

Functional and metabolic characterization of endothelial cells in chronic thromboembolic pulmonary hypertension

Smolders, V.F.E.D.

Citation

Smolders, V. F. E. D. (2020, December 3). *Functional and metabolic characterization of endothelial cells in chronic thromboembolic pulmonary hypertension*. Retrieved from https://hdl.handle.net/1887/138244

Note: To cite this publication please use the final published version (if applicable).

Cover Page

Universiteit Leiden

The handle<http://hdl.handle.net/1887/138244> holds various files of this Leiden University dissertation.

Author: Smolders, V.F.E.D. **Title**: Functional and metabolic characterization of endothelial cells in chronic thromboembolic pulmonary hypertension **Issue date**: 2020-12-03

CHAPTER 2

ENDOTHELIAL DYSFUNCTION IN PULMONARY HYPERTENSION: CAUSE OR CONSEQUENCE?

Valérie Françoise Smolders*, Kondababu Kurakula*, Olga Tura-Ceide,

J. Wouter Jukema, Paul H. A. Quax, Marie-José Goumans

*Both authors contributed equally

Submitted

Chapter 2 ǀ

ABSTRACT

Pulmonary arterial hypertension (PAH) is a rare, complex, and progressive disease characterized by abnormal remodelling of the pulmonary arteries that leads to right ventricular failure and death. Although our understanding of the causes for abnormal vascular remodelling in PAH is limited, accumulating evidence indicates that endothelial cell (EC) dysfunction is one of the first triggers initiating this process. EC dysfunction leads to the activation of several cellular signalling pathways in the endothelium, resulting in uncontrolled proliferation of ECs, pulmonary artery smooth muscle cells and fibroblasts, and eventually leads to vascular remodelling and occlusion of the pulmonary blood vessels. Other factors that are related to EC dysfunction in PAH are an increase in endothelial to mesenchymal transition, inflammation, apoptosis, and thrombus formation. In this review, we outline the latest advances on the role of EC dysfunction in PAH and other forms of pulmonary hypertension. We also elaborate on the molecular signals that orchestrate EC dysfunction in PAH. Understanding the role and mechanisms of EC dysfunction will unravel the therapeutic potential of targeting this process in PAH.

Keywords: Pulmonary hypertension – endothelial cell dysfunction – vasoactive factors – TGFβ – EndoMT – epigenetics

INTRODUCTION

Pulmonary hypertension (PH) is a condition defined by a mean pulmonary arterial pressure of more than 20 mmHg at rest and 30 mmHg during exercise. The range of genetic, molecular, and humoral causes that can lead to this increase in pressure is extensive. Therefore, PH is grouped into different classes based on clinical and pathological findings as well as therapeutic interventions $1,2$. The World Health Organization (WHO) classifies PH into five groups, namely: 1. Pulmonary arterial hypertension (PAH), 2. Pulmonary hypertension due to left heart disease (PH-LHD), 3. Pulmonary hypertension due to lung disease (PH-LD), 4. Chronic thromboembolic pulmonary hypertension (CTEPH), 5. Pulmonary hypertension due to unclear and/or multifactorial mechanisms $1,3,4$. PH is becoming more and more a global health issue due to the ageing population. Although PH-LHD and PH-LD are the most prevalent PH groups, research and drug development focuses mainly on PAH and CTEPH, which are rarer diseases that affect mainly younger people ⁵. Because of the amount of research conducted in PAH compared to the other four groups, this review will focus mostly on PAH.

PAH is characterized by remodelling of distal pulmonary arteries, causing a progressive increase in vascular resistance. Vascular remodelling is associated with alterations in vasoconstriction, pulmonary artery- endothelial cells (PAECs) and -smooth muscle cells (PASMCs) cell proliferation, inflammation, apoptosis, angiogenesis and thrombosis, which leads to muscularization and occlusion of the lumen of pulmonary arteries by formation of vascular lesions. Plexiform lesions are the most common lesions in PAH, characterized by deregulated endothelial cell (EC) proliferation. Other lesions in PH are thrombotic lesions and neointima formation, which form a layer of myofibroblasts and extracellular matrix between the endothelium and the external elastic lamina ⁶. One of the first triggers for development of PAH is EC injury triggering the activation of cellular signalling pathways that are not yet completely understood.

In normal conditions the endothelium is in a quiescent and genetically stable state. However, different types of injury can activate the endothelium. When activated, the endothelium secretes different growth factors and cytokines that affect EC and SMC proliferation, apoptosis, coagulation, attract inflammatory cells or affect vasoactivity to restore homeostasis. EC dysfunction, the loss of cellular functions leading to pathological changes, is crucial in the development of cardiovascular diseases and so too in PAH 7,8 . Many different

factors have been suggested to be involved in the initiation of EC dysfunction in PAH, like shear stress, hypoxia, inflammation, cilia length, and genetic factors (**Figure 1**) 6,9-11. In PAH the endothelium switches from a quiescent to an overactive state where it starts to secrete vasoconstrictive factors like endothelin-1 (ET-1) 12 and thromboxane 13 , and proliferative factors like vascular endothelial growth factor (VEGF) and reduce the secretion of vasodilators like nitric oxide (NO) and prostacyclin, indicating that EC dysfunction might play an central role in the pathogenesis of PAH 7,14.

The purpose of this review is to provide a state-of-the-art overview on EC dysfunction in PAH and to highlight current progress made in understanding this phenomenon. At last, this review discusses several models for studying EC dysfunction in PH and explores possible molecular targets and drugs for restoring EC function in PH.

Figure 1. Pulmonary artery remodelling, vascular resistance and pulmonary arterial hypertension (PAH) development. PAH results from a progressive increase in vascular resistance caused by pulmonary vascular remodelling. Molecular mechanisms behind the process of vascular remodelling are still not fully elucidated but endothelial cell (EC) injury is thought to be one of the early triggers. EC injury can be caused by shear stress, hypoxia and inflammation. Host factors such as genetic mutations and gender but also epigenetic factors and comorbidities are thought to play an important role in EC dysfunction. EC dysfunction leads to altered cell signalling that induces cellular processes such as EndoMT, apoptosis and proliferation. In addition, changes are found in cell metabolism and in the secretion of vasoactive, coagulation and thrombotic factors. Also vascular smooth muscle cells and fibroblasts are found to display a diseased cellular phenotype. EC dysfunction eventually promotes vasoconstriction, thrombus formation, neointima formation, muscularization and development of vascular lesions. As lumen size decreases, pulmonary vascular resistance increases and induces right ventricle (RV) hypertrophy with eventually RV failure.

FACTORS CONTRIBUTING TO EC DYSFUNCTION IN PAH

Approximately 80% of familial PAH (hPAH) and 20% of idiopathic cases of PAH (iPAH) are associated with mutations in the bone morphogenic type 2 receptor (BMPR2) but a penetrance of 20-30% suggests secondary stimuli such as endothelial to mesenchymal transition (EndoMT), inflammation, thrombosis, apoptosis and perturbations in vasoactivity as important contributors to EC dysfunction and PAH development 15-17.

Bone morphogenic type 2 receptor

BMPR2 encodes for a transmembrane serine/threonine kinase receptor belonging to the transforming growth factor-β (TGFβ) family of signalling proteins (**Figure 2**) 18. BMPR2 modulates cellular growth, apoptosis, inflammation and differentiation via binding of bone morphogenetic proteins (BMPs) to a heteromeric complex of a BMP type-I receptor and BMPR2, in a time, concentration and cell type dependent manner ¹⁹. Depending on the localization in the vascular bed, BMPR2 promotes survival of PAECs, while it has an antiproliferative effect on PASMCs 20-22.

To date over 380 PAH related mutations in *BMPR2* are known, mostly loss of function mutations 23,24. Low penetrance of disease development associated with *BMPR2* mutations observed in humans has also been confirmed in experimental models of PH, where *BMPR2* deletion alone does not induce PAH in the majority of the cases $25-27$. Interestingly, reduced levels of BMPR2 have also been found in PH patients without *BMPR2* mutations, suggestion additional involvement of genetic modifiers or environmental factors reducing BMPR2 dependent signaling 28-31.

BMPR2 is predominantly present in ECs lining the vascular lumen in the lung and expression is reduced in ECs from PH lung. Therefore mutated *BMPR2* is postulated to play a significant role in EC dysfunction in PAH $24,28$. Association between endothelial BMPR2 and PAH development was further supported by the observation that mice with endothelial specific deletion of *BMPR2* were prone to develop PAH 32,33. PAECs overexpressing a kinase-inactive BMPR2 mutant show increased susceptibility to apoptosis and conditioned medium from these PAECs stimulated proliferation of PASMCs via increased release of TGFβ1 and fibroblast growth factor (FGF)-2 34. BMP9 administration selectively enhanced endothelial *BMPR2* and reversed PH in rats 35. In line with these findings, several compounds attenuated EC dysfunction via increased BMPR2 signalling and reduced abnormal remodelling in experimental PH36-38. Moreover, BMPR2 acts as a gatekeeper to protect ECs from increased TGF β responses and integrin-mediated mechano-transduction 39 .

Loss of endothelial BMPR2 promotes release of pro-inflammatory cytokines in a SOD3 dependent manner, allowing leukocyte transmigration to underlying tissues, causing further vascular remodelling *in vivo* 25,40,41. Furthermore, loss of BMPR2 signalling in PAECs promotes a pro-inflammatory state during normoxia by enhancing mitochondrial biogenesis, mitochondrial potential and promoting glycolysis 42. BMPR2 deficiency in iPAH PAECs lacking BMPR2 are associated with loss of DNA damage control via reduced DNA repair related genes such as BRCA1. Increased DNA damage reciprocally leads to further reduction of BMPR2 expression and EC dysfunction 43. Transcriptome analysis of PAECs from iPAH patients revealed a correlation between reduced BMPR2 levels and downregulation of β-catenin, resulting in reduced Collagen-4 (COL4) and ephrinA1 (EFNA1) expression 44. Both COL4 and EFNA1 perform intertwining roles in endothelium structure. siRNA mediated silencing of *BMPR2* in PAECs resulted in increased PAEC proliferation, migration, and disruption of cytoskeletal architecture. One of the changes observed was increase in Ras/Raf/ERK signalling, and Ras inhibitors, like nintedanib 45 , reversed the enhanced proliferation and hypermotility of BMPR2 silencing in PAECs 46.

Carboxylesterase-1 (CES1) promotes BMP signalling by ensuring proper trafficking of BMPR2 from the endoplasmic reticulum (ER) to the plasma membrane 47 . CES1 is reduced in iPAH patients and impaired ER trafficking will result in decreased BMPR2 availability 47.

Pro-inflammatory cytokines, such as IL-6 and $TNF\alpha$, have also been found to downregulate BMPR2 expression in PAECs via a STAT3-miR-(Cluster 17/92) and NF-κB-p65 pathway,

respectively 29,48. Finally, miRNA-21, although primarily induced by BMPR2 signalling, negatively targets BMPR2 expression 49.

Figure 2. TGF-β superfamily signalling in PAH. The TGF-β superfamily is subdivided into the TGF-β group that include TGFβ, Nodal and activins and the BMP group that includes BMPs. Both groups signal through intracellular mediators, known as Smads. Receptor-regulated Smads (R-Smads) are phosphorylated by type-1 receptors (e.g. ALK1/2/3/6 and ALK4/5/7) and form complexes with a Common mediator Smad (Co-Smad). Subsequently, these complexes translocate into the nucleus where they induce transcriptional responses that alter gene expression of specific targets that influence apoptosis, cell differentiation, inflammation and proliferation. Inhibitory Smads (I-Smads) negatively regulate TGF-β and BMP signalling. Both TGF-β and BMP receptors can also signal independently from Smads and alter downstream cell-specific processes. It is know that TGF-β superfamily signalling plays an important role the initiation of EndoMT by triggering overexpression of genes like TWIST1, αSMA and phospho-vimentin.

Endothelial to mesenchymal transition

EndoMT is a phenomenon where ECs acquire a mesenchymal-like phenotype which is accompanied with loss of endothelial markers and gain of mesenchymal markers. In addition, ECs lose cell-cell contact, change their morphology and adopt a highly migratory and invasive phenotype (**Figure 3A**) 50,51. In lungs of human PAH patients and monocrotaline (MCT) and Sugen/hypoxia (SuHx) experimental PH rat models, EndoMT was observed whereby cells express high levels of α -SMA and activated phospho-vimentin and VE-cadherin, indicating their endothelial origin 52-54. Moreover, TWIST1, a key transcription factor in inducing EndoMT, is highly expressed in human PAH lungs compared to healthy lungs 52.

TGFβ treatment of PAECs induces expression of the EndoMT transcription factors TWIST1 and SNAIL1^{50,55} and the mesenchymal markers α -SMA and phospho-vimentin⁵⁶. TWIST1 increases expression of TGFβ, leading to enhanced TGFβ signalling57. In addition, reduced BMPR2 signalling promotes EndoMT via upregulation of the High Mobility Group AT-hook 1 and its target gene SLUG, independently of TGFβ signalling 58. More interestingly, BMP-7, a protein previously described as having anti-inflammatory and anti-tumour effects in several diseases, was attenuated hypoxia-induced EndoMT in PAECs both *in vivo* and *in vitro* by inhibiting the m-TORC1 signalling pathway59. BMPR2 favours EndoMT allowing cells of myo-fibroblastic character to create a vicious feed-forward process leading to hyperactivated TGFβ signalling39. In summary, alterations in TGFβ/BMP signalling are linked to the process of EndoMT observed in PAH 60.

Hypoxia is also an inducer of EndoMT through hypoxia-inducible transcription factor-1α (HIF-1α) and HIF-2α, and both transcription factors are increased in PAH $61,62$. PAH ECs display increased expression of HIF-2α, leading to SNAIL upregulation 54. In addition, HIF-1α knockdown alone effectively blocks hypoxia-induced EndoMT but also knockdown of its downstream target gene TWIST1 showed effective blockage of hypoxia-induced EndoMT in microvascular ECs (MVECs), however less pronounced⁶³. Nonetheless, microvascular endothelium may differ from arterial endothelial function.

Inflammation

Pulmonary arteries of PAH patients showed infiltration of macrophages, dendritic cells and lymphocytes into the plexiform lesions and an increased migration of monocytes^{9,64}. Increased levels of pro-inflammatory cytokines and chemokines, such as IL-1β, TNFα and IL-6, known activators of vascular endothelium, were found (**Figure 3B**) 37,65,66. IL-1β stimulates endothelial ET-1 production 67. Administration of IL-6 to experimentally induced PAH in a rat model and overexpression of IL-6 in transgenic mice led to occlusion of pulmonary arteries and RV hypertrophy 68,69 . IL-33 has a dual role as cytokine and a role in the nucleus 70 . Nuclear IL-33 is expressed in nuclei of healthy ECs but is less expressed in nuclei of ECs from iPAH lungs. Nuclear IL-33 modulates gene expression of pro-inflammatory cytokines and IL-33 knock-down in PAECs upregulates expression of IL-6. Therefore, loss of nuclear IL-33 could contribute to EC dysfunction in PAH 70 . Additionally IL-33 may contribute to inflammatory activation of the endothelium by promoting endothelial production of granulocyte macrophage-colony stimulating factor (GM-CSF) and macrophage-CSF 71 . Hypoxia induces expression of IL-33 and its receptor ST2 on ECs, leading to EC and SMC dysfunction with concomitant PH development 72.

Figure 3. Endothelial to mesenchymal transition (EndoMT) and endothelial dysfunction in PAH. **A)** Upon activation by transcriptional factors, hypoxia, haemodynamic forces, inflammation and TGF-β/BMP pathway signaling pulmonary endothelial cells (PAECs) undergo cellular transition to a mesenchymal phenotype. During transition, PAECs lose endothelial markers and gain mesenchymal markers such as αSMA and TWIST. These mesenchymal-like cells also gain mesenchymal characteristics that trigger vascular remodeling and PAH pathogenesis. **B)** Upon endothelial cell injury, PAECs become dysfunctional and alter their secretion of cytokines and other factors that regulate coagulation, thrombosis and vascular tone. A failure of PAECs in maintaining vessel homeostasis promotes vasoconstriction, thrombosis and inflammation that initiate PAH disease progression.

Thrombosis and coagulation

In situ pulmonary artery thrombosis is regularly found in PAH. However, it remains unclear if thrombosis due to EC dysfunction causes progression of PAH or whether it forms as a result of it. Various factors, e.g. von Willebrand factor, plasminogen activator inhibitor-1 and tissue factor (TF), secreted by EC, have been implicated in coagulation and are found differently expressed in PAH (**Figure 3B**) 73-75.

Recently TF has emerged as an interesting target involved in the pathogenesis of PAH. TF is a glycoprotein expressed on the cell surface of SMCs, macrophages, monocytes and ECs 76 . It plays a role in initiation of coagulation, facilitation of angiogenesis and mediation of arterial injury in the circulation $77,78$. Interestingly, TF is rarely expressed in healthy cells, but is highly expressed in PAECs in PAH, predominantly in plexiform lesions 79 . In PAH animal models, increased TF expression correlates with formation of plexiform-lesions 79 . Furthermore, in PAH patients elevated levels of thrombin, a downstream target of TF and essential for clot formation, are detected $74,80$. Elevated levels of fibrinopeptide-A (FPA) which increases thrombin activity (downstream of TF) are observed in PAH patients 74 . PAH patients have lower thrombomodulin levels, consistent with the hypercoagulable state observed in PAH patients⁸¹. However, the role of EC dysfunction in this is still unclear.

Apoptosis

EC apoptosis may also play a role in PH development via vascular dropout and selection pressure on ECs, contributing to the apoptosis-resistant phenotype of ECs in vascular lesions $82.$ Several attempts were made to elucidate the molecular pathways involved in regulation of PAEC apoptosis. The hypothesis is that disturbed responses to VEGF signalling in combination with hypoxia cause an initial increase in apoptosis in PAECs, leading to the emergence of aggressive apoptosis resistant and hyperproliferative ECs that cause formation of intimal lesions 83-85. A possible explanation for the initial increase in apoptosis of PAECs is that loss of BMPR2 signalling promotes mitochondrial dysfunction and subsequent PAEC apoptosis 42. White *et al*., interestingly, proposes a model in which the pro-apoptotic factor programmed cell death-4 (PDCD4) activates cleavage of caspase-3, inducing PAEC apoptosis. Interestingly, they show that reducing PDCD4 levels *in vivo* by overexpressing miRNA-21 prevents PH development in SuHx rats 86. Besides an initial increase in apoptosis, PAH is also characterized by PAECs that are hyperproliferative and apoptosis resistant 85 . PAECs from iPAH patients showed increased expression of pro-survival factors IL-15, BCL-2 and Mcl-1, together with persistent activation of the pro-survival STAT3 signalling pathway ⁸⁵. Furthermore, in lungs from iPAH patients and from SuHx rats Notch1 was elevated. Notch1 contributes to PAH pathogenesis by increasing EC proliferation and inhibiting apoptosis via p21downregulation and regulating BCL-2 and survivin expression. Furthermore, HIF1 α expression promotes Notch signalling human PAECs ⁸⁷.

VASOACTIVE FACTORS CONTRIBUTING TO EC DYSFUNCTION

PAH is characterized by an activated endothelium of which the balance between vasodilation and vasoconstriction, but also growth factor production, is altered, causing perturbations in pulmonary vascular homeostasis that promote vascular remodelling (**Figure 3B**).

Nitric oxide

NO is a fast-reacting endogenous free radical produced by endothelial NO Synthase (eNOS). NO is essential for vasorelaxation via PASMCs but also has antithrombotic effects and controls EC differentiation and growth $88-90$. NO has long been implicated in the pathogenesis of PAH, and lungs of PAH patients have reduced NO expression 91 . However, other studies reported contradictory results and some PH patients even show an increase in eNOS expression ^{91,92}. Furthermore, eNOS^{-/-} mice show reduced vascular remodelling after chronic hypoxia caused by reduced vascular proliferation ⁹³.

Next to hypoxia as a known regulator of eNOS expression, increasing evidence supports the involvement of epigenetic regulations such as histone acetylation and DNA methylation in expression eNOS. This is independent of the initial hypoxic environment. Experimental models of persistent pulmonary hypertension of the new-born (PPHN) and PPHN PAECs showed that epigenetic modifications can contribute to reduced eNOS expression and subsequent PPHN pathogenesis $94,95$. However, it is unclear whether such mechanisms exist in PAH pathogenesis 96-100.

Reduced NO availability can also be caused by freely circulating endogenous eNOS inhibitors 101 , such as asymmetric dimethylarginine (ADMA) 102 . The metabolism of this protein is facilitated by dimethylarginine dimethylaminohydrolase (DDAH)¹⁰³. Increased levels of ADMA are associated with the pathogenesis of PH 101,103,104 , and hypoxia-induced increase of miRNA-21 is found to reduce DDAH activity ^{49,104}.

Prostacyclin

Prostacyclin is another important vasodilator produced by EC with additional antithrombotic and antiproliferative properties $^{7,105-107}$. Prostacyclin is synthesized from arachidonic acid, by prostacyclin synthase and cyclo-oxygenase (COX) 108 . Decreased prostacyclin levels are measured in various patients with different forms of PAH, like iPAH and HIV-associated PAH $7,109$ explaining in part the increase in pulmonary vasoconstriction, SMC proliferation and

coagulation occurring in these patients. In experimental PAH models, mice overexpressing prostacyclin synthase are protected from developing chronic hypoxia-induced PAH 110.

Endothelin-1

ET-1 is a potent vasoconstrictor, mainly synthesized in EC, but also in smaller amounts in vSMCs, macrophages, fibroblasts, myocytes and epithelial cells $7,111,112$. The lungs show the highest level of ET-1 in the entire body ¹¹³. ET-1 stimulates vSMCs proliferation and platelet aggregation 7,106 . ET-1 exhibits its effects by binding to the ET_A and ET_B receptors, which activate signalling pathways in vSMCs regulating proliferation, vasorelaxation and vasoconstriction $107,113$. ET_A is predominantly expressed on vSMCs and is involved in vasoconstriction and proliferation of these cells, while ET_B is expressed on vSMCs and PAECs, and is involved in stimulating the release of vasodilators, like NO and prostacyclin, and inhibition of apoptosis ^{67,107,111,113,114}. Expression of ET-1 and its receptors is increased in lungs of PAH patients and experimental PH models 115-118. Furthermore, a correlation exists between expression of ET-1 and increase in pulmonary resistance in PAH 117. Multiple PAH associated factors are able to increase ET-1 expression including hypoxia, cytokines, growth factors, TGFB/BMP signalling and shear stress ¹¹⁹⁻¹²³. Increased synthesis of endothelial ET-1, accompanied with an increase in expression of ET_A on PASMCs likely contributes to the increased vasoconstriction and vascular remodelling observed in PAH 106,118,124.

Thromboxane

Thromboxane A2, produced by ECs and platelets, is a vasoconstrictor, inducer of platelet aggregation and a vSMCs mitogen 7,13 . Its production is increased by hypoxia and oxygen metabolites $125,126$. In PAH thromboxane A₂ is increased, creating an imbalance that might contribute to excessive platelet aggregation and vascular remodelling observed in PAH 13.

Vascular endothelial growth factor

VEGF is an angiogenic factor secreted by ECs. VEGF has multiple roles in maintaining lung structure and homeostasis but also is associated with several vascular disorders 7,127,128. The pulmonary endothelium does not secrete VEGF during normal homeostasis but iPAH ECs from plexiform lesions show increased expression of VEGF and VEGF receptor 2 129, and also VEGF plasma levels of PH patients are elevated 130. The relation between PAH and increased VEGF expression is still poorly understood. It is suggested that VEGF levels in PAECs are elevated in early stages of PAH as a protective response, while during disease progression VEGF keeps

promoting growth of PAECs, causing the formation of plexiform lesions ⁷. Rats treated with a VEGF receptor blocker in combination with hypoxia develop angio-obliterative PAH 131. Furthermore, overexpression of VEGFA slows down the development of hypoxia-induced PAH, and improves endothelial function by increasing eNOS activity among others 132.

EPIGENETICS

In recent years epigenetics has become a growing field of interest in PAH research. Currently the main focus of study for targeting PAH are the following three mechanisms of epigenetic regulation; DNA methylation, histone modifications and RNA interference (**Figure 4**) 14. DNA methylation profiling of PAECs from iPAH and hPAH patients revealed differences in expression of several genes involved in inflammatory processes, remodelling and lipid metabolism compared to controls 133. Among those genes ABCA1 was found most differently methylated/ downregulated in the discrimination between PAH and controls. ABCA1 belongs to the family of ATP binding cassette (ABC) transporters that are important for pulmonary homeostasis 133. Furthermore, ABCA1 is linked to PAH pathophysiology in a MCT animal model of PAH where activation of ABCA1 improved RV hypertrophy and pulmonary haemodynamics 14,133.

Figure 4. Epigenetics in PAH. In addition to genetic variations and other risk factors such as gender, comorbidities and environmental factors, epigenetic variations in PAH gain interest. Differences in DNA methylation profiles, increased histone acetylation and dysregulated miRNA expression in PAH patients point out a growing field in PAH research that provides better understanding of disease pathology.

Increased histone acetylation through histone-deacetylases (HDAC) is associated with vascular remodelling found in PAH 134,135. In humans, HDAC enzymes are divided into 4 classes: class-1 HDACs (HDAC-1, -2, -3 and -8), class-2a HDACs (HDAC-4, -5, -7 and -9), class-2b HDACs (HDAC-6 and -10), class-3 HDACs (Sir2-like proteins) and class-4 HDACs (HDAC-11) 136. HDAC-1 and -5 show increased expression in both lungs of iPAH patients and chronic hypoxic rats whereas HDAC-4 was only increased in human iPAH lungs 135. More recently HDAC-6 is linked to PAH pathogenesis, possibly through upregulation of HSP90¹³⁷. HDAC-6 was overexpressed in PAECs and PASMCs of PAH patients and PH experimental models 138. In the SuHx and MCT rat model pharmacological HDAC-6 inhibition improved PH 138. Several other studies showed that class-1 HDAC inhibitors attenuate PAH by suppressing arterial remodelling in a chronic hypoxia model and by reducing inflammation in PH-fibroblasts 135,139,140. In PAECs class-2a HDAC inhibitors restore the levels of myocyte-enhancer-factor-2 and attenuate PAH in both the MCT and SuHx PAH rat models ¹⁴¹.

The epigenetic regulator bromodomain-containing-protein-4 (BRD4) is linked to the pathogenesis of PAH 38. BRD4 is a member of the Bromodomain and Extra-Terminal (BET) motif family that binds histones to influence gene expression 142. BRD4 is overexpressed in lungs of PAH patients in a miR-204 dependent manner. It inhibits apoptosis by sending cell survival signals ^{38,143}, and stimulates proliferation of PAEC and PASMC proliferation at these sites ^{14,143}. Selective inhibition of BRD4 with RVX-208 restored EC function, reversed PAH in the MCT and SuHx rat models, and supported the RV function in pulmonary artery banding model of PAH 38.

EC DYSFUNCTION IN OTHER PH GROUPS

Group 2 PH

Group 2 PH is due to a complication of left heart disease and is most common in patients with heart failure (HF). Therefore research in group 2 PH focuses mostly on left ventricular dysfunction and not so much the lung vasculature. However, EC dysfunction is also associated with PH-LHD ¹⁴⁴. An experimental model of chronic HF showed reduced NO activity and responsiveness to NO in pulmonary arteries ¹⁴⁵. Moreover, ET-1 is elevated in certain PH-LHD phenotypes and ET-1 activity is increased in plasma of patients with chronic HF. Blocking the ET_A receptor caused pulmonary vasodilation in these patients 146,147 . Furthermore, polymorphisms found eNOS also contribute to PH development in patients with LHD 148 .

Unfortunately, treating PH-LHD patients with drugs used to treat PAH patients was not beneficial and even harmful 144,149 .

Group 3 PH

Chronic obstructive lung disease (COPD) associated PH is the best described form of PH in group 3. EC dysfunction is one of the causes for these patients to develop PH $¹⁵⁰$. Cigarette</sup> smoke decreases eNOS and prostacyclin expression in PAECs^{151,152}. COPD patients can show overexpression of VEGF and ET-1 in pulmonary arteries 153,154. These findings have led to the hypothesis that cigarette smoke may be one of the initiating factors for PH in COPD ¹⁵⁰. A role for HIF1 α and EndoMT has also been suggested in COPD ^{155,156}. Although there are similarities in EC dysfunction, drugs used to treat PAH are currently not recommend for group 3 PH, due to lack of evidence how these drugs may influence PH progression in combination with the underlying lung diseases 157.

Group 4 PH

CTEPH develops as a result of a pulmonary embolism (PE) that does not resolve 158 . These organized pulmonary thrombi in the lungs are associated with distal vascular remodelling of non-occluded vessels similar to the remodelling observed in PAH lungs 158. Activated platelets with a hyper-responsiveness to thrombin are likely to contribute to the CTEPH pathogenesis and progression via enhancing inflammatory responses of pulmonary ECs 159. EC dysfunctionassociated vascular remodelling has been suggested as a common mechanism between CTEPH and PAH 158,160. Primary cell cultures isolated from endarterectomized tissue coexpressed both EC and SMC markers, suggesting a role for EndoMT in intimal remodelling/lesion development in CTEPH 161 . The existence of endothelial dysfunction in CTEPH pathogenesis is further supported by the fact that conditioned medium from CTEPH derived PAECs, containing high levels of growth factors and inflammatory cytokines, increased PASMC proliferation and monocyte migration 162 . In addition, PAECs from CTEPH patients show an increased proliferation, altered angiogenic potential and metabolism, and apoptosis resistance ¹⁶³⁻¹⁶⁷. Increased levels of soluble intracellular adhesion molecule-1 (ICAM1) in PAECs from CTEPH patients and in endarterectomy may contribute to EC proliferation and apoptosis resistance through its effect on cell survival pathways ¹⁶⁶. Also FoxO1, in a PI3K/Akt dependent manner, is a possible contributor to the loss of balance between cell survival and death and was downregulated after PE in a rat model of CTEPH 168.

At last, PAECs isolated from CTEPH patients showed a significant rise in basal calcium levels, which is an important regulatory molecule for EC function 169. This imbalance in calcium homeostasis is caused by angiostatic factors such as PF4, IP-10 and collagen type 1, that are formed in the microenvironment created by the unresolved clot and eventually lead to EC dysfunction 169. So far, a soluble guanylate cyclase stimulator (Riociguat) is the only PAH based therapy that has been approved in patients with CTEPH that are not eligible for surgery 170 .

CURRENT AND FUTURE PERSPECTIVES

Although much progress has been made to understand EC dysfunction in PAH, to date there is still no definitive cure and patients only have a median survival rate of 2.8 years 171 . Current therapies for PAH, consisting of calcium channel blockers, ET-1 receptor antagonists, phosphodiesterase type 5 inhibitors, prostacyclin-derivatives and more recently also Riociguat, focus on SMC relaxation with limited or no effect on EC dysfunction and subsequent progressive pulmonary vascular remodelling $172-174$. The effects of EC dysfunction are neglected thus far. Therefore, research on EC dysfunction and its stimuli to target structural changes that narrow lumen size in PAH is vital to find a cure.

A first step towards reversing vascular remodelling in PAH is the use of apoptosis-inducing drugs, such as anthracyclines and proteasome inhibitors. They are already used in combination with cardio-protectants such as p53 inhibitors to reduce pulmonary pressure and restore blood flow in experimental models of PAH $175,176$. The combinatorial use is essential to circumvent the lack of cell-type/organ specificity of cell-killing drugs. Cancer patients but also experimental PAH animals treated with only cell-killing drugs show signs of cardiotoxicity which should be prevented in PAH patients that already suffer from reduced right heart function 175,177-179.

Another way to target progressive pulmonary vascular remodelling focuses on restoring signalling pathways and EC function, e.g. using TGFβ inhibitors, like kallistatin, known to inhibit EndoMT in HUVECs, stimulate eNOS expression and prevent TGFβ induced miRNA-21 synthesis¹⁸⁰. Blocking inflammation to restore normal EC function in PAH, however, was not successful. One explanation might be the complexity of the immune system and by inhibiting the bad side, one also suppresses beneficial inflammatory pathways 181,182.

Modulating BMPR2 has been proposed as therapeutic approach to reverse endothelial dysfunction in PAH too. A recent study comparing human induced pluripotent stem cellderived ECs (iPSC-ECs) from unaffected BMPR2-mutation carriers with iPSC-ECs from BMPR2 mutation carriers that present PAH identified several BMPR2 modifiers and differentially expressed genes in unaffected iPSC-ECs. These BMPR2 modifiers exert a protective response against PAH by improving downstream signalling, which compensates against BMPR2 mutation-induced EC dysfunction and offer insights towards new strategies to rescue BMPR2 signalling ¹⁸³. A potential therapy for stimulating BMPR2 signalling is through pharmaceuticals 184. Direct enhancement of endothelial BMPR2 signalling using recombinant BMP9 protein prevents and reverses established experimental PAH 35. However, in contrast to Long *et al*, Tu *et al* (2019) showed that deletion or inhibition of BMP9, protects against experimental PH via its effect on endothelial production of ET-1, apelin and adrenomedullin 185 . These studies show the BMP receptor family complexity as therapeutics in PAH. More recently, ACTRIIA-Fc, an activin and growth and differentiation factor (GDF) ligand trap, prevented and reversed existing PH in experimental PAH models. ACTRIIA-Fc inhibited SMAD2/3 activation and restored a favourable balance of BMP signalling versus TGFB/activin/GDF signalling. ACTRIIA-Fc is currently tested in a phase-2 clinical trial for efficacy and safety in PAH patients (NCT03496207) 186. Spiekerkoetter *et al*. uncovered a molecular mechanism where FK506 (tacrolimus) restores defective BMPR2 signalling in PAECs from iPAH patients, and reverses severe PAH in several rat models ¹⁸⁴. Based on improvements in clinical parameters and stabilization of cardiac function of end-stage PAH patients in a phase-2a clinical trial, low dose of FK506 was proposed as potential beneficial in the treatment of end-stage PAH 187.These findings open-up an area in which correcting BMPR2 mutations in combination with other therapies might be more successful in curing PAH. A proposed hypothesis to cure PAH describes collecting iPSCs from PAH patients, restoring the BMPR2 mutation with CRISPR/Cas9 and reinjecting those iPSCs in the patient to normalize EC function and signalling along with administration of drugs that could restore the protective gene expression profile of unaffected BMPR2 mutation carriers 188. 6-Mercaptopurine (MP), a well-established immunosuppressive drug, inhibits EC dysfunction and reverses development of PH in the SuHx rat model by restoring BMP signalling through upregulation of nuclear receptor Nur77¹⁸⁹. A recent proof-of-concept study with MP in a small group of PAH patients showed a significant reduction pulmonary vascular resistance, accompanied by increased BMPR2 mRNA expression in the patients' peripheral blood mononuclear cells. However, unexpected severe side-effects require further dose optimisation and/or use of other thiopurine analogues 36 . Chapter 2 ǀ

Transplantation of mesenchymal cells in rats from the SuHx model improved haemodynamic parameters but more interestingly reduced EndoMT (partially) through modulation of HIF2α expression ¹⁹⁰. Furthermore, mesenchymal stem cells are also suggested to reduce inflammation through secretion of paracrine factors and to attenuate vascular remodelling by lowering collagen deposition $190-192$. However the underlying mechanisms for this observation remain unclear 190**.**

At last, epigenetic modulation has received growing interests as potential therapeutic intervention. Especially specific HDAC inhibition shows great promise in reversing pulmonary remodelling and pressure 135. A problem with broad-spectrum HDAC drugs is that they show severe side effects on the right ventricle, which can have fatal consequences in PAH patients with RV failure ^{139,193,194}. Therefore, searches for more selective HDAC inhibitors that do not show cardiotoxicity are still being done. One example is MGCD0103, a HDAC inhibitor that selectively inhibits class-1 HDACs, which has been tested in a chronic hypoxia rat model. This inhibitor showed improved haemodynamics, reduced wall thickening while RV function was maintained 139. Also BET inhibitors such as RVX208 seem promising in the treatment of PAH through its beneficial effect on reducing the apoptosis-resistant and pro-inflammatory phenotype in PASMCs and MVECs isolated from PAH patients but also on vascular remodelling and the RV in several experimental models of PH 38 . Finally, miRNA-21 has been associated with multiple pathogenic features, such as TGFB signalling, EndoMT and apoptosis, central to PAH. Therefore, therapeutic modulation of miRNA-21 may be an important issue for future research to restore pathogenic signalling.

CONCLUSION

To date we still do not fully understand what triggers the onset and progression of PAH. We do know that BMPR2 mutations, epigenetics, physiological conditions, and inflammation are important triggers. EC dysfunction plays a central role in all of this, through EC proliferation, EndoMT and a misbalanced production of vasoactive factors resulting in the disorganized growth of PASMCs. However, the question still remains whether EC dysfunction is a cause or consequence of PAH. Despite advancements made in treating this disease, no focus on targeting PAH at its core. A better understanding of the molecular mechanisms involved in EC dysfunction in PAH is of utmost importance for developing successful therapies to save the lung as well as the heart, and maybe cure PAH in the future.

REFERENCES

- 1 Dumitrescu, D. *et al.* Definition, clinical classification and initial diagnosis of pulmonary hypertension: Updated recommendations from the Cologne Consensus Conference 2018. *International Journal of Cardiology* **272**, 11-19, doi:10.1016/j.ijcard.2018.08.083 (2018).
- 2 Simonneau, G. & Hoeper, M. M. The revised definition of pulmonary hypertension: exploring the impact on patient management. *European heart journal supplements : journal of the European Society of Cardiology* **21**, K4-k8, doi:10.1093/eurheartj/suz211 (2019).
- 3 Simonneau, G. *et al.* Haemodynamic definitions and updated clinical classification of pulmonary hypertension. *Eur Respir J* **53**, doi:10.1183/13993003.01913-2018 (2019).
- 4 Vonk Noordegraaf, A., Groeneveldt, J. A. & Bogaard, H. J. Pulmonary hypertension. *European Respiratory Review* **25**, 4, doi:10.1183/16000617.0096-2015 (2016).
- 5 Hoeper, M. M. *et al.* A global view of pulmonary hypertension. *Lancet Respir Med* **4**, 306-322, doi:10.1016/S2213-2600(15)00543-3 (2016).
- 6 Humbert, M. *et al.* Cellular and molecular pathobiology of pulmonary arterial hypertension. *J Am Coll Cardiol* **43**, 13S-24S, doi:10.1016/j.jacc.2004.02.029 (2004).
- 7 Budhiraja, R., Tuder, R. M. & Hassoun, P. M. Endothelial Dysfunction in Pulmonary Hypertension. *Circulation*, doi:10.1161/01.CIR.0000102381.57477.50 (2004).
- 8 Hadi, H. A., Carr, C. S. & Al Suwaidi, J. Endothelial dysfunction: cardiovascular risk factors, therapy, and outcome. *Vascular health and risk management* **1**, 183-198 (2005).
- 9 Humbert, M. *et al.* Endothelial cell dysfunction and cross talk between endothelium and smooth muscle cells in pulmonary arterial hypertension. *Vascular Pharmacology* **49**, 113-118, doi:10.1016/j.vph.2008.06.003 (2008).
- 10 Nicod, L. P. The endothelium and genetics in pulmonary arterial hypertension. *Swiss Medical Weekly* **137**, 437-442, doi:2007/31/smw-11668 (2007).
- 11 Dummer, A. *et al.* Endothelial dysfunction in pulmonary arterial hypertension: loss of cilia length regulation upon cytokine stimulation. *Pulm Circ* **8**, 2045894018764629, doi:10.1177/2045894018764629 (2018).
- 12 Stewart, D. J., Levy, R. D., Cernacek, P. & Langleben, D. Increased plasma endothelin-1 in pulmonary hypertension: Marker or mediator of disease? *Annals of Internal Medicine*, doi:10.7326/0003-4819-114-6-464 (1991).
- 13 Christman, B. W. *et al.* An Imbalance between the Excretion of Thromboxane and Prostacyclin Metabolites in Pulmonary Hypertension. *New England Journal of Medicine*, doi:10.1056/NEJM199207093270202 (1992).
- 14 Ranchoux, B. *et al.* Endothelial dysfunction in pulmonary arterial hypertension: An evolving landscape (2017 Grover Conference Series). *Pulmonary Circulation* **8**, doi:10.1177/2045893217752912 (2018).
- 15 Orriols, M., Gomez-Puerto, M. C. & Ten Dijke, P. BMP type II receptor as a therapeutic target in pulmonary arterial hypertension. *Cellular and Molecular Life Sciences* **74**, 2979-2995, doi:10.1007/s00018-017-2510-4 (2017).
- 16 Newman, J. H. *et al.* Mutation in the gene for bone morphogenetic protein receptor II as a cause of primary pulmonary hypertension in a large kindred. *The New England journal of medicine* **345**, 319-324, doi:10.1056/nejm200108023450502 (2001).
- 17 Larkin, E. K. *et al.* Longitudinal analysis casts doubt on the presence of genetic anticipation in heritable pulmonary arterial hypertension. *Am J Respir Crit Care Med* **186**, 892-896, doi:10.1164/rccm.201205-0886OC (2012).
- 18 Liu, F., Ventura, F., Doody, J. & Massagué, J. Human type II receptor for bone morphogenic proteins (BMPs): extension of the two-kinase receptor model to the BMPs. *Molecular and cellular biology* **15**, 3479-3486, doi:10.1128/mcb.15.7.3479 (1995).
- 19 Goumans, M. J., Zwijsen, A., Ten Dijke, P. & Bailly, S. Bone Morphogenetic Proteins in Vascular Homeostasis and Disease. *Cold Spring Harbor perspectives in biology* **10**, doi:10.1101/cshperspect.a031989 (2018).
- 20 Yang, X. *et al.* Dysfunctional Smad signaling contributes to abnormal smooth muscle cell proliferation in familial pulmonary arterial hypertension. *Circ Res* **96**, 1053-1063, doi:10.1161/01.Res.0000166926.54293.68 (2005).
- 21 Teichert-Kuliszewska, K. *et al.* Bone morphogenetic protein receptor-2 signaling promotes pulmonary arterial endothelial cell survival: Implications for loss-of-function mutations in the pathogenesis of pulmonary hypertension. *Circulation Research*, doi:10.1161/01.RES.0000200180.01710.e6 (2006).
- 22 Zhang, S. *et al.* Bone morphogenetic proteins induce apoptosis in human pulmonary vascular smooth muscle cells. *Am J Physiol Lung Cell Mol Physiol*, doi:10.1152/ajplung.00284.2002\r00284.2002 [pii] (2003).
- 23 Gräf, S. *et al.* Identification of rare sequence variation underlying heritable pulmonary arterial hypertension. *Nature communications* **9**, 1416, doi:10.1038/s41467-018-03672-4 (2018).
- 24 Frump, A., Prewitt, A. & de Caestecker, M. P. BMPR2 mutations and endothelial dysfunction in pulmonary arterial hypertension (2017 Grover Conference Series). *Pulmonary Circulation* **8**, doi:10.1177/2045894018765840 (2018).
- 25 Soon, E. *et al.* Bone morphogenetic protein receptor type II deficiency and increased inflammatory cytokine production: A gateway to pulmonary arterial hypertension. *American Journal of Respiratory and Critical Care Medicine*, doi:10.1164/rccm.201408-1509OC (2015).
- 26 Liu, D. *et al.* Dosage-dependent requirement of BMP type II receptor for maintenance of vascular integrity. *Blood*, doi:10.1182/blood-2006-11-058594 (2007).
- 27 Long, L. *et al.* Serotonin increases susceptibility to pulmonary hypertension in BMPR2 deficient mice. *Circ Res* **98**, 818-827, doi:10.1161/01.RES.0000215809.47923.fd (2006).
- 28 Atkinson, C. *et al.* Primary pulmonary hypertension is associated with reduced pulmonary vascular expression of type II bone morphogenetic protein receptor. *Circulation* **105**, 1672- 1678, doi:10.1161/01.cir.0000012754.72951.3d (2002).
- 29 Brock, M. *et al.* Interleukin-6 modulates the expression of the bone morphogenic protein receptor type II through a novel STAT3-microRNA cluster 17/92 pathway. *Circ Res* **104**, 1184- 1191, doi:10.1161/circresaha.109.197491 (2009).
- 30 Andruska, A. & Spiekerkoetter, E. Consequences of BMPR2 Deficiency in the Pulmonary Vasculature and Beyond: Contributions to Pulmonary Arterial Hypertension. *International journal of molecular sciences* **19**, doi:10.3390/ijms19092499 (2018).
- 31 Happé, C. *et al.* The BMP Receptor 2 in Pulmonary Arterial Hypertension: When and Where the Animal Model Matches the Patient. *Cells* **9**, doi:10.3390/cells9061422 (2020).
- 32 Hong, K. H. *et al.* Genetic ablation of the BMPR2 gene in pulmonary endothelium is sufficient to predispose to pulmonary arterial hypertension. *Circulation* **118**, 722-730, doi:10.1161/circulationaha.107.736801 (2008).
- 33 Majka, S. *et al.* Physiologic and molecular consequences of endothelial Bmpr2 mutation. *Respir Res* **12**, 84, doi:10.1186/1465-9921-12-84 (2011).
- 34 Yang, X., Long, L., Reynolds, P. N. & Morrell, N. W. Expression of Mutant BMPR-II in Pulmonary Endothelial Cells Promotes Apoptosis and a Release of Factors that Stimulate Proliferation of Pulmonary Arterial Smooth Muscle Cells. *Pulmonary Circulation*, doi:10.4103/2045- 8932.78100 (2011).
- 35 Long, L. *et al.* Selective enhancement of endothelial BMPR-II with BMP9 reverses pulmonary arterial hypertension. *Nat Med* **21**, 777-785, doi:10.1038/nm.3877 (2015).
- 36 Botros, L. *et al.* The Effects of Mercaptopurine on Pulmonary Vascular Resistance and BMPR2 Expression in Pulmonary Arterial Hypertension. *Am J Respir Crit Care Med*, doi:10.1164/rccm.202003-0473LE (2020).
- 37 Kurakula, K. *et al.* Prevention of progression of pulmonary hypertension by the Nur77 agonist 6-mercaptopurine: role of BMP signalling. *Eur Respir J* **54**, doi:10.1183/13993003.02400-2018 (2019).
- 38 Feen, D. E. V. D., Kurakula, K., Tremblay, E., Boucherat, O. & Bossers, G. P. L. Multicenter preclinical validation of BET inhibition for the treatment of pulmonary arterial hypertension. *Am J Respir Crit Care Med* **200**, 910-920 (2019).
- 39 Hiepen, C. *et al.* BMPR2 acts as a gatekeeper to protect endothelial cells from increased TGFβ responses and altered cell mechanics. *PLoS biology* **17**, e3000557, doi:10.1371/journal.pbio.3000557 (2019).
- 40 Prewitt, A. R. *et al.* Heterozygous null bone morphogenetic protein receptor type 2 mutations promote SRC kinase-dependent caveolar trafficking defects and endothelial dysfunction in pulmonary arterial hypertension. *The Journal of biological chemistry* **290**, 960-971, doi:10.1074/jbc.M114.591057 (2015).
- 41 Burton, V. J. *et al.* Bone morphogenetic protein receptor II regulates pulmonary artery endothelial cell barrier function. *Blood* **117**, 333-341, doi:10.1182/blood-2010-05-285973 (2011).
- 42 Diebold, I. *et al.* BMPR2 preserves mitochondrial function and DNA during reoxygenation to promote endothelial cell survival and reverse pulmonary hypertension. *Cell Metabolism*, doi:10.1016/j.cmet.2015.03.010 (2015).
- 43 Li, M. *et al.* Loss of bone morphogenetic protein receptor 2 is associated with abnormal DNA Repair in pulmonary arterial hypertension. *American Journal of Respiratory Cell and Molecular Biology*, doi:10.1165/rcmb.2013-0349OC (2014).
- 44 Rhodes, C. J. *et al.* RNA Sequencing Analysis Detection of a Novel Pathway of Endothelial Dysfunction in Pulmonary Arterial Hypertension. *Am.J.Respir.Crit Care Med.*, doi:10.1164/rccm.201408-1528OC (2015).
- 45 Rol, N. *et al.* Nintedanib improves cardiac fibrosis but leaves pulmonary vascular remodelling unaltered in experimental pulmonary hypertension. *Cardiovascular research* **115**, 432-439, doi:10.1093/cvr/cvy186 (2019).
- 46 Awad, K. S. *et al.* Raf/ERK drives the proliferative and invasive phenotype of BMPR2-silenced pulmonary artery endothelial cells. *American Journal of Physiology - Lung Cellular and Molecular Physiology* **310**, L187-L201, doi:10.1152/ajplung.00303.2015 (2016).
- 47 Orcholski, M. *et al.* Loss Of Carboxylesterase 1 Activity Is Associated With Reduced Bone Morphogenetic Protein Receptor 2 Activity And Membrane Localization In Pulmonary Endothelial Cells. *Am J Respir Crit Care Med* (2014).
- 48 Hurst, L. A. *et al.* TNFα drives pulmonary arterial hypertension by suppressing the BMP type-II receptor and altering NOTCH signalling. *Nature communications* **8**, 14079, doi:10.1038/ncomms14079 (2017).
- 49 Parikh, V. N. *et al.* MicroRNA-21 integrates pathogenic signaling to control pulmonary hypertension: Results of a network bioinformatics approach. *Circulation*, doi:10.1161/CIRCULATIONAHA.111.060269 (2012).
- 50 Sánchez-Duffhues, G., García de Vinuesa, A. & Ten Dijke, P. Endothelial-to-mesenchymal transition in cardiovascular diseases: Developmental signaling pathways gone awry. *Developmental dynamics : an official publication of the American Association of Anatomists* **247**, 492-508, doi:10.1002/dvdy.24589 (2018).
- 51 Medici, D. & Kalluri, R. Endothelial-mesenchymal transition and its contribution to the emergence of stem cell phenotype. *Seminars in Cancer Biology* **22**, 379-384, doi:10.1016/j.semcancer.2012.04.004 (2012).
- 52 Ranchoux, B. *et al.* Endothelial-to-mesenchymal transition in pulmonary hypertension. *Circulation* **131**, 1006-1018, doi:10.1161/CIRCULATIONAHA.114.008750 (2015).
- 53 Good, R. B. *et al.* Endothelial to Mesenchymal Transition Contributes to Endothelial Dysfunction in Pulmonary Arterial Hypertension. *American Journal of Pathology* **185**, 1850- 1858, doi:10.1016/j.ajpath.2015.03.019 (2015).
- 54 Tang, H. *et al.* Endothelial HIF-2α Contributes to Severe Pulmonary Hypertension by Inducing Endothelial-to-Mesenchymal Transition. *American Journal of Physiology-Lung Cellular and Molecular Physiology*, doi:10.1152/ajplung.00096.2017 (2017).
- 55 Goumans, M. J., van Zonneveld, A. J. & ten Dijke, P. Transforming growth factor beta-induced endothelial-to-mesenchymal transition: a switch to cardiac fibrosis? *Trends in cardiovascular medicine* **18**, 293-298, doi:10.1016/j.tcm.2009.01.001 (2008).
- 56 Ursoli Ferreira, F. *et al.* Endothelial Cells Tissue-Specific Origins Affects Their Responsiveness to TGF-β2 during Endothelial-to-Mesenchymal Transition. *International journal of molecular sciences* **20**, doi:10.3390/ijms20030458 (2019).
- 57 Mammoto, T., Muyleart, M., Konduri, G. G. & Mammoto, A. Twist1 in Hypoxia-induced Pulmonary Hypertension through Transforming Growth Factor-β–Smad Signaling. *American Journal of Respiratory Cell and Molecular Biology* **58**, 194-207, doi:10.1165/rcmb.2016- 0323OC (2018).
- 58 Hopper, R. K. *et al.* In pulmonary arterial hypertension, reduced bmpr2 promotes endothelialto-Mesenchymal transition via hmga1 and its target slug. *Circulation*, doi:10.1161/CIRCULATIONAHA.115.020617 (2016).
- 59 Zhang, H. *et al.* Bone morphogenetic protein-7 inhibits endothelial-mesenchymal transition in pulmonary artery endothelial cell under hypoxia. *Journal of Cellular Physiology*, doi:10.1002/jcp.26195 (2018).
- 60 Rol, N., Kurakula, K. B., Happé, C., Bogaard, H. J. & Goumans, M. J. TGF-β and BMPR2 Signaling in PAH: Two Black Sheep in One Family. *International journal of molecular sciences* **19**, doi:10.3390/ijms19092585 (2018).
- 61 Lei, W. *et al.* Expression and analyses of the HIF-1 pathway in the lungs of humans with pulmonary arterial hypertension. *Molecular medicine reports* **14**, 4383-4390, doi:10.3892/mmr.2016.5752 (2016).
- 62 Dai, Z. *et al.* Therapeutic Targeting of Vascular Remodeling and Right Heart Failure in Pulmonary Arterial Hypertension with a HIF-2α Inhibitor. *Am J Respir Crit Care Med* **198**, 1423- 1434, doi:10.1164/rccm.201710-2079OC (2018).
- 63 Zhang, B. *et al.* Hypoxia induces endothelial-mesenchymal transition in pulmonary vascular remodeling. *International Journal of Molecular Medicine*, doi:10.3892/ijmm.2018.3584 (2018).
- 64 Dorfmüller, P., Perros, F., Balabanian, K. & Humbert, M. Inflammation in pulmonary arterial hypertension. *Eur Respir J* **22**, 358-363, doi:10.1183/09031936.03.00038903 (2003).
- 65 Jasiewicz, M. *et al.* Enhanced IL-6 trans-signaling in pulmonary arterial hypertension and its potential role in disease-related systemic damage. *Cytokine* **76**, 187-192, doi:10.1016/J.CYTO.2015.06.018 (2015).
- 66 Groth, A. *et al.* Inflammatory cytokines in pulmonary hypertension. *Respir Res* **15**, 47, doi:10.1186/1465-9921-15-47 (2014).
- 67 Veyssier-Belot, C. & Cacoub, P. Role of endothelial and smooth muscle cells in the physiopathology and treatment management of pulmonary hypertension. *Cardiovascular research* **44**, 274-282, doi:10.1016/S0008-6363(99)00230-8 (1999).
- 68 Miyata, M. *et al.* Pulmonary hypertension in rats. 2. Role of interleukin-6. *International archives of allergy and immunology* **108**, 287-291, doi:10.1159/000237166 (1995).
- 69 Steiner, M. K. *et al.* Interleukin-6 overexpression induces pulmonary hypertension. *Circulation Research*, doi:10.1161/CIRCRESAHA.108.182014 (2009).
- 70 Shao, D. *et al.* Nuclear IL-33 regulates soluble ST2 receptor and IL-6 expression in primary human arterial endothelial cells and is decreased in idiopathic pulmonary arterial

hypertension. *Biochemical and Biophysical Research Communications* **451**, 8-14, doi:10.1016/j.bbrc.2014.06.111 (2014).

- 71 Montanari, E. *et al.* Interleukin-33 stimulates GM-CSF and M-CSF production by human endothelial cells. *Thrombosis and haemostasis* **116**, 317-327, doi:10.1160/th15-12-0917 (2016).
- 72 Liu, J. *et al.* IL-33 Initiates Vascular Remodelling in Hypoxic Pulmonary Hypertension by up-Regulating HIF-1α and VEGF Expression in Vascular Endothelial Cells. *EBioMedicine*, doi:10.1016/j.ebiom.2018.06.003 (2018).
- 73 Berger, G., Azzam, Z. S., Hoffman, R. & Yigla, M. Coagulation and anticoagulation in pulmonary arterial hypertension. *Isr Med Assoc J* **11**, 376-379 (2009).
- 74 Eisenberg, P. R. *et al.* Fibrinopeptide A levels indicative of pulmonary vascular thrombosis in patients with primary pulmonary hypertension. *Circulation*, doi:10.1161/01.CIR.82.3.841 (1990).
- 75 Geggel, R. L., Carvalho, A. C., Hoyer, L. W. & Reid, L. M. von Willebrand factor abnormalities in primary pulmonary hypertension. *The American review of respiratory disease*, doi:10.1164/arrd.1987.135.2.294 (1987).
- 76 Wilcox, J. N., Smith, K. M., Schwartz, S. M. & Gordon, D. Localization of tissue factor in the normal vessel wall and in the atherosclerotic plaque. *Proceedings of the National Academy of Sciences of the United States of America*, doi:10.1073/pnas.86.8.2839 (1989).
- 77 Mackman, N. Regulation of the Tissue Factor gene. *FASEB Journal* **9**, 883-889, doi:10.1096/fasebj.9.10.7615158.
- 78 Riewald, M. & Ruf, W. Orchestration of coagulation protease signaling by tissue factor. *Trends in cardiovascular medicine* **12**, 149-154, doi:10.1016/S1050-1738(02)00153-6 (2002).
- 79 White, R. J. *et al.* Plexiform-like lesions and increased tissue factor expression in a rat model of severe pulmonary arterial hypertension. *Am J Physiol Lung Cell Mol Physiol*, doi:10.1152/ajplung.00321.2006 (2007).
- 80 Lannan, K. L., Phipps, R. P. & White, R. J. Thrombosis, platelets, microparticles and PAH: More than a clot. *Drug Discov Today* **19**, 1230-1235, doi:10.1016/j.drudis.2014.04.001 (2014).
- 81 Sakamaki, F. *et al.* Increased plasma P-selectin and decreased thrombomodulin in pulmonary arterial hypertension were improved by continuous prostacyclin therapy. *Circulation* **102**, 2720-2725, doi:10.1161/01.cir.102.22.2720 (2000).
- 82 Tuder, R. M. *et al.* Relevant issues in the pathology and pathobiology of pulmonary hypertension. *J Am Coll Cardiol* **62**, D4-12, doi:10.1016/j.jacc.2013.10.025 (2013).
- 83 Taraseviciene-Stewart, L. *et al.* Inhibition of the VEGF receptor 2 combined with chronic hypoxia causes cell death-dependent pulmonary endothelial cell proliferation and severe pulmonary hypertension. *The FASEB Journal*, doi:10.1096/fj.00-0343com (2001).
- 84 Sakao, S. *et al.* Initial apoptosis is followed by increased proliferation of apoptosis-resistant endothelial cells. *The FASEB journal : official publication of the Federation of American Societies for Experimental Biology*, doi:10.1096/fj.04-3261fje (2005).
- 85 Masri, F. A. *et al.* Hyperproliferative apoptosis-resistant endothelial cells in idiopathic pulmonary arterial hypertension. *Am J Physiol Lung Cell Mol Physiol* **293**, 548-554, doi:10.1152/ajplung.00428.2006 (2007).
- 86 White, K. *et al.* Endothelial apoptosis in pulmonary hypertension is controlled by a microRNA/programmed cell death 4/caspase-3 axis. *Hypertension*, doi:10.1161/HYPERTENSIONAHA.113.03037 (2014).
- 87 Dabral, S. *et al.* Notch1 signalling regulates endothelial proliferation and apoptosis in pulmonary arterial hypertension. *European Respiratory Journal*, doi:10.1183/13993003.00773-2015 (2016).
- 88 Förstermann, U. & Sessa, W. C. Nitric oxide synthases: regulation and function. *European heart journal* **33**, 829-837, 837a-837d, doi:10.1093/eurheartj/ehr304 (2012).

- 89 Ziche, M. *et al.* Nitric oxide mediates angiogenesis in vivo and endothelial cell growth and migration in vitro promoted by substance P. *Journal of Clinical Investigation*, doi:10.1172/JCI117557 (1994).
- 90 Babaei, S. *et al.* Role of nitric oxide in the angiogenic response in vitro to basic fibroblast growth factor. *Circulation Research*, doi:10.1161/01.RES.82.9.1007 (1998).
- 91 Giaid, A. & Saleh, D. Reduced Expression of Endothelial Nitric Oxide Synthase in the Lungs of Patients with Pulmonary Hypertension. *New England Journal of Medicine*, doi:10.1056/NEJM199507273330403 (1995).
- 92 Xue C & A, J. R. Endothelial Nitric Oxide Synthase in the Lungs of Patients with Pulmonary Hypertension. *New England Journal of Medicine* **333**, 1642-1644, doi:10.1056/NEJM199512143332416 (1995).
- 93 Quinlan, T. R. *et al.* eNOS-deficient mice show reduced pulmonary vascular proliferation and remodeling to chronic hypoxia. *American journal of physiology. Lung cellular and molecular physiology*, doi:10.1023/A:1007183915921 (2000).
- 94 Xu, X. F. *et al.* Epigenetic regulation of the endothelial nitric oxide synthase gene in persistent pulmonary hypertension of the newborn rat. *Journal of Hypertension*, doi:10.1097/HJH.0b013e32833e08f1 (2010).
- 95 Ke, X. *et al.* Persistent pulmonary hypertension alters the epigenetic characteristics of endothelial nitric oxide synthase gene in pulmonary artery endothelial cells in a fetal lamb model. *Physiological genomics* **50**, 828-836, doi:10.1152/physiolgenomics.00047.2018 (2018).
- 96 Chan, Y. *et al.* The cell-specific expression of endothelial nitric-oxide synthase: A role for DNA methylation. *Journal of Biological Chemistry*, doi:10.1074/jbc.M405063200 (2004).
- 97 Fish, J. E. *et al.* The expression of endothelial nitric-oxide synthase is controlled by a cellspecific histone code. *Journal of Biological Chemistry*, doi:10.1074/jbc.M502115200 (2005).
- 98 Zhang, M.-x. *et al.* Effect of 27nt Small RNA on Endothelial Nitric-Oxide Synthase Expression. *Molecular biology of the cell*, doi:10.1091/mbc.E07-11-1186 (2007).
- 99 Gan, Y. *et al.* Role of histone deacetylation in cell-specific expression of endothelial nitric-oxide synthase. *Journal of Biological Chemistry*, doi:10.1074/jbc.M412960200 (2005).
- 100 Fish, J. E. *et al.* Hypoxic repression of endothelial nitric-oxide synthase transcription is coupled with eviction of promoter histones. *Journal of Biological Chemistry*, doi:10.1074/jbc.M109.067868 (2010).
- 101 Chester, A. H., Yacoub, M. H. & Moncada, S. Nitric oxide and pulmonary arterial hypertension. *Global Cardiology Science and Practice* **2017**, doi:10.21542/gcsp.2017.14 (2017).
- 102 Leone, A., Moncada, S., Vallance, P., Calver, A. & Collier, J. Accumulation of an endogenous inhibitor of nitric oxide synthesis in chronic renal failure. *The Lancet*, doi:10.1016/0140- 6736(92)90865-Z (1992).
- 103 Valkonen, V. P., Tuomainen, T. P. & Laaksonen, R. DDAH gene and cardiovascular risk. *Vascular Medicine*, doi:10.1191/1358863x05vm600oa (2005).
- 104 Iannone, L. *et al.* miR-21/DDAH1 pathway regulates pulmonary vascular responses to hypoxia. *Biochemical Journal* **462**, 103-112, doi:10.1042/BJ20140486 (2014).
- 105 Epstein, F. H., Vane, J. R., Änggård, E. E. & Botting, R. M. Regulatory Functions of the Vascular Endothelium. *New England Journal of Medicine*, doi:10.1056/nejm199007053230106 (2010).
- 106 Chen, Y.-F. & Oparil, S. Endothelial Dysfunction in the Pulmonary Vascular Bed. *The American Journal of the Medical Sciences* **320**, 223-232, doi:10.1016/S0002-9629(15)40831-6 (2000).
- 107 Humbert, M. & Sitbon, O. Treatment of Pulmonary Arterial Hypertension NEJM. *New England Journal of …*, 1425-1436, doi:10.1056/NEJMra040291 (2004).
- 108 Mitchell, J. A. *et al.* Role of prostacyclin in pulmonary hypertension. *Global cardiology science & practice* **2014**, 382-393, doi:10.5339/gcsp.2014.53 (2014).
- 109 Tuder, R. M. *et al.* Prostacyclin synthase expression is decreased in lungs from patients with severe pulmonary hypertension. *American Journal of Respiratory and Critical Care Medicine*, doi:10.1164/ajrccm.159.6.9804054 (1999).
- 110 Geraci, M. W. *et al.* Pulmonary prostacyclin synthase overexpression in transgenic mice protects against development of hypoxic pulmonary hypertension. *Journal of Clinical Investigation*, doi:10.1172/JCI5911 (1999).
- 111 Shao, D., Park, J. E. S. & Wort, S. J. The role of endothelin-1 in the pathogenesis of pulmonary arterial hypertension. *Pharmacol Res* **63**, 504-511, doi:10.1016/j.phrs.2011.03.003 (2011).
- 112 Howard, P. G., Plumpton, C. & Davenport, A. P. Anatomical localization and pharmacological activity of mature endothelins and their precursors in human vascular tissue. *Journal of Hypertension*, doi:10.1097/00004872-199211000-00010 (1992).
- 113 Chester, A. H. & Yacoub, M. H. The role of endothelin-1 in pulmonary arterial hypertension. *Global cardiology science & practice* **2014**, 62-78, doi:10.5339/gcsp.2014.29 (2014).
- 114 Shichiri, M., Kato, H., Marumo, F. & Hirata, Y. Endothelin-1 as an autocrine/paracrine apoptosis survival factor for endothelial cells. *Hypertension (Dallas, Tex. : 1979)* **30**, 1198-1203 (1997).
- 115 Giaid, A. *et al.* Expression of Endothelin-1 in the Lungs of Patients with Pulmonary Hypertension. *New England Journal of Medicine*, doi:10.1056/NEJM199306173282402 (1993).
- 116 Li, H. B. *et al.* Enhanced Endothelin-1 and Endothelin Receptor Gene-Expression in Chronic Hypoxia. *Journal of Applied Physiology* (1994).
- 117 Frasch, H. F., Marshall, C. & Marshall, B. E. Endothelin-1 is elevated in monocrotaline pulmonary hypertension. *American Journal of Physiology-Lung Cellular and Molecular Physiology*, doi:10.1152/ajplung.1999.276.2.l304 (1999).
- 118 Davie, N. *et al.* ET(A) and ET(B) receptors modulate the proliferation of human pulmonary artery smooth muscle cells. *Am J Respir Crit Care Med* **165**, 398-405, doi:10.1164/ajrccm.165.3.2104059 (2002).
- 119 Yamashita, K., Discher, D. J., Hu, J., Bishopric, N. H. & Webster, K. A. Molecular regulation of the endothelin-1 gene by hypoxia. Contributions of hypoxia-inducible factor-1, activator protein-1, GATA-2, AND p300/CBP. *The Journal of biological chemistry* **276**, 12645-12653, doi:10.1074/jbc.M011344200 (2001).
- 120 Rodríguez-Pascual, F., Redondo-Horcajo, M. & Lamas, S. Functional Cooperation Between Smad Proteins and Activator Protein-1 Regulates Transforming Growth Factor-β–Mediated Induction of Endothelin-1 Expression. *Circulation Research* **92**, 1288-1295, doi:10.1161/01.RES.0000078491.79697.7F (2003).
- 121 Kanse, S. M. *et al.* Cytokine stimulated endothelin release from endothelial cells. *Life sciences* **48**, 1379-1384, doi:10.1016/0024-3205(91)90434-d (1991).
- 122 Malek, A. M., Greene, A. L. & Izumo, S. Regulation of endothelin 1 gene by fluid shear stress is transcriptionally mediated and independent of protein kinase C and cAMP. *Proc Natl Acad Sci U S A* **90**, 5999-6003, doi:10.1073/pnas.90.13.5999 (1993).
- 123 Park, J. E. *et al.* BMP-9 induced endothelial cell tubule formation and inhibition of migration involves Smad1 driven endothelin-1 production. *PLoS One* **7**, e30075, doi:10.1371/journal.pone.0030075 (2012).
- 124 Galié, N., Manes, A. & Branzi, A. The endothelin system in pulmonary arterial hypertension. *Cardiovascular research* **61**, 227-237, doi:10.1016/j.cardiores.2003.11.026 (2004).
- 125 Tate, R. M., Morris, H. G., Schroeder, W. R. & Repine, J. E. Oxygen metabolites stimulate thromboxane production and vasoconstriction in isolated saline-perfused rabbit lungs. *The Journal of clinical investigation* **74**, 608-613, doi:10.1172/jci111458 (1984).
- 126 Fike, C. D., Zhang, Y. & Kaplowitz, M. R. Thromboxane inhibition reduces an early stage of chronic hypoxia-induced pulmonary hypertension in piglets. *Journal of applied physiology (Bethesda, Md. : 1985)* **99**, 670-676, doi:10.1152/japplphysiol.01337.2004 (2005).
- 127 Lee, S. H., Jeong, D., Han, Y. S. & Baek, M. J. Pivotal role of vascular endothelial growth factor pathway in tumor angiogenesis. *Annals of surgical treatment and research* **89**, 1-8, doi:10.4174/astr.2015.89.1.1 (2015).

- 128 Karaman, S., Leppänen, V. M. & Alitalo, K. Vascular endothelial growth factor signaling in development and disease. *Development (Cambridge, England)* **145**, doi:10.1242/dev.151019 (2018).
- 129 Tuder, R. M. *et al.* Expression of angiogenesis-related molecules in plexiform lesions in severe pulmonary hypertension: evidence for a process of disordered angiogenesis. *The Journal of pathology* **195**, 367-374, doi:10.1002/path.953 (2001).
- 130 Säleby, J., Bouzina, H., Ahmed, S., Lundgren, J. & Rådegran, G. Plasma receptor tyrosine kinase RET in pulmonary arterial hypertension diagnosis and differentiation. *ERJ Open Research* **5**, 00037-02019, doi:10.1183/23120541.00037-2019 (2019).
- 131 Voelkel, N. F. & Gomez-Arroyo, J. The role of vascular endothelial growth factor in pulmonary arterial hypertension: The angiogenesis paradox. *American Journal of Respiratory Cell and Molecular Biology* **51**, 474-484, doi:10.1165/rcmb.2014-0045TR (2014).
- 132 Partovian, C. *et al.* Adenovirus-mediated lung vascular endothelial growth factor overexpression protects against hypoxic pulmonary hypertension in rats. *American Journal of Respiratory Cell and Molecular Biology*, doi:10.1165/ajrcmb.23.6.4106 (2000).
- 133 Hautefort, A. *et al.* Pulmonary endothelial cell DNA methylation signature in pulmonary arterial hypertension. *Oncotarget* **8**, 52995-53016, doi:10.18632/oncotarget.18031 (2017).
- 134 Cavasin, M. A., Stenmark, K. R. & McKinsey, T. A. Emerging Roles for Histone Deacetylases in Pulmonary Hypertension and Right Ventricular Remodeling (2013 Grover Conference series). *Pulmonary Circulation*, doi:10.1086/679700 (2015).
- 135 Zhao, L. *et al.* Histone Deacetylation Inhibition in Pulmonary Hypertension: Therapeutic of Valproic Acid and SuPotentialberoylanilide Hydroxamic Acid. *Circulation* **126**, 455-467, doi:10.1161/CIRCULATIONAHA.112.103176 (2012).
- 136 Seto, E. & Yoshida, M. Erasers of histone acetylation: the histone deacetylase enzymes. *Cold Spring Harbor perspectives in biology* **6**, a018713, doi:10.1101/cshperspect.a018713 (2014).
- 137 Chabot, S. *et al.* HDAC6-HSP90 interplay in pulmonary arterial hypertension. *FASEB Journal* (2016).
- 138 Boucherat, O. *et al.* HDAC6: A Novel Histone Deacetylase Implicated in Pulmonary Arterial Hypertension. *Scientific Reports*, doi:10.1038/s41598-017-04874-4 (2017).
- 139 Cavasin, M. A. *et al.* Selective class i histone deacetylase inhibition suppresses hypoxiainduced cardiopulmonary remodeling through an antiproliferative mechanism. *Circulation Research*, doi:10.1161/CIRCRESAHA.111.258426 (2012).
- 140 Li, M. *et al.* Emergence of Fibroblasts with a Proinflammatory Epigenetically Altered Phenotype in Severe Hypoxic Pulmonary Hypertension. *The Journal of Immunology*, doi:10.4049/jimmunol.1100479 (2011).
- 141 Kim, J. *et al.* Restoration of Impaired Endothelial MEF2 Function Rescues Pulmonary Arterial Hypertension. *Circulation* **131**, 190-199, doi:10.1161/CIRCULATIONAHA.114.013339 (2015).
- 142 Devaiah, B. N., Gegonne, A. & Singer, D. S. Bromodomain 4: a cellular Swiss army knife. *Journal of leukocyte biology* **100**, 679-686, doi:10.1189/jlb.2RI0616-250R (2016).
- 143 Meloche, J. *et al.* Bromodomain-Containing Protein 4. *Circulation Research*, doi:10.1161/CIRCRESAHA.115.307004 (2015).
- 144 Fernández, A. I. *et al.* The Biological Bases of Group 2 Pulmonary Hypertension. *International journal of molecular sciences* **20**, doi:10.3390/ijms20235884 (2019).
- 145 Ontkean, M., Gay, R. & Greenberg, B. Diminished endothelium-derived relaxing factor activity in an experimental model of chronic heart failure. *Circ Res* **69**, 1088-1096, doi:10.1161/01.res.69.4.1088 (1991).
- 146 Givertz, M. M. *et al.* Acute endothelin A receptor blockade causes selective pulmonary vasodilation in patients with chronic heart failure. *Circulation*, doi:10.1161/01.CIR.101.25.2922 (2000).
- 147 Meoli, D. F. *et al.* The transpulmonary ratio of endothelin 1 is elevated in patients with preserved left ventricular ejection fraction and combined pre- and post-capillary pulmonary hypertension. *Pulm Circ* **8**, 2045893217745019, doi:10.1177/2045893217745019 (2018).
- 148 Duarte, J. D. *et al.* Endothelial nitric oxide synthase genotype is associated with pulmonary hypertension severity in left heart failure patients. *Pulmonary Circulation* **8**, 204589401877304-204589401877304, doi:10.1177/2045894018773049 (2018).
- 149 Vachiéry, J. L. *et al.* Pulmonary hypertension due to left heart disease. *Eur Respir J* **53**, doi:10.1183/13993003.01897-2018 (2019).
- 150 Barberà, J. A. Mechanisms of development of chronic obstructive pulmonary diseaseassociated pulmonary hypertension. *Pulmonary circulation* **3**, 160-164, doi:10.4103/2045- 8932.109949 (2013).
- 151 BarberÀ, J. A. *et al.* Reduced Expression of Endothelial Nitric Oxide Synthase in Pulmonary Arteries of Smokers. *American Journal of Respiratory and Critical Care Medicine* **164**, 709-713, doi:10.1164/ajrccm.164.4.2101023 (2001).
- 152 Nana-Sinkam, S. P. *et al.* Prostacyclin prevents pulmonary endothelial cell apoptosis induced by cigarette smoke. *American Journal of Respiratory and Critical Care Medicine*, doi:10.1164/rccm.200605-724OC (2007).
- 153 Santos, S. *et al.* Enhanced expression of vascular endothelial growth factor in pulmonary arteries of smokers and patients with moderate chronic obstructive pulmonary disease. *American Journal of Respiratory and Critical Care Medicine*, doi:10.1164/rccm.200210- 1233OC (2003).
- 154 Carratu, P. *et al.* Exhaled and arterial levels of endothelin-1 are increased and correlate with pulmonary systolic pressure in COPD with pulmonary hypertension. *BMC Pulmonary Medicine*, doi:10.1186/1471-2466-8-20 (2008).
- 155 Xiong, P. Y., Potus, F., Chan, W. & Archer, S. L. Models and Molecular Mechanisms of World Health Organization Group 2 to 4 Pulmonary Hypertension. *Hypertension* **71**, 34-55, doi:10.1161/hypertensionaha.117.08824 (2018).
- 156 Reimann, S. *et al.* Increased S100A4 expression in the vasculature of human COPD lungs and murine model of smoke-induced emphysema. *Respir Res* **16**, 127, doi:10.1186/s12931-015- 0284-5 (2015).
- 157 Olschewski, H. *et al.* in *International Journal of Cardiology* (2018).
- 158 Simonneau, G., Torbicki, A., Dorfmüller, P. & Kim, N. The pathophysiology of chronic thromboembolic pulmonary hypertension. *European respiratory review : an official journal of the European Respiratory Society* **26**, doi:10.1183/16000617.0112-2016 (2017).
- 159 Yaoita, N. *et al.* Platelets are highly activated in patients of chronic thromboembolic pulmonary hypertension. *Arteriosclerosis, Thrombosis, and Vascular Biology*, doi:10.1161/ATVBAHA.114.304404 (2014).
- 160 Humbert, M. Pulmonary arterial hypertension and chronic thromboembolic pulmonary hypertension: Pathophysiology. *European respiratory review : an official journal of the European Respiratory Society* **19**, 59-63, doi:10.1183/09059180.00007309 (2010).
- 161 Sakao, S. *et al.* Endothelial-like cells in chronic thromboembolic pulmonary hypertension: Crosstalk with myofibroblast-like cells. *Respiratory Research*, doi:10.1186/1465-9921-12-109 (2011).
- 162 Mercier, O. *et al.* Abnormal pulmonary endothelial cells may underlie the enigmatic pathogenesis of chronic thromboembolic pulmonary hypertension. *The Journal of heart and lung transplantation : the official publication of the International Society for Heart Transplantation* **36**, 305-314, doi:10.1016/j.healun.2016.08.012 (2017).
- 163 Tura-Ceide, O. *et al.* suppl 60 edn PA3606-PA3606 (European Respiratory Society).
- 164 Naito, A. *et al.* Endothelial cells from pulmonary endarterectomy specimens possess a high angiogenic potential and express high levels of hepatocyte growth factor. *BMC pulmonary medicine* **18**, 197, doi:10.1186/s12890-018-0769-3 (2018).

- 165 Quarck, R., Wynants, M., Verbeken, E., Meyns, B. & Delcroix, M. Contribution of inflammation and impaired angiogenesis to the pathobiology of chronic thromboembolic pulmonary hypertension. *Eur Respir J* **46**, 431-443, doi:10.1183/09031936.00009914 (2015).
- 166 Arthur Ataam, J. *et al.* ICAM-1 PROMOTES THE ABNORMAL ENDOTHELIAL CELL PHENOTYPE IN CHRONIC THROMBOEMBOLIC PULMONARY HYPERTENSION. *The Journal of Heart and Lung Transplantation* **0**, doi:10.1016/j.healun.2019.06.010 (2019).
- 167 Smolders, V. F. *et al.* Decreased glycolysis as metabolic footprint of endothelial cells in chronic thromboembolic pulmonary hypertension. *European Respiratory Journal* **54**, OA5167, doi:10.1183/13993003.congress-2019.OA5167 (2019).
- 168 Deng, C. *et al.* Role of FoxO1 and apoptosis in pulmonary vascular remolding in a rat model of chronic thromboembolic pulmonary hypertension. *Scientific Reports* **7**, 1-10, doi:10.1038/s41598-017-02007-5 (2017).
- 169 Zabini, D. *et al.* Angiostatic factors in the pulmonary endarterectomy material from chronic thromboembolic pulmonary hypertension patients cause endothelial dysfunction. *PLoS ONE* **7**, doi:10.1371/journal.pone.0043793 (2012).
- 170 Conole, D. & Scott, L. J. Riociguat: first global approval. *Drugs* **73**, 1967-1975, doi:10.1007/s40265-013-0149-5 (2013).
- 171 Prins, K. W. & Thenappan, T. WHO Group I Pulmonary Hypertension: Epidemiology and Pathophysiology. *Cardiol Clin*, doi:10.1016/j.ccl.2016.04.001 (2016).
- 172 Hoeper, M. M. *et al.* in *International Journal of Cardiology* (2018).
- 173 Lan, N. S. H., Massam, B. D., Kulkarni, S. S. & Lang, C. C. Pulmonary Arterial Hypertension: Pathophysiology and Treatment. *Diseases (Basel, Switzerland)* **6**, doi:10.3390/diseases6020038 (2018).
- 174 Humbert, M. & Ghofrani, H. A. The molecular targets of approved treatments for pulmonary arterial hypertension. *Thorax* **71**, 73-83, doi:10.1136/thoraxjnl-2015-207170 (2016).
- 175 Suzuki, Y. J., Ibrahim, Y. F. & Shults, N. V. Apoptosis-based therapy to treat pulmonary arterial hypertension. *Journal of Rare Diseases Research & Treatment* **1**, 17-24 (2016).
- 176 Ibrahim, Y. F. *et al.* Mechanism of the susceptibility of remodeled pulmonary vessels to druginduced cell killing. *Journal of the American Heart Association* **3**, e000520, doi:10.1161/jaha.113.000520 (2014).
- 177 Kim, S. Y. *et al.* Bortezomib alleviates experimental pulmonary arterial hypertension. *Am J Respir Cell Mol Biol* **47**, 698-708, doi:10.1165/rcmb.2011-0331OC (2012).
- 178 Jain, D., Russell, R. R., Schwartz, R. G., Panjrath, G. S. & Aronow, W. Cardiac Complications of Cancer Therapy: Pathophysiology, Identification, Prevention, Treatment, and Future Directions. *Current cardiology reports* **19**, 36, doi:10.1007/s11886-017-0846-x (2017).
- 179 Voelkel, N. F. *et al.* Right ventricular function and failure: report of a National Heart, Lung, and Blood Institute working group on cellular and molecular mechanisms of right heart failure. *Circulation* **114**, 1883-1891, doi:10.1161/circulationaha.106.632208 (2006).
- 180 Guo, Y. *et al.* Kallistatin inhibits TGF-β-induced endothelial-mesenchymal transition by differential regulation of microRNA-21 and eNOS expression. *Experimental Cell Research* **337**, 103-110, doi:10.1016/j.yexcr.2015.06.021 (2015).
- 181 Marsh, L. M. *et al.* The inflammatory cell landscape in the lungs of patients with idiopathic pulmonary arterial hypertension. *Eur Respir J* **51**, doi:10.1183/13993003.01214-2017 (2018).
- 182 Kumar, R. & Graham, B. How does inflammation contribute to pulmonary hypertension? *Eur Respir J* **51**, doi:10.1183/13993003.02403-2017 (2018).
- 183 Gu, M. *et al.* Patient-Specific iPSC-Derived Endothelial Cells Uncover Pathways that Protect against Pulmonary Hypertension in BMPR2 Mutation Carriers. *Cell stem cell* **20**, 490-504.e495, doi:10.1016/j.stem.2016.08.019 (2017).
- 184 Spiekerkoetter, E. *et al.* FK506 activates BMPR2, rescues endothelial dysfunction, and reverses pulmonary hypertension. *Journal of Clinical Investigation*, doi:10.1172/JCI65592 (2013).
- 185 Tu, L. *et al.* Selective BMP-9 Inhibition Partially Protects Against Experimental Pulmonary Hypertension. *Circ Res* **124**, 846-855, doi:10.1161/circresaha.118.313356 (2019).
- 186 Yung, L. M. *et al.* ACTRIIA-Fc rebalances activin/GDF versus BMP signaling in pulmonary hypertension. *Sci Transl Med* **12**, doi:10.1126/scitranslmed.aaz5660 (2020).
- 187 Spiekerkoetter, E. *et al.* Low-Dose FK506 (Tacrolimus) in End-Stage Pulmonary Arterial Hypertension. *Am J Respir Crit Care Med* **192**, 254-257, doi:10.1164/rccm.201411-2061LE (2015).
- 188 Quarck, R. & Perros, F. Rescuing BMPR2-driven endothelial dysfunction in PAH: a novel treatment strategy for the future? *Stem Cell Investigation* **4**, 56-56, doi:10.21037/sci.2017.05.11 (2017).
- 189 Kurakula, K. *et al.* 6-mercaptopurine, an agonist of Nur77, reduces progression of pulmonary hypertension by enhancing BMP signalling. *The European respiratory journal*, 1802400- 1802400, doi:10.1183/13993003.02400-2018 (2019).
- 190 Huang, J. *et al.* Transplantation of Mesenchymal Stem Cells Attenuates Pulmonary Hypertension by Normalizing the EndMT. *American Journal of Respiratory Cell and Molecular Biology*, doi:10.1165/rcmb.2018-0165oc (2019).
- 191 de Mendonça, L. *et al.* Mesenchymal stromal cell therapy reduces lung inflammation and vascular remodeling and improves hemodynamics in experimental pulmonary arterial hypertension. *Stem cell research & therapy* **8**, 220, doi:10.1186/s13287-017-0669-0 (2017).
- 192 Martire, A. *et al.* Mesenchymal stem cells attenuate inflammatory processes in the heart and lung via inhibition of TNF signaling. *Basic research in cardiology* **111**, 54, doi:10.1007/s00395- 016-0573-2 (2016).
- 193 Bogaard, H. J. *et al.* Suppression of histone deacetylases worsensright ventricular dysfunction after pulmonary artery banding in rats. *American Journal of Respiratory and Critical Care Medicine*, doi:10.1164/rccm.201007-1106OC (2011).
- 194 Wang, Y. *et al.* Epigenetic Regulation and Its Therapeutic Potential in Pulmonary Hypertension. *Frontiers in pharmacology* **9**, 241, doi:10.3389/fphar.2018.00241 (2018).