



Universiteit
Leiden
The Netherlands

Wrapping up : nidovirus membrane structures and innate immunity

Oudshoorn, D.

Citation

Oudshoorn, D. (2017, December 28). *Wrapping up : nidovirus membrane structures and innate immunity*. Retrieved from <https://hdl.handle.net/1887/59466>

Version: Not Applicable (or Unknown)

License: [Licence agreement concerning inclusion of doctoral thesis in the Institutional Repository of the University of Leiden](#)

Downloaded from: <https://hdl.handle.net/1887/59466>

Note: To cite this publication please use the final published version (if applicable).

Cover Page



Universiteit Leiden



The following handle holds various files of this Leiden University dissertation:
<http://hdl.handle.net/1887/59466>

Author: Oudshoorn, D.

Title: Wrapping up : nidovirus membrane structures and innate immunity

Issue Date: 2017-11-28

REFERENCES

1. Zirkel F, Roth H, Kurth A, Drosten C, Ziebuhr J, Junglen S. 2013. Identification and characterization of genetically divergent members of the newly established family Mesoniviridae. *Journal of virology* 87:6346-6358.
2. Gorbalenya AE, Snijder EJ, Cowley JA, Ziebuhr J, Faaberg K, Enjuanes L, Rottier PJ, de Groot RJ, Perlman S. 2011. Virus Taxonomy: The Classification and Nomenclature of Viruses – The 9th Report of the ICTV. Elsevier.
3. Lauber C, Goeman JJ, Parquet Mdel C, Nga PT, Snijder EJ, Morita K, Gorbalenya AE. 2013. The footprint of genome architecture in the largest genome expansion in RNA viruses. *PLoS pathogens* 9:e1003500.
4. Snijder EJ, Kikkert M, Fang Y. 2013. Arterivirus molecular biology and pathogenesis. *The Journal of general virology* 94:2141-2163.
5. Bryans JT, Crowe ME, Doll ER, McCollum WH. 1957. Isolation of a filterable agent causing arteritis of horses and abortion by mares; its differentiation from the equine abortion (influenza) virus. *The Cornell veterinarian* 47:3-41.
6. Balasuriya UB, Go YY, MacLachlan NJ. 2013. Equine arteritis virus. *Veterinary microbiology* 167:93-122.
7. Neumann EJ, Kliebenstein JB, Johnson CD, Mabry JW, Bush EJ, Seitzinger AH, Green AL, Zimmerman JJ. 2005. Assessment of the economic impact of porcine reproductive and respiratory syndrome on swine production in the United States. *J Am Vet Med Assoc* 227:385-392.
8. Han J, Zhou L, Ge X, Guo X, Yang H. 2017. Pathogenesis and control of the Chinese highly pathogenic porcine reproductive and respiratory syndrome virus. *Veterinary microbiology*.
9. Lunney JK, Fang Y, Ladwig A, Chen N, Li Y, Rowland B, Renukaradhya GJ. 2016. Porcine Reproductive and Respiratory Syndrome Virus (PRRSV): Pathogenesis and Interaction with the Immune System. *Annu Rev Anim Biosci* 4:129-154.
10. van Dinten LC, den Boon JA, Wassenaar AL, Spaan WJ, Snijder EJ. 1997. An infectious arterivirus cDNA clone: identification of a replicase point mutation that abolishes discontinuous mRNA transcription. *Proceedings of the National Academy of Sciences of the United States of America* 94:991-996.
11. Ge XY, Li JL, Yang XL, Chmura AA, Zhu G, Epstein JH, Mazet JK, Hu B, Zhang W, Peng C, Zhang YJ, Luo CM, Tan B, Wang N, Zhu Y, Crameri G, Zhang SY, Wang LF, Daszak P, Shi ZL. 2013. Isolation and characterization of a bat SARS-like coronavirus that uses the ACE2 receptor. *Nature* 503:535-538.
12. Zaki AM, van Boheemen S, Bestebroer TM, Osterhaus AD, Fouchier RA. 2012. Isolation of a novel coronavirus from a man with pneumonia in Saudi Arabia. *The New England journal of medicine* 367:1814-1820.
13. de Wit E, van Doremale N, Falzarano D, Munster VJ. 2016. SARS and MERS: recent insights into emerging coronaviruses. *Nature reviews. Microbiology* 14:523-534.
14. Graham RL, Donaldson EF, Baric RS. 2013. A decade after SARS: strategies for controlling emerging coronaviruses. *Nature reviews. Microbiology* 11:836-848.
15. Zumla A, Chan JF, Azhar EI, Hui DS, Yuen KY. 2016. Coronaviruses – drug discovery and therapeutic options. *Nature reviews. Drug discovery* 15:327-347.
16. Angelini MM, Neuman BW, Buchmeier MJ. 2014. Untangling membrane rearrangement in the *nidovirales*. *DNA and cell biology* 33:122-127.
17. den Boon JA, Diaz A, Ahlquist P. 2010.

- Cytoplasmic viral replication complexes. *Cell host & microbe* 8:77-85.
- 18. Miller S, Krijnse-Locker J. 2008. Modification of intracellular membrane structures for virus replication. *Nature reviews. Microbiology* 6:363-374.
 - 19. Romero-Brey I, Bartenschlager R. 2016. Endoplasmic Reticulum: The Favorite Intracellular Niche for Viral Replication and Assembly. *Viruses* 8.
 - 20. Koops K, Barcena M, Limpens RW, Koster AJ, Mommaas AM, Snijder EJ. 2012. Ultrastructural characterization of arterivirus replication structures: reshaping the endoplasmic reticulum to accommodate viral RNA synthesis. *Journal of virology* 86:2474-2487.
 - 21. Koops K, Kikkert M, Worm SH, Zevenhoven-Dobbe JC, van der Meer Y, Koster AJ, Mommaas AM, Snijder EJ. 2008. SARS-coronavirus replication is supported by a reticulovesicular network of modified endoplasmic reticulum. *PLoS biology* 6:e226.
 - 22. Maier HJ, Hawes PC, Cottam EM, Mantell J, Verkade P, Monaghan P, Wileman T, Britton P. 2013. Infectious bronchitis virus generates spherules from zippered endoplasmic reticulum membranes. *mBio* 4:e00801-00813.
 - 23. Romero-Brey I, Merz A, Chiramel A, Lee JY, Chlanda P, Haselman U, Santarella-Mellwig R, Habermann A, Hoppe S, Kallis S, Walther P, Antony C, Krijnse-Locker J, Bartenschlager R. 2012. Three-dimensional architecture and biogenesis of membrane structures associated with hepatitis C virus replication. *PLoS pathogens* 8:e1003056.
 - 24. Snijder EJ, van Tol H, Roos N, Pedersen KW. 2001. Non-structural proteins 2 and 3 interact to modify host cell membranes during the formation of the arterivirus replication complex. *The Journal of general virology* 82:985-994.
 - 25. Suhy DA, Giddings TH, Jr., Kirkegaard K. 2000. Remodeling the endoplasmic reticulum by po-
- liovirus infection and by individual viral proteins: an autophagy-like origin for virus-induced vesicles. *Journal of virology* 74:8953-8965.
- 26. Salonen A, Vasiljeva L, Merits A, Magden J, Jokitalo E, Kaariainen L. 2003. Properly folded nonstructural polyprotein directs the semliki forest virus replication complex to the endosomal compartment. *Journal of virology* 77:1691-1702.
 - 27. Schwartz M, Chen J, Janda M, Sullivan M, den Boon J, Ahlquist P. 2002. A positive-strand RNA virus replication complex parallels form and function of retrovirus capsids. *Molecular cell* 9:505-514.
 - 28. Angelini MM, Akhlaghpour M, Neuman BW, Buchmeier MJ. 2013. Severe acute respiratory syndrome coronavirus nonstructural proteins 3, 4, and 6 induce double-membrane vesicles. *mBio* 4.
 - 29. Bowie AG, Unterholzner L. 2008. Viral evasion and subversion of pattern-recognition receptor signalling. *Nature reviews. Immunology* 8:911-922.
 - 30. Gurtler C, Bowie AG. 2013. Innate immune detection of microbial nucleic acids. *Trends in microbiology* 21:413-420.
 - 31. Yoneyama M, Onomoto K, Jogi M, Akaboshi T, Fujita T. 2015. Viral RNA detection by RIG-I-like receptors. *Current opinion in immunology* 32:48-53.
 - 32. Sancho-Shimizu V, Perez de Diego R, Jouanguy E, Zhang SY, Casanova JL. 2011. Inborn errors of anti-viral interferon immunity in humans. *Current opinion in virology* 1:487-496.
 - 33. Chan YK, Gack MU. 2016. Viral evasion of intracellular DNA and RNA sensing. *Nature reviews. Microbiology* 14:360-373.
 - 34. Nielsen R, Bustamante C, Clark AG, Glanowski S, Sackton TB, Hubisz MJ, Fledel-Alon A, Tanenbaum DM, Civello D, White TJ, J JS, Adams MD, Cargill M. 2005. A scan for positively selected genes in the genomes of humans and chim-

- panzees. PLoS biology 3:e170.
35. Paul D, Bartenschlager R. 2013. Architecture and biogenesis of plus-strand RNA virus replication factories. World journal of virology 2:32-48.
 36. Romero-Brey I, Bartenschlager R. 2014. Membranous replication factories induced by plus-strand RNA viruses. Viruses 6:2826-2857.
 37. Neufeldt CJ, Joyce MA, Van Buuren N, Levin A, Kirkegaard K, Gale M, Jr., Tyrrell DL, Wozniak RW. 2016. The Hepatitis C Virus-Induced Membranous Web and Associated Nuclear Transport Machinery Limit Access of Pattern Recognition Receptors to Viral Replication Sites. PLoS pathogens 12:e1005428.
 38. Egger D, Wolk B, Gosert R, Bianchi L, Blum HE, Moradpour D, Bienz K. 2002. Expression of hepatitis C virus proteins induces distinct membrane alterations including a candidate viral replication complex. Journal of virology 76:5974-5984.
 39. Romero-Brey I, Berger C, Kallis S, Kolovou A, Paul D, Lohmann V, Bartenschlager R. 2015. NS5A Domain 1 and Polyprotein Cleavage Kinetics Are Critical for Induction of Double-Membrane Vesicles Associated with Hepatitis C Virus Replication. mBio 6:e00759.
 40. Nagy PD, Pogany J. 2012. The dependence of viral RNA replication on co-opted host factors. Nature reviews. Microbiology 10:137-149.
 41. Belov GA, van Kuppeveld FJ. 2012. (+)RNA viruses rewire cellular pathways to build replication organelles. Current opinion in virology 2:740-747.
 42. Reiss S, Rebhan I, Backes P, Romero-Brey I, Erfle H, Matula P, Kaderali L, Poenisch M, Blankenburg H, Hiet MS, Longerich T, Diehl S, Ramirez F, Balla T, Rohr K, Kaul A, Buhler S, Pepperkok R, Lengauer T, Albrecht M, Eils R, Schirmacher P, Lohmann V, Bartenschlager R. 2011. Recruitment and activation of a lipid kinase by hepatitis C virus NS5A is essential for integrity of the membranous replication compartment.
 43. Berger KL, Kelly SM, Jordan TX, Tartell MA, Randall G. 2011. Hepatitis C virus stimulates the phosphatidylinositol 4-kinase III alpha-dependent phosphatidylinositol 4-phosphate production that is essential for its replication. Journal of virology 85:8870-8883.
 44. Hsu NY, Illytska O, Belov G, Santiana M, Chen YH, Takvorian PM, Pau C, van der Schaaf H, Kaushik-Basu N, Balla T, Cameron CE, Ehrenfeld E, van Kuppeveld FJ, Altan-Bonnet N. 2010. Viral reorganization of the secretory pathway generates distinct organelles for RNA replication. Cell 141:799-811.
 45. Diaz A, Ahlquist P. 2012. Role of host reticulon proteins in rearranging membranes for positive-strand RNA virus replication. Current opinion in microbiology 15:519-524.
 46. Diaz A, Zhang J, Ollwerther A, Wang X, Ahlquist P. 2015. Host ESCRT proteins are required for bromovirus RNA replication compartment assembly and function. PLoS pathogens 11:e1004742.
 47. Barcena M, Koster AJ. 2009. Electron tomography in life science. Seminars in cell & developmental biology 20:920-930.
 48. Kopek BG, Perkins G, Miller DJ, Ellisman MH, Ahlquist P. 2007. Three-dimensional analysis of a viral RNA replication complex reveals a virus-induced mini-organelle. PLoS biology 5:e220.
 49. Maier HJ, Cottam EM, Stevenson-Leggett P, Wilkinson JA, Harte CJ, Wileman T, Britton P. 2013. Visualizing the autophagy pathway in avian cells and its application to studying infectious bronchitis virus. Autophagy 9:496-509.
 50. Welsch S, Miller S, Romero-Brey I, Merz A, Bleck CK, Walther P, Fuller SD, Antony C, Krijnse Locker J, Bartenschlager R. 2009. Composition and three-dimensional architecture of the dengue virus replication and assembly sites. Cell host & microbe 5:365-375.

51. Gillespie LK, Hoenen A, Morgan G, Mackenzie JM. 2010. The endoplasmic reticulum provides the membrane platform for biogenesis of the flavivirus replication complex. *Journal of virology* 84:10438-10447.
52. Offerdahl DK, Dorward DW, Hansen BT, Bloom ME. 2012. A three-dimensional comparison of tick-borne flavivirus infection in mammalian and tick cell lines. *PloS one* 7:e47912.
53. Miorin L, Romero-Brey I, Maiuri P, Hoppe S, Krijnse-Locker J, Bartenschlager R, Marcello A. 2013. Three-dimensional architecture of tick-borne encephalitis virus replication sites and trafficking of the replicated RNA. *Journal of virology* 87:6469-6481.
54. Junjhon J, Pennington JG, Edwards TJ, Perera R, Lanman J, Kuhn RJ. 2014. Ultrastructural characterization and three-dimensional architecture of replication sites in dengue virus-infected mosquito cells. *Journal of virology* 88:4687-4697.
55. Fontana J, Lopez-Iglesias C, Tzeng WP, Frey TK, Fernandez JJ, Risco C. 2010. Three-dimensional structure of Rubella virus factories. *Virology* 405:579-591.
56. Limpens RW, van der Schaar HM, Kumar D, Koster AJ, Snijder EJ, van Kuppeveld FJ, Barcena M. 2011. The transformation of enterovirus replication structures: a three-dimensional study of single- and double-membrane compartments. *mBio* 2.
57. Cao X, Jin X, Zhang X, Li Y, Wang C, Wang X, Hong J, Wang X, Li D, Zhang Y. 2015. Morphogenesis of Endoplasmic Reticulum Membrane-Invaginated Vesicles during Beet Black Scorch Virus Infection: Role of Auxiliary Replication Protein and New Implications of Three-Dimensional Architecture. *Journal of virology* 89:6184-6195.
58. Grimley PM, Berezesky IK, Friedman RM. 1968. Cytoplasmic structures associated with an arbovirus infection: loci of viral ribonucleic acid synthesis. *Journal of virology* 2:1326-1338.
59. Froshauer S, Kartenbeck J, Helenius A. 1988. Alphavirus RNA replicase is located on the cytoplasmic surface of endosomes and lysosomes. *The Journal of cell biology* 107:2075-2086.
60. Kujala P, Ikaheimonen A, Ehsani N, Vihinen H, Auvinen P, Kaariainen L. 2001. Biogenesis of the Semliki Forest virus RNA replication complex. *Journal of virology* 75:3873-3884.
61. Frolova EI, Gorchakov R, Pereboeva L, Atasheva S, Frolov I. 2010. Functional Sindbis virus replicative complexes are formed at the plasma membrane. *Journal of virology* 84:11679-11695.
62. Laliberte JF, Sanfacon H. 2010. Cellular remodeling during plant virus infection. *Annual review of phytopathology* 48:69-91.
63. Diaz A, Gallei A, Ahlquist P. 2012. Bromovirus RNA replication compartment formation requires concerted action of 1a's self-interacting RNA capping and helicase domains. *Journal of virology* 86:821-834.
64. Ahlquist P. 2006. Parallels among positive-strand RNA viruses, reverse-transcribing viruses and double-stranded RNA viruses. *Nature reviews. Microbiology* 4:371-382.
65. Belov GA, Nair V, Hansen BT, Hoyt FH, Fischer ER, Ehrenfeld E. 2012. Complex dynamic development of poliovirus membranous replication complexes. *Journal of virology* 86:302-312.
66. Ferraris P, Beaumont E, Uzbekov R, Brand D, Gaillard J, Blanchard E, Roingeard P. 2013. Sequential biogenesis of host cell membrane rearrangements induced by hepatitis C virus infection. *Cell Mol Life Sci* 70:1297-1306.
67. Paul D, Hoppe S, Saher G, Krijnse-Locker J, Bartenschlager R. 2013. Morphological and biochemical characterization of the membranous hepatitis C virus replication compartment. *Journal of virology* 87:10612-10627.
68. Cowley JA, Dimmock CM, Walker PJ. 2002. Gill-associated nidovirus of *Penaeus monodon* prawns transcribes 3'-coterminal subgenomic

- mRNAs that do not possess 5'-leader sequences. *The Journal of general virology* 83:927-935.
69. Knoops K, Swett-Tapia C, van den Worm SH, Te Velthuis AJ, Koster AJ, Mommaas AM, Snijder EJ, Kikkert M. 2010. Integrity of the early secretory pathway promotes, but is not required for, severe acute respiratory syndrome coronavirus RNA synthesis and virus-induced remodeling of endoplasmic reticulum membranes. *Journal of virology* 84:833-846.
 70. Cavanagh D. 1997. *Nidovirales*: a new order comprising Coronaviridae and Arteriviridae. *Archives of virology* 142:629-633.
 71. Lauck M, Sibley SD, Hyeroba D, Tumukunde A, Weny G, Chapman CA, Ting N, Switzer WM, Kuhn JH, Friedrich TC, O'Connor DH, Goldberg TL. 2013. Exceptional simian hemorrhagic fever virus diversity in a wild African primate community. *Journal of virology* 87:688-691.
 72. Dunowska M, Biggs PJ, Zheng T, Perrott MR. 2012. Identification of a novel nidovirus associated with a neurological disease of the Australian brushtail possum (*Trichosurus vulpecula*). *Veterinary microbiology* 156:418-424.
 73. Kuhn JH, Lauck M, Bailey AL, Shchetinin AM, Vishnevskaya TV, Bao Y, Ng TF, LeBreton M, Schneider BS, Gillis A, Tamoufe U, Diffo JL, Takuo JM, Kondov NO, Coffey LL, Wolfe ND, Delwart E, Clawson AN, Postnikova E, Bollinger L, Lackemeyer MG, Radoshitzky SR, Palacios G, Wada J, Shevtsova ZV, Jahrling PB, Lapin BA, Deriabin PG, Dunowska M, Alkhovsky SV, Rogers J, Friedrich TC, O'Connor DH, Goldberg TL. 2015. Reorganization and expansion of the nidoviral family Arteriviridae. *Archives of virology*.
 74. Brierley I, Digard P, Inglis SC. 1989. Characterization of an efficient coronavirus ribosomal frameshifting signal: requirement for an RNA pseudoknot. *Cell* 57:537-547.
 75. den Boon JA, Snijder EJ, Chirnside ED, de Vries AA, Horzinek MC, Spaan WJ. 1991. Equine arte-
 - ritis virus is not a togavirus but belongs to the coronaviruseslike superfamily. *Journal of virology* 65:2910-2920.
 76. Ziebuhr J, Snijder EJ, Gorbunova AE. 2000. Virus-encoded proteinases and proteolytic processing in the *Nidovirales*. *The Journal of general virology* 81:853-879.
 77. Li Y, Tas A, Sun Z, Snijder EJ, Fang Y. 2014. Proteolytic processing of the porcine reproductive and respiratory syndrome virus replicase. *Virus research*.
 78. Gorbunova AE, Enjuanes L, Ziebuhr J, Snijder EJ. 2006. *Nidovirales*: evolving the largest RNA virus genome. *Virus research* 117:17-37.
 79. Fang Y, Snijder EJ. 2010. The PRRSV replicase: exploring the multifunctionality of an intriguing set of nonstructural proteins. *Virus research* 154:61-76.
 80. Lehmann KC, Snijder EJ, Posthuma CC, Gorbunova AE. 2015. What we know but do not understand about nidovirus helicases. *Virus research* 202:12-32.
 81. Lehmann KC, Gulyaeva A, Zevenhoven-Dobbe JC, Janssen GM, Ruben M, Overkleeft HS, van Veelen PA, Samborski DV, Kravchenko AA, Leontovich AM, Sidorov IA, Snijder EJ, Posthuma CC, Gorbunova AE. 2015. Discovery of an essential nucleotidylating activity associated with a newly delineated conserved domain in the RNA polymerase-containing protein of all nidoviruses. *Nucleic acids research* 43:8416-8434.
 82. Gorbunova AE, Koonin EV, Donchenko AP, Blinov VM. 1989. Coronavirus genome: prediction of putative functional domains in the non-structural polyprotein by comparative amino acid sequence analysis. *Nucleic acids research* 17:4847-4861.
 83. Vatter HA, Di H, Donaldson EF, Radu GU, Maines TR, Brinton MA. 2014. Functional analyses of the three simian hemorrhagic fever virus nonstructural protein 1 papain-like proteases. *Journal of*

- virology 88:9129-9140.
84. Wassenaar AL, Spaan WJ, Gorbalenya AE, Snijder EJ. 1997. Alternative proteolytic processing of the arterivirus replicase ORF1a polyprotein: evidence that NSP2 acts as a cofactor for the NSP4 serine protease. *Journal of virology* 71:9313-9322.
85. Li Y, Tas A, Snijder EJ, Fang Y. 2012. Identification of porcine reproductive and respiratory syndrome virus ORF1a-encoded non-structural proteins in virus-infected cells. *The Journal of general virology* 93:829-839.
86. van Aken D, Zevenhoven-Dobbe J, Gorbalenya AE, Snijder EJ. 2006. Proteolytic maturation of replicase polyprotein pp1a by the nsp4 main proteinase is essential for equine arteritis virus replication and includes internal cleavage of nsp7. *The Journal of general virology* 87:3473-3482.
87. Faaberg KS, Plagemann PG. 1996. Membrane association of the C-terminal half of the open reading frame 1a protein of lactate dehydrogenase-elevating virus. *Archives of virology* 141:1337-1348.
88. Pedersen KW, van der Meer Y, Roos N, Snijder EJ. 1999. Open reading frame 1a-encoded subunits of the arterivirus replicase induce endoplasmic reticulum-derived double-membrane vesicles which carry the viral replication complex. *Journal of virology* 73:2016-2026.
89. Posthuma CC, Pedersen KW, Lu Z, Joosten RG, Roos N, Zevenhoven-Dobbe JC, Snijder EJ. 2008. Formation of the arterivirus replication/transcription complex: a key role for nonstructural protein 3 in the remodeling of intracellular membranes. *Journal of virology* 82:4480-4491.
90. Kappes MA, Miller CL, Faaberg KS. 2015. Porcine reproductive and respiratory syndrome virus nonstructural protein 2 (nsp2) topology and selective isoform integration in artificial membranes. *Virology* 481:51-62.
91. Snijder EJ, Wassenaar AL, Spaan WJ, Gorbalenya AE. 1995. The arterivirus Nsp2 protease. An unusual cysteine protease with primary structure similarities to both papain-like and chymotrypsin-like proteases. *The Journal of biological chemistry* 270:16671-16676.
92. Kanjanahaluethai A, Baker SC. 2000. Identification of mouse hepatitis virus papain-like proteinase 2 activity. *Journal of virology* 74:7911-7921.
93. Ziebuhr J, Thiel V, Gorbalenya AE. 2001. The autocatalytic release of a putative RNA virus transcription factor from its polyprotein precursor involves two paralogous papain-like proteases that cleave the same peptide bond. *The Journal of biological chemistry* 276:33220-33232.
94. Harcourt BH, Jukneliene D, Kanjanahaluethai A, Bechill J, Severson KM, Smith CM, Rota PA, Baker SC. 2004. Identification of severe acute respiratory syndrome coronavirus replicase products and characterization of papain-like protease activity. *Journal of virology* 78:13600-13612.
95. Oostra M, te Lintelo EG, Deijs M, Verheije MH, Rottier PJ, de Haan CA. 2007. Localization and membrane topology of coronavirus nonstructural protein 4: involvement of the early secretory pathway in replication. *Journal of virology* 81:12323-12336.
96. Hagemeijer MC, Monastyrskaya I, Griffith J, van der Sluijs P, Voortman J, van Bergen en Henegouwen PM, Vonk AM, Rottier PJ, Reggiori F, de Haan CA. 2014. Membrane rearrangements mediated by coronavirus nonstructural proteins 3 and 4. *Virology* 458-459:125-135.
97. van Dinten LC, Wassenaar AL, Gorbalenya AE, Spaan WJ, Snijder EJ. 1996. Processing of the equine arteritis virus replicase ORF1b protein: identification of cleavage products containing the putative viral polymerase and helicase domains. *Journal of virology* 70:6625-6633.
98. van der Meer Y, van Tol H, Locker JK, Snijder EJ. 1998. ORF1a-encoded replicase subunits are

- involved in the membrane association of the arterivirus replication complex. *Journal of virology* 72:6689-6698.
99. van Hemert MJ, de Wilde AH, Gorbalyena AE, Snijder EJ. 2008. The *in vitro* RNA synthesizing activity of the isolated arterivirus replication/transcription complex is dependent on a host factor. *The Journal of biological chemistry* 283:16525-16536.
100. Denison MR, Spaan WJ, van der Meer Y, Gibson CA, Sims AC, Prentice E, Lu XT. 1999. The putative helicase of the coronavirus mouse hepatitis virus is processed from the replicase gene polyprotein and localizes in complexes that are active in viral RNA synthesis. *Journal of virology* 73:6862-6871.
101. Shi ST, Schiller JJ, Kanjanahaluethai A, Baker SC, Oh JW, Lai MM. 1999. Colocalization and membrane association of murine hepatitis virus gene 1 products and De novo-synthesized viral RNA in infected cells. *Journal of virology* 73:5957-5969.
102. van der Meer Y, Snijder EJ, Dobbe JC, Schleich S, Denison MR, Spaan WJ, Locker JK. 1999. Localization of mouse hepatitis virus nonstructural proteins and RNA synthesis indicates a role for late endosomes in viral replication. *Journal of virology* 73:7641-7657.
103. Bost AG, Carnahan RH, Lu XT, Denison MR. 2000. Four proteins processed from the replicase gene polyprotein of mouse hepatitis virus colocalize in the cell periphery and adjacent to sites of virion assembly. *Journal of virology* 74:3379-3387.
104. Brockway SM, Clay CT, Lu XT, Denison MR. 2003. Characterization of the expression, intracellular localization, and replication complex association of the putative mouse hepatitis virus RNA-dependent RNA polymerase. *Journal of virology* 77:10515-10527.
105. Prentice E, McAuliffe J, Lu X, Subbarao K, Denison MR. 2004. Identification and characterization of severe acute respiratory syndrome coronavirus replicase proteins. *Journal of virology* 78:9977-9986.
106. Snijder EJ, van der Meer Y, Zevenhoven-Dobbe J, Onderwater JJ, van der Meulen J, Koerten HK, Mommaas AM. 2006. Ultrastructure and origin of membrane vesicles associated with the severe acute respiratory syndrome coronavirus replication complex. *Journal of virology* 80:5927-5940.
107. van Hemert MJ, van den Worm SH, Knoops K, Mommaas AM, Gorbalyena AE, Snijder EJ. 2008. SARS-coronavirus replication/transcription complexes are membrane-protected and need a host factor for activity *in vitro*. *PLoS pathogens* 4:e1000054.
108. Breese SSJ, McCollum WH. 1970. Electron microscopy characterization of equine arteritis virus, p. 133-139. In Bryans JT, Gerber H (ed.), *Proceedings of the 2nd International Conference on Equine Infectious Diseases*. S. Karger, Basel.
109. Wood O, Tauraso N, Liebhaber H. 1970. Electron microscopic study of tissue cultures infected with simian haemorrhagic fever virus. *The Journal of general virology* 7:129-136.
110. Stueckemann JA, Ritzl DM, Holth M, Smith MS, Swart WJ, Cafruny WA, Plagemann GW. 1982. Replication of lactate dehydrogenase-elevating virus in macrophages. 1. Evidence for cytocidal replication. *The Journal of general virology* 59:245-262.
111. Wada R, Fukunaga Y, Kondo T, Kanemaru T. 1995. Ultrastructure and immuno-cytochemistry of BHK-21 cells infected with a modified Bucyrus strain of equine arteritis virus. *Archives of virology* 140:1173-1180.
112. Weiland FH, Granzow M, Wieczorek-Krohmer M, Weiland E. 1995. Electron microscopic studies on the morphogenesis of PRRSV in infected cells -comparative studies, p. 499-502. In Schwyzler M, Ackermann M, Bertoni G, Kocherhanss R,

- McCullough K, Engels M, Wittek R, Zanoni R (ed.), Immunobiology of viral infections. Proceedings of the 3rd Congress of the European Society of Veterinary Virology.
113. Pol JM, Wagenaar F, Reus JE. 1997. Comparative morphogenesis of three PRRS virus strains. *Veterinary microbiology* 55:203-208.
114. Metwally S, Mohamed F, Faaberg K, Burrage T, Prarat M, Moran K, Bracht A, Mayr G, Berninger M, Koster L, To TL, Nguyen VL, Reising M, Landgraf J, Cox L, Lubroth J, Carrillo C. 2010. Pathogenicity and molecular characterization of emerging porcine reproductive and respiratory syndrome virus in Vietnam in 2007. *Transboundary and emerging diseases* 57:315-329.
115. Wieringa R, de Vries AA, van der Meulen J, Godeke GJ, Onderwater JJ, van Tol H, Koerten HK, Mommaas AM, Snijder EJ, Rottier PJ. 2004. Structural protein requirements in equine arteritis virus assembly. *Journal of virology* 78:13019-13027.
116. McDonald K. 2007. Cryopreparation methods for electron microscopy of selected model systems. *Methods in cell biology* 79:23-56.
117. Leapman RD, Aronova MA. 2007. Localizing specific elements bound to macromolecules by EFTEM. *Methods in cell biology* 79:593-613.
118. Friedman RM, Sreevalsan T. 1970. Membrane binding of input arbovirus ribonucleic acid: effect of interferon or cycloheximide. *Journal of virology* 6:169-175.
119. Kujala P, Ahola T, Ehsani N, Auvinen P, Vihinen H, Kaariainen L. 1999. Intracellular distribution of rubella virus nonstructural protein P150. *Journal of virology* 73:7805-7811.
120. Quinkert D, Bartenschlager R, Lohmann V. 2005. Quantitative analysis of the hepatitis C virus replication complex. *Journal of virology* 79:13594-13605.
121. Svoboda D, Nielson A, Werber A, Higginson J. 1962. An electron microscopic study of viral hepatitis in mice. *The American journal of pathology* 41:205-224.
122. David-Ferreira JF, Manaker RA. 1965. An Electron Microscope Study of the Development of a Mouse Hepatitis Virus in Tissue Culture Cells. *The Journal of cell biology* 24:57-78.
123. Orenstein JM, Banach B, Baker SC. 2008. Morphogenesis of Coronavirus HCoV-NL63 in Cell Culture: A Transmission Electron Microscopic Study. *The open infectious diseases journal* 2:52-58.
124. Lundin A, Dijkman R, Bergstrom T, Kann N, Adamiak B, Hannoun C, Kindler E, Jonsdottir HR, Muth D, Kint J, Forlenza M, Muller MA, Drosten C, Thiel V, Trybala E. 2014. Targeting membrane-bound viral RNA synthesis reveals potent inhibition of diverse coronaviruses including the middle East respiratory syndrome virus. *PLoS pathogens* 10:e1004166.
125. Gosert R, Kanjanahaluethai A, Egger D, Bienz K, Baker SC. 2002. RNA replication of mouse hepatitis virus takes place at double-membrane vesicles. *Journal of virology* 76:3697-3708.
126. Goldsmith CS, Tatti KM, Ksiazek TG, Rollin PE, Comer JA, Lee WW, Rota PA, Bankamp B, Bellini WJ, Zaki SR. 2004. Ultrastructural characterization of SARS coronavirus. *Emerging infectious diseases* 10:320-326.
127. Ulasli M, Verheije MH, de Haan CA, Reggiori F. 2010. Qualitative and quantitative ultrastructural analysis of the membrane rearrangements induced by coronavirus. *Cellular microbiology* 12:844-861.
128. de Wilde AH, Raj VS, Oudshoorn D, Bestebroer TM, van Nieuwkoop S, Limpens RW, Posthuma CC, van der Meer Y, Barcena M, Haagmans BL, Snijder EJ, van den Hoogen BG. 2013. MERS-coronavirus replication induces severe *in vitro* cytopathology and is strongly inhibited by cyclosporin A or interferon-alpha treatment. *The Journal of general virology* 94:1749-1760.
129. Hagemeijer MC, Vonk AM, Monastyrska I, Rottier

- PJ, de Haan CA. 2012. Visualizing coronavirus RNA synthesis in time by using click chemistry. *Journal of virology* 86:5808-5816.
130. Neupert W, Herrmann JM. 2007. Translocation of proteins into mitochondria. *Annu Rev Biochem* 76:723-749.
131. Al-Mulla HM, Turrell L, Smith NM, Payne L, Balaji S, Zust R, Thiel V, Baker SC, Siddell SG, Neuman BW. 2014. Competitive fitness in coronaviruses is not correlated with size or number of double-membrane vesicles under reduced-temperature growth conditions. *mBio* 5:e01107-01113.
132. Diaz A, Wang X, Ahlquist P. 2010. Membrane-shaping host reticulon proteins play crucial roles in viral RNA replication compartment formation and function. *Proceedings of the National Academy of Sciences of the United States of America* 107:16291-16296.
133. Diaz A, Zhang J, Ollwerther A, Wang X, Ahlquist P. 2015. Correction: Host ESCRT Proteins Are Required for Bromovirus RNA Replication Compartment Assembly and Function. *PLoS pathogens* 11:e1004845.
134. Snijder EJ, Wassenaar AL, Spaan WJ. 1994. Proteolytic processing of the replicase ORF1a protein of equine arteritis virus. *Journal of virology* 68:5755-5764.
135. Stachowiak JC, Brodsky FM, Miller EA. 2013. A cost-benefit analysis of the physical mechanisms of membrane curvature. *Nature cell biology* 15:1019-1027.
136. de Wilde AH, Wanhee KF, Scholte FE, Goeman JJ, Ten Dijke P, Snijder EJ, Kikkert M, van Hemert MJ. 2015. A Kinome-Wide Small Interfering RNA Screen Identifies Proviral and Antiviral Host Factors in Severe Acute Respiratory Syndrome Coronavirus Replication, Including Double-Stranded RNA-Activated Protein Kinase and Early Secretory Pathway Proteins. *Journal of virology* 89:8318-8333.
137. Stanley RE, Ragusa MJ, Hurley JH. 2014. The beginning of the end: how scaffolds nucleate autophagosome biogenesis. *Trends in cell biology* 24:73-81.
138. Yang Z, Klionsky DJ. 2010. Mammalian autophagy: core molecular machinery and signaling regulation. *Current opinion in cell biology* 22:124-131.
139. Stoltz A, Ernst A, Dikic I. 2014. Cargo recognition and trafficking in selective autophagy. *Nature cell biology* 16:495-501.
140. Chiramel AI, Brady NR, Bartenschlager R. 2013. Divergent roles of autophagy in virus infection. *Cells* 2:83-104.
141. Kudchodkar SB, Levine B. 2009. Viruses and autophagy. *Reviews in medical virology* 19:359-378.
142. Kirkegaard K, Taylor MP, Jackson WT. 2004. Cellular autophagy: surrender, avoidance and subversion by microorganisms. *Nature reviews Microbiology* 2:301-314.
143. Prentice E, Jerome WG, Yoshimori T, Mizushima N, Denison MR. 2004. Coronavirus replication complex formation utilizes components of cellular autophagy. *The Journal of biological chemistry* 279:10136-10141.
144. Cottam EM, Maier HJ, Manifava M, Vaux LC, Chandra-Schoenfelder P, Gerner W, Britton P, Ktistakis NT, Wileman T. 2011. Coronavirus nsP6 proteins generate autophagosomes from the endoplasmic reticulum via an omegasome intermediate. *Autophagy* 7:1335-1347.
145. Chen Q, Fang L, Wang D, Wang S, Li P, Li M, Luo R, Chen H, Xiao S. 2012. Induction of autophagy enhances porcine reproductive and respiratory syndrome virus replication. *Virus research* 163:650-655.
146. Liu Q, Qin Y, Zhou L, Kou Q, Guo X, Ge X, Yang H, Hu H. 2012. Autophagy sustains the replication of porcine reproductive and respiratory virus in host cells. *Virology* 429:136-147.
147. Sun MX, Huang L, Wang R, Yu YL, Li C, Li PP, Hu XC, Hao HP, Ishag HA, Mao X. 2012. Porcine reproductive and respiratory syndrome virus in-

- duces autophagy to promote virus replication. *Autophagy* 8:1434-1447.
148. Monastyrská I, Ulasli M, Rottier PJ, Guan JL, Reggiori F, de Haan CA. 2013. An autophagy-independent role for LC3 in equine arteritis virus replication. *Autophagy* 9:164-174.
149. Zhao Z, Thackray LB, Miller BC, Lynn TM, Becker MM, Ward E, Mizushima NN, Denison MR, Virgin HWT. 2007. Coronavirus replication does not require the autophagy gene ATG5. *Autophagy* 3:581-585.
150. Reggiori F, Monastyrská I, Verheij MH, Cali T, Ulasli M, Bianchi S, Bernasconi R, de Haan CA, Molinari M. 2010. Coronaviruses Hijack the LC3-I-positive EDEMosomes, ER-derived vesicles exporting short-lived ERAD regulators, for replication. *Cell host & microbe* 7:500-508.
151. Jackson WT, Giddings TH, Jr., Taylor MP, Mulinyawe S, Rabinovitch M, Kopito RR, Kirkegaard K. 2005. Subversion of cellular autophagosomal machinery by RNA viruses. *PLoS biology* 3:e156.
152. Richards AL, Jackson WT. 2012. Intracellular vesicle acidification promotes maturation of infectious poliovirus particles. *PLoS pathogens* 8:e1003046.
153. Chen YH, Du W, Hagemeijer MC, Takvorian PM, Pau C, Cali A, Brantner CA, Stempinski ES, Connelly PS, Ma HC, Jiang P, Wimmer E, Altan-Bonnet G, Altan-Bonnet N. 2015. Phosphatidylserine vesicles enable efficient en bloc transmission of enteroviruses. *Cell* 160:619-630.
154. Cottam EM, Whelband MC, Wileman T. 2014. Coronavirus NSP6 restricts autophagosome expansion. *Autophagy* 10:1426-1441.
155. Zuber C, Cormier JH, Guhl B, Santimaria R, Hebert DN, Roth J. 2007. EDEM1 reveals a quality control vesicular transport pathway out of the endoplasmic reticulum not involving the COPII exit sites. *Proceedings of the National Academy of Sciences of the United States of America* 104:4407-4412.
156. Cali T, Galli C, Olivari S, Molinari M. 2008. Segregation and rapid turnover of EDEM1 by an autophagy-like mechanism modulates standard ERAD and folding activities. *Biochem Biophys Res Commun* 371:405-410.
157. Spuul P, Balistreri G, Kaariainen L, Ahola T. 2010. Phosphatidylinositol 3-kinase-, actin-, and microtubule-dependent transport of Semliki Forest Virus replication complexes from the plasma membrane to modified lysosomes. *Journal of virology* 84:7543-7557.
158. Bredenbeek PJ, Frolov I, Rice CM, Schlesinger S. 1993. Sindbis virus expression vectors: packaging of RNA replicons by using defective helper RNAs. *Journal of virology* 67:6439-6446.
159. de Boer P, Hoogenboom JP, Giepmans BN. 2015. Correlated light and electron microscopy: ultrastructure lights up! *Nature methods* 12:503-513.
160. Kukulski W, Schorb M, Welsch S, Picco A, Kaksonen M, Briggs JA. 2011. Correlated fluorescence and 3D electron microscopy with high sensitivity and spatial precision. *The Journal of cell biology* 192:111-119.
161. Pan J, Peng X, Gao Y, Li Z, Lu X, Chen Y, Ishaq M, Liu D, Dediego ML, Enjuanes L, Guo D. 2008. Genome-wide analysis of protein-protein interactions and involvement of viral proteins in SARS-CoV replication. *PloS one* 3:e3299.
162. Hagemeijer MC, Ulasli M, Vonk AM, Reggiori F, Rottier PJ, de Haan CA. 2011. Mobility and interactions of coronavirus nonstructural protein 4. *Journal of virology* 85:4572-4577.
163. Wang J, Ptacek JB, Kirkegaard K, Bullitt E. 2013. Double-membraned liposomes sculpted by poliovirus 3AB protein. *The Journal of biological chemistry* 288:27287-27298.
164. Yao F, Svensjo T, Winkler T, Lu M, Eriksson C, Eriksson E. 1998. Tetracycline repressor, tetR, rather than the tetR-mammalian cell transcription factor fusion derivatives, regulates induci-

- ble gene expression in mammalian cells. *Human gene therapy* 9:1939-1950.
165. van Kasteren PB, Bailey-Elkin BA, James TW, Ninaber DK, Beugeling C, Khajehpour M, Snijder EJ, Mark BL, Kikkert M. 2013. Deubiquitinase function of arterivirus papain-like protease 2 suppresses the innate immune response in infected host cells. *Proceedings of the National Academy of Sciences of the United States of America* 110:E838-847.
166. Schonborn J, Oberstrass J, Breyel E, Tittgen J, Schumacher J, Lukacs N. 1991. Monoclonal antibodies to double-stranded RNA as probes of RNA structure in crude nucleic acid extracts. *Nucleic acids research* 19:2993-3000.
167. Weber F, Wagner V, Rasmussen SB, Hartmann R, Paludan SR. 2006. Double-stranded RNA is produced by positive-strand RNA viruses and DNA viruses but not in detectable amounts by negative-strand RNA viruses. *Journal of virology* 80:5059-5064.
168. Kukulski W, Schorb M, Welsch S, Picco A, Kaksonen M, Briggs JA. 2012. Precise, correlated fluorescence microscopy and electron tomography of lowicryl sections using fluorescent fiducial markers. *Methods in cell biology* 111:235-257.
169. Mastronarde DN. 2005. Automated electron microscope tomography using robust prediction of specimen movements. *Journal of structural biology* 152:36-51.
170. Kremer JR, Mastronarde DN, McIntosh JR. 1996. Computer visualization of three-dimensional image data using IMOD. *Journal of structural biology* 116:71-76.
171. Drosten C, Gunther S, Preiser W, van der Werf S, Brodt HR, Becker S, Rabenau H, Panning M, Kolesnikova L, Fouchier RA, Berger A, Burguiere AM, Cinatl J, Eickmann M, Escriou N, Grywna K, Kramme S, Manuguerra JC, Muller S, Rickerts V, Sturmer M, Vieth S, Klenk HD, Osterhaus AD, Schmitz H, Doerr HW. 2003. Identification of a novel coronavirus in patients with severe acute respiratory syndrome. *The New England journal of medicine* 348:1967-1976.
172. Ksiazek TG, Erdman D, Goldsmith CS, Zaki SR, Peret T, Emery S, Tong S, Urbani C, Comer JA, Lim W, Rollin PE, Dowell SF, Ling AE, Humphrey CD, Shieh WJ, Guarner J, Paddock CD, Rota P, Fields B, DeRisi J, Yang JY, Cox N, Hughes JM, LeDuc JW, Bellini WJ, Anderson LJ, Group SW. 2003. A novel coronavirus associated with severe acute respiratory syndrome. *The New England journal of medicine* 348:1953-1966.
173. van Boheemen S, de Graaf M, Lauber C, Bestebroer TM, Raj VS, Zaki AM, Osterhaus AD, Haagmans BL, Gorbalenya AE, Snijder EJ, Fouchier RA. 2012. Genomic characterization of a newly discovered coronavirus associated with acute respiratory distress syndrome in humans. *mBio* 3.
174. Perlman S, Netland J. 2009. Coronaviruses post-SARS: update on replication and pathogenesis. *Nature reviews. Microbiology* 7:439-450.
175. Snijder EJ, Decroly E, Ziebuhr J. 2016. The Nonstructural Proteins Directing Coronavirus RNA Synthesis and Processing. *Advances in virus research* 96:59-126.
176. Baez-Santos YM, St John SE, Mesecar AD. 2015. The SARS-coronavirus papain-like protease: structure, function and inhibition by designed antiviral compounds. *Antiviral research* 115:21-38.
177. Hilgenfeld R. 2014. From SARS to MERS: crystallographic studies on coronaviral proteases enable antiviral drug design. *The FEBS journal* 281:4085-4096.
178. Mielech AM, Chen Y, Mesecar AD, Baker SC. 2014. Nidovirus papain-like proteases: multifunctional enzymes with protease, deubiquitinating and deSGylating activities. *Virus research* 194:184-190.
179. Steuber H, Hilgenfeld R. 2010. Recent advances in targeting viral proteases for the discovery

- of novel antivirals. Current topics in medicinal chemistry 10:323-345.
- 180.van der Hoeven B, Oudshoorn D, Koster AJ, Snijder EJ, Kikkert M, Barcena M. 2016. Biogenesis and architecture of arterivirus replication organelles. *Virus research*.
- 181.Avila-Perez G, Rejas MT, Rodriguez D. 2016. Ultrastructural characterization of membranous torovirus replication factories. *Cellular microbiology* 18:1691-1708.
- 182.Neuman BW. 2016. Bioinformatics and functional analyses of coronavirus nonstructural proteins involved in the formation of replicative organelles. *Antiviral research* 135:97-107.
- 183.Oostra M, Hagemeijer MC, van Gent M, Bekker CP, te Lintelo EG, Rottier PJ, de Haan CA. 2008. Topology and membrane anchoring of the coronavirus replication complex: not all hydrophobic domains of nsp3 and nsp6 are membrane spanning. *Journal of virology* 82:12392-12405.
- 184.Baliji S, Cammer SA, Sobral B, Baker SC. 2009. Detection of nonstructural protein 6 in murine coronavirus-infected cells and analysis of the transmembrane topology by using bioinformatics and molecular approaches. *Journal of virology* 83:6957-6962.
- 185.Clementz MA, Kanjanahaluethai A, O'Brien TE, Baker SC. 2008. Mutation in murine coronavirus replication protein nsp4 alters assembly of double membrane vesicles. *Virology* 375:118-129.
- 186.Beachboard DC, Anderson-Daniels JM, Denison MR. 2015. Mutations across murine hepatitis virus nsp4 alter virus fitness and membrane modifications. *Journal of virology* 89:2080-2089.
- 187.Sparks JS, Lu X, Denison MR. 2007. Genetic analysis of Murine hepatitis virus nsp4 in virus replication. *Journal of virology* 81:12554-12563.
- 188.Gadlage MJ, Sparks JS, Beachboard DC, Cox RG, Doyle JD, Stobart CC, Denison MR. 2010. Murine hepatitis virus nonstructural protein 4 regulates virus-induced membrane modifications and replication complex function. *Journal of virology* 84:280-290.
- 189.Miyazaki J, Takaki S, Araki K, Tashiro F, Tominaga A, Takatsu K, Yamamura K. 1989. Expression vector system based on the chicken beta-actin promoter directs efficient production of interleukin-5. *Gene* 79:269-277.
- 190.Bailey-Elkin BA, Knaap RC, Johnson GG, Dalebout TJ, Ninaber DK, van Kasteren PB, Bredenbeek PJ, Snijder EJ, Kikkert M, Mark BL. 2014. Crystal structure of the Middle East respiratory syndrome coronavirus (MERS-CoV) papain-like protease bound to ubiquitin facilitates targeted disruption of deubiquitinating activity to demonstrate its role in innate immune suppression. *The Journal of biological chemistry* 289:34667-34682.
- 191.Yang X, Chen X, Bian G, Tu J, Xing Y, Wang Y, Chen Z. 2014. Proteolytic processing, deubiquitinase and interferon antagonist activities of Middle East respiratory syndrome coronavirus papain-like protease. *The Journal of general virology* 95:614-626.
- 192.Norholm MH, Light S, Virkki MT, Elofsson A, von Heijne G, Daley DO. 2012. Manipulating the genetic code for membrane protein production: what have we learnt so far? *Biochim Biophys Acta* 1818:1091-1096.
- 193.Almsherqi ZA, Kohlwein SD, Deng Y. 2006. Cubic membranes: a legend beyond the Flatland* of cell membrane organization. *The Journal of cell biology* 173:839-844.
- 194.Kilianski A, Mielech AM, Deng X, Baker SC. 2013. Assessing activity and inhibition of Middle East respiratory syndrome coronavirus papain-like and 3C-like proteases using luciferase-based biosensors. *Journal of virology* 87:11955-11962.
- 195.Nagy PD, Pogany J. 2011. The dependence of viral RNA replication on co-opted host factors. *Nature reviews. Microbiology* 10:137-149.
- 196.Oudshoorn D, van der Hoeven B, Limpens RW, Beugeling C, Snijder EJ, Barcena M, Kikkert

- M. 2016. Antiviral Innate Immune Response Interferes with the Formation of Replication-Associated Membrane Structures Induced by a Positive-Strand RNA Virus. *mBio* 7.
197. Deng Y, Almsherqi ZA, Ng MM, Kohlwein SD. 2010. Do viruses subvert cholesterol homeostasis to induce host cubic membranes? *Trends in cell biology* 20:371-379.
198. Lu XT, Sims AC, Denison MR. 1998. Mouse hepatitis virus 3C-like protease cleaves a 22-kilodalton protein from the open reading frame 1a polyprotein in virus-infected cells and *in vitro*. *Journal of virology* 72:2265-2271.
199. Ziebuhr J, Siddell SG. 1999. Processing of the human coronavirus 229E replicase polyproteins by the virus-encoded 3C-like proteinase: identification of proteolytic products and cleavage sites common to pp1a and pp1ab. *Journal of virology* 73:177-185.
200. Ruggli N, Tratschin JD, Mittelholzer C, Hofmann MA. 1996. Nucleotide sequence of classical swine fever virus strain Alfort/187 and transcription of infectious RNA from stably cloned full-length cDNA. *Journal of virology* 70:3478-3487.
201. Pott J, Mahlakoiv T, Mordstein M, Duerr CU, Michiels T, Stockinger S, Staeheli P, Hornef MW. 2011. IFN-lambda determines the intestinal epithelial antiviral host defense. *Proceedings of the National Academy of Sciences of the United States of America* 108:7944-7949.
202. Muller U, Steinhoff U, Reis LF, Hemmi S, Pavlovic J, Zinkernagel RM, Aguet M. 1994. Functional role of type I and type II interferons in antiviral defense. *Science* 264:1918-1921.
203. Jiang X, Chen ZJ. 2012. The role of ubiquitylation in immune defence and pathogen evasion. *Nature reviews. Immunology* 12:35-48.
204. Maelfait J, Beyaert R. 2012. Emerging role of ubiquitination in antiviral RIG-I signaling. *Microbiol Mol Biol Rev* 76:33-45.
205. Harhaj EW, Dixit VM. 2012. Regulation of NF- κ B by deubiquitinases. *Immunol Rev* 246:107-124.
206. Malynn BA, Ma A. 2010. Ubiquitin makes its mark on immune regulation. *Immunity* 33:843-852.
207. Xu P, Duong DM, Seyfried NT, Cheng D, Xie Y, Robert J, Rush J, Hochstrasser M, Finley D, Peng J. 2009. Quantitative proteomics reveals the function of unconventional ubiquitin chains in proteasomal degradation. *Cell* 137:133-145.
208. Hershko A, Ciechanover A, Heller H, Haas AL, Rose IA. 1980. Proposed role of ATP in protein breakdown: conjugation of protein with multiple chains of the polypeptide of ATP-dependent proteolysis. *Proceedings of the National Academy of Sciences of the United States of America* 77:1783-1786.
209. Kerscher O, Felberbaum R, Hochstrasser M. 2006. Modification of proteins by ubiquitin and ubiquitin-like proteins. *Annu Rev Cell Dev Biol* 22:159-180.
210. Zhao S, Ulrich HD. 2010. Distinct consequences of posttranslational modification by linear versus K63-linked polyubiquitin chains. *Proceedings of the National Academy of Sciences of the United States of America* 107:7704-7709.
211. Dammer EB, Na CH, Xu P, Seyfried NT, Duong DM, Cheng D, Gearing M, Rees H, Lah JJ, Levey AI, Rush J, Peng J. 2011. Polyubiquitin linkage profiles in three models of proteolytic stress suggest the etiology of Alzheimer disease. *The Journal of biological chemistry* 286:10457-10465.
212. Tokunaga F, Sakata S, Saeki Y, Satomi Y, Kirisako T, Kamei K, Nakagawa T, Kato M, Murata S, Yamaoka S, Yamamoto M, Akira S, Takao T, Tanaka K, Iwai K. 2009. Involvement of linear polyubiquitylation of NEMO in NF- κ B activation. *Nature cell biology* 11:123-132.
213. Xia ZP, Sun L, Chen X, Pineda G, Jiang X, Adhikari A, Zeng W, Chen ZJ. 2009. Direct activation of protein kinases by unanchored polyubiquitin chains. *Nature* 461:114-119.
214. Hay RT. 2005. SUMO: a history of modification.

- Molecular cell 18:1-12.
- 215.Haas AL, Ahrens P, Bright PM, Ankel H. 1987. Interferon induces a 15-kilodalton protein exhibiting marked homology to ubiquitin. *The Journal of biological chemistry* 262:11315-11323.
- 216.Hochstrasser M. 2000. Evolution and function of ubiquitin-like protein-conjugation systems. *Nature cell biology* 2:E153-157.
- 217.Lenschow DJ, Lai C, Frias-Staheli N, Giannakopoulos NV, Lutz A, Wolff T, Osiak A, Levine B, Schmidt RE, Garcia-Sastre A, Leib DA, Pekosz A, Knobeloch KP, Horak I, Virgin HWt. 2007. IFN-stimulated gene 15 functions as a critical antiviral molecule against influenza, herpes, and Sindbis viruses. *Proceedings of the National Academy of Sciences of the United States of America* 104:1371-1376.
- 218.Durfee LA, Lyon N, Seo K, Huibregtse JM. 2010. The ISG15 conjugation system broadly targets newly synthesized proteins: implications for the antiviral function of ISG15. *Molecular cell* 38:722-732.
- 219.Werneke SW, Schilte C, Rohatgi A, Monte KJ, Michault A, Arenzana-Seisdedos F, Vanlandingham DL, Higgs S, Fontanet A, Albert ML, Lenschow DJ. 2011. ISG15 is critical in the control of Chikungunya virus infection independent of UbE1L mediated conjugation. *PLoS pathogens* 7:e1002322.
- 220.Tatsumi K, Yamamoto-Mukai H, Shimizu R, Waguri S, Sou YS, Sakamoto A, Taya C, Shitara H, Hara T, Chung CH, Tanaka K, Yamamoto M, Komatsu M. 2011. The Ufm1-activating enzyme Uba5 is indispensable for erythroid differentiation in mice. *Nat Commun* 2:181.
- 221.Lemaire K, Moura RF, Granvik M, Igoillo-Esteve M, Hohmeier HE, Hendrickx N, Newgard CB, Waelkens E, Cnop M, Schuit F. 2011. Ubiquitin fold modifier 1 (UFM1) and its target UFBP1 protect pancreatic beta cells from ER stress-induced apoptosis. *PloS one* 6:e18517.
- 222.Schmidtke G, Kalveram B, Groettrup M. 2009. Degradation of FAT10 by the 26S proteasome is independent of ubiquitylation but relies on NUB1L. *FEBS Lett* 583:591-594.
- 223.Watson IR, Irwin MS, Ohh M. 2011. NEDD8 pathways in cancer, Sine Quibus Non. *Cancer Cell* 19:168-176.
- 224.Raasi S, Schmidtke G, Groettrup M. 2001. The ubiquitin-like protein FAT10 forms covalent conjugates and induces apoptosis. *The Journal of biological chemistry* 276:35334-35343.
- 225.Schmid D, Munz C. 2007. Innate and adaptive immunity through autophagy. *Immunity* 27:11-21.
- 226.Pickart CM. 2001. Mechanisms underlying ubiquitination. *Annu Rev Biochem* 70:503-533.
- 227.Hershko A, Ciechanover A. 1998. The ubiquitin system. *Annu Rev Biochem* 67:425-479.
- 228.Schulman BA, Harper JW. 2009. Ubiquitin-like protein activation by E1 enzymes: the apex for downstream signalling pathways. *Nature reviews. Molecular cell biology* 10:319-331.
- 229.Love KR, Catic A, Schlieker C, Ploegh HL. 2007. Mechanisms, biology and inhibitors of deubiquitinating enzymes. *Nat Chem Biol* 3:697-705.
- 230.Nijman SM, Luna-Vargas MP, Velds A, Brummelkamp TR, Dirac AM, Sixma TK, Bernards R. 2005. A genomic and functional inventory of deubiquitinating enzymes. *Cell* 123:773-786.
- 231.Li S, Zheng H, Mao AP, Zhong B, Li Y, Liu Y, Gao Y, Ran Y, Tien P, Shu HB. 2010. Regulation of virus-triggered signaling by OTUB1- and OTUB2-mediated deubiquitination of TRAF3 and TRAF6. *The Journal of biological chemistry* 285:4291-4297.
- 232.Kayagaki N, Phung Q, Chan S, Chaudhari R, Quan C, O'Rourke KM, Eby M, Pietras E, Cheng G, Bazan JF, Zhang Z, Arnott D, Dixit VM. 2007. DUBA: a deubiquitinase that regulates type I interferon production. *Science* 318:1628-1632.
- 233.Boone DL, Turer EE, Lee EG, Ahmad RC, Wheeler MT, Tsui C, Hurley P, Chien M, Chai S, Hitotsumatsu O, McNally E, Pickart C, Ma A. 2004. The ubiquitin-modifying enzyme A20 is

- required for termination of Toll-like receptor responses. *Nature immunology* 5:1052-1060.
234. Friedman CS, O'Donnell MA, Legarda-Addison D, Ng A, Cardenas WB, Yount JS, Moran TM, Basler CF, Komuro A, Horvath CM, Xavier R, Ting AT. 2008. The tumour suppressor CYLD is a negative regulator of RIG-I-mediated antiviral response. *EMBO Rep* 9:930-936.
235. Pichlmair A, Reis e Sousa C. 2007. Innate recognition of viruses. *Immunity* 27:370-383.
236. Chen G, Shaw MH, Kim YG, Nunez G. 2009. NOD-like receptors: role in innate immunity and inflammatory disease. *Annu Rev Pathol* 4:365-398.
237. Gay NJ, Gangloff M. 2007. Structure and function of Toll receptors and their ligands. *Annu Rev Biochem* 76:141-165.
238. Ostuni R, Zanoni I, Granucci F. 2010. Deciphering the complexity of Toll-like receptor signaling. *Cell Mol Life Sci* 67:4109-4134.
239. Kawai T, Akira S. 2011. Toll-like receptors and their crosstalk with other innate receptors in infection and immunity. *Immunity* 34:637-650.
240. Yoneyama M, Kikuchi M, Natsukawa T, Shinobu N, Imaizumi T, Miyagishi M, Taira K, Akira S, Fujita T. 2004. The RNA helicase RIG-I has an essential function in double-stranded RNA-induced innate antiviral responses. *Nature immunology* 5:730-737.
241. Loo YM, Gale M, Jr. 2011. Immune signaling by RIG-I-like receptors. *Immunity* 34:680-692.
242. Hausmann S, Marq JB, Tapparel C, Kolakofsky D, Garcin D. 2008. RIG-I and dsRNA-induced IFNbeta activation. *PloS one* 3:e3965.
243. Luthra P, Sun D, Silverman RH, He B. 2011. Activation of IFN-β expression by a viral mRNA through RNase L and MDA5. *Proceedings of the National Academy of Sciences of the United States of America* 108:2118-2123.
244. Kato H, Takeuchi O, Sato S, Yoneyama M, Yamamoto M, Matsui K, Uematsu S, Jung A, Kawai T, Ishii KJ, Yamaguchi O, Otsu K, Tsujimura T, Koh CS, Reis e Sousa C, Matsuura Y, Fujita T, Akira S. 2006. Differential roles of MDA5 and RIG-I helicases in the recognition of RNA viruses. *Nature* 441:101-105.
245. Unterholzner L, Keating SE, Baran M, Horan KA, Jensen SB, Sharma S, Sirois CM, Jin T, Latz E, Xiao TS, Fitzgerald KA, Paludan SR, Bowie AG. 2010. IFI16 is an innate immune sensor for intracellular DNA. *Nature immunology* 11:997-1004.
246. Rathinam VA, Jiang Z, Waggoner SN, Sharma S, Cole LE, Waggoner L, Vanaja SK, Monks BG, Ganesan S, Latz E, Hornung V, Vogel SN, Szomolanyi-Tsuda E, Fitzgerald KA. 2010. The AIM2 inflammasome is essential for host defense against cytosolic bacteria and DNA viruses. *Nature immunology* 11:395-402.
247. Ishikawa H, Barber GN. 2008. STING is an endoplasmic reticulum adaptor that facilitates innate immune signalling. *Nature* 455:674-678.
248. Matsuzawa A, Tseng PH, Vallabhapurapu S, Luo JL, Zhang W, Wang H, Vignali DA, Gallagher E, Karin M. 2008. Essential cytoplasmic translocation of a cytokine receptor-assembled signaling complex. *Science* 321:663-668.
249. Sharma S, tenOever BR, Grandvaux N, Zhou GP, Lin R, Hiscock J. 2003. Triggering the interferon antiviral response through an IKK-related pathway. *Science* 300:1148-1151.
250. Honda K, Takaoka A, Taniguchi T. 2006. Type I interferon [corrected] gene induction by the interferon regulatory factor family of transcription factors. *Immunity* 25:349-360.
251. Schoggins JW, Wilson SJ, Panis M, Murphy MY, Jones CT, Bieniasz P, Rice CM. 2011. A diverse range of gene products are effectors of the type I interferon antiviral response. *Nature* 472:481-485.
252. Kash JC, Tumpey TM, Proll SC, Carter V, Perwitasari O, Thomas MJ, Basler CF, Palese P, Taubenberger JK, Garcia-Sastre A, Swayne DE, Katze MG. 2006. Genomic analysis of increased host immune and cell death responses induced

- by 1918 influenza virus. *Nature* 443:578-581.
253. Hugot JP, Chamaillard M, Zouali H, Lesage S, Cezard JP, Belaiche J, Almer S, Tysk C, O'Morain CA, Gassull M, Binder V, Finkel Y, Cortot A, Modigliani R, Laurent-Puig P, Gower-Rousseau C, Macry J, Colombel JF, Sahbatou M, Thomas G. 2001. Association of NOD2 leucine-rich repeat variants with susceptibility to Crohn's disease. *Nature* 411:599-603.
254. Crow MK. 2010. Type I interferon in organ-targeted autoimmune and inflammatory diseases. *Arthritis Res Ther* 12 Suppl 1:S5.
255. Miceli-Richard C, Lesage S, Rybojad M, Prieur AM, Manouvrier-Hanu S, Hafner R, Chamaillard M, Zouali H, Thomas G, Hugot JP. 2001. CARD15 mutations in Blau syndrome. *Nat Genet* 29:19-20.
256. Hooks JJ, Moutsopoulos HM, Geis SA, Stahl NI, Decker JL, Notkins AL. 1979. Immune interferon in the circulation of patients with autoimmune disease. *The New England journal of medicine* 301:5-8.
257. Li Q, Xu B, Michie SA, Rubins KH, Schreiber RD, McDevitt HO. 2008. Interferon-alpha initiates type 1 diabetes in nonobese diabetic mice. *Proceedings of the National Academy of Sciences of the United States of America* 105:12439-12444.
258. Oshiumi H, Matsumoto M, Hatakeyama S, Seya T. 2009. Riplet/RNF135, a RING finger protein, ubiquitinates RIG-I to promote interferon-beta induction during the early phase of viral infection. *The Journal of biological chemistry* 284:807-817.
259. Gack MU, Kirchhofer A, Shin YC, Inn KS, Liang C, Cui S, Myong S, Ha T, Hopfner KP, Jung JU. 2008. Roles of RIG-I N-terminal tandem CARD and splice variant in TRIM25-mediated antiviral signal transduction. *Proceedings of the National Academy of Sciences of the United States of America* 105:16743-16748.
260. Gack MU, Shin YC, Joo CH, Urano T, Liang C, Sun L, Takeuchi O, Akira S, Chen Z, Inoue S, Jung JU. 2007. TRIM25 RING-finger E3 ubiquitin ligase is essential for RIG-I-mediated antiviral activity. *Nature* 446:916-920.
261. Zeng W, Sun L, Jiang X, Chen X, Hou F, Adhikari A, Xu M, Chen ZJ. 2010. Reconstitution of the RIG-I pathway reveals a signaling role of unanchored polyubiquitin chains in innate immunity. *Cell* 141:315-330.
262. Jiang X, Kinch LN, Brautigam CA, Chen X, Du F, Grishin NV, Chen ZJ. 2012. Ubiquitin-Induced Oligomerization of the RNA Sensors RIG-I and MDA5 Activates Antiviral Innate Immune Response. *Immunity*.
263. Reyes-Turcu FE, Horton JR, Mullally JE, Heroux A, Cheng X, Wilkinson KD. 2006. The ubiquitin binding domain ZnF UBP recognizes the C-terminal diglycine motif of unanchored ubiquitin. *Cell* 124:1197-1208.
264. Gao D, Yang YK, Wang RP, Zhou X, Diao FC, Li MD, Zhai ZH, Jiang ZF, Chen DY. 2009. REUL is a novel E3 ubiquitin ligase and stimulator of retinoic-acid-inducible gene-I. *PloS one* 4:e5760.
265. Xu L, Xiao N, Liu F, Ren H, Gu J. 2009. Inhibition of RIG-I and MDA5-dependent antiviral response by gC1qR at mitochondria. *Proceedings of the National Academy of Sciences of the United States of America* 106:1530-1535.
266. Moore CB, Bergstrahl DT, Duncan JA, Lei Y, Morrison TE, Zimmermann AG, Accavitti-Loper MA, Madden VJ, Sun L, Ye Z, Lich JD, Heise MT, Chen Z, Ting JP. 2008. NLRX1 is a regulator of mitochondrial antiviral immunity. *Nature* 451:573-577.
267. Oshiumi H, Miyashita M, Inoue N, Okabe M, Matsumoto M, Seya T. 2010. The ubiquitin ligase Riplet is essential for RIG-I-dependent innate immune responses to RNA virus infection. *Cell host & microbe* 8:496-509.
268. Arimoto K, Takahashi H, Hishiki T, Konishi H, Fujita T, Shimotohno K. 2007. Negative regulation of the RIG-I signaling by the ubiquitin ligase

- RNF125. *Proceedings of the National Academy of Sciences of the United States of America* 104:7500-7505.
269. Inn KS, Gack MU, Tokunaga F, Shi M, Wong LY, Iwai K, Jung JU. 2011. Linear ubiquitin assembly complex negatively regulates RIG-I- and TRIM25-mediated type I interferon induction. *Molecular cell* 41:354-365.
270. Zou W, Wang J, Zhang DE. 2007. Negative regulation of ISG15 E3 ligase EFP through its autoISGylation. *Biochem Biophys Res Commun* 354:321-327.
271. Wang C, Deng L, Hong M, Akkaraju GR, Inoue J, Chen ZJ. 2001. TAK1 is a ubiquitin-dependent kinase of MKK and IKK. *Nature* 412:346-351.
272. Ninomiya-Tsuji J, Kishimoto K, Hiyama A, Inoue J, Cao Z, Matsumoto K. 1999. The kinase TAK1 can activate the NIK-I kappaB as well as the MAP kinase cascade in the IL-1 signalling pathway. *Nature* 398:252-256.
273. Yamamoto M, Okamoto T, Takeda K, Sato S, Sanjo H, Uematsu S, Saitoh T, Yamamoto N, Sakurai H, Ishii KJ, Yamaoka S, Kawai T, Matsuura Y, Takeuchi O, Akira S. 2006. Key function for the Ubc13 E2 ubiquitin-conjugating enzyme in immune receptor signaling. *Nature immunology* 7:962-970.
274. Deng L, Wang C, Spencer E, Yang L, Braun A, You J, Slaughter C, Pickart C, Chen ZJ. 2000. Activation of the IkappaB kinase complex by TRAF6 requires a dimeric ubiquitin-conjugating enzyme complex and a unique polyubiquitin chain. *Cell* 103:351-361.
275. Bhoj VG, Chen ZJ. 2009. Ubiquitylation in innate and adaptive immunity. *Nature* 458:430-437.
276. Xu LG, Wang YY, Han KJ, Li LY, Zhai Z, Shu HB. 2005. VISA is an adapter protein required for virus-triggered IFN-beta signaling. *Molecular cell* 19:727-740.
277. Chang M, Jin W, Sun SC. 2009. Peli1 facilitates TRIF-dependent Toll-like receptor signaling and proinflammatory cytokine production. *Nature immunology* 10:1089-1095.
278. Meylan E, Burns K, Hofmann K, Blancheteau V, Martinon F, Kelliher M, Tschopp J. 2004. RIP1 is an essential mediator of Toll-like receptor 3-induced NF-kappa B activation. *Nature immunology* 5:503-507.
279. Mikkelsen SS, Jensen SB, Chiliveru S, Melchjorsen J, Julkunen I, Gaestel M, Arthur JS, Flavell RA, Ghosh S, Paludan SR. 2009. RIG-I-mediated activation of p38 MAPK is essential for viral induction of interferon and activation of dendritic cells: dependence on TRAF2 and TAK1. *The Journal of biological chemistry* 284:10774-10782.
280. Pertel T, Hausmann S, Morger D, Zuger S, Guerra J, Lascano J, Reinhard C, Santoni FA, Uchil PD, Chatel L, Bisiaux A, Albert ML, Strambio-De-Castillia C, Mothes W, Pizzato M, Grutter MG, Luban J. 2011. TRIM5 is an innate immune sensor for the retrovirus capsid lattice. *Nature* 472:361-365.
281. Tseng PH, Matsuzawa A, Zhang W, Mino T, Vignali DA, Karin M. 2010. Different modes of ubiquitination of the adaptor TRAF3 selectively activate the expression of type I interferons and proinflammatory cytokines. *Nature immunology* 11:70-75.
282. Wertz IE, O'Rourke KM, Zhou H, Eby M, Aravind L, Seshagiri S, Wu P, Wiesmann C, Baker R, Boone DL, Ma A, Koonin EV, Dixit VM. 2004. Deubiquitination and ubiquitin ligase domains of A20 downregulate NF-kappaB signalling. *Nature* 430:694-699.
283. Shi M, Deng W, Bi E, Mao K, Ji Y, Lin G, Wu X, Tao Z, Li Z, Cai X, Sun S, Xiang C, Sun B. 2008. TRIM30 alpha negatively regulates TLR-mediated NF-kappa B activation by targeting TAB2 and TAB3 for degradation. *Nature immunology* 9:369-377.
284. Reyes-Turcu FE, Ventii KH, Wilkinson KD. 2009. Regulation and cellular roles of ubiquitin-specific deubiquitinating enzymes. *Annu Rev Biochem* 78:363-397.

- 285.Maniatis T. 1999. A ubiquitin ligase complex essential for the NF-kappaB, Wnt/Wingless, and Hedgehog signaling pathways. *Genes Dev* 13:505-510.
- 286.Rahighi S, Ikeda F, Kawasaki M, Akutsu M, Suzuki N, Kato R, Kensche T, Uejima T, Bloor S, Komander D, Randow F, Wakatsuki S, Dikic I. 2009. Specific recognition of linear ubiquitin chains by NEMO is important for NF-kappaB activation. *Cell* 136:1098-1109.
- 287.Laplantine E, Fontan E, Chiaravalli J, Lopez T, Lakisic G, Veron M, Agou F, Israel A. 2009. NEMO specifically recognizes K63-linked polyubiquitin chains through a new bipartite ubiquitin-binding domain. *The EMBO journal* 28:2885-2895.
- 288.Hacker H, Tseng PH, Karin M. 2011. Expanding TRAF function: TRAF3 as a tri-faced immune regulator. *Nature reviews. Immunology* 11:457-468.
- 289.Nakhaei P, Mesplede T, Solis M, Sun Q, Zhao T, Yang L, Chuang TH, Ware CF, Lin R, Hiscott J. 2009. The E3 ubiquitin ligase Triad3A negatively regulates the RIG-I/MAVS signaling pathway by targeting TRAF3 for degradation. *PLoS pathogens* 5:e1000650.
- 290.Tsuchida T, Zou J, Saitoh T, Kumar H, Abe T, Matsuura Y, Kawai T, Akira S. 2010. The ubiquitin ligase TRIM56 regulates innate immune responses to intracellular double-stranded DNA. *Immunity* 33:765-776.
- 291.Zhang J, Hu MM, Wang YY, Shu HB. 2012. TRIM32 modulates type I interferon induction and cellular antiviral response by targeting MITA/STING for K63-linked ubiquitination. *The Journal of biological chemistry*.
- 292.Zhong B, Zhang L, Lei C, Li Y, Mao AP, Yang Y, Wang YY, Zhang XL, Shu HB. 2009. The ubiquitin ligase RNF5 regulates antiviral responses by mediating degradation of the adaptor protein MITA. *Immunity* 30:397-407.
- 293.Zeng W, Xu M, Liu S, Sun L, Chen ZJ. 2009. Key role of Ubc5 and lysine-63 polyubiquitination in viral activation of IRF3. *Molecular cell* 36:315-325.
- 294.Tenoever BR, Ng SL, Chua MA, McWhirter SM, Garcia-Sastre A, Maniatis T. 2007. Multiple functions of the IKK-related kinase IKKepsilon in interferon-mediated antiviral immunity. *Science* 315:1274-1278.
- 295.Wild P, Farhan H, McEwan DG, Wagner S, Rogov VV, Brady NR, Richter B, Korac J, Waidmann O, Choudhary C, Dotsch V, Bumann D, Dikic I. 2011. Phosphorylation of the autophagy receptor optineurin restricts Salmonella growth. *Science* 333:228-233.
- 296.Wang C, Chen T, Zhang J, Yang M, Li N, Xu X, Cao X. 2009. The E3 ubiquitin ligase Nrdp1 'preferentially' promotes TLR-mediated production of type I interferon. *Nature immunology* 10:744-752.
- 297.Li S, Wang L, Berman M, Kong YY, Dorf ME. 2011. Mapping a dynamic innate immunity protein interaction network regulating type I interferon production. *Immunity* 35:426-440.
- 298.Chau TL, Gioia R, Gatot JS, Patrascu F, Carpenterier I, Chapelle JP, O'Neill L, Beyaert R, Piette J, Chariot A. 2008. Are the IKKs and IKK-related kinases TBK1 and IKK-epsilon similarly activated? *Trends Biochem Sci* 33:171-180.
- 299.Arimoto K, Funami K, Saeki Y, Tanaka K, Okawa K, Takeuchi O, Akira S, Murakami Y, Shimotohno K. 2010. Polyubiquitin conjugation to NEMO by tripartite motif protein 23 (TRIM23) is critical in antiviral defense. *Proceedings of the National Academy of Sciences of the United States of America* 107:15856-15861.
- 300.Zhao T, Yang L, Sun Q, Arguello M, Ballard DW, Hiscott J, Lin R. 2007. The NEMO adaptor bridges the nuclear factor-kappaB and interferon regulatory factor signaling pathways. *Nature immunology* 8:592-600.
- 301.Belghaoui S, Paz S, Goulet M, Sun Q, Iwai K, Dikic I, Hiscott J, lin R. 2012. Linear ubiquitination of

- NEMO negatively regulates the IFN antiviral response through the disruption of the MAVS signalosome. *Cell host & microbe* 12:211-222.
302. Clark K, Takeuchi O, Akira S, Cohen P. 2011. The TRAF-associated protein TANK facilitates cross-talk within the IkappaB kinase family during Toll-like receptor signaling. *Proceedings of the National Academy of Sciences of the United States of America* 108:17093-17098.
303. Schmid S, Mordstein M, Kochs G, Garcia-Sastre A, Tenover BR. 2010. Transcription factor redundancy ensures induction of the antiviral state. *The Journal of biological chemistry* 285:42013-42022.
304. Yang K, Shi HX, Liu XY, Shan YF, Wei B, Chen S, Wang C. 2009. TRIM21 is essential to sustain IFN regulatory factor 3 activation during antiviral response. *Journal of immunology* 182:3782-3792.
305. Higgs R, Lazzari E, Wynne C, Ni Gabhann J, Espinosa A, Wahren-Herlenius M, Jefferies CA. 2010. Self protection from anti-viral responses--Ro52 promotes degradation of the transcription factor IRF7 downstream of the viral Toll-Like receptors. *PloS one* 5:e11776.
306. Yoshimi R, Chang TH, Wang H, Atsumi T, Morse HC, 3rd, Ozato K. 2009. Gene disruption study reveals a nonredundant role for TRIM21/Ro52 in NF-kappaB-dependent cytokine expression in fibroblasts. *Journal of immunology* 182:7527-7538.
307. Kubota T, Matsuoka M, Chang TH, Tailor P, Sasaki T, Tashiro M, Kato A, Ozato K. 2008. Virus infection triggers SUMOylation of IRF3 and IRF7, leading to the negative regulation of type I interferon gene expression. *The Journal of biological chemistry* 283:25660-25670.
308. Liang Q, Deng H, Li X, Wu X, Tang Q, Chang TH, Peng H, Rauscher FJ, 3rd, Ozato K, Zhu F. 2011. Tripartite motif-containing protein 28 is a small ubiquitin-related modifier e3 ligase and negative regulator of IFN regulatory factor 7. *Journal of immunology* 187:4754-4763.
309. Zhang J, Xu LG, Han KJ, Wei X, Shu HB. 2004. PIASy represses TRIF-induced ISRE and NF-kappaB activation but not apoptosis. *FEBS Lett* 570:97-101.
310. Shuai K, Liu B. 2005. Regulation of gene-activation pathways by PIAS proteins in the immune system. *Nature reviews. Immunology* 5:593-605.
311. Balakirev MY, Jaquinod M, Haas AL, Chroboczek J. 2002. Deubiquitinating function of adenovirus proteinase. *Journal of virology* 76:6323-6331.
312. Gonzalez CM, Wang L, Damania B. 2009. Kaposi's sarcoma-associated herpesvirus encodes a viral deubiquitinase. *Journal of virology* 83:10224-10233.
313. Frias-Staheli N, Giannakopoulos NV, Kikkert M, Taylor SL, Bridgen A, Paragas J, Richt JA, Rowland RR, Schmaljohn CS, Lenschow DJ, Snijder EJ, Garcia-Sastre A, Virgin HW. 2007. Ovarian tumor domain-containing viral proteases evade ubiquitin- and ISG15-dependent innate immune responses. *Cell host & microbe* 2:404-416.
314. Clementz MA, Chen Z, Banach BS, Wang Y, Sun L, Ratia K, Baez-Santos YM, Wang J, Takayama J, Ghosh AK, Li K, Mesecar AD, Baker SC. 2010. Deubiquitinating and interferon antagonism activities of coronavirus papain-like proteases. *Journal of virology* 84:4619-4629.
315. Wang D, Fang L, Li P, Sun L, Fan J, Zhang Q, Luo R, Liu X, Li K, Chen H, Chen Z, Xiao S. 2011. The leader proteinase of foot-and-mouth disease virus negatively regulates the type I interferon pathway by acting as a viral deubiquitinase. *Journal of virology* 85:3758-3766.
316. van Kasteren PB, Beugeling C, Ninaber DK, Frias-Staheli N, van Boheemen S, Garcia-Sastre A, Snijder EJ, Kikkert M. 2011. Arterivirus and nairovirus ovarian tumor domain-containing deubiquitinases target activated RIG-I to control innate immune signaling. *Journal of virology*.
317. Gack MU, Albrecht RA, Urano T, Inn KS, Huang IC,

- Carnero E, Farzan M, Inoue S, Jung JU, Garcia-Sastre A. 2009. Influenza A virus NS1 targets the ubiquitin ligase TRIM25 to evade recognition by the host viral RNA sensor RIG-I. *Cell host & microbe* 5:439-449.
- 318.Lilley CE, Chaurushiya MS, Boutell C, Landry S, Suh J, Panier S, Everett RD, Stewart GS, Durocher D, Weitzman MD. 2010. A viral E3 ligase targets RNF8 and RNF168 to control histone ubiquitination and DNA damage responses. *The EMBO journal* 29:943-955.
- 319.Nerenberg BT, Taylor J, Bartee E, Gouveia K, Barry M, Fruh K. 2005. The poxviral RING protein p28 is a ubiquitin ligase that targets ubiquitin to viral replication factories. *Journal of virology* 79:597-601.
- 320.Kim W, Bennett EJ, Huttlin EL, Guo A, Li J, Possemato A, Sowa ME, Rad R, Rush J, Comb MJ, Harper JW, Gygi SP. 2011. Systematic and quantitative assessment of the ubiquitin-modified proteome. *Molecular cell* 44:325-340.
- 321.Ting L, Rad R, Gygi SP, Haas W. 2011. MS3 eliminates ratio distortion in isobaric multiplexed quantitative proteomics. *Nature methods* 8:937-940.
- 322.Dikic I, Wakatsuki S, Walters KJ. 2009. Ubiquitin-binding domains – from structures to functions. *Nature reviews. Molecular cell biology* 10:659-671.
- 323.Neuman BW, Angelini MM, Buchmeier MJ. 2014. Does form meet function in the coronavirus replicative organelle? *Trends in microbiology* 22:642-647.
- 324.Nagy PD, Barajas D, Pogany J. 2012. Host factors with regulatory roles in tombusvirus replication. *Current opinion in virology* 2:691-698.
- 325.Schneider WM, Chevillotte MD, Rice CM. 2014. Interferon-stimulated genes: a complex web of host defenses. *Annual review of immunology* 32:513-545.
- 326.Anggakusuma, Romero-Brey I, Berger C, Colpitts CC, Boldanova T, Engelmann M, Todd D, Perin PM, Behrendt P, Vondran FW, Xu S, Goffinet C, Schang LM, Heim MH, Bartenschlager R, Pietschmann T, Steinmann E. 2015. Interferon-inducible cholesterol-25-hydroxylase restricts hepatitis C virus replication through blockage of membranous web formation. *Hepatology* 62:702-714.
- 327.Helbig KJ, Eyre NS, Yip E, Narayana S, Li K, Fiches G, McCartney EM, Jangra RK, Lemon SM, Beard MR. 2011. The antiviral protein viperin inhibits hepatitis C virus replication via interaction with nonstructural protein 5A. *Hepatology* 54:1506-1517.
- 328.Melen K, Keskinen P, Lehtonen A, Julkunen I. 2000. Interferon-induced gene expression and signaling in human hepatoma cell lines. *Journal of hepatology* 33:764-772.
- 329.van den Born E, Posthuma CC, Knoops K, Snijder EJ. 2007. An infectious recombinant equine arteritis virus expressing green fluorescent protein from its replicase gene. *The Journal of general virology* 88:1196-1205.
- 330.Faas FG, Avramut MC, van den Berg BM, Mommaas AM, Koster AJ, Ravelli RB. 2012. Virtual nanoscopy: generation of ultra-large high resolution electron microscopy maps. *The Journal of cell biology* 198:457-469.
- 331.Helbig KJ, Beard MR. 2014. The role of viperin in the innate antiviral response. *Journal of molecular biology* 426:1210-1219.
- 332.Sahu SK, Gummadi SN, Manoj N, Aradhya GK. 2007. Phospholipid scramblases: an overview. *Archives of biochemistry and biophysics* 462:103-114.
- 333.Cyster JG, Dang EV, Reboldi A, Yi T. 2014. 25-Hydroxycholesterols in innate and adaptive immunity. *Nature reviews. Immunology* 14:731-743.
- 334.Schoggins JW, MacDuff DA, Imanaka N, Gainey MD, Shrestha B, Eitson JL, Mar KB, Richardson RB, Ratushny AV, Litvak V, Dabelic R, Manicassamy B, Aitchison JD, Aderem A, Elliott

- RM, Garcia-Sastre A, Racaniello V, Snijder EJ, Yokoyama WM, Diamond MS, Virgin HW, Rice CM. 2014. Pan-viral specificity of IFN-induced genes reveals new roles for cGAS in innate immunity. *Nature* 505:691-695.
335. Diamond MS, Farzan M. 2013. The broad-spectrum antiviral functions of IFIT and IFITM proteins. *Nature reviews. Immunology* 13:46-57.
336. Choi J, Park S, Biering SB, Selleck E, Liu CY, Zhang X, Fujita N, Saitoh T, Akira S, Yoshimori T, Sibley LD, Hwang S, Virgin HW. 2014. The parasitophorous vacuole membrane of Toxoplasma gondii is targeted for disruption by ubiquitin-like conjugation systems of autophagy. *Immunity* 40:924-935.
337. Park S, Choi J, Biering SB, Dominici E, Williams LE, Hwang S. 2016. Targeting by Autophagy proteins (TAG): Targeting of IFNG-inducible GTPases to membranes by the LC3 conjugation system of autophagy. *Autophagy*:0.
338. van Kasteren PB, Beugeling C, Ninaber DK, Frias-Staheli N, van Boheemen S, Garcia-Sastre A, Snijder EJ, Kikkert M. 2012. Arterivirus and nairovirus ovarian tumor domain-containing Deubiquitinases target activated RIG-I to control innate immune signaling. *Journal of virology* 86:773-785.
339. MacLachlan NJ, Balasuriya UB, Hedges JF, Schweidler TM, McCollum WH, Timoney PJ, Hullinger PJ, Patton JF. 1998. Serologic response of horses to the structural proteins of equine arteritis virus. *Journal of veterinary diagnostic investigation : official publication of the American Association of Veterinary Laboratory Diagnosticicians, Inc* 10:229-236.
340. Carlotti F, Bazuine M, Kekarainen T, Seppen J, Pognonec P, Maassen JA, Hoeben RC. 2004. Lentiviral vectors efficiently transduce quiescent mature 3T3-L1 adipocytes. *Molecular therapy : the journal of the American Society of Gene Therapy* 9:209-217.
341. Shalem O, Sanjana NE, Hartenian E, Shi X, Scott DA, Mikkelsen TS, Heckl D, Ebert BL, Root DE, Doench JG, Zhang F. 2014. Genome-scale CRISPR-Cas9 knockout screening in human cells. *Science* 343:84-87.
342. Sanjana NE, Shalem O, Zhang F. 2014. Improved vectors and genome-wide libraries for CRISPR screening. *Nature methods* 11:783-784.
343. Kusano S, Eizuru Y. 2012. Human phospholipid scramblase 1 interacts with and regulates transactivation of HTLV-1 Tax. *Virology* 432:343-352.
344. V'Kovski P, Al-Mulla H, Thiel V, Neuman BW. 2015. New insights on the role of paired membrane structures in coronavirus replication. *Virus research* 202:33-40.
345. Ratia K, Saikatendu KS, Santarsiero BD, Barretto N, Baker SC, Stevens RC, Mesecar AD. 2006. Severe acute respiratory syndrome coronavirus papain-like protease: structure of a viral deubiquitinating enzyme. *Proceedings of the National Academy of Sciences of the United States of America* 103:5717-5722.
346. Sulea T, Lindner HA, Purisima EO, Menard R. 2005. Deubiquitination, a new function of the severe acute respiratory syndrome coronavirus papain-like protease? *Journal of virology* 79:4550-4551.
347. Barretto N, Jukneliene D, Ratia K, Chen Z, Mesecar AD, Baker SC. 2005. The papain-like protease of severe acute respiratory syndrome coronavirus has deubiquitinating activity. *Journal of virology* 79:15189-15198.
348. Lindner HA, Fotouhi-Ardakani N, Lytvyn V, Lachance P, Sulea T, Menard R. 2005. The papain-like protease from the severe acute respiratory syndrome coronavirus is a deubiquitinating enzyme. *Journal of virology* 79:15199-15208.
349. Devaraj SG, Wang N, Chen Z, Chen Z, Tseng M, Barretto N, Lin R, Peters CJ, Tseng CT, Baker SC, Li K. 2007. Regulation of IRF-3-dependent innate immunity by the papain-like protease domain of the severe acute respiratory syndrome coronavirus. *The Journal of biological chemistry*

- try 282:32208-32221.
- 350.Sun Z, Chen Z, Lawson SR, Fang Y. 2010. The cysteine protease domain of porcine reproductive and respiratory syndrome virus nonstructural protein 2 possesses deubiquitinating and interferon antagonism functions. *Journal of virology* 84:7832-7846.
- 351.Lemm JA, Rumenapf T, Strauss EG, Strauss JH, Rice CM. 1994. Polypeptide requirements for assembly of functional Sindbis virus replication complexes: a model for the temporal regulation of minus- and plus-strand RNA synthesis. *The EMBO journal* 13:2925-2934.
- 352.Shirako Y, Strauss JH. 1994. Regulation of Sindbis virus RNA replication: uncleaved P123 and nsP4 function in minus-strand RNA synthesis, whereas cleaved products from P123 are required for efficient plus-strand RNA synthesis. *Journal of virology* 68:1874-1885.
- 353.Kaur J, Debnath J. 2015. Autophagy at the crossroads of catabolism and anabolism. *Nature reviews. Molecular cell biology* 16:461-472.
- 354.D'Angelo G, Vicinanza M, Di Campi A, De Matteis MA. 2008. The multiple roles of PtdIns(4)P -- not just the precursor of PtdIns(4,5)P₂. *Journal of cell science* 121:1955-1963.
- 355.Fritsch SD, Weichhart T. 2016. Effects of Interferons and Viruses on Metabolism. *Frontiers in immunology* 7:630.
- 356.Xue B, Yang D, Wang J, Xu Y, Wang X, Qin Y, Tian R, Chen S, Xie Q, Liu N, Zhu H. 2016. ISG12a restricts hepatitis C virus infection through ubiquitination-dependent degradation pathway. *Journal of virology*.
- 357.Maier HJ, Neuman BW, Bickerton E, Keep SM, Alrashedi H, Hall R, Britton P. 2016. Extensive coronavirus-induced membrane rearrangements are not a determinant of pathogenicity. *Sci Rep* 6:27126.

Samenvatting

Curriculum vitae

List of publications

SAMENVATTING

Virussen kunnen levensbedreigende infecties veroorzaken, ondanks het feit dat ze relatief simpel in elkaar zitten. Het virusdeeltje bestaat uit het genoom dat is ingepakt in een of meerdere eiwitten. Bij veel virussen bevat het deeltje ook een membraan, met daarin verankerd de eiwitten waarmee het virusdeeltje aan een receptor kan binden om een gastheercel te infecteren. Na dit moment van infectie, interacteren virussen op allerlei vlakken met de gastheercel om uiteindelijk nieuwe virusdeeltjes te kunnen maken. Dit stapsgewijze proces wordt de replicatiecyclus genoemd, en tussen virusfamilies en -groepen bestaat aanzienlijke variatie waar het de details van deze cyclus betreft. In het geval van de positiefstrengige (+) RNA-virussen bestaat het virale genoom uit een enkelstrengs mRNA molecuul dat direct na het binnendringen in de cel wordt afgelezen door ribosomen om zo de eerste virale eiwitten te produceren. Deze eiwitten zorgen ervoor dat het genoom wordt vermenigvuldigd en kan worden verpakt in nieuwe virusdeeltjes.

Dit proefschrift richt zich op de een specifiek aspect van de replicatie van de arterivirussen en de coronavirussen (twee +RNA-virusfamilies), namelijk de vorming van zogenaamde replicatieorganellen in het cytoplasma van de geïnfecteerde cel. Arterivirussen en coronavirussen zijn evolutionair verwant en behoren tot de orde van de nidovirussen. Arterivirussen veroorzaken vooral problemen in de veterinaire wereld, terwijl coronavirussen ook humane infecties kunnen veroorzaken. De bekendste coronavirussen zijn het severe acute respiratory syndrome coronavirus (SARS-CoV) en Middle East respiratory syndrome coronavirus (MERS-CoV), die beiden dodelijke respiratoire infecties in de mens kunnen veroorzaken. Hoewel het genoom van coronavirussen twee tot drie keer groter is dan dat van arterivirussen, is de genoomorganisatie en replicatiecyclus van deze twee virusfamilies in grote lijnen vergelijkbaar. De voor mensen ongevaarlijke arterivirussen worden daarom mede gebruikt als model om de replicatiecyclus van levensbedreigende coronavirussen in kaart te brengen.

+RNA virale replicatieorganellen zijn structuren die aangemaakt worden in de geïnfecteerde gastheercel op basis van een samenspel tussen virale en cellulaire spelers. De basis van de replicatieorganellen wordt gevormd door membranen die 'gestolen' worden van de gastheercel. De transformatie van deze membranen tot unieke membraanstructuren wordt aangestuurd door specifieke +RNA virale transmembraaneiwitten, vermoedelijk in nauwe samenwerking met gastheercel eiwitten. De omvangrijke wetenschappelijke literatuur over +RNA-virale replicatieorganellen wordt samengevat in **hoofdstuk 2**. In de wetenschappelijke literatuur is veel gespeculeerd over de functie van deze replicatieorganellen tijdens virusinfectie. Ten eerste wordt gedacht dat deze structuren een "micromilieu" vormen ter bevordering van de enzymatische reacties die nodig zijn tijdens de reproductie van het virale genoom. Ook zouden deze structuren kunnen bijdragen aan de regulatie van de replicatiecyclus, door verschillende processen te compartmentaliseren. Tot slot wordt aangenomen dat de replicatieorganellen ervoor zorgen dat de gastheercel het binnengedrongen virus niet (of pas later) kan herkennen. De structuren zouden een soort schild vormen om het virale genoom af te schermen van het zogenaamde aangeboren

immuunsysteem. Replicatieorganellen worden niet alleen in nidovirus- geïnfecteerde cellen gevormd, maar in alle eukaryote cellen die geïnfecteerd zijn met een +RNA-virus, hoewel de soort membraanstructuren en het als membraandonor gebruikte cellulaire compartiment sterk kunnen verschillen. De vorming van deze membraanstructuren is daarmee één van de meest karakteristieke elementen van de replicatiecyclus van deze grote virusgroep. Voorbeelden van de replicatieorganellen van arterivirussen en coronavirussen zijn afgebeeld in **hoofdstuk 2** figuur 2 en 4. Arterivirus-geïnduceerde replicatieorganellen bestaan uit "double-membrane vesicles" (DMVs), gesloten blaasjes die zijn omgeven met twee gepaarde membranen (**hoofdstuk 2**, figuur 2B). Deze DMVs zijn via hun buitenmembraan met elkaar verbonden en vormen zo een netwerk. Coronavirussen vormen naast DMVs ook nog enkele andere membraanstructuren zoals "convoluted membranes", "spherules" en "vesicle packets".

Bij nidovirussen wordt de vorming van de replicatieorganellen geïnduceerd door de virale eiwitten die gemaakt worden door translatie van het virale genoom. Deze eiwitten worden gemaakt als onderdeel van twee lange poly-eiwitten, die intern gekliefd worden tot losse eenheden (nonstructural proteins, nsps) door proteases die in deze poly-eiwitten aanwezig zijn. Doordat de replicatiecyclus van een virus een complex proces is, is het lastig om specifieke onderdelen van de virale replicatiemachine te modificeren zonder dat de replicatie zelf verstoord wordt of verloren gaat. Door bijvoorbeeld mutaties aan te brengen in de nsps die de vorming van replicatieorganellen induceren zou men kunnen bestuderen hoe de organellen gevormd worden en welke gastheercelfactoren daarvoor nodig zijn. Maar als door zulke mutaties essentiële aspecten van de replicatiecyclus niet meer functioneren worden de virale eiwitten die de vorming van membraanstructuren aansturen überhaupt niet meer gemaakt, waardoor er geen replicatieorganellen worden gevormd. **hoofdstuk 3** beschrijft onderzoek naar de vorming van arterivirus replicatieorganellen. Sinds 2001 was bekend dat de expressie van twee arterivirale nsps (nsp2 en nsp3) voldoet om - buiten de context van virusreplicatie - de vorming van membraanstructuren te induceren. Deze 'surrogaat'structuren lijken sterk op de replicatieorganellen die gevormd worden tijdens arterivirusinfectie. Omdat nsp5, naast nsp2 en nsp3, het enige andere arterivirale transmembraan nsp is, was het aannemelijk dat nsp5 ook een rol speelt in de vorming van DMVs, en ook dit is in dit hoofdstuk bestudeerd. De DMVs die werden gevormd na gezamenlijke expressie van nsp2, nsp3 en nsp5 waren kleiner en uniformer, maar de globale structuur van de DMVs veranderde niet. Dit wijst erop dat nsp5 waarschijnlijk een regulerende rol heeft. Ook is in dit onderzoek een driedimensionale reconstructie gemaakt van de 'surrogaat' DMVs, om meer inzicht te krijgen in hun biogenese. Daaruit bleek dat waarschijnlijk eerst twee membranen worden gepaard, die daarna tot DMVs worden getransformeerd.

Het was lang onbekend welke coronavirale eiwitten nodig zijn om DMVs (en de andere structuren van het coronavirus replicatieorganel) te vormen. In 2013 beschreven Angelini *et al.* dat voor SARS-CoV nsp3, nsp4 en nsp6 nodig zijn voor de vorming van DMVs. Coronavirus nsp3, nsp4 en nsp6 zijn te vergelijken met arterivirus nsp2, nsp3 en nsp5 (zie ook **hoofdstuk 2**, figuur 1). In **hoofdstuk 4** laten we echter zien dat voor zowel SARS-CoV

als MERS-CoV expressie van nsp3 en nsp4 voldoende is om DMV-vorming te induceren, en dat MERS-CoV nsp6 geen rol lijkt te spelen bij DMV-vorming. Ook dit onderzoek werd uitgevoerd met behulp van gedetailleerde driedimensionale reconstructies op basis van elektronentomografie, en laat zien dat arterivirussen en coronavirussen vergelijkbare mechanismen gebruiken om DMVs te vormen. Zoals eerder vermeld maken de coronavirus nsps deel uit van een groot poly-eiwit, dat door interne proteases tot individuele nsps wordt gekliefd. Deze proteases bevinden zich in nsp3 en nsp5. In dit onderzoek is ook gekeken naar de rol in DMV-vorming van de klieving tussen nsp3 en nsp4 door het protease in nsp3. De klieving tussen nsp3 en nsp4 bleek essentieel om DMV-vorming op gang te brengen. Als deze klieving werd geblokkeerd, werden de membranen nog wel gepaard, maar in plaats van DMVs accumuleerden dubbele membranen, vaak in de vorm van concentrische structuren. Deze resultaten geven aan dat de klieving van het virale poly-eiwit een integrale rol kan spelen in de regulatie van de vorming van +RNA virale replicatieorganellen.

Cellen hebben een breed scala aan sensoren om de aanwezigheid van een virus te detecteren. Deze sensoren herkennen bijvoorbeeld het virale genoom, virale eiwitten of virusdeeltjes. Herkenning is gebaseerd op bepaalde ongebruikelijke moleculaire patronen, en is niet gericht op een specifiek pathogenen, zoals dat wel het geval is bij de adaptieve immuunrespons. Als een cel een binnengedrongen virus heeft gesigneerd, wordt het aangeboren immuunsysteem geactiveerd. Dit zorgt ervoor dat zowel in de geïnfecteerde cel als in nabijgelegen cellen een antivirale modus aangeschakeld wordt, die deze cellen moeilijker te infecteren maakt en virusreplicatie remt. Dit signaal wordt onder meer overgedragen door interferon eiwitten. Ook zorgt de antivirale modus ervoor dat witte bloedcellen worden gerekruteerd om het adaptieve immuunsysteem te activeren. Deze mechanismen, en hoe ze gereguleerd worden, worden toegelicht in **hoofdstuk 5**. Om toch te kunnen repliceren hebben virussen verschillende methoden ontwikkeld om te voorkomen dat ze herkend worden of om de cellulaire respons te blokkeren of te vertragen. Dit resulteert in een constante wapenwedloop tussen het aangeboren immuunsysteem en virussen.

De resultaten beschreven in **hoofdstuk 6** toonden aan dat het aangeboren immuunsysteem arterivirus DMV-vorming, door expressie van virale eiwitten geïnduceerd, kan remmen. In dit onderzoek zijn cellen die arterivirus nsp2 en nsp3 tot expressie brengen behandeld met interferon om de antivirale modus te activeren. Aangezien interferonbehandeling virusreplicatie op meerdere fronten remt, is dit onderzoek uitgevoerd buiten de context van virusinfectie om specifiek een effect van het aangeboren immuunsysteem op DMV-vorming te kunnen onderzoeken. In deze proefopzet zorgde interferonbehandeling ervoor dat minder cellen DMVs vormden. Daarnaast werden er na interferon behandeling meer gepaarde membranen gevonden (die geen DMVs vormden) in vergelijking met onbehandelde cellen. Deze bevindingen wijzen op remming van DMV-biogenese door het aangeboren immuunsysteem. De antivirale respons die door interferon wordt geactiveerd leidt tot verhoogde expressie van minstens driehonderd verschillende genen. Van drie van deze genen (CH25H, viperin en PLSCR1) is getest of ze invloed hebben op DMV-vorming door arterivirus nsp2 en nsp3, omdat elders vergelijkbare effecten van hun

genproducten waren beschreven. CH25H, viperin en PLSCR1 bleken echter niet betrokken bij de remming van arterivirale DMV-vorming door het aangeboren immuunsysteem, wat suggereert dat dit gebeurt via een tot op heden onbekend mechanisme.

Het werk in dit proefschrift verschaft nieuwe inzichten in de biogenese van replicatieorganellen van arterivirussen en coronavirussen. Ook is nu duidelijk dat cellen actief proberen de vorming van deze replicatieorganellen tegen te gaan. Als in toekomstig onderzoek het mechanisme van de remming van DMV-vorming door het aangeboren immuunsysteem opgehelderd wordt, kan deze kennis bijvoorbeeld gebruikt worden om selectieve antivirale strategieën te ontwikkelen.

CURRICULUM VITAE

Diede Oudshoorn werd 5 februari 1989 geboren te Warmond. In 2005 behaalde hij zijn gymnasiumdiploma aan het Stedelijk Gymnasium in Leiden. Vervolgens begon hij aan de opleiding Biomedische Wetenschappen aan de Universiteit Leiden. Tijdens de bachelor fase nam hij deel aan een uitwisselingsprogramma met het Karolinska Instituut in Stockholm, Zweden. Tijdens de onderzoeksspecialisatie in de master fase heeft Diede zijn eerste stageproject uitgevoerd bij de afdeling Medische Microbiologie (LUMC) onder begeleiding van Kazimier Wannee, M.Sc. en dr. ir. Marjolein Kikkert. De afstudeerstage werd uitgevoerd in New York bij het Department of Microbiology van de Icahn School of Medicine at Mount Sinai, onder supervisie van dr. Gijs Versteeg en prof. dr. Adolfo García-Sastre. Daar werkte hij aan ISG15 (interferon-stimulated gene 15) van de muis. Direct na het behalen van zijn masterdiploma (*cum laude*) begon hij aan een promotietraject bij de LUMC-afdeling Medische Microbiologie, onder begeleiding van promotor prof. dr. Eric Snijder en zijn voormalige stagebegeleider dr. ir. Marjolein Kikkert. Zijn onderzoek richtte zich in eerste instantie op de interactie tussen het aangeboren immuunsysteem en virale replicatie en breidde zich gedurende het project uit naar de vorming van virusgeïnduceerde membraanstructuren onder begeleiding van dr. Montserrat Bárcena.

LIST OF PUBLICATIONS

1. **Oudshoorn D**, Rijs K, Limpens RWAL, Groen K, Koster AJ, Snijder EJ, Kikkert M, Bárcena M. 2017. Expression and Cleavage of Middle East Respiratory Syndrome Coronavirus nsp3-4 Polyprotein Induce the Formation of Double-Membrane Vesicles That Mimic Those Associated with Coronaviral RNA Replication. *mBio* 8(6) e01658-17.
2. **Oudshoorn D**, van der Hoeven B, Limpens RW, Beugeling C, Snijder EJ, Bárcena M, Kikkert M. 2016. Antiviral Innate Immune Response Interferes with the Formation of Replication-Associated Membrane Structures Induced by a Positive-Strand RNA Virus. *mBio* 7(6) e01991-16.
3. van der Hoeven B*, **Oudshoorn D***, Koster AJ, Snijder EJ, Kikkert M, Bárcena M. 2016. Biogenesis and architecture of arterivirus replication organelles. *Virus Res* 220:70-90.
4. de Wilde AH, Raj VS, **Oudshoorn D**, Bestebroer TM, van Nieuwkoop S, Limpens RW, Posthuma CC, van der Meer Y, Bárcena M, Haagmans BL, Snijder EJ, van den Hoogen BG. 2013. MERS-coronavirus replication induces severe *in vitro* cytopathology and is strongly inhibited by cyclosporin A or interferon-alpha treatment. *J Gen Virol* 94:1749-1760.
5. **Oudshoorn D**, Versteeg GA, Kikkert M. 2012. Regulation of the innate immune system by ubiquitin and ubiquitin-like modifiers. *Cytokine Growth Factor Rev* 23:273-282.
6. **Oudshoorn D**, van Boheemen S, Sanchez-Aparicio MT, Rajsbaum R, Garcia-Sastre A, Versteeg GA. 2012. HERC6 is the main E3 ligase for global ISG15 conjugation in mouse cells. *PLoS One* 7:e29870.

*Authors contributed equally to this study

