ceramides measured with LC/MS were incorporated into the SC. Some formulated ceramides could still be present in folds of the SC. In cultured skin, it was shown that ceramides stay in these folds.<sup>23</sup> Even though the increase of the formulated ceramides was observed at both depths, tape-strips obtained lipid material from various depth of SC<sup>44</sup>, and thus, tape-strips at both depths can contain material from these folds. Nevertheless, it has been demonstrated that when treating stripped *ex vivo* SC with deuterated ceramides once, small ceramide amounts were incorporated into the SC lipid matrix after regeneration.<sup>45</sup> Others have shown that dimeric ceramides are able to diffuse into the skin.<sup>46</sup> Together with the relatively large increases of the formulated ceramides, it is likely that a fraction of the applied ceramides did incorporate into the SC lipid matrix and actively enhance barrier recovery.

Multiple studies have used SC lipid containing formulations, either for long or short term treatment<sup>18-21</sup> and it was shown that the SC lipids ratio in formulations is essential for short term barrier recovery.<sup>22,47</sup> Visscher et al. tested the effect of 5 days VC treatment on TEWL and hydration of tape-stripped SC, but they observed no changes in TEWL when comparing with untreated stripped SC.<sup>28</sup> In comparison to our study, the barrier was less extensively disrupted. Therefore, incomplete SC removal may lead to a lower efficacy of the formulation to improve barrier repair. Here, a detailed description of the recovery process is provided and, for the first time, insights are given into changes in the SC ceramide composition due to treatment. Below, a mechanism is suggested on how a VC based formulation affects ceramide synthesis.

At the stripped (non-treated) sites, two prominent changes were observed in the endogenous ceramides: I) the amounts of S subclass were increased, and II) C34 ceramides were increased. The amounts of these ceramide groups correlated to a decrease in barrier function. However, from these data it could not yet be deduced if these increases were causal or contingent with a decreased barrier function. A plausible reasoning is that in a direct response to barrier disruption, the epidermal calcium gradient changed<sup>2</sup> and a subsequent modulation in gene expression occurred.<sup>29</sup> These changes in epidermal homeostasis might have affected the SC ceramide processing in two ways.

First, in the epidermal lipid biosynthesis, ceramides containing a dihydrosphingosine are either converted into sphingosine or phytosphingosine containing ceramide<sup>48,49</sup>

(Figure 6) (biosynthesis of subclass H ceramides has not yet been reported). When defining the ratio between the subclasses as:  $(dS+P+H)/S$ , this ratio could only change if either of the two known lipid biosynthesis pathways changed. The present study showed that this ratio was substantially decreased at the stripped sites, indicating barrier disruption influenced the specific synthesis pathways of subclasses S and P. Importantly, the ratio between the subclasses correlated strongly to TEWL after barrier recovery, which suggests that an optimal ratio is important for an optimal skin barrier repair. Furthermore, this correlation was site independent, thus, at stripped sites with a higher TEWL, synthesis of sphingosine ceramides was preferred over phytosphingosine containing ceramides. Treatment was able to favorably change the subclass ratio towards a lipid composition that correlated to a more efficient skin barrier recovery, by reducing the amount of NS and AS subclasses.

The second change in lipid biosynthesis due to barrier disruption was the increased amount of C34 ceramide. These ceramides are produced from the sphingoid and acyl building blocks (a C16 or C18 acyl chain and a C16 or C18 sphingoid chain<sup>50</sup>) and do not require additional elongation steps.<sup>51</sup> Although these C34 ceramides can be produced as a quick first response to mitigate an impaired SC barrier, C34 ceramides are “short” chain ceramides which have been shown to reduce skin barrier function.<sup>6,52</sup> Altogether, the C34 ceramides and the ratio of ceramide subclasses could function as indicators of reduced barrier function and changes in barrier lipid homeostasis. Here, we showed that treatment was able to partly diminish the amount of C34 ceramides and, by diminishing the amount of NS and AS, able to change the subclass ratio and thus the preceding ceramide synthesis. This reduction could be attributed to I) the semi-occlusive effect of the formulation, in which the formulation acted as an additional barrier, II) the presence of excess formulated ceramides reducing the response to replenish the ceramides. As a whole, these data show that the formulation partially normalized the lipid biosynthesis and directed the SC towards a more functional lipid homeostasis. The lipid organization was used to verify if changes in ceramides composition were causal to changes in barrier function.

An important characteristic of the SC ceramide fraction is the mean chain length. The chain length was reduced when the barrier function is impaired e.g.: in stripped sites and AD.<sup>8,17</sup> Here, treatment was able to restore the mean chain length at stripped sites (excluding the added ceramides). The mean chain length correlated strongly to the fraction of EO ceramides and treatment was able to increase the EO fraction. At the treated sites, the spacing was slightly increased indicating an increase in the repeat distances of the lamellar phases. Furthermore, the spacing correlated to the mean chain length and weakly correlated to the EO fraction (data not shown). Taken together, the data suggest that treatment influenced the lamellar lipid organization through increasing the EO fraction thereby increasing the mean chain length (Figure 6).

As far as the lateral packing was concerned, our studies showed that a high fraction of lipids formed an orthorhombic packing and this was not affected by stripping. For this reason the fraction of lipids forming an orthorhombic packing could not be increased due to treatment. Changes due to treatment might become more apparent in affected skin of AD patients where the fraction of lipids forming an orthorhombic organization is decreased compared to that in healthy skin.<sup>43</sup>

To summarize the effects of the VC based formulation on barrier recovery in healthy SC, it was observed that treatment: I) accelerated barrier recovery, II) changed the lipid synthesis to a more favorable lipid composition by decreasing the amount of C34 ceramides and changing the subclass ratio (dS+P+H)/S, and increasing the fraction of EO ceramides, III) these changes in lipid composition affected the SC lipid organization and correlated to the SC barrier function. Interestingly, similar increases in S subclass, and increases in the fraction of C34 chain length ceramides were observed in AD.<sup>8,17</sup> An essential application of this formulation could be in a particular group of AD patients, where barrier restoring lipophilic formulations are commonly used.<sup>53</sup> Moreover, the treatment presented here modulated the alterations in lipid composition observed in AD, towards a healthy lipid homeostasis.

## Acknowledgments

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## References

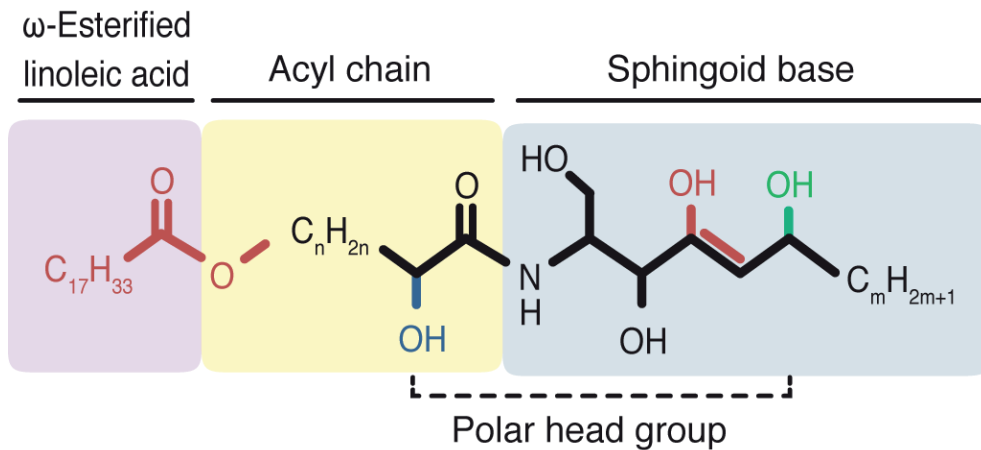
- Eckhart L, Lippens S, Tschachler E, et al. Cell death by cornification. *Biochimica et biophysica acta* **2013**; 1833: 3471-3480.
- Feingold KR, Elias PM. Role of lipids in the formation and maintenance of the cutaneous permeability barrier. *Biochimica et biophysica acta* **2014**; 1841: 280-294.
- van Smeden J, Janssens M, Kaye EC, et al. The importance of free fatty acid chain length for the skin barrier function in atopic eczema patients. *Exp Dermatol* **2014**; 23: 45-52.
- Sahle FF, Gebre-Mariam T, Dobner B, et al. Skin diseases associated with the depletion of stratum corneum lipids and stratum corneum lipid substitution therapy. *Skin Pharmacol Physiol* **2015**; 28: 42-55.
- t'Kindt R, Jorge L, Dumont E, et al. Profiling and characterizing skin ceramides using reversed-phase liquid chromatography-quadrupole time-of-flight mass spectrometry. *Analytical chemistry* **2012**; 84: 403-411.
- Mojumdar EH, Kariman Z, van Kerckhove L, et al. The role of ceramide chain length distribution on the barrier properties of the skin lipid membranes. *Biochim Biophys Acta* **2014**; 1838: 2473-2483.
- Stahlberg S, Lange S, Dobner B, et al. Probing the Role of Ceramide Headgroup Polarity in Short-Chain Model Skin Barrier Lipid Mixtures by <sup>2</sup>H Solid-State NMR Spectroscopy. *Langmuir* **2016**; 32: 2023-2031.
- Janssens M, van Smeden J, Gooris GS, et al. Increase in short-chain ceramides correlates with an altered lipid organization and decreased barrier function in atopic eczema patients. *Journal of lipid research* **2012**; 53: 2755-2766.
- Matsui T, Amagai M. Dissecting the formation, structure and barrier function of the stratum corneum. *Int Immunol* **2015**; 27: 269-280.
- Rawlings AV. Molecular basis for stratum corneum maturation and moisturization. *Br J Dermatol* **2014**; 171 Suppl 3: 19-28.
- Tanaka M, Zhen YX, Tagami H. Normal recovery of the stratum corneum barrier function following damage induced by tape stripping in patients with atopic dermatitis. *Br J Dermatol* **1997**; 136: 966-967.
- Wolf R, Parish LC. Barrier-repair prescription moisturizers: do we really need them? Facts and controversies. *Clin Dermatol* **2013**; 31: 787-791.
- Hon KL, Leung AK, Barankin B. Barrier repair therapy in atopic dermatitis: an overview. *Am J Clin Dermatol* **2013**; 14: 389-399.
- Corazza M, Minghetti S, Bianchi A, et al. Barrier creams: facts and controversies. *Dermatitis* **2014**; 25: 327-333.
- van Zuuren EJ, Fedorowicz Z, Christensen R, et al. Emollients and moisturisers for eczema. *Cochrane Database Syst Rev* **2017**; 2: CD012119.
- Ito S, Ishikawa J, Naoe A, et al. Ceramide synthase 4 is highly expressed in involved skin of patients with atopic dermatitis. *J Eur Acad Dermatol Venereol* **2016**.
- Ishikawa J, Narita H, Kondo N, et al. Changes in the ceramide profile of atopic dermatitis patients. *The Journal of investigative dermatology* **2010**; 130: 2511-2514.
- Meckfessel MH, Brandt S. The structure, function, and importance of ceramides in skin and their use as therapeutic agents in skin-care products. *J Am Acad Dermatol* **2014**; 71: 177-184.
- Lynde CW, Andriessen A. A cohort study on a ceramide-containing cleanser and moisturizer used for atopic dermatitis. *Cutis* **2014**; 93: 207-213.
- Oh MJ, Nam JJ, Lee EO, et al. A synthetic C16 omega-hydroxyphytoceramide improves skin barrier functions from diversely perturbed epidermal conditions. *Arch Dermatol Res* **2016**; 308: 563-574.
- Barba C, Parra JL, Coderch L, et al. In vivo and in vitro evaluation of topical formulations containing physiological lipid mixture for replacement of skin barrier function. *G Ital Dermatol Venereol* **2014**; 149: 347-353.
- Man MM, Feingold KR, Thornfeldt CR, et al. Optimization of physiological lipid mixtures for barrier repair. *The Journal of investigative dermatology* **1996**; 106: 1096-1101.
- Zhang Q, Flach CR, Mendelsohn R, et al. Topically applied ceramide accumulates in skin glyphs. *Clin Cosmet Invest Dermatol* **2015**; 8: 329-337.
- Rhodes LE, O'Farrell S, Jackson MJ, et al. Dietary fish-oil supplementation in humans reduces UVB-erythema sensitivity but increases epidermal lipid peroxidation. *The Journal of investigative dermatology* **1994**; 103: 151-154.
- Rissmann R, Oudshoorn MH, Zwier R, et al. Mimicking vernix caseosa--preparation and characterization of synthetic biofilms. *Int J Pharm* **2009**; 372: 59-65.
- Oudshoorn MH, Rissmann R, van der Coelen D, et al. Effect of synthetic vernix biofilms on barrier recovery of damaged mouse skin. *Exp Dermatol* **2009**; 18: 695-703.
- Oudshoorn MH, Rissmann R, van der Coelen D, et al. Development of a murine model to evaluate the effect of vernix caseosa on skin barrier recovery. *Exp Dermatol* **2009**; 18: 178-184.
- Visscher MO, Barai N, LaRuffa AA, et al.

- Epidermal barrier treatments based on vernix caseosa. *Skin Pharmacol Physiol* **2011**: 24: 322-329.
29. Sextius P, Marionnet C, Bon FX, et al. Large scale study of epidermal recovery after stratum corneum removal: dynamics of genomic response. *Experimental dermatology* **2010**: 19: 259-268.
  30. Bandier J, Carlsen BC, Rasmussen MA, et al. Skin reaction and regeneration after single sodium lauryl sulfate exposure stratified by filaggrin genotype and atopic dermatitis phenotype. *Br J Dermatol* **2015**: 172: 1519-1529.
  31. O'Connor RJ, Ogle J, Odio M. Induction of epidermal damage by tape stripping to evaluate skin mildness of cleansing regimens for the premature epidermal barrier. *Int J Dermatol* **2016**: 55 Suppl 1: 21-27.
  32. Breternitz M, Flach M, Prassler J, et al. Acute barrier disruption by adhesive tapes is influenced by pressure, time and anatomical location: integrity and cohesion assessed by sequential tape stripping. A randomized, controlled study. *Br J Dermatol* **2007**: 156: 231-240.
  33. Pinkus H. Examination of the epidermis by the strip method of removing horny layers. I. Observations on thickness of the horny layer, and on mitotic activity after stripping. *The Journal of investigative dermatology* **1951**: 16: 383-386.
  34. Boiten W, Absalah S, Vreeken R, et al. Quantitative analysis of ceramides using a novel lipidomics approach with three dimensional response modelling. *Biochimica et biophysica acta* **2016**: 1861: 1652-1661.
  35. Masukawa Y, Narita H, Sato H, et al. Comprehensive quantification of ceramide species in human stratum corneum. *Journal of lipid research* **2009**: 50: 1708-1719.
  36. van Smeden J, Boiten WA, Hankemeier T, et al. Combined LC/MS-platform for analysis of all major stratum corneum lipids, and the profiling of skin substitutes. *Biochimica et biophysica acta* **2014**: 1841: 70-79.
  37. Motta S, Monti M, Sesana S, et al. Ceramide composition of the psoriatic scale. *Biochimica et biophysica acta* **1993**: 1182: 147-151.
  38. Boncheva M, Damien F, Normand V. Molecular organization of the lipid matrix in intact Stratum corneum using ATR-FTIR spectroscopy. *Biochimica et biophysica acta* **2008**: 1778: 1344-1355.
  39. Damien F, Boncheva M. The extent of orthorhombic lipid phases in the stratum corneum determines the barrier efficiency of human skin in vivo. *The Journal of investigative dermatology* **2010**: 130: 611-614.
  40. Checa A, Bedia C, Jaumot J. Lipidomic data analysis: tutorial, practical guidelines and applications. *Anal Chim Acta* **2015**: 885: 1-16.
  41. Vaz FM, Pras-Raves M, Bootsma AH, et al. Principles and practice of lipidomics. *J Inherit Metab Dis* **2015**: 38: 41-52.
  42. De Livera AM, Zaloumis S, Simpson JA. Models for the analysis of repeated continuous outcome measures in clinical trials. *Respirology* **2014**: 19: 155-161.
  43. van Smeden J, Janssens M, Kaye EC, et al. The importance of free fatty acid chain length for the skin barrier function in atopic eczema patients. *Exp Dermatol* **2014**: 23: 45-52.
  44. van der Molen RG, Spies F, van 't Noordende JM, et al. Tape stripping of human stratum corneum yields cell layers that originate from various depths because of furrows in the skin. *Arch Dermatol Res* **1997**: 289: 514-518.
  45. Berkers T, Visscher D, Gooris GS, et al. Topically Applied Ceramides Interact with the Stratum Corneum Lipid Matrix in Compromised Ex Vivo Skin. *Pharmaceutical research* **2018**: 35: 48.
  46. Neubert RH, Sonnenberger S, Dobner B, et al. Controlled Penetration of a Novel Dimeric Ceramide into and across the Stratum Corneum Using Microemulsions and Various Types of Semisolid Formulations. *Skin Pharmacol Physiol* **2016**: 29: 130-134.
  47. Yang L, Mao-Qiang M, Taljebini M, et al. Topical stratum corneum lipids accelerate barrier repair after tape stripping, solvent treatment and some but not all types of detergent treatment. *Br J Dermatol* **1995**: 133: 679-685.
  48. Gault CR, Obeid LM, Hannun YA. An overview of sphingolipid metabolism: from synthesis to breakdown. *Advances in experimental medicine and biology* **2010**: 688: 1-23.
  49. Uchida Y, Hara M, Nishio H, et al. Epidermal sphingomyelins are precursors for selected stratum corneum ceramides. *Journal of lipid research* **2000**: 41: 2071-2082.
  50. Masukawa Y, Narita H, Shimizu E, et al. Characterization of overall ceramide species in human stratum corneum. *Journal of lipid research* **2008**: 49: 1466-1476.
  51. Kihara A. Synthesis and degradation pathways, functions, and pathology of ceramides and epidermal acylceramides. *Prog Lipid Res* **2016**: 63: 50-69.
  52. Paloncyova M, Vavrova K, Sovova Z, et al. Structural Changes in Ceramide Bilayers Rationalize Increased Permeation through Stratum Corneum Models with Shorter Acyl Tails. *J Phys Chem B* **2015**: 119: 9811-9819.
  53. Giam YC, Hebert AA, Dizon MV, et al. A review on the role of moisturizers for atopic dermatitis. *Asia Pac Allergy* **2016**: 6: 120-128.

## Supplementary data

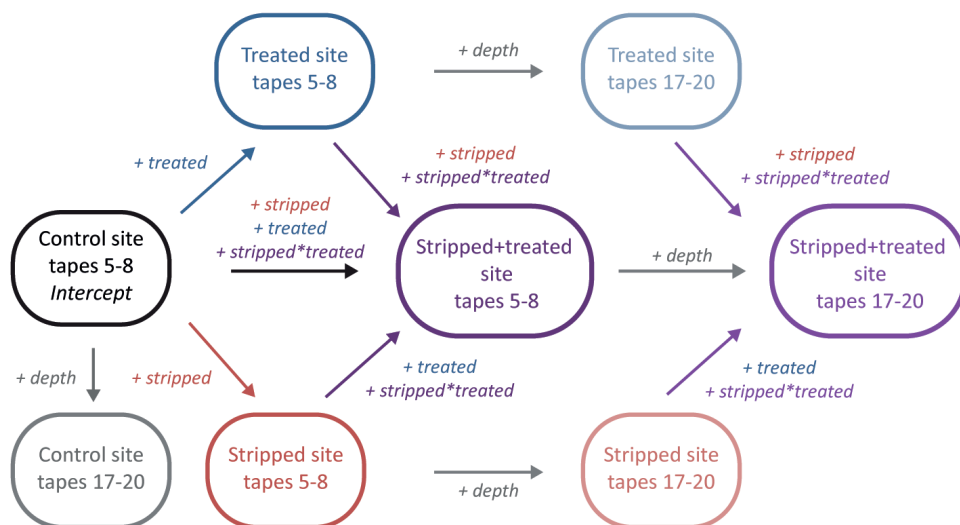


**Figure S1. Indication of TEWL measurement sites.** The TEWL measurements at the sites were performed at three sections of the sites. Here the stripped site is shown which can be identified by the erythroderma. TEWL measurements were performed at the sections indicated with black circles. At every time-point TEWL was measured at the right, middle, and left section.

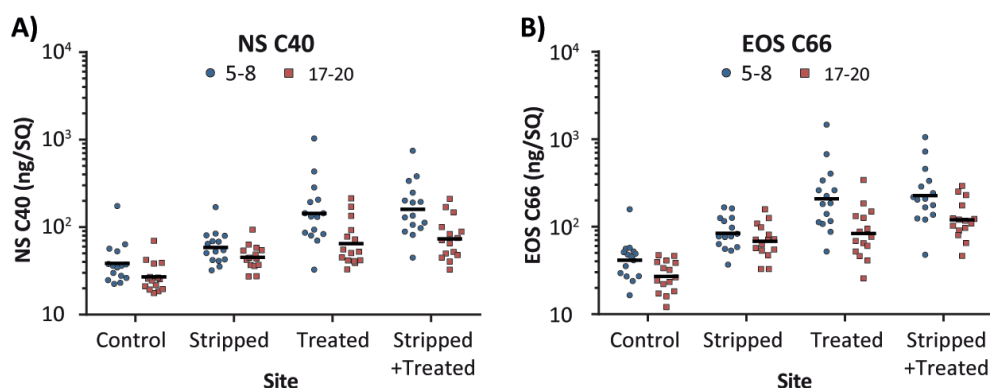


**Figure S2. General structure of a ceramide.** A ceramide can be composed of three parts which determine the subclass of the ceramide. The polar head group consists of a sphingoid base and an acyl chain. There are 4 different sphingoid bases in human SC: the sphingoid (S), a dihydro-sphingoid (dS), a 4-hydroxy or phyto-sphingoid (P), and a 6-hydroxy-sphingoid (H). The acyl chain of human SC occurs in 3 different forms. A non-hydroxyl acyl chain (N), an  $\alpha$ -hydroxy-acyl, and a  $\omega$ -hydroxy-acyl chain (O). In SC a specific type of ceramide occurs where the  $\omega$ -hydroxyl group becomes esterified with a linoleic acid called the EO subclass of ceramides. Together these form 16 different subclasses. Both the acyl and sphingoid chain can vary in carbon chain length.

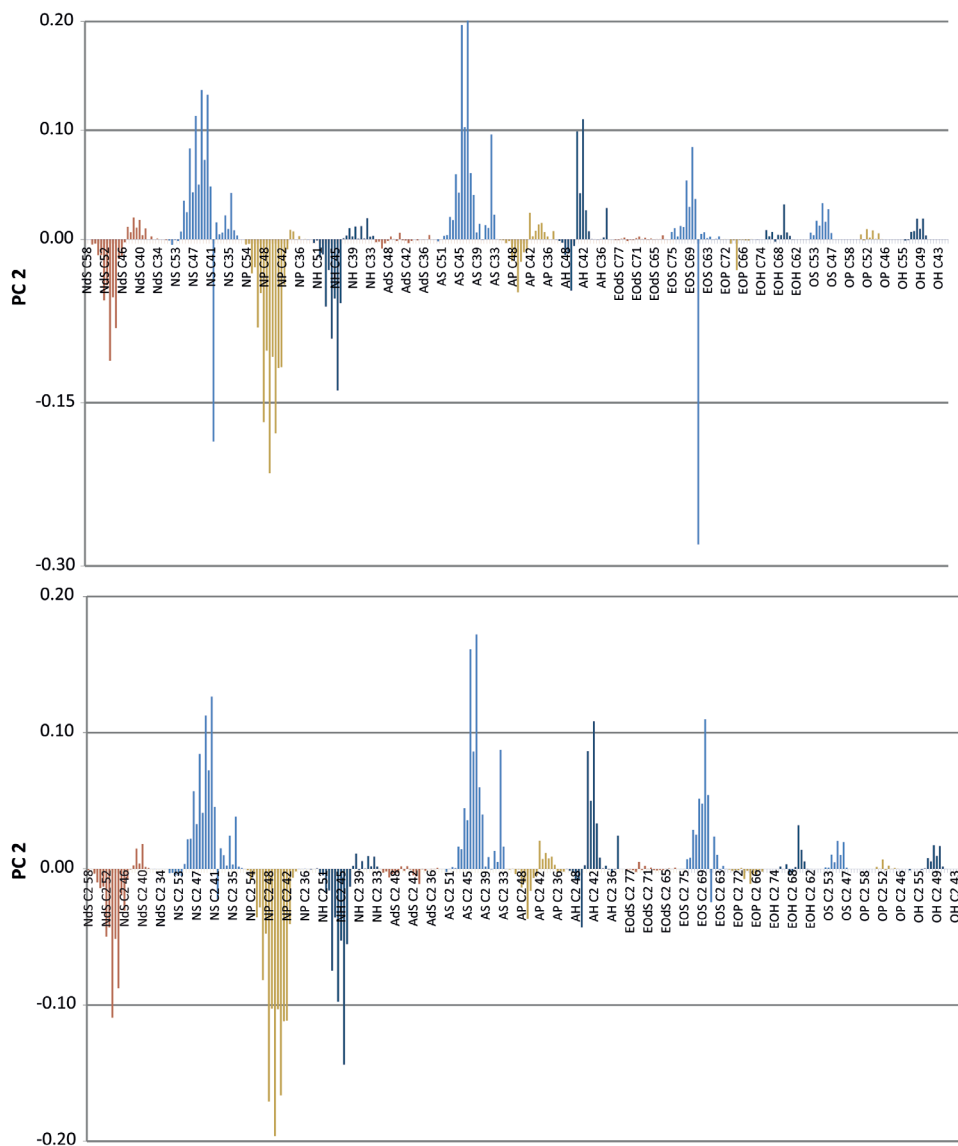




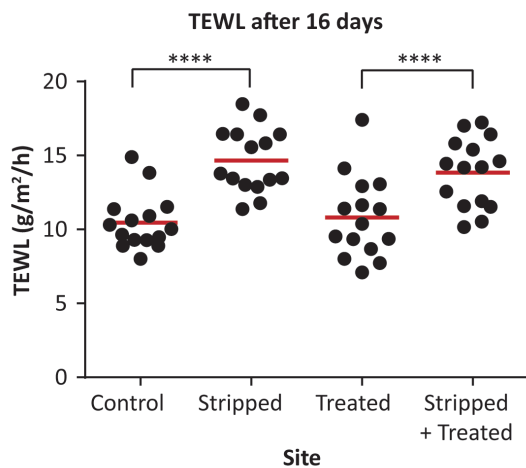
**Figure S3. Linear mixed modelling.** The LLMs used here compare the effects of treatment, stripping, and depth to the control site of the subject. Using subject as a random factor, gives each subjects control site (starting point) a different value, so it can start at any random point. It then determines the effect size of the fixed factors (treatment, stripping, and depth). ‘Fixed’ means that the model will say that all these effects have the same size for each subject and for each time this effect occurs, so also the effect at treatment at the stripped site (difference between stripped and stripped+treated). Because a stripped site can respond differently to treatment, an interaction term is added to the model, in this case *stripped\*treated*. In this model the interaction is the effect size of treatment additional to the effects of stripping and treatment (the fixed factors). For most data we were interested in the effect of treatment on the stripped sites. This is the sum of the effect sizes of treatment and *stripped\*treated* (illustrated above as the arrow from stripped site to Stripped+treated site). In the schematic above it is shown which steps are required to calculate the predicted values of the mixed model. The effect sizes are indicated in italics. Here, we were not interested in the mean predicted by the model but in the effect sizes of the different factors, as these calculate how the experimental conditions changed and if these were significant.



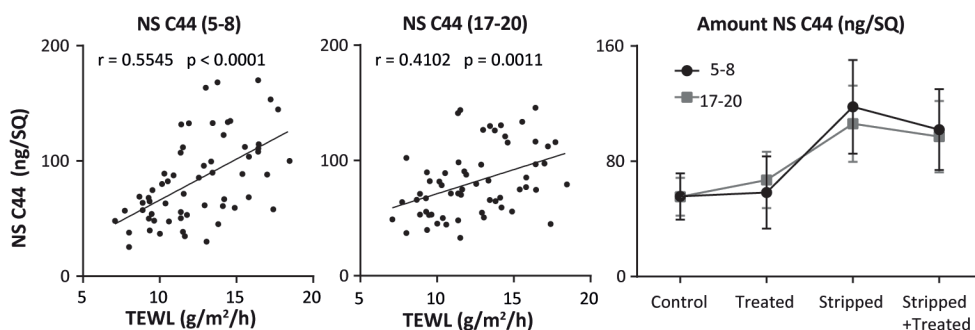
**Figure S4. LLM of the formulation ceramides. A,B** Plots of the amounts of NS C40 and EOS C66 (log (ng/SQ)) in all quantified samples. Lines are geometric means. Statistical outputs of the LMMs are reported in supplementary Table S4.



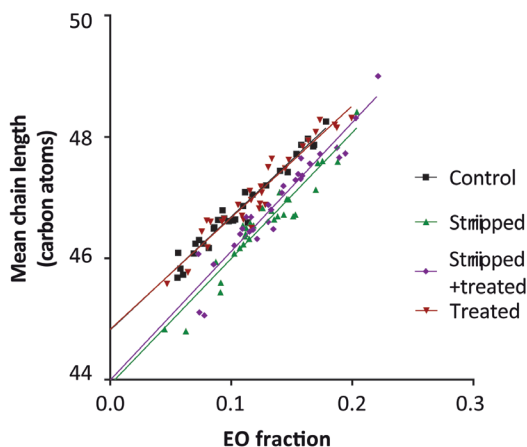
**Figure S5. PC2 plot.** The two graphs depict the scores/correlation of all ceramides to PC2. The first graph shows all ceramides detected in samples 5-8 and the second graph all ceramides detected in samples 17-20. Subclasses were given different colors. High positive scores indicate a correlation to the stripped sites and a high negative score to the control sites.



**Figure S6. TEWL at day 16.** The TEWL (g/m<sup>2</sup>/h) of all 4 sites after 16 days of treatment and/or barrier recovery. The red line is the mean value and the significance levels that are indicated were tested with a 1-way ANOVA with Bonferroni post-test.



**Figure S7. The amount of NS C44 and the correlation to the TEWL.** Pearson  $r$  and the corresponding  $p$ -value are indicated above the data. The amount in ng/SQ of NS C44 at the 4 site and at both depths (5-8 and 17-20 tape-strips).



**Figure S8. Correlation between mean chain length and EO fraction.** The correlations between the mean chain length and the fraction of EO ceramides are depicted per site. Each site showed significant Pearson correlations coefficients: control (0.9817), stripped (0.9565), treated (0.9600), and stripped+treated (0.9568).

**Table S1. List of the ceramides standards used with the corresponding supplier.**

Ceramide	Supplier	Ceramide	Supplier
N(24)dS(18)	Avanti*	A(24)S(18)	Avanti*
N(18)dS(18)	Avanti*	A(R)(22)S(18)	Avanti*
N(16)dS(18)	Avanti*	A(S)(22)S(18)	Avanti*
N(24)S(18)	Evonik**	A(R)(16)S(18)	Avanti*
N(22)S(18)	Avanti*	A(S)(16)S(18)	Avanti*
N(20)S(18)	Avanti*	A(R)(24)P(18)	Avanti*
N(18)S(18)	Avanti*	E(18:2)O(30)S(18)	Evonik**
N(16)S(18)	Avanti*	E(18:2)O(27)S(18)	Evonik**
N(24)P(18)	Evonik**	E(18:2)O(30)P(S18)	Evonik**
N(22)P(18)	Evonik**	E(18:2)O(27)P(18)	Evonik**
N(16)P(18)	Evonik**	N(24deu)S(18)	Evonik**

\* Avanti polar lipids (Alabaster AL, USA), \*\* Evonik Industries (Essen, Germany)

**Table S2. Ingredients of the formulation**

Ingredient	Supplier	Proportion
Super sterol esters Crodalan SWL	Croda, UK	49.72% w/w
Crodamol GTCC miglyol	Croda, UK	36.98% w/w
Squalane	Galderma	6.63% w/w
Cholesterol USP/NF	Croda, UK	3.63% w/w
Palmitic acid (FFAC16:0)	Emery	0.83% w/w
Stearic acid (FFA 18:0))	Berg + Schmidt	0.10% w/w
Ceramide NS (C40)	Evonik, Germany	1.18% w/w
Ceramide EOS (C66)	Evonik, Germany	0.83% w/w
Oxyplex LM (anti-oxidant; protection of Ceramide EOS) containing:	Merck, Germany	0.10% w/w
DL-γ-Tocopherol (Vitamin E)		0.025% w/w
Lecithin		0.025% w/w
Ascorbyl palmitate		0.020% w/w
Glycerin monostearate		0.020% w/w
Glycerin monooleate		0.0025%w/w
Citric acid		0.0025%w/w

**Table S3. Correlations to amount of barrier disruption.** The table depicts the Pearson r and p-value of the correlation between the TEWL directly after barrier disruption and AUC of the barrier recovery curve. It also contains the correlations between the difference in TEWLs directly after barrier disruption and the  $\Delta T$  at different recovery %.

	TEWL Start vs. AUC	TEWL Dif. vs. 5%	TEWL Dif. vs. 10%	TEWL Dif. vs. 15%	TEWL Dif. vs. 20%	TEWL Dif. vs. 25%	TEWL Dif. vs. 30%	TEWL Dif. vs. 35%	TEWL Dif. vs. 40%	TEWL Dif. vs. 45%
Pearson r	-0.26	0.05	0.15	0.20	0.17	-0.15	-0.26	-0.27	-0.34	-0.43
p-value	0.17	0.85	0.60	0.47	0.54	0.60	0.35	0.34	0.22	0.11

	TEWL Dif. vs. 50%	TEWL Dif. vs. 55%	TEWL Dif. vs. 60%	TEWL Dif. vs. 65%	TEWL Dif. vs. 70%	TEWL Dif. vs. 75%	TEWL Dif. vs. 80%	TEWL Dif. vs. 85%	TEWL Dif. vs. 90%	TEWL Dif. vs. 95%
Pearson r	-0.50	-0.40	-0.34	-0.34	-0.22	-0.09	-0.06	-0.23	-0.11	0.86
p-value	0.06	0.14	0.21	0.22	0.44	0.76	0.82	0.42	0.71	0.06

**Table S4. LLM of the formulation ceramides.** The table (A) shows the output of the LLM of the logarithm of the amount of NS C40 (ng/SQ) and EOS C66 (ng/SQ). The effect sizes are the changes to the intercept (control site tape-strips 5-8), due to the depth, stripping, and treatment. The second part of the table (B) shows the amount in ng/SQ. This was calculated by exponentiation of the mean log value outputted by the LLM.

A)

		Log NS C40		Log EOS C66 <sup>1</sup>	
		Estimate	p-value	Estimate	p-value
Effect size	Intercept	1.575	<0.001	1.50	<0.001
	Depth	-0.135	0.011	-0.135	0.011
	Stripped	+0.204	<0.001	+0.356	<0.001
	Treated	+0.580	<0.001	+0.697	<0.001
Interactions	Depth*Treated	-0.207	0.006	-0.200	0.006
	Stripped*Treated	-0.152	0.040	-0.260	0.040

B)

Calculated amount	NS C40 (ng/SQ)		EOS C66 (ng/SQ)	
	Depth 5-8	Depth 17-20	Depth 5-8	Depth 17-20
Control	37.62	27.57	38.93	28.54
Stripped	60.13	44.09	88.32	64.76
Treated	142.92	65.12	193.64	89.64
Stripped Treated	160.87	73.29	241.68	111.87

**Table S5. LLM of the effects on the lateral lipid packing.** The output of linear mixed models for the FWHM of the scissoring peak. The intercept is the estimated amount at the control site at a depth of 2-10 tape-strips. Estimates are in  $\text{cm}^{-1}$ . The effect size is the amount in  $\text{cm}^{-1}$  with which the variable changes the intercept. Significance levels of all effect sizes and interactions are given.

		FWHM	
		Estimate	p-value
	Intercept	11.86	<0.001
Effect size	Stripped	-0.37	0.19
	Treated	-0.43	0.08
	Depth 12-20	-0.59	0.02
Interactions	Stripped*Treated	0.44	0.23
	Stripped*Depth 12-20	0.67	0.07
	Treated*Depth 12-20	0.47	0.17
	Stripped*Treated*Depth 12-20	-1.32	0.01