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Targeting and exploiting cytomegalovirus for vaccine development

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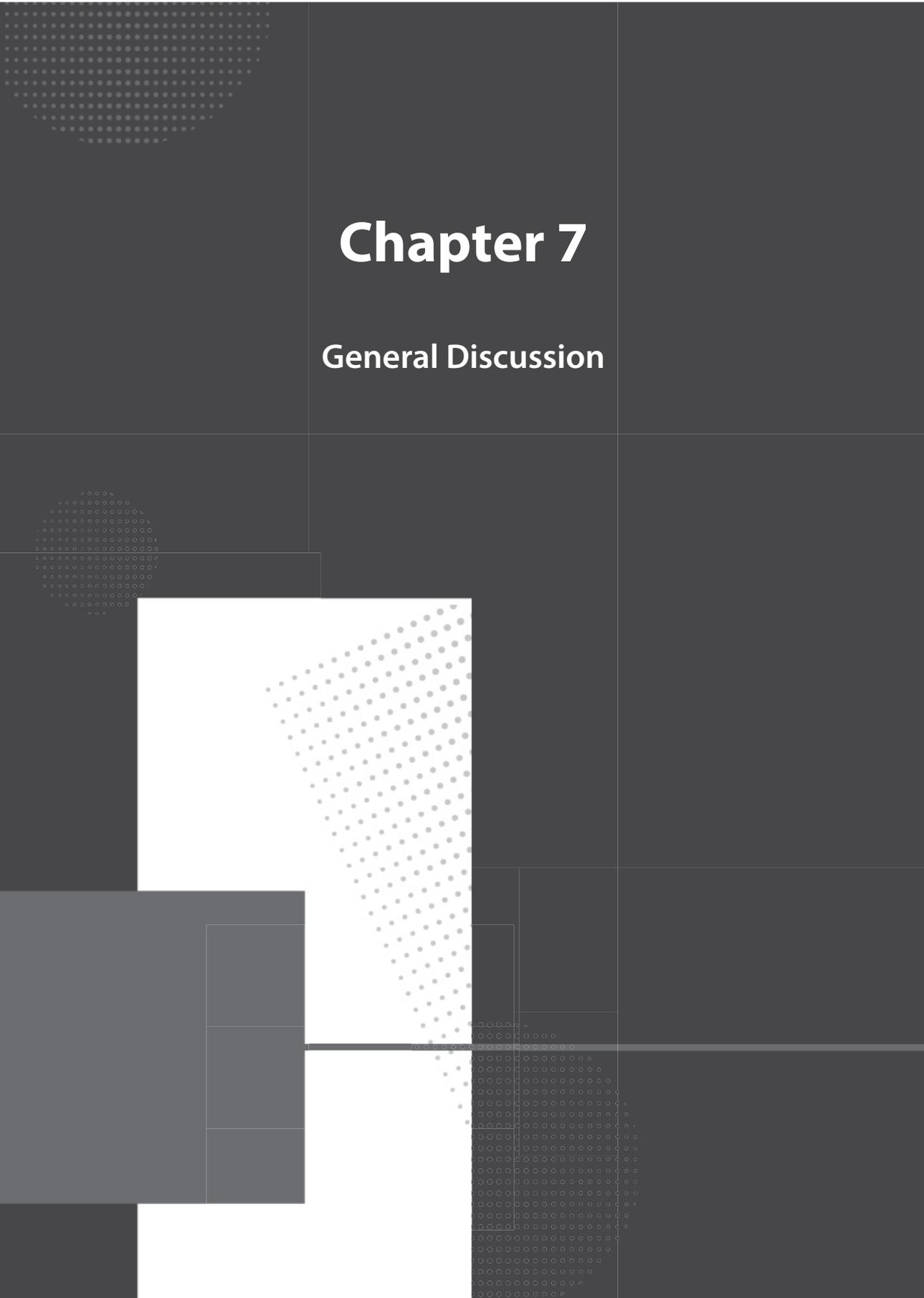
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Chapter 7

General Discussion



GENERAL DISCUSSION

T cells are essential components of naturally acquired protective immune responses in many diseases. Although various ways of inducing potent T cell response by vaccination have been assessed, the majority result in low level, non-protective immune responses. Vaccine development has broadened and currently vaccine platforms range from replication-deficient (attenuated) or killed micro-organisms to viral vectored vaccines and more recently to protein subunit vaccines. Many highly effective vaccines designed to induce protective CD4⁺ and/or CD8⁺ T cells against chronic infections and cancer have been developed. Although their precise mechanisms of protection depend on the complexity and type of antigen, the quality of the vaccine-induced T cell response is gaining increasing attention.

In this doctoral thesis, the SLP T cell-based vaccination strategy alone or in combination with agonistic antibodies triggering the costimulatory TNFR superfamily member OX40 were tested for their capacity to induce potent CD4⁺ and CD8⁺ T cell responses and to confer protection against lytic MCMV infection. Vaccination with MHC class I- and II-restricted MCMV epitope containing SLPs successfully eliminated virus spread in naïve mice but failed to boost host's antiviral immune responses and treat established MCMV infection. Direct correlates of protection were thoroughly investigated. Another vaccination model tested was the MCMV-based vector vaccine against virus-induced cancer (HPV). Injection of the MCMV vectored vaccine expressing an immunodominant MHC class-I restricted HPV E7 epitope led to long term protection against tumor outgrowth in naïve hosts but exhibited limited efficacy in hosts with a strong pre-existing immunity to MCMV. To optimise both vaccination strategies, we explored essential components required to shape protective immune responses and ways to subvert existing immune responses and improve immune recognition.

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COMBINATION IMMUNOTHERAPY AGAINST MCMV

Several approaches can be used to improve immunity to MCMV-associated disease. Concomitant stimulation of CD8⁺ and CD4⁺ T cell responses via MHC class I and II epitope-containing SLPs, respectively, and augmentation of costimulatory signals mediated by the TNFR family member OX40 are very promising tools, which will be extensively discussed below.

Maximum virus control through activation of CD4⁺ and CD8⁺ T cells

The last decades, numerous immunotherapeutic platforms mainly focused at the induction of neutralizing antibodies have been tested against CMV. The majority of these vaccines exhibited low or short-term efficacy [1]. Whether sterile immunity against CMV is an achievable goal remains questionable mainly due to the numerous immune evasion mechanisms exploited by the virus. T cell-based vaccines designed to

induce CD4⁺ or/and CD8⁺ T cell responses is a rapidly growing field. Many studies have demonstrated the importance of T cell immunity against CMV. There is solid evidence that CD4⁺ T cells are crucial for controlling CMV replication during the acute phase of the infection while CD8⁺ T cells play an essential role during latency and virus reactivation [2-4]. SLP T cell based immunotherapy is a safe and well-explored vaccination platform, which results in the activation of CD4⁺ and CD8⁺ T cells and has shown remarkable potency to treat HPV associated disease in both preclinical and clinical studies [5,6].

In **chapter 2**, MHC class I epitope containing SLPs along with TLR9 agonists were tested in a prime-boost vaccination setting in various mouse strains for their ability to contain high load systemic MCMV infection. Vaccination led to the induction of vigorous and poly-functional (IFN- γ ⁺/ TNF⁺/ IL-2⁺) cytotoxic CD8⁺ T cell subsets mediating potent and long-term protective immunity against MCMV infection. Vaccine-induced CD8⁺ T cells slowly converted to a unique memory T cell subset sharing features from both effector and central memory T cells while their functionality was significantly improved throughout time. Notably, the size of each distinct SLP vaccine-induced CD8⁺ T cell response was found unrelated to the functional avidity and proportional to the naïve T cell precursor frequency. This finding suggests that T cell precursor frequencies may be considered as a powerful model to predict the subsequent size of the T cell response induced upon peptide vaccination. In shared antigen or neo-antigen cancer vaccine trials *in vitro* stimulation with the targeted antigen is used to provide insight into the patient's immune system ability to respond to potential vaccination [7,8]. However, determination of the targeted antigen/peptide precursor frequency will possibly ease classic *in vitro* antigen selection processes and may form an important parameter to be considered in immunotherapeutic strategies that their efficacy is based on the selection of antigens.

Despite the importance of CD4⁺ T cells in the immune system, the role of CMV-specific CD4⁺ T cell responses in CMV infection has not been completely understood. CD4⁺ T cell "help" is likely to be crucial for increasing the effectiveness of candidate vaccines for CMV. Therefore, in **chapter 3** the efficacy of MCMV-specific CD4⁺ T cells to control MCMV was studied using a vaccine that comprises MHC class II epitopes. In addition, the capacity of the CD4⁺ T cell responses to enhance the efficacy of a MCMV-specific CD8⁺ T cell response was studied by simultaneous injection of MHC class I and II epitope SLPs. CD4⁺ T cells induced after vaccination with various MHC class II SLPs and OX40 ligation elicited broad antiviral Th1 cytokine responses and showed direct antiviral function against MCMV infection. Interestingly, vaccine induced CD4⁺ T cell responses conferred moderate protection against lytic MCMV infection in both lymphoid and non-lymphoid organ tissues. These findings advance the findings of other reports showing direct antiviral effector function of CMV specific CD4⁺ T cells in both mice and human [2,9-12].

Moreover, inclusion of CD4⁺ T cell "help" during vaccination with a combination of MHC class I SLP vaccines significantly enhanced CD8⁺ T cell expansion and remarkably

improved the overall prophylactic vaccine efficacy. Notably, CD4⁺ T cell signals increased priming of vaccine-evoked CD8⁺ T cells suggesting a direct synergy between these T cell subsets. The concept that CD4⁺ T cell “help” to license DC for proper CD8⁺ CTL priming must be antigen specific has long been concluded [13-16]. Thus, MHC class II epitopes are likely to add value if included in the design of epitope-based vaccines against CMV. Consequently, in **chapter 5** a plethora of highly immunogenic IE2 MHC class II restricted T cell epitopes were identified following traditional *in silico* screening methods. Whereas the T cell response to the IE1 HCMV protein is dominated by MHC class I T cell responses, limited CD8⁺ T cell reactivity was measured against the IE2 protein. Since T cell responses to IE antigens predominate the lytic phase of the CMV infection, a vaccine formulation that will comprise IE1-specific MHC class I epitopes and IE2-specific MHC class II epitopes targeting immune dominant CD8⁺ and CD4⁺ T cell viral regions respectively is possible to act synergistically and inhibit the establishment of CMV latency.

The need for enforced OX40 co-stimulation

Therapeutic targeting of immune check point inhibitors (PD-1/ PD-L1 and CTLA-4) or costimulatory receptors (41BB/ 41BBL and OX40/ OX40L) has been beneficial against many chronic viral infections and cancer [17-24]. Costimulatory molecules provide critical interactions for inducing and maintaining adaptive immune response against CMV. Antibodies can specifically bind to costimulatory receptors and boost or inhibit an immune response.

In **chapter 3** agonistic OX40 antibodies in combination with various MHC class I and MHC class II epitope containing SLPs of MCMV-encoded antigens were tested in a prime-boost vaccination schedule. OX40 ligation synergized with the SLP-based vaccines and empowered overall vaccine efficacy but did not exhibit any therapeutic or prophylactic reaction when provided as monotherapy. Enforced OX40 activation strongly increased the number of-vaccine-induced CD4⁺ T cell and CD8⁺ T cell responses, especially when provided during booster vaccination. The effect of OX40 triggering was more pronounced on CD4⁺ T cells presumably due to the higher expression of the OX40 molecule on this T cell subset [25,26]. OX40 mediated signals were not short lived or limited on effector T cells but influenced also memory T cell formation. Specifically, memory vaccine-derived CD8⁺ T cells treated with OX40 agonists during booster SLP vaccination exhibited improved functionality, survival and secondary clonal expansion compared to the untreated subjects.

Interestingly, the timing and the number of doses OX40 agonistic antibody was administered regulated both the size and the quality of the subsequent vaccine-mediated T cell response. Specially, when agonistic OX40 antibody was provided during both prime and booster SLP vaccination all the positive effects of the OX40 triggering on T cell induction were diminished or utterly lost. A possible explanation

for this outcome is that short time intervals between the two vaccinations did not allow adequate memory T cell development and upon secondary OX40 stimulation apoptotic cell death events were dramatically accelerated. T cell susceptibility to activation-induced cell death (AICD) can occur in a cell-autonomous manner and is regulated by previous T cell activation history and the stage of the T cell maturity [27]. Repeated TCR activation and co-stimulation may promote the Fas/CD95 pro-apoptotic pathway whereas correct timing of co-stimulation can promote the Bcl-2 anti-apoptotic pathway [28]. Consistent with this, upregulation of the Bcl-2 molecule, known as a target of OX40, was measured when agonistic anti-OX40 antibody was provided only during booster vaccination and correlated with prolonged T cell survival. In contrast, repeated enforced OX40 stimulation during both prime and booster SLP vaccination downregulated Ki67 and Bcl-2 expression leading to decreased T cell proliferation and survival. Additionally, it has been previously reported that Bcl-2 upregulation accompanied with increased expression of IL-2 inhibits AICD of previously activated T cells [28,29]. OX40 stimulation during booster SLP activation dramatically escalated autocrine IL-2 cytokine production levels and AICD events were delayed. Whether further treatment with IL-2 at the time of stimulation with OX40 agonists could overcome T cell susceptibility to AICD remains to be explored.

Another interesting observation was that IL-2 secretion was tightly regulated by OX40 costimulatory signals. Enforced OX40 stimulation mainly during booster SLP vaccination promoted "Th1" cytokine production by both CD4⁺ and CD8⁺ vaccine-specific T cell subsets. Strikingly, IL-2⁺ CD4⁺ and/or CD8⁺ cytokine T cell responses were significantly bolstered during the effector phase of the vaccine response by OX40 stimulation and maintained at high levels throughout time. As a future development, it would be particularly intriguing to test whether IL-2 induction, currently provided in many combination immunotherapeutic settings through laborious adoptive transfers or high toxicity direct infusions could be complemented or even replaced by providing OX40 co-stimulation using agonistic antibodies.

Due to its capacity to regulate both CD4⁺ and CD8⁺ T cells, OX40 is considered a promising candidate in immunotherapy of persistent viral infections and cancer. However, there is preclinical evidence that anti-OX40 antibodies could induce off-target toxicity causing deleterious immunosuppressive side effects by promoting the accumulation of MDSCs and the production of Th2 cytokines leading to autoimmunity or inflammatory diseases [30-32]. In **chapter 3**, NK cell augmentation and Th2 cytokine activity associated with pro-inflammatory and autoimmune conditions were not triggered by OX40 stimulation. Presumably, combination immunotherapy with peptide-vaccines and low dose schedules of OX40 stimulation did not allow development of off target toxicity events. Moreover, agonistic OX40 antibody treatment has exhibited mild toxicity and no expansion of Tregs, when applied as monotherapy for the treatment of solid tumours [24], compared to the FDA approved CTLA-4 (ipilimumab) or PD-1/ PD-L1 mAb blockade therapies [33,34].

Despite all the positive effects of OX40 agonists it is unlikely that anti-OX40 as single agent will be sufficient to cure patients with different tumour types or viral infections. However, there is great promise that combination immunotherapy incorporating OX40 stimulation and vaccination may be able to increase efficacy. Finally, it would be particularly informative to investigate whether enforced OX40 triggering could complement the therapeutic activity of other costimulatory antibody (anti-4-1BB), checkpoint inhibitor (PD-1/ PD-L1/ CDLA-4) and conventional treatments (i.e. radiotherapy, chemotherapy, cytokine infusions). Encouraging results come from recent studies where anti-OX40 antibody treatment uniquely synergized with PD1-blockade to promote tumour regression in experimental models [34,35].

DIVERSITY IN SPECIFICITY

A valuable observation in **chapter 2** was that the prophylactic efficacy of the distinct MHC class I MCMV epitope containing SLP vaccines was remarkably potentiated when all individual SLPs were combined and administered as a mixture. The efficacy of the MHC class I SLP vaccines to protect against lytic MCMV infection was primarily driven by the breadth of the CD8⁺ T cell responses rather than the magnitude of the response to all individual SLPs. Although the magnitude of the individual antigen-specific CD8⁺ T cell responses was significantly reduced when provided within a mixture and the overall T cell response size was approximately similar to individual SLP vaccines, the ability to contain virus spread was drastically enhanced.

An interesting observation of the present work was the superior capacity of the M38 and m139 MCMV MHC class I epitope containing SLPs to reduce viral titers upon lytic MCMV challenge. M38 and m139 antigens elicit strong inflationary CD8⁺ T cell responses during MCMV infection and presumably play an important role in regulating virus replication and persistence [36,37]. Conceivably, induction of strong CD8⁺ T cell vaccine responses to the M38 and m139 viral antigens in combination with high expression of these antigens on MCMV-infected cells may explain the advanced efficacy of the M38 and m139 specific SLP vaccines in controlling virus spread.

Associations of potent T cell responses with increased breadth leading to decreased viremia or complete viral eradication has been observed in other chronic viral infections [38-41]. A possible explanation for this finding is that development of a robust cytotoxic T cell response with a diverse repertoire of specificities, targeting viral antigens which are expressed at different stages in the viral life cycle increases the likelihood of immune recognition of viral infected cells and the probability of virus dissemination control. Especially, there is evidence that potential encounter of viral infected cells with vaccine-induced T cells of different specificities boost cytokine-mediated direct cell killing and promote CTL cooperation events [42].

Although the exact mechanisms through which immune T cell repertoire diversity influences anti-viral immunity are not yet defined, it is important to consider induction

of a T cell response with increased breadth when designing prophylactic T cell-based vaccines against MCMV infection. Based on the results obtained from the chapter 2 the breadth of the vaccine response may be an important goal in vaccine formulations and a determinant of vaccine efficacy.

SHAKING DOWN THE RIGID WALLS OF PRE-EXISTING IMMUNITY

In many infection models, immunological memory to a viral vector is considered a hindrance for subsequent induction of cell-mediated and humoral immune responses. Various components of the immune system, including neutralizing antibodies, vector-specific T cells and type I IFN-activated NK cells contribute to seriously compromised immune responses against the delivered heterologous antigen [43]. Additionally, pre-existing immunity to a viral vector impacts viral vector's expression level, virus trafficking and alters homing patterns of vaccine induced T cells. Similarly, pre-existing immunity may modulate the magnitude, breadth and immune system's antibody and T cell response to the inserted antigen [44]. CMV-based vectors are endowed with the capacity to stimulate robust CTL and humoral immune responses and have shown unprecedented efficacy against persistent viral infections and cancer [45-47]. Due to the high prevalence rate of CMV in the population, pre-existing immune responses in the host may exist. However, compared to other viruses and traditional viral vector systems (i.e. adenoviruses, lentiviruses), CMV has the ability to re-infect or even superinfect the same host and initiate a second cycle of immune responses.

Thus, in **chapter 5**, the capacity of MCMV-based vector vaccines to induce antitumor responses against HPV-induced cancer was explored. Whether the presence of pre-existing immune responses to MCMV is detrimental or can augment response to the vectored antigen was also investigated. MCMV vectors encoded a dominant MHC class I HPV16 E7 epitope in either inflationary or non-inflationary MCMV epitope regions were tested for both prophylactic and therapeutic efficacy against HPV⁺ tumors in mice. Notably, systemic or subcutaneous MCMV-vectored vaccine administration stimulated vigorous HPV-specific T cell responses recapitulating MCMV response pattern and provided complete and long term protection against tumor challenge in naïve mice.

The therapeutic testing of the MCMV vector vaccine prolonged survival of challenged mice but exhibited moderate immunogenicity when compared to mice with no immunological memory to the vector, eventually leading to minor therapeutic effect. Interestingly, the efficacy of the vaccine was associated with the level of pre-existing humoral immunity to MCMV. Specifically, the initial viral infection dose determined the magnitude of the subsequent antibody and T cell vaccine-specific response. Mice initially infected through the systemic or subcutaneous routes developed strong anti-viral immunity, which severely diminished viral vector's impeding vaccine efficacy. In addition, there is supporting evidence that the therapeutic efficacy of the CMV vectors expressing tumour or viral antigens is significantly attenuated when the host harbours

latent CMV [47]. We and others have previously reported that the initial viral inoculum dose impacts virus immune response [48,49]. Hence, we tested whether different levels of pre-existing immunity to MCMV can influence the subsequent MCMV-vector vaccine efficacy. Strikingly, when initial infection administered orally, weak T cell and antibody responses were elicited, which were significantly boosted following systemic or subcutaneous MCMV- vector vaccine administration. Consistently, the vaccine efficacy was escalated leading to almost complete eradication of tumor-bearing mice. In contrast, strong pre-existing immunity to MCMV was not surpassed by any of the conventional vaccine injection routes tested.

It is not surprising that MCMV – when applied as a vaccine vector containing tumor antigens – is not capable to overcome its own immunogenicity. Even therapeutic targeting of persistent MCMV infection using the strong SLP T cell-based vaccine concept in **appendix of chapter 3** was not capable to boost the considerable high levels of pre-existing virus-specific T cells present 8 weeks after viral infection. Whether the efficacy of the peptide vaccines in therapeutic settings would have been improved in case of low dose pre-exposure to the virus remains to be tested.

Our results suggest that T cell-based vaccine vectors must be designed to generate sufficient quantities of antigen and induce broad cytotoxic CD4⁺ and CD8⁺ T cell responses. Key outstanding research challenges in the use of CMV vectors are dealing with the diversity in the level of pre-existing immunity of pre-exposed individuals and in discovering ways to minimize this response to ensure that all CMV vectored vaccines will reach the threshold levels of protective immunity needed for efficacy. Several strategies, such as augmentation of viral vector dose, immunization route and timing are important factors to circumvent pre-existing immunity to CMV and need to be considered. Moreover, strategies to lower pre-existing immunity through dynamic targeting of critical genes/proteins for virus replication and latency are likely to be particularly informative. Importantly, deletion of virally encoded inhibitors of MHC class I antigen presentation may be essential for blocking establishment of persistent secondary infection or superinfection in CMV rhesus macaques [50]. Finally, the design of distinct CMV serotypes to overcome immunological memory might be a feasible future direction. Analysis of the components involved at the regulation of pre-existing MCMV immunity will help further improve the development of therapeutic CMV vaccines and vector delivery systems for animal and human use. [51]

CONCLUDING REMARKS

T cell mediated protection is multifaceted and driven by several factors. Despite differences among viral infectious diseases, common determinants of the vaccine efficacy is the magnitude, breadth, availability of co-stimulation, tissue location and functionality of CD8⁺ T cells (**chapter 2, 3, 4 and 6**). The role of CD4⁺ T cell “help” in providing direct effector function and regulating the magnitude of the CD8⁺ T cell

response is crucial (**chapter 3**). Therapeutic interventions with MCMV vectors appear to be promising for chronic viral infections (**chapter 5**). A fundamental research question which the findings of this thesis put forward is whether other factors such as innate immunity and B cells are required for sufficient vaccine function and how these factors can be co-manipulated to optimize vaccination.

REFERENCES

1. McCormick AL, Mocarski ES (2015) The immunological underpinnings of vaccinations to prevent cytomegalovirus disease. *Cell Mol Immunol* 12: 170-179.
2. Gamadia LE, Remmerswaal EB, Weel JF, Bemelman F, van Lier RA, et al. (2003) Primary immune responses to human CMV: a critical role for IFN-gamma-producing CD4+ T cells in protection against CMV disease. *Blood* 101: 2686-2692.
3. Polic B, Hengel H, Krmpotic A, Trgovcich J, Pavic I, et al. (1998) Hierarchical and redundant lymphocyte subset control precludes cytomegalovirus replication during latent infection. *J Exp Med* 188: 1047-1054.
4. Reddehase MJ, Mutter W, Munch K, Buhning HJ, Koszinowski UH (1987) CD8-positive T lymphocytes specific for murine cytomegalovirus immediate-early antigens mediate protective immunity. *J Virol* 61: 3102-3108.
5. Kenter GG, Welters MJ, Valentijn AR, Lowik MJ, Berends-van der Meer DM, et al. (2009) Vaccination against HPV-16 oncoproteins for vulvar intraepithelial neoplasia. *N Engl J Med* 361: 1838-1847.
6. Welters MJ, van der Sluis TC, van Meir H, Loof NM, van Ham VJ, et al. (2016) Vaccination during myeloid cell depletion by cancer chemotherapy fosters robust T cell responses. *Sci Transl Med* 8: 334ra352.
7. Wang RF, Wang HY (2017) Immune targets and neoantigens for cancer immunotherapy and precision medicine. *Cell Res* 27: 11-37.
8. Rajasagi M, Shukla SA, Fritsch EF, Keskin DB, DeLuca D, et al. (2014) Systematic identification of personal tumor-specific neoantigens in chronic lymphocytic leukemia. *Blood* 124: 453-462.
9. Jeitziner SM, Walton SM, Torti N, Oxenius A (2013) Adoptive transfer of cytomegalovirus-specific effector CD4+ T cells provides antiviral protection from murine CMV infection. *Eur J Immunol* 43: 2886-2895.
10. Verma S, Weiskopf D, Gupta A, McDonald B, Peters B, et al. (2015) Cytomegalovirus-Specific CD4 T Cells Are Cytolytic and Mediate Vaccine Protection. *J Virol* 90: 650-658.
11. Casazza JP, Betts MR, Price DA, Precopio ML, Ruff LE, et al. (2006) Acquisition of direct antiviral effector functions by CMV-specific CD4+ T lymphocytes with cellular maturation. *J Exp Med* 203: 2865-2877.
12. Pachnio A, Ciaurris M, Begum J, Lal N, Zuo J, et al. (2016) Cytomegalovirus Infection Leads to Development of High Frequencies of Cytotoxic Virus-Specific CD4+ T Cells Targeted to Vascular Endothelium. *PLoS Pathog* 12: e1005832.
13. Ossendorp F, Mengede E, Camps M, Filius R, Melief CJ (1998) Specific T helper cell requirement for optimal induction of cytotoxic T lymphocytes against major histocompatibility complex class II negative tumors. *J Exp Med* 187: 693-702.
14. Shirai M, Pendleton CD, Ahlers J, Takeshita T, Newman M, et al. (1994) Helper-cytotoxic T lymphocyte (CTL) determinant linkage required for priming of anti-HIV CD8+ CTL in vivo with peptide vaccine constructs. *J Immunol* 152: 549-556.
15. Gao FG, Khammanivong V, Liu WJ, Leggatt GR, Frazer IH, et al. (2002) Antigen-specific CD4+ T-cell help is required to activate a memory CD8+ T cell to a fully functional tumor killer cell. *Cancer Res* 62: 6438-6441.

16. Zhang S, Zhang H, Zhao J (2009) The role of CD4 T cell help for CD8 CTL activation. *Biochem Biophys Res Commun* 384: 405-408.
17. Ma SD, Xu X, Jones R, Delecluse HJ, Zumwalde NA, et al. (2016) PD-1/CTLA-4 Blockade Inhibits Epstein-Barr Virus-Induced Lymphoma Growth in a Cord Blood Humanized-Mouse Model. *PLoS Pathog* 12: e1005642.
18. Hodi FS, O'Day SJ, McDermott DF, Weber RW, Sosman JA, et al. (2010) Improved survival with ipilimumab in patients with metastatic melanoma. *N Engl J Med* 363: 711-723.
19. Topalian SL, Taube JM, Anders RA, Pardoll DM (2016) Mechanism-driven biomarkers to guide immune checkpoint blockade in cancer therapy. *Nat Rev Cancer* 16: 275-287.
20. Kaufmann DE, Walker BD (2009) PD-1 and CTLA-4 inhibitory cosignaling pathways in HIV infection and the potential for therapeutic intervention. *J Immunol* 182: 5891-5897.
21. Vezys V, Penaloza-MacMaster P, Barber DL, Ha SJ, Konieczny B, et al. (2011) 4-1BB signaling synergizes with programmed death ligand 1 blockade to augment CD8 T cell responses during chronic viral infection. *J Immunol* 187: 1634-1642.
22. Humphreys IR, Loewendorf A, de Trez C, Schneider K, Benedict CA, et al. (2007) OX40 costimulation promotes persistence of cytomegalovirus-specific CD8 T Cells: A CD4-dependent mechanism. *J Immunol* 179: 2195-2202.
23. Salek-Ardakani S, Moutaftsi M, Sette A, Croft M (2011) Targeting OX40 promotes lung-resident memory CD8 T cell populations that protect against respiratory poxvirus infection. *J Virol* 85: 9051-9059.
24. Curti BD, Kovacsovics-Bankowski M, Morris N, Walker E, Chisholm L, et al. (2013) OX40 is a potent immune-stimulating target in late-stage cancer patients. *Cancer Res* 73: 7189-7198.
25. al-Shamkhani A, Birkeland ML, Puklavec M, Brown MH, James W, et al. (1996) OX40 is differentially expressed on activated rat and mouse T cells and is the sole receptor for the OX40 ligand. *Eur J Immunol* 26: 1695-1699.
26. Croft M, So T, Duan W, Soroosh P (2009) The significance of OX40 and OX40L to T-cell biology and immune disease. *Immunol Rev* 229: 173-191.
27. Green DR, Droin N, Pinkoski M (2003) Activation-induced cell death in T cells. *Immunol Rev* 193: 70-81.
28. Maher S, Toomey D, Condron C, Bouchier-Hayes D (2002) Activation-induced cell death: the controversial role of Fas and Fas ligand in immune privilege and tumour counterattack. *Immunol Cell Biol* 80: 131-137.
29. Pender MP (1999) Activation-induced apoptosis of autoreactive and alloreactive T lymphocytes in the target organ as a major mechanism of tolerance. *Immunol Cell Biol* 77: 216-223.
30. Ueno H, Blanco P (2015) OX40/OX40L axis: not a friend in autoimmunity. *Oncotarget* 6: 21779-21780.
31. Webb GJ, Hirschfield GM, Lane PJ (2016) OX40, OX40L and Autoimmunity: a Comprehensive Review. *Clin Rev Allergy Immunol* 50: 312-332.
32. Gaspal F, Withers D, Saini M, Bekiaris V, McConnell FM, et al. (2011) Abrogation of CD30 and OX40 signals prevents autoimmune disease in FoxP3-deficient mice. *J Exp Med* 208: 1579-1584.
33. Kavanagh B, O'Brien S, Lee D, Hou Y, Weinberg V, et al. (2008) CTLA4 blockade expands FoxP3+ regulatory and activated effector CD4+ T cells in a dose-dependent fashion. *Blood* 112: 1175-1183.
34. Linch SN, McNamara MJ, Redmond WL (2015) OX40 agonists and combination immunotherapy: putting the pedal to the metal. *Frontiers in Oncology* 5.

35. Guo Z, Wang X, Cheng D, Xia Z, Luan M, et al. (2014) PD-1 blockade and OX40 triggering synergistically protects against tumor growth in a murine model of ovarian cancer. *PLoS One* 9: e89350.
36. Menard C, Wagner M, Ruzsics Z, Holak K, Brune W, et al. (2003) Role of murine cytomegalovirus US22 gene family members in replication in macrophages. *J Virol* 77: 5557-5570.
37. Munks MW, Cho KS, Pinto AK, Sierro S, Klenerman P, et al. (2006) Four distinct patterns of memory CD8 T cell responses to chronic murine cytomegalovirus infection. *J Immunol* 177: 450-458.
38. Abdel-Hakeem MS, Shoukry NH (2014) Protective immunity against hepatitis C: many shades of gray. *Front Immunol* 5: 274.
39. Radebe M, Gounder K, Mokgoro M, Ndhlovu ZM, Mncube Z, et al. (2015) Broad and persistent Gag-specific CD8+ T-cell responses are associated with viral control but rarely drive viral escape during primary HIV-1 infection. *Aids* 29: 23-33.
40. Hu X, Valentin A, Dayton F, Kulkarni V, Alicea C, et al. (2016) DNA Prime-Boost Vaccine Regimen To Increase Breadth, Magnitude, and Cytotoxicity of the Cellular Immune Responses to Subdominant Gag Epitopes of Simian Immunodeficiency Virus and HIV. *J Immunol* 197: 3999-4013.
41. Barouch DH, O'Brien KL, Simmons NL, King SL, Abbink P, et al. (2010) Mosaic HIV-1 vaccines expand the breadth and depth of cellular immune responses in rhesus monkeys. *Nat Med* 16: 319-323.
42. Halle S, Keyser KA, Stahl FR, Busche A, Marquardt A, et al. (2016) In Vivo Killing Capacity of Cytotoxic T Cells Is Limited and Involves Dynamic Interactions and T Cell Cooperativity. *Immunity* 44: 233-245.
43. Saxena M, Van TT, Baird FJ, Coloe PJ, Smooker PM (2013) Pre-existing immunity against vaccine vectors--friend or foe? *Microbiology* 159: 1-11.
44. McCoy K, Tatsis N, Koriath-Schmitz B, Lasaro MO, Hensley SE, et al. (2007) Effect of preexisting immunity to adenovirus human serotype 5 antigens on the immune responses of nonhuman primates to vaccine regimens based on human- or chimpanzee-derived adenovirus vectors. *J Virol* 81: 6594-6604.
45. Hansen SG, Piatak M, Jr., Ventura AB, Hughes CM, Gilbride RM, et al. (2013) Immune clearance of highly pathogenic SIV infection. *Nature* 502: 100-104.
46. Qiu Z, Huang H, Grenier JM, Perez OA, Smilowitz HM, et al. (2015) Cytomegalovirus-Based Vaccine Expressing a Modified Tumor Antigen Induces Potent Tumor-Specific CD8(+) T-cell Response and Protects Mice from Melanoma. *Cancer Immunol Res* 3: 536-546.
47. Xu G, Smith T, Grey F, Hill AB (2013) Cytomegalovirus-based cancer vaccines expressing TRP2 induce rejection of melanoma in mice. *Biochem Biophys Res Commun* 437: 287-291.
48. Akondy RS, Johnson PL, Nakaya HI, Edupuganti S, Mulligan MJ, et al. (2015) Initial viral load determines the magnitude of the human CD8 T cell response to yellow fever vaccination. *Proc Natl Acad Sci U S A* 112: 3050-3055.
49. Redeker A, Welten SP, Arens R (2014) Viral inoculum dose impacts memory T-cell inflation. *Eur J Immunol* 44: 1046-1057.
50. Hansen SG, Powers CJ, Richards R, Ventura AB, Ford JC, et al. (2010) Evasion of CD8+ T cells is critical for superinfection by cytomegalovirus. *Science* 328: 102-106.
51. Fausther-Bovendo H, Kobinger GP (2014) Pre-existing immunity against Ad vectors: humoral, cellular, and innate response, what's important? *Hum Vaccin Immunother* 10: 2875-2884.
52. Panagioti E, Redeker A, van Duikeren S, Franken KL, Drijfhout JW, et al. (2016) The Breadth of Synthetic Long Peptide Vaccine-Induced CD8+ T Cell Responses Determines the Efficacy against Mouse Cytomegalovirus Infection. *PLoS Pathog* 12: e1005895.

