

MRI and histologic studies on early markers of Alzheimer's disease Duijn, S. van

Citation

Duijn, S. van. (2018, October 10). *MRI and histologic studies on early markers of Alzheimer's disease*. Retrieved from https://hdl.handle.net/1887/66118

Version:	Not Applicable (or Unknown)
License:	<u>Licence agreement concerning inclusion of doctoral thesis in the</u> <u>Institutional Repository of the University of Leiden</u>
Downloaded from:	https://hdl.handle.net/1887/66118

Note: To cite this publication please use the final published version (if applicable).

Cover Page



Universiteit Leiden



The handle <u>http://hdl.handle.net/1887/66118</u> holds various files of this Leiden University dissertation.

Author: Duijn, S. van Title: MRI and histologic studies on early markers of Alzheimer's disease Issue Date: 2018-10-10

Chapter 1

General Introduction

This thesis addresses a variety of aspects of the early diagnosis of Alzheimer's disease (AD). AD is a complex clinical syndrome characterized by a cluster of symptoms and signs comprising difficulties in memory, changes in behaviour, disturbance in language and other cognitive functions causing impairments in activities of daily living. (Ferri et al., 2005; Qiu et al., 2009; Villemagne et al., 2013). In 2010, 5,4% of the European population at the age of 60+ had dementia, which was (at that time) 6.3 million people (Wittchen et al., 2011). AD accounts for 75% of all dementia cases, implying 4.7 million people in Europe had AD in 2010 and this number is further increasing. Brookmeyer et al., predicted an AD incidence of 1 in 85 persons worldwide in 2050 (Brookmeyer et al., 2007). As a result, AD represents an important socio-economic and public health concern. Results from human studies suggest that females are at higher risk for developing AD than men (Andersen et al., 1999; Corder et al., 2004; Grimm et al., 2012; Janicki and Schupf, 2010; Musicco, 2009)

Due to an incomplete understanding of the pathophysiology of the disease, an effective therapy is currently not available. Limitations in studying the early stages of the disease during life, have been partly responsible for lack of knowledge about the pathophysiology of AD. However, there are promising strategies, some of which are already effective in animal models and some are tested in clinical trials including immunotherapy (Lambracht-Washington and Rosenberg, 2015; Landlinger et al., 2015), inhibition of A β production (Howell et al., 2015; Wang et al., 2012) or tau aggregation (Harrington et al., 2015; Richter et al., 2014; Wischik et al., 2014; Wischik et al., 2015). The ability to detect the disease in an early stage would help increasing the knowledge on the pathophysiology of AD, would improve the chances for developing effective treatments, and would widen the therapeutic window for effective treatment.

However, at the moment AD is difficult to diagnose at an early stage and even at advanced stages of the disease a definitive diagnosis of AD still requires an autopsy. Therefore, diagnostic methods are needed allowing early in vivo detection of AD pathology (Jack, Jr. et al., 2010).

Pathology

The main pathological hallmarks of AD are atrophy, neurofibrillary degeneration and extracellular amyloid plaques (figure 1 and 2) (Doens and Fernandez, 2014; Dore et al., 2013; Ma et al., 2014; Price et al., 1991; Takahashi et al., 2010; Villemagne et al., 2013). All these can also be demonstrated in non-demented elderly subjects and therefore, are not specific for AD but their quantity and distribution in relation to the clinical symptoms is specific (Thal et al., 2014). It has been demonstrated that plaques and tangles lead to synaptic dysfunction, mitochondrial damage, inflammation and neuronal death (Doens and Fernandez, 2014; Takahashi et al., 2010). It still remains unknown how these pathological hallmarks are related to each other. Another frequent finding in AD is cerebral amyloid angiopathy (CAA) that can also contribute to the cognitive decline (Weller et al., 2009). More recently, changes in iron distribution have been noted (Bartzokis, 2011; Crichton et al., 2002; Haacke et al., 2005; Meadowcroft et al., 2015a; Meadowcroft et al., 2015b).

Atrophy

Shrinkage of the brain associated with AD is regarded as a valid marker of disease state and progression. Brain atrophy is correlated to neurofibrillary tangles (NFT) and neuropsychological deficits. It starts, in the majority of AD patients, in the hippocampus and the enthorinal cortex, extending to the temporal, parietal and frontal neocortices during the disease progression (Frisoni et al., 2010).

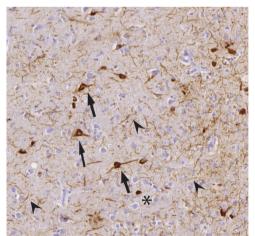


Figure 1: Examples of neurofibrillary tangles (arrows), neuropil threads (arrowheads) and dystrophic neurites (asterix).

Neurofibrillary degeneration

Neurofibrillary tangles (NFT) are one of the manifestations of neurofibrillary degeneration, the other being neuropil threads (NT) and dystrophic neurites (DN) (figure 1). In all these lesions there is intracellular accumulation of hyper-phosphorylated tau protein, forming soluble aggregates and paired helical filaments (PHF) (Kidd, 1963). NFT are neuronal cell bodies filled with PHF whereas in NT these PHF are present in the neuronal processes. DN are NT with irregular, dilated and distorted shapes. Normally tau proteins are involved in structural and regulatory function of the cytoskeleton where they promote the assembly of microtubule and their stability (Alonso et al., 2008; Grundke-Iqbal et al., 1986). However, when tau becomes hyper phosphorylated it exerts the exact opposite effect, leading to the dismantling of the same microtubule.

The resulting loss of neuronal structure impairs axonal transport, leading to disturbed proper synaptic, neuronal signalling and eventually leads to neuronal death (Ballatore et al., 2007). The degree of tau pathology correlates very well with dementia but neurofibrillary degeneration is not specific for AD: it is also seen in other neurodegenerative diseases, although with a different distribution in the brain.

Plaques

Plaques represent a wide array of lesions that contain extracellular deposits of amyloid β protein (A β) of which variable amounts are present as amyloid. Histologically, plaques are classified as diffuse, compact, classical and neuritic plaques. The type of plaque depends on the density and circumscription of A β (diffuse vs. compact plaques), the presence of an A β amyloid core (classical plaque) and the coexistence of dystrophic neurites (neuritic plaque) (figure 2) (Duyckaerts et al., 2009). A β plaques in an extensive amount is typical for AD but there is poor correlation between the amount of plaques and the degree of dementia.

Cerebral A β is generally cleaved by a-secretase and either degraded or cleared from the brain across the blood-brain barrier. A β peptide is generated by β - and γ -secretase induced cleavage of the amyloid precursor protein (APP), a transmembrane protein, forming predominantly A β 1-40 or A β 1-42. According to the amyloid cascade hypothesis, AD is initiated by an imbalance in A β production and clearance (Hardy, 2009; Hardy and Selkoe, 2002). This hypothesis is supported by the finding that APP gene mutations around the a, β - and γ -cleavage sites and gene mutations in proteins involved in cleavage at the APP γ -site lead to increased A β production and often early onset AD with an autosomal dominant pattern of inheritance. However, these mutations account for only < 5% of all AD cases. The other 95% of sporadic AD probably has a more complex multifactorial etiology (Minati et al., 2009).

Due to its fibrillogenic nature, high local concentrations of A β 1-42 aggregate into soluble oligomers. These oligomers cluster into larger insoluble A β fibrils that allow the formation of β -sheet structures, which are characteristic for amyloid. This clustering of oligomers triggers the misfolding

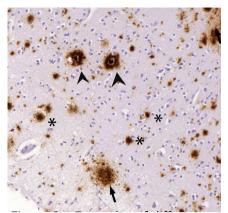


Figure 2: Examples of diffuse plaques (arrows), classical plaques (arrowheads) and compact plaques (asterix).

of other A β species, including the more soluble A β 1-40, forming plaques (Duyckaerts et al., 2009).

The study of Corder et al. suggested an acceleration of amyloid deposition in women of late middle age associated with APOE4 (Corder et al., 2004). In some AD mouse models, a similar sex-related difference was found, showing more A β accumulation in female mice (Callahan et al., 2001; Wang et al., 2003).

CAA

In the majority of AD cases different amounts of CAA are found in the brain (Natte et al., 2001; van Rooden et al., 2009; Weller et al., 2009). CAA is caused by the same A β deposits as in plaques, mainly A β 1-40 and always forms amyloid which leads to stiffness and a loss of structure of the vesselwall. CAA can occur as a sporadic disease with little or no parenchymal A β deposits and is considered a major cause of cerebral microbleeds, haemorrhages and cognitive loss.

Inflammation

In AD, microglia may play an important role in disease progression by activating different inflammatory cytokines, causing neuronal damage and cell death (Doens and Fernandez, 2014; Hardy and Selkoe, 2002; Kettenmann et al., 2011). Microglia cluster especially around plaques and CAA.

Iron

Iron was recently identified as one of the pathological changes in the AD brain (Bartzokis, 2011; Meadowcroft et al., 2009; van Duijn et al., 2013). There are two hypotheses on the role of iron in AD. The first hypothesis claims that iron would directly contribute to the development of AD, due to its neurotoxic characteristics when not properly regulated (Bartzokis, 2011). The second is the idea of iron deposits being secondary to the formation of plaques and tangles (Peters et al., 2015); not playing a leading role in the development of AD, but following the formation of plaques. Iron also has relevance in AD research because it can be detected with

high sensitivity by magnetic resonance imaging (MRI) and may serve as an in vivo marker for AD.

Magnetic resonance imaging (MRI) and spectroscopy (MRS) in AD

The in vivo diagnosis of AD is now based on clinical and neuropsychological criteria, with additional techniques such as neuroimaging and cerebrospinal fluid biomarkers playing a supportive role, resulting in "probable" AD at best. This diagnosis is not always accurate and needs post mortem histological confirmation (Fox et al., 1996; Hyman et al., 2012; Jack, Jr. et al., 2010). An MR-based hallmark for AD is hippocampal atrophy. This measurement, however, is neither conclusive nor specific for AD and consequently of limited use in clinical setting (Nasrallah and Wolk, 2014). Furthermore, cerebral atrophy in AD is found in a late stage of the disease (Jack, Jr. et al., 2010) and therefore intrinsically a poor candidate for early diagnosis. Iron is a potential interesting target for the detection of early changes in AD. MRI is particularly sensitive to iron deposition in tissues, due to the changes it induces in the magnetic field.

Earlier studies demonstrated that increased iron accumulation in amyloid plaques induces a magnetic susceptibility effect. This is visible as hypointens foci on T2*-weighted or susceptibility-weighted (SW) MRI in the cerebral cortex of transgenic AD mouse models and in human post-mortem brain slices (Chamberlain et al., 2011; Meadowcroft et al., 2009; van Rooden et al., 2009). The high magnetic field strengths needed to obtain these results only recently became available for in vivo human use. These high field human MRI systems (> 7 Tesla) may offer new possibilities to specifically detect the neuropathological hallmarks of AD, with iron as main field of focus. Perhaps changes in iron distribution can be detected even at an earlier stage than the traditional hippocampal atrophy.

MRS is a non-invasive tool which can be used to measure the concentration of various brain metabolites in vivo (Marjanska et al., 2005; Oberg et al., 2008; Rupsingh et al., 2011). MRS uses MR to study the quantity of metabolites by measuring the interaction of a radiofrequency electromagnetic field with molecular nuclei inside an external high magnetic field (Azevedo et al., 2008). Measuring these metabolic changes in vivo, could help identify AD at an early stage since metabolic levels are believed to precede structural changes (Jack, Jr. et al., 2010).

Numerous MRS studies have been performed in transgenic (tg) mouse models of AD. Several tg mouse models are available that develop sim-

ilar, but not identical, pathology as compared to human AD. The results of previous MRS studies have shown AD-related abnormalities for several metabolites (Braakman et al., 2008; Chen et al., 2009; Choi et al., 2010; Dedeoglu et al., 2004; Marjanska et al., 2005; Oberg et al., 2008; von Kienlin M. et al., 2005; Westman et al., 2009; Xu et al., 2010). N-acetylaspartate (NAA) in the brain is predominantly present in neuronal cell bodies. Decreased NAA levels, indicating neuronal damage, have been found in tq mice in comparison to wild type (wt) mice (Chen et al., 2009; Choi et al., 2010; Dedeoglu et al., 2004; Marjanska et al., 2005; Oberg et al., 2008; von Kienlin M. et al., 2005). Myo inositol (mIns) and taurine play a role in osmoregulation and are mainly found in astrocytes of brain tissue. These metabolites were found to be higher in to mice than in wt mice (Chen et al., 2009; Choi et al., 2010; Dedeoglu et al., 2004; Marjanska et al., 2005; Westman et al., 2009). Glutamate (glu) is an excitatory neurotransmitter, involved in learning, memory formation, and cognition, which is found to be decreased in mice with AD (Braakman et al., 2008; Choi et al., 2010; Dedeoglu et al., 2004; Marjanska et al., 2005; Oberg et al., 2008; von Kienlin M. et al., 2005).

Tg mouse models allow monitoring of the pathological and metabolic changes from the onset of AD in a longitudinal study, which is an effective way to investigate the early changes in the AD brain. However, no longitudinal study has been performed on AD mouse models using the non-invasive technique of MRS. Following mice from birth and investigating metabolic changes using MRS, the early start of AD might be detected making treatment more effective and giving us more insight in the pathogenesis of this disease.

Scope of this thesis

The overall aim of this thesis was to investigate MRI-based early markers of AD. We focused on correlation of radiological findings in AD with histology and we used MRS to study metabolic changes in brains of transgenic mice with AD.

Chapter 2 describes the effects of prolonged formalin fixation on MRI signal of brain tissue. This is important because such long fixed material is more readily available than brain tissue which is fixed for less than a year. In chapter 3 we compare different histological techniques to visualize iron in human brain tissue. Selection of the best techniques is crucial for reliable histological-radiological studies to assess the value of brain iron as an MRI-based biomarker for AD.

Chapter 4 illustrates the relation between AD pathology and iron distribution in brain tissue of AD patients compared with normal aging subjects in different age groups. The difference of iron distribution between AD patients and aging was investigated in the frontal cortex.

Chapter 5 demonstrates a disturbed iron accumulation and myelin architecture in AD using MRI with histological correlation on ex vivo brain tissue.

In Chapter 6 we describe the first systematic longitudinal MRS study to investigate the differences in metabolic changes during development of AD in a transgenic mouse model.

Results of this thesis and recommendations for future studies are discussed in chapter 7.

Reference List

Alonso, A.C., Li, B., Grundke-Iqbal, I., Iqbal, K., 2008. Mechanism of tau-induced neurodegeneration in Alzheimer disease and related tauopathies. Curr. Alzheimer Res. 5, 375-384.

Andersen, K., Launer, L.J., Dewey, M.E., Letenneur, L., Ott, A., Copeland, J.R., Dartigues, J.F., Kragh-Sorensen, P., Baldereschi, M., Brayne, C., Lobo, A., Martinez-Lage, J.M., Stijnen, T., Hofman, A., 1999. Gender differences in the incidence of AD and vascular dementia: The EURODEM Studies. EURODEM Incidence Research Group. Neurology 53, 1992-1997.

Azevedo, D., Tatsch, M., Hototian, S.R., Bazzarella, M.C., Castro, C.C., Bottino, C.M., 2008. Proton spectroscopy in Alzheimer's disease and cognitive impairment no dementia: a community-based study. Dement. Geriatr. Cogn Disord. 25, 491-500.

Ballatore, C., Lee, V.M., Trojanowski, J.Q., 2007. Tau-mediated neurodegeneration in Alzheimer's disease and related disorders. Nat .Rev. Neurosci. 8, 663-672.

Bartzokis, G., 2011. Alzheimer's disease as homeostatic responses to age-related myelin breakdown. Neurobiol. Aging 32, 1341-1371.

Braakman, N., Oerther, T., de Groot, H.J., Alia, A., 2008. High resolution localized two-dimensional MR spectroscopy in mouse brain in vivo. Magn Reson. Med. 60, 449-456.

Brookmeyer, R., Johnson, E., Ziegler-Graham, K., Arrighi, H.M., 2007. Forecasting the global burden of Alzheimer's disease. Alzheimer's Dement. 3, 186-191.

Callahan, M.J., Lipinski, W.J., Bian, F., Durham, R.A., Pack, A., Walker, L.C., 2001. Augmented senile plaque load in aged female beta-amyloid precursor protein-transgenic mice. Am. J. Pathol. 158, 1173-1177.

Chamberlain, R., Wengenack, T.M., Poduslo, J.F., Garwood, M., Jack, C.R., Jr., 2011. Magnetic resonance imaging of amyloid plaques in transgenic mouse models of Alzheimer's disease. Curr. Med. Imaging Rev. 7, 3-7.

Chen, S.Q., Wang, P.J., Ten, G.J., Zhan, W., Li, M.H., Zang, F.C., 2009. Role of myo-inositol by magnetic resonance spectroscopy in early diagnosis of Alzheimer's disease in APP/PS1 transgenic mice. Dement. Geriatr. Cogn Disord. 28, 558-566.

Choi, J.K., Jenkins, B.G., Carreras, I., Kaymakcalan, S., Cormier, K., Kowall, N.W., Dedeoglu, A., 2010. Anti-inflammatory treatment in AD mice protects against neuronal pathology. Exp. Neurol. 223, 377-384.

Corder, E.H., Ghebremedhin, E., Taylor, M.G., Thal, D.R., Ohm, T.G., Braak, H., 2004. The biphasic relationship between regional brain senile plaque and neurofibrillary tangle distributions: modification by age, sex, and APOE polymorphism. Ann. N. Y. Acad. Sci. 1019, 24-28.

Crichton, R.R., Wilmet, S., Legssyer, R., Ward, R.J., 2002. Molecular and cellular mechanisms of iron homeostasis and toxicity in mammalian cells. J. Inorg. Biochem. 91, 9-18.

Dedeoglu, A., Choi, J.K., Cormier, K., Kowall, N.W., Jenkins, B.G., 2004. Magnetic resonance spectroscopic analysis of Alzheimer's disease mouse brain that express mutant human APP shows altered neurochemical profile. Brain Res. 1012, 60-65.

Doens, D., Fernandez, P.L., 2014. Microglia receptors and their implications in the response to amyloid beta for Alzheimer's disease pathogenesis. J. Neuroinflammation. 11, 48.

Dore, V., Villemagne, V.L., Bourgeat, P., Fripp, J., Acosta, O., Chetelat, G., Zhou, L., Martins, R., Ellis, K.A., Masters, C.L., Ames, D., Salvado, O., Rowe, C.C., 2013. Cross-sectional and longitudinal analysis of the relationship between Abeta deposition, cortical thickness, and memory in cognitively unimpaired individuals and in Alzheimer disease. JAMA Neurol. 70, 903-911.

Duyckaerts, C., Delatour, B., Potier, M.C., 2009. Classification and basic pathology of Alzheimer disease. Acta Neuropathol. 118, 5-36.

Ferri, C.P., Prince, M., Brayne, C., Brodaty, H., Fratiglioni, L., Ganguli, M., Hall, K., Hasegawa, K., Hendrie, H., Huang, Y., Jorm, A., Mathers, C., Me-

nezes, P.R., Rimmer, E., Scazufca, M., 2005. Global prevalence of dementia: a Delphi consensus study. Lancet 366, 2112-2117.

Fox, N.C., Freeborough, P.A., Rossor, M.N., 1996. Visualisation and quantification of rates of atrophy in Alzheimer's disease. Lancet 348, 94-97.

Frisoni, G.B., Fox, N.C., Jack, C.R., Jr., Scheltens, P., Thompson, P.M., 2010. The clinical use of structural MRI in Alzheimer disease. Nat. Rev. Neurol. 6, 67-77.

Grimm, A., Lim, Y.A., Mensah-Nyagan, A.G., Gotz, J., Eckert, A., 2012. Alzheimer's disease, oestrogen and mitochondria: an ambiguous relationship. Mol. Neurobiol. 46, 151-160.

Grundke-Iqbal, I., Iqbal, K., Quinlan, M., Tung, Y.C., Zaidi, M.S., Wisniewski, H.M., 1986. Microtubule-associated protein tau. A component of Alzheimer paired helical filaments. J. Biol. Chem. 261, 6084-6089.

Haacke, E.M., Cheng, N.Y., House, M.J., Liu, Q., Neelavalli, J., Ogg, R.J., Khan, A., Ayaz, M., Kirsch, W., Obenaus, A., 2005. Imaging iron stores in the brain using magnetic resonance imaging. Magn Reson. Imaging 23, 1-25.

Hardy, J., 2009. The amyloid hypothesis for Alzheimer's disease: a critical reappraisal. J. Neurochem. 110, 1129-1134.

Hardy, J., Selkoe, D.J., 2002. The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. Science 297, 353-356.

Harrington, C.R., Storey, J.M., Clunas, S., Harrington, K.A., Horsley, D., Ishaq, A., Kemp, S.J., Larch, C.P., Marshall, C., Nicoll, S.L., Rickard, J.E., Simpson, M., Sinclair, J.P., Storey, L.J., Wischik, C.M., 2015. Cellular Models of Aggregation-dependent Template-directed Proteolysis to Characterize Tau Aggregation Inhibitors for Treatment of Alzheimer Disease. J. Biol. Chem. 290, 10862-10875.

Howell, M.D., Bailey, L.A., Cozart, M.A., Gannon, B.M., Gottschall, P.E., 2015. Hippocampal administration of chondroitinase ABC increases

plaque-adjacent synaptic marker and diminishes amyloid burden in aged APPswe/PS1dE9 mice. Acta Neuropathol. Commun. 3, 54.

Hyman, B.T., Phelps, C.H., Beach, T.G., Bigio, E.H., Cairns, N.J., Carrillo, M.C., Dickson, D.W., Duyckaerts, C., Frosch, M.P., Masliah, E., Mirra, S.S., Nelson, P.T., Schneider, J.A., Thal, D.R., Thies, B., Trojanowski, J.Q., Vinters, H.V., Montine, T.J., 2012. National Institute on Aging-Alzheimer's Association guidelines for the neuropathologic assessment of Alzheimer's disease. Alzheimers Dement. 8, 1-13.

Jack, C.R., Jr., Knopman, D.S., Jagust, W.J., Shaw, L.M., Aisen, P.S., Weiner, M.W., Petersen, R.C., Trojanowski, J.Q., 2010. Hypothetical model of dynamic biomarkers of the Alzheimer's pathological cascade. Lancet Neurol. 9, 119-128.

Janicki, S.C., Schupf, N., 2010. Hormonal influences on cognition and risk for Alzheimer's disease. Curr. Neurol. Neurosci. Rep. 10, 359-366.

Kettenmann, H., Hanisch, U.K., Noda, M., Verkhratsky, A., 2011. Physiology of microglia. Physiol Rev. 91, 461-553.

Kidd, M., 1963. Paired helical filaments in electron microscopy of Alzheimer's disease. Nature 197, 192-193.

Lambracht-Washington, D., Rosenberg, R.N., 2015. A noninflammatory immune response in aged DNA Abeta42-immunized mice supports its safety for possible use as immunotherapy in AD patients. Neurobiol. Aging 36, 1274-1281.

Landlinger, C., Oberleitner, L., Gruber, P., Noiges, B., Yatsyk, K., Santic, R., Mandler, M., Staffler, G., 2015. Active immunization against complement factor C5a: a new therapeutic approach for Alzheimer's disease. J. Neuroinflammation. 12, 150.

Ma, Y., Zhang, S., Li, J., Zheng, D.M., Guo, Y., Feng, J., Ren, W.D., 2014. Predictive accuracy of amyloid imaging for progression from mild cognitive impairment to Alzheimer disease with different lengths of follow-up: a systematic review. Medicine (Baltimore) 93, e150. Marjanska, M., Curran, G.L., Wengenack, T.M., Henry, P.G., Bliss, R.L., Poduslo, J.F., Jack, C.R., Jr., Ugurbil, K., Garwood, M., 2005. Monitoring disease progression in transgenic mouse models of Alzheimer's disease with proton magnetic resonance spectroscopy. Proc. Natl. Acad. Sci. U.S.A 102, 11906-11910.

Meadowcroft, M.D., Connor, J.R., Smith, M.B., Yang, Q.X., 2009. MRI and Histological Analysis of Beta-Amyloid Plaques in Both Human Alzheimer's Disease and APP/PS1 Transgenic Mice. Journal of Magnetic Resonance Imaging 29, 997-1007.

Meadowcroft, M.D., Connor, J.R., Yang, Q.X., 2015a. Cortical iron regulation and inflammatory response in Alzheimer's disease and APPSWE/ PS1DeltaE9 mice: a histological perspective. Front Neurosci. 9, 255.

Meadowcroft, M.D., Peters, D.G., Dewal, R.P., Connor, J.R., Yang, Q.X., 2015b. The effect of iron in MRI and transverse relaxation of amyloid-beta plaques in Alzheimer's disease. NMR Biomed. 28, 297-305.

Minati, L., Edginton, T., Bruzzone, M.G., Giaccone, G., 2009. Current concepts in Alzheimer's disease: a multidisciplinary review. Am. J. Alzheimers Dis. Other Demen. 24, 95-121.

Musicco, M., 2009. Gender differences in the occurrence of Alzheimer's disease. Funct. Neurol. 24, 89-92.

Nasrallah, I.M., Wolk, D.A., 2014. Multimodality imaging of Alzheimer disease and other neurodegenerative dementias. J. Nucl. Med. 55, 2003-2011.

Natte, R., Maat-Schieman, M.L., Haan, J., Bornebroek, M., Roos, R.A., van Duinen, S.G., 2001. Dementia in hereditary cerebral haemorrhage with amyloidosis-Dutch type is associated with cerebral amyloid angiopathy but is independent of plaques and neurofibrillary tangles. Ann. Neurol. 50, 765-772.

Oberg, J., Spenger, C., Wang, F.H., Andersson, A., Westman, E., Skoglund, P., Sunnemark, D., Norinder, U., Klason, T., Wahlund, L.O., Lindberg, M., 2008. Age related changes in brain metabolites observed by 1H MRS in

APP/PS1 mice. Neurobiol. Aging 29, 1423-1433.

Peters, D.G., Connor, J.R., Meadowcroft, M.D., 2015. The relationship between iron dyshomeostasis and amyloidogenesis in Alzheimer's disease: Two sides of the same coin. Neurobiol. Dis. 81, 49-65.

Price, J.L., Davis, P.B., Morris, J.C., White, D.L., 1991. The distribution of tangles, plaques and related immunohistochemical markers in healthy aging and Alzheimer's disease. Neurobiol. Aging 12, 295-312.

Qiu, C., Kivipelto, M., von, S.E., 2009. Epidemiology of Alzheimer's disease: occurrence, determinants, and strategies toward intervention. Dialogues. Clin. Neurosci. 11, 111-128.

Richter, M., Mewes, A., Fritsch, M., Krugel, U., Hoffmann, R., Singer, D., 2014. Doubly Phosphorylated Peptide Vaccines to Protect Transgenic P301S Mice against Alzheimer's Disease Like Tau Aggregation. Vaccines. (Basel) 2, 601-623.

Rupsingh, R., Borrie, M., Smith, M., Wells, J.L., Bartha, R., 2011. Reduced hippocampal glutamate in Alzheimer disease. Neurobiol. Aging 32, 802-810.

Smith, M.A., Harris, P.L., Sayre, L.M., Perry, G., 1997. Iron accumulation in Alzheimer disease is a source of redox-generated free radicals. Proc. Natl. Acad. Sci. U.S.A 94, 9866-9868.

Takahashi, R.H., Capetillo-Zarate, E., Lin, M.T., Milner, T.A., Gouras, G.K., 2010. Co-occurrence of Alzheimer's disease amyloid and tau pathologies at synapses. Neurobiol.Aging 31, 1145-1152.

Thal, D.R., Attems, J., Ewers, M., 2014. Spreading of amyloid, tau, and microvascular pathology in Alzheimer's disease: findings from neuro-pathological and neuroimaging studies. J. Alzheimers Dis. 42 Suppl 4, S421-S429.

van Duijn, S., Nabuurs, R.J., van Duinen, S.G., Natte, R., 2013. Comparison of histological techniques to visualize iron in paraffin-embedded brain tissue of patients with Alzheimer's disease. J. Histochem. Cytochem. 61,

785-792.

van Rooden, S., Maat-Schieman, M.L., Nabuurs, R.J., van der Weerd, L., van Duijn, S., van Duinen, S.G., Natte, R., van Buchem, M.A., van der Grond, J., 2009. Cerebral amyloidosis: postmortem detection with human 7.0-T MR imaging system. Radiology 253, 788-796.

Villemagne, V.L., Burnham, S., Bourgeat, P., Brown, B., Ellis, K.A., Salvado, O., Szoeke, C., Macaulay, S.L., Martins, R., Maruff, P., Ames, D., Rowe, C.C., Masters, C.L., 2013. Amyloid beta deposition, neurodegeneration, and cognitive decline in sporadic Alzheimer's disease: a prospective cohort study. Lancet Neurol. 12, 357-367.

von Kienlin M., Kunnecke, B., Metzger, F., Steiner, G., Richards, J.G., Ozmen, L., Jacobsen, H., Loetscher, H., 2005. Altered metabolic profile in the frontal cortex of PS2APP transgenic mice, monitored throughout their life span. Neurobiol. Dis. 18, 32-39.

Wang, H., Megill, A., He, K., Kirkwood, A., Lee, H.K., 2012. Consequences of inhibiting amyloid precursor protein processing enzymes on synaptic function and plasticity. Neural Plast. 2012, 272374.

Wang, J., Tanila, H., Puolivali, J., Kadish, I., van, G.T., 2003. Gender differences in the amount and deposition of amyloidbeta in APPswe and PS1 double transgenic mice. Neurobiol. Dis. 14, 318-327.

Weller, R.O., Preston, S.D., Subash, M., Carare, R.O., 2009. Cerebral amyloid angiopathy in the aetiology and immunotherapy of Alzheimer disease. Alzheimers Res. Ther. 1, 6.

Westman, E., Spenger, C., Oberg, J., Reyer, H., Pahnke, J., Wahlund, L.O., 2009. In vivo 1H-magnetic resonance spectroscopy can detect metabolic changes in APP/PS1 mice after donepezil treatment. BMC. Neurosci. 10, 33.

Wischik, C.M., Harrington, C.R., Storey, J.M., 2014. Tau-aggregation inhibitor therapy for Alzheimer's disease. Biochem. Pharmacol. 88, 529-539. Wischik, C.M., Staff, R.T., Wischik, D.J., Bentham, P., Murray, A.D., Storey, J.M., Kook, K.A., Harrington, C.R., 2015. Tau aggregation inhibitor therapy: an exploratory phase 2 study in mild or moderate Alzheimer's disease. J. Alzheimers Dis. 44, 705-720.

Wittchen, H.U., Jacobi, F., Rehm, J., Gustavsson, A., Svensson, M., Jonsson, B., Olesen, J., Allgulander, C., Alonso, J., Faravelli, C., Fratiglioni, L., Jennum, P., Lieb, R., Maercker, A., van, O.J., Preisig, M., Salvador-Carulla, L., Simon, R., Steinhausen, H.C., 2011. The size and burden of mental disorders and other disorders of the brain in Europe 2010. Eur. Neuropsychopharmacol. 21, 655-679.

Xu, W., Zhan, Y.Q., Huang, W., Wang, X.X., Zhang, S.M., Lei, H., 2010. Reduction of Hippocampal N-Acetyl Aspartate Level in Aged APP(Swe)/ PS1(dE9) Transgenic Mice Is Associated With Degeneration of CA3 Pyramidal Neurons. Journal of Neuroscience Research 88, 3155-3160.