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CHAPTER 5

Single molecule as a local acoustic detector for mechanical oscillators

A single molecule can serve as a nanometer-sized detector of acoustic strain. Such a nanomicrophone has the great advantage that it can be placed very close to acoustic signal sources and high sensitivities can be achieved. We demonstrate this scheme by monitoring the fluorescence intensity of a single dibenzoterrylene molecule in an anthracene crystal attached to an oscillating tuning fork. The characterization of the vibration amplitude and of the detection sensitivity is a first step towards detection and control of nanomechanical oscillators through optical detection and feedback.

The content of this chapter is published.

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5.1.Introduction

Acoustic vibrations are central to communication in our daily lives, but also form the basis of many technologies such as sonar, seismography or ultrasound medical imaging. The main advantage of acoustic waves is their faculty to propagate in media which absorb or scatter electromagnetic waves. A major drive to achieving smaller, faster and, more sensitive sound detectors is the perspective of generating, detecting and controlling sound at nanometer scales, which would open new avenues for acoustic microscopy^{1,2}, as well as for manipulation and cooling of acoustic degrees of freedom down to the quantum regime³. Single gold nanoparticles have been shown to be sensitive vibration detectors in solution⁴. In recent years, thanks to spectacular progress in nanoscience, nanometer-scale oscillators have been applied to accurate mass measurements down to the mass of single atoms, and to bacterium screening^{5,6}. Nanomechanical oscillators are promising candidates as quantum systems that can be manipulated. The vibration amplitude and phase, and even the quantum state of a nanomechanical oscillator could be read through coupling to a quantum system such as a qubit, an optical cavity, a single-electron transistor, a SQUID or a point contact between two conductors⁷⁻¹⁰. Recently, Puller *et al.* have put forward the theoretical possibility to detect the displacement and to manipulate the state of a nanomechanical oscillator through the optical fluorescence signal of a single molecule¹¹. The aim of this chapter is to demonstrate such detection experimentally, and to provide measurements of the sensitivity in a well-controlled and well-understood case. The oscillator will be a quartz crystal tuning fork, causing mechanical deformations of the host crystal around the molecule under optical study.

To selectively detect individual molecules, the molecules have to be separated from each other^{12,13}. At cryogenic temperature and in suitable rigid matrixes, the absorption spectrum of a molecule presents an extremely narrow electronic transition that occurs without any creation or annihilation of phonons, and is therefore called the zero-phonon-line (ZPL). Its linewidth is chiefly determined by the lifetime of the excited state and lies in the range of 10-50 MHz for many well-studied host-guest systems¹³⁻¹⁹. Because of its sharpness, the ZPL is extremely sensitive to the molecule's local environment. The frequency of the ZPL can be shifted by mechanical strains or electric fields, including those caused by local degrees of freedom still

active at the cryogenic temperature of the experiment. Such dynamics lead to spectral diffusion or to spectral jumps of the ZPL^{20-22} , and should therefore be eliminated or minimized for sensing applications. For example, the ZPL is very sensitive to librations of any methyl groups present in the host matrix23. Static hydrostatic pressure is also a well-known factor which shifts the ZPLs of single molecules²⁴⁻²⁷. The present work also relates to an earlier discovery by our group of acoustic modes localized at defects of the anthracene crystal²⁸. Here, the tuning fork may be regarded as a well-known and well-controlled replacement for one of these localized modes. As we will see, its effects on the single-molecule lines are very comparable to those of the localized defect oscillators described previously.

The sensitivity of a molecular ZPL to the environment has its origin in the short-range interactions between the guest molecules and its first shells of host neighbors. Any variation of the respective positions of the guest and the host molecules induces a shift of the ZPL. Acoustic waves generated by a mechanical oscillator in contact with the sample will couple to the molecular transition and shift the ZPL of the guest molecule. We can thus use a single molecule to detect the local vibrations by monitoring the instantaneous frequency of its ZPL. In the present work, we chose single dibenzoterrylene (DBT) molecules embedded in an anthracene (Ac) crystal because of the stability of their ZPL, of their lifetime-limited linewidth and of their convenient wavelength 16 .

5.2.Experimental

The Ac single crystal doped with DBT molecules was glued to a quartz crystal tuning fork with a well-defined resonant oscillation frequency and high quality factor (Q factor), as shown in Fig. 5.1a. By electrically driving the tuning fork, the crystal is stretched or compressed periodically at the driving frequency. Such periodic vibrations change the average distance between molecules inside the crystal. The ZPLs of DBT molecules thus shift correspondingly, as shown in the cartoon of Fig. 5.1. Of course, the deformations of the molecular surroundings are much more complicated in reality and may involve fork's bending, shearing, and molecular

distortions. Any of these deformations, however, will give rise to a periodic shift of the molecular ZPL at the wave's frequency.

When the excitation laser is tuned to the wing of the ZPL as indicated by the dashed vertical line in Fig. 5.1b, a shift of the ZPL changes the molecular absorption and thereby the measured fluorescence intensity. In this way, the molecule's fluorescence can be used as a probe to read out the vibration amplitude, phase and frequency of the tuning fork, by monitoring timedependent changes in the fluorescence intensity.

Figure 5.1: (a) Sample mounting: an anthracene crystal doped with DBT was attached to the quartz crystal tuning fork; (b) the ZPL of a single DBT molecule is shifted upon deformation of its surrounding host crystal, as shown in (c) and (d). The real deformations are threedimensional and much more complicated, as molecules can also rotate and be distorted.

First, we characterized the tuning fork optically using the experimental set up described in Scheme 5.1. The excitation laser was focused on the edge of one of the tuning fork prongs. The reflected light was collected and detected by a photodiode, providing a signal modulated at the oscillating frequency of the tuning for k^{29} . The lock-in signal thus provides the modulation amplitude of the reflected light at the oscillation frequency of the tuning fork.

For the bare tuning fork in vacuum at 1.5 K, a sharp peak was obtained at frequency of 32.709 kHz with high *Q* factor of about 40,000 (Fig.5.2d), corresponding to the fundamental flexion mode (symmetrical oscillation of the two prongs)³⁰. When the sample crystal was attached, the oscillation of the tuning fork was strongly shifted and damped. Depending on the details of the contact between

the crystal and the fork, the frequency shift could be positive or negative. The *Q*-factor was always lowered by the crystal. Surprisingly, three peaks rather than a single peak were observed in the range from 10 kHz to 50 kHz, as shown in Fig. 5.2c. Certain crystal defects can present acoustic modes at these frequencies, as was reported in reference²⁸.

Scheme 5.1: Diagram of the confocal microscope used for the reflection measurements. The laser beam spot was focused on the edge of the tuning fork prong as shown in the insert. PD stands for photodiode.

However, our present experiments directly monitor the oscillation of the tuning fork and are therefore less sensitive to localized crystal modes. In addition, we did not observe any significant temperature dependence for these additional modes. We thus assign them to additional deformations of the tuning fork-crystal system, in addition to the strongest resonant peak located at 20.091 kHz which we attribute to the damped fundamental flexion mode. Theoretical calculation and modeling would be needed for a better understanding of the additional modes.

98 *Figure 5.2: a) Spectral trail of a single DBT molecule as a function of the driving frequency (a) from 19.820 kHz to 19.850 kHz or (b) from 20.060 kHz to 20.130 kHz. The driving voltage was 0.8 V. Lock-in signal of the reflection light intensity from the tuning fork with sample attached (c) and from the bare tuning fork (d). The asymmetry of the main resonance around 20.090 kHz (c) is due to the anharmonicity of the tuning fork's oscillation. All the measurements were done in vacuum at 1.5 K.*

5.3.Results and Discussion

Spectral trails of a single DBT molecule were obtained by repeatedly scanning the laser frequency (2 GHz, 3 s/scan) while slowly varying the driving frequency on the tuning fork (19.820 kHz to 19.850 kHz and 20.060 kHz to 20.130 kHz). Because of the limited fluorescence rate and time resolution, we could not directly monitor the oscillation of the ZPL shift. Instead, the periodical shift of the ZPL was observed as a broadening of the molecular line. As shown in Fig. 5.2a and 5.2b, the broadening of the molecular ZPL resonated at the specific frequencies found by monitoring the displacement of the tuning fork (Fig.5.2c). These results are evidence of the coupling between the DBT molecule and the vibrations generated by the tuning fork in the anthracene crystal. The slight asymmetry of the main peak is observed both on the lock-in signal from the fork (Fig. 5.2c) and on the molecular fluorescence (Fig.5.2b). It is due to a frequency shift at high oscillation amplitudes and indicates anharmonicity of the tuning fork's oscillations. We studied the distortions of the resonance line with the oscillation amplitude, and found that the anharmonic effect is much more pronounced by the presence of the crystal. Similar observations were made on the localized acoustic defect modes²⁸.

Figure 5.3: (a) Fluorescence intensity trace of a single DBT molecule (3,300 counts/s) and (b) Fourier transforms of such recordings when the tuning fork was driven at its resonant frequency (20.091 kHz), for various driving voltages (1 mV to 20 mV). The sharp peak at 20.091 kHz

indicates the modulation due to coupling to the tuning fork. This signal becomes comparable to noise at a driving voltage of 1 mV.

To estimate the detection sensitivity of a single molecule, we fixed the laser frequency to the half maximum position of the ZPL (as shown by the dashed line in Fig. 5.1b) and monitored the fluorescence photons from the molecule by time-tagged single-photon-counting when the tuning fork was driven at resonant frequency (20.091 kHz). The conted photons with bin size of 10μs over 10ms, 100ms and 1s are shown in Fig.5.3a. Because of the low fluorescence signal, we cannot directly see the modulation of the fluorescence intensity trace, not even for the largest oscillation amplitude. However, a fast-Fourier transform (FFT) of the fluorescence intensity trace reveals the weak modulation, as shown in Fig. 5.3b for different oscillation amplitudes. Note that, in spite of the random distribution of single photon detection events, the FFT picks up the weak modulated component unambiguously, just as a lock-in detection would filter an analog signal. The sharp peaks represent the driving frequency (their linewidth is determined by the function generator). The FFT signal amplitude varies linearly with the driving voltage, i.e., with the tuning fork's oscillation amplitude. As a consequence, the FFT signal can no longer be distinguished from noise, for about 1 mV driving voltage with an integration time of 1 s.

Hereafter, we discuss the detection sensitivity following the reasoning and notations of Puller et al.¹¹. We measure the oscillator's displacement in relative deformation of the host lattice $\varepsilon = \delta x / x$ rather than the absolute displacement of the nanotube's tip as reported¹¹. The sensitivity is determined by the smallest deformation detectable over a time *t* against shot noise fluctuations of the fluorescence signal, \sqrt{It} , where *I* is the fluorescence rate.

The shift of the molecular frequency is given by

$$
\delta v = \varepsilon \frac{dv}{d\varepsilon}.
$$

It gives rise to an intensity change

100

$$
\delta I = I \delta v / \Gamma \;,
$$

where Γ is the linewidth (30 MHz in previous experiments, 80 MHz in our case).

The minimum deformation value at which the signal overcomes the photon noise is thus:

$$
\varepsilon_m = \Gamma \frac{d\varepsilon}{d\nu} \sqrt{\frac{t}{I}}.
$$

The elastic modulus of anthracene crystal is $E \sim 10^{10} \text{ N/m}^2$ corresponding to 10^5 bar. $31,32$ The frequency shift induced by pressure change is reported²⁴ to be ~ 1 GHz/bar, i.e. a 1 GHz shift corresponds to a lattice deformation $\varepsilon = 10^{-5}$.

The coupling constant, corresponding to a change of frequency of the molecular transition per relative deformation of the crystal lattice is thus about

$$
\frac{dV}{d\varepsilon} = 10^{14} \text{ Hz}.
$$

Therefore, the typical values in single-molecule spectroscopy give, a deformation of 1 ppb/ \sqrt{Hz}

$$
\varepsilon_m = 10^{-9} \sqrt{t}
$$

With a low fluorescent rate of 3300 counts/s and a broad linewidth in our experiment, we would expect a sensitivity of about 1.4×10^{-8} Hz^{-1/2}. However, the experimental detection limit is rather 7×10^{-7} Hz^{-1/2}. We attribute this difference of a factor 50 to the background mechanical vibrations generated by the noisy environment of our cryostat. Indeed, a noise background appears in the FFT of the intensity trace. This noise background was also observed in the reflection measurement where no molecular spectroscopy was involved. Therefore, the oscillations of the tuning fork are driven by these noise sources.

We estimated the energy of these background oscillations. For this, we need the minimum detectable oscillation amplitude of the prongs, which was 0.1 nm. So the oscillation energy associated with this amplitude, can be deduced from the spring constant of the tuning fork $(k = 10000 \text{ N/m})$, to be around 10^{-17} J and found them to be much larger than thermal fluctuations (10⁻²³ J) expected at our experimental temperature (1.5 K). These vibrations ought to be eliminated to reach the shot-noise limited sensitivity of our single-molecule acoustic detector.

 We now discuss the theoretical detection limits for various oscillators. But first, we need to estimate the amplitude of the tuning fork with crystal under the experimental conditions. Assuming that the edges of the prongs are similar, the displacement of the prong would be linear with the lock-in signal. We then estimate the displacement of 0.14 nm/mV from the slope of the linear fit, by comparing the lock-in signal for the bare tuning fork and the tuning fork with crystal.

 It is worth to note that the edge of the tuning fork is not perfectly sharp at the nanometer scale. In this experiment, we have made some assumptions: 1) the focal spot has the same size for the bare tuning fork and for the tuning fork with crystal; 2) the edges of the prongs for both cases are the same; 3) by keeping the reflection intensity the same, we assume that the effect of laser power is excluded; 4) the lock-in signal is linearly dependent on displacement and linearly dependent on the driving voltage in this range.

Now that we know the vibration amplitude of the tuning fork's prongs, and that linearly depends on the driving voltage³³⁻³⁵, we can estimate the experimental sensitivity of the fork prong's displacement to be 0.14 nm/ \sqrt{Hz} .

Thus, in our experiments, the detection sensitivity is 0.14 nm/ \sqrt{Hz} , corresponding to a relative deformation of the host lattice of 7×10^{-7} (crystal size 200 µm).

If the shot-noise-limited sensitivity is achieved, much weaker displacements still are detectable with the molecule. Assuming the oscillation to be a mode delocalized over the whole crystal, typically 100 μ m in size (as presented in previous work²⁸), the detectable displacement would be about 1.4 pm/ \sqrt{Hz} in our present conditions, and would reach 100 fm/ \sqrt{Hz} with improvements in the fluorescence collection efficiency.

A nano-oscillator with 10 nm in size located around the molecule would give a sensitivity of 10 am/ \sqrt{Hz} . This sensitivity exceeds that of the device proposed by Puller et al. based on an oscillating carbon nanotube placed in the vicinity of the molecule¹¹.

Another important feature of our acoustic detector is its time resolution. The frequency of nanomechanical oscillators covers a large range from MHz to $THz^{36,37}$. The time resolution of our current setup is limited to 25 MHz by the dead time of our photodetector. Even with faster photodetector, the time resolution will be limited to some hundreds of MHz by the fluorescence lifetime of the molecules. Higher bandwidths would require faster emitters, for example molecules coupled to plasmonic antennas.

As a final remark, we rule out the possibility of a Stark shift of the molecular transition due to the applied electric field for driving the tuning for $k^{38,39}$. Indeed, Fig. 5.2 shows that the resonant acoustic effect is much larger than the frequency-independent Stark effect. Moreover, we can estimate the electric field created around the molecule to be 5 kV/m, for an applied voltage of 0.8 V on the tuning fork. Such a weak field cannot induce a significant Stark effect on the molecule^{28,39}. The observed ZPL shift of the DBT molecule is therefore exclusively due to the mechanical coupling between the molecule and the tuning fork.

5.4.Conclusion

In conclusion, we measured the coupling of a single organic molecule to acoustic strain generated by a macroscopic mechanical oscillator. Exciting the fluorescence at half-maximum gave rise to an intensity modulation of the fluorescence intensity trace. Such a weak was successfully detected trough a fast Fourier transform that works as a lock-in amplifier for a digital signal. The sensitivity threshold to relative deformation reached in our current experiments was about 7×10^{-7} Hz^{-1/2}, limited by mechanical experimental noise. However, for a shot-noise-limited detection of the vibrations of a nanomechanical oscillator, the sensitivity should reach the level of 1 fm/ \sqrt{Hz} , comparable to the theoretical sensitivity proposed by Puller $et al¹¹$. Such a high sensitivity is promising for reading out the quantum states of nanomechanical devices. Even though the detection sensitivity is not exceptionally high compared to other techniques, the main advantage of single-molecule detection is the small size of the sensor (sub-

nanometer in size). These small probes can be placed in the elastic strain field of the oscillator, enabling high coupling to artificial nanomechanical oscillators¹¹ or to natural oscillators found around defects in crystals²⁸ and defects in disordered solids⁴².

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Reference List

- 1. Dunn, F. Ultrasonic Absorption Microscope. *J. Acoust. Soc. Am.* **1959,** 31**,** 632.
- 2. R. G. Maev, *Acoustic Microscopy* (Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim, Germany, **2008**).
- 3. J. Gieseler, B. Deutsch, R. Quidant, and L. Novotny, *Phys. Rev. Lett*. **109,** 103603 (**2012**).
- 4. A. Ohlinger, A. Deak, A. A. Lutich, and J. Feldmann, Phys. Rev. Lett. **108,** 018101 (**2012**).
- 5. Calleja, and J. Tamayo, *Nat. Nanotechnol*. **2010,** 5, 641.
- 6. G. Longo, L. Alonso-Sarduy, L. M. Rio, A. Bizzini, A. Trampuz, J. Notz, G. Dietler, and S. Kasas, *Nat. Nanotechnol*. **2013,** 8, 522.
- 7. N. Flowers-Jacobs, D. Schmidt, and K. Lehnert, *Phys. Rev. Lett*. **98,** 096804 (**2007**).
- 8. S. Etaki, M. Poot, I. Mahboob, K. Onomitsu, H. Yamaguchi, and H. S. J. van der Zant, *Nat. Phys.* **2008,** 4, 785.
- 9. R. G. Knobel and A. N. Cleland, *Nature***. 2003,** 424, 291.
- 10. M. D. LaHaye, J. Suh, P. M. Echternach, K. C. Schwab, and M. L. Roukes, *Nature*. **2009,** 459, 960.
- 11. V. Puller, B. Lounis, and F. Pistolesi, *Phys. Rev. Lett*. **110,** 125501 (**2013**).
- 12. W. E. Moerner and M. Orrit, *Science*. **1999,** 283, 1670.
- 104
- 13. M. Orrit and J. Bernard, *Phys. Rev. Lett*. **65**, 2716 (**1990**).
- 14. A.-M. Boiron, F. Jelezko, Y. Durand, B. Lounis, and M. Orrit, *Mol. Cryst. Liq. Cryst. Sci. Technol. Sect. A.* **1996,** 291, 41.
- 15. F. Jelezko, P. Tamarat, B. Lounis, and M. Orrit, *J. Phys. Chem*. **1996,**100, 13892 ().
- 16. A. A. L. Nicolet, C. Hofmann, M. A. Kol'chenko, B. Kozankiewicz, and M. Orrit, *ChemPhysChem* **2007,** 8, 1215.
- 17. A. A. Gorshelev, A. V Naumov, I. Y. Eremchev, Y. G. Vainer, L. Kador, and J. Köhler, *ChemPhysChem*. **2010,** 11, 182.
- 18. A. Walser, A. Renn, S. Götzinger, and V. Sandoghdar, *Chem. Phys. Lett*. **2009,** 472, 44.
- 19. P. Navarro, Y. Tian, M. van Stee, and M. Orrit, *Chem.Phys.Chem.* **2014,** 15, 3032-3039*.*
- 20. W. P. Ambrose and W. E. Moerner, *Nature*. **1991,** 349, 225.
- 21. T. Basché, S. Kummer, and C. Bräuchle, *Nature*. **1995,** 373, 132.
- 22. F. Kulzer, S. Kummer, R. Matzke, C. Brauchle, and T. Basché, *Nature.* **1997,** 387, 688.
- 23. Y. Tian, P. Navarro, B. Kozankiewicz, and M. Orrit, *ChemPhysChem***. 2012,** 13, 3510.
- 24. A. A. L. Nicolet, P. Bordat, C. Hofmann, M. A. Kol'chenko, B. Kozankiewicz, R. Brown, and M. Orrit, *ChemPhysChem*. **2007,** 8, 1929.
- 25. M. Croci, H.-J. Müschenborn, F. Güttler, A. Renn, and U. P. Wild, *Chem. Phys. Lett.* **1993,** 212, 71.
- 26. T. Iwamoto, A. Kurita, and T. Kushida, *Chem. Phys. Lett.* **1998,** 284, 147.
- 27. A. Müller, W. Richter, and L. Kador, *Chem. Phys. Lett.* **1995**, 241, 547.
- 28. M. A. Kol'chenko, A. A. L. Nicolet, M. D. Galouzis, C. Hofmann, B. Kozankiewicz, and M. Orrit, *New J. Phys.* **2009,** 11, 023037.
- 29. D. van Vörden, M. Lange, M. Schmuck, N. Schmidt, and R. Möller, Beilstein *J. Nanotechnol.* **2012,** 3, 809.
- 30. J.-M. Friedt and E. Carry, *Am. J. Phys.* **2007,** 75, 415.
- 31. R. C. Dye and C. J. Eckhardt, *J. Chem. Phys.* **1989,** 90, 2090.
- 32. H. B. Huntington, *J. Chem. Phys.* **1969,** 50, 3844.
- 33. A. Castellanos-Gomez, C. R. Arroyo, N. Agraït, and G. Rubio-Bollinger, *Microsc. Microanal.* **2012,** 18, 353.
- 34. P. Sandoz, J.-M. Friedt, and E. Carry, *Rev. Sci. Instrum.* **2008,** 79, 086102.
- 35. A. G. Ruiter, K. O. van der Werf, J. A. Veerman, M. F. Garcia-Parajo, W. H. Rensen, and N. F. van Hulst, *Ultramicroscopy.* **1998,** 71, 149.
- 36. G. Anetsberger, O. Arcizet, Q. P. Unterreithmeier, R. Rivière, A. Schliesser, E. M. Weig, J. P. Kotthaus, and T. J. Kippenberg, *Nat. Phys.* **2009,** 5, 909.
- 37. V. V. Temnov, *Nat. Photonics.* **2012,** 6, 728.
- 38. M. Orrit, J. Bernard, A. Zumbusch, and R. I. Personov, *Chem. Phys. Lett.* **1992,**196, 595.
- 39. C. Brunel, P. Tamarat, B. Lounis, J. C. Woehl, and M. Orrit, *J. Phys. Chem. A*. **1999**, 103, 2429.
- 40. Y. Vainer, A. Naumov, M. Bauer, and L. Kador, *Phys. Rev. Lett.* **97**, 185501 **(2006).**