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## Reducing uncertainties in image-guided radiotherapy of rectal cancer

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# Chapter 4

## Applicator visualization using ultrashort echo time MRI for high-dose-rate endorectal brachytherapy

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## ABSTRACT

### Purpose

The individual channels in an endorectal applicator for high-dose-rate endorectal brachytherapy are not visible on standard MRI sequences. The aim of this study was to test whether an ultrashort echo time (UTE) MRI sequence could be used to visualize the individual channels to enable MR-only treatment planning for rectal cancer.

### Methods and materials

We used a radial 3D UTE pulse sequence and acquired images of phantoms and two rectal cancer patients. We rigidly registered a UTE image and CT scan of an applicator phantom, based on the outline of the applicator. One observer compared channel positions on the UTE image and CT scan in five slices spaced 25 mm apart. To quantify geometric distortions, we scanned a commercial 3D geometric QA phantom and calculated the difference between detected marker positions on the UTE image and corresponding marker positions on two 3D T<sub>1</sub>-weighted images with opposing readout directions.

### Results

On the UTE images, there is sufficient contrast to discern the individual channels. The difference in channel positions on the UTE image compared to the CT was on average  $-0.1 \pm 0.1$  mm (LR) and  $0.1 \pm 0.3$  mm (AP). After rigid registration to the 3D T<sub>1</sub>-weighted sequences, the residual 95<sup>th</sup> percentile of the geometric distortion inside a 550 mm diameter sphere was 1.0 mm (LR), 0.9 mm (AP) and 0.9 mm (CC).

### Conclusions

With a UTE sequence, the endorectal applicator and individual channels can be adequately visualized in both phantom and patients. The geometrical fidelity is within acceptable range.

## INTRODUCTION

For rectal cancer patients, high-dose rate endorectal brachytherapy (HDREBT) can be used to deliver high doses to the tumor while sparing surrounding organs at risk due to a steep dose gradient [1]. HDREBT may be delivered using an intracavitary mold applicator set, such as the flexible eight-channel applicator (Elekta, Veenendaal, the Netherlands). Applicator reconstruction for the flexible eight-channel applicator is currently performed on CT [2,3], because the individual channels of the applicator are not visible on conventional MR images due to the short T<sub>2</sub>-relaxation time of the applicator. However, CT suffers from limited soft-tissue contrast, which makes it challenging to delineate the target volume accurately [4,5]. MRI is the primary imaging modality for tumor visualization because of its superior soft-tissue contrast and it is therefore currently registered to CT imaging to aid in the tumor delineation on CT. However, acquiring both CT and MRI on the same day can be time consuming and changes in

applicator positioning between the scans may occur because of patient movement and/or differences in organ filling. An MRI-only approach in which applicator reconstruction, delineation of target volume and organs at risk, and treatment planning is performed on MRI is therefore preferred.

MRI-only brachytherapy treatment planning is already the standard for cervical cancer in Europe [6]. To perform applicator reconstruction on MRI, models of rigid applicators have been made available in commercial treatment planning systems and these can be rigidly registered to the applicator on MR images. However, for non-rigid applicators such as the flexible endorectal applicator, such models are not available. In addition, since the individual channels of the flexible endorectal applicator are not visible on MRI, the rotation of the applicator cannot be determined because of its cylindrical shape. In brachytherapy for cervical cancer, dummy catheters can be used to aid in applicator reconstruction. The dummy catheter is filled with a fluid that produces high signal intensity on MRI. However, such dummy catheters are not available for the flexible endorectal applicator. As an alternative, an ultrashort echo time (UTE) sequence [7,8] may be used to visualize the applicator and the individual channels. UTE uses very short echo times on the order of <0.5 ms and allows visualization of materials with very short  $T_2$ -relaxation times.

The aim of this study was to test if a UTE sequence can be used to visualize the individual channels within the flexible endorectal applicator for HDREBT treatment planning. To this end, we first evaluated the visibility of the individual channels in a phantom and determined the geometric fidelity of the UTE sequence. Finally, we acquired UTE images from two rectal cancer patients with applicator *in situ* to evaluate the visibility of the individual channels in an applicator *in situ*.

## METHODS AND MATERIALS

### Applicator

We used a commercial intracavitary mold applicator (OncoSmart, Elekta, Veenendaal, The Netherlands). This is a flexible cylindrical applicator made of silicon with a diameter of 20 mm and a length of 280 mm. The applicator has eight channels radially spaced along its circumference, which allows for an asymmetric dose distribution [9].

### UTE sequence

All MRI scans in this study were performed on a 3T scanner (Ingenia, Philips Healthcare, Best, The Netherlands). We used a 3D stack of radials UTE pulse sequence for radial sampling of free induction decays, enabled by clinical science functionality under a research agreement. A cylindrical encoding scheme was performed with radial sampling in-plane and cartesian sampling through-plane. Radial sampling was used as it allows for very short echo times in the order of <0.5 ms. The sequence is similar to the stack of spirals UTE sequence described in Qian *et al.* [10]. For UTE excitation, a non-selective

hard radiofrequency pulse was applied for a very short duration (0.05 ms) and was followed by a phase-encoding gradient in craniocaudal (CC) direction for slice encoding. Immediately after the phase-encoding gradient, a radial read-out was performed to quickly sample the k-space in the two directions that are perpendicular to the slice direction. The radial k-space data acquisition started already during the ramp of the gradients [7]. The scan parameters for the UTE sequences used for imaging the applicator phantom, the two patients, and the geometric QA phantom are shown in Table 1. To ensure clinical feasibility, the scanning times for patients were kept below 6 minutes.

## Applicator phantom

To optimize and evaluate the UTE sequence for applicator visualization, we prepared a phantom. We first filled a box (400 x 300 x 190 mm<sup>3</sup>) halfway with agarose gel. The agarose gel consisted of 0.2 g Dotarem 0.5 mM (Guerbet, Villepinte, France), 10 grams agar (A1296, Sigma Aldrich, Saint Louis, USA), and 3 grams NaCl per liter water, aiming for a T<sub>1</sub> (spin-lattice relaxation time) of 1-2 seconds and a T<sub>2</sub> (spin-spin relaxation time) of >30 ms [11], which is on the same order of magnitude as the relaxation properties of human soft-tissue around the rectum. A balloon was placed around the applicator and subsequently filled with 20cc of water with 5% Telebrix Gastro (Guerbet, Villepinte, France), as in our clinical procedure. The applicator was then placed on top of the first layer of agarose gel and a second layer of agarose gel was applied to fill the box.

## Individual channel visualization

To evaluate the visibility of the individual channels within the applicator, we acquired an axial UTE image of the phantom. To test whether the visualized channels on the UTE image actually represent the individual channels within the applicator, we acquired an axial CT scan (Brilliance Big Bore, Philips Healthcare, Best, The Netherlands) of the phantom (voxel size 0.63 x 0.63 x 1 mm<sup>3</sup>, 120 kVp, tube current 346 mA, exposure time 887 ms). We performed a rigid registration of the CT scan to the UTE

**Table 1.** Scan parameters for the UTE sequences used for imaging the applicator phantom, the patients, and the geometric fidelity phantom.

Parameter	Applicator phantom	Patient 1	Patient 2	Geometric QA phantom
Voxel size (mm <sup>3</sup> )	1.0x1.0x2.5	1.0x1.0x3.5	0.98x0.98x2.5	1.94x1.94x1.94
Echo time (ms)	0.14	0.14	0.14	0.14
Repetition time (ms)	5.26	4.97	5.22	4.22
Acquisition grid	376x376x126	376x376x90	384x384x90	288x288x206
Field of view (mm <sup>3</sup> )	376x376x315	376x376x315	376x376x225	560x560x400
Readout bandwidth (Hz/mm)	886	886	886	895
SENSE factor	1.4	1.4	1.4	1.0
Flip angle (deg)	10	10	10	10
Acquisition duration (s)	499	337	353	643

image based on the outline of the applicator using Elastix, a toolbox for intensity-based medical image registration [12]. To determine channel positions, one observer manually aligned a 2D template of the channel configuration on the UTE image and the registered CT scan in five slices spaced 25 mm apart. We then calculated the in-plane difference in channel positions between the UTE image and the CT scan. To evaluate the visibility of the channels in a clinical setting, we acquired informed consent from two rectal cancer patients undergoing HDREBT within a clinical trial. In addition to the MRI sequences acquired for HDREBT treatment planning, we acquired an additional axial UTE image for these two patients with the applicator *in situ*. As part of sequence optimization, we acquired a UTE image with a slice thickness of 3.5 mm in the first patient and a slice thickness of 2.5 mm in the second patient. For one slice in each UTE image, we determined the contrast-to-noise ratio ( $\text{CNR} = |S_C - S_A|/\sigma$ ) of the channels relative to the applicator. Here,  $S_C$  and  $S_A$  are the average signal intensities of the channels and the applicator, respectively, and  $\sigma$  is the standard deviation of the image noise. Average signal intensities were extracted from regions of interest of  $2 \times 2$  mm placed on and between the channels. In a similar fashion,  $\sigma$  was estimated from signal intensity variations inside a region of interest in the balloon surrounding the applicator.

## Geometric fidelity

To quantify geometric distortions, we acquired MRI images of a commercial 3D geometric QA phantom (Philips Healthcare, Best, The Netherlands) [13,14]. The phantom was placed on the treatment table centered around the isocenter. It consists of seven plastic plates, placed 55 mm apart, each containing 276 spheres (markers) filled with oil with a diameter of 10 mm. Within each plate, the markers are located on a grid with a spacing of 25 mm. In total, the phantom is 330 mm long and has a diameter of 500 mm.

To determine the actual position of the markers within the phantom, we used two axial 3D  $T_1$ -weighted gradient-echo sequences (voxel size: 1.94 mm  $\times$  1.94 mm  $\times$  1.94 mm, field of view: 560  $\times$  560  $\times$  400 mm $^3$ , TE: 3.40 ms, TR: 6.90 ms, readout bandwidth: 828 Hz/mm) with opposing readout directions (anterior and posterior) to be able to correct for magnetic-field inhomogeneity. Subsequently, we acquired a UTE image with the same isotropic voxel size as the two axial 3D  $T_1$ -weighted images (Table 1). The marker detection algorithm described in Keesman *et al.* [13] was used to detect the markers, which produced lists of marker positions, one for each image.

To correct for magnetic-field inhomogeneity, the corresponding detected marker positions in the two 3D  $T_1$ -weighted images were averaged. To assess the distortion of the averaged 3D  $T_1$ -weighted marker positions, a regular reference grid (containing the ideal marker positions, defined according to the known geometry of the phantom) was rigidly registered per plate to the averaged 3D  $T_1$ -weighted marker positions. Residuals between the averaged 3D  $T_1$ -weighted marker positions and the registered reference grid ( $\text{ref\_grid}_{T_1,3D}$ ) were calculated.

To assess the geometric distortion of the UTE image relative to the 3D T<sub>1</sub>-weighted images, we calculated the residuals between detected markers in the UTE image and ref\_grid<sub>T<sub>1</sub>,3D</sub>. The 5<sup>th</sup>, 50<sup>th</sup> and 95<sup>th</sup> percentiles for the residuals are presented for various diameters of spherical volume (DSV) to evaluate the geometric fidelity for various distances from the isocenter.

## RESULTS

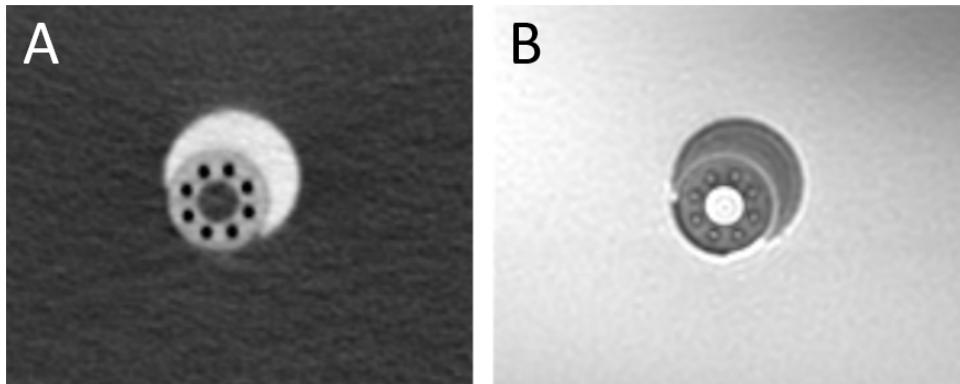
### Individual channel visualization

On the UTE image that was acquired of the applicator phantom, the channels have sufficient contrast relative to the applicator itself to be able to discern the individual channels (Figure 1). The difference in channel positions on the UTE image compared to the CT was on average -0.1 ± 0.1 mm (left-right (LR)) and 0.1 ± 0.3 mm (anteroposterior (AP)). In addition, individual channels are visible on the UTE patient images (Figure 2). CNR was calculated from the slices that contained the balloon (Figure 2, A2 and B2). The CNR was 1.9 for patient 1 and 2.4 for patient 2.

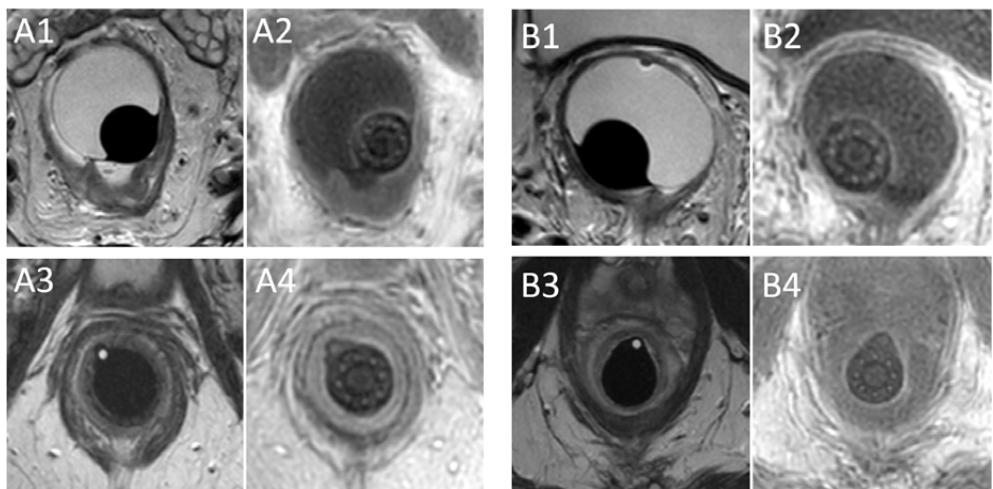
### Geometric distortion

In total, 1428 corresponding markers out of the 1932 markers in the geometric QA phantom were detected by the marker detection algorithm in both the 3D T<sub>1</sub>-weighted images and the UTE image. Marker appearance on the UTE image and the 3D T<sub>1</sub>-weighted images with opposing readout directions is shown in Figure 3. The mean and standard deviation of the residuals between the averaged 3D T<sub>1</sub>-weighted marker positions and ref\_grid<sub>T<sub>1</sub>,3D</sub> was 0.01 ± 0.42 mm (LR), -0.03 ± 0.36 mm (AP), and 0.02 ± 0.33 mm (CC). The 95<sup>th</sup> percentile of the residuals was 0.81 mm (LR), 0.68 mm (AP), and 0.65 mm (CC) within a DSV of 550 mm, which includes all detected markers. In Figure 4, the residuals are plotted as a function of distance to the isocenter in the LR, AP and CC directions, respectively.

The mean and standard deviation of the residuals between detected markers in the UTE image and ref\_grid<sub>T<sub>1</sub>,3D</sub> were 0.46 ± 0.46 mm (LR), -1.69 ± 0.50 mm (AP), and 0.07 ± 0.48 mm (CC). This indicates a systematic shift, mostly in the AP direction. In a clinical scenario, a rigid registration is performed between the UTE image and an anatomical image based on the outline of the applicator to correct for any changes in patient and/or applicator positioning that can occur within the scan session. To investigate effects of higher-order distortion patterns in the UTE image, we corrected this systematic shift by means of a global translation. Subsequently, we calculated the remaining residuals. The 5<sup>th</sup>, 50<sup>th</sup> and 95<sup>th</sup> percentiles for these residuals within various DSVs are presented in Table 2. A 95<sup>th</sup> percentile of 1.0 mm or lower is observed in all directions within a DSV of 550 mm. The magnitude of the distortions increases with increasing distance from the isocenter. This can also be observed in Figure 4, where the residuals are plotted as a function of distance to the isocenter in the LR, AP and CC directions, respectively.



**Figure 1.** A CT scan (A) and UTE MR image (B) of the applicator phantom.



**Figure 2.** Axial slices of T2-TSE images (A1, A3, B1, B3) and UTE images (A2, A4, B2, B4) in two patients (A and B) with the applicator in situ.

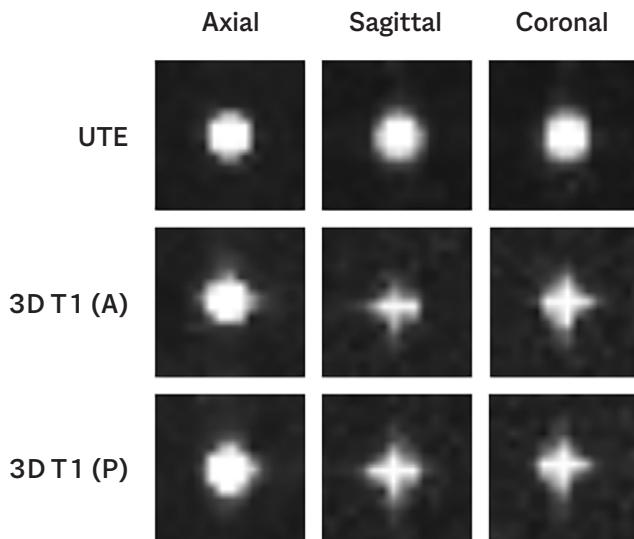
## DISCUSSION

Treatment planning for HDREBT for rectal cancer using the flexible endorectal applicator is currently not performed on MRI alone, as the individual channels within the applicator are not visible on MRI images. The aim of this study was to test if a UTE sequence can be used to visualize the individual channels within the flexible endorectal applicator for HDREBT treatment planning. We have shown that the individual channels are visible on UTE images, both in a phantom and in patients. The CNR of the channels and the

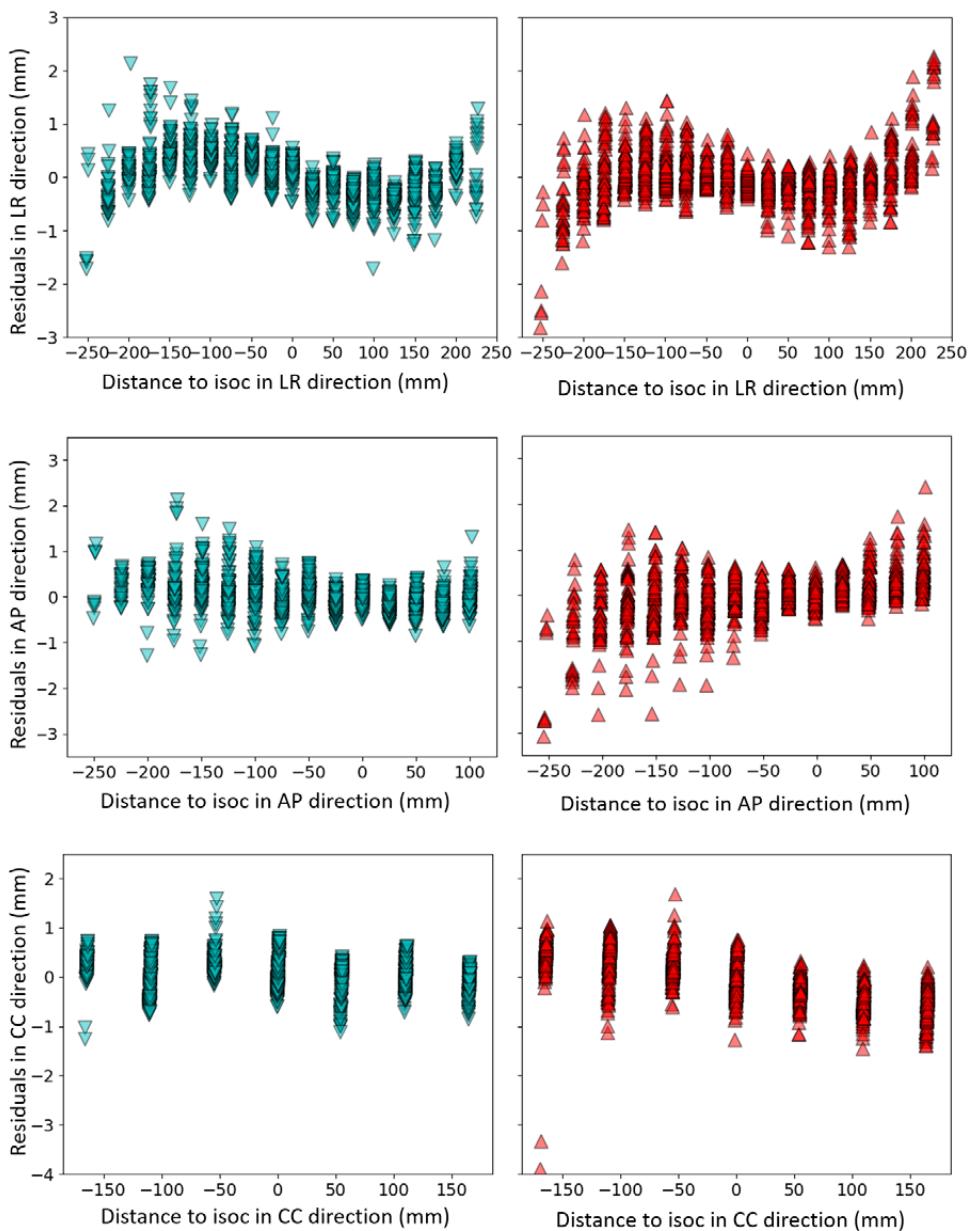
applicator was higher in patient 2. This may be explained by a partial volume effect due to a difference in slice thickness, which was 3.5 mm for patient 1 and 2.5 mm for patient 2.

We observed a systematic shift of the detected marker positions in the UTE image relative to ref\_grid<sub>T<sub>1</sub>,3D</sub>, especially in the AP direction. In a clinical application, an anatomical image is used for delineation, while the UTE image would be used to visualize the individual channels within the applicator. A rigid registration would be performed based on the outline of the applicator and surrounding balloon, negating any systematic offsets. After correcting for the systematic shift, the 95<sup>th</sup> percentile of the residuals is 1.0 mm or lower in all directions within a DSV of 550 mm. In addition, within a volume that is typically of interest for applicator reconstruction (i.e., a DSV of 300 mm), the 95<sup>th</sup> percentile of the residuals was 0.3 mm (LR), 0.4 mm (AP) and 0.7 mm (CC), which is acceptable for HDREBT treatment planning.

In a UTE sequence, k-space data acquisition starts quickly after the radiofrequency excitation and is performed during the ramp up of the gradients [7]. This makes the UTE sequence prone to degraded image quality due to eddy currents and unbalanced hardware time delays, that lead to undesired k-space trajectory deviations [15]. This could have contributed to the systematic shift we have observed. Although for our application it suffices to correct for this shift by using a rigid registration, different techniques can be used to measure the actual k-space trajectory to improve the image reconstruction [15-17].



**Figure 3.** Appearance of a single marker of the 3D geometric QA phantom in the axial, sagittal and coronal plane on a UTE image and two 3D T<sub>1</sub>-weighted images with opposing readout directions (A = anterior readout direction, P = posterior readout direction).



**Figure 4.** Residuals of the average detected marker positions of the 3D  $T_1$ -weighted images (blue) and residuals of detected markers in the UTE image (red) with respect to ref\_grid <sub>$T_1,3D$</sub>  in the LR-, AP- and CC-direction as a function of distance to isocenter (isoc) in the LR-, AP- and CC-direction, respectively.

**Table 2.** Measured residuals of detected marker positions in the UTE image relative to  $\text{ref\_grid}_{\text{T1,3D}}$  in the LR, AP and CC direction after rigid registration of the detected markers in the UTE image to  $\text{ref\_grid}_{\text{T1,3D}}$ .

DSV (mm)	Percentile	LR (mm)	AP (mm)	CC (mm)	No. of markers
100	5 <sup>th</sup>	0.0	0.0	0.0	11
	50 <sup>th</sup>	0.1	0.1	0.1	
	95 <sup>th</sup>	0.2	0.4	0.7	
200	5 <sup>th</sup>	0.0	0.0	0.0	121
	50 <sup>th</sup>	0.1	0.1	0.2	
	95 <sup>th</sup>	0.3	0.4	0.6	
300	5 <sup>th</sup>	0.0	0.0	0.0	387
	50 <sup>th</sup>	0.1	0.2	0.3	
	95 <sup>th</sup>	0.3	0.4	0.7	
400	5 <sup>th</sup>	0.0	0.0	0.0	862
	50 <sup>th</sup>	0.1	0.2	0.3	
	95 <sup>th</sup>	0.5	0.6	0.8	
550	5 <sup>th</sup>	0.0	0.0	0.0	1428
	50 <sup>th</sup>	0.2	0.3	0.3	
	95 <sup>th</sup>	1.0	0.9	0.9	

## CONCLUSIONS

The endorectal applicator and the individual channels can be adequately visualized using a UTE sequence in both a phantom and patients. After a rigid registration to an anatomical image, the geometric fidelity of the UTE sequence is within acceptable range. The UTE sequence is therefore suitable for HDREBT treatment planning.

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