

Diagnostic and prognostic markers of cutaneous lymphomas Schrader, A.M.R.

Citation

Schrader, A. M. R. (2020, August 27). *Diagnostic and prognostic markers of cutaneous lymphomas*. Retrieved from https://hdl.handle.net/1887/136020

Note: To cite this publication please use the final published version (if applicable).

Cover Page

Universiteit Leiden

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Author: Schrader, A.M.R. **Title**: Diagnostic and prognostic markers of cutaneous lymphomas **Issue date**: 2020-08-27

PERSISTENT ONCOGENIC PATHWAYS DRIVE PRIMARY CUTANEOUS DIFFUSE LARGE B-CELL LYMPHOMA, LEG TYPE AND THEIR RELAPSES

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Submitted

ABSTRACT

Primary cutaneous diffuse large B‑cell lymphoma, leg type (PCDLBCL‑LT) is an aggressive cutaneous lymphoma with a 5‑year disease‑specific survival (DSS) of 56%. To improve biological understanding, disease classification, and risk stratification, we aimed to gain more insight into the molecular profiles of primary and relapsed disease in PCDLBCL-LT patients. Fifty-five PCDLBCL-LT patients were analysed with targeted next-generation sequencing for 52 B-cell-lymphoma-related genes and 'triple' fluorescence *in situ* hybridization (*MYC*/*BCL2*/*BCL6*). This included 14 cases with paired analysis of diagnostic and relapse/refractory disease. The most frequent alterations were mutations in *MYD88* (72%), *CD79B* (51%), *PIM1* (36%), *TBL1XR1* (25%), and *KMT2D* (24%), loss of *CDKN2A* (65%), and *MYC* rearrangements (26%). Paired analysis showed largely identical driver alterations in relapse/ refractory disease. Additionally, disease evolution was characterized by subclonal variations due to ongoing somatic hypermutation (SHM), such as in *PIM1*. Employing survival analysis, only *MYC* rearrangements showed an inferior overall survival (p=0.045; HR:2.23; 95%CI:1.00‑4.98), DSS (p<0.001; HR:4.16; 95%CI:1.63‑10.60), and disease-free survival (p=0.003; HR:2.85; 95%CI:1.39-5.86). In conclusion, this study demonstrates that *MYD88*/*CD79B* mutations, loss of *CDKN2A*, and *MYC* rearrangements are early events and persistent oncogenic drivers in primary and relapsed PCDLBCL‑LT. In addition, disease evolution is characterized by ongoing SHM. From the prominent drivers, only *MYC* rearrangements were associated with an inferior survival. These results support the idea that activation of the NF-KB pathway, *MYC* signaling, and CDKN2A loss play a critical role in pathogenesis of PCDLBCL-LT and could provide attractive targets for novel therapeutic approaches.

INTRODUCTION

Primary cutaneous diffuse large B-cell lymphoma, leg type (PCDLBCL-LT) is a rare, extranodal variant of diffuse large B‑cell lymphoma (DLBCL) defined by skin localization and no extracutaneous manifestations at time of diagnosis. It is an aggressive type of primary cutaneous lymphoma with a 5‑year disease-specific survival (DSS) of only 56%.¹ First-line treatment consists of immune‑polychemotherapy (rituximab, cyclophosphamide, doxorubicin, vincristine, and prednisone; R-CHOP). Unfortunately, immune-polychemotherapy has a high toxicity and is often poorly tolerated by this elderly and frail patient population. As such, in case a patient has a single lesion and/or poor clinical condition, local radiotherapy only can be considered.¹ Additionally, despite high complete remission rates after initial therapy, over two-thirds of the patients early relapse within months to years.²

Currently, standardized second‑line and third‑line treatment guidelines are lacking and no routine classifiers are available to predict patients at risk for relapsed/ refractory disease. At time of diagnosis, few factors are associated with an inferior survival: multiple lesions, loss of 9p.21 (*CDKN2A/B*), and mutated *MYD88* L265P.²⁻⁴ In 2018, our group identified an association between *MYC* rearrangements and a higher risk for disease progression and lymphoma-related death.⁵ More recently, Ducharme et al.⁶ reported that mutations in genes affecting the B-cell receptor (BCR) pathway were associated with a reduced disease-free period. However, as PCDLBCL‑LT is a rare disease entity, the mentioned studies were performed in small patient cohorts and observations have not been confirmed by other studies. At present, it remains difficult to predict which patients relapse, and, given the poor outcomes, an urgent need for development of novel therapeutic approaches in patients with PCDLBCL‑LT exists. Therefore, our aim was to perform a comprehensive molecular analysis of patients with PCDLBCL‑LT at time of diagnosis and relapse/refractory disease, to provide insight into the molecular profile and disease evolution, and to correlate this profile with disease outcome.

MATERIALS AND METHODS

All patients diagnosed with PCDLBCL-LT (2000-2019) in the Leiden University Medical Center (LUMC), The Netherlands (n=56), and the University Hospitals, Leuven (UZL), Belgium (n=9), were retrospectively selected. Diagnosis was made in accordance with the current classification system of the World Health Organization (WHO)⁷ and the WHO - European Organisation for Research and Treatment of Cancer (EORTC)¹ and centrally reviewed by A.M.R.S., P.M.J., and R.W.. At time of diagnosis, presence of extracutaneous disease was excluded by standard staging procedures, consisting of a PET‑CT scan or a CT‑scan in combination with a bone marrow biopsy. Formalin-fixed and paraffin-embedded or fresh frozen skin biopsies of pre‑treatment lesions and relapsed/refractory disease were collected from the archives of the Pathology Departments. Patients were excluded in case no tissue samples were available. Clinical characteristics were collected from the Dutch Cutaneous Lymphoma Database and from medical records. The study was performed in accordance with the Code Proper Secondary Use of Human Tissue established by the Dutch Federation of Medical Sciences and the CuraRata Biobank protocol (3.5162), and approved by the medical ethics committees of the LUMC (B19.011) and the UZL (S62445).

All cases were studied for protein expression of CD20 and/or CD79a, CD10, BCL6, MUM1, BCL2, IgM, and MYC, rearrangement status of *MYC*, and only in case of a *MYC* rearrangement for double hit status of *BCL2* and *BCL6*, with break apart rearrangement probes, as described in a previous study.⁵ The rearrangement status of 39 patients was previously reported.⁵

Genomic DNA from FFPE biopsies (n=55) was microdissected from deparaffinised 10µm sections and fully automatically isolated with the Tissue Preparation System (TPS) robot (Siemens Healthcare Diagnostics), as described previously.⁸ Genomic DNA from fresh frozen biopsies (n=7) was isolated from 25µm cryosections with the QIAamp DNA Mini Kit (Qiagen). DNA concentrations were quantified with the Qubit dsDNA HS Fluorometer (Life Technologies).

For targeted next‑generation sequencing (tNGS), libraries were prepared either with the Ion Chef System (ThermoFisher Scientific) or manually for sequencing with the LYMFv1 panel. The LYMFv1 panel is a validated Ion Ampliseq panel that was developed in-house and contains 1362 amplicons, subdivided into 2 primer pools, that cover >95% of 52 B-cell lymphoma-related genes. The panel was composed based on a comprehensive review of >130 original scientific reports for frequency and clinical relevance of genetic mutations in B-cell lymphomas. This panel has a high overlap (73%) with the proposed consensus tNGS panel for mature B-cell malignancies by Sujobert et al.⁹ The generated sequencing data was aligned against the human reference genome (GRCh37/hg19) using the TMAP 5.0.7 software with default parameters and variants were called by the Torrent Variant Caller (ThermoFisher Scientific). All variants with ≥100 reads and a variant allele frequency of ≥10% were functionally annotated in the Geneticist Assistant NGS Interpretive Workbench (SoftGenetics). Variants with a population frequency of >1% in the 1000 Genomes Project¹⁰, variants previously identified in sequencing runs with 'in-house' DNA samples isolated from blood of ~300 healthy controls, and variants in homopolymeric regions or variants with a strand bias of >90% were excluded from further analysis. The remaining variants were categorized for their pathogenicity as class 1 (benign), class 2 (likely benign), class 3 (unknown significance), class 4 (likely pathogenic), and class 5 (pathogenic).¹¹ This classification was done based on information retrieved from dbSNP¹², Clinvar¹³, Cosmic¹⁴, Alamut Focus (Version 1.0; Interactive Biosoftware, Rouen, France), and literature. In addition, a strategy was developed to further categorize all class 3 variants. This strategy is based on the CADD-Phred score¹⁵ of the variants and their notation in SIFT, Polyphen2_HDIV, LRT, and MutationTaster. Finally, all class 4 and class 5 variants, as well as all class 3 variants with a CADD‑Phred score >25, or a CADD‑Phred score between 10 and 25 with ≥2 additional deleterious notations in SIFT, Polyphen2_HDIV, LRT, and MutationTaster were included. These variants are further referred to as 'mutations'. Quality control (QC) consisted of assessment of the coverage with a minimal count of 100 reads and a ratio of transitions versus transversions (Ts:Tv ratio) <5. Patients were excluded in case these quality criteria were not met. The average read count per patient is listed in Supplemental Table 1.

Copy‑number changes were also assessed. In short, the median base coverage per amplicon was calculated and normalized using the median value of all amplicons in that sample. Samples with low read count or with a high coverage variability among amplicons in the same gene were marked so that extra care should be taken while interpreting the final results. Systematic differences between amplicons were normalized using a set of 18 libraries prepared with DNA extracted from 10 non‑neoplastic tonsils. A gene was considered lost or gained if the normalized coverage of more than 2 consecutive amplicons was below or above the estimated 99% confidence intervals (CIs) of these amplicons, respectively. Copy-number analysis and visualization of results, as well as loss of heterozygosity (LOH) was done using the Next‑Generation Sequence Expert (NGSE) Shiny app (https://git. lumc.nl/druano/NGSE).

Statistical analysis was performed using IBM SPSS statistics (version 23) and RStudio (version 1.1442)*.* The median follow‑up time was determined using the reverse Kaplan-Meier method.¹⁶ Survival was calculated from the date of first histological diagnosis to the date of death by any cause for overall survival (OS), the date of death by lymphoma for DSS, and the date of disease relapse or progression of disease or death by any cause (whichever came first) for disease-free survival (DFS). Patients without an event at the end of follow-up were censored. For OS and DFS, survival curves were plotted using the Kaplan‑Meier method and compared with the Log‑rank test. For DSS, cumulative incidences were estimated with the competing risk method and compared using the Gray's test, with non-lymphoma-related death considered as competing risk. In case of a statistically significant p-value (0.05) , the corresponding hazard ratios (HRs) and 95% CIs were calculated with the Cox proportional‑hazards model. Based on literature, the following parameters were included in analysis: age at diagnosis (≤70 years *vs* >70 years)2 , sex (male *vs* female), disease extension (single vs multiple lesions)², MYD88 L265P mutations (mutant vs wild type)⁴, *CDKN2A* status (loss vs no loss)³, mutations in the BCR pathways (mutant CD79A, CD79B or CARD11 vs wild type)⁶, MYC status (rearranged vs wild type)⁵, MCD (mutant *MYD88* and *CD79B* vs wild type)17, and other frequent molecular alterations in our cohort. In case the molecular profile of primary disease was not available, the profile of relapse/refractory disease was included in the survival analysis (n=5).

RESULTS

Patient characteristics

For this study, 65 patients with PCDLBCL-LT were selected. Three patients were excluded because no material was available and an additional 7 patients because the obtained sequencing data did not pass the quality control. In total, 69 skin biopsies were successfully sequenced from a total of 55 patients with PCDLBCL-LT, including paired analysis of diagnostic and relapsed/refractory disease in 14 patients (Supplemental Figure 1). The patient characteristics are presented in Table 1. The cohort consisted of 28 (51%) females and the median age at diagnosis was 78 (range, 47 to 92) years. Patients presented with single lesions in 13 of 55 (24%) cases and with multiple lesions located in 1 body region (regional disease) in 34 of 55 (62%) cases or more body regions (multifocal disease) in 8 of 55 (15%) cases.² The legs were involved in the vast majority (46/55; 84%).

Twenty‑seven (49%) patients were systemically treated with immunepolychemotherapy, consisting of (R-)CHOP or CHOP-like regimens (n=23). In 8 of these patients, systemic treatment was combined with local radiotherapy. Another 26 patients (47%) received local treatment, consisting of radiotherapy (n=25), surgical excision (n=1), or a combination (n=1). No treatment was given in 2 of 55 (4%) patients because of spontaneous remission of a solitary lesion after biopsy (n=1) and sudden cardiac death (n=1). After initial treatment, 51 of 53 (96%) patients showed a complete response, while 2 of 53 (4%) patients had refractory/ progressive disease, both after R‑CHOP treatment. From the complete responders, 33 of 51 (63%) patients developed disease relapses. In second-line and/or third-line treatment, regimens were highly heterogeneous, including local radiotherapy, monotherapy with rituximab or lenalidomide, (immune‑)polychemotherapy with different regimens, or autologous/allogeneic stem-cell transplantation. The median follow‑up duration of all patients was 7.9 (range, 0.08 to 14.3) years. For systemically-treated patients only, the median follow-up duration was 9.1 (range, 0.7 to 14.3) years and in the subgroup of locally‑treated patients this was 7.7 (range, 0.5 to 8.9) years.

Table 1. Clinical characteristics and immunophenotype of 55 patients with primary cutaneous diffuse large B‑cell lymphoma, leg type

Table 1. Clinical characteristics and immunophenotype of 55 patients with primary cutaneous diffuse large B‑cell lymphoma, leg type (continued)

a Local therapy consisted of radiotherapy in 24 patients, surgical excision combined with radiotherapy in 1 patient, and surgical excision in 1 patient.

b (Immune‑)polychemotherapy consisted of a combination of rituximab plus cyclophosphamide, doxorubicin, vincristine, and prednisone (R‑CHOP) in 21 patients, rituximab plus cyclophosphamide, etoposide, vincristine, and prednisone (R‑CEOP) in 2 patients, CHOP in 3 patients, and vincristine and prednisone (OP) in 1 patient. In 7 patients, immune‑polychemotherapy was combined with local radiotherapy.

c No treatment was given in 2 patients due to spontaneous remission of a single lesion (n=1) and sudden cardiac death (n=1).

d Data is missing in 1 case.

e Data is missing in 2 cases.

Protein expression and rearrangement status

Nearly all cases were positive for BCL2 (53/55; 96%) and IgM (50/53; 94%), the majority expressed MUM1 (46/55; 84%), MYC (36/54; 67%), and BCL6 (32/55; 58%), and few cases expressed CD10 (7/55; 13%). In total, 14 of 53 (26%) PCDLBCL‑LT patients showed a *MYC* rearrangement. An additional *BCL6* rearrangement was detected in 1 case at time of diagnosis (as described previously⁵) and in 1 case with an acquired double hit at time of relapsed disease that was initially absent. Unfortunately, the sequencing data of the diagnostic sample from this patient failed the quality control and was excluded from paired analysis. None of the *MYC*‑rearranged patients had a second hit in *BCL2*.

Figure 1. Oncoprint plot presenting the detected molecular alterations with an occurrence in over 10% of the 55 included patients with (PCDLBCL-LT). Our results demonstrate that activation of the NF‑κB pathway, *MYC* signaling, and loss of *CDKN2A* are the main drivers of lymphomagenesis in PCDLBCL-LT patients.

Molecular profile

The detected molecular alterations with an occurrence in over 10% of the 55 patients with PCDLBCL-LT are presented in Figure 1. The involved pathways are visualized in a schematic diagram in Figure 2. The median number of mutations per patient was 5 (range, 0 to 26). The majority of the patients $(40/55; 72%)$ harbored a *MYD88* mutation, of which the L265P hotspot was the most frequently present (35/40; 88%). Non-L265P variants were detected in 5 of 40 (13%) patients, including S243N (n=3), an additional T84I (n=1), M232T (n=1), and Y240S (n=1). *MYD88* mutations result in activation of the NF-KB pathway, thereby promoting cell survival and proliferation.¹⁸ Other gene mutations that result in NF- κ B activation were detected in *CD79B* in 28 of 55 (51%) patients and *CARD11* in 6 of 55 (11%) patients, both components of the B-cell receptor pathway, and mutations in *PIM1* in 20 of 55 (36%) patients, a gene involved in the affiliated JAK/STAT pathway. *CD79B* mutations comprised the hotspot Y196 variant in 23 of 28 (82%) cases. Non-Y196 variants (5/28; 18%) included splice-site variants (n=3), L199R/G40fs (n=1), and

I201fs (n=1). In 6 of 23 (26%) patients with *CD79B* Y196, additional *CD79B* non‑Y196 mutations were detected, including splice‑site variants (n=3) and I109, E229A, and M190fs/E208A (n=1 each). In 21 of 55 (38%) patients concurrent mutations in *MYD88* L265P and *CD79B* were detected, similar to the MCD genotype in DLBCL, as defined by Schmitz et al.¹⁷

Figure 2. Schematic diagram of predominantly involved pathways in lymphomagenesis of primary cutaneous diffuse large B‑cell lymphoma, leg type (PCDLBCL‑LT). The central pathway that shows molecular alterations in these patients is the NF‑κB pathway, with recurrent mutations in *MYD88* (72%), *CD79B* (51%), *PIM1* (36%), and *CARD11* (11%), and the affiliated *PRDM1* gene (11%). Other commonly affected pathways are involved in cell cycle arrest/apoptosis, with *CDKN2A* loss in 65%, *MYC* rearrangements in 26%, and mutations in the transcription factors *BTG1* (20%) and *BTG2* (13%). Less frequent mutations were seen in several epigenetic modifiers, such as *TBL1XR1* (25%), *KMT2D* (24%), *ETV6* (16%), *MEF2B* (16%), and *EP300* (11%). It is likely that patients with PCDLBCL‑LT will be treated with a more targeted approach in the near future, with the NF‑κB pathway, downstream CDKN2A and MYC as central targets for novel therapeutic approaches, instead of the current one-size-fits-all treatment with immune-polychemotherapy.

In addition to activation of the NF-KB pathway, NF-KB itself promotes plasma cell differentiation via *IRF4* and *PRDM119*, with mutations in 4 of 55 (7%) patients and 6 of 55 (11%) patients, respectively. Also frequently affected were the epigenetic modifiers *TBL1XR1* (14/55; 25%), *KMT2D* (13/55; 24%), *MEF2B* (9/55; 16%), *ETV6* (9/55; 16%) and *EP300* (6/55; 11%), as well as the regulators of cell cycle/apoptosis *MYC* (9/55; 16%) and the tumor suppressor *CDKN2A* (6/55; 11%). Finally, the transcription factors *BTG1* (11/55; 20%) and *BTG2* (7/55; 13%) were commonly mutated. *MYC* mutations co‑occurred with *MYC* rearrangements in 8 of 9 (89%) patients. In contrast, patients without a *MYC* mutation only harbored *MYC* rearrangements in 6 of 44 (14%) patients.

More often than *CDKN2A* mutations, copy‑number loss of *CDKN2A* was observed (36/55; 65%). While it is difficult to reliably distinguish a hemizygous from a homozygous loss, 4 patients concomitantly presented with a *CDKN2A* mutation, suggesting hemizygous loss of *CDKN2A*. Other copy-number alterations observed were only present in few patients, including loss of *BCL7A* (6%), *CD70* (6%), and *PCLG2* (4%), and gain of *HIST1H1E* (6%) and *BCL2* (6%) (data not shown).

Figure 3. Side-by-side overview of the results of comprehensive molecular analysis of paired diagnostic and relapsed/refractory disease in 14 patients with primary cutaneous diffuse large B‑cell lymphoma, leg type (PCDLBCL‑LT). Oncoprint plot representing the molecular alterations of PCDLBCL‑LT, demonstrating that the dominant drivers of lymphomagenesis, i.e. *MYD88*/*CD79B*, *CDKN2A*, and *MYC*, are early oncogenic events in disease evolution. In addition, mutations in SHM motifs were present in *TBL1XR1*, *KMT2D*, *BTG2*, *ETV6*, *MYC*, and *MYD88* (S243N variant), with an occurrence ranging from 8% to 100% of the variants per gene. Remark: In all cases with a discrepancy in the mutational profile between diagnostic and relapse/refractory disease, it was ruled out that the variants were already present in low frequency (<10%) in the diagnostic and relapse samples, respectively. Abbreviations: FU, follow-up; D+, death by lymphoma; Do, death by unrelated cause; A+, alive w disease; Ao, alive w/o disease; CR, complete remission; PD, progressive disease.

*mutation in SHM motif (WRCY‑RGYW)

Disease evolution

Paired molecular analysis of diagnostic and relapsed/refractory disease was available in 14 patients (Figure 3). A conceptual representation of disease evolution is presented in Figure 4. Treatment consisted of local radiotherapy in 4 of 14 (29%) patients and immune‑polychemotherapy in 10 of 14 (71%) patients, which was combined with local radiotherapy in 5 patients. Thirteen patients achieved a complete remission after initial treatment and 1 patient had refractory/progressive disease under immune‑polychemotherapy. In case of treatment with local radiotherapy only, disease relapse was present in another body site in 2 patients. The median time between diagnostic and relapse/refractory disease was 14 (range, 4 to 80) months.

Figure 4. Conceptual representation of lymphomagenesis in patients with primary cutaneous diffuse large B-cell lymphoma, leg type (PCDLBCL-LT). Overall, our results suggest that activation of the NF-KB pathway, *MYC* signaling, and loss of *CDKN2A* are early drivers of PCDLBCL‑LT and are persistently present in primary and relapsed/refractory disease. In addition, subclonal variations in disease evolution are largely characterized by passenger mutations and ongoing somatic hypermutation (SHM).

Overall, the molecular profile at time of diagnosis was predominantly similar with the profile in relapse/refractory disease. In 4 of 14 (29%) patients, the molecular profile was identical at both time points. In the other 10 patients, the profile varied slightly, with acquired mutations (n=4), lost mutations (n=3), or both (n=3). Interestingly, from the supposed driver alterations, *MYD88* mutations and *MYC* rearrangements were uniformly present in both diagnostic and relapse/refractory disease. This was also the case for the less frequently mutated genes *TBL1XR1*, *KMT2D*, *SGK1*, and *ATM*.

Acquired mutations were detected in several other genes, of which *PIM1* was most remarkable, with acquired mutations in 3 of 14 (21%) cases, ranging from 1 to 3 mutations per patient. This increase can be explained by the fact that *PIM1* is a known target of ongoing somatic hypermutation $(SHM)^{20}$, and indeed, these variants were all present in SHM motifs (WRCY‑RGYW). Other genes with variants in SMH motifs were *TBL1XR1*, *KMT2D*, *BTG2*, *ETV6*, *MYC*, and *MYD88* (S243N variant), with frequencies ranging from 8%‑100% of the variants per gene. On the other hand, for several genes, such as *BTG1*, *CARD11*, and *ETV6*, some mutations were present at time of diagnosis but were completely absent in the relapsed/refractory samples.

Table 2. Survival analysis of molecular factors in patients with primary cutaneous diffuse large B‑cell lymphoma, leg type

Abbreviations: WT, wild type; BCR, B‑cell receptor; HR, hazard ratio; CI, confidence interval. Bold values indicate a statistically significant result.

**Results are less reliable because of <5 events in 1 of the groups.*

Figure 5. Survival curves for *MYC* status in all patients and the subgroup of systemically-treated patients with primary cutaneous diffuse large B-cell lymphoma, leg type. Patients with a MYC rearrangement demonstrate an inferior overall survival in (A) all patients (p=0.045; HR:2.23; 95%CI:1.00-4.98) and (B) systemically-treated patients (p=0.005; HR:4.47; 95%CI:1.43-13.99), an inferior disease-specific survival in (C) all patients (p<0.001; HR:4.16; 95%CI:1.63-10.60) and (D) systemically-treated patients (p<0.001; HR:7.51; 95%CI:1.95-28.87), and a shorter disease-free survival in (E) all patients (p=0.003; HR:2.85; 95%CI:1.39-5.86) and (F) systemically-treated patients (p<0.001; HR:13.77; 95%CI:3.59-52.85) compared with patients without a MYC rearrangement.

Survival analysis

To identify risk classifiers for relapsed/refractory disease and survival, we correlated the molecular profiles with the patient outcomes. In all 55 patients, survival analysis (Table 2 and Figure 5) demonstrated that only *MYC* rearrangements were associated with an inferior OS (p=0.045; HR:2.23; 95%CI:1.00‑4.98), DSS (p<0.001; HR:4.16; 95%CI:1.63‑10.60), and shorter DFS (p=0.003; HR:2.85; 95%CI:1.39‑5.86). To assess the influence of treatment heterogeneity, the subgroup of systemically‑treated patients only was also evaluated. This analysis revealed similar results, with only *MYC* rearrangements to be associated with an inferior OS (p=0.005; HR:4.47; 95%CI:1.43‑13.99), DSS (p<0.001; HR:7.51; 95%CI:1.95‑28.87), DFS (p<0.001; HR:13.77; 95%CI:3.59‑52.85). Interestingly, none of the adverse molecular factors described in literature (*MYD88* L265P, loss of *CDKN2A*, and mutations in the BCR pathway) were associated with an inferior survival. In addition to the molecular alterations, age at diagnosis >70 years and male gender were associated with a significantly inferior survival (Supplemental Table 2).

DISCUSSION

To provide insight into the genetic background and disease evolution of patients with PCDLBCL-LT, we comprehensively analysed the molecular profile of a relatively large cohort of patients (n=55), performed paired analysis of diagnostic and relapsed/refractory disease in 14 of these patients, and studied the association of recurrent molecular alterations with disease outcome.

As PCDLBCL‑LT is a rare disease, previously, only smaller cohorts of patients were analysed for molecular alterations, including whole‑exome sequencing in 31 patients and targeted sequencing in an additional 20 patients.^{21,22} Consistent with these studies, we found a high frequency of mutations in *MYD88* (72%), *CD79B* (51%), *PIM1* (36%), *TBL1XR1* (25%), and *KMT2D* (24%), and loss of *CDKN2A* (65%). This profile predominantly contributes to constitutive activation of the NF‑κB‑signalling pathway and overlaps with the activated B-cell (ABC) subtype of DLBCL and, in particular, extranodal DLBCL, such as intravascular large B‑cell lymphoma, primary central nervous system lymphoma (PCNSL), and primary testicular lymphoma

(PTL).²³⁻²⁵ This profile was identified as the C5 signature by Chapuy et al.²⁶, with the major alterations being mutations in *MYD88* and *CD79B*, additional mutations in *ETV6*, *PIM1*, *TBL1XR1*, and *BTG1*, and ongoing SHM. At the same time, Schmitz et al.17 described the MCD group in DLBCL, with co‑occurrence of mutations in *MYD88* and *CD79B*, further characterized by inactivating mutations in *PRDM1*, as well as alterations in the tumor suppressor genes *CDKN2A*, *ETV6*, *BTG1*, *BTG2*, *TBL1XR1*, and *KLHL14*. In our cohort of PCDLBCL-LT patients, we detected molecular features of both the C5 signature and the MCD genotype.

Additionally to the molecular similarities with ABC‑DLBCL, we also identified *MYC* rearrangements in 26% of the PCDLBCL‑LT patients, which are more frequently found in GCB‑DLBCL.27‑29 However, the association of *MYC* rearrangements with the GCB subtype accounts especially for double hits with *BCL2* and triple hits whereas single *MYC* rearrangements and double hits with *BCL6* show a rather equal distribution between the GCB and ABC subtypes.³⁰ In previous studies, which included small cohorts up to 25 PCDLBCL‑LT patients, the frequency of *MYC* rearrangements was reported between 0% and 43%.³¹⁻³⁴ In addition to our previous study5 , we currently describe 14 new patients of which 3 (21%) patients had a *MYC* rearrangement. In our cohort, the majority of the MYC-rearranged patients (10/14; 71%) also harbored *MYD88* and/or *CD79B* mutations.

In addition, Mareschal et al.²¹ report MYC mutations in 26% of patients with PCDLBCL-LT compared with 16% in our cohort.²¹ Interestingly, in our cohort, *MYC* mutations often co‑occurred with *MYC* rearrangements (89%) and only 14% of the *MYC* rearrangements were present in patients without a *MYC* mutation. This corresponds to DLBCL, as described by Reddy et al.35, with *MYC* rearrangements in >80% of the cases with a *MYC* mutation, while only 10% of the cases without a *MYC* mutation were *MYC*‑rearranged.

Our results further indicate that PCDLBCL‑LT patients with a *MYC* rearrangement may experience a more aggressive disease course, despite absence of double hits in *BCL2* and *BCL6*. In survival analysis, *MYC* rearrangements were the only molecular feature associated with an inferior OS, DSS, and DFS, in both all patients and the

subgroup of systemically-treated patients. These results support our previously published data on an association of *MYC* rearrangements with an inferior survival in PCDLBCL‑LT.5 Recently, we demonstrated a prognostic importance of *MYD88* mutations in systemic DLBCL³⁶, as was previously also reported in PCDLBCL-LT by Pham-Ledard et al⁴. However, in our current study, MYD88 mutations, including the subgroup of *MYD88* L265P, were not associated with an adverse outcome. Accordingly, other previously described adverse molecular factors, i.e. loss of *CDKN2A* and mutations in the BCR pathway, were not associated with an inferior survival in our cohort.^{3,4,6} Results of survival analysis in relatively small patient cohorts, including the results of the current study, should always be interpreted with caution and should be confirmed by other studies. In PCDLBCL-LT, so far, none of the reported adverse molecular factors have been independently confirmed.

Besides their prognostic effect, the predominant drivers of lymphomagenesis in PCDLBCL‑LT patients, i.e. *MYD88/CD79B* mutations, *MYC* rearrangements, and, to a lesser extent, *CDKN2A* loss were persistently present in diagnostic and relapse/ refractory disease. These findings suggest that these are early events in disease evolution of PCDLBCL-LT, and that, despite large reductions in lymphoma volume by initial therapy in the vast majority of the patients, minimally residual disease persisted that was not detected by our currently standard methods of clinical examination and PET‑CT scanning. Such minimal residual disease may be detected with targeted analysis of liquid biopsies for circulating tumor DNA harbouring hotspot mutations like MYD88 L265P in the future.³⁷ In addition, subclonal variations between diagnosis and relapse were present and commonly caused by ongoing SHM. This corresponds with the recent study by Ducharme et al.⁶, demonstrating mutations in SHM motifs in 33% to 100% of several affected genes, including *PIM1*, *IRF4*, *MYC*, *BCL2*, and *CARD11*, and expression of activation‑induced cytidine‑deaminase, the enzyme that is responsible for SHM, as was reported by Dijkman et al.³⁸

The increasing knowledge of the molecular landscape of PCDLBCL-LT, including 55 patients in the current study, provides a rationale for novel, more targeted treatment strategies in the near future. So far, several case reports and a phase

6

II clinical trial demonstrate promising results of treatment with lenalidomide in relapsed/refractory PCDLBCL-LT patients.³⁹⁻⁴² Additionally, 1 patient in our cohort with progressive disease under immune‑polychemotherapy and radiotherapy achieved an enduring complete remission for over a year on lenalidomide monotherapy. Also, several relapsed/refractory PCDLCBL‑LT cases have been reported with response to ibrutinib⁴³⁻⁴⁵, an inhibitor of the BCR pathway. Given the importance of *MYD88*/*CD79B* as drivers of lymphomagenesis in the majority of PCDLBCL‑LT patients, the NF‑κB pathway and its affiliated signaling pathways might be targeted in several alternative ways by future systemic therapies, as reviewed by De Groen et al.¹⁸ In addition, *MYC* and the cyclin-dependent kinases downstream of CDKN2A could be considered as targets for novel therapeutic approaches, either as monotherapy or in combination with inhibition of the NF-κB pathway.^{46,47} Despite a large spread in reported frequency of *MYC* rearrangements, MYC protein expression is reported in the majority of patients with PCDLBCL-LT, with 67% in our cohort and 83% in the cohort reported by Menguy et al.³² It seems plausible that not only the *MYC*‑rearranged patients, but also the MYC expressors, may benefit from MYC-directed treatment strategies.⁴⁶ Finally, as mutations in epigenetic modifiers were present in a significant subgroup of patients (64%), these genes could also provide attractive targets for novel therapeutic approaches.48 Currently, we stand at the beginning of precision medicine in PCDLBCL‑LT, and it will be highly interesting to see the efficacy of these and other targeted treatment strategies in clinical trials for relapse/refractory disease but also as first-line therapy. In case of novel targeted treatments, ongoing SHM that results in subclonal variation may become clinically relevant, as for example *CARD11* mutations that may cause resistance to ibrutinib.18

In conclusion, our data corroborate the importance of *MYD88*/*CD79B* mutations, *CDKN2A* loss, and *MYC* rearrangements as early events and persistent oncogenic drivers of disease evolution of PCDLBCL‑LT. In addition, disease evolution is characterized by ongoing SHM. From these predominant drivers, only *MYC* rearrangements are associated with an inferior survival. These results support the idea that activation of the NF‑κB pathway, *MYC* signaling, and loss of *CDKN2A* play a critical role in pathogenesis of PCDLBCL‑LT and that these molecular alterations could provide attractive targets for novel therapeutic approaches.

REFERENCES

- 1. Willemze R, Cerroni L, Kempf W, et al. The 2018 update of the WHO‑EORTC classification for primary cutaneous lymphomas. *Blood*. 2019;133(16):1703‑1714.
- 2. Senff NJ, Hoefnagel JJ, Jansen PM, et al. Reclassification of 300 primary cutaneous B‑Cell lymphomas according to the new WHO‑EORTC classification for cutaneous lymphomas: comparison with previous classifications and identification of prognostic markers. *J Clin Oncol*. 2007;25(12):1581‑1587.
- 3. Senff NJ, Zoutman WH, Vermeer MH, et al. Fine‑mapping chromosomal loss at 9p21: correlation with prognosis in primary cutaneous diffuse large B‑cell lymphoma, leg type. *J Invest Dermatol*. 2009;129(5):1149‑1155.
- 4. Pham‑Ledard A, Beylot‑Barry M, Barbe C, et al. High frequency and clinical prognostic value of MYD88 L265P mutation in primary cutaneous diffuse large B‑cell lymphoma, leg‑type. *JAMA Dermatol*. 2014;150(11):1173‑1179.
- 5. Schrader AMR, Jansen PM, Vermeer MH, Kleiverda JK, Vermaat JSP, Willemze R. High Incidence and Clinical Significance of MYC Rearrangements in Primary Cutaneous Diffuse Large B‑Cell Lymphoma, Leg Type. *Am J Surg Pathol*. 2018;42(11):1488‑1494.
- 6. Ducharme O, Beylot‑Barry M, Pham‑Ledard A, et al. Mutations of the B‑Cell Receptor Pathway Confer Chemoresistance in Primary Cutaneous Diffuse Large B-Cell Lymphoma Leg Type. *J Invest Dermatol*. 2019;139(11):2334‑2342 e2338.
- 7. Swerdlow SH ed WHO Classification of Tumours of Haematopoietic and Lymphoid Tissues. In: Campo ELH, N; Jaffe, ES; Pileri, SA; Stein, H; Thiele, J; Vardiman, JW ed (ed Revised 4th). Lyon: IARC; 2017.
- 8. van Eijk R, Stevens L, Morreau H, van Wezel T. Assessment of a fully automated high-throughput DNA extraction method from formalin-fixed, paraffin-embedded tissue for KRAS, and BRAF somatic mutation analysis. *Exp Mol Pathol*. 2013;94(1):121‑125.
- 9. Sujobert P, Le Bris Y, de Leval L, et al. The Need for a Consensus Next-generation Sequencing Panel for Mature Lymphoid Malignancies. *Hemasphere*. 2019;3(1):e169.
- 10. Genomes Project C, Auton A, Brooks LD, et al. A global reference for human genetic variation. *Nature*. 2015;526(7571):68‑74.
- 11. Plon SE, Eccles DM, Easton D, et al. Sequence variant classification and reporting: recommendations for improving the interpretation of cancer susceptibility genetic test results. *Hum Mutat*. 2008;29(11):1282‑1291.
- 12. Sherry ST, Ward M, Sirotkin K. dbSNP‑database for single nucleotide polymorphisms and other classes of minor genetic variation. *Genome Res*. 1999;9(8):677‑679.

- 13. Landrum MJ, Lee JM, Benson M, et al. ClinVar: improving access to variant interpretations and supporting evidence. *Nucleic Acids Res*. 2018;46(D1):D1062‑D1067.
- 14. Tate JG, Bamford S, Jubb HC, et al. COSMIC: the Catalogue Of Somatic Mutations In Cancer. *Nucleic Acids Res*. 2019;47(D1):D941‑D947.
- 15. Kircher M, Witten DM, Jain P, O'Roak BJ, Cooper GM, Shendure J. A general framework for estimating the relative pathogenicity of human genetic variants. *Nat Genet*. 2014;46(3):310‑315.
- 16. Schemper M, Smith TL. A note on quantifying follow-up in studies of failure time. *Control Clin Trials*. 1996;17(4):343‑346.
- 17. Schmitz R, Wright GW, Huang DW, et al. Genetics and Pathogenesis of Diffuse Large B‑Cell Lymphoma. *N Engl J Med*. 2018;378(15):1396‑1407.
- 18. de Groen RAL, Schrader AMR, Kersten MJ, Pals ST, Vermaat JSP. MYD88 in the driver's seat of B-cell lymphomagenesis: from molecular mechanisms to clinical implications. *Haematologica*. 2019;104(12):2337‑2348.
- 19. Basso K, Dalla‑Favera R. Germinal centres and B cell lymphomagenesis. *Nat Rev Immunol*. 2015;15(3):172‑184.
- 20. Khodabakhshi AH, Morin RD, Fejes AP, et al. Recurrent targets of aberrant somatic hypermutation in lymphoma. *Oncotarget*. 2012;3(11):1308‑1319.
- 21. Mareschal S, Pham‑Ledard A, Viailly PJ, et al. Identification of somatic mutations in primary cutaneous diffuse large B-cell lymphoma, leg-type by massive parallel sequencing. *J Invest Dermatol*. 2017;137(9):1984‑1994.
- 22. Zhou XA, Louissaint A, Jr., Wenzel A, et al. Genomic Analyses Identify Recurrent Alterations in Immune Evasion Genes in Diffuse Large B‑Cell Lymphoma, Leg Type. *J Invest Dermatol*. 2018;138(11):2365‑2376.
- 23. Schrader AMR, Jansen PM, Willemze R, et al. High prevalence of MYD88 and CD79B mutations in intravascular large B‑cell lymphoma. *Blood*. 2018;131(18):2086‑2089.
- 24. Fontanilles M, Marguet F, Bohers E, et al. Non‑invasive detection of somatic mutations using next‑generation sequencing in primary central nervous system lymphoma. *Oncotarget*. 2017;8(29):48157‑48168.
- 25. Chapuy B, Roemer MG, Stewart C, et al. Targetable genetic features of primary testicular and primary central nervous system lymphomas. *Blood*. 2016;127(7):869‑881.
- 26. Chapuy B, Stewart C, Dunford AJ, et al. Molecular subtypes of diffuse large B cell lymphoma are associated with distinct pathogenic mechanisms and outcomes. *Nat Med*. 2018;24(5):679‑690.
- 27. Pasqualucci L, Dalla‑Favera R. The genetic landscape of diffuse large B‑cell lymphoma. *Semin Hematol*. 2015;52(2):67‑76.
- 28. Copie‑Bergman C, Cuilliere‑Dartigues P, Baia M, et al. MYC‑IG rearrangements are negative predictors of survival in DLBCL patients treated with immunochemotherapy: a GELA/LYSA study. *Blood*. 2015;126(22):2466‑2474.
- 29. Rosenwald A, Bens S, Advani R, et al. Prognostic Significance of MYC Rearrangement and Translocation Partner in Diffuse Large B‑Cell Lymphoma: A Study by the Lunenburg Lymphoma Biomarker Consortium. *J Clin Oncol*. 2019;37(35):3359‑3368.
- 30. Scott DW, King RL, Staiger AM, et al. High grade B‑cell lymphoma with MYC and BCL2 and/or BCL6 rearrangements with diffuse large B‑cell lymphoma morphology. *Blood*. 2018;131(18):2060‑2064.
- 31. Wiesner T, Streubel B, Huber D, Kerl H, Chott A, Cerroni L. Genetic aberrations in primary cutaneous large B‑cell lymphoma: a fluorescence in situ hybridization study of 25 cases. *Am J Surg Pathol*. 2005;29(5):666‑673.
- 32. Menguy S, Frison E, Prochazkova‑Carlotti M, et al. Double‑hit or dual expression of MYC and BCL2 in primary cutaneous large B‑cell lymphomas. *Mod Pathol*. 2018;31(8):1332‑1342.
- 33. Pham‑Ledard A, Prochazkova‑Carlotti M, Andrique L, et al. Multiple genetic alterations in primary cutaneous large B-cell lymphoma, leg type support a common lymphomagenesis with activated B‑cell‑like diffuse large B‑cell lymphoma. *Mod Pathol*. 2014;27(3):402‑411.
- 34. Hallermann C, Kaune KM, Gesk S, et al. Molecular cytogenetic analysis of chromosomal breakpoints in the IGH, MYC, BCL6, and MALT1 gene loci in primary cutaneous B‑cell lymphomas. *J Invest Dermatol*. 2004;123(1):213‑219.
- 35. Reddy A, Zhang J, Davis NS, et al. Genetic and Functional Drivers of Diffuse Large B Cell Lymphoma. *Cell*. 2017;171(2):481‑494 e415.
- 36. Vermaat JS, Somers SF, de Wreede LC, et al. MYD88 mutations identify a molecular subgroup of diffuse large B-cell lymphoma with an unfavourable prognosis. *Haematologica*. 2020;105(2):424‑434.
- 37. Kurtz DM, Scherer F, Jin MC, et al. Circulating Tumor DNA Measurements As Early Outcome Predictors in Diffuse Large B‑Cell Lymphoma. *J Clin Oncol*. 2018;36(28):2845‑2853.
- 38. Dijkman R, Tensen CP, Buettner M, Niedobitek G, Willemze R, Vermeer MH. Primary cutaneous follicle center lymphoma and primary cutaneous large B‑cell lymphoma, leg type, are both targeted by aberrant somatic hypermutation but demonstrate differential expression of AID. *Blood*. 2006;107(12):4926‑4929.
- 39. Beylot‐Barry M, Mermin D, Maillard A, et al. A Single‐Arm Phase II Trial of Lenalidomide in Relapsing or Refractory Primary Cutaneous Large B‑Cell Lymphoma, Leg Type. *J Invest Dermatol*. 2018;138(9):1982‑1989.

- 40. Savini P, Lanzi A, Foschi FG, Marano G, Stefanini GF. Lenalidomide monotherapy in relapsed primary cutaneous diffuse large B cell lymphoma‑leg type. *Ann Hematol*. 2014;93(2):333‑334.
- 41. Swaika A, Menke DM, Jain MK, Sher T. Remission induction with lenalidomide in a patient with relapsed diffuse large B cell lymphoma of the leg type. *Ann Hematol*. 2015;94(5):895‑896.
- 42. Al Dhafiri M, Sicre de Fontbrune F, Marinho E, et al. Effectiveness of lenalidomide in relapsed primary cutaneous diffuse large B‑cell lymphoma, leg type. *Clin Case Rep*. 2019;7(5):964‑967.
- 43. Gupta E, Accurso J, Sluzevich J, Menke DM, Tun HW. Excellent Outcome of Immunomodulation or Bruton's Tyrosine Kinase Inhibition in Highly Refractory Primary Cutaneous Diffuse Large B‑Cell Lymphoma, Leg Type. *Rare Tumors*. 2015;7(4):6067.
- 44. Deng AL, Kim YR, Lichtenstein EA, O'Connor OA, Deng C. Combination of ibrutinib and chemotherapy produced a durable remission in multiply relapsed diffuse large B‑cell lymphoma leg type with mutant MYD88 and wildtype CD79. *Haematologica*. 2017;102(7):e275‑e277.
- 45. Pang A, Au‑Yeung R, Leung RYY, Kwong YL. Addictive response of primary cutaneous diffuse large B cell lymphoma leg type to low‑dose ibrutinib. *Ann Hematol*. 2019;98(10):2433‑2436.
- 46. Schick M, Habringer S, Nilsson JA, Keller U. Pathogenesis and therapeutic targeting of aberrant MYC expression in haematological cancers. *Br J Haematol*. 2017;179(5):724‑738.
- 47. Bose P, Simmons GL, Grant S. Cyclin-dependent kinase inhibitor therapy for hematologic malignancies. *Expert Opin Investig Drugs*. 2013;22(6):723‑738.
- 48. Kuhnl A, Cunningham D, Chau I. Beyond genomics ‑ Targeting the epigenome in diffuse large B‑cell lymphoma. *Cancer Treat Rev*. 2017;59:132‑137.

Supplemental Figure 1. Flow‑chart of study inclusion. Abbreviations: QC, quality control.

Supplemental Table 1. Average read count per patient with primary cutaneous diffuse large B-cell lymphoma, leg type

Abbreviations: P, primary sample; R, relapse sample.

Supplemental Table 2. Survival analysis of clinical factors in patients with primary cutaneous diffuse large B‑cell lymphoma, leg type

Abbreviations: HR, hazard ratio; CI, confidence interval; NR, not reliable.

**Results are less reliable because of <5 events in 1 of the groups.*

Remark: disease extension was excluded from analysis in disease‑specific survival for all patients and in overall‑survival, disease‑specific survival, and disease‑free survival for the systemically‑treated patients because of ≤1 event in the single lesion arm.