

Synthesis & biological applications of glycosylated iminosugars Duivenvoorden, B.A.

Citation

Duivenvoorden, B. A. (2011, December 15). *Synthesis & biological applications of glycosylated iminosugars*. Retrieved from https://hdl.handle.net/1887/18246

Version:	Corrected Publisher's Version
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B Design and Synthesis of Three Novel Human Chitinase Fluorogenic Substrates*

3.1 Introduction

Until the end of the last century it was believed that man lacked the ability to process chitin, a linear polymer of β -1,4-linked *N*-acetyl-D-glucosamines which is found on cell walls and coating of many organisms. The first mammalian chitinase was serendipitously discovered in the search for elevated serum glycosidase activity in patients suffering from Gaucher disease, a rare lysosomal storage disease in which glucosylceramide (GC) is inefficiently processed by mutant β -glucocerebrosidase (GBA1).^{1–4}

This enzyme, identified as human chitotriosidase (CHIT1), is nowadays used as marker to reflect the total body burden on Gaucher cells.⁵ A drawback in the marker assays is the disproportional fluorophore to enzyme ratio (Figure 3.1 A), which is found when using high concentrations of 4-methylumbelliferyl-chitobiose **126** (Figure 2.1) as substrate.⁶ This is caused by the transglycosylase activity of CHIT1 forming larger chito-oligomers as substrates, resulting in inefficient release

^{*} Duivenvoorden, B. A.; Ghauharali, K.; Bussink, A. P.; Codée, J. D. C.; van der Marel, G. A.; Scheij, S.; Verhoek, M.; Overkleeft, H. S.; Groener, J. E.; Aerts, J. M. F. G.; Boot, R. G. *Manuscript in preparation*.

of 4-methylumbelliferyl (4-MU), the fluorophore. To circumvent the possibility of CHIT1 mediated transglycosylation a modified fluorogenic substrate **125** was synthesized (**Chapter 2**), in which the 4'-OH is removed.⁶ This derivative follows Michaelis-Menten kinetics giving rise to a proportional 4-MU to enzyme ratio (Figure 3.1 A).^{6,7} More recent studies⁸ showed that **125** is also the substrate of choice when dealing with Gaucher patients having the common polymorphism (G102S) in CHIT1, which shows a slightly impaired catalytic activity toward 4-MU-chitobioside substrate **126** as compared to the wild type CHIT1. However, G102S-CHIT1 activity is normal when using 4-methylumbelliferyl 4'-deoxychitobioside **125** as substrate. It should further be mentioned that increased plasma chitotriosidase activity is increased, albeit much more modestly, in several lysosomal and nonlysosomal diseases, such as sarcoidosis, visceral leishmaniasis, leprosy, arthritis, multiple sclerosis, thalassemia, chronic obstructive pulmonary disease (COPD), malaria, and atherosclerosis.^{2,9–17}

To gain a bigger pool of effective CHIT1 fluorescent substrates, this chapter describes the synthesis of three novel human chitinase substrates **136,137** and **138** bearing an anomeric 4-MU for fluorometric read-out. The three substrates have a different modification on the 4'-OH of the non-reducing sugar, going from the relative small *O*-methyl group (OMe) to the more sterically demanding *O*-isopropyl (*i*OPr) and *O*-methyl cyclohexane group (OMCH). A 1,6-anhydro building block is used as a common precursor for the synthesis of the donors and acceptors which after condensation and further modifications, will be evaluated as substrates for CHIT1.

3.2 Results and discusion

The use of an 1,6-anhydro building block for the synthesis of the donors and acceptors helps to overcome the low reactivity of the 4-OH function of *N*-acetyl-glucosamine, which is well recognized.¹⁸ These constrained 1,6-anhydro sugars are know to exhibit enhanced reactivity of the 4-OH function over there unconstrained counterparts.¹⁹ Further enhancement of the reactivity of the 4-OH can be gained by the use of an azide group on the 2-position of glucosamine, resulting in a 10 fold increase of reactivity as compared to *N*-acetyl or *N*-phthalimido-protected acceptors, as shown by the group of Crich.¹⁸ By using 1,6-anhydro glucosamine **148** as the key building block a route was developed for the synthesis of three novel fluorogenic 4-methylumbelliferyl chiotobiose substrates, bearing a modification on the 4'-OH of the non-reducing end the retrosynthesis of which is outlined in Scheme 3.1.

First acceptor **148** was synthesized *via* a modified literature procedure.^{20–22} Deacetylation of tri-*O*-acetyl-D-glucal and treatment with bis(tributyltin)oxide in



Scheme 3.1: Route of synthesis of three novel human chitinase fluorogenic substrates.

refluxing acetonitrile, followed by 1,6-iodocyclization^{21,23,24} gave compound **150** (Scheme 3.2). Compound **150** was then heated in a DMF:H₂O mixture and treated with a mild base (NaHCO₃) to yield 1,6:2,3-bis-anhydro- β -D-glucopyranose **151**. Cerny epoxide **151** was carefully purified to prevent the formation of the unwanted regio-isomer in the next step. This side reaction was attributed to the presence of trace amounts of tin. Opening of the epoxide of **151** with sodium azide, followed by benzylation of the diol and subsequently regioselective debenzylation using TiCl₄ gave acceptor **148**.²⁵

Scheme 3.2: Synthesis of key building block 148.



Reagents and conditions: a) $MeOH/H_2O/Et_3N$; b) (1) $(Bu_3Sn)_2O$, Δ , ACN; (2) I_2 , DCM, 4 °C; c) NaHCO₃, DMF/H₂O, 98% over three steps; d) NaN₃, DMF/H₂O, 120 °C, 52%; e) BnBr, NaH, DMF, 0 °C, 80%; f) TiCl₄, DCM, 75%.

Alkylation of the 4-position of anhydro sugar **148** using sodium hydride and MeI in DMF yielded compound **154** (Scheme 3.3) in a good yield. Under similar conditions the application of *Ii*Pr and BrMCH (bromomethyl cyclohexane) gave rise to compounds **155** and **156** in slightly reduced yields. Subsequently all three 1,6-anhydro sugars were opened under acidic conditions followed by *in situ* acetylation, using Ac₂O and TFA. Selective deprotection of anomeric position and treatment with trichloroacenitrile and DBU led to the isolation of anomeric mixture of

imidates 145, 146 and 147 with α as the major isomer.





Reagents and conditions: a) MeI, NaH, DMF, 91%; b) *Ii*Pr, NaH, DMF, 60%; c) BrMCH, NaH, DMF, 75%; d) Ac₂O, TFA (10% v/v), **157**: 90%, **158**: 86%, **159**: 72%; e)THF, piperidine (6%v/v), **160**: 71%, **161**: quant., **162**: 98%; f) CCl₃CN, DBU, DCM, **145**: 83%, **146**: 70%, **147**: 71%.

Couplings of the imidates (145, 146, 147) with acceptor 148 (Scheme 3.4) were performed at -80 °C, in dry toluene under influence of BF₃ · OEt₂ yielding dimers 142, 143 and 144 in high yield and high β -selectivity.²⁶ After deacetylation, pure β -anomers (163, 164, 165) were obtained by silica gel chromatography. Trifluoroacetic acid and Ac₂O mediated opening of the 1,6-anhydro sugar in 163, 164, 165 resulted in the formation of dimers 166, 167 and 168. The final steps towards the target substrates involved several protective group manipulations. Starting off with a Staudinger reduction of the azides, the released amines were acetylated using Ac₂O and pyridine. The benzyl groups were removed by hydrogenolysis using Pearlman's catalyst in MeOH:2,2,2-trifluoroethanol (TFE). Preceding the introduction of the fluorophore at the anomeric center the free hydroxyls were protected with acetyl groups yielding intermediate 139, 140 and 141.

Fluorophore 4-MU was selected by virtue of its easy quantification in fluorometric assays. Introduction of 4-MU can be attained by phase transfer conditions (PTC) which involves a halide donor in combination with a phenolate also known as the Michael procedure.^{27,28} Conversion of the anomeric acetate in **139**, **140** and **141** into the corresponding α -chloride was found to be troublesome. Several methods were explored including ZnCl₂ and α, α -2,2-dichloromethyl methyl ether (DCMME)²⁹, AcCl and HCl³⁰ however, these did not give satisfactory results. A combination of AcOH and Ac₂O at 0 °C and dry HCl gas, produced using Kipp conditions (HCl and H₂SO₄), gave the highest yields and most reproducible results. ^{31,32} The obtained chlorides were coupled with fluorophore 4-MU *via* a optimized Michael procedure using NaHCO₃ as a base, an excess of 4-MU sodium salt and TBAHS gave the best results to yield compounds **169**, **170** and **171**. Deacetylation under Zemplén conditions and HPLC purification yielded the final products **136**, **137** and **138** as white powders.

Scheme 3.4: Synthesis of three humane fluorogenic chitinase substrates 136, 137 and 138.



Reagents and conditions: a) $BF_3 \cdot Et_2O$, Tol, -80 °C, **142**: 83%, **143**: 81 %, **144**: 65%; b) NaOMe, MeOH, **163**: 65%, **164**: 50%, **165**: 52%; c) Ac_2O , TFA (15% v/v), **166**: 90%, **167**: 80%, **168**: 87%; d) (1) PMe_3, Tol:H₂O:dioxane; (2) Ac_2O , Pyr. (3) Pd(OH)₂, H₂, MeOH, TFE; (4) Ac_2O , Pyr., **139**: 48%, **140**: 40%, **141**: 38%; e) (1) AcOH, Ac_2O , HCl_g, 0 °C; (2) Na 4-MU, TBHS, NaHCO₃ 0.2M, CHCl₃, **169**: 25%, **170**: 17%, **171**: 50%; f) NaOMe, MeOH, **136**: 37%, **137**: 60%, **138**: 22%.

The three human chitinase substrates (**136**, **137** and **138**) were tested for their ability to be processed by human chitinase CHIT1. CHIT1 is able to degrade 4-MU-chitotriose and 4-MU-chitobiose by removal of the oligosaccharide moiety and concomitant release of fluorescent 4-MU. However, the ongoing transglycosylation of the substrates, results in a reduced 4-MU release at higher substrate concentrations (Figure 3.1 A). As described in **Chapter 2** 4-MU-deoxychitobiose **125**

is an improved fluorogenic substrate, which indeed allows a superior fluorometric assay of chitinase activity since the interfering transglycosylation of substrate does not occur (Figure 3.1 A). Compounds **136**, **137** and **138** all showed Michaelis-Menten kinetics like the parent 4'-deoxy substrate **125** (Figure 3.1 B).



Figure 3.1: (A) Michaelis-Menten kinetics of 4-MU-deoxychitobiose; \circ :125, \bullet :4-MU-chitobiose 126, \Box :4-MU-chitotriose; (B) Michaelis-Menten kinetics chitinase substrates bearing a modification at the 4-position of the non-reducing end; \circ :125, \triangle :136, ∇ :137, \star :138.

Km values of the new 4-MU-substrates become higher with a more bulky substituent: 37, 50, 88, and 200 μ M for **125**, **136**, **137** and **138**, respectively. V_{max} values are quite similar for **125**, **136** and **137**: 5.0, 4.7 and 4.4 mmol/mg CHIT1/h, respectively. The V_{max} value for **138**, (1.7 mmol/mg chitotriosidase/h) is clearly lower. Nevertheless none of the three novel compounds was superior to the 4-deoxy derivative **125**.

In plasma or tissue extracts a stepwise degradation of chitinase substrates can occur. This reaction is catalyzed by β -hexosaminidase which slowly and stepwise removes a GluNAc moiety from the non-reducing end, resulting in undesired background release of 4-MU.³³ Therefore, the ability of jack bean (*Canavalia*) β -hexosaminidase to sequentially hydrolyze the modified 4-MU-chitobioses was examined, its enzymatic activity towards 4-MU-GlcNAc (0.135 mM) was first determined at the optimal pH of 4.0. The assay showed to be linear in time over 60 minutes (Figure 3.2 A).

An identical amount of enzyme was incubated with 0.135 mM of 4-MU-chitobioses with different modifications at 4-position of the non-reducing end (**136**, **137** and **138**) as well as the 4'-deoxy derivative **125** and the unmodified substrate. Both the unmodified 4-MU-chitobiose substrate and **125** are relatively good substrates, resulting in gradual release of the fluorescent leaving group (Figure 3.2 B). After 1 hour incubation with 4-MU-chitbiose the amount of 4-MU released by jack bean β -hexosaminidase was about 4% of that released from 4-MU-GlcNac under similar conditions. In contrast, in the case of **125** this was about 2%. Figure 3.2 B shows that substrates bearing a modification at the 4-position of the



Figure 3.2: (A) Hydrolysis of 4-MU-GlcNAc (0.135 mM) over 60 minutes by jack bean β -hexosaminidase; **I**:4-MU-GlcNAc; (B) Hydrolysis of chitobiose and derivatives bearing a modification at the 4-position of the non-reducing end; •:4-MU-chitobiose **126**, •:**125**, \triangle :**136**, **V**:**137**, *****:**138**.

non-reducing end, particularly 4'-isopropyloxychitobiosyl umbelliferone **137** and 4'-cyclohexylmethoxychitobiosyl umbelliferone **138**, are much more resistant towards jack bean β -hexosaminidase mediated hydrolysis, most likely due to the steric bulk which precludes binding of a other sugar to the active site of this β -hexosaminidase.

3.3 Conclusion

This chapter describes the synthesis of three novel human chitinase fluorogenic substrates, using 1,6-anhydrosugar **148** as a common building block. Anhydro sugar **148** was not only transformed into a glycosyl acceptor but was also used as precursor in the synthesis of the three different imidate donors (**145**, **146**, **147**). Key step entailed the coupling under phase transfer conditions of the fluorophore and the dimeric α -chlorides.

The newly designed compounds **136**, **137** and **138** do not act as acceptors in transglycosylation and offer substrates for CHIT1 that are hydrolysed according to Michaelis-Menten kinetics. An additional advantage is that the novel compounds are lesser substrates for β -hexosaminidases. In situations where significant β -hexosaminidase activity is suspected in a sample, next to chitinase activity, and where one aims to monitor specifically chitinase activity, the latter two substrates (4'-isopropyloxychitobiosyl umbelliferone **137** and 4'-cyclohexylmethoxychitobiosyl umbelliferone **138**) may be the reagents of choice.

3.4 Experimental section

All reagent were of commercial grade and used as received (Acros, Fluka, Merck, Schleicher & Schuell) unless stated otherwise. Diethyl ether (Et₂O), light petroleum ether (PE 40-60), en toluene (Tol) were purchased from Riedel-de Haën. Dichloromethane (DCM), *N*,*N*-

dimethylformamide (DMF), methanol (MeOH), pyridine (pyr) and tetrahydrofuran (THF) were obtained from Biosolve. THF was distilled over LiAlH₄ before use. Dichloromethane was boiled under reflux over P₂O₅ for 2 h and distilled prior to use. Molecular sieves 3Å were flame dried under vacuum before use. All reactions sensitive to moisture or oxygen were performed under an inert atmosphere of argon unless stated otherwise. Solvents used for flash chromatography were of pro analysis quality. Flash chromatography was performed on Screening Devices silica gel 60 (0.004 - 0.063 mm). TLC-analysis was conducted on DC-alufolien (Merck, Kieselgel60, F245) with detection by UV-absorption (254 nm) for UV-active compounds and by spraying with 20% H₂SO₄ in ethanol or with a solution of $(NH_4)_6 Mo_7 O_{24} \cdot 4 H_2 O 25 g/L$, $(NH_4)_4 Ce(SO_4)_4 \cdot 2 H_2 O 10 g/L$, 10% $H_2 SO_4$ in $H_2 O$ followed by charring at \sim 150 °C. ¹H and ¹³C NMR spectra were recorded on a Bruker DMX-400 (400/100 MHz), a Bruker AV 400 (400/100 MHz), a Bruker AV 500 (500/125 MHz) or a Bruker DMX-600 (600/150 MHz) spectrometer. Chemical shifts (δ) are given in ppm relative to the chloroform residual solvent peak or tetramethylsilane as internal standard. Coupling constants are given in Hz. All given ¹³C spectra are proton decoupled. High resolution mass spectra were recorded on a LTQ-Orbitrap (Thermo Finnigan) Mass spectrometer. LC/MS analysis was performed on a Jasco HPLC-system (detection simultaneous at 214 nm and 245 nm) equipped with an analytical Alltima C_{18} column (Alltech, 4.6 mmD x 50 mmL, 3μ particle size) in combination with buffers A: H₂O, B: MeCN and C: 0.5% aq. TFA and coupled to a Perkin Almer Sciex API 165 mass spectrometer. Optical rotations were measured on a Propol automatic polarimeter. IR spectra were recorded on a Shimadzu FTIR-8300 and are reported in cm⁻¹.

1,6-Anhydro-2-deoxy-2-iodo- β -D-glucopyranose (150):



A solution of the commercial available tri-*O*-acetyl-D-glucal (2.72 g, 10 mmol) was dissolved in $MeOH:H_2O:Et_3N$ (10:10:1, 125 mL) was stirred for 1 h at ambient temperature, then concentrated. The residue was dried by coevaporation with dioxane (3x 50 mL). The clear oil was use further without any purifications. Crude deprotected D-glucal (1.46 g, 10 mmol) was

dissolved in 100 mL MeCN. The solution was boiled under reflux with 4.08 mL bis(tributyl stannyl) oxide (4.77g, 8 mmol) and molsieves 4Å for 2.5 h. Subsequently the reaction was cooled to 0 °C, followed by portion wise addition of 3.8 g I₂ (15 mmol, 1.5 equiv). The dark brown mixture was stirred overnight at 4 °C. TLC showed complete conversion of D-glucal into **150**. The mixture was filtered through Celite and concentrated. To the residue were added Na₂S₂O₃ (50 mL) and PE (50 mL), and the biphasic mixture was vigorously stirred for several h until the mixture became colorless. The aqueous phase was than washed several times with EtOAc(4x 40 mL). The combined organic layers were dried (Na₂SO₄) and concentrated *in vacuo*. The crude product was used without further purification in the next step.

1,6:2,3-Bis-anhydro- β -D-mannopyranose (151):



A heterogeneous solution of compound **150** (2.71 g, 10 mmol) and NaHCO₃ (2.5 g, 25 mmol, 2.5 equiv) in DMF:H₂O (10:1) 25 mL was heated to 120 °C. After 4 h, the reaction mixture was cooled, concentrated (*in vacuo*) and silica gel purification (MeOH/EtOAc 10%) yielded 1.42 g (9.85 mmol, 98% over 3 steps) of the title compound **151** as light yellow oil. TLC:

EtOAc/PE 50%; ¹H NMR (200 MHz, $CDCl_3$) δ 3.15 (d, J = 3.7 Hz, 1H, CH, C'-2), 3.25 (d, J =

8.8 Hz, 1H, CH, C'-4), 3.44 (t, J = 2.9, 1H, CH, C'-3), 3.69-3.95 (m, 2H, CH₂, C'-6), 5.70 (d, J = 2.9 Hz, CH, C'-1), ¹³C NMR (50 MHz, CDCl₃) δ 48.49 (CH, C'-2), 53.23 (CH, C'-3), 64.69 (CH₂, C'-6), 65.84 (CH, C'-4), 73.21 (CH, C'-5), 96.56 (CH, C'-1).

1,6-Anhydro-2-azido-2-deoxy- β -D-glucopyranose (152):



The bis-anhydro sugar **151** (1.15 g, 8 mmol) was heated to reflux temperature in a 10:1 MeOH:H₂O (40 mL) solution containing 5.20 g NaN₃ (80 mmol, 10 equiv), and 4.24 g NH₄Cl (80 mmol, 10 equiv). After ¹H NMR showed complete conversion to compound **152** (4.5 days), the solution was cooled, filtered through Celite and concentrated under reduced pres-

sure. Silica gel purification (EtOAc/PE 80%) yielded 0.99 g (5.32 mmol, 66.5%) of the title compound **152** as off-white solid. TLC: EtOAc/PE 50%; ¹H NMR (200 MHz, CDCl₃) δ 3.17 (s, 1H, CH, C'-2), 3.53 (s, 1H, CH, C'-4), 3.32-3.69 (m, 2H, CH, CH2, C'-3, C'-6), 4.03 (d, *J* = 7.3 Hz, 1H, CH₂, C'-6), 4.46 (d, *J* = 4.3 Hz, 1H, CH, C'-5), 5.36 (s, 1H, CH, C'-1); ¹³C NMR (50 MHz, CDCl₃) δ 64.07 (CH, C'-2), 66.55 (CH₂, C'-6), 72.92 (CH, C'-3, C'-4), 77.99 (CH, C'-5), 101.79 (CH, C'-1); ESI-MS: 209.9 (M⁺Na⁺)

1,6-Anhydro-3,4-di-O-benzyl-2-azido-2-deoxy-β-D-glucopyranose (153):



The anhydro sugar **152** (9.35 g, 50 mmol) was dissolved in DMF and cooled to 0 °C. Benzyl bromide (15 mL, 125 mmol, 2.5 equiv) was added followed by portion wise addition of NaH (6 g, 150 mmol, 3 equiv). The reaction was stirred for 3 h, allowing the mixture to warm to rT, after which it was cooled (0 °C) and quenched by addition of MeOH. The mixture was

concentrated *in vacuo* and the oily residue was taken up in Et₂O and washed with 1M HCl. The organic layer was dried, filtered and concentrated under reduced pressure. Purification by silica gel chromatography (EtOAc/PE 20%) yielded compound **153** in 65% (11.93 g, 32.5 mmol). TLC: EtOAc/PE 30%; ¹H NMR (400 MHz, $CDCl_3$) δ 7.42 - 7.22 (m, 12H), 5.51 - 5.43 (s, 1H), 4.65 - 4.44 (m, 5H), 4.03 - 3.96 (d, J = 7.3 Hz, 1H), 3.73 - 3.68 (m, 1H), 3.67 - 3.62 (s, 1H), 3.38 - 3.34 (s, 1H), 3.29 - 3.25 (s, 1H).; ¹³C NMR (100 MHz, d_4), MeOD) δ 137.50, 137.36, 128.66, 128.65, 128.16, 128.11, 128.00, 127.89, 100.65, 76.05, 74.49, 72.47, 71.44, 65.45, 60.03.

1,6-Anhydro-3-O-benzyl-2-azido-2-deoxy- β -D-glucopyranose (148):



Compound **153** (10.28 g, 28 mmol) was coevaporated thrice with Tol, after which it was dissolved in dry DCM (450 mL). Next TiCl_4 (3.2 mL, 29.4 mmol, 1.05 equiv) was carefully added, the mixture was stirred for 1.5 h at rT. The reaction mixture was than poured into cooled (0 °C) H₂O after which the layers were separated. The organic layer was washed with

NaHCO₃ and H₂O and dried using MgSO₄. Concentration and purification by silicagel chromatography (EtOAc/PE 25%) gave **148** in 75% yield (5.9 g, 21.3 mmol). TLC: EtOAc/PE 40%; ¹H NMR (400 MHz, CDCl₃) δ 7.43 - 7.26 (m, 5H), 5.47 - 5.43 (s, 1H), 4.71 - 4.57 (m, 2H), 4.56 - 4.50 (d, *J* = 5.6 Hz, 1H), 4.32 - 4.15 (d, *J* = 7.3 Hz, 1H), 3.83 - 3.72 (m, 1H), 3.70 - 3.63 (d, *J* = 7.3 Hz, 1H), 3.62 - 3.58 (m, 1H), 3.55 - 3.50 (s, 1H), 2.70 - 2.57 (d, *J* = 9.8 Hz, 1H).; ¹³C NMR (100 MHz, MeOD) δ 137.27, 128.76, 128.27, 127.84, 100.21, 78.17, 76.43, 72.66, 69.01, 65.21, 59.78.

1,6-Anhydro-2-azido-3-O-benzyl-2-deoxy-4-O-methyl- β -D-glucopyranose (154):



Compound **148** (8.15 mmol, 2.26 g) was dissolved in DMF (25 mL) and cooled using an ice-bath, after stirring for 15 minutes NaH (60% dispersion in mineral oil) (0.49 g, 12 mmol, 1.5 equiv) was added portion wise. After 30 minutes the gas development stopped and MeI (0.61 mL, 9.8 mmol 1.2 equiv) was added dropwise. After 1.5 h the reaction was

quenched with MeOH. The reaction mixture was concentrated *in vacuo* and purified using a short silica column (EtOAc/PE 20%) which gave product **154** as clear oil (91%, 2.16 g, 7.42 mmol). TLC: EtOAc/PE 40%; ¹H NMR (400 MHz, CDCl₃) δ 7.41-7.27 (m, 5H, CH_{arom} Bn), 5.48 (s, 1H, H-1, 4.64 (m, 3H, CH₂ Bn, H-5), 4.08 (d, *J* = 7.2 Hz, 1H, H-6), 3.76 (d, *J* = 6.4 Hz, 1H, H-6), 3.59 (s, 1H, H-4), 3.39 (s, 3H, CH₃), 3.29 (s, 1H, H-3), 3.20 (s, 1H, H-2); ¹³C NMR (100 MHz, CDCl₃) δ 137.4 (C_q), 128.7-127.9 (CH_{arom} Bn), 100.6 (C-1), 78.8 (C-2), 75.9 (C-3), 73.8 (C-4), 72.5 (C-6), 65.3 (CH₂ Bn), 59.9 (C-5), 57.2 (CH₃ Me); IR (neat) *v* 2096.5, 1718.5, 1244.0, 1099.3, 1004.8, 929.6, 867.9; HRMS: C₁₄H₁₇N₃O₄ + Na⁺ requires 314.1111, found 314.1113; [*a*]²_p + 21.7 °(c = 2, CHCl₃).

1,6-Di-O-acetyl-2-azido-3-O-benzyl-2-deoxy-4-O-methyl- α/β -D-glucopyranose (157):



Anhydro compound **154** (2.16 g, 7.42 mmol) was taken up in Ac_2O (37 mL) and cooled with an ice bath. To this cooled solution TFA (3.7 mL, 10% v/v) was added and the reaction was stirred overnight at room temperature. After complete conversion of the starting material the

reaction was diluted with toluene and coevaporated. The resulting oil was purified using a short silica column (EtOAc/PE 30%) yielding **157** (2.62 g, 6.66 mmol, 90%) as a white solid. TLC: EtOAc/PE 50%; ¹H NMR (400 MHz, CDCl₃) δ 7.43-7.23 (m, 5H, CH_{arom} Bn_{a/β}), 6.22 (d, *J* = 3.6 Hz, 1H, H-1_a), 5.45 (d, *J* = 8.1 Hz, 1H, H-1_β), 4.88-4.81 (m, 2H, CH₂ Bn_{a/β}), 4.30-4.21 (m, 2H, CH₂, H-6_{a/β}), 3.87-3.78 (m, 2H, H-5_{a/β} H-4_{a/β}), 3.55-3.45 (m, 4H, CH₃ Me, H-3_{a/β}), 3.30 (m, 1H, H-2_{a/β}), 2.09 (s, 3H, CH₃, OAc), 2.04 (s, 3H CH₃, OAc); ¹³C NMR (100 MHz, CDCl₃) δ 170.1 (Cq, Ac), 168.3 (Cq, Ac), 137.4 (Cq), 128.2-127.7 (CH_{arom} Bn), 92.2 (C-1_β), 90.0 (C-1_α), 82.4 (C-4_β), 79.9 (C-4_α), 79.5 (C-3_a), 79.1 (C-3_β), 75.1 (CH₂ Bn), 73.5 (C-5_β), 71.0 (C-5_α), 64.5 (CH₃ Me_β), 62.1 (C-6_{a/β}), 62.1 (CH₃ Me_α), 60.6 (C-2_α), 60.4 (C-2_β), 20.4 (CH₃ OAc), 20.3 (CH₃ OAc); IR (neat) *v* 2106.1, 1753.2, 1733.9, 1373.2, 1136.0, 1109.0, 1004.8, 935.4, 906.5, 740.6; HRMS: C₁₈H₂₃N₃O₇ + Na⁺ requires 416.1428, found 416.1427.

6-O-Acetyl-2-azido-3-O-benzyl-2-deoxy-4-O-methyl- α/β -D-glucopyranose (160):



Compound **157** (2.62 g, 6.66 mmol) was dissolved in THF (30 mL) and piperidine (1.8 mL, 6% v/v) was added. The clear solution was stirred overnight at room temperature. After complete conversion to a lower running spot on TLC the reaction was diluted with EtOAc (100 mL)

and poured in 1M HCl (100 mL). The layers were separated and the organic layer was washed twice with H₂O and once with brine. Subsequently the EtOAc layer was dried and concentrated *in vacuo*. Flash silica column purification (EtOAc/PE 20%) yielded compound **160** in 71% as a white foam (1.65 g, 4.70 mmol). TLC: EtOAc/PE 50%; ¹H NMR (400 MHz, CDCl₃) δ 7.46-7.28 (m, 8H, CH_{arom} Bn α/β), 5.26 (d, J = 3.4 Hz, 1H, H-1 $_{\alpha}$), 4.90-4.77 (m, 3H, CH₂ Bn α/β), 4.56 (d, J = 7.5 Hz, 1H, H-1 $_{\beta}$), 4.37 (m, 2H, CH₂, H-6 $_{\alpha/\beta}$), 4.07-4.00 (m, 1H, H-5 $_{\alpha}$), 3.93 (dd, J = 10.1, 8.9 Hz, 1H, H-4 $_{\alpha}$), 3.69 (s, 1H, OH), 3.55 (s, 3H, CH₃, Me $_{\alpha}$), 3.53 (s, 1H, CH₃ Me $_{\beta}$), 3.47-3.41 (m, 1H, H-4 $_{\beta}$), 3.40-3.30 (m, 3H, H-2 $_{\alpha/\beta}$, H-5 $_{\beta}$), 3.30-3.19 (m, 2H, H-3 $_{\alpha/\beta}$), 2.09 (s, J = 5.5 Hz, 6H CH₃, Ac α/β); ¹³C

NMR (100 MHz, CDCl₃) δ 171.2 (C_q, Ac), 171.1 (C_q, Ac), 137.6 (C_q), 137.6 (C_q Bn), 128.4-127.9 (CH_{arom} Bn), 95.9 (C-1_β), 91.7 (C-1_α), 82.6 (C-3,4_β), 80.4 (C-3,4_α), 79.6 (C-3,4_{α/β}), 75.4 (CH₂, Bn_{α/β}), 73.0 (C-5_β), 68.9 (C-5_α), 67.1 (C-2_β), 63.6 (C-2_α), 62.97 (C-6_{α/β}), 62.95 (C-6_{α/β}), 60.8 (OMe_{α/β}), 60.7 (OMe_{α/β}) 20.7 (CH₃ Ac_{α/β}), 20.7 (CH₃ Ac_{α/β}); IR (neat) v 2937.4, 2106.1, 1739.7, 1456.2, 1319.2, 1238.2, 1120.6, 1085.8, 1035.7, 995.2, 746.4, 698.2; HRMS: C₁₆H₂₁N₃O₆ + Na⁺ requires 374.1323, found 374.1321.

6-O-Acetyl-2-azido-3-O-benzyl-2-deoxy-4-O-methyl-1-O-(*N*-trichloroacetimidoyl) -*α*-D-glucopyranoside (145):



A solution of compound **160** (1.6 g, 4.6 mmol) and trichloroacetonitrile (1.37 mL, 13.7 mmol, 3 equiv) in dry DCM (25 mL) was treated with 0.2 equivalents of DBU (0.12 mL, 0.91 mmol) for 18 h at ambient temperature. The dark brown solution was concentrated under reduce pressure and directly purified using a silica gel column (20% EtOAc/PE and 2.5% TEA) to obtain the title compound

145 in a good yield (1.88 g, 3.80 mmol, 83%). TLC: EtOAc/PE 30% + 2.5% TEA; ¹H NMR (400 MHz, CDCl₃) δ 8.74 (s, 1H, NH), 7.45-7.29 (m, 5H, CH_{arom} Bn $_{\alpha/\beta}$), 6.37 (d, J = 3.5 Hz, 1H, H-1 $_{\alpha}$), 4.89 (d, J = 10.4 Hz, 2H, CH₂ Bn), 4.37-4.21 (m, 2H, H-6), 4.01-3.90 (m, 2H, H-4,5), 3.67-3.59 (m, 1H, H-3), 3.56 (s, J = 11.7 Hz, 3H, CH₃ Me), 3.36 (dd, J = 18.4, 8.5 Hz, 1H, H-2), 2.06 (s, J = 5.0 Hz, 3H, CH₃ OAc); ¹³C NMR (100 MHz, CDCl₃) δ 170.4 (C_q, C=N)160.6 (C_q, Ac), 137.6 (C_q), 128.6-128.1 (CH_{arom} Bn) 94.6 (C-1) 90.7 (C_q, CCl₃) 79.9 (C-3,4,5), 75.6 (CH₂ Bn), 71.8 (C-3,4,5), 62.8 (C-3,4,5), 62.4 (C-6), 61.1 (C-2), 20.8 (CH₃ Ac); IR (neat) v 2110.0, 1733.9, 1678.0, 146.2, 1373.2 1228.6, 1016.4, 986.2, 906.5, 789.5, 734.8, 696.3.

1,6-Anhydro-2-azido-3-O-benzyl-2-deoxy-4-O-(6-O-acetyl-2-azido-3-O-benzyl-2-deoxy-4-O-methyl- β -D-glucopyranosyl)- β -D-glucopyranose (142):



A mixture of imidate **145** (1.8 g, 3.6 mmol) and alcohol **148** (2.0 g, 7.3 mmol, 2 equiv) were coevaporated thrice with toluene and dissolved in dry toluene (18 mL). To this mixture activated molecular sieves (4Å) were added and the solution was cooled to -78 °C. After 10 minutes a solution of BF₃·Et₂O (90 μ L, 0.72 mmol, 0.2 equiv) in dry toluene (3 mL) was added and

the temperature was allowed to rise to -20 °C in 90 minutes. TLC analysis showed complete conversion of imidate **145** into a lower running spot. The reaction was quenched using TEA (0.5 mL), filtered and concentrated *in vacuo*. The excess of acceptor was acetylated, using an Ac₂O-pyridine cocktail (1 mL/3 mL), after which the reaction was quenched using MeOH and concentrated. The oily residue was directly purified using a silica gel column (EtOAc/PE 40%). Compound **142** was obtained in 83% in 1:3 α : β ratio (1.84 g, 3.02 mmol). TLC: EtOAc/Tol 60%; ¹H NMR (400 MHz CDCl₃) δ 7.44-7.14 (m, 10H, CH_{arom} Bn_{α/β}), 5.55 (s, 1H, H-1_{α}), 5.47 (s, 1H, H-1_{β}), 4.83 (dt, *J* = 18.1, 10.8 Hz, 2H, CH₂, Bn), 4.73-4.52 (m, 3H, CH₂, H-6, H-5'), 4.41-4.24 (m, 2H, CH₂ Bn, H-1'), 4.16 - 4.01 (m, 2H, CH₂ Bn, H-6'), 3.95 (dd, *J* = 6.4, 5.0 Hz, 1H, H-3'), 3.85-3.69 (m, 2H, CH₂ H-6', H-5), 3.58-3.52 (m, 3H, CH₃ Me), 3.46 (dt, *J* = 18.9, 9.5 Hz, 1H, H-2'), 3.34-3.12 (m, 4H, H-2/3/4/4'), 2.01 (s, 3H, CH₃ Ac_{β}); ¹³C NMR (100 MHz, CDCl₃) δ 170.7 (C_q, Ac), 137.8 (C_q), 128.7-127.8 (CH_{arom} Bn), 102.3 (C-1), 100.7 (C-1'), 82.7 (C-4), 79.3 (C-3'), 77.5 (C-5'), 76.6 (C-4'), 75.6 (CH₂ Bn), 75.5 (C-3), 74.8 (C-5), 73.4 (C-6'), 72.9 (C-2), 72.5 (C-6), 65.8 (CH₂ Bn), 61.1 (CH₃ Me), 59.3 (C-2), 20.9 (CH₃ Ac); IR (neat) v 2100.3, 1739.7, 1456.2, 1363.6, 1232.4, 1066.6, 1001.0, 1026.0, 931.6,

898.8, 738.7, 696.3; HRMS: $C_{29}H_{34}N_6O_9 + Na^+$ requires 633.2279, found 633.2279; $[\alpha]_D^{23} + 11.4$ °(c = 1, CHCl₃).

1,6-Anhydro-2-azido-3-*O*-benzyl-2-deoxy-4-*O*-(2-azido-3-*O*-benzyl-2-deoxy-4-*O*-me-thyl- β -D-glucopyranosyl)- β -D-glucopyranose (163):



Disaccharide **142** (1.84 g, 3.02 mmol) was dissolved in MeOH (15 mL) and a catalytic amount of NaOMe (30% in MeOH) was added. The reaction mixture was stirred for 1 h at ambient temperature, after which it was neutralized $(pH\sim7)$ using Amberlite[®] IR-120 H⁺ resin. Filtering off the resin, concentration and purification using a short silica col-

umn (EtOAc/PE) yielded title compound **163** as a clear oil (1.0 g, 1.8 mmol, 59%). TLC: EtOAc/PE 35%; ¹H NMR (400 MHz, CDCl₃) δ 7.48-7.30 (m, 10H, CH_{arom} 2xBn), 5.52 (s, 1H, H-1), 4.87 (q, *J* = 10.9 Hz, 2H, CH₂ Bn), 4.73 (m, 2H, CH2, H-6', H-5), 4.61 (d, *J* = 12.3 Hz, 1H, CH₂ H-6'), 4.26 (d, *J* = 8.0 Hz, 1H, H-1'), 4.20-4.10 (m, 1H, CH₂ H-6), 3.94 (d, *J* = 1.3 Hz, 1H, H-3), 3.81 (m, 1H, CH₂ H-6), 3.75 (s, 1H, H-5), 3.69 (s, 2H, CH₂ Bn), 3.58 (s, 3H, CH₃, OMe), 3.49-3.44 (m, 1H, H-2'), 3.36 (s, 1H, H-2), 3.34-3.28 (m, 2H, H-3', H-4), 3.13-3.03 (m, 1H, H-4); ¹³C NMR (100 MHz, CDCl₃) δ 137.9 (C_q), 137.3 (C_q), 128.7-127.7 (CH_{arom} 2xBn), 102.1 (C-1'), 100.3 (C-1), 82.5 (C-4), 78.9 (C-3'), 77.9 (C-5'), 75.6 (C-4', C-3), 75.5 (CH₂ Bn), 75.3 (C-5), 74.7 (C-6'), 72.2 (C-2'), 65.8 (C-6), 65.0 (CH₂ Bn), 61.4 (CH₃, OMe), 59.0 (C-2); IR (neat) *v* 2108.1, 1454.2, 1261.4, 1141.8, 1074.3, 1006.8, 740.6, 698.2; HRMS: C₂₇H₃₂N₆O₈ + Na⁺ requires 591.2174, found 591.2172; [α]²³/₂-16.67°(c = 0.6, CHCl₃).

2-Azido-1,6-di-O-acetyl-3-O-benzyl-2-deoxy-4-O-(2-azido-3-O-benzyl-2-deoxy-4-O-methyl- β -D-glucopyranosyl)- α/β -D-glucopyranose (166):



To a solution of compound **163** (1.0 g, 1.7 mmol) in Ac₂O (8.5 mL) TFA (1.28 mL, 15% v/v) was added. The reaction was stirred for 18 h after which it was diluted with toluene and concentrated. After purification using silica gel chromatograph (EtOAc/PE 30%) disaccharide **166** was obtained

in 90% yield (1.09 g, 1.53 mmol) as transparent foam. TLC: EtOAc/PE 45%; ¹H NMR (400 MHz, CDCl₃) δ 7.42-7.27 (m, 15H, CH_{arom} Bn_{a/β}), 6.20 (d, J = 3.7 Hz, 1H, H-1_a), 5.49-5.43 (m, 1H, H-1_β), 5.05 (dd, J = 33.7, 11.2 Hz, 1H, CH₂ H_{6'}), 4.89-4.59 (m, 4H, CH₂ Bn, CH₂ H-6'), 4.52-4.44 (m, 2H, CH₂ Bn), 4.39-4.28 (m, 1H, H-1'), 4.19 (m, 1H, CH₂ H-6), 4.15-4.07 (m, 1H, CH₂ H-6), 4.00-3.86 (m, 3H, H-3, H-4, H-5), 3.78 (dd, J = 7.2, 6.0 Hz, 1H),3.74-3.68 (m, 1H), 3.58-3.50 (m, 1H, H-2), 3.49 (s, 3H, CH₃, Me), 3.42-3.31 (m, 2H, H-2', H-4'), 3.31-3.19 (m, 2H, H-3', H-5'), 2.19-1.90 (m, 13H, CH₃, Ac_{a/β}); ¹³C NMR (100 MHz, CDCl₃) δ 170.7-168.9 (3x C_q, Ac), 138.3-127.5 (Bn), 101.5 (C-1'), 90.23(C-1), 83.3 (C-3'), 79.9 (C-5'), 78.6 (C-4, C-5), 77.5 (C-4, C-5), 75.9 (C-6'), 75.1 (CH₂ Bn), 73.5 (C-4), 71.1 (C-3), 66.8 (C-2'), 62.9 (C-6), 62.4 (CH₂ Bn), 62.2 (C-2), 61.0 (CH₃, OMe), 21.1-20.8 (3x CH₃ Ac); IR (neat) ν 2111.9, 1743.5, 1234.4, 1029.9, 741.6, 689.5; HRMS: C₃₃H₄₀N₆O₁₂ + Na⁺ requires 735.2596, found 735.2595.

2-Acetamido-1,3,6-tri-O-acetyl-2-deoxy4-O-(2-acetamido-2-deoxy-3,6-O-di-acetyl-4–O-methyl- β -D-glucopyranosyl)- α/β -D-glucopyranose (139):



Compound **166** (71 mg, 0.1 mmol) was taken up in a mixture of dioxane:toluene: $H_2O(5/2/1, v/v/v, 2 mL)$ and cooled with an ice-bath. After 10 minutes 1M PMe₃ (0.5 mL) in toluene was added and the reaction was stirred for 18 h at 4 °C. After TLC analysis showed conversion towards lower run-

ning, ninhydrin positive spot, the reaction was coevaporated thrice with toluene and subsequently acetylated using an Ac₂O-pyridine cocktail (0.5 mL/1.5 mL). The mixture was stirred for 20 h after which it was quenched with MeOH and concentrated under reduced pressure. The resulting white solid was dissolved in a 1:1 mixture of MeOH and TFE (4 mL) and purged with argon. A catalytic amount of Pd(OH)₂ spiked with Pd-black was added and the mixture was purged with H₂. Reduction of the benzyl-groups was continued for 5 h followed by filtration and concentration. The residue was again taken up in an Ac_2O pyridine cocktail (0.5 mL/1.5 mL) with a catalytic amount of DMAP and stirred at ambient temperature for 18 h. After complete acetylation of the disaccharide the mixture was quenched with MeOH and concentrated under reduced pressure. Silica gel purification (MeOH/DCM 3%) yielded 31 mg (47 μ mol, 48%) of the title compound **139** as an off-white solid. TLC: MeOH/DCM 5%; ¹H NMR (400 MHz, CDCl₃) δ 6.31 (d, J = 9.8 Hz, 1H, NH), 6.11 (m, 2H, NH, H-1_a), 5.74 (s, 1H), 5.61 (d, J = 7.5 Hz, 1H, H-1_b), 5.26-5.17 (m, 1H, H-1_b) 4'), 5.09-5.01 (m, 1H), 5.01-4.93 (m, 1H H-4), 4.45-4.37 (m, 1H, H-6), 4.30 (m, 5H, H-1' $_{\alpha}$, H-1'_β, H-2', H-5), 4.25-4.18 (m, 1H, H-6), 4.17-4.07 (m, 1H, H-2), 4.06-3.94 (m, 1H, H-2'), 3.93-3.75 (m, 2H), 3.45 (m, 1H, H-3'), 3.41 (d, J = 3.0 Hz, 4H, CH₃, OMe), 3.35 (d, J = 9.2Hz, 1H, H-3), 2.21-1.90 (m, 33H, CH₃ Ac and NHAc); 13 C NMR (100 MHz, CDCl₃) δ 171.4, 171.2, 170.9, 170.88, 170.81, 170.6, 170.5, 170.47, 170.4, 170.3, 170.2, 169.4, 168.9, 102.0, 100.8, 92.5, 90.5, 77.4, 77.1, 77.0, 76.7, 75.9, 75.2, 74.9, 73.9, 73.7, 73.0, 71.9, 71.0, 70.9, 70.7, 62.9, 62.8, 62.4, 61.6, 60.4, 60.4, 54.2, 54.1, 51.4, 51.2, 29.7, 23.2, 23.1, 23.0, 21.0, 20.97, 20.9, 20.88, 20.77, 20.70, 20.6; IR (neat) v 3282.6, 1741.6, 1662.5, 1544.9, 1434.9, 1373.2, 1226.6, 1112.9, 1033.8, 943.1, 732.9; HRMS: C₂₇H₄₀N₂O₁₆ + H⁺ requires 649.2451, found 649.2452.

2-Acetamido-1,3,6-tri-*O*-acetyl-2-deoxy-4-*O*-(2-acetamido-2-deoxy-3,6-*O*-di-acetyl-4--*O*-methyl- β -D-glucopyranosyl)-1-*O*-4-methylumbelliferyl- β -D-glucopyranoside (169).



Dimer **139** (100 mg, 154 μ mol) was dissolved in AcOH (2 mL) and Ac₂O (1 mL). At 0 °C dry HCl_(g) was bubbled through (liberated under Kipp conditions) for 3h. The reaction mixture was then placed at 5 °C for 42 h at which TLC analyses (DCM-acetone 60-40) showed com-

plete consumption of starting material. The reaction was diluted with CHCl₃ (10 mL, 0 °C) and washed twice with H₂O (15 mL, 0 °C) and twice with NaHCO₃ (15 ml, 0 °C). The organic layer was dried over MgSO₄ and concentrated *in vacuo* yielding the anomeric *a*-chloride as an amorphous solid of which purity was evaluated by ¹H-NMR. The resulting solid was dissolved in CHCl₃ (5 mL) and added to a solution of NaHCO₃ 0.2M (5 mL), 4-methylumbelliferyl sodium salt^{34,35} (152mg, 770 μ mol) and TBAHS (105 mg, 310 μ mol). The biphasic mixture was stirred overnight with the exclusion of light. The phases were separated and the organic layer was washed twice with NaHCO₃ (0.2 M) and twice with

 H_2O . The organic layer was dried over MgSO₄ and concentrated *in vacuo*. Purification by column chromatography (MeOH/CHCl₃ 3%) yielded 169 (27 mg, 25%) as a white amorphous solid. TLC: MeOH/DCM 5%; ¹H NMR (600 MHz, (d_4), MeOD) δ 8.06 (d, J = 9.2 Hz, 1H, NH), 7.62 (d, J = 8.7 Hz, 1H, CH, 4-methylumbelliferyl), 6.99-6.94 (m, 2H, CH_{arom} 4methylumbelliferyl), 6.19 (s, J = 9.6 Hz, 1H, CH_{arom} 4-methylumbelliferyl), 5.27 (d, J = 8.710.2 Hz, 1H, H-6'), 4.33 (dt, J = 20.2, 7.2 Hz, 1H, CH₂, H-6'), 4.23-4.17 (m, 1H, H-2), 4.14 (dd, J = 11.8, 7.3 Hz, 2H, CH₂ H-6), 3.95 (t, J = 7.7 Hz, 1H, H-5), 3.86 (t, J = 9.2 Hz, 1H, H-4), 3.79- $3.72 \text{ (m, 1H, H-2')}, 3.59 \text{ (dd, } J = 8.6, 2.9 \text{ Hz}, 1\text{H}, \text{H-5')}, 3.45-3.40 \text{ (m, 3H, CH}_3 \text{ Me)}, 3.37 \text{ (dd, J)}$ J = 12.4, 6.4 Hz, 1H, H-4'), 2.44 (s, 3H, CH₃, 4-methylumbelliferyl), 2.17-2.05 (m, 12H, CH₃) 4xAc), 1.97-1.92 (m, 6H, CH₃ 2xAc); ¹³C NMR (151 MHz, (d₄), MeOD)δ 173.3-171.6 (C_q, Ac), 162.9-154.9 (Cq, 4-methylumbelliferyl), 127.01 (Carom, 4-methylumbelliferyl) 116.2 (Cq, 4-methylumbelliferyl),115.1-104.6 (Carom, 4-methylumbelliferyl), 101.9 (C-1'), 98.9 (C-1), 77.4 (C-4'), 75.9 (C-4), 74.1 (C-3), 74.1 (C-5), 73.7 (C-5'), 64.0 (C-6'), 63.7 (C-6), 60.8 (CH₃) Me), 55.9 (C-2'), 54.9 (C-2), 23.1-19.0 (CH₃ Ac); IR (neat) v 1739.7, 1660.6, 1612.4, 1612.4, 1371.3, 1222.8, 1066.6, 1033.8, 623.0; HRMS: C₃₅H₄₄N₂O₁₇ + H⁺ requires 765.2713, found 765.2717; $[\alpha]_{p}^{23}$ -32.4 °(c = 0.5, CHCl₃/MeOH).

2-Acetamido-2-deoxy-4-O-(2-acetamido-2-deoxy-4-O-methyl- β -D-glucopyranosyl)-1--O-4-methylumbelliferyl- β -D-glucopyranoside (136).



To a suspension of **169** (12mg, 15.70 μ mol) in MeOH (1 mL) was added NaOMe (30 wt% in MeOH) (173 μ L, 0.40 μ mol). The reaction was stirred with the exclusion of light. When LCMS (gradient 0 to 50% MeOH) showed complete conversion to product the mixture

was quenched with AcOH (5 μL, 80 μmol). The reaction was diluted with H₂O (2 ml) the MeOH was evaporated *in vacuo* and the remaining H₂O was lyophilized. Purification by HPLC (gradient H₂O-MeOH + 0.1% TFA 80-20 → 60-40) evaporation of MeOH and lyophilizing H₂O yielded **136** (3.5 mg, 5.9 μmol, 37%) as white fluffy solid. ¹H NMR (600 MHz, (*d*₆), DMSO) δ 7.90 (d, *J* = 9.0 Hz, 1H, NH), 7.75 (d, *J* = 9.1 Hz, 1H, NH), 7.69 (d, *J* = 8.8 Hz, 1H, CH, 4-methylumbelliferyl), 7.03 (d, *J* = 2.2 Hz, 1H, CH, 4-methylumbelliferyl), 6.95 (dd, *J* = 8.8, 2.3 Hz, 1H, CH, 4-methylumbelliferyl), 6.26 (s, 1H, CH, 4-methylumbelliferyl), 5.16 (dd, *J* = 10.8, 7.4 Hz, 2H, H-1, OH), 4.83 (t, *J* = 5.3 Hz, 1H, OH), 4.74 (d, *J* = 2.1 Hz, 1H, OH), 4.68 (t, *J* = 5.9 Hz, 1H, OH), 4.36 (d, *J* = 8.5 Hz, 1H, H-1'_β), 3.77 (d, *J* = 9.6 Hz, 1H), 3.70-3.13 (m, 14H), 2.94 (t, *J* = 9.2 Hz, 1H), 2.40 (s, 3H, CH₃ 4-methylumbelliferyl), 1.84 (s, 3H, CH₃ Ac), 1.80 (s, 3H, CH₃ Ac); ¹³C NMR (151 MHz, (*d*₆), DMSO) δ 169.2, 160.1, 159.9, 154.4, 153.3, 126.5, 114.3, 113.4, 103.1, 101.9, 98.2, 80.7, 80.0, 75.6, 75.1, 73.8, 72.2, 59.8, 55.7, 54.4, 23.0, 18.1; HRMS: C₂₇H₃₆N₂O₁₃ + H⁺ requires 597.2290, found 597.2283.

1,6-Anhydro-2-azido-3-O-benzyl-2-deoxy-4-O-isopropyl-β-D-glucopyranose (155):



Compound **148** (1.5 g, 5.3 mmol) was dissolved in 25 mL DMF and cooled using an ice-bath and after stirring for 5 minutes NaH (60% dispersion in mineral oil) (234 mg, 5.86 mmol, 1.1 equiv) was added. After 30 minutes the gas formation stopped and 2-iodopropane (586 μ L, 5.86 mmol, 1.1 equiv) was added dropwise. After 3 h the reaction was quenched with

MeOH. The mixture was concentrated and the residue was taken up in Et₂O and washed twice with 1M HCl_(*aq*). The organic layer was dried with MgSO₄ and concentrated *in vacuo*. The oily residue was re-dissolved in 25 mL DMF and cooled using an ice-bath. The alkylation step was repeated three times. After four cycles the oil compound was purified using a short silica column (EtOAc/PE 20%) which gave product **155** as yellow oil (1.0 g, 3.1 mmol, 60%). TLC: EtOAc/PE 40%; ¹H NMR (400 MHz, CDCl₃) δ 7.31 (s, 5H, CH_{arom} Bn), 5.43 (s, 1H, H-1), 4.64-4.44 (m, 3H, CH₂ Bn,H-5), 4.02 (d, *J* = 6.7 Hz, 1H, H-6), 3.67 (s, 1H, H-6), 3.57 (dd, *J* = 12.9, 9.5 Hz, 2H, CH, *i*Prop, H-3), 3.33 (s, 1H, H-2), 3.16 (s, 1H, H-4), 1.14 (d, *J* = 5.3 Hz, 6H, 2xCH₃ *i*Prop); ¹³C NMR (100 MHz, CDCl₃) δ 137.2 (C_q), 128.2-127.5 (CH_{arom} Bn), 100.3 (C-1), 76.8 (C-3), 75.1 (C-5), 74.2 (C-4), 72.1 (CH₂ Bn), 70.1 (*i*Prop), 65.1 (C-6), 59.4 (C-2), 22.1 (CH₃ *i*Prop), 21.9 (CH₃ *i*Prop); IR (neat) v 2970.2, 2898.8, 2098.4, 1454.2, 1369.4, 1070.4, 1004.8, 964.3, 867.9, 736.8; HRMS: C₁₆H₂₁N₃O₄ + Na⁺ requires 342.1424, found 342.1425; $[\alpha]_{c3}^{2}$ 28.33 °(c = 0.6, CHCl₃).

1,6-Di-O-acetyl-2-azido-3-O-benzyl-2-deoxy-4-O-isopropyl- α/β -D-glucopyranose (158):

The yellow oil **155** (1.0 g, 3.1 mmol) was taken up in 20 mL Ac_2O and cooled with an ice bath. To this cooled solution 2 mL TFA (10% v/v) was added and the reaction was stirred overnight at room temperature. After complete conversion of the starting material the reaction

was diluted with toluene and coevaporated to yellow oil. Purification using a short silica column (EtOAc/PE 30%) yielded compound **158** in a good yield 86% (1.13 g, 2.69 mmol). TLC: EtOAc/PE 50%; ¹H NMR (400 MHz, CDCl₃) δ 7.43-7.28 (m, 6H, CH_{arom} Bn), 6.22 (d, J = 3.7 Hz, 1H, H-1 $_{\alpha}$), 5.47 (d, J = 8.4 Hz, 1H, H-1 $_{\beta}$), 4.90-4.80 (m, 2H, CH₂ Bn), 4.34-4.16 (m, 2H, CH-2, H-6), 3.98-3.90 (m, 1H, CH, *i*Prop), 3.87-3.81 (m, 2H, H-3, H-5), 3.56 (m, 2H, H-4, H-2), 2.19-2.13 (m, 4H, CH₃ Ac), 2.07 (s, 4H, CH₃ Ac), 1.21 (t, J = 5.4 Hz, 3H, CH₃ *i*Prop), 1.12 (t, J = 6.3 Hz, 3H, CH₃ *i*Prop); ¹³C NMR (100 MHz, CDCl₃) δ 170.7 (C_q Ac), 168.9 (C_q Ac), 137.6 (C_q), 128.6-128.1 (CH_{arom} Bn), 92.9 (C-1 $_{\beta}$), 90.5 (C-1 $_{\alpha}$), 83.3 (C-3/5 $_{\beta}$), 80.8 (C-3/5 $_{\alpha}$), 75.9 (CH₂ Bn), 74.4 (C-4 $_{\beta}$), 74.2 (C-4 $_{\alpha}$), 73.3 (C-3/5 $_{\beta}$), 73.3 (C-3/5 $_{\alpha}$), 71.7 (*i*Prop), 65.3 (C-2 $_{\beta}$), 63.0 (C-2 $_{\alpha}$), 62.5 (C-6), 23.2 (Ac), 22.0 (Ac), 21.1 (*i*Prop), 20.9 (*i*Prop); IR (neat) v 3423.4, 2358.8, 2343.4, 2102.3, 1745., 1456.2, 1371.3, 1234.4, 1143.7, 1124.4, 1026.1, 968.2, 997.1, 948.9, 740.6, 694.3; HRMS: C₂₀H₂₇N₃O₇ + Na⁺ requires 444.1741, found 444.1740.

6-O-Acetyl-2-azido-3-O-benzyl-2-deoxy-4-O-isopropyl- α/β -D-glucopyranose (161):



Compound **158** (1.13 g, 2.69 mmol) was dissolved in 20 mL THF and 1.5 mL piperidine (6% v/v) was added. The clear solution was stirred overnight at room temperature. After complete conversion to a lower running spot on TLC the reaction was diluted with 100 mL EtOAc and poured in 100 mL 1M HCl. The layers were separated and the or-

ganic layer was washed twice with H_2O and once with brine. Subsequently the EtOAc layer was dried and concentrated *in vacuo*. Flash silica column purification (EtOAc/PE

30%) yielded compound **161** in quantitative yield (1.02 g, 2.69 mmol) as a white foam. TLC: EtOAc/PE 40%; ¹H NMR (400 MHz, CDCl₃) δ 7.43-7.26 (m, 10H, CH_{arom} Bn), 5.26 (d, J = 2.9 Hz, 1H, H-1_{α}), 4.90-4.75 (m, 4H, CH₂ Bn), 4.58-4.53 (m, 1H, H-1_{β}), 4.42-4.33 (m, 3H, CH₂ H-6), 4.14 (m, 1H, CH₂ H-6), 4.03 (m, 2H, H-5), 3.98-3.87 (m, 2H, H-3), 3.51-3.39 (m, 4H, H-4, CH, *i*Prop.), 3.36 (m, 2H, H-2), 2.08 (m, 6H, CH₃ Ac), 1.23-1.16 (m, 6H, CH₃ *i*Prop), 1.13 (m, 6H, CH₃ *i*Prop); ¹³C NMR (100 MHz, CDCl₃) δ 171.3 (C_q Ac), 137.7 (C_q), 137.64 (C_q), 128.5-127.9 (CH_{arom} Bn), 96.2 (C-1_{β}), 91.8 (C-1_{α}), 83.2 (CH_{β} *i*Prop), 80.3 (CH_{α} *i*Prop), 75.8 (CH₂ Bn), 75.7 (CH₂ Bn), 75.1 (C-4_{α}, 74.5 (C-4_{β}), 73.4 (C-3_{β}), 73.2 (C-3_{α}), 69.2 (C-5_{β}), 67.6 (C-2_{β}), 64.1 (C-2_{α}), 63.0 (C-6_{α/β}), 62.9 (C-6_{α/β}), 23.1 (CH₃ Ac_{α/β}), 23.1 (CH₃ Ac_{α/β}), 22.0 (CH₃ *i*Prop), 20.9 (CH₃ *i*Prop); IR (neat) v 2922.0, 2108.1, 1739.7, 1363.6, 1224.7, 1026.1, 1010.6, 966.3, 914.2, 740.6, 696.3; HRMS: C₁₈H₂₅N₃O₆ + Na⁺ requires 402.1636, found 402.1634.

6-O-Acetyl-2-azido-3-O-benzyl-2-deoxy-4-O-isopropyl-1-O-(N-trichloroacetimidoyl) -α-D-glucopyranoside (146):



A solution of compound **161** (1.02 g, 2.69 mmol) and trichloroacetonitrile (0.809 mL, 8.07 mmol, 3 equiv) in dry 15 mL DCM was treated with 1,8-diazabicyclo[5.4.0]undec-7-ene 83 μ L (0.54 mmol, 0.2 equiv) for 18 h at ambient temperature. The dark brown solution was concentrated under reduce pressure and directly purified using a silica gel column (EtOAc/PE 12.5% + 2.5% TEA) to obtain

the title compound **146** as a yellow oil (0.986 g, 1.88 mmol, 70%). TLC: EtOAc/PE 30%; ¹H NMR (400 MHz, (d_4), MeOD) δ 8.77 (s, 1H, NH), 7.42-7.25 (m, 5H, CH_{arom} Bn), 6.40 (d, J = 3.5 Hz, 1H, H-1_a), 4.89 (q, J = 10.6 Hz, 2H, CH₂ Bn), 4.34 (dd, J = 12.1, 2.1 Hz, 1H, H-5, 4.18 (dd, J = 12.1, 4.4 Hz, 1H, H-6), 3.97 (m, 1H, CH, *i*Prop), 3.92 (m, 2H, H-3, H-4), 3.63 (dd, J = 10.2, 3.6 Hz, 1H, H-6), 3.60-3.53 (m, 1H, H-2), 2.03 (s, 3H, CH₃ Ac), 1.21 (d, J = 6.1 Hz, 3H, CH₃ *i*Prop), 1.13 (d, J = 6.1 Hz, 3H, CH₃ *i*Prop); ¹³C NMR (100 MHz, (d_4), MeOD) δ 170.3 (C_q, C=N), 160.4 (C_q, Ac), 137.4 (C_q), 128.3-127.8 (CH_{arom} Bn), 94.4 (C-1), 90.7 (CCl₃), 80.2 (*i*Prop), 75.6 (C_q, CH₂ Bn), 74.5 (C-4), 73.2 (C-3), 72.0 (C-5), 63.0 (C-2), 62.2 (C-6), 22.9 (CH₃ *i*Prop), 21.9 (CH₃ *i*Prop), 20.6 (CH₃ Ac); IR (neat) v 2110.0, 1741.6, 1674.1, 1234.4, 1139.9, 1018.3, 966.3, 908.4, 794.6, 729.0, 644.2.

1,6-Anhydro-2-azido-3-O-benzyl-2-deoxy-4-O-(6-O-acetyl-2-azido-3-O-benzyl-2-deoxy-4-O-isopropyl- β -D-glucopyranosyl)- β -D-glucopyranose (143):



A mixture of imidate **146** (761 mg, 1.45 mmol) and alcohol **148** (803 mg, 2.9 mmol, 2 equiv) was coevaporated thrice with toluene and dissolved in 7.25 mL of dry toluene. To this mixture activated molecular sieves (4Å) were added and the solution was cooled to -78 °C. After 10 minutes a solution of BF₃ · Et₂O (37 μ L, 0.3 mmol, 0.2 equiv) in dry toluene (0.7 mL) was added and the

temperature was allowed to rise to -20 °C in 90 minutes. TLC analysis showed complete conversion of imidate **146** into a lower running spot. The reaction was quenched using TEA (0.5 mL), filtered and concentrated *in vacuo*. The excess of acceptor was acetylated, using an Ac₂O-pyridine cocktail (1 mL/3 mL), after which the reaction was quenched using MeOH and concentrated. The oily residue was directly purified using a silica gel column (EtOAc/PE 40%). Compound **143** was obtained in 81% in 1:5 α : β ratio (755 mg, 1.18 mmol). TLC: EtOAc/Tol 40%; ¹H NMR (400 MHz, CDCl₃) δ 7.39-7.12 (m, 12H, CH_{arom} Bn),

5.52 (s, 1H, H-1_{*a*}), 5.49 (s, 1H, H-1_{*β*}), 4.87-4.75 (m, 2H, CH₂ Bn), 4.69 (t, J = 5.7 Hz, 1H, H-5_{*β*}), 4.63 (dd, J = 14.8, 7.5 Hz, 2H, CH₂ Bn), 4.40-4.32 (m, 2H, CH₂, H-6'_{*β*}, H-1_{*β*}), 4.12 (d, J = 7.1 Hz, 2H, CH₂, H-6_{*β*}), 4.09-4.02 (m, 1H, CH₂, H-6'_{*β*}), 4.00-3.86 (m, 2H, H-4'_{*β*}, H-3_{*β*}), 3.77 (m, 2H, CH₂, H-6_{*β*}, H-4_{*β*}), 3.50-3.37 (m, 2H, H-2'_{*β*}, H-5'_{*β*}), 3.33-3.15 (m, 3H, H-3'_{*β*}, H_{*β*}, CH, *i*Prop), 2.05 (m, 3H, CH₃ Ac), 1.99 (s, 3H, CH₃ Ac), 1.18 (m, 3H, CH₃ *i*Prop), 1.14-1.07 (m, 3H, CH₃ *i*Prop); ¹³C NMR (100 MHz, CDCl₃) δ 170.7 (Ac), 137.8 (C_q), 128.7-127.7 (CH_{arom} Bn), 102.4 (C-1'), 100.7 (C-1), 83.2 (*i*Prop), 77.3 (C-4) 76.9 (C-4'), 75.9 (CH₂ Bn), 74.8 (C-5), 74.2 (C-5'), 73.7 (C-3'), 73.3 (C-3), 72.5 (CH₂ Bn), 66.1 (C-2'), 65.1 (C-6), 62.6 (C-6'), 59.3 (C-2), 23.2 (CH₃ *i*Prop), 22.1 (CH₃ *i*Prop), 20.8 (CH₃ Ac); IR (neat) *v* 2102.3, 1735.8, 1238.2, 1068.5, 1028.0, 906.5, 727.1, 646.1, 624.9; HRMS: C₃₁H₃₈N₆O₉+Na⁺ requires 661.2592; found 661.2592; [*a*]₂²³ 19.4 °(c = 1, CHCl₄).

1,6-Anhydro-2-azido-3-O-benzyl-2-deoxy-4-O-(2-azido-3-O-benzyl-2-deoxy-4-O-isopropyl- β -D-glucopyranosyl)- β -D-glucopyranose (164):



Disaccharide **143** (755 mg, 1.18 mmol) was dissolved in MeOH (7 mL) and a catalytic amount of NaOMe (30% in MeOH) was added. The reaction mixture was stirred for 1 h at ambient temperature, after which it was neutralized $(pH\sim7)$ using Amberlite[®] IR-120 H⁺ resin. Filtering off the resin, concentration and purification using a short silica col-

umn (EtOAc/PE 25%) yielded title compound **164** as a clear oil (415 mg, 0.70 mmol, 59%). TLC: EtOAc/Tol 40%; ¹H NMR (400 MHz, CDCl₃) δ 7.40-7.26 (m, 10H, CH_{arom} Bn), 5.48 (s, 1H, H-1' $_{\beta}$), 4.86-4.76 (m, 2H, CH₂ Bn), 4.73-4.66 (m, 2H, CH₂ Bn), 4.57 (d, *J* = 12.3 Hz, 1H, H-5), 4.26 (d, *J* = 8.0 Hz, 1H, H-1), 4.13 (d, *J* = 7.3 Hz, 1H, H-6), 3.96-3.90 (m, 1H, H-3), 3.89 (s, 1H, H-4'), 3.81-3.75 (m, 1H, H-4), 3.72 (s, 1H, H-6), 3.64 (d, *J* = 2.9 Hz, 2H, CH₂, H6'), 3.47-3.38 (m, 2H, H₂, H_{5'}), 3.31 (s, 1H, CH, *i*Prop), 3.28-3.21 (m, 1H, H-3'), 3.06 (dt, *J* = 9.6, 3.2 Hz, 1H, H-2'), 1.28-1.23 (m, 6H, 2x CH₃ *i*Prop); ¹³C NMR (100 MHz, CDCl₃) δ 137.9 (Cq), 137.3 (Cq), 128.8-127.8 (CH_{arom} Bn), 102.3 (C-1'), 100.4 (C-1), 83.0 *(i*Prop), 78.0 (C-4), 75.9 (C-4', C-5), 75.8 (CH₂ Bn), 74.8 (C-5), 74.2 (C-3), 73.3 (C-3'), 72.4 (CH₂ Bn), 66.3 (C-2'), 65.2 (C-6), 61.4 (C-6), 59.1 (C-2), 23.0 (CH₃ *i*Prop), 22.4 (CH₃ *i*Prop); IR (neat) ν 2972.1, 212.3, 1454.2, 1259.4, 1070.4, 1026.1, 1004.8, 964.3, 896.8, 867.9, 696.3; HRMS: C₂₉H₃₆N₆O₈ + Na⁺ requires 619.2487, found 619.2484; [α]₂₃²³ -10.8 °(c = 0.5, CHCl₃).

2-Azido-3-O-benzyl-2-deoxy-1,6-di-O-acetyl-4-O-(2-azido-3-O-benzyl-2-deoxy-4-O-isopropyl- β -D-glucopyranosyl)- α/β -D-glucopyranose (167):



To a solution of compound **164** (0.95 mg, 160 μ mol) in Ac₂O (1.5 mL) was treated TFA (0.30 mL, 15% v/v). The reaction was stirred for 18 h after which it was diluted with toluene and concentrated. After purification using silica gel chromatograph (EtOAc/PE 30%) disaccharide

167 was obtained in 96% yield (92 mg, 154 μ mol) as transparent foam. TLC: EtOAc/PE 60%; ¹H NMR (400 MHz, CDCl₃) δ 7.40-7.24 (m, 15H, CH_{arom} Bn_{α/β}), 6.19 (d, J = 3.7 Hz, 1H, H-1_{α}), 5.45 (d, J = 8.1, Hz, 1H, H-1_{β}), 5.14-4.98 (m, 1H, CH₂ H-6'), 4.90-4.70 (m, 3H, CH₂ Bn, H-6'), 4.54-4.35 (m, 2H, CH₂ Bn), 4.35-4.29 (m, 1H, H-1'), 4.26-4.19 (m, 1H, CH₂ H-6), 4.10-4.01 (m, 1H, CH₂ H-6), 3.95 (dd, J = 9.8, 5.0 Hz, 1H, H-5), 3.94-3.84 (m, 2H, H-3, H-4), 3.58-3.51 (m, 1H, H-2), 3.47-3.26 (m, 5H, CH, *i*Prop, H-2'/3'/4'/5'), 2.19-2.14

(m, 3H, CH₃ Ac), 2.11 (s, 3H, CH₃ Ac), 1.88 (s, 3H, CH₃ Ac), 1.17 (d, J = 6.1 Hz, 3H, CH₃ *i*Prop), 1.09 (d, J = 6.1 Hz, 3H, CH₃ *i*Prop); ¹³C NMR (100 MHz, CDCl₃) δ 170.6 (C_q, Ac), 170.5 (C_q, Ac), 168.8 (C_q, Ac), 138.2 (C_q), 137.6 (C_q), 128.6-127.5 (CH_{arom} Bn), 101.5 (C-1'), 90.2 (C-1), 83.6 (C-3'), 78.4 (C-4/5), 77.1 (C-4/5), 76.0 (C-6'), 75.1 (CH₂ Bn), 74.3 (C-5'), 73.7 (C-4'), 73.1 (*i*Prop), 71.1 (C-3), 67.1 (C-2'), 62.8 (C-6), 62.3 (C-2), 62.2 (CH₂ Bn), 23.1 (CH₃ *i*Prop), 22.1 (CH₃ *i*Prop), 21.0 (CH₃ Ac), 20.9 (CH₃ Ac); IR (neat) v 2110.0, 1737.7, 1363.6, 1222.8, 1026.1, 1008.7, 931.6, 914.2, 82.5, 696.3; HRMS: C₃₅H₄₄N₆O₁₂+Na⁺ requires 763.2909, found 763.2913.

2-Acetamido-2-deoxy-1,3,6-tri-O-acetyl-4-O-(2-acetamido-2-deoxy-3,6-O-di-acetyl-4--O-isopropyl- β -D-glucopyranosyl)- α/β -D-glucopyranose (140):



Compound **167** (90 mg, 0.15 mmol) was taken up in a mixture of dioxane/toluene/ H_2O (5/2/1, v/v/v, 3 mL) and cooled with an ice-bath. After 10 minutes 1M PMe₃ (0.7 mL) intoluene was added and the reaction was stirred for 18 h at +4 °C. After TLC analysis showed con-

version towards lower running, ninhydrin positive spot, the reaction was coevaporated thrice with toluene and subsequently acetylated using an Ac₂O-pyridine cocktail (1 mL/3 mL). The mixture was stirred for 20 h after which it was quench with MeOH and concentrated under reduced pressure. The resulting white solid was dissolved in a 1:1 mixture of MeOH and TFE (6 mL) and purged with argon. A catalytic amount of Pd(OH)₂ spiked with Pd-black was added and the mixture was purged with H₂. Reduction of the benzyl-groups was continued for 5 h followed by filtration and concentration. The residue was again taken up in an Ac₂O-pyridine cocktail (0.5 mL/1.5 mL) with a catalytic amount of DMAP and stirred at ambient temperature for 18 h. After complete acetylation of the disaccharide the mixture was quenched with MeOH and concentrated under reduced pressure. Silica gel purification (MeOh/DCM 3%) yielded 37 mg (55 μ mol, 36%, 71% per step) of the title compound **140** as an off-white solid. TLC: MeOH/DCM 5%; ¹H NMR (400 MHz, (d_4) , MeOD) δ 7.83-7.61 (m, 2H, NH), 6.04 (d, J = 3.6 Hz, 1H, $H_{1'\alpha}$), 5.65 (d, J = 8.6 Hz, 1H, $H_{1'\beta}$), 5.24 (dd, J = 10.9, 8.9 Hz, 1H), 5.13 (dd, J = 9.9, 8.4 Hz, 1H), 5.04 (dd, J = 10.5, 8.2 Hz, 1H), 4.61-4.54 (m, 2H), 4.47-4.27 (m, 4H), 4.28-4.07 (m, 4H), 3.97 (m, 1H), 3.83 (m, 4H), 3.73-3.65 (m, 2H), 3.57-3.48 (m, 3H), 3.37-3.32 (m, 2H), 2.20 (s, 3H, CH₃), 2.17-2.01 (m, 3H, CH₃), 1.95-1.90 (m, 9H, CH₃), 1.10 (dd, *J* = 6.1, 2.1 Hz, 8H, CH₃ iProp); ¹³C NMR (100 MHz, (*d*₄), MeOD) δ 171.75, 171.61, 171.04, 170.82, 170.53, 170.47, 170.15, 169.32, 169.26, 100.67, 100.29, 91.70, 90.05, 75.26, 74.98, 74.78, 74.65, 73.27, 73.20, 73.14, 72.79, 72.56, 70.71, 70.15, 62.60, 62.12, 61.83, 54.16, 52.20, 50.46, 48.73, 48.52, 48.30, 48.09, 47.88, 47.66, 47.45, 22.17, 21.93, 21.61, 21.51, 20.04, 19.98; IR (neat) v 1733.9, 1652.9, 1558.4, 1396.4, 1224.7, 1031.8, 1031.8, 908.4, 727.1, 646.1; HRMS: C₂₉₆H₄₄N₂O₁₆ + Na⁺ requires 699.2583, found 699.2582.

2-Acetamido-1,3,6-tri-O-acetyl-2-deoxy-4-O-(2-acetamido-2-deoxy-3,6-O-di-acetyl -4- O-isopropyl- β -D-glucopyranosyl)-1-O-4-methylumbelliferyl- β -D-glucopy-ranoside

(170).



Dimer **140** (30 mg, 44 μ mol) was dissolved in AcOH (2 ml) and Ac₂O (1 ml). At 0 °C dry HCl_(g) was bubbled through (liberated under Kipp conditions) for 3h. The reaction mixture was then placed at 5 °C for 42 h at which TLC analyses (DCM-acetone

60-40) showed complete consumption of starting material. The reaction diluted with CHCl₃ (10 ml, 0 °C) and washed twice with H₂O (15 ml, 0 °C) and twice with NaHCO₃ (15 ml, 0 °C). The organic layer was dried over MgSO₄ and concentrated *in vacuo* yielding an amorphous solid of which purity was evaluated by ¹H-NMR. The resulting solid α -chloride was dissolved in CHCl₃ (2 ml) and added to a solution of NaHCO₃ 0.2M (1 ml), 4-methylumbelliferyl sodium salt34,35 (43mg, 220 µmol, 5 equiv) and TBAHS (29 mg, 88 μ mol, 2 equiv). The biphasic mixture was stirred overnight with the exclusion of light. The phases were separated and the organic layer was washed twice with $NaHCO_3$ (0.2 M) and twice with H₂O. The organic layer was dried over MgSO₄ and concentrated in vacuo. Purification by column chromatography (MeOH/CHCl₃ 3%)yielded 170 (6 mg, 17 %) as a white amorphous solid. TLC: MeOH/DCM 5%; ¹H NMR (400 MHz, (d_4), MeOD) δ 7.62 (s, 1H, CH, 4-methylumbelliferyl), 7.01-6.95 (m, 2H, CH_{arom} 4-methylumbelliferyl), 6.21 (s, 1H, CH_{arom} 4-methylumbelliferyl), 5.25 (t, J = 9.3 Hz, 2H, H-1_{1 β}, H-3), 5.14-5.00 (m, 1H, H-3'), 4.61 (d, J = 8.4 Hz, 1H, H-1'_{β}), 4.49 (dd, J = 11.2, 9.5 Hz, 1H, CH₂ H-6'), 4.34 (dd, *J* = 19.6, 7.1 Hz, 1H, CH₂ H-6'), 4.27-4.11 (m, 3H, CH₂ H-6, H-2), 3.96-3.90 (m, 1H, H-5), 3.83 (dd, *J* = 18.6, 9.5 Hz, 2H, H-4, H-2'), 3.71 (dt, *J* = 12.1, 6.0 Hz, 1H, CH, *i*Prop), 3.54 (t, J = 7.2 Hz, 2H, H-4', H-5'), 2.46 (s, 3H, CH₃ 4-methylumbelliferyl), 2.17-2.03 (m, 12H, CH₃ 4xAc), 1.94 (d, J = 6.4 Hz, 6H, CH₃ 2xAc), 1.10 (dt, J = 13.1, 6.5 Hz, 6H, CH₃ iProp); ¹³C NMR (100 MHz, (d_4), MeOD) δ 171.8-170.1 (C_q, 6xAc), 161.5-153.3 (C_q, 4-methylumbelliferyl), 125.5 (CH_{arom} 4-methylumbelliferyl), 114.7 (C_q, 4-methylumbelliferyl), 113.7 (CH_{arom} 4methylumbelliferyl), 111.8 (Carom, 4-methylumbelliferyl), 103.2 (C-1'), 100.4 (C-1), 97.4 (C-4'), 75.7 (C-4), 74.6- 72.6 (C-3/3'/5/5'/CH), 62.5 (C-6'), 62.3 (C-6), 54.2 (C-2'), 53.3 (C-2), 22.0-21.4 (CH₃ 4xAc), 19.8 (CH₃ iProp, Ac), 17.7 (CH₃ Ac); IR (neat) v 2933.5, 1743.5, 1643.2, 1618.2, 1369.4, 1232.4, 1070.4, 1041.5; HRMS: $C_{37}H_{48}N_2O_{17} + H^+$ requires 793.3026, found 793.3031.

2-Acetamido-2-deoxy-4-O-(2-acetamido-2-deoxy-4-O-isopropyl- β -D-glucopyranosyl)-1-O- 4-methylumbelliferyl- β -D-glucopyranoside (137).



To a suspension of **170** (6mg, 7 μ mol) in MeOH (1 mL) was added NaOMe (30 wt% in MeOH) (86 μ L, 0.4 μ mol). The reaction was stirred with the exclusion of light. When LCMS (gradient 0 to 50 % MeOH) showed complete conversion to product

the mixture was quenched with AcOH (20 μ L, 350 μ mol). The reaction was diluted with H₂O (2 mL) the MeOH was evaporated *in vacuo* and the remaining H₂O was lyophilized.

Purification by HPLC (gradient H₂O-MeOH + 0.1% TFA 80-20 \rightarrow 60-40) evaporation of MeOH and lyophilizing H₂O yielded **137** (2.6 mg, 60%) as white fluffy solid. ¹H NMR (600 MHz, (*d*₆) DMSO) δ 7.9 (d, *J* = 9.1 Hz, 1H, NH), 7.8 (d, *J* = 9.2 Hz, 1H, NH), 7.7 (d, *J* = 8.8 Hz, 1H, 4-methylumbelliferyl), 7.0 (d, *J* = 2.4 Hz, 1H, 4-methylumbelliferyl), 6.9 (dd, *J* = 8.8, 2.4 Hz, 1H, 4-methylumbelliferyl), 6.3 (d, *J* = 1.1 Hz, 1H, 4-methylumbelliferyl), 5.2 (d, *J* = 8.5 Hz, 1H, H_{1 β}), 5.1 (d, *J* = 6.5 Hz, 1H, OH), 4.8 (s, 1H, OH), 4.8 (d, *J* = 2.3 Hz, 1H, OH), 4.7 (s, 1H, OH), 4.4 (s, 1H, OH), 3.4 (m, 13H), 2.4 (d, *J* = 0.9 Hz, 3H, CH₃ 4-methylumbelliferyl), 1.9 (s, 3H, CH₃ Ac), 1.8 (s, 3H, Ac), 1.1 (d, *J* = 6.1 Hz, 3H, CH₃ *i*Prop), 1.1 (d, *J* = 6.1 Hz, 3H, CH₃ *i*Prop); ¹³C NMR (151 MHz, (*d*₆, DMSO) δ 169.3, 169.3, 160.2, 159.9, 154.5, 153.4, 126.6, 114.4, 113.5, 111.9, 103.2, 102.1, 98.3, 80.9, 76.0, 75.9, 75.1, 74.2, 72.3, 71.9, 55.8, 54.4, 23.2, 23.1, 22.3, 18.2; HRMS: C₂₉H₄₀N₂O₁₃ + Na⁺ requires 625.2603, found 625.2599.

1,6-Anhydro-2-azido-3-O-benzyl-2-deoxy-4-O-methyl-cyclohexane- β -D-glucopyranose (156):



Compound **148** (1.7 g, 6.0 mmol) was dissolved in 30 mL DMF and cooled using an ice-bath and after stirring for 5 minutes NaH (60% dispersion in mineral oil) (0.6 g, 15 mmol, 2.5 equiv) was added. After 30 minutes the gas evolvement stopped and (bromomethyl)cyclohexane (4.18 mL, 30 mmol, 5 equiv) was added drop-

wise. The reaction was quenched with MeOH after 3 h. The reaction mixture was concentrated. The residue was taken up in Et₂O and washed twice with 1M HCl_(aq). The organic layer was dried with MgSO₄ and concentrated *in vacuo*. The oily residue was re-dissolved in 25 mL DMF and cooled using an ice-bath. The alkylation step was repeated three times. After four cycles the oil compound was purified using a short silica column (EtOAc/PE 10%) which gave product **156** as yellow oil (1.7 g, 4.5 mmol, 75%). TLC: EtOAc/PE 30%; ¹H NMR (400 MHz, CDCl₃) δ 7.35 (s, J = 29.7 Hz, 5H, CH_{arom} Bn), 5.47 (s, 1H, H-1), 4.64 (q, J = 12.1 Hz, 3H, CH₂ Bn, H-5), 4.04 (d, J = 5.7 Hz, 1H, CH₂, H-6), 3.76 (s, 1H, CH₂ H-6), 3.60 (s, 1H, H-3), 3.30-3.15 (m, 4H, CH₂, MCH, H-2, H-4), 1.66 (t, J = 26.2 Hz, 3H, CH₂, CH, MCH), 1.32-1.08 (m, 2H, CH₂, MCH), 0.97-0.87 (m, 2H, CH₂, MCH); ¹³C NMR (100 MHz, CDCl₃) δ 137.5 (C_q, Bn), 128.7-127.9 (CH_{arom} Bn), 100.7 (C-1), 78.0 (C-4), 76.4 (C-3), 75.8 (CH₂, MCH), 25.9 (CH₂, MCH); IR (neat) v 2927.1, 2357.3, 2101.4, 1731.9, 1275.7, 1091.6, 710.4, 316.1; HRMS: C₂₀H₂₇N₃O₄ + Na⁺ requires 396.1894, found 396.1892; $[\alpha]_D^{23} + 3.5$ °(c = 0.4, CHCl₃).

1,6-Di-*O*-acetyl-2-azido-3-*O*-benzyl-2-deoxy-4-*O*-methyl-cyclohexane- α/β -D-glucopy-anose (159):



Oil **156** (1.50g, 4 mmol) was taken up in Ac_2O (20 mL) and cooled with an ice bath. To this cooled solution TFA (2mL, 10% v/v) was added and the reaction was stirred overnight at room temperature. After complete conversion of the starting material the reaction was diluted with toluene and coevaporated. Purifi-

cation using a short silica column (EtOAc/PE 7%) gave compound **159** in 72% yield (1.37 g, 2.88 mmol). TLC: EtOAc/PE 25%; ¹H NMR (400 MHz,CDCl₃) δ 7.46 - 7.28 (m, 6H, CH_{arom} Bn_{α/β}), 6.20 (d, *J* = 3.6 Hz, 1H, H-1_{α}), 5.45 (d, *J* = 8.2 Hz, 1H, H-1_{β}), 4.89-4.79 (m, 3H, CH₂ Bn), 4.31-4.19 (m, 3H, H-6_{α/β}), 3.90-3.80 (m, 2H, H-5_{α/β}), H-3_{α/β}, H-4_{α/β}), 3.72-3.64 (m, 1H, CH₂, MCH), 3.58-3.30 (m, 3H, H-2_{α/β}, H-4_{α/β}, H-4_{α/β}), 3.24 (dt, *J* = 14.7, 7.4 Hz, 1H, CH₂),

MCH), 2.17 (s, 1H, CH₃ OAc_β), 2.16 (s, 3H, CH₃ Ac_α), 2.08 (s, 4H, Ac_{α/β}), 1.82-0.83 (m, 16H, CH₂, MCH); ¹³C NMR (100 MHz, CDCl₃) δ 170.7 (C_q, Ac), 169.0 (C_q, Ac), 168.9 (C_q, Ac), 137.7 (C_q), 137.7 (C_q), 128.6-128.2 (CH_{arom} Bn), 92.8 (C-1_β), 90.5(C-1_α), 83.1 (C-4_β), 80.4 (C-4_α), 79.5 (CH₂, MCH), 79.3 (CH₂, OMCH), 78.0 (C-3_α), 77.7 (C-3_β), 75.8 (CH₂ Bn), 75.7 (CH₂ Bn), 74.3 (C-5_β), 71.6 (C-5_α), 65.1 (C-2_β), 62.7 (C-2_α), 62.7 (C-6_β), 62.6 (C-6_α), 38.9 (CH), 38.8 (CH), 30.3-25.9 (CH₂, MCH), 21.1 (CH₃), 21.1 (CH₃ Ac), 21.0 (CH₃ Ac), 20.9 (CH₃ Ac); IR (neat) ν 2922.0, 2108.1, 1743.5, 1369.4, 1217.0, 1136.0, 1026.1, 1008.7, 931.6, 734.8, 698.2; HRMS: C₂₄H₃₃N₂O₇ + Na⁺ requires 498.2214, found 498.2207.

6-O-Acetyl-2-azido-3-O-benzyl-2-deoxy-4-O-methyl-cyclohexane- α/β -D- glucopyranose (162):



Compound **159** (1.31 g, 2.77 mmol) was dissolved in THF (14 mL) and piperidine (840 μ L, 6% v/v) was added. The clear solution was stirred overnight at room temperature. After complete conversion to a lower running spot on TLC the reaction was diluted with EtOAc (100 mL) and poured in 1M HCl (100 mL). The layers

were separated and the organic layer was washed twice with H₂O and once with brine. Subsequently the EtOAc layer was dried and concentrated in vacuo. Flash silica column purification (EtOAc/PE 8%) yielded compound 162 in 98% yield (1.23 g, 2.83 mmol) as a white foam. TLC: EtOAc/PE 25%; ¹H NMR (400 MHz, CDCl₃) δ 7.44-7.28 (m, 10H, CH_{arom} $Bn_{\alpha/\beta}$), 5.27 (t, J = 3.1 Hz, 1H, H-1_{α}), 4.89-4.77 (m, 4H, CH₂Bn_{α/β}), 4.57 (d, J = 4.3 Hz, 1H, H-1_{β}), 4.38-4.31 (m, 2H, H-6_{α/β}), 4.25-4.13 (m, 3H, H-6_{α/β}, H-5_{β}), 4.07 (dtd, J = 6.4, 4.5, 2.3 Hz, 1H, H-5_{α}), 3.98-3.90 (m, 2H, H-4_{α/β}), 3.68 (dt, *J* = 14.4, 6.5 Hz, 2H, CH₂), 3.56-3.51 $(m, 1H, CH_2), 3.47 (m, 1H, CH), 3.41-3.21 (m, 4H, CH_2, H-2_{\alpha/\beta}), 2.09 (s, 2H, H-3_{\alpha/\beta}), 2.08 (s, 2H, H-3_{\alpha/\beta}),$ 3H, CH₃ OAc), 1.90-0.77 (m, 3H, CH₃ OAc), 1.80-0.83 (m, 22H, CH₂, CH, MCH); ¹³C NMR $(100 \text{ MHz}, \text{CDCl}_3) \delta$ 171.0 (C_q, OAc), 137.9 (C_q, OBn), 137.9 (C_q), 128.6-128.1 (CH_{arom} Bn), 96.2 (C-1_{*β*}), 92.1 (C-1_{*α*}), 83.1 (C-4_{*β*}), 80.0 (C-4_{*α*}), 79.24 (CH₂, MCH), 78.71 (C-3_{*α*}), 78.05 (C- (3_{β}) , 75.67 (CH₂ Bn), 75.60 (C- (5_{β})), 73.5 (C- (5_{α})), 69.4 (C- (2_{β})), 67.5 (C- (2_{β})), 64.1 (C- $(6_{\alpha/\beta})$), 63.2 (CH₂), 63.1 (CH₂), 47.6 (CH), 42.8- 24.5 (CH₂, MCH), 21.4 (CH₃ Ac), 21.0 (CH₃ Ac): Peak assignment based on chemical shift because couplings are inconclusive; IR (neat) v 2922.0, 2852.5, 2106.1, 1741.6, 1616.2, 1450.4, 1363.6, 1234.4, 1116.7, 1082.0, 1033.8, 910.3, 736.8, 698.2; HRMS: $C_{22}H_{31}N_3O_6 + Na^+$ requires 456.2105, found 456.2102.

6-O-Acetyl-2-azido-3-O-benzyl-2-deoxy-4-O-methyl-cyclohexane-1-O-(N- trichloro-acetimidoyl)-α-D-glucopyranoside (147):



A solution of compound **162** (1.18 g, 2.72 mmol) and trichloroacetonitrile (1.03 mL, 8.16 mmol, 3 equiv) in dry DCM (14 mL) was treated with 1,8-diazabicyclo[5.4.0]undec-7-ene 81 μ L (0.54 mmol, 0.2 equiv) for 18 h at ambient temperature. The dark brown solution was concentrated under reduced pressure and the brown oil was directly purified using a sil-

ica gel column (EtOAc/PE 7.5% + 2.5% TEA) to obtain the title compound **147** as a yellow oil (1.11 g, 1.93 mmol, 71%). TLC: EtOAc/PE 30%; ¹H NMR (400 MHz, CDCl₃) δ 8.77 (s, 1H, NH), 7.43-7.25 (m, 5H, CH_{arom} Bn), 6.38 (d, *J* = 3.5 Hz, 1H, H-1_{1α}), 5.62-5.58 (m, 1H, H-1_β), 4.91-4.82 (m, 2H, CH₂ Bn), 4.35-4.19 (m, 2H, CH₂ H-6), 4.01-3.90 (m, 2H, CH₂, MCH), 3.70 (dt, *J* = 10.9, 5.5 Hz, 1H, H-4), 3.64-3.57 (m, 1H, H-5), 3.49-3.38 (m, 1H, H-2), 3.25 (dt, *J* = 12.7, 6.3 Hz, 1H, H-3), 2.08-2.02 (m, 3H, CH₃ Ac), 1.82-1.61 (m, 4H, CH₂, MCH), 1.60-1.46

(m, 1H, CH, MCH), 1.28-1.11 (m, 4H, CH₂, MCH), 1.01-0.86 (m, 2H, CH₂, MCH); ¹³C NMR (100 MHz, CDCl₃) δ 170.2 (C_q, C=N), 160.4 (C_q, Ac), 137.5 (C_q), 128.3-127.8 (C-1), 94.5 (C_q, CCl₃), 79.8 (C-3/4/5), 79.2 (CH₂, MCH), 77.9 (C-3/4/5), 75.3 (CH₂ Bn), 71.8 (C-3/4/5), 62.7 (C-2), 62.2 (C-6), 38.5 (MCH), 30.0-25.6 (CH₂, MCH), 20.6 (CH₃ Ac); IR (neat) *v* 2106.1, 1739.7, 1652.9, 1506.3, 1232.4, 110.9, 1004.8, 904.6, 835.1, 698.2.

1,6-Anhydro-2-azido-3-O-benzyl-2-deoxy-4-O-(6-O-acetyl-2-azido-3-O -benzyl-2-deoxy-4-O-methyl-cyclohexane- β -D-glucopyranosyl)- β -D-glucopyranose (144):



A mixture of imidate **147** (1.09 g, 1.87 mmol) and alcohol **148** (1.04 g, 3.76 mmol, 2 equiv) was coevaporated thrice with toluene and dissolved in dry toluene (9.5 mL). To this mixture activated molecular sieves (4Å) were added and the solution was cooled to -78 °C. After 10 minutes a solution of $BF_3 \cdot Et_2O$ (47 μ L, 0.374 mmol, 0.2 equiv) in 0.25 mL dry toluene was added and the temperature was allowed to rise

to -20 °C in 90 minutes. TLC analysis showed complete conversion of imidate 147 into a lower running spot. The reaction was quenched using TEA (0.5 mL), filtered and concentrated in vacuo. The excess of acceptor was acetylated, using an Ac₂O-pyridine cocktail (1 mL/3 mL), after which the reaction was quenched using MeOH and concentrated. The oily residue was directly purified using a silica gel column (EtOAc/PE 10%). Compound 144 was obtained in 65% in 1:8 α : β ratio (0.845 g, 1.22 mmol). TLC: EtOAc/Tol 25%; ¹H NMR (400 MHz, CDCl₃) δ 7.42-7.23 (m, 10H, CH_{arom} Bn), 5.55 (s, 1H, H-1_{α}), 5.49 (s, 1H, H-1_β), 4.87-4.74 (m, 2H, CH₂ Bn), 4.72-4.56 (m, 3H, CH₂ H-6', H-5'), 4.37-4.30 (m, 3H, CH₂ Bn, H-1), 4.20-3.99 (m, 2H, CH₂ Bn, H-6), 3.96 (s, 1H, H-3), 3.80-3.44 (m, 3H, CH₂ H-6, MCH, H-5'), 3.34-3.20 (m, 5H, CH₂, MCH, H-2, H-3', H-4, H-4'), 2.00 (s, 3H, CH₃ Ac), 1.78-1.45 (m, 5H, CH₂, CH, MCH), 1.29-1.09 (m, 4H, CH₂, MCH), 0.99-0.84 (m, 2H, CH₂, MCH); 13 C NMR (100 MHz, CDCl₃) δ 170.7 (C_q, Ac), 137.9 (C_q), 137.6 (C_q), 128.8-127.8 (CH_{arom} Bn), 102.3 (C-1'), 100.7 (C-1), 82.9 (C-3'/4'), 80.1 (C-4'/3'), 79.4 (CH₂. MCH), 77.6 (C-5'), 77.4 (C-3), 75.7 (CH₂ Bn), 75.0, 74.8 (C-5), 73.6 (C-4), 72.6 (C-6'), 65.9 (C-2'), 65.2 (C-6), 62.7 (CH₂ Bn), 59.3 (C-2), 38.8 (MCH), 30.3-25.9 (CH₂, MCH), 20.9 (CH₃ Ac). IR (neat) v 2922.0, 2100.3, 1739.7, 1454.2, 1363.6, 1236.3, 1070.4, 1026.1, 1006.8, 964.3, 902.6, 736.8, 696.3; HRMS: $C_{35}H_{44}N_6O_9 + Na^+$ requires 715.3062, found 715.3064; $[\alpha]_{D}^{23}$ 5 °(c = 0.2, CHCl₃).

1,6-Anhydro-2-azido-3-*O*-benzyl-2-deoxy-4-*O*-(2-azido-3-*O*-benzyl-2-deoxy-4-*O*-methyl- cyclohexane-β-D-glucopyranosyl)-β-D-glucopyranose (165):



Disaccharide **144** (785 mg, 1.13 mmol) was dissolved in MeOH (5 mL) and a catalytic amount of NaOMe (30% in MeOH) was added. The reaction mixture was stirred for 1 h at ambient temperature, after which it was neutralized ($pH\sim7$) using Amberlite^(R) IR-120 H⁺ resin. Filtering off the resin, concentration and purification using a short silica column (EtOAc/PE 10%) yielded title compound **165** as

a clear oil (377 mg, 580 μ mol, 52%). TLC: EtOAc/PE 30%; ¹H NMR (400 MHz, d_4 , MeOD) δ .43-7.25 (m, 10H, CH_{arom} Bn), 5.46 (s, 1H, H-1), 4.89-4.78 (m, 2H, CH₂ Bn), 4.75-4.62 (m, 3H, CH₂ Bn, H-5), 4.56 (d, J = 7.4 Hz, 1H, H-1_{β}), 4.11 (d, J = 7.3 Hz, 1H, CH₂ H-6), 3.93 (d, J = 8.3 Hz, 2H, H-5', H-3), 3.86-3.67 (m, 3H, CH₂, H-6', H-6), 3.62 (dt, J = 14.0, 7.0 Hz, 1H, CH₂, MCH), 3.44-3.28 (m, 5H, CH₂, MCH, H-2', H-3', H-4', H-4), 3.23 (s, 1H,

H-2), 1.75 (dd, J = 30.5, 13.3 Hz, 4H, CH₂, MCH), 1.53 (s, J = 2.9 Hz, 1H, CH, MCH), 1.33-1.11 (m, 4H, CH₂, MCH), 1.03-0.87 (m, 2H, CH₂, MCH); ¹³C NMR (100 MHz, d_4 , MeOD) δ 139.7 (C_q), 139.4 (C_q), 129.5-128.8 (CH_{arom} Bn), 102.7 (C-1'), 101.9 (C-1), 84.4 (C-3'/4'/4), 79.9 (CH₂, MCH), 78.9 (C-3'/4'/4), 78.7 (C-3'/4'/4), 77.4 (C-5'/3), 77.3 (C-5'/3), 76.4 (CH₂ Bn), 76.0 (C-5), 73.4 (CH₂ Bn), 67.6 (C-2'), 66.2 (C-2), 62.0 (C-6), 61.0 (C-2), 40.1 (MCH), 31.3-27.0 (CH₂, MCH); IR (neat) v 2927.8, 2356.1, 2106.1, 1733.4, 1278.3, 1113.0, 1069.4, 708.8, 504.4; HRMS: C₃₃H₄₂N₆O₈ + Na⁺ requires 673.2956, found 673.2957; $[\alpha]_{\rm D}^{23}$ -4 °(c = 0.2, CHCl₃).

2-Azido-3-O-benzyl-2-deoxy-1,6-di-O-acetyl-4-O-(2-azido-3-O-benzyl-2-deoxy-4-O-methyl-cyclohexane- β -D-glucopyranosyl)- α/β -D-glucopyranose (168):



To a solution of compound **165** (377 mg, 580 μ mol) in 3 mL Ac₂O 0.45 mL (15% v/v) TFA was added. The reaction was stirred for 18 h after which it was diluted with toluene and concentrated. After purification using silica gel chromatograph (EtOAc/PE 10%) disac-

charide **168** was obtained in 87% yield (400 mg, 505 μ mol) as transparent foam. TLC: EtOAc/PE 25%; ¹H NMR (400 MHz, CDCl₃) δ 7.44-7.19 (m, 10H, CH_{arom} Bn), 6.19 (d, J = 3.7 Hz, 1H, H_{1a}), 5.50-5.41 (m, 1H, H-1_β), 5.17-4.98 (m, 1H, CH₂ Bn), 4.87-4.71 (m, 3H, CH₂ Bn), 4.52-4.43 (m, 1H, CH₂ H-6), 4.35 (m, 2H, CH₂ H-6, H-1), 4.23-4.17 (m, 1H, CH₂ H-6'), 4.14-4.07 (m, 2H, CH₂ H-6', H-3'), 4.00-3.85 (m, 2H, H-3, H-4), 3.64 (dd, J = 8.5, 5.8 Hz, 1H, CH₂, MCH), 3.57-3.50 (m, 1H, H-2), 3.44-3.27 (m, 4H, H-2', H-4, H-5, H-5'), 3.20 (dd, J = 8.4, 7.0 Hz, 1H, CH₂, MCH), 2.16 (d, J = 2.4 Hz, 3H, CH₃ Ac), 2.10 (s, 3H, CH₃ Ac), 1.90 (s, 3H, CH₃ Ac), 1.75-1.60 (m, 4H, CH₂, MCH), 1.54-1.43 (m, 1H, CH, MCH), 1.27-1.09 (m, 4H, CH₂, MCH), 0.96-0.83 (m, 2H, MCH); ¹³C NMR (100 MHz, CDCl₃) δ 170.5 (Cq, Ac), 168.8 (Cq, Ac), 138.2 (Cq), 137.6 (Cq), 128.5-127.4 (CH_{arom} Bn), 101.4 (C-1'), 90.1 (C-1), 83.3 (C-4/5/'4), 79.1 (CH₂, MCH), 78.4 (C-3), 77.8 (C-4/5/'4), 77.1 (C-4), 7.7 (CH₂ Bn), 75.0 (CH₂ Bn), 73.5 (C-4/5/'4), 71.0 (C-3'), 66.8 (C-2'), 62.8 (C-6), 62.2 (C-2), 62.1 (C-6'), 38.7 (MCH), 30.1-25.8 (CH₂, MCH), 21.0 (CH₃ Ac), 20.8 (CH₃ Ac), 20.7 (CH₃ Ac); IR (neat) ν 2922.0, 2356.9, 2341.4, 2108.1, 1737.7, 1452.3, 1363.6, 1222.8, 1105.1, 1026.1, 1008.7, 931.6, 823.5, 734.8, 696.3; HRMS: C₃₉H₅₀N₆O₁₂ + Na⁺ requires 817.3379, found 817.3382.

2-Acetamido-2-deoxy-1,3,6-tri-O-acetyl-4-O-(2-acetamido-2-deoxy-3,6-O-di-acetyl-4--O-methyl-cyclohexane- β -D-glucopyranosyl)- α/β -D-glucopyranose (141):



Compound **168** (365 mg, 0.46 mmol) was taken up in a mixture of dioxane:toluene; H_2O (5:2:1, v/v/v, 8 mL) and cooled with an ice-bath. After 10 minutes PMe₃ (2.30 mL, 5 equiv) 1M intoluene was added and the reaction was stirred for 18 h at +4 °C. After TLC anal-

ysis showed conversion towards lower running, ninhydrin positive spot, the reaction was coevaporated thrice with toluene and subsequently acetylated using an Ac_2O -pyridine cocktail (2 mL/6 mL). The mixture was stirred for 20 h after which it was quench with MeOH and concentrated under reduced pressure. The resulting white solid was dissolved in a 1:1 mixture of MeOH and TFE (8 mL) and purged with argon. A catalytic amount of Pd(OH)₂ spiked with Pd-black was added and the mixture was purged with H₂. Reduction of the benzyl-groups was continued for 5 h followed by filtration and concentration. The residue was again taken up in an Ac₂O-pyridine cocktail (2 mL:6 mL) with a catalytic

amount of DMAP and stirred at ambient temperature for 18 h. After complete acetylation of the disaccharide the mixture was quenched with MeOH and concentrated under reduced pressure. Silica gel purification (MeOH/DCM 3%) yielded 122 mg (170 μ mol, 39%, 73% per step) of the title compound **141** as an off-white foam. TLC: MeOH/DCM 5%; ¹H NMR (400 MHz, CDCl₃) δ 7.44-7.19 (m, 14H), 6.19 (d, J = 3.7 Hz, 1H), 5.50-5.41 (m, 1H), 5.17-4.98 (m, 1H), 4.87 - 4.71 (m, 4H), 4.52-4.43 (m, 1H), 4.35 (m, 3H), 4.23-4.17 (m, 1H), 4.14-4.07 (m, 2H), 4.00-3.85 (m, 3H), 3.64 (dd, J = 8.5, 5.8 Hz, 1H), 3.57-3.50 (m, 2H), 3.44-3.27 (m, 5H), 3.20 (dd, J = 8.4, 7.0 Hz, 1H), 2.16 (d, J = 2.4 Hz, 3H), 2.10 (s, 4H), 1.90 (s, 3H), 1.75-1.60 (m, 7H), 1.54-1.43 (m, 2H), 1.27-1.09 (m, 6H), 0.96-0.83 (m, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 171.3, 171.0-169.09, 102.0, 101.9, 100.9, 92.4, 90.4, 90.3, 82.5, 79.0, 78.8, 76.6, 75.8, 75.6, 75.1, 75.0, 74.8, 73.6, 73.4, 73.1, 72.1, 70.9, 70.7, 69.8, 69.7, 63.0, 62.9, 62.2, 61.7, 54.1, 53.6, 51.9, 51.2, 38.4, 29.9, 29.8, 29.7, 29.7, 26.3, 25.7, 25.7, 23.1, 23.1, 22.9, 21.0, 20.9, 20.8, 20.83, 20.79, 20.77, 20.7, 20.6, 20.6; IR (neat) v 2925.8, 1739.7, 1652.9, 1558.4, 1521.7, 1506.3, 1369.4, 1220.9, 1112.9, 1012.6, 939.3, 906.5, 715.5; HRMS: $C_{33}H_{50}N_2O_{16} + Na^+$ requires 753.3053, found 753.3055

2-Acetamido-1,3,6-tri-*O*-acetyl-2-deoxy-4-*O*-(2-acetamido-2-deoxy-3,6-*O*-di-acetyl-4-*O*-methyl-cyclohexane- β -D-glucopyranosyl)-1-*O*-4-methylumbelliferyl- β -D-glucopyranoside (171):



Dimer **141** (70 mg, 95 μ mol) was dissolved in AcOH (2 mL) and Ac₂O (1 mL). At 0 °C dry HCl_(g) was bubbled through (liberated under Kipp conditions) for 3h. The reaction mixture was then placed at 5 °C for 42 h at

which TLC analyses (DCM-acetone 60-40) showed complete consumption of starting material. The reaction diluted with CHCl₃ (10 ml, 0 °C) and washed twice with H₂O (15 ml, 0 °C) and twice with NaHCO₃ (15 mL, 0 °C). The organic layer was dried over MgSO₄ and concentrated *in vacuo* yielding an amorphous solid of which the purity was evaluated by ¹H-NMR. The resulting solid was dissolved in CHCl₃ (5 ml) and added to a solution of NaHCO₃ 0.2M (5 ml), 4-methylumbelliferyl sodium salt 34,35 (94 mg, 480 μ mol) and TBAHS (64 mg, 191 μ mol). The biphasic mixture was stirred overnight with the exclusion of light. The phases were separated and the organic layer was washed twice with NaHCO₃ (0.2 M) and twice with H_2O . The organic layer was dried over MgSO₄ and concentrated in vacuo. Purification by column chromatography (MeOH/CHCl₃ 3%) yielded 171 (41 mg, 48 μ mol, 50%) as a white amorphous solid. TLC: MeOH/DCM 5%; ¹H NMR (400 MHz, d_4 , MeOD) δ 7.61 (d, J = 8.4 Hz, 1H, CH, 4-methylumbelliferyl), 7.01-6.95 (m, 2H, CH_{arom} 4-methylumbelliferyl), 6.20 (s, 1H, CH_{arom} 4-methylumbelliferyl), 5.25 (t, J = 8.3 Hz, 2H, $H-1_{\beta}$, H-3), 5.17-5.10 (m, 1H, H-3'), 4.63 (d, J = 8.3 Hz, 1H, $H-1'_{\beta}$), 4.51 (d, J = 10.2 Hz, 1H, CH₂ H-6), 4.31 (d, J = 2.7 Hz, 2H, CH₂ H-6'), 4.24-4.11 (m, 2H, CH₂ H-6, H-2), 3.93 (t, *J* = 8.3 Hz, 1H, H-5), 3.87-3.74 (m, 2H, H-4, H-2), 3.58 (dd, *J* = 8.0, 4.9 Hz, 1H, H-5'), 3.48-3.25 (m, 3H, CH₂, MCH, H-4'), 2.45 (s, 3H, CH₃ 4-methylumbelliferyl), 2.14 (s, 6H, CH_3 Ac), 2.10-2.04 (m, 6H, CH_3 Ac), 1.96 (d, J = 5.8 Hz, 6H, CH_3 Ac), 1.68 (t, J = 11.4Hz, 4H, CH₂, MCH), 1.44 (d, J = 11.2 Hz, 1H, CH, MCH), 1.30-1.09 (m, 4H, CH₂, MCH), 0.91 (dd, J = 20.8, 11.9 Hz, 2H, CH₂, MCH); ¹³C NMR (100 MHz, d_4 , MeOD) δ 171.7-170.1 (Cq, Ac), 161.4 (Cq, 4-methylumbelliferyl), 159.4 (Cq, 4-methylumbelliferyl), 154.2 (Cq, 4methylumbelliferyl), 153.3 (Cq, 4-methylumbelliferyl), 125.5 (CH_{arom} 4-methylumbelliferyl), 114.7 (C_q, 4-methylumbelliferyl), 113.6 (CH_{arom} 4-methylumbelliferyl), 111.8 (CH_{arom} 4-methylumbelliferyl), 103.3 (CH_{arom} 4-methylumbelliferyl), 100.4 (C-1'), 97.5 (C-1), 78.2 (CH₂, MCH), 75.7-75.7 (C-4/4'), 74.4 (C-3'), 72.6-72.4 (C-3/5/5'), 62.6 (C-6'), 62.3 (C-6), 54.3 (C-2'), 53.3 (C-2), 38.0 (MCH), 29.4-25.2 (CH₂, MCH), 21.7-17.6 (CH₃ Ac); IR (neat) ν 1739.7, 1652.9, 1539.1, 1521.7, 1488.9, 1473.5, 1369.4, 1222.8, 1012.6, 939.3, 902.6, 628.8; HRMS:C₄₁H₅₄N₂O₁₇ + Na⁺ requires 869.3315, found 869.3317.

2-Acetamido-2-deoxy-4-O-(2-acetamido-2-deoxy-4-O-methyl-cyclohexane- β -D-gluco-pyranosyl)-1-O-4-methylumbelliferyl- β -D-glucopyranoside (138):



To a suspension of **171** (20 mg, 23 μ mol) in MeOH (1 mL) was added NaOMe (30 wt% in MeOH) (414 μ L, 2.30 μ mol). The reaction was stirred with the exclusion of light. When LCMS (gradient 0 to 50 % MeOH)

showed complete conversion to product the mixture was quenched with AcOH (5 μ L, 80 μ mol). The reaction was diluted with H₂O (2 mL) the MeOH was evaporated *in vacuo* and the remaining H_2O was lyophilized. Purification by HPLC (gradient H_2O -MeOH + 0.1% TFA 80-20 \rightarrow 60-40) evaporation of MeOH and lyophilizing H₂O yielded **138** (3.5 mg, 5.16 μ mol, 22%) as white fluffy solid. ¹H NMR (600 MHz, (d_6), DMSO) δ 7.90 (d, J = 9.2 Hz, 1H, NH), 7.74 (d, *J* = 9.0 Hz, 1H, NH), 7.70 (d, *J* = 8.8 Hz, 1H, CH, 4-methylumbelliferyl), 7.04 (d, J = 2.3 Hz, 1H, CH, 4-methylumbelliferyl), 6.96 (dd, J = 8.8, 2.3 Hz, 1H CH, 4methylumbelliferyl), 6.27 (s, 1H, CH, 4-methylumbelliferyl), 5.18 (d, J = 8.4 Hz, 1H, H-1), 5.06 (d, J = 6.3 Hz, 1H, OH), 4.81 (dd, J = 5.7, 5.0 Hz, 1H, OH), 4.77 (d, J = 2.1 Hz, 1H, OH), 4.69 (t, J = 6.0 Hz, 1H, OH), 4.38 (d, J = 8.4 Hz, 1H, H₁'), 3.33 (s, 14H), 3.00 (t, J = 1.09.2 Hz, 1H), 2.41 (s, J = 9.4 Hz, 3H, CH₃ 4-methylumbelliferyl), 1.85 (s, 3H, CH₃ Ac), 1.81 (s, 3H, CH₃ Ac), 1.70-1.59 (m, 4H, CH₂, MCH), 1.51-1.44 (m, 1H, CH, MCH), 1.24-1.07 (m, 4H, CH₂, MCH), 0.89 (d, J = 12.2 Hz, 2H, CH₂, MCH); ¹³C NMR (151 MHz, (d_6), DMSO) δ 169.2, 169.1, 160.0, 159.9, 154.4, 153.3, 126.5, 114.3, 113.4, 111.9, 103.1, 101.9, 98.2, 80.8, 78.6, 77.6, 75.7, 75.0, 74.0, 72.3, 55.8, 54.4, 38.0, 29.7, 29.4, 26.2, 25.4, 25.3, 23.0, 18.1; HRMS: $C_{33}H_{46}N_2O_{13} + Na^+$ requires 701.2892, found 701.2890

3.5 Biological Evaluation

Enzymes: Human chitotriosidase was expressed in BHK cells and purified from culture medium as described earlier.³⁶ Jack bean β -hexosaminidase was purchased from Sigma. The tissue was homogenized in 4 volumes of 0.1 M potassium phosphate buffer pH 6.5. The soluble fraction was isolated following ultracentrifugation at 50.000 g for 20 minutes and next β -hexosaminidiase in the supernatant was enriched using Concanavalin. A chromotagraphy and elution of bound enzyme with 0.1 mM methylmannose 0.1 M potassium phosphate buffer pH 6.5.

Enzyme activity measurements using artificial 4-*MU-substrates:* For activity measurements of β -hexosaminidases, 4-MU-GlcNac (Sigma) was used as substrate. Briefly, samples were incubated for 20 minutes at 37 °C with 2 mM 4-MU-GlcNac in McIlvaine buffer (100 mM citric acid, 200 mM sodium phosphate, pH 4.0). The reactions were stopped by the addi-

tion of excess 0.3 M glycine-NaOH, pH 10.3. Formed 4-methylumbelliferone was detected fluorimetrically (excitation at 366 nm; emission at 445 nm). Activity of recombinant chitotriosidase towards 4-MU-oligosaccharide substrates was determined by incubation at 37 °C in McIlvaine buffer (100 mM citric acid, 200 mM sodium phosphate, pH 5.2). Substrate concentrations in various experiments were different as indicated.

References

- Boot, R. G.; Renkema, G. H.; Strijland, A.; van Zonneveld, A. J.; Aerts, J. M. F. G. J. Biol. Chem. 1995, 270, 26252–26256.
- [2] Hollak, C. E. M.; van Weely, S.; van Oers, M. H. J.; Aerts, J. M. F. G. J. Clin. Invest. 1994, 93, 1288–1292.
- [3] Renkema, G. H.; Boot, R. G.; Muijsers, A. O.; Donkerkoopman, W. E.; Aerts, J. M. F. G. J. Biol. Chem. 1995, 270, 2198–2202.
- [4] Brady, R. O.; Kanfer, J. N.; Shapiro, D. Biochem. Biophys. Res. Commun. 1965, 18, 221–225.
- [5] Schoonhoven, A.; Rudensky, B.; Elstein, D.; Zimran, A.; Hollak, C. E. M.; Groener, J. E.; Aerts, J. M. F. G. *Clin. Chim. Acta* 2007, *381*, 136–139.
- [6] Aguilera, B.; Ghauharali-van der Vlugt, K.; Helmond, M. T. J.; Out, J. M. M.; Donker-Koopman, W. E.; Groener, J. E. M.; Boot, R. G.; Renkema, G. H.; van der Marel, G. A.; van Boom, J. H.; Overkleeft, H. S.; Aerts, J. M. F. G. *J. Biol. Chem.* **2003**, *278*, 40911–40916.
- [7] Aerts, J. M. F. G.; Hollak, C. E. M. Baillieres Clin. Haem. 1997, 10, 691-709.
- [8] Bussink, A. P.; Verhoek, M.; Vreede, J.; Ghauharali-van der Vlugt, K.; Donker-Koopman, W. E.; Sprenger, R. R.; Hollak, C. E.; Aerts, J. M. F. G.; Boot, R. G. FEBS J. 2009, 276, 5678–5688.
- [9] Guo, Y.; He, Y.; Boer, A.; Wevers, R.; de Bruijn, R.; Groener, J.; Hollak, C.; Aerts, J.; Galjaard, H.; van Diggelen, H. J. Inherit. Metab. Dis. 1995, 18, 717–722.
- [10] Brinkman, J.; Wijburg, F.; Hollak, C.; Groener, J.; Verhoek, M.; Scheij, S.; Aten, J.; Boot, R.; Aerts, J. J. Inherit. Metab. Dis. 2005, 28, 13–20.
- [11] vom Dahl, S.; Harzer, K.; Rolfs, A.; Albrecht, B.; Niederau, C.; Vogt, C.; van Weely, S.; Aerts, J.; Müller, G.; Häussinger, D. J. Hepatol. 1999, 31, 741 – 746.
- [12] Vedder, A.; Cox-Brinkman, J.; Hollak, C.; Linthorst, G.; Groener, J.; Helmond, M.; Scheij, S.; Aerts, J. Mol. Genet. Metab. 2006, 89, 239 – 244.
- [13] Boot, R. G.; Hollak, C.; Verhoek, M.; Alberts, C.; Jonkers, R. E.; Aerts, J. M. F. G. *Clin. Chim. Acta* 2010, 411, 31 – 36.
- [14] Iyer, A.; van Eijk, M.; Silva, E.; Hatta, M.; Faber, W.; Aerts, J.; Kumar Das, P. Clin. Immunol. 2009, 131, 501 – 509.
- [15] Boven, L. A.; Van Meurs, M.; Van Zwam, M.; Wierenga-Wolf, A.; Hintzen, R. Q.; Boot, R. G.; Aerts, J. M. F. G.; Amor, S.; Nieuwenhuis, E. E.; Laman, J. D. *Brain* **2006**, *129*, 517–526.
- [16] Boot, R. G.; van Achterberg, T. A. E.; van Aken, B. E.; Renkema, G. H.; Jacobs, M. J. H. M.; Aerts, J. M. F. G.; de Vries, C. J. M. Arterioscler., Thromb., Vasc. Biol. 1999, 19, 687–694.
- [17] Labadaridis, J.; Dimitriou, E.; Costalos, C.; Aerts, J. M. F. G.; van Weely, S.; Donker-Koopman, W.; Michelakakis, H. Acta Paediatr. 1998, 87, 605–606.
- [18] Crich, D.; Dudkin, V. J. Am. Chem. Soc. 2001, 123, 6819–6825.
- [19] Shapiro, D.; Rabinsoh, Y.; Acher, A. J.; Diverhab, A. J. Org. Chem. 1970, 35, 1464–1467.
- [20] van Leeuwen, S. H. Ph.D. thesis, Leiden University, 1997.

- [21] Tailler, D.; Jacquinet, J. C.; Noirot, A. M.; Beau, J. M. J. Chem. Soc., Perkin Trans. 1 1992, 3163– 3164.
- [22] Hawley, J.; Bampos, N.; Aboitiz, N.; Jimenez-Barbero, J.; de la Paz, M. L.; Sanders, J. K. M.; Carmona, P; Vicent, C. *Eur. J. Org. Chem.* **2002**, 1925–1936.
- [23] Czernecki, S.; Leteux, C.; Veyrieres, A. Tetrahedron Lett. 1992, 33, 221-224.
- [24] Leteux, C.; Veyrieres, A. J. Chem. Soc., Perkin Trans. 1 1994, 2647-2655.
- [25] Hori, H.; Nishida, Y.; Ohrui, H.; Meguro, H. J. Org. Chem. 1989, 54, 1346–1353.
- [26] Tailler, D.; Jacquinet, J. C.; Beau, J. M. J. Chem. Soc., Chem. Commun. 1994, 1827–1828.
- [27] Roy, R.; Tropper, F. D.; Grandmaitre, C. Can. J. Chem. 1991, 69, 1462–1467.
- [28] Carriere, D.; Meunier, S. J.; Tropper, F. D.; Cao, S.; Roy, R. J. Mol. Catal. A: Chem. 2000, 154, 9–22.
- [29] Pavliak, V.; Kovac, P. Carbohydr. Res. 1991, 210, 333-337.
- [30] Lee, J.; Coward, J. J. Org. Chem. 1992, 57, 4126-4135.
- [31] Oguri, S.; Tejima, S. Chem. Pharm. Bull. 1981, 29, 1629–1635.
- [32] Inaba, T.; Ohgushi, T.; Iga, Y.; Hasegawa, E. Chem. Pharm. Bull. 1984, 32, 1597–1603.
- [33] Ikonne, J. U.; Ellis, R. B. Biochem. J. 1973, 135, 457-462.
- [34] Kostova, I. P.; Manolov, I. I.; Nicolova, I. N.; Danchev, N. D. Farmaco 2001, 56, 707–713.
- [35] Kostova, I. P.; Manolov, I. I.; Radulova, M. K. Acta Pharm. 2004, 54, 37–47.
- [36] Fusetti, F; von Moeller, H.; Houston, D.; Rozeboom, H. J.; Dijkstra, B. W.; Boot, R. G.; Aerts, J. M. F. G.; van Aalten, D. M. F. J. Biol. Chem. 2002, 277, 25537–25544.