

Physiology and pathophysiology of the ileal brake in humans $\ensuremath{\text{Vu, M.K.}}$

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Chapter 1

INTRODUCTION, AIMS AND OUTLINES OF THE THESIS

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INTRODUCTION

Ileal brake history

Digestion and absorption of nutrients are complex processes that involve various functions of the gastrointestinal (GI) tract. This includes the interplay between nutrients, digestive enzymes and gut surface area. Motility, transport of intraluminal content, secretion of enzymes and fluids are regulated by hormonal, neural (enteric and central nervous system) and local regulatory mechanisms. Intraluminal nutrients by themselves have a major role in controlling gastrointestinal transport, digestion and absorption. The presence of nutrients in the small intestine stimulates pancreatic enzyme secretion, gallbladder contraction and converts intestinal motility from the fasted into a fed motility pattern (1). On the other hand, intestinal nutrients also trigger feedback inhibitory mechanisms that will modify gastrointestinal transport, digestion and absorption. For instance, duodenal nutrients inhibit gastric acid secretion and delay gastric emptying. This phenomenon is called the duodenal brake, a negative feedback loop from the proximal gut on gastric functions (1-3).

A nutrient-triggered inhibitory feedback loop from the more distal to the proximal gut was first described by Spiller et al and Read *et al* (4,5). These two groups of researchers showed that transit of a meal through the small intestine was significantly delayed when a lipid emulsion was administered into the ileum. This phenomenon is called the ileal brake. Since then, evidence has increased showing in both humans and animals that intraileal nutrients alter intestinal motility, reduce transit time, delay gastric emptying and inhibit gastric acid and exocrine pancreatic secretion (6-8,15-20,25-28).

One may conclude that non-absorbed nutrients reaching the distal small bowel bring the process of transport, secretion, digestion and absorption to an end.

1. Ileal brake and gastric emptying

Earlier studies in both humans and dogs have demonstrated that infusion of nutrients into the ileum delays gastric emptying of both solid and liquid meals (6,7). These findings have been further extended by Fone et al, who showed that ileal fat inhibits antral and duodenal motility while stimulating pyloric contractions. Especially the latter may be responsible for the delayed gastric emptying (8). However, this is the only study demonstrating an association between antropyloroduodenal motility changes and delayed gastric emptying induced by ileal nutrients in humans. Not only the distal stomach, but also the proximal stomach contributes to gastric emptying. The motor function of the proximal stomach is characterised by receptive and adaptive relaxation (9,10). Receptive relaxation is induced by pharyngeal stimulation (swallowing) and distention of the eosophagus by the bolus of food. Adaptive relaxation or accommodation is the ability of the proximal stomach to distend in response to an intragastric load with only minimal changes in intragastric pressure (gastric tone) (11). A relationship between proximal gastric tone and gastric emptying has been described previously (12-14). Up till now it is not known whether nutrients in the distal gut affect proximal gastric motor functions.

2. Ileal brake and intestinal motility and transit

The inhibitory effect of ileal nutrients on small intestinal transit has been

confirmed by several studies since the original publications of Spiller et al and Read et al (4,5,15,16). On the other hand, data on the effect of ileal nutrients on digestive intestinal motility patterns are scarce and the various studies differ in methodology. Welch et al demonstrated the early occurrence of phase III after meal ingestion in 3 of 14 healthy subjects by infusion of lipid into the ileum (17). A study by Layer et al, using duodenal perfusion of a mixture of essential amino acids instead of a meal to induce a fed-like motor pattern, showed that ileal infusion of fat or carbohydrate induces premature phase III-like activity in 12 of 14 healthy subjects (18). With respect to the effects of ileal nutrients on fasting motor patterns, results are contrasting. Ileal fatty acids in dogs prolong interdigestive cycles and inhibit jejunal motility (19) whereas in humans, ileal perfusion of carbohydrates or fat during interdigestive phase I markedly decreases the duration of phase II motor activity, induces premature phase III motility and shortens the length of the interdigestive cycle (20). Although contradictory, these results nonetheless suggest that fasting motor activities may be modulated by the presence of nutrients in the distal small intestine. However, further research is needed to exactly define the effect of ileal nutrients on intestinal motility patterns in humans.

3. Ileal brake and gastric acid and exocrine pancreatic secretion

It is known that nutrients in the proximal small intestine, especially fat, potently inhibit gastric acid secretion (21,22). However, there is also evidence suggesting that gastric acid secretory function is regulated by the distal intestine. For instance, colonic perfusion of lipids or protein decreases exogenously stimulated gastric acid secretion in humans and dogs (23, 24). In addition, ileal perfusion of lipids or carbohydrates inhibits both unstimulated and endogenously stimulated

gastric acid secretion in humans (25). The effects of ileal nutrients on the secretory function have been more extensively studied in humans and animals with respect to exocrine pancreatic secretion (26-28, 18, 20). However, the obtained results are contradictory. While in rats and cats, ileal fat decreases pancreatic enzyme secretion this is not the case in dogs (26-28). Layer et al showed in humans that ileal perfusion with either carbohydrates or triglycerides inhibits the secretion of all exocrine pancreatic enzymes to an equal extent (18). On the other hand, results presented by Jain *et al* indicate that ileal carbohydrates do not inhibit but increase the release of the pancreatic enzyme amylase when compared to trypsin (29). Thus, the question whether ileal nutrients inhibit exocrine pancreatic secretion still remains to be answered.

4. Ileal brake and satiety

Although the role of the stomach and intraduodenal nutrients in the regulation of food intake and satiety is well established (30-33), effects from the distal gut are poorly defined because of the limited number of studies on this subject up till now. There are two human studies, both by Welch *et al*, who have demonstrated that ileal nutrients significantly reduce the total amount of food intake (34, 35). In addition, one study in rats showed that while ileal infusion of glucose reduces both meal frequency and size, ileal free fatty acids reduce only the latter (36). Although scarce, these consistent data imply that activation of the ileal brake decreases food intake and may induce early satiety. However, further studies are necessary to prove this theory.

Triggers of the ileal brake

The ileal brake is an intraluminal nutrient-triggered feedback control from the distal to the proximal gut. The inhibitory response of upper gastrointestinal motility differs with respect to the type of the nutrients administered into the ileum. In both humans and dogs, infusion of fat into the ileum delays gastric emptying and increases intestinal transit time (4-8; 37, 38). Within the range of different lipid emulsions, free fatty acids and digested triglycerides have been shown to be more potent than neutral triglycerides in eliciting the ileal brake effect (15). Intraileal carbohydrates, on the other hand, delay gastric emptying only at high concentrations (15). Data concerning intraileal proteins and the activation of the ileal brake are contradictory. Read at al found that ileal proteins delay small intestinal transit (5) whereas other investigators were not able to demonstrate any inhibitory effects of ileal proteins on gastrointestinal motility in humans (6, 15). These contrasting data may result from methodological differences. Nevertheless, it is generally accepted, based on consistent results from numerous studies, that fat is the most potent trigger of the ileal brake (39). However, it is important to bear in mind that species differences exist concerning triggers for the ileal brake. For instance, free fatty acid, a potent trigger of the ileal brake in humans and dogs, has no effect on the pig (40).

Mediators of the ileal brake

The mechanisms involved in the control of the ileal brake remain to be explored. The ileal brake may be mediated by hormonal and/or neural factors. *Hormonal factors:* A number of gut peptides, including glucagon-like peptide 1 (GLP-1), neurotensin and peptide YY (PYY) have been

hypothesized as possible humoral mediators of the ileal brake. Attention has been focused mainly on these peptides because of the localisation of their secretory cells, mainly in the distal gut.

GLP-1 is synthesized within the endocrine L cells in the intestine, primarily in the ileum and colon (41, 42). The release of GLP-1 is stimulated by the direct contact of the L cells to luminal nutrients (25, 43). However, given the rapid release of GLP-1 after meal intake, there is also evidence suggesting that GLP-1 release results from an indirect neural or humoral signals arising from the proximal gut (44, 45). GLP-1 plasma levels have been shown to increase in parallel with the inhibitory effects of the ileal brake on antral motility, gastric acid and exocrine pancreatic secretion (18, 25). However, up to now, there are no studies using a specific GLP-1 antagonist to clearly define the role of endogenous GLP-1 as a hormonal mediator of the ileal brake.

Neurotensin is produced by the mucosal endocrine N cells which are distributed throughout the small intestine, with the highest concentration found in the ileum (46-48). There is only scarce, indirect evidence available suggesting the role of neurotensin as a hormonal mediator of the ileal brake. Spiller *et al* have shown that ileal fat perfusion inhibits jejunal motility and significantly increases plasma neurotensin concentrations (5, 15).

PYY is, like GLP-1, also synthesized and secreted by L cells in the distal ileum and colon (49). The presence of nutrients, especially fats, in the ileum stimulates PYY release (4-8, 15). In contrast to the poorly established role of GLP-1 and neurotensin as hormonal mediators of the ileal brake, associations between plasma PYY concentrations and delayed small intestinal transit and gastric emptying have been shown by numerous investigators (5, 15, 37, 38).

In addition, the role of endogenous PYY as a mediator of the ileal brake has been elucidated using a PYY antagonist. In dogs, administration of PYY antibodies abolishes the prolonged small intestinal transit induced by intraileal fat (38). Although similar studies in humans are lacking, this direct evidence nonetheless suggests that PYY is very likely the humoral mediator of the ileal brake.

Neural factors: Several neural pathways have been suggested to contribute to the regulation of the ileal brake. The role of the extrinsic nervous system, in particular the vagus nerve, has been suggested but evidence is mostly indirect. It has been shown that intraileal fats increase vagal afferent activity in rats (50). In animals, the inhibitory effect of PYY and GLP-1 on meal stimulated gastric acid secretion and gastric motility was significantly reduced or even abolished after vagotomy (51, 52). These results suggest that both the candidate ileal brake hormones PYY and GLP-1 act through vagal innervation but a direct relationship between vagal cholinergic control and the fat induced ileal brake has not yet been proven. On the other hand, direct evidence exists demonstrating the involvement of the sympatho-adrenergic pathway in the inhibitory effect of the ileal brake. In dogs, administration of a combined α - and β -adrenergic blockade totally abolishes the inhibitory action of exogenous PYY en endogenous PYY release by ileal fat on exocrine pancreatic secretion (53). In addition, an adrenoceptor antagonist reverses the delayed intestinal transit induced by intraileal fat (54). Furthermore, evidence exists showing that the intrinsic nervous system (myenteric en submucosal plexus) also plays an important role the regulation of the ileal brake. The fatinduced ileal brake in dogs is abolished when ondansetron, a 5-HT3-receptor antagonist was administered into the proximal but not the distal small bowel (55). Similarly, the prolonged intestinal transit induced by fat in the ileum was blocked when naloxon, an opioid receptor antagonist was infused into the proximal small intestine (56). These findings suggest that both peripheral serotonergic and also opioid pathways are involved in the regulation of the ileal brake. However, it is plausible to assume that all the abovementioned different neural pathways interact and regulate the ileal brake.

Clinical implications of altered ileal brake function

Theoretically, the feedback function of the ileal brake could be impaired due to mucosal defects as in Crohn's ileitis and celiac disease or could be absent following resection of the distal small intestine. It is plausible to hypothesize that when the inhibitory feedback mechanism of the ileal brake is impaired or absent, gastric emptying and small intestinal transit are accelerated, resulting in increased concentrations of undigested and unabsorbed nutrients in the distal gut. This in turn could contribute to the development of symptoms such as diarrhea and malabsorption seen in inflammatory bowel diseases and after small intestine resection.

On the other hand, malabsorption and/or accelerated intestinal transit, irrespective of its cause, may also alter the ileal brake function. Given that PYY is a candidate hormonal mediator of the ileal brake, plasma PYY release could be considered as a marker for the activation of this feedback mechanism. Plasma PYY levels have been found to be increased in chronic pancreatitis and in patients with dumping syndrome after (partial) gastric resection (57-59). These findings suggest that the activation of the ileal brake is enhanced in these disease states due to the increased amount of unabsorbed nutrients reaching the distal small intestine. Based on the above made

assumptions, altered ileal brake function could be a primary defect, thereby giving rise to disease manifestations or secondary, in response to changes caused by the disease.

AIMS AND OUTLINES OF THE THESIS

Given its important role as a nutrient-triggered feedback control mechanism, increasing knowledge and understanding of the ileal brake is relevant for physiology and pathophysiology and may help to develop novel strategies in treating patients with malabsorption and maldigestion. The studies presented in this thesis have been designed to gain further insight into the physiology and pathophysiology of the ileal brake. The following issues were addressed:

Physiology of the ileal brake

It has been shown in dogs that the ileal brake is a more potent feedback mechanism compared to the more proximal, so called jejunal brake. However, a comparative study between jejunal and ileal brake has not been performed in humans. Chapter II was therefore designed to compare the effect of intraileal versus intrajejunal fat on digestive and interdigestive gastrointestinal motor patterns and postprandial gallbladder motility. The latter plays a role in delivering bile acids into the duodenum for the digestion of dietery fats. Furthermore, impaired gallbladder motility contributes to the pathogenesis of sludge and stone formation. Up til now little is known about the effect of the ileal brake on gallbladder motility. Gallbladder volumes were measured by real time ultrasonography and gastrointestinal motility was measured by means of the stationary water perfusion manometry system.

- The study in Chapter III was designed to investigate whether pancreatic and biliary secretion will be affected when nutrients are administered more distally into the small intestine than usual during enteral feeding. Duodenal outputs of pancreatic enzymes and bilirubin were measured by aspiration using a recovery marker in healthy volunteers.
- It has been shown in recent years that the distal gut hormone PYY has an important role in satiety and eating behaviour. However, little is known about the effect of the ileal brake on satiety and proximal gastric motor function. **Chapter IV** was undertaken to compare the effects of ileal brake activation with ileal fat (endogenous PYY release) versus exogenous PYY₃. ₃₆ infusion on satiety and on proximal gastric motor function. Two experimental protocols were used. In the first protocol, the effect of ileal fat and subsequent endogenous PYY release was studied and in the second protocol, the dose-response relationship of exogenous PYY was investigated. In both protocols satiety and motor function of the proximal stomach were monitored using Visual Analog Scale (VAS) and an electronic barostat, respectively. Plasma PYY levels were measured by radioimmunoassay.
- Medium chain triglycerides (MCT) are thought to be hydrolysed and absorbed more rapidly and completely compared to long chain triglycerides (LCT). However, patients receiving MCT frequently complain of nausea, cramps, abdominal pain and diarrhoea. We hypothesized that MCT are less rapidly absorbed and cause these gastrointestinal side effects. The release of the distal gut hormone PYY induced by intraduodenal MCT was used as evidence for the hypothesized malabsorption of MCT. Results are described in **Chapter V.**

• **Chapter VI** investigates whether artificially induced malabsorption in healthy volunteers affects gastrointestinal and gallbladder motility through activation of the ileal brake. The osmotic laxative magnesium sulphate was used to induce malabsorption after ingestion of a fatty meal.

Pathophysiology of the ileal brake

- Altered gastrointestinal motility has been observed in patients with chronic pancreatitis with impaired exocrine function. However, the reported results are conflicting. In Chapter VII we therefore investigated digestive and interdigestive antroduodenal motility and secretion of several relevant gut hormones in a large group of patients with chronic pancreatitis. Differences in gut motility and hormone secretion were compared between patients with and without exocrine pancreatic insufficiency. Gastrointestinal motility and also studied after hormone secretion were pancreatic enzyme supplementation in order to further elucidate the role of exocrine pancreatic insufficiency and subsequent maldigestion in patient with chronic pancreatitis
- **Chapter VIII** deals with patients with systemic sclerosis. This systemic disorder may give rise to various complications within the gastrointestinal tract. In this chapter we focused on antroduodenojejunal motility and proximal and distal gut hormone release in patients with diffuse and limited type of systemic sclerosis. The obtained data were related to esophageal manometry findings and gastrointestinal symptoms.
- It is known that patients with Crohn's disease have an increased risk of developing gallstones. When considering the possible mechanisms that contribute to gallstone formation in these patients the questions are: 1)

whether gallbladder motility plays a role in the pathogenesis of gallstone formation in Crohn's disease and 2) whether changes in gallbladder motility are explained by altered ileal brake function due to disease localization and bowel resection. These items have been investigated and results are described in **chapter IX**.

REFERENCES

- 1. Text book of Gastroenterology. Tadataka Yamada, 2003.
- 2. Parr NJ, Grime JS, Baxter JN, Critchley M, Mackie CR. Small bowel resistances and the gastroduodenal brake. Gut. 1987 Aug; 28(8):950-4.
- Shahidullah M, Kennedy TL, Parks TG. The vagus, the duodenal brake, and gastric emptying. Gut. 1975 May;16(5):331-6.
- Spiller RC, Trotman IF, Higgins BE, Ghatei MA, Grimble GK, Lee YC, Bloom SR, Misiewicz JJ, Silk DB. The ileal brake--inhibition of jejunal motility after ileal fat perfusion in man. Gut. 1984 Apr;25(4):365-74.
- Read NW, McFarlane A, Kinsman RI, Bates TE, Blackhall NW, Farrar GB, Hall JC, Moss G, Morris AP, O'Neill B. Effect of infusion of nutrient solutions into the ileum on gastrointestinal transit and plasma levels of neurotensin and enteroglucagon.. Gastroenterology. 1984 Feb;86(2):274-80.
- 6. Welch IM, Cunningham KM, Read NW. Regulation of gastric emptying by ileal nutrients in humans. Gastroenterology. 1988 Feb;94(2):401-4.
- Holgate AM, Read NW. Effect of ileal infusion of intralipid on gastrointestinal transit, ileal flow rate, and carbohydrate absorption in humans after ingestion of a liquid meal. Gastroenterology. 1985 Apr;88(4):1005-11.
- 8. Fone DR, Horowitz M, Read NW, Dent J, Maddox A. The effect of terminal

ileal triglyceride infusion on gastroduodenal motility and the intragastric distribution of a solid meal. Gastroenterology. 1990 Mar;98(3):568-75.

- Jahnberg T, Abrahamsson H, Jansson G, Martinson J. Gastric relaxatory response to feeding before and after vagotomy. Scand J Gastroenterol. 1977;12(2):225-28.
- Jahnberg T. Gastric adaptive relaxation. Effects of vagal activation and vagotomy. An experimental study in dogs and in man. Scand J Gastroenterol Suppl. 1977;46:1-32.
- Azpiroz F, Malagelada JR. Gastric tone measured by an electronic barostat in health and postsurgical gastroparesis. Gastroenterology. 1987 Apr;92(4):934-43.
- Azpiroz F. Control of gastric emptying by gastric tone. Dig Dis Sci. 1994 Dec;39(12 Suppl):18S-19S.
- Moragas G, Azpiroz F, Pavia J, Malagelada JR. Relations among intragastric pressure, postcibal perception, and gastric emptying. Am J Physiol. 1993 Jun;264(6 Pt 1):G1112-7.
- Ropert A, des Varannes SB, Bizais Y, Roze C, Galmiche JP. Simultaneous assessment of liquid emptying and proximal gastric tone in humans. Gastroenterology. 1993 Sep;105(3):667-74.
- 15. Spiller RC, Trotman IF, Adrian TE, Bloom SR, Misiewicz JJ, Silk DB. Further characterisation of the 'ileal brake' reflex in man--effect of ileal infusion of partial digests of fat, protein, and starch on jejunal motility and release of neurotensin, enteroglucagon, and peptide YY. Gut. 1988 Aug;29(8):1042-51.
- 16. Lin HC, Zhao XT, Wang L. Intestinal transit is more potently inhibited by fat in the distal (ileal brake) than in the proximal (jejunal brake) gut. Dig Dis

Sci. 1997 Jan;42(1):19-25.

- Welch IM, Davison PA, Worlding J, Read NW. Effect of ileal infusion of lipid on jejunal motor patterns after a nutrient and nonnutrient meal. Am J Physiol. 1988 Dec;255:G800-6.
- Layer P, Peschel S, Schlesinger T, Goebell H. Human pancreatic secretion and intestinal motility: effects of ileal nutrient perfusion. Am J Physiol. 1990;258(2 Pt 1):G196-201.
- Wen J, Phillips SF, Sarr MG, Kost LJ, Holst JJ. PYY and GLP-1 contribute to feedback inhibition from the canine ileum and colon. Am J Physiol. 1995 Dec;269(6 Pt 1):G945-52.
- 20. Layer P, Schlesinger T, Groger G, Goebell H. Modulation of human periodic interdigestive gastrointestinal motor and pancreatic function by the ileum. Pancreas. 1993 Jul;8(4):426-32.
- Hopert R, Liehr RM, Emde C, Riecken EO. Reduction of 24-hour gastric acidity by different dietary regimens: a randomized controlled study in healthy volunteers. JPEN J Parenter Enteral Nutr. 1989 May-Jun;13(3):292-5.
- Lloyd KC, Wang J, Solomon TE. Acid inhibition by intestinal nutrients mediated by CCK-A receptors but not plasma CCK. Am J Physiol Gastrointest Liver Physiol. 2001 Oct;281(4):G924-30.
- Jian R, Besterman HS, Sarson DL, Aymes C, Hostein J, Bloom SR, Rambaud JC. Colonic inhibition of gastric secretion in man. Dig Dis Sci. 1981 Mar;26(3):195-201.
- Seal AM, Debas HT. Colonic inhibition of gastric acid secretion in the dog. Gastroenterology. 1980 Nov;79(5 Pt 1):823-6.
- 25. Layer P, Holst JJ, Grandt D, Goebell H. Ileal release of glucagon-like

peptide-1 (GLP-1). Association with inhibition of gastric acid secretion in humans. Dig Dis Sci. 1995 May;40(5):1074-82.

- Harper AA, Hood AJ, Mushens J, Smy JR. Inhibition of external pancreatic secretion by intracolonic and intraileal infusions in the cat. J Physiol. 1979 Jul;292:445-54.
- Laugier R, Sarles H. Action of oleic acid on the exocrine pancreatic secretion of the conscious rat: evidence for an anti-cholecystokininpancreozymin factor. J Physiol. 1977 Sep;271(1):81-92.
- Hage G, Tiscornia O, Palasciano G, Sarles HInhibition of pancreatic exocrine secretion by intra-colonic oleic acid infusion in the dog. Biomedicine. 1974 Jun 10;21(6):263-7.
- 29. Jain NK, Boivin M, Zinsmeister AR, DiMagno EP. The ileum and carbohydrate-mediated feedback regulation of postprandial pancreaticobiliary secretion in normal humans. Pancreas. 1991 Sep;6(5):495-505.
- Deutsch JA. The role of the stomach in eating. Am J Clin Nutr. 1985 Nov;42(5 Suppl):1040-3.
- Phillips RJ, Powley TL. Gastric volume rather than nutrient content inhibits food intake. Am J Physiol. 1996 Sep;271(3 Pt 2):R766-9.
- 32. Matzinger D, Gutzwiller JP, Drewe J, Orban A, Engel R, D'Amato M, Rovati L, Beglinger C. Inhibition of food intake in response to intestinal lipid is mediated by cholecystokinin in humans. Am J Physiol. 1999 Dec;277(6 Pt 2):R1718-24.
- 33. Lieverse RJ, Jansen JB, Masclee AA, Rovati LC, Lamers CB. Effect of a low dose of intraduodenal fat on satiety in humans: studies using the type A cholecystokinin receptor antagonist loxiglumide. Gut.1994 Apr;35(4):501-5.

- Welch I, Saunders K, Read NW. Effect of ileal and intravenous infusions of fat emulsions on feeding and satiety in human volunteers. Gastroenterology. 1985 Dec;89(6):1293-7.
- 35. Welch IM, Sepple CP, Read NW. Comparisons of the effects on satiety and eating behaviour of infusion of lipid into the different regions of the small intestine. Gut. 1988 Mar;29(3):306-11.
- 36. Woltman T, Reidelberger R. Effects of duodenal and distal ileal infusions of glucose and oleic acid on meal patterns in rats. Am J Physiol. 1995 Jul;269(1 Pt 2):R7-14.
- 37. Pironi L, Stanghellini V, Miglioli M, Corinaldesi R, De Giorgio R, Ruggeri E, Tosetti C, Poggioli G, Morselli Labate AM, Monetti N. Fat-induced ileal brake in humans: a dose-dependent phenomenon correlated to the plasma levels of peptide YY. Gastroenterology. 1993 Sep;105(3):733-9.
- Lin HC, Zhao XT, Wang L, Wong H. Fat-induced ileal brake in the dog depends on peptide YY. Gastroenterology. 1996 May;110(5):1491-5.
- Van Citters GW, Lin HC. The ileal brake: a fifteen-year progress report. Curr Gastroenterol Rep. 1999 Oct;1(5):404-9.
- 40. Dobson CL, Hinchcliffe M, Davis SS, Chauhan S, Wilding IR. Is the pig a good animal model for studying the human ileal brake? J Pharm Sci. 1998 May;87(5):565-8.
- Schirra J, Goke B. The physiological role of GLP-1 in human: incretin, ileal brake or more? Regul Pept. 2005 Jun 15;128(2):109-15.
- 42. Eissele R, Goke R, Willemer S, Harthus HP, Vermeer H, Arnold R, Goke B. Glucagon-like peptide-1 cells in the gastrointestinal tract and pancreas of rat, pig and man. Eur J Clin Invest. 1992 Apr;22(4):283-91.
- 43. Herrmann-Rinke C, Voge A, Hess M, Goke B. Regulation of glucagon-like

peptide-1 secretion from rat ileum by neurotransmitters and peptides. J Endocrinol. 1995 Oct;147(1):25-31.

- 44. Rocca AS, Brubaker PL. Role of the vagus nerve in mediating proximal nutrient-induced glucagon-like peptide-1 secretion. Endocrinology. 1999 Apr;140(4):1687-94.
- 45. Balks HJ, Holst JJ, von zur Muhlen A, Brabant G. Rapid oscillations in plasma glucagon-like peptide-1 (GLP-1) in humans: cholinergic control of GLP-1 secretion via muscarinic receptors. J Clin Endocrinol Metab. 1997 Mar;82(3):786-90.
- 46. Bloom SR, Polak JM. Aspects of neurotensin physiology and pathology. Ann N Y Acad Sci 1982;400:105-116.
- 47. Flaten O, Hanssen LE. Concentration of neurotensin in human plasma after glucose, meals and lipids. Acta Physiol Scand. 1982 Feb;114(2):311-3.
- 48. Ferris CF, George JK, Eastwood G, Potegal M, Carraway RE. Plasma levels of human neurotensin: methodological and physiological considerations. Peptides. 1991 Mar-Apr;12(2):215-20.
- Adrian TE, Ferri GL, Bacarese-Hamilton AJ, Fuessl HS, Polak JM, Bloom SR. Human distribution and release of a putative new gut hormone, peptide YY. Gastroenterology. 1985 Nov;89(5):1070-7.
- 50. Randich A, Tyler WJ, Cox JE, Meller ST, Kelm GR, Bharaj SS. Responses of celiac and cervical vagal afferents to infusions of lipids in the jejunum or ileum of the rat. Am J Physiol Regul Integr Comp Physiol. 2000 Jan;278(1):R34-43.
- 51. Chen CH, Rogers RC. Central inhibitory action of peptide YY on gastric motility in rats. Am J Physiol. 1995 Oct;269(4 Pt 2):R787-92.
- 52. Lloyd KC, Amirmoazzami S, Friedik F, Heynio A, Solomon TE, Walsh JH.

Candidate canine enterogastrones: acid inhibition before and after vagotomy. Am J Physiol. 1997 May;272(5 Pt 1):G1236-42.

- 53. Konturek SJ, Bilski J, Pawlik W, Tasler J, Domschke W. Adrenergic pathway in the inhibition of pancreatic secretion by peptide YY in dogs. Gastroenterology. 1988 Feb;94(2):266-73.
- 54. Lin HC, Neevel C, Chen PS, Suh G, Chen JH. Slowing of intestinal transit by fat or peptide YY depends on beta-adrenergic pathway. Am J Physiol Gastrointest Liver Physiol. 2003 Dec;285(6):G1310-6.
- 55. Lin HC, Chen JH. Slowing of intestinal transit by fat depends on an ondansetron - sensitive, efferent serotonergic pathway. Neurogastroenterol Motil. 2003 Jun;15(3):317-22.
- Zhao XT, Wang L, Lin HC. Slowing of intestinal transit by fat depends on naloxone-blockable efferent, opioid pathway. Am J Physiol Gastrointest Liver Physiol. 2000 Jun;278(6):G866-70.
- 57. Adrian TE, Savage AP, Bacarese-Hamilton AJ, Wolfe K, Besterman HS, Bloom SR. Peptide YY abnormalities in gastrointestinal diseases. Gastroenterology. 1986 Feb;90(2):379-84.
- Adrian TE, Long RG, Fuessl HS, Bloom SR. Plasma peptide YY (PYY) in dumping syndrome. Dig Dis Sci. 1985 Dec;30(12):1145-8.
- 59. El-Salhy M, Suhr O, Danielsson A. Peptide YY in gastrointestinal disorders. Peptides. 2002 Feb;23(2):397-402.