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Modulation of VLDL triglyceride metabolism

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Citation

Bijland, S. (2010, December 16). *Modulation of VLDL triglyceride metabolism*. Retrieved from <https://hdl.handle.net/1887/16248>

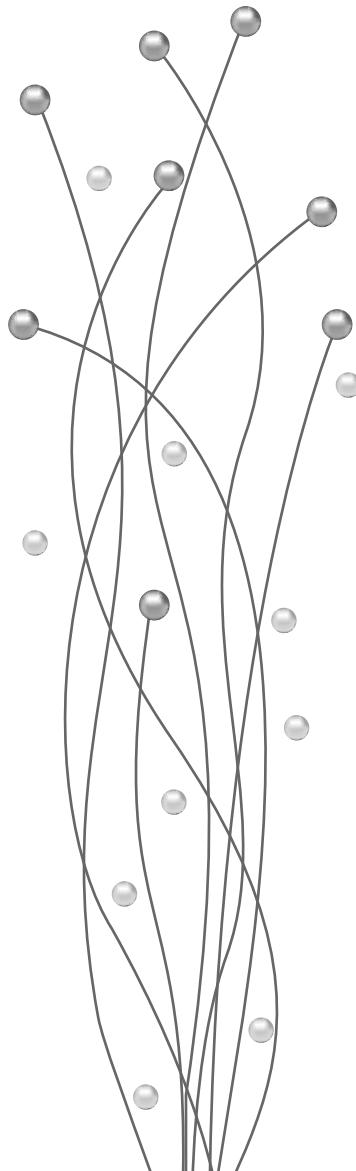
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CHAPTER 8



References

Summary

Nederlandse samenvatting
voor niet-ingewijden

List of publications

Curriculum vitae

1. Caballero, B. *The global epidemic of obesity: an overview*. Epidemiol. Rev. 29, 1-5 (2007).
2. Haslam, D. W. and James, W. P. *Obesity*. Lancet 366, 1197-1209 (2005).
3. Malhi, H. and Gores, G. J. *Molecular mechanisms of lipotoxicity in nonalcoholic fatty liver disease*. Semin. Liver Dis. 28, 360-369 (2008).
4. Yki-Jarvinen, H. *Fat in the liver and insulin resistance*. Ann. Med. 37, 347-356 (2005).
5. Capeau, J. *Insulin resistance and steatosis in humans*. Diabetes Metab 34, 649-657 (2008).
6. Mahley, R. W., Innerarity, T. L., Rall, S. C., Jr. et al. *Plasma lipoproteins: apolipoprotein structure and function*. J. Lipid Res. 25, 1277-1294 (1984).
7. Mu, H. and Hoy, C. E. *The digestion of dietary triacylglycerols*. Prog. Lipid Res. 43, 105-133 (2004).
8. Hussain, M. M., Kancha, R. K., Zhou, Z. et al. *Chylomicron assembly and catabolism: role of apolipoproteins and receptors*. Biochim. Biophys. Acta 1300, 151-170 (1996).
9. Green, P. H. and Riley, J. W. *Lipid absorption and intestinal lipoprotein formation*. Aust. N. Z. J. Med. 11, 84-90 (1981).
10. Ginsberg, H. N. *Lipoprotein physiology*. Endocrinol. Metab Clin. North Am. 27, 503-519 (1998).
11. Mahley, R. W., Hui, D. Y., Innerarity, T. L. et al. *Chylomicron remnant metabolism. Role of hepatic lipoprotein receptors in mediating uptake*. Arteriosclerosis 9, I14-I18 (1989).
12. Mahley, R. W. and Ji, Z. S. *Remnant lipoprotein metabolism: key pathways involving cell-surface heparan sulfate proteoglycans and apolipoprotein E*. J. Lipid Res. 40, 1-16 (1999).
13. Lusis, A. J. *Atherosclerosis*. Nature 407, 233-241 (2000).
14. Glass, C. K. and Witztum, J. L. *Atherosclerosis. the road ahead*. Cell 104, 503-516 (2001).
15. Zannis, V. I., Chroni, A., and Krieger, M. *Role of apoA-I, ABCA1, LCAT, and SR-BI in the biogenesis of HDL*. J. Mol. Med. 84, 276-294 (2006).
16. Huuskonen, J., Olkkonen, V. M., Jauhainen, M. et al. *The impact of phospholipid transfer protein (PLTP) on HDL metabolism*. Atherosclerosis 155, 269-281 (2001).
17. Tall, A. R. *An overview of reverse cholesterol transport*. Eur. Heart J. 19 Suppl A, A31-A35 (1998).
18. O'Brien, P. J., Alborn, W. E., Sloan, J. H. et al. *The novel apolipoprotein A5 is present in human serum, is associated with VLDL, HDL, and chylomicrons, and circulates at very low concentrations compared with other apolipoproteins*. Clin. Chem. 51, 351-359 (2005).

19. Out, R., Hoekstra, M., Spijkers, J. A. et al. *Scavenger receptor class B type I is solely responsible for the selective uptake of cholesteryl esters from HDL by the liver and the adrenals in mice.* J. Lipid Res. 45, 2088-2095 (2004).
20. Acton, S., Rigotti, A., Landschulz, K. T. et al. *Identification of scavenger receptor SR-BI as a high density lipoprotein receptor.* Science 271, 518-520 (1996).
21. Krieger, M. *Scavenger receptor class B type I is a multiligand HDL receptor that influences diverse physiologic systems.* J. Clin. Invest 108, 793-797 (2001).
22. Chiang, J. Y. *Regulation of bile acid synthesis: pathways, nuclear receptors, and mechanisms.* J. Hepatol. 40, 539-551 (2004).
23. Morton, R. E. and Zilversmit, D. B. *Inter-relationship of lipids transferred by the lipid-transfer protein isolated from human lipoprotein-deficient plasma.* J. Biol. Chem. 258, 11751-11757 (1983).
24. Drayna, D., Jarnagin, A. S., McLean, J. et al. *Cloning and sequencing of human cholesteryl ester transfer protein cDNA.* Nature 327, 632-634 (1987).
25. Gibbons, G. F. *Assembly and secretion of hepatic very-low-density lipoprotein.* Biochem. J. 268, 1-13 (1990).
26. Tietge, U. J., Bakillah, A., Maugeais, C. et al. *Hepatic overexpression of microsomal triglyceride transfer protein (MTP) results in increased in vivo secretion of VLDL triglycerides and apolipoprotein B.* J. Lipid Res. 40, 2134-2139 (1999).
27. Fisher, E. A. and Ginsberg, H. N. *Complexity in the secretory pathway: the assembly and secretion of apolipoprotein B-containing lipoproteins.* J. Biol. Chem. 277, 17377-17380 (2002).
28. Barrows, B. R. and Parks, E. J. *Contributions of different fatty acid sources to very low-density lipoprotein-triacylglycerol in the fasted and fed states.* J. Clin. Endocrinol. Metab 91, 1446-1452 (2006).
29. Donnelly, K. L., Smith, C. I., Schwarzenberg, S. J. et al. *Sources of fatty acids stored in liver and secreted via lipoproteins in patients with nonalcoholic fatty liver disease.* J. Clin. Invest 115, 1343-1351 (2005).
30. Minahk, C., Kim, K. W., Nelson, R. et al. *Conversion of low density lipoprotein-associated phosphatidylcholine to triacylglycerol by primary hepatocytes.* J. Biol. Chem. 283, 6449-6458 (2008).
31. Vance, D. E. *Role of phosphatidylcholine biosynthesis in the regulation of lipoprotein homeostasis.* Curr. Opin. Lipidol. 19, 229-234 (2008).
32. Wiggins, D. and Gibbons, G. F. *Origin of hepatic very-low-density lipoprotein triacylglycerol: the contribution of cellular phospholipid.* Biochem. J. 320 (Pt 2), 673-679 (1996).
33. Zechner, R. *The tissue-specific expression of lipoprotein lipase: implications for energy and lipoprotein metabolism.* Curr. Opin. Lipidol. 8, 77-88 (1997).
34. Goldberg, I. J. *Lipoprotein lipase and lipolysis: central roles in lipoprotein metabolism and atherosclerosis.* J. Lipid Res. 37, 693-707 (1996).

35. Beigneux, A. P., Weinstein, M. M., Davies, B. S. et al. *GPIHBP1 and lipolysis: an update*. Curr. Opin. Lipidol. 20, 211-216 (2009).
36. Sugden, M. C., Holness, M. J., and Howard, R. M. *Changes in lipoprotein lipase activities in adipose tissue, heart and skeletal muscle during continuous or interrupted feeding*. Biochem. J. 292 (Pt 1), 113-119 (1993).
37. Ruge, T., Wu, G., Olivecrona, T. et al. *Nutritional regulation of lipoprotein lipase in mice*. Int. J. Biochem. Cell Biol. 36, 320-329 (2004).
38. Wang, C. S. *Structure and functional properties of apolipoprotein C-II*. Prog. Lipid Res. 30, 253-258 (1991).
39. Schaap, F. G., Rensen, P. C., Voshol, P. J. et al. *ApoAV reduces plasma triglycerides by inhibiting very low density lipoprotein-triglyceride (VLDL-TG) production and stimulating lipoprotein lipase-mediated VLDL-TG hydrolysis*. J. Biol. Chem. 279, 27941-27947 (2004).
40. Merkel, M., Loeffler, B., Kluger, M. et al. *Apolipoprotein AV accelerates plasma hydrolysis of triglyceride-rich lipoproteins by interaction with proteoglycan-bound lipoprotein lipase*. J. Biol. Chem. 280, 21553-21560 (2005).
41. Ginsberg, H. N., Le, N. A., Goldberg, I. J. et al. *Apolipoprotein B metabolism in subjects with deficiency of apolipoproteins CIII and AI. Evidence that apolipoprotein CIII inhibits catabolism of triglyceride-rich lipoproteins by lipoprotein lipase in vivo*. J. Clin. Invest. 78, 1287-1295 (1986).
- 142 42. Havel, R. J., Fielding, C. J., Olivecrona, T. et al. *Cofactor activity of protein components of human very low density lipoproteins in the hydrolysis of triglycerides by lipoproteins lipase from different sources*. Biochemistry 12, 1828-1833 (1973).
43. Lichtenstein, L. and Kersten, S. *Modulation of plasma TG lipolysis by Angiopoietin-like proteins and GPIHBP1*. Biochim. Biophys. Acta 1801, 415-420 (2010).
44. Medh, J. D., Bowen, S. L., Fry, G. L. et al. *Lipoprotein lipase binds to low density lipoprotein receptors and induces receptor-mediated catabolism of very low density lipoproteins in vitro*. J. Biol. Chem. 271, 17073-17080 (1996).
45. Zimmermann, R., Strauss, J. G., Haemmerle, G. et al. *Fat mobilization in adipose tissue is promoted by adipose triglyceride lipase*. Science 306, 1383-1386 (2004).
46. Zimmermann, R., Lass, A., Haemmerle, G. et al. *Fate of fat: the role of adipose triglyceride lipase in lipolysis*. Biochim. Biophys. Acta 1791, 494-500 (2009).
47. Watt, M. J. and Steinberg, G. R. *Regulation and function of triacylglycerol lipases in cellular metabolism*. Biochem. J. 414, 313-325 (2008).
48. Su, X. and Abumrad, N. A. *Cellular fatty acid uptake: a pathway under construction*. Trends Endocrinol. Metab 20, 72-77 (2009).
49. Stremmel, W., Pohl, L., Ring, A. et al. *A new concept of cellular uptake and intracellular trafficking of long-chain fatty acids*. Lipids 36, 981-989 (2001).
50. van der Vusse, G. J., van Bilsen, M., Glatz, J. F. et al. *Critical steps in cellular fatty acid uptake and utilization*. Mol. Cell Biochem. 239, 9-15 (2002).

51. Briscoe, C. P., Tadayyon, M., Andrews, J. L. et al. *The orphan G protein-coupled receptor GPR40 is activated by medium and long chain fatty acids.* J. Biol. Chem. 278, 11303-11311 (2003).
52. Brown, A. J., Goldsworthy, S. M., Barnes, A. A. et al. *The Orphan G protein-coupled receptors GPR41 and GPR43 are activated by propionate and other short chain carboxylic acids.* J. Biol. Chem. 278, 11312-11319 (2003).
53. Itoh, Y., Kawamata, Y., Harada, M. et al. *Free fatty acids regulate insulin secretion from pancreatic beta cells through GPR40.* Nature 422, 173-176 (2003).
54. Oh Da, Y., Talukdar, S., Bae, E. J. et al. *GPR120 is an omega-3 fatty acid receptor mediating potent anti-inflammatory and insulin-sensitizing effects.* Cell 142, 687-698 (2010).
55. Xiong, Y., Miyamoto, N., Shibata, K. et al. *Short-chain fatty acids stimulate leptin production in adipocytes through the G protein-coupled receptor GPR41.* Proc. Natl. Acad. Sci. U. S. A 101, 1045-1050 (2004).
56. Hirasawa, A., Tsumaya, K., Awaji, T. et al. *Free fatty acids regulate gut incretin glucagon-like peptide-1 secretion through GPR120.* Nat. Med. 11, 90-94 (2005).
57. Le Poul, E., Loison, C., Struyf, S. et al. *Functional characterization of human receptors for short chain fatty acids and their role in polymorphonuclear cell activation.* J. Biol. Chem. 278, 25481-25489 (2003).
58. Sina, C., Gavrilova, O., Forster, M. et al. *G protein-coupled receptor 43 is essential for neutrophil recruitment during intestinal inflammation.* J. Immunol. 183, 7514-7522 (2009).
59. Glass, C. K. *Going nuclear in metabolic and cardiovascular disease.* J. Clin. Invest 116, 556-560 (2006).
60. Germain, P., Staels, B., Dacquet, C. et al. *Overview of nomenclature of nuclear receptors.* Pharmacol. Rev. 58, 685-704 (2006).
61. Daynes, R. A. and Jones, D. C. *Emerging roles of PPARs in inflammation and immunity.* Nat. Rev. Immunol. 2, 748-759 (2002).
62. Lefebvre, P., Chinetti, G., Fruchart, J. C. et al. *Sorting out the roles of PPAR alpha in energy metabolism and vascular homeostasis.* J. Clin. Invest 116, 571-580 (2006).
63. Staels, B., Vu-Dac, N., Kosykh, V. A. et al. *Fibrates downregulate apolipoprotein C-III expression independent of induction of peroxisomal acyl coenzyme A oxidase. A potential mechanism for the hypolipidemic action of fibrates.* J. Clin. Invest 95, 705-712 (1995).
64. Schoonjans, K., Peinado-Onsurbe, J., Lefebvre, A. M. et al. *PPARalpha and PPARgamma activators direct a distinct tissue-specific transcriptional response via a PPRE in the lipoprotein lipase gene.* EMBO J. 15, 5336-5348 (1996).
65. Martin, G., Schoonjans, K., Lefebvre, A. M. et al. *Coordinate regulation of the expression of the fatty acid transport protein and acyl-CoA synthetase genes by PPARalpha and PPARgamma activators.* J. Biol. Chem. 272, 28210-28217 (1997).

- 144
66. Motojima, K., Passilly, P., Peters, J. M. et al. *Expression of putative fatty acid transporter genes are regulated by peroxisome proliferator-activated receptor alpha and gamma activators in a tissue- and inducer-specific manner.* J. Biol. Chem. 273, 16710-16714 (1998).
 67. Brandt, J. M., Djouadi, F., and Kelly, D. P. *Fatty acids activate transcription of the muscle carnitine palmitoyltransferase I gene in cardiac myocytes via the peroxisome proliferator-activated receptor alpha.* J. Biol. Chem. 273, 23786-23792 (1998).
 68. Barak, Y., Nelson, M. C., Ong, E. S. et al. *PPAR gamma is required for placental, cardiac, and adipose tissue development.* Mol. Cell 4, 585-595 (1999).
 69. Rosen, E. D., Sarraf, P., Troy, A. E. et al. *PPAR gamma is required for the differentiation of adipose tissue in vivo and in vitro.* Mol. Cell 4, 611-617 (1999).
 70. Vidal-Puig, A., Jimenez-Linan, M., Lowell, B. B. et al. *Regulation of PPAR gamma gene expression by nutrition and obesity in rodents.* J. Clin. Invest 97, 2553-2561 (1996).
 71. Olswang, Y., Cohen, H., Papo, O. et al. *A mutation in the peroxisome proliferator-activated receptor gamma-binding site in the gene for the cytosolic form of phosphoenolpyruvate carboxykinase reduces adipose tissue size and fat content in mice.* Proc. Natl. Acad. Sci. U. S. A 99, 625-630 (2002).
 72. Kliewer, S. A., Forman, B. M., Blumberg, B. et al. *Differential expression and activation of a family of murine peroxisome proliferator-activated receptors.* Proc. Natl. Acad. Sci. U. S. A 91, 7355-7359 (1994).
 73. Wang, Y. X., Lee, C. H., Tiep, S. et al. *Peroxisome-proliferator-activated receptor delta activates fat metabolism to prevent obesity.* Cell 113, 159-170 (2003).
 74. Tanaka, T., Yamamoto, J., Iwasaki, S. et al. *Activation of peroxisome proliferator-activated receptor delta induces fatty acid beta-oxidation in skeletal muscle and attenuates metabolic syndrome.* Proc. Natl. Acad. Sci. U. S. A 100, 15924-15929 (2003).
 75. Brown, A. J., Sun, L., Feramisco, J. D. et al. *Cholesterol addition to ER membranes alters conformation of SCAP, the SREBP escort protein that regulates cholesterol metabolism.* Mol. Cell 10, 237-245 (2002).
 76. Repa, J. J., Liang, G., Ou, J. et al. *Regulation of mouse sterol regulatory element-binding protein-1c gene (SREBP-1c) by oxysterol receptors, LXRAalpha and LXRBeta.* Genes Dev. 14, 2819-2830 (2000).
 77. Watanabe, M., Houten, S. M., Wang, L. et al. *Bile acids lower triglyceride levels via a pathway involving FXR, SHP, and SREBP-1c.* J. Clin. Invest 113, 1408-1418 (2004).
 78. Horton, J. D., Goldstein, J. L., and Brown, M. S. *SREBPs: activators of the complete program of cholesterol and fatty acid synthesis in the liver.* J. Clin. Invest 109, 1125-1131 (2002).
 79. Ueda, A., Hamadeh, H. K., Webb, H. K. et al. *Diverse roles of the nuclear orphan receptor CAR in regulating hepatic genes in response to phenobarbital.* Mol. Pharmacol. 61, 1-6 (2002).

80. Kassam, A., Winrow, C. J., Fernandez-Rachubinski, F. et al. *The peroxisome proliferator response element of the gene encoding the peroxisomal beta-oxidation enzyme enoyl-CoA hydratase/3-hydroxyacyl-CoA dehydrogenase is a target for constitutive androstane receptor beta/9-cis-retinoic acid receptor-mediated transactivation.* J. Biol. Chem. 275, 4345-4350 (2000).
81. Nakamura, K., Moore, R., Negishi, M. et al. *Nuclear pregnane X receptor cross-talk with FoxA2 to mediate drug-induced regulation of lipid metabolism in fasting mouse liver.* J. Biol. Chem. 282, 9768-9776 (2007).
82. Zhou, J., Zhai, Y., Mu, Y. et al. *A novel pregnane X receptor-mediated and sterol regulatory element-binding protein-independent lipogenic pathway.* J. Biol. Chem. 281, 15013-15020 (2006).
83. Rezen, T., Tamasi, V., Lovgren-Sandblom, A. et al. *Effect of CAR activation on selected metabolic pathways in normal and hyperlipidemic mouse livers.* BMC. Genomics 10, 384- (2009).
84. Maglich, J. M., Lobe, D. C., and Moore, J. T. *The nuclear receptor CAR (NR1I3) regulates serum triglyceride levels under conditions of metabolic stress.* J. Lipid Res. 50, 439-445 (2009).
85. de Haan, W., de Vries-van der Weij, Mol, I. M. et al. *PXR agonism decreases plasma HDL levels in ApoE3-Leiden.CETP mice.* Biochim. Biophys. Acta 1791, 191-197 (2009).
86. Morere, P., Nouvet, G., Stain, J. P. et al. [Information obtained by liver biopsy in 100 tuberculous patients]. Sem. Hop. 51, 2095-2102 (1975).
87. Grieco, A., Forgirone, A., Miele, L. et al. *Fatty liver and drugs.* Eur. Rev. Med. Pharmacol. Sci. 9, 261-263 (2005).
88. Vega, R. B., Huss, J. M., and Kelly, D. P. *The coactivator PGC-1 cooperates with peroxisome proliferator-activated receptor alpha in transcriptional control of nuclear genes encoding mitochondrial fatty acid oxidation enzymes.* Mol. Cell Biol. 20, 1868-1876 (2000).
89. Bhalla, S., Ozalp, C., Fang, S. et al. *Ligand-activated pregnane X receptor interferes with HNF-4 signaling by targeting a common coactivator PGC-1alpha. Functional implications in hepatic cholesterol and glucose metabolism.* J. Biol. Chem. 279, 45139-45147 (2004).
90. Shiraki, T., Sakai, N., Kanaya, E. et al. *Activation of orphan nuclear constitutive androstane receptor requires subnuclear targeting by peroxisome proliferator-activated receptor gamma coactivator-1 alpha. A possible link between xenobiotic response and nutritional state.* J. Biol. Chem. 278, 11344-11350 (2003).
91. Lin, J., Yang, R., Tarr, P. T. et al. *Hyperlipidemic effects of dietary saturated fats mediated through PGC-1beta coactivation of SREBP.* Cell 120, 261-273 (2005).
92. Puigserver, P., Rhee, J., Donovan, J. et al. *Insulin-regulated hepatic gluconeogenesis through FOXO1-PGC-1alpha interaction.* Nature 423, 550-555 (2003).
93. Yoon, J. C., Puigserver, P., Chen, G. et al. *Control of hepatic gluconeogenesis through the transcriptional coactivator PGC-1.* Nature 413, 131-138 (2001).

94. Saltiel, A. R. and Kahn, C. R. *Insulin signalling and the regulation of glucose and lipid metabolism*. Nature 414, 799-806 (2001).
95. Bollen, M., Keppens, S., and Stalmans, W. *Specific features of glycogen metabolism in the liver*. Biochem. J. 336 (Pt 1), 19-31 (1998).
96. Exton, J. H. and Park, C. R. *Control of gluconeogenesis in liver. I. General features of gluconeogenesis in the perfused livers of rats*. J. Biol. Chem. 242, 2622-2636 (1967).
97. Reaven, G. M. *The insulin resistance syndrome: definition and dietary approaches to treatment*. Annu. Rev. Nutr. 25, 391-406 (2005).
98. Dekker, J. M., Girman, C., Rhodes, T. et al. *Metabolic syndrome and 10-year cardiovascular disease risk in the Hoorn Study*. Circulation 112, 666-673 (2005).
99. Jeppesen, J., Hansen, T. W., Rasmussen, S. et al. *Metabolic syndrome, low-density lipoprotein cholesterol, and risk of cardiovascular disease: a population-based study*. Atherosclerosis 189, 369-374 (2006).
100. Manninen, V., Tenkanen, L., Koskinen, P. et al. *Joint effects of serum triglyceride and LDL cholesterol and HDL cholesterol concentrations on coronary heart disease risk in the Helsinki Heart Study. Implications for treatment*. Circulation 85, 37-45 (1992).
101. Kim, J. K., Fillmore, J. J., Chen, Y. et al. *Tissue-specific overexpression of lipoprotein lipase causes tissue-specific insulin resistance*. Proc. Natl. Acad. Sci. U. S. A 98, 7522-7527 (2001).
102. Seppala-Lindroos, A., Vehkavaara, S., Hakkinen, A. M. et al. *Fat accumulation in the liver is associated with defects in insulin suppression of glucose production and serum free fatty acids independent of obesity in normal men*. J. Clin. Endocrinol. Metab 87, 3023-3028 (2002).
103. Virkamaki, A., Korsheninnikova, E., Seppala-Lindroos, A. et al. *Intramyocellular lipid is associated with resistance to in vivo insulin actions on glucose uptake, antilipolysis, and early insulin signaling pathways in human skeletal muscle*. Diabetes 50, 2337-2343 (2001).
104. den Boer, M. A., Voshol, P. J., Kuipers, F. et al. *Hepatic glucose production is more sensitive to insulin-mediated inhibition than hepatic VLDL-triglyceride production*. Am. J. Physiol Endocrinol. Metab 291, E1360-E1364 (2006).
105. den Boer, M., Voshol, P. J., Kuipers, F. et al. *Hepatic steatosis: a mediator of the metabolic syndrome. Lessons from animal models*. Arterioscler. Thromb. Vasc. Biol. 24, 644-649 (2004).
106. Rader, D. J. *Effect of insulin resistance, dyslipidemia, and intra-abdominal adiposity on the development of cardiovascular disease and diabetes mellitus*. Am. J. Med. 120, S12-S18 (2007).
107. Getz, G. S. and Reardon, C. A. *Diet and murine atherosclerosis*. Arterioscler. Thromb. Vasc. Biol. 26, 242-249 (2006).
108. van den Maagdenberg, A. M., Hofker, M. H., Krimpenfort, P. J. et al. *Transgenic mice carrying the apolipoprotein E3-Leiden gene exhibit hyperlipoproteinemia*. J. Biol. Chem. 268, 10540-10545 (1993).

109. de Knijff, P., van den Maagdenberg, A. M., Stalenhoef, A. F. et al. *Familial dysbeta lipoproteinemia associated with apolipoprotein E3-Leiden in an extended multigeneration pedigree.* J. Clin. Invest 88, 643-655 (1991).
110. van Vlijmen, B. J., van den Maagdenberg, A. M., Gijbels, M. J. et al. *Diet-induced hyperlipoproteinemia and atherosclerosis in apolipoprotein E3-Leiden transgenic mice.* J. Clin. Invest 93, 1403-1410 (1994).
111. Verschuren, L., Kleemann, R., Offerman, E. H. et al. *Effect of low dose atorvastatin versus diet-induced cholesterol lowering on atherosclerotic lesion progression and inflammation in apolipoprotein E³-Leiden transgenic mice.* Arterioscler. Thromb. Vasc. Biol. 25, 161-167 (2005).
112. Kleemann, R., Princen, H. M., Emeis, J. J. et al. *Rosuvastatin reduces atherosclerosis development beyond and independent of its plasma cholesterol-lowering effect in APOE³-Leiden transgenic mice: evidence for antiinflammatory effects of rosuvastatin.* Circulation 108, 1368-1374 (2003).
113. Delsing, D. J., Post, S. M., Groenendijk, M. et al. *Rosuvastatin reduces plasma lipids by inhibiting VLDL production and enhancing hepatobiliary lipid excretion in ApoE³-leiden mice.* J. Cardiovasc. Pharmacol. 45, 53-60 (2005).
114. van der Hoogt, C. C., de Haan, W., Westerterp, M. et al. *Fenofibrate increases HDL-cholesterol by reducing cholesteryl ester transfer protein expression.* J. Lipid Res. 48, 1763-1771 (2007).
115. Zadelaar, S., Kleemann, R., Verschuren, L. et al. *Mouse models for atherosclerosis and pharmaceutical modifiers.* Arterioscler. Thromb. Vasc. Biol. 27, 1706-1721 (2007).
116. Westerterp, M., van der Hoogt, C. C., de Haan, W. et al. *Cholesteryl ester transfer protein decreases high-density lipoprotein and severely aggravates atherosclerosis in APOE³-Leiden mice.* Arterioscler. Thromb. Vasc. Biol. 26, 2552-2559 (2006).
117. van der Hoorn, J. W., de Haan, W., Berbee, J. F. et al. *Niacin increases HDL by reducing hepatic expression and plasma levels of cholesteryl ester transfer protein in APOE³Leiden.CETP mice.* Arterioscler. Thromb. Vasc. Biol. 28, 2016-2022 (2008).
118. de Haan, W., van der Hoogt, C. C., Westerterp, M. et al. *Atorvastatin increases HDL cholesterol by reducing CETP expression in cholesterol-fed APOE³-Leiden. CETP mice.* Atherosclerosis 197, 57-63 (2008).
119. de Vries-van der Weij, de Haan, W., Hu, L. et al. *Bexarotene induces dyslipidemia by increased very low-density lipoprotein production and cholesteryl ester transfer protein-mediated reduction of high-density lipoprotein.* Endocrinology 150, 2368-2375 (2009).
120. Boden, W. E. *High-density lipoprotein cholesterol as an independent risk factor in cardiovascular disease: assessing the data from Framingham to the Veterans Affairs High-Density Lipoprotein Intervention Trial.* Am. J. Cardiol. 86, 19L-22L (2000).
121. Nordestgaard, B. G., Benn, M., Schnohr, P. et al. *Nonfasting triglycerides and risk of myocardial infarction, ischemic heart disease, and death in men and women.* JAMA 298, 299-308 (2007).

122. Barter, P. J., Caulfield, M., Eriksson, M. et al. *Effects of torcetrapib in patients at high risk for coronary events.* N. Engl. J. Med. 357, 2109-2122 (2007).
123. Bloomfield, D., Carlson, G. L., Sarep, A. et al. *Efficacy and safety of the cholesteryl ester transfer protein inhibitor anacetrapib as monotherapy and coadministered with atorvastatin in dyslipidemic patients.* Am. Heart J. 157, 352-360 (2009).
124. Rennings, A. J. and Stalenhoef, A. F. *JTT-705: is there still future for a CETP inhibitor after torcetrapib?* Expert. Opin. Investig. Drugs 17, 1589-1597 (2008).
125. Clark, R. W., Sutfin, T. A., Ruggeri, R. B. et al. *Raising high-density lipoprotein in humans through inhibition of cholesteryl ester transfer protein: an initial multidose study of torcetrapib.* Arterioscler. Thromb. Vasc. Biol. 24, 490-497 (2004).
126. de Groot, G. J., Kuivenhoven, J. A., Stalenhoef, A. F. et al. *Efficacy and safety of a novel cholesteryl ester transfer protein inhibitor, JTT-705, in humans: a randomized phase II dose-response study.* Circulation 105, 2159-2165 (2002).
127. Hermann, F., Enseleit, F., Spieker, L. E. et al. *Cholesterylestertransfer protein inhibition and endothelial function in type II hyperlipidemia.* Thromb. Res. 123, 460-465 (2009).
128. Rashid, S., Trinh, D. K., Uffelman, K. D. et al. *Expression of human hepatic lipase in the rabbit model preferentially enhances the clearance of triglyceride-enriched versus native high-density lipoprotein apolipoprotein A-I.* Circulation 107, 3066-3072 (2003).
129. Jiang, X. C., Agellon, L. B., Walsh, A. et al. *Dietary cholesterol increases transcription of the human cholesteryl ester transfer protein gene in transgenic mice. Dependence on natural flanking sequences.* J. Clin. Invest 90, 1290-1295 (1992).
130. Berbee, J. F., vander Hoogt, C. C., Sundararaman, D. et al. *Severe hypertriglyceridemia in human APOC1 transgenic mice is caused by apoC-I-induced inhibition of LPL.* J. Lipid Res. 46, 297-306 (2005).
131. Aalto-Setala, K., Fisher, E. A., Chen, X. et al. *Mechanism of hypertriglyceridemia in human apolipoprotein (apo) CIII transgenic mice. Diminished very low density lipoprotein fractional catabolic rate associated with increased apo CIII and reduced apo E on the particles.* J. Clin. Invest 90, 1889-1900 (1992).
132. Pietzsch, J., Subat, S., Nitzsche, S. et al. *Very fast ultracentrifugation of serum lipoproteins: influence on lipoprotein separation and composition.* Biochim. Biophys. Acta 1254, 77-88 (1995).
133. Li, X., Catalina, F., Grundy, S. M. et al. *Method to measure apolipoprotein B-48 and B-100 secretion rates in an individual mouse: evidence for a very rapid turnover of VLDL and preferential removal of B-48- relative to B-100-containing lipoproteins.* J. Lipid Res. 37, 210-220 (1996).
134. Egusa, G., Brady, D. W., Grundy, S. M. et al. *Isopropanol precipitation method for the determination of apolipoprotein B specific activity and plasma concentrations during metabolic studies of very low density lipoprotein and low density lipoprotein apolipoprotein B.* J. Lipid Res. 24, 1261-1267 (1983).

135. Rensen, P. C., Herijgers, N., Netscher, M. H. et al. *Particle size determines the specificity of apolipoprotein E-containing triglyceride-rich emulsions for the LDL receptor versus hepatic remnant receptor in vivo.* J. Lipid Res. 38, 1070-1084 (1997).
136. Teusink, B., Voshol, P. J., Dahlmans, V. E. et al. *Contribution of fatty acids released from lipolysis of plasma triglycerides to total plasma fatty acid flux and tissue-specific fatty acid uptake.* Diabetes 52, 614-620 (2003).
137. Rensen, P. C., Jong, M. C., van Vark, L. C. et al. *Apolipoprotein E is resistant to intracellular degradation in vitro and in vivo. Evidence for retroendocytosis.* J. Biol. Chem. 275, 8564-8571 (2000).
138. Voshol, P. J., Rensen, P. C., van Dijk, K. W. et al. *Effect of plasma triglyceride metabolism on lipid storage in adipose tissue: studies using genetically engineered mouse models.* Biochim. Biophys. Acta 1791, 479-485 (2009).
139. Escola-Gil, J. C., Julve, J., Marzal-Casacuberta, A. et al. *ApoA-II expression in CETP transgenic mice increases VLDL production and impairs VLDL clearance.* J. Lipid Res. 42, 241-248 (2001).
140. Salerno, A. G., Patricio, P. R., Berti, J. A. et al. *Cholesteryl ester transfer protein (CETP) increases postprandial triglyceridemia and delays triglyceride plasma clearance in transgenic mice.* Biochem. J. 419, 629-634 (2009).
141. Harada, L. M., Amigo, L., Cazita, P. M. et al. *CETP expression enhances liver HDL-cholesteryl ester uptake but does not alter VLDL and biliary lipid secretion.* Atherosclerosis 191, 313-318 (2007).
142. Lamarche, B., Uffelman, K. D., Carpentier, A. et al. *Triglyceride enrichment of HDL enhances in vivo metabolic clearance of HDL apo A-I in healthy men.* J. Clin. Invest 103, 1191-1199 (1999).
143. Melchior, G. W., Castle, C. K., Murray, R. W. et al. *Apolipoprotein A-I metabolism in cholesteryl ester transfer protein transgenic mice. Insights into the mechanisms responsible for low plasma high density lipoprotein levels.* J. Biol. Chem. 269, 8044-8051 (1994).
144. Rashid, S., Barrett, P. H., Uffelman, K. D. et al. *Lipolytically modified triglyceride-enriched HDLs are rapidly cleared from the circulation.* Arterioscler. Thromb. Vasc. Biol. 22, 483-487 (2002).
145. Chapman, M. J. *Fibrates in 2003: therapeutic action in atherogenic dyslipidaemia and future perspectives.* Atherosclerosis 171, 1-13 (2003).
146. Schoonjans, K., Staels, B., and Auwerx, J. *The peroxisome proliferator activated receptors (PPARS) and their effects on lipid metabolism and adipocyte differentiation.* Biochim. Biophys. Acta 1302, 93-109 (1996).
147. Staels, B., Dallongeville, J., Auwerx, J. et al. *Mechanism of action of fibrates on lipid and lipoprotein metabolism.* Circulation 98, 2088-2093 (1998).
148. Hogue, J. C., Lamarche, B., Deshaies, Y. et al. *Differential effect of fenofibrate and atorvastatin on in vivo kinetics of apolipoproteins B-100 and B-48 in subjects with type 2 diabetes mellitus with marked hypertriglyceridemia.* Metabolism 57, 246-254 (2008).

149. Watts, G. F., Ji, J., Chan, D. C. et al. Relationships between changes in plasma lipid transfer proteins and apolipoprotein B-100 kinetics during fenofibrate treatment in the metabolic syndrome. *Clin. Sci. (Lond)* 111, 193-199 (2006).
150. Linden, D., Alsterholm, M., Wennbo, H. et al. PPAR α deficiency increases secretion and serum levels of apolipoprotein B-containing lipoproteins. *J. Lipid Res.* 42, 1831-1840 (2001).
151. Tordjman, K., Bernal-Mizrachi, C., Zemany, L. et al. PPAR α deficiency reduces insulin resistance and atherosclerosis in apoE-null mice. *J. Clin. Invest* 107, 1025-1034 (2001).
152. Kersten, S. Peroxisome proliferator activated receptors and lipoprotein metabolism. *PPAR. Res.* 2008, 132960- (2008).
153. Hahn, S. E. and Goldberg, D. M. Modulation of lipoprotein production in Hep G2 cells by fenofibrate and clofibrate. *Biochem. Pharmacol.* 43, 625-633 (1992).
154. Lamb, R. G., Koch, J. C., and Bush, S. R. An enzymatic explanation of the differential effects of oleate and gemfibrozil on cultured hepatocyte triacylglycerol and phosphatidylcholine biosynthesis and secretion. *Biochim. Biophys. Acta* 1165, 299-305 (1993).
155. Zechner, R. Rapid and simple isolation procedure for lipoprotein lipase from human milk. *Biochim. Biophys. Acta* 1044, 20-25 (1990).
156. Bligh, E. G. and Dyer, W. J. A rapid method of total lipid extraction and purification. *Can. J. Biochem. Physiol* 37, 911-917 (1959).
157. de Groot, P. J., Reiff, C., Mayer, C. et al. NuGO contributions to GenePattern. *Genes Nutr.* 3, 143-146 (2008).
158. Storey, J. D. and Tibshirani, R. Statistical significance for genomewide studies. *Proc. Natl. Acad. Sci. U. S. A* 100, 9440-9445 (2003).
159. Boorsma, A., Foat, B. C., Vis, D. et al. T-profiler: scoring the activity of predefined groups of genes using gene expression data. *Nucleic Acids Res.* 33, W592-W595 (2005).
160. Reich, M., Liefeld, T., Gould, J. et al. GenePattern 2.0. *Nat. Genet.* 38, 500-501 (2006).
161. Adiels, M., Taskinen, M. R., Packard, C. et al. Overproduction of large VLDL particles is driven by increased liver fat content in man. *Diabetologia* 49, 755-765 (2006).
162. Guerin, M., Bruckert, E., Dolphin, P. J. et al. Fenofibrate reduces plasma cholesteryl ester transfer from HDL to VLDL and normalizes the atherogenic, dense LDL profile in combined hyperlipidemia. *Arterioscler. Thromb. Vasc. Biol.* 16, 763-772 (1996).
163. Heller, F. and Harvengt, C. Effects of clofibrate, bezafibrate, fenofibrate and probucol on plasma lipolytic enzymes in normolipaemic subjects. *Eur. J. Clin. Pharmacol.* 25, 57-63 (1983).

164. Malmendier, C. L., Lontie, J. F., Delcroix, C. et al. *Apolipoproteins C-II and C-III metabolism in hypertriglyceridemic patients. Effect of a drastic triglyceride reduction by combined diet restriction and fenofibrate administration.* Atherosclerosis 77, 139-149 (1989).
165. Aoyama, T., Peters, J. M., Iritani, N. et al. *Altered constitutive expression of fatty acid-metabolizing enzymes in mice lacking the peroxisome proliferator-activated receptor alpha (PPAR α).* J. Biol. Chem. 273, 5678-5684 (1998).
166. Gulick, T., Cresci, S., Caira, T. et al. *The peroxisome proliferator-activated receptor regulates mitochondrial fatty acid oxidative enzyme gene expression.* Proc. Natl. Acad. Sci. U. S. A 91, 11012-11016 (1994).
167. Shiri-Sverdlov, R., Wouters, K., van Gorp, P. J. et al. *Early diet-induced non-alcoholic steatohepatitis in APOE2 knock-in mice and its prevention by fibrates.* J. Hepatol. 44, 732-741 (2006).
168. Ameen, C., Edvardsson, U., Ljungberg, A. et al. *Activation of peroxisome proliferator-activated receptor α increases the expression and activity of microsomal triglyceride transfer protein in the liver.* J. Biol. Chem. 280, 1224-1229 (2005).
169. Duval, C., Muller, M., and Kersten, S. *PPAR α and dyslipidemia.* Biochim. Biophys. Acta 1771, 961-971 (2007).
170. Watts, G. F., Barrett, P. H., Ji, J. et al. *Differential regulation of lipoprotein kinetics by atorvastatin and fenofibrate in subjects with the metabolic syndrome.* Diabetes 52, 803-811 (2003).
171. Duivenvoorden, I., Teusink, B., Rensen, P. C. et al. *Acute inhibition of hepatic beta-oxidation in APOE $^{\star}3$ Leiden mice does not affect hepatic VLDL secretion or insulin sensitivity.* J. Lipid Res. 46, 988-993 (2005).
172. Linden, D., Lindberg, K., Oscarsson, J. et al. *Influence of peroxisome proliferator-activated receptor α agonists on the intracellular turnover and secretion of apolipoprotein (Apo) B-100 and ApoB-48.* J. Biol. Chem. 277, 23044-23053 (2002).
173. Srivastava, R. A., Jahagirdar, R., Azhar, S. et al. *Peroxisome proliferator-activated receptor- α selective ligand reduces adiposity, improves insulin sensitivity and inhibits atherosclerosis in LDL receptor-deficient mice.* Mol. Cell Biochem. 285, 35-50 (2006).
174. Hall, R. G., Leff, R. D., and Gumbo, T. *Treatment of active pulmonary tuberculosis in adults: current standards and recent advances. Insights from the Society of Infectious Diseases Pharmacists.* Pharmacotherapy 29, 1468-1481 (2009).
175. Austerhoff, A., Kindler, U., Knop, P. et al. *[Liver toxicity of combined rifampicin-isoniazid-ethambutol medication (author's transl)].* Dtsch. Med. Wochenschr. 99, 1182- (1974).
176. Pilheu, J. A., De Salvo, M. C., and Barcat, J. A. *[Effect of isoniazid and rifampicin regimens on the liver of tuberculosis patients].* Medicina (B Aires) 39, 298-304 (1979).
177. Taranger, J., Girbal, J. P., and Giacchero, G. *[Rifampicin and liver function (72 punctures biopsies)].* Rev. Tuberc. Pneumol. (Paris) 34, 717-720 (1970).

178. Khogali, A. M., Chazan, B. I., Metcalf, V. J. et al. *Hyperlipidaemia as a complication of rifampicin treatment.* *Tubercle.* 55, 231-233 (1974).
179. Lehmann, J. M., McKee, D. D., Watson, M. A. et al. *The human orphan nuclear receptor PXR is activated by compounds that regulate CYP3A4 gene expression and cause drug interactions.* *J. Clin. Invest.* 102, 1016-1023 (1998).
180. Piriou, A., Warnet, J. M., Jacqueson, A. et al. *Fatty liver induced by high doses of rifampicin in the rat: possible relation with an inhibition of RNA polymerases in eukaryotic cells.* *Arch. Toxicol. Suppl* 333-337 (1979).
181. Piriou, A., Maissiat, R., Jacqueson, A. et al. *Ultrastructural changes in the parenchymal liver cells of rats treated with high doses of rifampicin.* *Br. J. Exp. Pathol.* 68, 201-207 (1987).
182. Hoekstra, M., Lammers, B., Out, R. et al. *Activation of the nuclear receptor PXR decreases plasma LDL-cholesterol levels and induces hepatic steatosis in LDL receptor knockout mice.* *Mol. Pharm.* 6, 182-189 (2009).
183. Berbee, J. F., Havekes, L. M., and Rensen, P. C. *Apolipoproteins modulate the inflammatory response to lipopolysaccharide.* *J. Endotoxin. Res.* 11, 97-103 (2005).
184. Speijer, H., Groener, J. E., van, Ramshorst E. et al. *Different locations of cholestryly ester transfer protein and phospholipid transfer protein activities in plasma.* *Atherosclerosis* 90, 159-168 (1991).
185. de Haan, W., de Vries-van der Weij, J., van der Hoorn, J. W. et al. *Torcetrapib does not reduce atherosclerosis beyond atorvastatin and induces more proinflammatory lesions than atorvastatin.* *Circulation* 117, 2515-2522 (2008).
186. Gautier, T., Tietge, U. J., Boverhof, R. et al. *Hepatic lipid accumulation in apolipoprotein C-I-deficient mice is potentiated by cholestryly ester transfer protein.* *J. Lipid Res.* 48, 30-40 (2007).
187. van Eck, M., Twisk, J., Hoekstra, M. et al. *Differential effects of scavenger receptor BI deficiency on lipid metabolism in cells of the arterial wall and in the liver.* *J. Biol. Chem.* 278, 23699-23705 (2003).
188. Williamson, R., Lee, D., Hagaman, J. et al. *Marked reduction of high density lipoprotein cholesterol in mice genetically modified to lack apolipoprotein A-I.* *Proc. Natl. Acad. Sci. U. S. A* 89, 7134-7138 (1992).
189. Sporstol, M., Tapia, G., Malerod, L. et al. *Pregnane X receptor-agonists down-regulate hepatic ATP-binding cassette transporter A1 and scavenger receptor class B type I.* *Biochem. Biophys. Res. Commun.* 331, 1533-1541 (2005).
190. Sakai, N., Vaisman, B. L., Koch, C. A. et al. *Targeted disruption of the mouse lecithin:cholesterol acyltransferase (LCAT) gene. Generation of a new animal model for human LCAT deficiency.* *J. Biol. Chem.* 272, 7506-7510 (1997).
191. Jiang, X. C., Bruce, C., Mar, J. et al. *Targeted mutation of plasma phospholipid transfer protein gene markedly reduces high-density lipoprotein levels.* *J. Clin. Invest.* 103, 907-914 (1999).

192. Le Goff, W., Guerin, M., and Chapman, M. J. *Pharmacological modulation of cholesteryl ester transfer protein, a new therapeutic target in atherogenic dyslipidemia.* Pharmacol. Ther. 101, 17-38 (2004).
193. Rigotti, A., Trigatti, B. L., Penman, M. et al. *A targeted mutation in the murine gene encoding the high density lipoprotein (HDL) receptor scavenger receptor class B type I reveals its key role in HDL metabolism.* Proc. Natl. Acad. Sci. U. S. A 94, 12610-12615 (1997).
194. Kiss E. *Fluorinated surfactants and repellents.* (2001).
195. Calafat, A. M., Wong, L. Y., Kuklenyik, Z. et al. *Polyfluoroalkyl chemicals in the U.S. population: data from the National Health and Nutrition Examination Survey (NHANES) 2003-2004 and comparisons with NHANES 1999-2000.* Environ. Health Perspect. 115, 1596-1602 (2007).
196. Xu, L., Krenitsky, D. M., Seacat, A. M. et al. *Biotransformation of N-ethyl-N-(2-hydroxyethyl)perfluoroctanesulfonamide by rat liver microsomes, cytosol, and slices and by expressed rat and human cytochromes P450.* Chem. Res. Toxicol. 17, 767-775 (2004).
197. Xu, L., Krenitsky, D. M., Seacat, A. M. et al. *N-glucuronidation of perfluoroctanesulfonamide by human, rat, dog, and monkey liver microsomes and by expressed rat and human UDP-glucuronosyltransferases.* Drug Metab Dispos. 34, 1406-1410 (2006).
198. D'eon, J. C., Hurley, M. D., Wallington, T. J. et al. *Atmospheric chemistry of N-methyl perfluorobutane sulfonamidoethanol, C4F9SO2N(CH₃)CH₂CH₂OH: kinetics and mechanism of reaction with OH.* Environ. Sci. Technol. 40, 1862-1868 (2006).
199. Giesy, J. P. and Kannan, K. *Global distribution of perfluoroctane sulfonate in wildlife.* Environ. Sci. Technol. 35, 1339-1342 (2001).
200. Hansen, K. J., Clemen, L. A., Ellefson, M. E. et al. *Compound-specific, quantitative characterization of organic fluoroochemicals in biological matrices.* Environ. Sci. Technol. 35, 766-770 (2001).
201. Olsen, G. W., Burris, J. M., Ehresman, D. J. et al. *Half-life of serum elimination of perfluoroctanesulfonate, perfluorohexanesulfonate, and perfluoroctanoate in retired fluoroochemical production workers.* Environ. Health Perspect. 115, 1298-1305 (2007).
202. Lau, C., Anitole, K., Hodes, C. et al. *Perfluoroalkyl acids: a review of monitoring and toxicological findings.* Toxicol. Sci. 99, 366-394 (2007).
203. Olsen, G. W., Chang, S. C., Noker, P. E. et al. *A comparison of the pharmacokinetics of perfluorobutanesulfonate (PFBS) in rats, monkeys, and humans.* Toxicology 256, 65-74 (2009).
204. Nelson, J. W., Hatch, E. E., and Webster, T. F. *Exposure to polyfluoroalkyl chemicals and cholesterol, body weight, and insulin resistance in the general U.S. population.* Environ. Health Perspect. 118, 197-202 (2010).
205. Steenland, K., Tinker, S., Frisbee, S. et al. *Association of perfluoroctanoic acid and perfluoroctane sulfonate with serum lipids among adults living near a chemical plant.* Am. J. Epidemiol. 170, 1268-1278 (2009).

206. Olsen, G. W., Burris, J. M., Burlew, M. M. et al. *Epidemiologic assessment of worker serum perfluorooctanesulfonate (PFOS) and perfluorooctanoate (PFOA) concentrations and medical surveillance examinations*. *J. Occup. Environ. Med.* 45, 260-270 (2003).
207. Seacat, A. M., Thomford, P. J., Hansen, K. J. et al. *Subchronic toxicity studies on perfluorooctanesulfonate potassium salt in cynomolgus monkeys*. *Toxicol. Sci.* 68, 249-264 (2002).
208. Curran, I., Hierlihy, S. L., Liston, V. et al. *Altered fatty acid homeostasis and related toxicologic sequelae in rats exposed to dietary potassium perfluorooctanesulfonate (PFOS)*. *J. Toxicol. Environ. Health A* 71, 1526-1541 (2008).
209. Haughom, B. and Spydevold, O. *The mechanism underlying the hypolipemic effect of perfluorooctanoic acid (PFOA), perfluorooctane sulphonic acid (PFOSA) and clofibrate acid*. *Biochim. Biophys. Acta* 1128, 65-72 (1992).
210. Martin, M. T., Brennan, R. J., Hu, W. et al. *Toxicogenomic study of triazole fungicides and perfluoroalkyl acids in rat livers predicts toxicity and categorizes chemicals based on mechanisms of toxicity*. *Toxicol. Sci.* 97, 595-613 (2007).
211. Manal, A., Abd El-Nasser, M. A., Shaaban, A. A. et al. *Toxicological effects of perfluoroalkyl acids on pregnant female mice*. *Ass. Univ. Environ. Res.* 12, 23-39 (2009).
212. Butenhoff, J. L., Chang, S. C., Ehresman, D. J. et al. *Evaluation of potential reproductive and developmental toxicity of potassium perfluorohexanesulfonate in Sprague Dawley rats*. *Reprod. Toxicol.* 27, 331-341 (2009).
213. Lieder, P. H., Chang, S. C., York, R. G. et al. *Toxicological evaluation of potassium perfluorobutanesulfonate in a 90-day oral gavage study with Sprague-Dawley rats*. *Toxicology* 255, 45-52 (2009).
214. Ehresman, D. J., Froehlich, J. W., Olsen, G. W. et al. *Comparison of human whole blood, plasma, and serum matrices for the determination of perfluorooctanesulfonate (PFOS), perfluorooctanoate (PFOA), and other fluorocompounds*. *Environ. Res.* 103, 176-184 (2007).
215. Post, S. M., Groenendijk, M., Solaas, K. et al. *Cholesterol 7alpha-hydroxylase deficiency in mice on an APOE*3-Leiden background impairs very-low-density lipoprotein production*. *Arterioscler. Thromb. Vasc. Biol.* 24, 768-774 (2004).
216. Post, S. M., de Roos, B., Vermeulen, M. et al. *Cafestol increases serum cholesterol levels in apolipoprotein E*3-Leiden transgenic mice by suppression of bile acid synthesis*. *Arterioscler. Thromb. Vasc. Biol.* 20, 1551-1556 (2000).
217. Post, S. M., de Crom, R., van Haperen, R. et al. *Increased fecal bile acid excretion in transgenic mice with elevated expression of human phospholipid transfer protein*. *Arterioscler. Thromb. Vasc. Biol.* 23, 892-897 (2003).
218. Moreau, A., Vilarem, M. J., Maurel, P. et al. *Xenoreceptors CAR and PXR activation and consequences on lipid metabolism, glucose homeostasis, and inflammatory response*. *Mol. Pharm.* 5, 35-41 (2008).

219. Bijland, S., Pieterman, E. J., Maas, A. C. et al. *Fenofibrate increases very low density lipoprotein-triglyceride production despite reducing plasma triglyceride levels in APOE*3-Leiden.CETP mice.* *J. Biol. Chem.* 285, 25168-25175 (2010).
220. Berthiaume, J. and Wallace, K. B. *Perfluorooctanoate, perflourooctanesulfonate, and N-ethyl perfluorooctanesulfonamido ethanol; peroxisome proliferation and mitochondrial biogenesis.* *Toxicol. Lett.* 129, 23-32 (2002).
221. Seacat, A. M., Thomford, P. J., Hansen, K. J. et al. *Sub-chronic dietary toxicity of potassium perfluorooctanesulfonate in rats.* *Toxicology* 183, 117-131 (2003).
222. Shipley, J. M., Hurst, C. H., Tanaka, S. S. et al. *Trans-activation of PPAR α and induction of PPAR α target genes by perfluorooctane-based chemicals.* *Toxicol. Sci.* 80, 151-160 (2004).
223. Bjork, J. A., Lau, C., Chang, S. C. et al. *Perfluorooctane sulfonate-induced changes in fetal rat liver gene expression.* *Toxicology* 251, 8-20 (2008).
224. Ikeda, T., Aiba, K., Fukuda, K. et al. *The induction of peroxisome proliferation in rat liver by perfluorinated fatty acids, metabolically inert derivatives of fatty acids.* *J. Biochem.* 98, 475-482 (1985).
225. Chateau-Degat, M. L., Pereg, D., Dallaire, R. et al. *Effects of perfluorooctanesulfonate exposure on plasma lipid levels in the Inuit population of Nunavik (Northern Quebec).* *Environ. Res.* (2010).
226. Tilton, S. C., Orner, G. A., Benninghoff, A. D. et al. *Genomic profiling reveals an alternate mechanism for hepatic tumor promotion by perfluorooctanoic acid in rainbow trout.* *Environ. Health Perspect.* 116, 1047-1055 (2008).
227. Alberti, K. G., Zimmet, P., and Shaw, J. *International Diabetes Federation: a consensus on Type 2 diabetes prevention.* *Diabet. Med.* 24, 451-463 (2007).
228. Lonardo, A., Lombardini, S., Scaglioni, F. et al. *Hepatic steatosis and insulin resistance: does etiology make a difference?* *J. Hepatol.* 44, 190-196 (2006).
229. Heijboer, A. C., Donga, E., Voshol, P. J. et al. *Sixteen hours of fasting differentially affects hepatic and muscle insulin sensitivity in mice.* *J. Lipid Res.* 46, 582-588 (2005).
230. Heijboer, A. C., Voshol, P. J., Donga, E. et al. *High fat diet induced hepatic insulin resistance is not related to changes in hypothalamic mRNA expression of NPY, AgRP, POMC and CART in mice.* *Peptides* 26, 2554-2558 (2005).
231. Zambon, A., Hashimoto, S. I., and Brunzell, J. D. *Analysis of techniques to obtain plasma for measurement of levels of free fatty acids.* *J. Lipid Res.* 34, 1021-1028 (1993).
232. 't Hoen, P. A., de Kort, F., van Ommen, G. J. et al. *Fluorescent labelling of cRNA for microarray applications.* *Nucleic Acids Res.* 31, e20- (2003).
233. Smyth, G. K. *Linear models and empirical bayes methods for assessing differential expression in microarray experiments.* *Stat. Appl. Genet. Mol. Biol.* 3, Article3- (2004).
234. Zhang, B., Kirov, S., and Snoddy, J. *WebGestalt: an integrated system for exploring gene sets in various biological contexts.* *Nucleic Acids Res.* 33, W741-W748 (2005).

235. Newgard, C. B., Lu, D., Jensen, M. V. et al. *Stimulus/secretion coupling factors in glucose-stimulated insulin secretion: insights gained from a multidisciplinary approach.* Diabetes 51 Suppl 3, S389-S393 (2002).
236. Brown, M. S. and Goldstein, J. L. *The SREBP pathway: regulation of cholesterol metabolism by proteolysis of a membrane-bound transcription factor.* Cell 89, 331-340 (1997).
237. Eberle, D., Hegarty, B., Bossard, P. et al. *SREBP transcription factors: master regulators of lipid homeostasis.* Biochimie 86, 839-848 (2004).
238. Goldstein, J. L. and Brown, M. S. *Regulation of the mevalonate pathway.* Nature 343, 425-430 (1990).
239. Tomkins, G. M. and Chaikoff, I. L. *Cholesterol synthesis by liver. I. Influence of fasting and of diet.* J. Biol. Chem. 196, 569-573 (1952).
240. Chawla, A., Repa, J. J., Evans, R. M. et al. *Nuclear receptors and lipid physiology: opening the X-files.* Science 294, 1866-1870 (2001).
241. Konno, Y., Negishi, M., and Kodama, S. *The roles of nuclear receptors CAR and PXR in hepatic energy metabolism.* Drug Metab Pharmacokinet. 23, 8-13 (2008).
242. Yu, S., Matsusue, K., Kashireddy, P. et al. *Adipocyte-specific gene expression and adipogenic steatosis in the mouse liver due to peroxisome proliferator-activated receptor gamma1 (PPARgamma1) overexpression.* J. Biol. Chem. 278, 498-505 (2003).
243. Heikkinen, S., Auwerx, J., and Argmann, C. A. *PPARgamma in human and mouse physiology.* Biochim. Biophys. Acta 1771, 999-1013 (2007).
244. Tanaka, T., Masuzaki, H., Ebihara, K. et al. *Transgenic expression of mutant peroxisome proliferator-activated receptor gamma in liver precipitates fasting-induced steatosis but protects against high-fat diet-induced steatosis in mice.* Metabolism 54, 1490-1498 (2005).
245. Gross, D. N., van den Heuvel, A. P., and Birnbaum, M. J. *The role of FoxO in the regulation of metabolism.* Oncogene 27, 2320-2336 (2008).
246. Shimomura, I., Bashmakov, Y., and Horton, J. D. *Increased levels of nuclear SREBP-1c associated with fatty livers in two mouse models of diabetes mellitus.* J. Biol. Chem. 274, 30028-30032 (1999).
247. Ide, T., Shimano, H., Yahagi, N. et al. *SREBPs suppress IRS-2-mediated insulin signalling in the liver.* Nat. Cell Biol. 6, 351-357 (2004).
248. Ntambi, J. M., Miyazaki, M., Stoehr, J. P. et al. *Loss of stearoyl-CoA desaturase-1 function protects mice against adiposity.* Proc. Natl. Acad. Sci. U. S. A 99, 11482-11486 (2002).
249. Sampath, H., Miyazaki, M., Dobrzyn, A. et al. *Stearoyl-CoA desaturase-1 mediates the pro-lipogenic effects of dietary saturated fat.* J. Biol. Chem. 282, 2483-2493 (2007).
250. Inazu, A., Nakajima, K., Nakano, T. et al. *Decreased post-prandial triglyceride response and diminished remnant lipoprotein formation in cholesteryl ester transfer protein (CETP) deficiency.* Atherosclerosis 196, 953-957 (2008).

251. Kastelein, J. J., van Leuven, S. I., Burgess, L. et al. *Effect of torcetrapib on carotid atherosclerosis in familial hypercholesterolemia.* N. Engl. J. Med. 356, 1620-1630 (2007).
252. Nicholls, S. J., Tuzcu, E. M., Brennan, D. M. et al. *Cholesteryl ester transfer protein inhibition, high-density lipoprotein raising, and progression of coronary atherosclerosis: insights from ILLUSTRATE (Investigation of Lipid Level Management Using Coronary Ultrasound to Assess Reduction of Atherosclerosis by CETP Inhibition and HDL Elevation).* Circulation 118, 2506-2514 (2008).
253. Krishna, R., Anderson, M. S., Bergman, A. J. et al. *Effect of the cholesteryl ester transfer protein inhibitor, anacetrapib, on lipoproteins in patients with dyslipidaemia and on 24-h ambulatory blood pressure in healthy individuals: two double-blind, randomised placebo-controlled phase I studies.* Lancet 370, 1907-1914 (2007).
254. Arai, T., Yamashita, S., Hirano, K. et al. *Increased plasma cholesteryl ester transfer protein in obese subjects. A possible mechanism for the reduction of serum HDL cholesterol levels in obesity.* Arterioscler. Thromb. 14, 1129-1136 (1994).
255. Dullaart, R. P., Sluiter, W. J., Dikkeschei, L. D. et al. *Effect of adiposity on plasma lipid transfer protein activities: a possible link between insulin resistance and high density lipoprotein metabolism.* Eur. J. Clin. Invest 24, 188-194 (1994).
256. Magkos, F., Mohammed, B. S., and Mittendorfer, B. *Plasma lipid transfer enzymes in non-diabetic lean and obese men and women.* Lipids 44, 459-464 (2009).
257. Ebenbichler, C. F., Laimer, M., Kaser, S. et al. *Relationship between cholesteryl ester transfer protein and atherogenic lipoprotein profile in morbidly obese women.* Arterioscler. Thromb. Vasc. Biol. 22, 1465-1469 (2002).
258. Laimer, M. W., Engl, J., Tschaner, A. et al. *Effects of weight loss on lipid transfer proteins in morbidly obese women.* Lipids 44, 1125-1130 (2009).
259. Ritsch, A. and Patsch, J. R. *Cholesteryl ester transfer protein: gathering momentum as a genetic marker and as drug target.* Curr. Opin. Lipidol. 14, 173-179 (2003).
260. Boekholdt, S. M. and Thompson, J. F. *Natural genetic variation as a tool in understanding the role of CETP in lipid levels and disease.* J. Lipid Res. 44, 1080-1093 (2003).
261. Borggreve, S. E., Hillege, H. L., Wolffenbuttel, B. H. et al. *The effect of cholesteryl ester transfer protein -629C->A promoter polymorphism on high-density lipoprotein cholesterol is dependent on serum triglycerides.* J. Clin. Endocrinol. Metab 90, 4198-4204 (2005).
262. Yki-Jarvinen, H. *Thiazolidinediones and the liver in humans.* Curr. Opin. Lipidol. 20, 477-483 (2009).
263. Fabbrini, E., Mohammed, B. S., Korenblat, K. M. et al. *Effect of fenofibrate and niacin on intrahepatic triglyceride content, very low-density lipoprotein kinetics, and insulin action in obese subjects with nonalcoholic fatty liver disease.* J. Clin. Endocrinol. Metab 95, 2727-2735 (2010).

264. Jeong, S. and Yoon, M. *Fenofibrate inhibits adipocyte hypertrophy and insulin resistance by activating adipose PPAR α in high fat diet-induced obese mice.* Exp. Mol. Med. 41, 397-405 (2009).
265. Guerre-Millo, M., Gervois, P., Raspe, E. et al. *Peroxisome proliferator-activated receptor α activators improve insulin sensitivity and reduce adiposity.* J. Biol. Chem. 275, 16638-16642 (2000).
266. Duan, S. Z., Usher, M. G., and Mortensen, R. M. *PPARs: the vasculature, inflammation and hypertension.* Curr. Opin. Nephrol. Hypertens. 18, 128-133 (2009).
267. Babaev, V. R., Ishiguro, H., Ding, L. et al. *Macrophage expression of peroxisome proliferator-activated receptor- α reduces atherosclerosis in low-density lipoprotein receptor-deficient mice.* Circulation 116, 1404-1412 (2007).
268. Marx, N., Kehrle, B., Kohlhammer, K. et al. *PPAR activators as antiinflammatory mediators in human T lymphocytes: implications for atherosclerosis and transplantation-associated arteriosclerosis.* Circ. Res. 90, 703-710 (2002).
269. Madej, A., Okopien, B., Kowalski, J. et al. *Effects of fenofibrate on plasma cytokine concentrations in patients with atherosclerosis and hyperlipoproteinemia IIb.* Int. J. Clin. Pharmacol. Ther. 36, 345-349 (1998).
270. Ip, E., Farrell, G. C., Robertson, G. et al. *Central role of PPAR α -dependent hepatic lipid turnover in dietary steatohepatitis in mice.* Hepatology 38, 123-132 (2003).
- 158 271. Fernandez-Miranda, C., Perez-Carreras, M., Colina, F. et al. *A pilot trial of fenofibrate for the treatment of non-alcoholic fatty liver disease.* Dig. Liver Dis. 40, 200-205 (2008).
272. Zhou, C., King, N., Chen, K. Y. et al. *Activation of pregnane X receptor induces hypercholesterolemia in wild-type and accelerates atherosclerosis in apolipoprotein E deficient mice.* J. Lipid Res. (2009).
273. Gibson, J. C., Lee, W. H., and Piccolo, J. R. *The ansamycins: hypolipidemic agents stimulating cholesterol removal by nonclassical mechanisms.* J. Lipid Res. 35, 1524-1534 (1994).
274. Gibson, J. C., Lee, W. H., and Stephan, Z. F. *The ansamycins: a novel class of hypolipidemic agents with a high affinity for lipoproteins.* Atherosclerosis 112, 47-57 (1995).
275. Grun, F. and Blumberg, B. *Perturbed nuclear receptor signaling by environmental obesogens as emerging factors in the obesity crisis.* Rev. Endocr. Metab Disord. 8, 161-171 (2007).
276. Grun, F. and Blumberg, B. *Endocrine disrupters as obesogens.* Mol. Cell Endocrinol. 304, 19-29 (2009).
277. Miao, J., Fang, S., Bae, Y. et al. *Functional inhibitory cross-talk between constitutive androstane receptor and hepatic nuclear factor-4 in hepatic lipid/glucose metabolism is mediated by competition for binding to the DR1 motif and to the common coactivators, GRIP-1 and PGC-1 α .* J. Biol. Chem. 281, 14537-14546 (2006).

278. Guo, D., Sarkar, J., Suino-Powell, K. et al. *Induction of nuclear translocation of constitutive androstane receptor by peroxisome proliferator-activated receptor alpha synthetic ligands in mouse liver.* J. Biol. Chem. 282, 36766-36776 (2007).
279. Wieneke, N., Hirsch-Ernst, K. I., Kuna, M. et al. *PPARalpha-dependent induction of the energy homeostasis-regulating nuclear receptor NR1i3 (CAR) in rat hepatocytes: potential role in starvation adaptation.* FEBS Lett. 581, 5617-5626 (2007).
280. Randle, P. J., Garland, P. B., Hales, C. N. et al. *The glucose fatty-acid cycle. Its role in insulin sensitivity and the metabolic disturbances of diabetes mellitus.* Lancet 1, 785-789 (1963).
281. Randle, P. J., Garland, P. B., Newsholme, E. A. et al. *The glucose fatty acid cycle in obesity and maturity onset diabetes mellitus.* Ann. N. Y. Acad. Sci. 131, 324-333 (1965).
282. Randle, P. J. *Regulatory interactions between lipids and carbohydrates: the glucose fatty acid cycle after 35 years.* Diabetes Metab Rev. 14, 263-283 (1998).
283. Peterson, L. R., Herrero, P., Schechtman, K. B. et al. *Effect of obesity and insulin resistance on myocardial substrate metabolism and efficiency in young women.* Circulation 109, 2191-2196 (2004).
284. Kelley, D. E. and Mandarino, L. J. *Fuel selection in human skeletal muscle in insulin resistance: a reexamination.* Diabetes 49, 677-683 (2000).
285. Kelley, D. E., Goodpaster, B. H., and Storlien, L. *Muscle triglyceride and insulin resistance.* Annu. Rev. Nutr. 22, 325-346 (2002).
286. Sugden, M. C., Zariwala, M. G., and Holness, M. J. *PPARs and the orchestration of metabolic fuel selection.* Pharmacol. Res. 60, 141-150 (2009).
287. Tremblay, F., Gagnon, A., Veilleux, A. et al. *Activation of the mammalian target of rapamycin pathway acutely inhibits insulin signaling to Akt and glucose transport in 3T3-L1 and human adipocytes.* Endocrinology 146, 1328-1337 (2005).
288. Kamagate, A., Qu, S., Perdomo, G. et al. *FoxO1 mediates insulin-dependent regulation of hepatic VLDL production in mice.* J. Clin. Invest 118, 2347-2364 (2008).
289. Accili, D. and Arden, K. C. *FoxOs at the crossroads of cellular metabolism, differentiation, and transformation.* Cell 117, 421-426 (2004).
290. Barthel, A., Schmoll, D., and Unterman, T. G. *FoxO proteins in insulin action and metabolism.* Trends Endocrinol. Metab 16, 183-189 (2005).
291. Kamagate, A. and Dong, H. H. *FoxO1 integrates insulin signaling to VLDL production.* Cell Cycle 7, 3162-3170 (2008).
292. Maury, E. and Brichard, S. M. *Adipokine dysregulation, adipose tissue inflammation and metabolic syndrome.* Mol. Cell Endocrinol. 314, 1-16 (2010).
293. Hotamisligil, G. S. *Inflammatory pathways and insulin action.* Int. J. Obes. Relat Metab Disord. 27 Suppl 3, S53-S55 (2003).
294. Moller, D. E. *Potential role of TNF-alpha in the pathogenesis of insulin resistance and type 2 diabetes.* Trends Endocrinol. Metab 11, 212-217 (2000).

295. Hundal, R. S., Petersen, K. F., Mayerson, A. B. *et al.* Mechanism by which high-dose aspirin improves glucose metabolism in type 2 diabetes. *J. Clin. Invest* 109, 1321-1326 (2002).
296. Yuan, M., Konstantopoulos, N., Lee, J. *et al.* Reversal of obesity- and diet-induced insulin resistance with salicylates or targeted disruption of Ikk β . *Science* 293, 1673-1677 (2001).
297. Jarvis, D., Chinn, S., Potts, J. *et al.* Association of body mass index with respiratory symptoms and atopy: results from the European Community Respiratory Health Survey. *Clin. Exp. Allergy* 32, 831-837 (2002).
298. Schachter, L. M., Peat, J. K., and Salome, C. M. Asthma and atopy in overweight children. *Thorax* 58, 1031-1035 (2003).
299. Tantisira, K. G., Litonjua, A. A., Weiss, S. T. *et al.* Association of body mass with pulmonary function in the Childhood Asthma Management Program (CAMP). *Thorax* 58, 1036-1041 (2003).
300. Xu, B., Jarvelin, M. R., and Pekkanen, J. Body build and atopy. *J. Allergy Clin. Immunol.* 105, 393-394 (2000).
301. Visness, C. M., London, S. J., Daniels, J. L. *et al.* Association of obesity with IgE levels and allergy symptoms in children and adolescents: results from the National Health and Nutrition Examination Survey 2005-2006. *J. Allergy Clin. Immunol.* 123, 1163-9, 1169 (2009).

Summary

Obesity is characterized by excessive fat storage and is associated with various diseases like cardiovascular disease (CVD) and type 2 diabetes (DM2), thereby being a serious problem of public health. Excessive energy intake is an important cause of obesity since excess energy is primarily stored as fat. The stored fat is mobilized again during fasting in the form of fatty acids (FA). These FA are re-esterified in the liver in triglycerides (TG) that are secreted in VLDL particles to deliver FA to peripheral tissues where they can be used for energy.

One of the current views of the cause of diseases related to obesity is the (mis) handling of TG derived FA. Therefore it is important to understand pathways involved in the uptake, distribution, oxidation and storage of TG. In this thesis we have evaluated the effect of different interventions on VLDL-TG metabolism to gain a better understanding of its complex regulation. For these studies we used APOE^{*3}-Leiden (E3L) and E3L.CETP transgenic mice that have a human-like lipoprotein metabolism and respond to lipid-modifying drugs in a ways similar to humans.

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The first part of this thesis focuses on the effect of cholesteryl ester transfer protein (CETP) on VLDL-TG metabolism. CETP transfers cholesteryl esters and TG between lipoproteins and has been shown to have a major impact on cholesterol metabolism. Previous studies in E3L mice have shown that expression of CETP reduces HDL-C, increases LDL-C and thereby increases the risk to develop atherosclerosis. Whether CETP also has impact on TG metabolism was explored in **chapter 2**. Expression of CETP hardly affected VLDL-TG metabolism and did not influence high fat diet induced obesity in E3L mice. We concluded that CETP inhibitors are not likely to have adverse health effects related to TG metabolism.

The second part of this thesis addressed the effect of different pharmaceutical interventions on the regulation of VLDL-TG metabolism by nuclear receptors. Intermediates of lipid metabolism and hormones activate nuclear receptors after which they are transported to the nucleus where they regulate amongst other the expression of genes involved in energy homeostasis. These nuclear receptors include peroxisome proliferators activated receptors (PPARs) and the xenobiotic receptors pregnane X receptor (PXR) and constitutive androstane receptor (CAR).

In **chapter 3** we investigated the effect of the pharmacologic PPAR α activator

fenofibrate on VLDL-TG metabolism. To this end, E3L.CETP mice were fed a western-type diet without or with fenofibrate. Treatment with fenofibrate lowered total plasma TG levels by increasing lipolysis of VLDL-TG. Unexpectedly, fenofibrate increased VLDL-TG production which is partly explained by increased FA turnover.

The effect of rifampicin on VLDL metabolism was studied in **chapter 4**. E3L.CETP mice were treated with the PXR agonist rifampicin, an antibiotic prescribed for the treatment of tuberculosis. Treatment with this drug is associated with hepatic steatosis and dyslipidemia. E3L.CETP mice were fed a western-type diet for 3 weeks followed by an additional 3 weeks diet without or with increasing doses of rifampicin. The highest dose of rifampicin used (0.10%) showed a decrease of both HDL and VLDL cholesterol levels whereas total TG levels were unaltered. This decrease in cholesterol was mainly explained by lowering of both HDL and VLDL particle production by the liver. However, the VLDL particles secreted were enriched in TG explaining why total TG levels were not affected by rifampicin.

In addition to drugs affecting nuclear receptors and lipid metabolism, chemical pollutants might also act as agonists for nuclear receptors. In **chapter 5** we describe the hypolipidemic effects of perfluoroalkyl sulfonates (PFAS), a group of chemicals used for stain repellence and coatings. These compounds are highly resistant to degradation and bioaccumulate. PFAS activate PPAR α as well as the xenobiotic receptors PXR and CAR, altogether reducing HDL and VLDL production by the liver as well as increasing VLDL-TG clearance by increasing lipolysis.

In **chapter 6** we examined the differences in hepatic gene expression in response to fasting or a high fat diet, both known to induce hepatic steatosis. However, only after high fat diet feeding this steatosis is associated with hepatic insulin resistance. To gain insight in the transcriptional processes leading to steatosis associated insulin resistance, C57Bl6/J mice were fed standard chow diet or a high fat diet for 2 weeks. After 2 weeks half of the mice fed chow were fasted for 16 hours whereas the other part of the chow fed mice and the high fat diet fed mice were fasted for 4 hours. Strikingly, fasting affected far more genes compared to control than feeding a high fat diet. Furthermore, hardly any overlap in gene expression profile was seen between fasting and a high

fat diet, suggesting completely different gene programmes are activated. High fat diet feeding was especially associated with activation of PPAR α whereas fasting activated xenobiotic receptors PXR and CAR.

Chapter 7 discusses the observation that nuclear receptors have a major impact on lipoprotein metabolism and play a key role in the pathology associated with obesity. Nuclear receptors are used as targets for the development of drugs to treat the metabolic syndrome. However, although we only focussed on the effect of nuclear receptors on lipoprotein metabolism in this thesis, inflammation is also regulated by these nuclear receptors. Development of new drugs that target nuclear receptors focuses on organ-specificity as well as gene-specificity. However, a focus on dyslipidemia only might neglect the anti-inflammatory effects of novel potential drugs that could provide added benefit.

Nederlandse samenvatting voor niet-ingewijden

Ongezond overgewicht

Zwaarlijvigheid (**obesitas**) is het overmatig opslaan van vet in ons lichaam en leidt tot ziektes als hart- en vaatziekten en suikerziekte en is daarom een groot probleem in de gezondheidszorg. Of iemand obees is wordt bepaald aan de hand van je Body Mass Index (**BMI**): het lichaamsgewicht in kilogram, gedeeld door de lichaamslengte in meters in het kwadraat (kg/m^2). Een BMI tussen 18,5 en 25 wordt beschouwd als ideaal voor een gezond individu; een BMI van meer dan 25 wordt beschouwd als te zwaar (overgewicht); een BMI van meer dan 30 wordt beschouwd als obesitas. Overmatige inname van calorieën is één van de belangrijkste oorzaken van obesitas omdat de extra calorieën voornamelijk als vet worden opgeslagen. Wanneer we vasten wordt dit vet weer vrijgemaakt in de vorm van **vetzuren** zodat we het vet kunnen gebruiken als energie bron. De meeste van deze vetzuren gaan direct naar de lever waar ze worden omgezet in **triglyceriden** (TG). Deze TG bestaan uit een glycerol keten met drie vetzuren. TG worden vanuit de lever vervolgens weer vrijgelaten in de bloedbaan.

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Transport van vet in ons lichaam

Omdat vet slecht oplost in water en bloed, moet vet 'oplosbaar' verpakt worden om getransporteerd te worden in ons lichaam. De verpakking van vet wordt ook wel een **lipoproteïne** genoemd. Afhankelijk van hun samenstelling hebben deze lipoproteïnen een ander dichtheid en worden ze als volgt ingedeeld: zeer lage dichtheid lipoproteïnen (**VLDL**) met voornamelijk TG, lage dichtheid lipoproteïnen (**LDL**) met TG en cholesterol en hoge dichtheid lipoproteïnen (**HDL**) met voornamelijk cholesterol. Het hebben van veel LDL is een risicofactor voor het ontstaan van aderverkalking terwijl HDL er juist voor beschermd. LDL is in de volksmond ook wel bekend als slecht cholesterol en HDL als goed cholesterol.

TG uit de lever komen in de bloedbaan terecht in VLDL om bij andere weefsels vetzuren af te leveren die als energie bron gebruikt kunnen worden. Omdat TG zeer groot is moeten deze eerst weer omgezet worden in losse vetzuren voordat ze door de weefsels opgenomen kunnen worden. Dit proces heet **lipolyse**. De reden dat vetzuren eerst via de lever in TG worden omgezet heeft te maken met het feit dat vetzuren zelf toxicisch zijn. In weefsels worden ze daarom zo snel mogelijk óf verbrand óf weer omgezet in TG zodat ze geen schade kunnen aanrichten.

Verstoring TG transport in obesitas

Er wordt momenteel heel veel onderzoek gedaan naar de reden waarom mensen met obesitas een hoog risico lopen om hart- en vaatziekten en suikerziekte te ontwikkelen. Men denkt dat één van de oorzaken hiervoor is dat het lichaam verkeerd omgaat met TG en de vetzuren hiervan. Daarom is het belangrijk dat we meer inzicht krijgen in de stofwisseling processen betrokken bij de opname, het transport, de verbranding en de opslag van TG en vetzuren (**TG metabolisme**). In dit proefschrift onderzoeken we het effect van verschillende interventies op het metabolisme van VLDL-TG om zo beter inzicht te krijgen in de regulatie hiervan.

In het eerste deel van dit proefschrift hebben we gekeken naar het effect van cholesteryl ester transfer proteïne (**CETP**) op het VLDL-TG metabolisme. CETP is betrokken bij het verplaatsen van cholesteryl esters en TG tussen lipoproteïnen en we hebben eerder al laten zien dat CETP een grote invloed heeft op het cholesterol metabolisme. De werking van CETP leidt tot verlaagde niveaus van HDL cholesterol (goed cholesterol) en een toename van LDL cholesterol (slecht cholesterol) waardoor de kans op aderverkalking toeneemt. Of CETP ook effect heeft op TG is onderzocht in **hoofdstuk 2**. De werking van CETP heeft nauwelijks effect op het VLDL-TG metabolisme en heeft geen effect op de ontwikkeling van obesitas door het aangeboden dieet. Omdat CETP cholesterol verplaatst van HDL naar LDL en het goede cholesterol hierdoor afneemt, worden er medicijnen ontwikkeld om CETP te remmen en zo het goede cholesterol te verhogen. Het is belangrijk dat het remmen van CETP geen negatieve bijwerkingen heeft omdat CETP ook TG verplaatst tussen lipoproteïnen. We concluderen dan ook dat CETP remmers waarschijnlijk geen negatieve bijwerkingen zullen hebben met betrekking tot het TG metabolisme.

Regulatie van TG metabolisme

In het tweede deel van dit proefschrift hebben we gekeken naar de effecten van verschillende farmacologische interventies op de regulatie van het VLDL-TG metabolisme. **Nucleaire receptoren** spelen hierbij een grote rol. Dit zijn eiwitten in de cel die zijn betrokken bij de communicatie in de cel. Nucleaire receptoren herkennen specifieke moleculen in de cel, bijvoorbeeld vetzuren, en verplaatsen zich vervolgens naar de kern van de cel waar ze de **werking (expressie) van genen** beïnvloeden. Genen zijn onderdeel van ons DNA en bevatten de receptuur om 'machines' te maken. Deze machines zijn de werktuigen in onze cellen en hebben allemaal een eigen functie. Wanneer

de expressie van een gen toeneemt wordt er meer van die machine gemaakt. Op deze manier worden processen in onze cellen, en dus in ons lichaam, beïnvloed. Andere moleculen die herkend kunnen worden door nucleaire receptoren zijn hormonen en xenobiotica. Xenobiotica zijn moleculen die van nature niet in het lichaam voorkomen, bijvoorbeeld medicijnen die we slikken. Tot de nucleaire receptoren behoren de peroxisoom proliferator geactiveerde receptoren (**PPAR**), en de xenobioticum receptoren pregnaan X receptor (**PXR**) en constitutieve androstaan receptor (**CAR**).

In **hoofdstuk 3** hebben we het effect van de farmacologische PPAR α activator fenofibraat op het VLDL-TG metabolisme onderzocht. Fenofibraat behoort tot de groep van de fibraten en hebben als belangrijkste functie het verlaging van TG in het bloed. Er werd altijd vanuit gegaan dat deze verlaging wordt veroorzaakt door minder productie van VLDL-TG én meer opname van VLDL-TG. Onze studie laat zien dat de verlaging van TG door fenofibraat geheel wordt verklaard door meer opname van VLDL-TG. Onverwacht veroorzaakt fenofibraat zelfs een verhoging van de productie van VLDL-TG door de lever. Deze toename wordt deels veroorzaakt door een toename in de omvorming en verplaatsing van vetzuren, oftewel meer **vetzuur omzet**. Verhoogde omzetting van vetzuren in het lichaam wordt als één van de oorzaken beschouwd voor het ontstaan van ziekten gerelateerd aan obesitas. Het is dus belangrijk dat we niet alleen weten wat medicijnen op totale niveau's doen van TG en cholesterol maar ook hoe medicijnen dit doen. Waarschijnlijk is de verhoogde omzetting van vetzuren met fenofibraat niet schadelijk omdat fenofibraat ook ontsteking remt. Ontsteking is ook belangrijk bij het ontstaan van hart- en vaatziekten en suikerziekte in mensen met obesitas.

Het effect van rifampicine op het VLDL metabolisme is onderzocht in **hoofdstuk 4**. Rifampicine is een antibioticum dat wordt voorgeschreven voor de behandeling van tuberculose. Maar rifampicine activeert ook PXR. Bij behandeling met dit medicijn zijn bijwerkingen als vervetting van de lever en verstoring van TG en cholesterol in het bloed bekend. In onze studie laten we zien dat rifampicine inderdaad PXR activeert en leidt tot een verlaging zowel HDL als VLDL cholesterol terwijl de TG niveau's onveranderd blijven. De afname van het cholesterol wordt veroorzaakt door minder productie van voornamelijk VLDL door de lever. Omdat de VLDL deeltjes uitgescheiden door de lever meer TG bevatten heeft rifampicine geen effect op het totale TG niveau.

Chemische verontreinigende stoffen hebben ook invloed op nucleaire receptoren. In **hoofdstuk 5** beschrijven we het effect van perfluoroakylnafonaten (PFAS) op het TG en cholesterol metabolisme. PFAS zijn chemische stoffen die worden gebruikt in anti-vlek middelen en in coatings. Nadeel van deze stoffen is dat ze biologisch slecht afbreekbaar zijn en daardoor ophopen in het milieu. We laten zien dat PFAS de nucleaire receptoren PPAR α , PXR en CAR activeren, wat leidt tot remming van zowel de HDL als VLDL productie door de lever. Daarnaast stimuleren PFAS de lipolyse van VLDL-TG. Bij elkaar verklaart dit waarom cholesterol en TG sterk verlagen in onze studie.

In **hoofdstuk 6** hebben we gekeken naar de verschillen in lever gen expressie in reactie op langdurig vasten of een hoog vet dieet. Zowel langdurig vasten als een hoog vet dieet leiden tot vervetting van de lever maar alleen bij een hoog vet dieet is er ook sprake van dat de lever niet langer reageert op insuline. Dit wordt ook wel insuline resistantie genoemd en kan leiden tot suikerziekte. We hebben deze studie uitgevoerd om meer inzicht te krijgen in de regulatie van het vet metabolisme bij zowel lever vervetting als insuline resistantie. Opvallend is dat vasten de werking van veel meer genen beïnvloed vergeleken met controle dieren dan een hoog vet dieet. Daarnaast was er ook nauwelijks overlap in de genen die tot werking kwamen vergeleken tussen vasten en hoog vet dieet. Dit suggereert dat er compleet andere gen expressie programma's betrokken zijn bij het ontstaan van een vette lever tussen vasten en een hoog vet dieet. Hoog vet dieet geïnduceerde vette levers waren geassocieerd met activatie van PPAR α terwijl na vasten vooral de xenobioticum receptoren PXR en CAR werden geactiveerd.

In dit proefschrift laten we zien dat nucleaire receptoren een grote invloed hebben op het VLDL-TG metabolisme en ook betrokken zijn bij het ontstaan van ziektes gerelateerd aan obesitas. Momenteel wordt er veel aandacht besteed aan het ontwikkelen van medicijnen die nucleaire receptoren beïnvloeden om zo ziekten in obese mensen te behandelen. Onze onderzoeken laten vooral zien wat het effect is van nucleaire receptoren op VLDL-TG maar ze hebben vaak ook invloed op ontsteking. Bij het ontstaan van ziektes bij obesitas zijn vaak vele verschillende processen betrokken waaronder het TG metabolisme en ontsteking. Door geïntegreerd onderzoek te doen naar deze verschillende processen kunnen we een beter beeld krijgen waarom mensen met obesitas ziek worden en goede medicijnen ontwikkelen.

List of publications

Bijland S, van den Berg SAA, Voshol PJ, van den Hoek AM, Princen HM, Havekes LM, Rensen PCN, Willems van Dijk K. CETP does not affect triglyceride production or clearance in APOE*3-Leiden mice.

Journal of Lipid Research 2010;51(1):97-102

van den Berg SAA, Guigas B, Bijland S, Ouwens M, Voshol PJ, Frants RR, Havekes LM, Romijn JA, Willems van Dijk K. High levels of dietary stearate promote adiposity and deteriorate hepatic insulin sensitivity.

Nutrition & Metabolism 2010;7:24

van den Berg SAA*, Nabben M*, Bijland S, Voshol PJ, van Klinken JB, Havekes LM, Romijn JA, Hoeks J, Hesselink MK, Schrauwen P, Willems van Dijk K. High levels of whole body energy expenditure are associated with skeletal muscle mitochondrial uncoupling in C57Bl/6 mice.

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Metabolism 2010 May 20

Bijland S, Pieterman EJ, Maas ACE, van der Hoorn JWA, van Erk MJ, van Klinken JB, Havekes LM, Willems van Dijk K, Princen HMG, Rensen PCN. Fenofibrate increases VLDL-triglyceride production despite reducing plasma triglyceride levels in APOE*3-Leiden.CETP mice.

The Journal of Biological Chemistry 2010;285(33):25168-75

de Wilde J, Smit E, Mohren R, Boekschoten MV, de Groot P, van den Berg SAA, Bijland S, Voshol PJ, Willems van Dijk K, de Wit NW, Bunschoten A, Schaart G, Hulshof MF, Mariman EC. An 8-week high-fat diet induces obesity and insulin resistance with small changes in the muscle transcriptome of C57BL/6J mice.

Journal of Nutrigenetics and Nutrigenomics. 2009;2(6):280-91

de Vogel-van den Bosch J*, van den Berg SAA*, Bijland S, Voshol PJ, Havekes LM, Romijn HA, Hoeks J, van Beurden D, Hesselink MKC, Schrauwen P, Willems van Dijk K. High fat diets rich in medium- versus long-chain fatty acids induce distinct patterns of tissue specific insulin resistance.
The Journal of Nutritional Biochemistry 2010

Bijland S*, Rensen PCN*, Pieterman EJ, Maas ACE, van der Hoorn JW, van Erk MJ, Havekes LM, Willems van Dijk K, Butenhoff JL, Princen HMG. Perfluoroalkyl sulfonates cause alkyl chain-dependent hepatic steatosis and combined hypolipidemia in ApoE^{*3}-Leiden.CETP mice.

Submitted.

Langeveld M*, van den Berg SAA*, Bijl N, Bijland S, van Roomen C, Houben-Weerts JH, Ottenhoff R, Houten SM, Willems van Dijk K, Romijn JA, Groen AK, Aerts JM, Voshol PJ. N-(5-adamantane-1-yl-methoxy-pentyl)-deoxynojirimycin (AMP-DNM) reduces body weight by decreasing energy intake and increasing fat oxidation in ob/ob mice.

Submitted

Bijland S*, de Haan W*, Smit JWA, Havekes LM, Princen HMG, Willems van Dijk K, Rensen PCN. Rifampicin decreases plasma cholesterol by impairing HDL and VLDL particle production in ApoE^{*3}-LEIDEN.CETP mice.

Manuscript in preparation.

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Curriculum Vitae

Silvia Bijland werd geboren op 13 juli 1980 te Zaanstad. Na het doorlopen van de Mavo en Havo behaalde ze haar VWO diploma in juni 2000 aan het Bertrand Russell College te Krommenie. In september dat jaar begon zij haar studie Biomedische Wetenschappen aan de Universiteit van Utrecht. Ze behaalde haar propedeutisch examen in 2001 en haar Bachelor diploma in 2003. Tijdens haar Master Developmental Biology and Biomedical Genetics heeft zij een drietal stages doorlopen. Tijdens haar hoofdvakstage heeft ze onderzoek verricht binnen de vakgroep Fysiologische Chemie van het Universitair Medisch Centrum Utrecht onder leiding van Drs. JA Riedl en Prof. Dr. JL Bos met als onderwerp ‘Small GTPase Rap1’. Haar tweede onderzoeksstage heeft ze verricht onder leiding van Drs. MJ Adjobo-Hermans en Prof. Dr. D. Gadella Jr. bij de afdeling Molecular Cytology van het Swammerdam Institute for Lifescience te Amsterdam met als onderwerp ‘Visualisation of dimerization C terminal domain PLC β '. Daarnaast heeft ze nog een onderzoeksstage verricht bij de afdeling Endocrinologie van het Leids Universitair Medisch Centrum Leiden onder leiding van Drs. JFP Berbée en Dr. PCN Rensen met als onderwerp ‘Role of apolipoprotein AIV in inflammation’. In 2006 behaalde zij haar Master diploma en aansluitend startte zij als promovenda met haar promotieonderzoek op de afdelingen Humane Genetica en Endocrinologie onder begeleiding van haar promotor Prof. Dr. Ir. LM Havekes en copromotores Dr. Ir. K. Willems van Dijk en Dr. PCN Rensen.

