



Universiteit  
Leiden  
The Netherlands

## **The role of homologous recombination in mitotic and meiotic double-strand break repair**

Vries, Femke Adriana Theodora de

### **Citation**

Vries, F. A. T. de. (2007, January 17). *The role of homologous recombination in mitotic and meiotic double-strand break repair*. Retrieved from <https://hdl.handle.net/1887/8784>

Version: Corrected Publisher's Version

License: [Licence agreement concerning inclusion of doctoral thesis in the Institutional Repository of the University of Leiden](#)

Downloaded from: <https://hdl.handle.net/1887/8784>

**Note:** To cite this publication please use the final published version (if applicable).



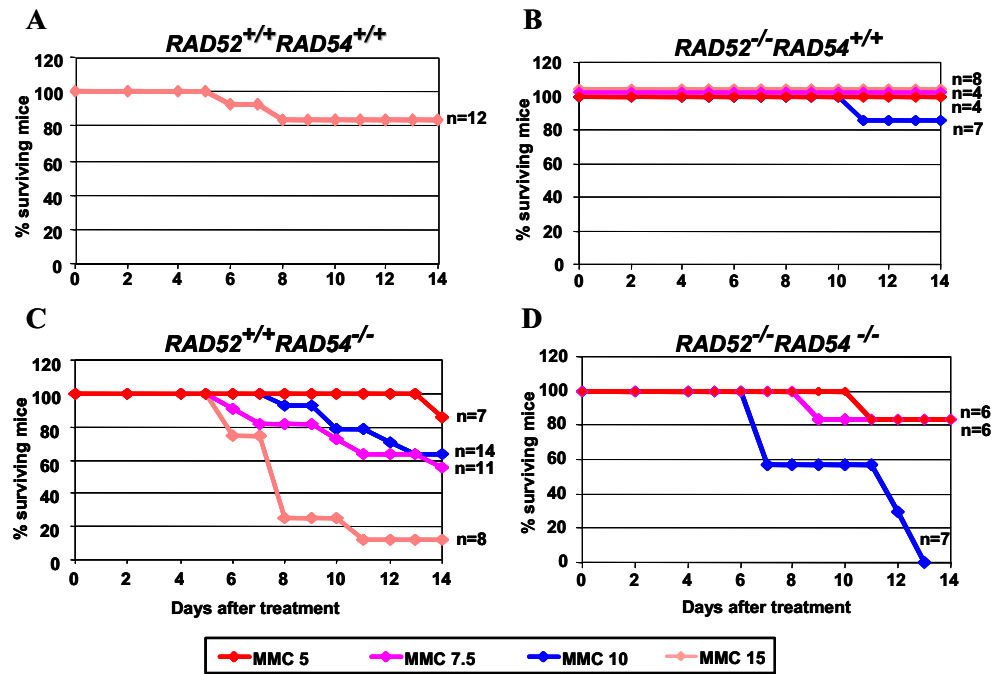
## Appendix

### Selected coloured figures





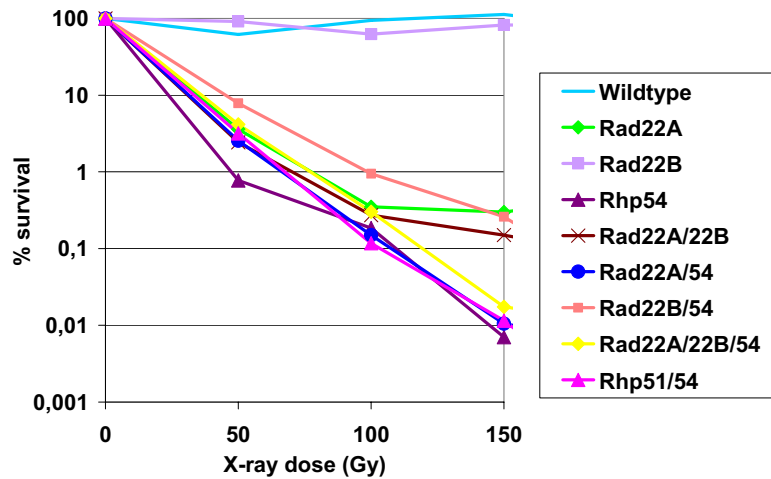
## Chapter 3 Figure 2



**Figure 2:** Survival of mice following mitomycin C treatment. Animals were injected intraperitoneally with 5, 7.5, 10 or 15 mg/kg bodyweight MMC. Total numbers of individually treated mice are indicated per dose. Control (A) *RAD52<sup>-/-</sup>* (B), *RAD54<sup>-/-</sup>* (C) and *RAD52<sup>-/-</sup> / RAD54<sup>-/-</sup>* (D) mice.

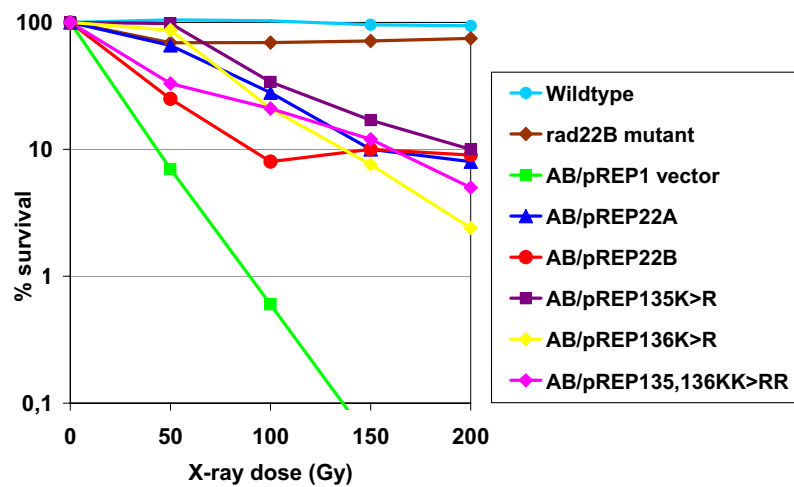
Coloured figures

### Chapter 3 Figure 4



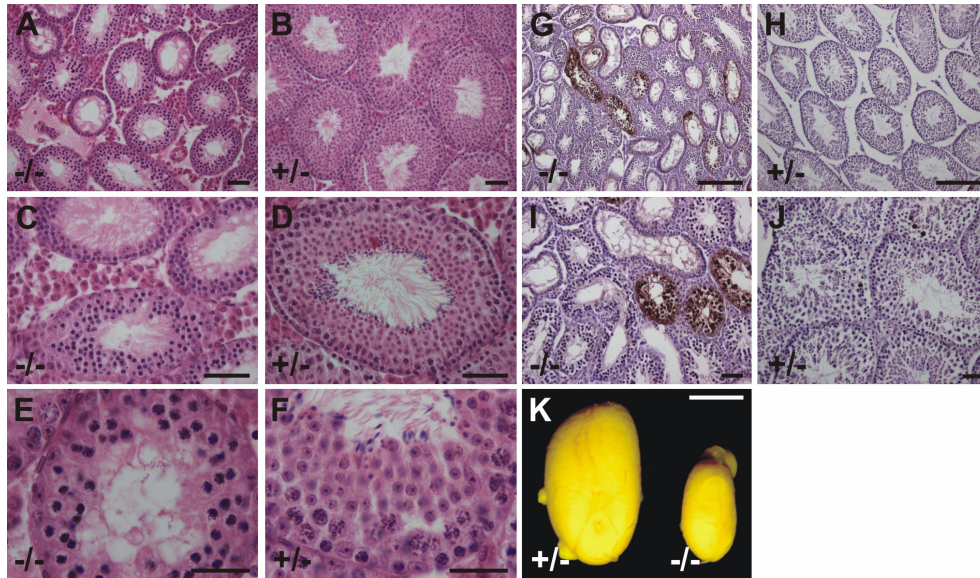
**Figure 4:** X-ray survival of wildtype and single, double and triple mutant *S. pombe* strains. Exponentially growing cells were harvested, irradiated and appropriate dilutions were plated in triplicate on YES media. After 3 days of incubation at 30°C the colonies were counted. Each survival experiment was repeated at least twice.

### Chapter 4 Figure 4



**Figure 4:** Survival of *S. pombe* strains after irradiation with X-rays. After irradiation of exponentially growing cells, appropriate dilutions were plated and colonies were counted after incubation of the plates for 3 days at 30°C. Each survival experiment was repeated at least twice. Strains used in this experiment: wildtype (Y4), *rad22B*<sup>-/-</sup> (*rad22B* mutant), and double mutant *rad22A*<sup>-/-</sup>*rad22B*<sup>-/-</sup> strains containing expression vectors without insert (AB/pREP-), with Rad22A insert (AB/pREP22A), with Rad22B insert (AB/pREP22B) and with Rad22A inserts in which the putative SUMO acceptor site has been mutated (AB/pREP135 K>R, AB/pREP136 K>R, AB/pREP135,136 KK>RR). For details see Materials and methods.

## Chapter 5 Figure 2

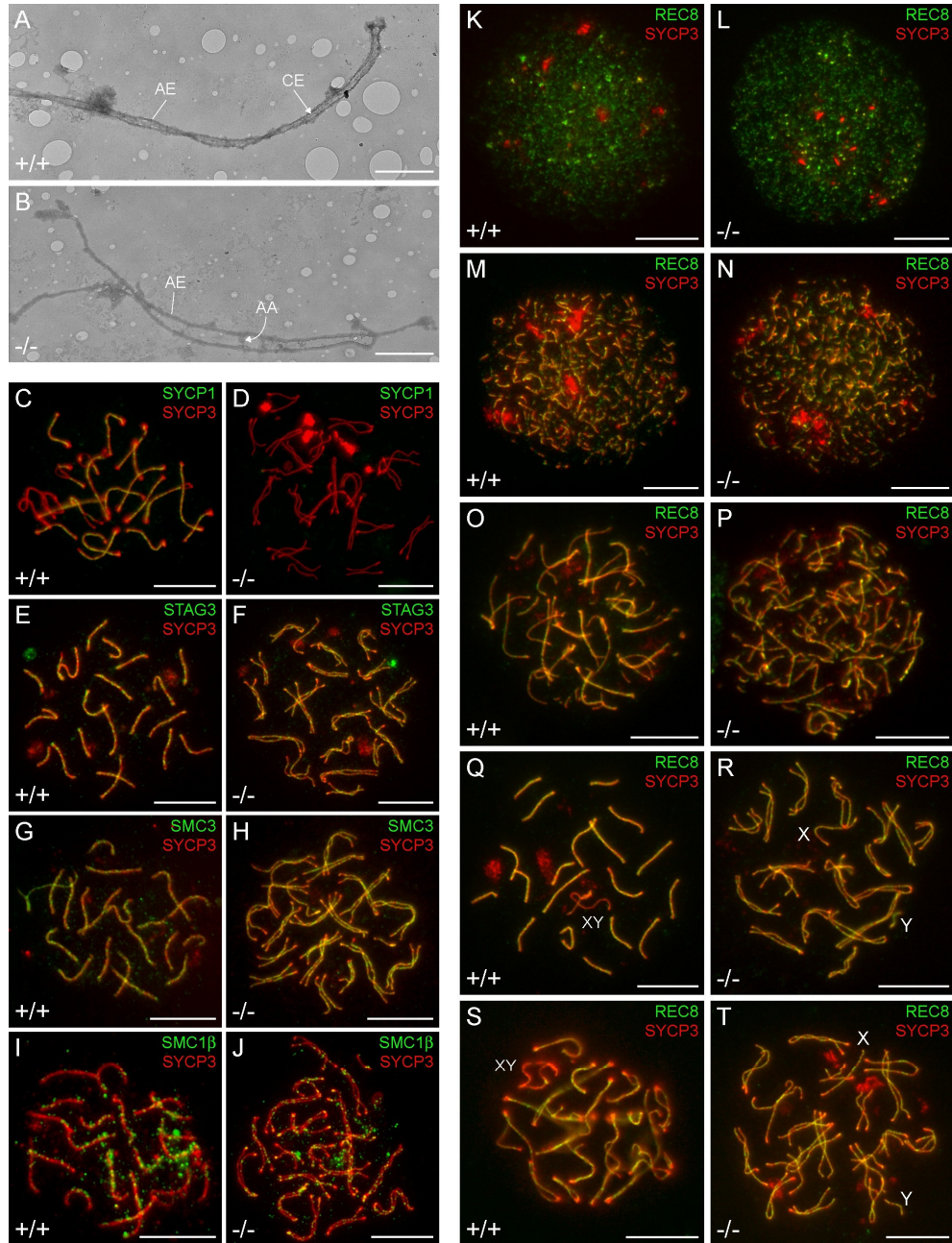


**Figure 2:** Morphology, histology and TUNEL analysis of testes from *Sycp1*<sup>-/-</sup> mice. The histological sections were stained with haematoxylin and eosin. **(A-F)** Testicular histology of adult *Sycp1*<sup>-/-</sup> (-/-, **A,C,E**) and *Sycp1*<sup>+/-</sup> (+/-, **B,D,F**) mice. Note the total absence of postmeiotic germ cells in *Sycp1*<sup>-/-</sup> sections. Pachytene nuclei are abundant, but show aberrant nuclear morphology. **(G-J)** TUNEL analysis of testis sections of *Sycp1*<sup>-/-</sup> (-/-, **G,I**) and *Sycp1*<sup>+/-</sup> (+/-, **H,J**) mice. Tubule sections with numerous TUNEL-positive nuclei occur only in *Sycp1*<sup>-/-</sup> mice. A few apoptotic nuclei are visible in tubule sections from *Sycp1*<sup>+/-</sup> mice. **(K)** Testes from *Sycp1*<sup>+/-</sup> (+/-) and *Sycp1*<sup>-/-</sup> (-/-) mice. Bars: **(A-D,I,J)** 50 µm; **(E-F)** 25 µm; **(G-H)** 100 µm; **(K)** 2 mm.

**Figure 3:** Assembly of AEs in *Sycp1*<sup>-/-</sup> mice.

**(A-B)** Electron micrographs of AEs and SCs from wildtype (+/+) and *Sycp1*<sup>-/-</sup> (-/-) male mice; **(A)** wildtype SC with closely apposed axial elements (AE) and a central element (CE); **(B)** homologously aligned axial elements (AE) from a *Sycp1*<sup>-/-</sup> spermatocyte, connected by axial associations (AA). **(C-J)** Components of AEs and SCs in wildtype (+/+) and *Sycp1*<sup>-/-</sup> (-/-) diplotene **(C-D)** or pachytene **(E-J)** spermatocytes; LE/AE protein SYCP3 and all analyzed cohesins are present in LEs/AEs of wildtype and mutant, whereas SYCP1 is not detectable in mutant spermatocytes. **(K-T)** formation of AEs/LEs, as shown by REC8/SYCP3 double labelling, in wildtype (+/+) and *Sycp1*<sup>-/-</sup> (-/-) spermatocytes; **(K,L)** early leptoneuma; **(M,N)** late leptoneuma; **(O,P)** zygonema; **(Q,R)** pachynema; **(S,T)** diplonema; note the XY bivalent (XY) in wildtype cells **(Q,S)**, and separate X and Y chromosomes in the *Sycp1*<sup>-/-</sup> cells **(R,T)**. Bars in **(A-B)** 1 µm; bars in **(C-T)** 10 µm.

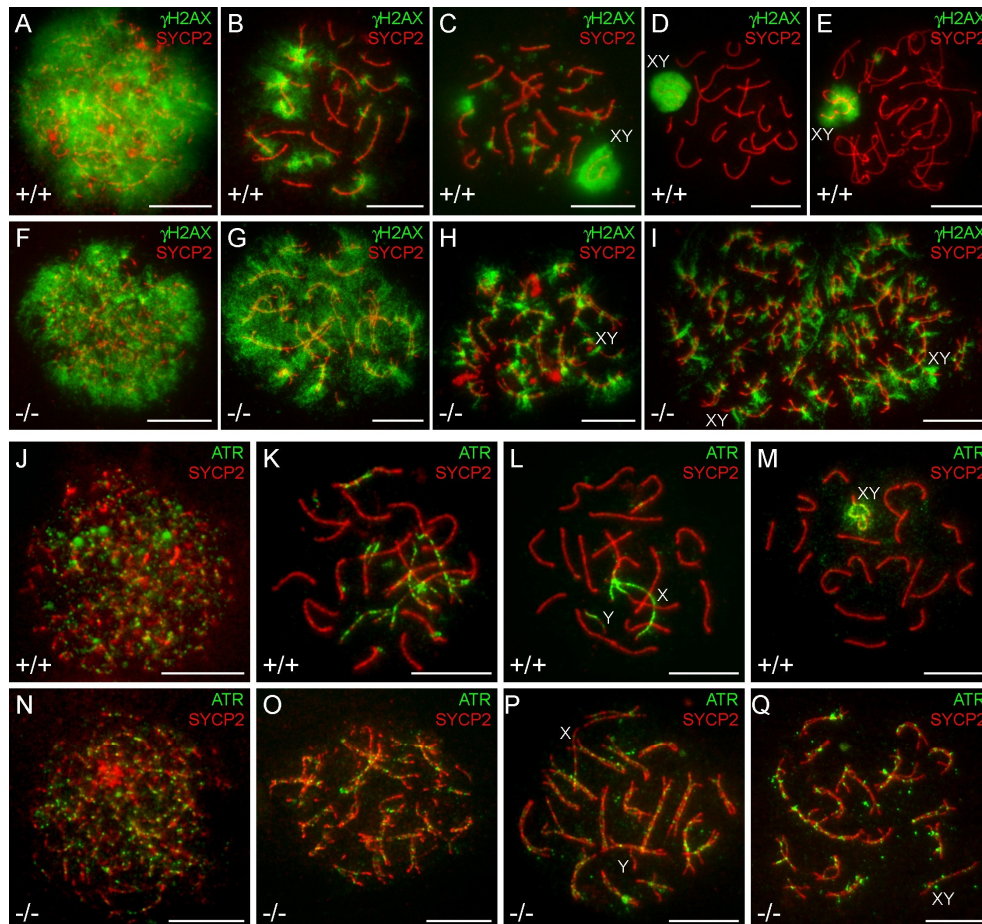
**Chapter 5 Figure 3**



**Legends figure 3:** see page 175



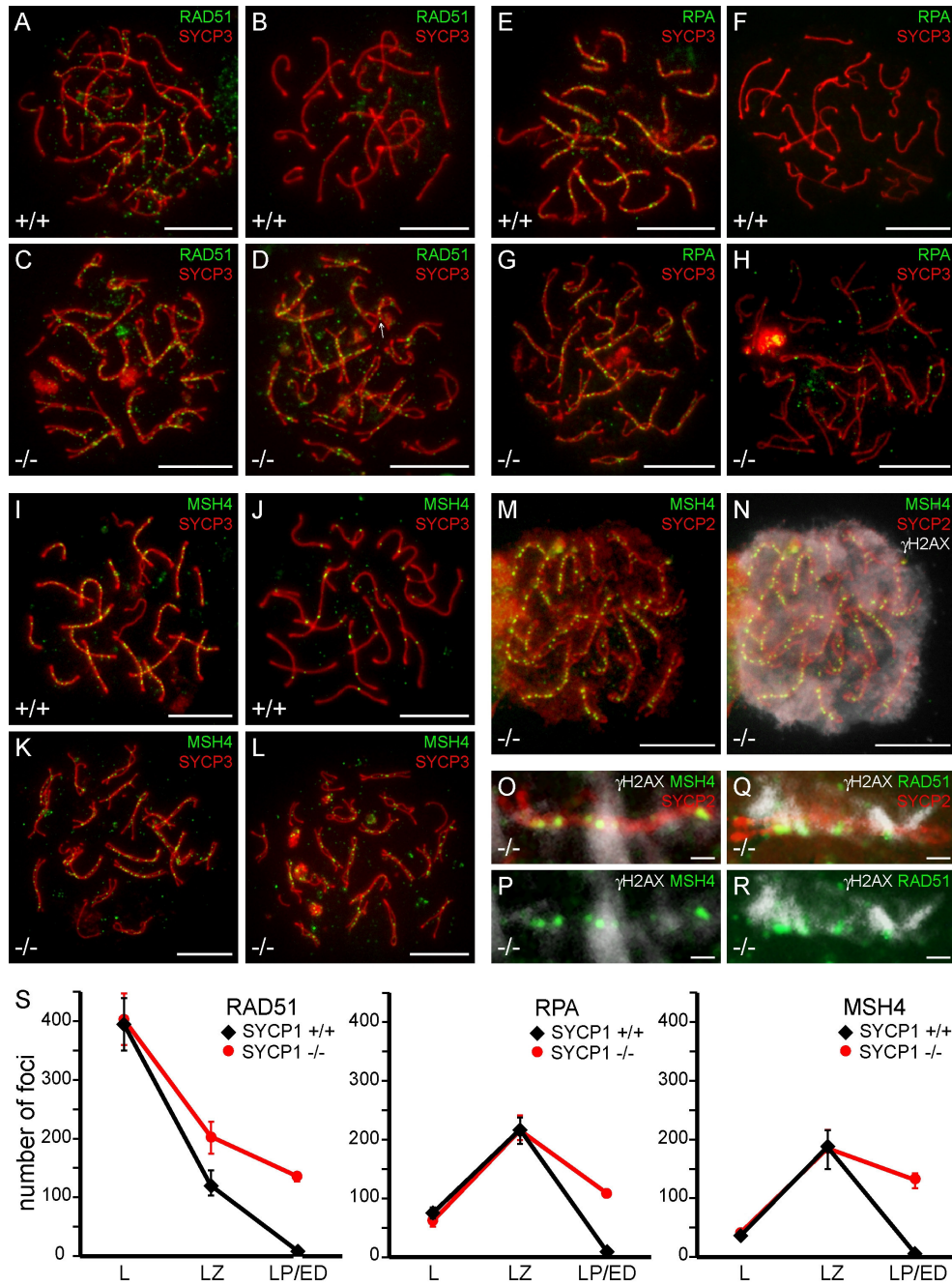
## Chapter 5 Figure 4



**Figure 4:**  $\gamma$ H2AX and ATR in wildtype (+/+) and *Sycp1*<sup>-/-</sup> (-/-) spermatocytes. **(A-I)**  $\gamma$ H2AX ; **(A,F)** leptone-; **(B,G)** zygonema; **(C)** early pachynema; **(D,H)** mid-pachynema; **(E,I)** diplonema; the sex chromosomes (XY) form an XY-body in wildtype spermatocytes **(C-E)**, but not in *Sycp1*<sup>-/-</sup> spermatocytes, even though the X and Y chromosomes are associated in the cells in **(H)** and **(I)**. **(J-Q)** ATR; **(J,N)** leptone-; **(K,O)** zygonema; **(L)** early pachynema and **(M)** and **(P)** mid-pachynema; **(Q)** diplonema; ATR is present throughout the chromatin of the XY bivalent in wildtype spermatocytes **(M)**, but forms foci and distinct domains along the X and Y chromosomes in *Sycp1*<sup>-/-</sup> cells **(P-Q)**. Insets in **(J)** and **(N)** show the close association of ATR with the ends of AE fragments in wildtype (+/+) and *Sycp1*<sup>-/-</sup> leptone-. Bars 10  $\mu$ m.



**Chapter 5 Figure 5**

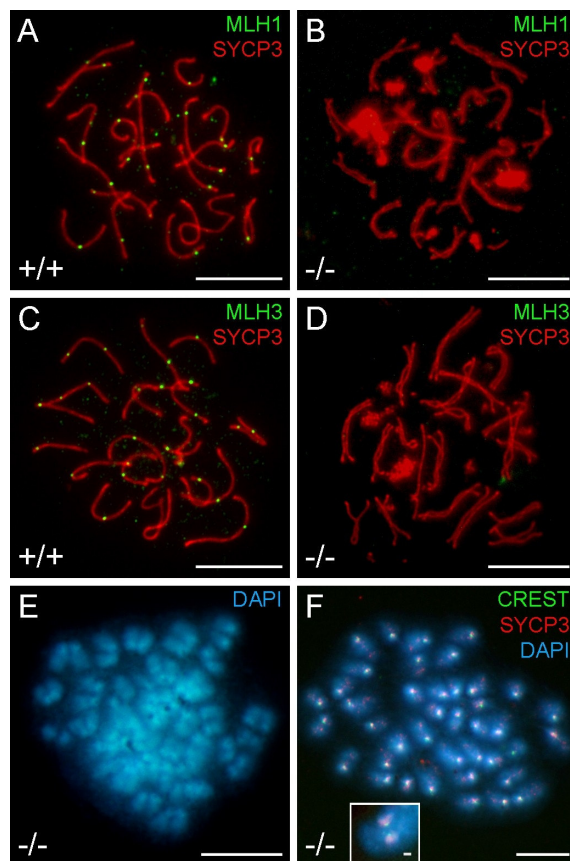


**Legends figure 5:** see page 179

**Figure 5:** Recombination-related proteins along AEs and SCs in wildtype (+/+) and *Sycp1*<sup>-/-</sup> (-/-) spermatocytes.

(A-D) RAD51/DMC1; (A,C) late zygonema; (B,D) late pachynema. (E-H) RPA; (E,G) late zygonema; (F,H) diplonema. (I-L) MSH4; (I,K) late zygonema; (J), mid-pachynema; (L), diplonema. (M-N) MSH4/SYCP2/ $\gamma$ H2AX triple labelling of a zygotene *Sycp1*<sup>-/-</sup> spermatocyte; the number and localization of MSH4 foci appears normal, but the persistence of  $\gamma$ H2AX throughout the chromatin is abnormal. (O-P) MSH4/ SYCP3/ $\gamma$ H2AX triple labelling of a late pachytene *Sycp1*<sup>-/-</sup> bivalent, to show that part of the  $\gamma$ H2AX domains co-localize with an MSH4 focus. (Q-R) RAD51/SYCP2/ $\gamma$ H2AX triple labelling of a late pachytene *Sycp1*<sup>-/-</sup> bivalent, to show that part of the  $\gamma$ H2AX domains co-localize with a RAD51 focus. (S) Counts of RAD51, RPA and MSH4 foci in successive stages of meiotic prophase; the vertical axes represent the number of AE or SC associated foci per cell; the vertical bars represent the observed range of the counts of the number of foci per cell in a given spermatocyte stage. For more details of the counts, see Supplementary Information, Fig. S4. Bars in (A-N) 10  $\mu$ m; bars in (O-R) 1  $\mu$ m.

### Chapter 5 Figure 6



**Figure 6:** Formation of crossovers and chiasmata.

(A,B) MLH1 labelling and (C,D) MLH3 labelling of wildtype (+/+) or *Sycp1*<sup>-/-</sup> (-/-) pachytene spermatocytes. The *Sycp1*<sup>-/-</sup> spermatocytes do not assemble MLH1 or MLH3 foci. (E,F) A natural (E) and an okadaic acid-induced (F) metaphase I spermatocyte of *Sycp1*<sup>-/-</sup>. In the cells shown here, only univalents can be identified; the inset in (F) shows a bivalent found in another OA-induced *Sycp1*<sup>-/-</sup> metaphase I. Bars in (A-F) 10  $\mu$ m; bar in the inset in (F) 1  $\mu$ m.

