

Vulvar squamous cell carcinoma : genetics, morphology and clinical behaviour

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Citation

Trietsch, M. D. (2017, November 9). *Vulvar squamous cell carcinoma : genetics, morphology and clinical behaviour*. Retrieved from https://hdl.handle.net/1887/54945

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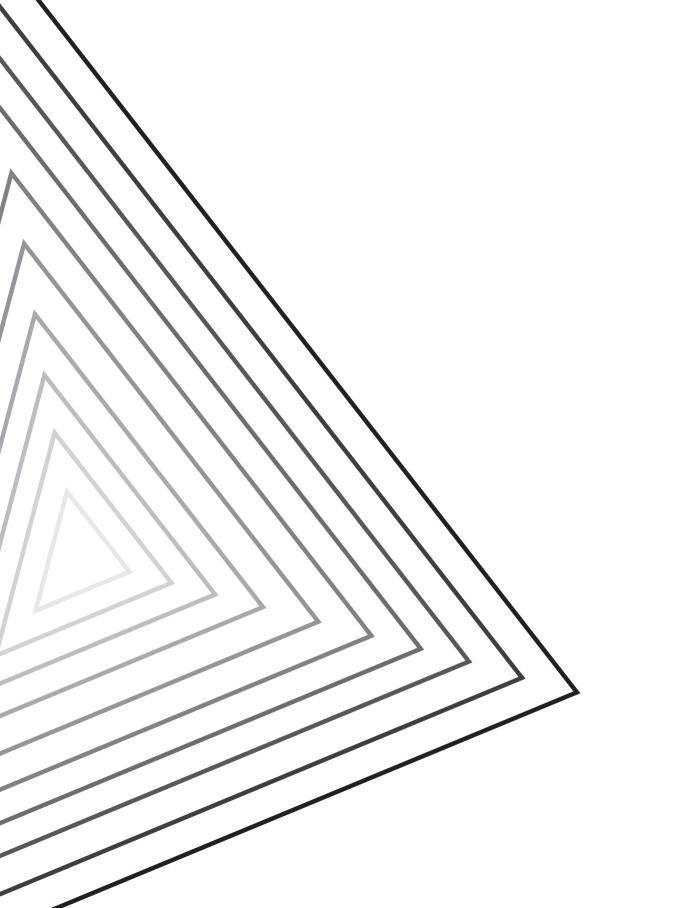


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Title: Vulvar squamous cell carcinoma: genetics, morphology and clinical behaviour

Issue Date: 2017-11-09



Chapter 1

General introduction



General introduction

Although vulvar cancer is a rare gynaecologic malignancy with 2 new cases per 100,000 women each year, its impact on the often older aged patients can be devastating (1-7). When detected at an early stage, vulvar cancer can be treated curatively by surgical excision (8). Some patients however, suffer from rapid recurrences and progression (9). Vulvar cancer that has spread to the urethra, anus or lymph nodes requires larger or wider surgery, or a combination of surgery and radio- and chemotherapy, which in turn results in higher morbidity rates. This sometimes mutilating treatment has a major impact on the quality of life, because patients suffer from wound healing problems, lymph oedema and nerve damage that can result in sexual dysfunction and incontinence (10-12).

Currently, the aetiology of vulvar cancer has been far from unravelled. The majority of vulvar cancers are squamous cell carcinomas. It is well accepted that approximately 40% of all new cases of vulvar squamous cell carcinomas (VSCCs) are associated with a persistent infection by a high risk variant of the human papilloma virus (HPV) and that the other 60% are HPV negative and associated with mutations in the *TP53* gene (3;13;14). The HPV positive vulvar cancers are preceded by usual vulvar intraepithelial neoplasia (uVIN), a condition that has a malignant potential of 9-16% in untreated women (1;15) (figure met HE coupe). This type of vulvar cancer affects younger women and is associated with smoking and a higher number of sexual partners (1:3:16).

Differentiated vulvar intraepithelial neoplasia (dVIN) is the precursor lesion of the non HPV related vulvar squamous cell carcinoma (figure 1). This type of VIN is very hard to recognise and is mainly seen adjacent to vulvar cancer. Whether this means that it is a precursor lesion that rapidly progresses to cancer before it can be detected as a solitary lesion is still being debated (17;18).

Markers that can predict patient outcome other than lymph node metastases have not been established yet (19;20). This thesis contains work to further understand and predict why vulvar cancer can behave as aggressively as it sometimes does.

Treatment of vulvar cancer

The International Federation of Gynaecology and Obstetrics (FIGO) staging system uses clinical and histological parameters to subdivide patients into risk categories (table 1 FIGO staging) (21). Early stage vulvar cancer has a fairly good prognosis, but the prognosis rapidly declines with increasing stage (table 2 FIGO stage and survival) (22).

Surgical excision is the primary treatment for low stage vulvar cancer (12). Depending on the size and spread of the tumour, other treatment modalities come to play. In the Netherlands, patients with unifocal VSCC with a diameter <4 cm

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Table 1 FIGO staging in vulvar cancer:

Stage	TNM Classification	Description
0	Tis NO MO	Carcinoma in situ, intraepithelial carcinoma
1	T1 N0 M0	Confined to the vulva or perineum; no nodal metastasis
Α	T1a N0 M0	Lesions =< 2 cm with stromal invasion, =< 1 mm
В	T1b N0 M0	Lesions > 2 cm in size or stromal invasion, > 1 mm
II	T2 N0 M0	Adjacent spread to the lower urethra, the vagina, or the anus, no nodal metastasis
III	T1,2 N1a,b N2a,b,c M0	Tumour confined to vulva or adjacent spread to the lower urethra, the vagina, or the anus and positive inguinofemoral lymph nodes
Α	T1,2 N1a,b M0	One lymph node metastasis >= 5mm or 1-2 lymph node metastases < 5 mm
В	T1,2 N2a,b M0	Three or more lymph nodes < 5mm or 2 or more lymph nodes >= 5mm
С	T1,2 N2c M0	Lymph nodes with extracapsular spread
	T1,2 N3 M0	Tumour with fixed or ulcerated lymph nodes
IV A	T3 anyN M0	Tumour with spread into upper urethra/vagina, bladder, rectal mucosa, bone or fixed to pelvic bone
IV B	Any T Any N M1	Any distant metastasis, including pelvic lymph nodes

(FIGO 2009 and UICC 7th ed.)

Table 2 Five year survival according to FIGO Annual Report 2006, using the 1995 FIGO staging system:

Stage I: 79%	
Stage II: 59%	
Stage III: 43%	
Stage IV: 13%	

without suspicious groin nodes are generally treated with radical local excision and a sentinel node procedure (23). Tumours with a diameter exceeding 4 cm or multifocal tumours are generally treated with radical local excision and unilateral or bilateral inguinofemoral lymphadenectomy. In patients with tumours in FIGO stage III or higher but with contraindications for extensive surgery, such as high age and comorbidity, radical local excision is performed without inguinofemoral lymphadenectomy, followed by (chemo)radiation.

All of these treatments can be mutilating and have high morbidity rates (10;11). Wound healing problems, lymph oedema, and sexual dysfunction regularly occur. These negative effects of treatment can have a major impact on the patient's quality of life (10).

Aetiology of vulvar squamous cell carcinoma: two different pathways

Cancer is the uncontrolled growth of abnormal cells, which can occur when the DNA in our cells is damaged and fails to be repaired. It can arise in virtually any cell type within the body. Skin cancer is one of the most prevalent types of cancer worldwide and is most often caused by DNA damage through ultraviolet irradiation

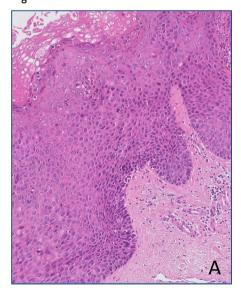


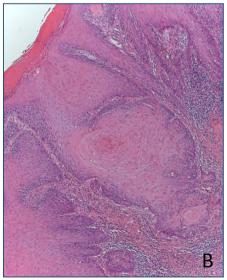
from sunlight. Well known skin cancer types are, amongst others, melanomas, basal cell carcinomas and squamous cell carcinomas.

The skin of the outer parts of the female genital tract, the vulva, can also be affected by cancer, but usually through a completely different etiologic pathway than in skin cancer on other locations of the body. As said before, most vulvar cancers are squamous cell carcinomas, a type of cancer arising from the squamous epithelium of the vulvar skin.

The pathogenesis of vulvar cancer can be divided into two different pathways: an HPV dependent, and an HPV independent route. Approximately 40% of all vulvar cancers are caused by a persistent infection with high risk HPV (3;13;14). HPV's are common and contagious viruses that can cause abnormal cell growth which can result in warts, and in some cases, cancer. More than 100 types of HPV have been described, of which 15 are 'high risk' types that can cause malignancies. These high risk HPV's are the main cause of cervical cancer and its precursor lesion cervical intra-epithelial neoplasia (CIN) (24). They can also cause the much less prevalent vulvar cancer and usual vulvar intraepithelial neoplasia (uVIN) (1;2;15;25). CIN usually goes unnoticed by its host, but VIN itself can be an unpleasant condition, resulting in itching and pain. Of VIN patients not receiving treatment, 9-16% will progress towards vulvar cancer (1;15). uVIN patients that do receive treatment have a risk of developing cancer from their VIN lesion of approximately 3% (figure 1a).

Figure 1 Usual VIN and differentiated VIN





HE slides of a usual VIN (A) and differentiated VIN (B). Note the uniform cells with hyperchromatic nuclei and little maturation in uVIN and the mature cells with eosinophilic cytoplasm in dVIN.

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It is estimated that over 80% of women will have an HPV infection at least once in her life, whereas only a very small number of these women will suffer from cancer caused by HPV infection. In most cases, the HPV infection is transient, and the body is able to clear the virus by itself within 1 to 2 years (24). Some women, however, are unable to rid the virus, which finds a way to integrate itself in the human DNA and causes cell growth and, in some cases, eventually cancer. Smoking and a compromised immune system (for example by HIV/AIDS) are factors that have been proven to result in a higher risk of persistent HPV infection and therefore a higher risk of HPV associated cancer (24). Other than that, little is known about why some women are unable to clear the HPV infection.

The other 60% of vulvar cancers are not associated with HPV infections, but with lichen sclerosis, a chronic, autoimmune inflammatory disease of the skin. Lichen sclerosis progresses towards cancer in 3-5% of patients (1;15). This pathway has a worse prognosis than the HPV positive pathway, and affects older patients than the HPV positive pathway, with an average age at diagnosis of 70 versus 40 years (1-3;15;25). Approximately 3-5% of women with lichen sclerosis will develop vulvar squamous cell carcinoma. Differentiated VIN (dVIN) is thought to be the precursor lesion of HPV negative vulvar cancers, but it is hardly ever diagnosed before invasive cancer has developed. This is probably due to its subtle clinical and histological appearance, which makes it hard to recognise by clinicians and pathologists, but it could also mean that dVINs progress very rapidly into vulvar cancer (3;15) (figure 1b).

Somatic mutations in the HPV negative pathway

The HPV negative pathway is associated with lichen sclerosis, but also with somatic mutations. The most frequently mutated and well-studied gene in vulvar cancer, and in fact in any type of cancer, is the TP53 (tumour protein 53) gene. This gene is the guardian of the cell cycle: it stops cells from dividing when aberrations in the DNA are detected. TP53 malfunctioning because of a somatic mutation leads to an uncontrolled cell cycle and chromosomal instability, which can lead to the formation of tumours (26). Studies on somatic mutations in vulvar cancer other than in TP53 are limited in number and size. Holway and Growden have found that quite a large percentage of the vulvar cancers and carcinomas in situ they studied carry mutations in PTEN (Phosphatase and tensin homolog), but the study sizes are rather small (27;28). O'Nions, Soufir and Gasco have reported mutations in CDKN2A (cyclin-dependent kinase inhibitor 2A) (29-31). Both PTEN and CDKN2A are also tumour suppressor genes that are involved in signalling pathways that control the cell cycle and stop cells from dividing or even cause cells to undergo apoptosis (32;33). Studies of melanoma, lung, colorectal, and breast carcinomas have shown that the somatic mutation status can be used to predict prognosis and guide tumour-spe-



cific treatment strategies (34-37). Clinical trials have shown promising outcomes of targeted therapies, such as in targeting the PI3K/AKT/mTOR pathway in colorectal cancer (35).

Epigenetic alterations

In addition to genetic mutations, in which the DNA sequence is changed, VSCC might also develop under influence of epigenetic changes. Epigenetic changes, such as hypermethylation, are heritable changes in gene expression without changes in the DNA sequence. Hypermethylation functions as a switch that can turn genes on or off, thus inactivating tumour suppressor genes (38-43). Hypermethylation of the promoters of RASSF2A, MGMT, and TSP1 has been described in vulvar cancer (40).

Predictive morphological factors in vulvar cancer

As mentioned in the first part of the general introduction, clinical and histological parameters that constitute the FIGO staging system have been studied carefully and have proven their prognostic and clinical value in large cohorts of patients. Identifying more risk factors for poor survival is important to further reduce the mortality and morbidity of vulvar cancer patients, but few studies have found new factors of clinical importance (19). To date, lymph node involvement is the only accurate prognostic factor for survival and recurrence (20).

Recent publications have focussed on the potential prognostic value of morphological characteristics in vulvar squamous cell carcinoma (44). Vulvar squamous cell carcinomas with a spray patterned or spindle cell morphology are thought to represent a subgroup of vulvar cancers with a worse prognosis than 'conventional', solid vulvar cancers. Spindle cells are thin, slender, elongated cancer cells that infiltrate stromal tissue and that occur either as single cells or as cords of cells rather than as groups or islands. Spindle cell carcinoma is seen in many different types of cancer and is associated with a worse prognosis in cancers of the oral cavity, oesophagus and lung (45-49). Spindle-shaped epithelial cancer cells have lost their typical epithelial characteristics and seem to gain the ability to infiltrate the underlying stromal and lymphvascular tissue and bud from the primary tumour (50).

Epithelial to Mesenchymal Transition

Research by the Brazilian group of Rocha/AC Camargo Cancer Center (44) has pointed out that this invasive growth pattern in vulvar cancer might be caused by a process called Epithelial to Mesenchymal Transition (EMT). EMT is a process

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that naturally occurs during embryonic development, but can also be seen during cancer progression (51). Cells undergoing EMT lose their epithelial characteristics, such as their cobble stone shape, cell-cell-adhesion and basal cell polarity, and gain mesenchymal traits that provide them with the capacity to migrate as single cells through the extracellular matrix (51). This way, EMT supports the process of solid tumours converting to invasive separate cells. Several signalling pathways control EMT, amongst others Wnt and NOTCH signalling. One of the major inducers of EMT is Transforming Growth Factor-β (TGF-β). In healthy cells, TGF-β has a tumour suppressor role, but in malignant cells, its role changes and TGF-B will induce cell motility and angiogenesis (52;53). Another cell motility promoter that is thought to be involved in EMT is L1-cell adhesion molecule (L1CAM), which is an emerging prognostic factor for metastasis in many cancer subtypes. L1CAM is a membrane alvcoprotein involved in neural development where it has two roles: a cell adhesion function, and a cell motility promoting function. It is expressed by normal nerve tissue, but it is sometimes detected on tumour cell surface (54). The role and relevance of L1CAM has been studied in gynaecological cancers such as endometrial and ovarian cancer, but not yet in vulvar cancer.

Thesis outline

the etiology of HPV positive vulvar cancer is quite well understood, but the HPV independent axis remains to be unravelled. This thesis aims to gain knowledge on the origin of this type of vulvar cancer through the study of two mechanisms: genetic and morphological alterations in vulvar cancer.

In order to study somatic mutations in vulvar cancer, suitable techniques and methods had to be selected, that match the small numbers of patients, the relatively low quality and quantity of available DNA and the lack of prior knowledge we have from the available literature on somatic mutations in vulvar cancer other than TP53. Chapter 2 describes the design of a somatic mutations profiling panel using mass spectrometry that is created especially for gynaecological cancers, focussing on the mutations that are most relevant in cervical, endometrial, ovarian and vulvar cancer. Chapter 3 contains the results from applying this panel to a cohort of 108 vulvar cancer patients that were treated for primary VSCC in the Leiden University Medical Center between 2000 and 2009. In a review of the literature (chapter 4), these data are compared to the current knowledge on genetic and epigenetic changes in vulvar cancer and its precursors.

The majority of vulvar cancers are squamous cell carcinomas, but within this cancer type, morphological sub classifications can be made that can help us understand and predict the progression of vulvar cancers. **Chapter 5** describes a vulvar squamous cell carcinoma type called squamous cell carcinoma with spindle



cell morphology. This type of cancer progression is in some cases thought to develop through the process of EMT, which is reflected by the expression of EMT inducing molecule L1CAM. **Chapter 6** reports the prevalence and prognostic value of L1CAM expression in separate groups of vulvar cancer patients from the Leiden and the Groningen University Medical Centers. In **Chapter 7**, the relation between mutational and morphological data is studied by testing solid and spindle cell carcinomas and their recurrences and metastases for somatic mutations. **Chapter 8** gives an overview of the findings of this thesis and sheds some light on possible future research.

Reference List

- (1) Del Pino M., Rodriguez-Carunchio L, Ordi J. Pathways of vulvar intraepithelial neoplasia and squamous cell carcinoma. Histopathology 2013 Jan;62(1):161-75.
- (2) Gadducci A, Tana R, Barsotti C, Guerrieri ME, Genazzani AR. Clinico-pathological and biological prognostic variables in squamous cell carcinoma of the vulva. Crit Rev Oncol Hematol 2012 Jul;83(1):71-83.
- (3) van der Avoort I, Shirango H, Hoevenaars BM, Grefte JM, de Hullu JA, de Wilde PC, et al. Vulvar squamous cell carcinoma is a multifactorial disease following two separate and independent pathways. Int J Gynecol Pathol 2006 Jan;25(1):22-9.
- (4) Schuurman MS, van den Einden LC, Massuger LF, Kiemeney LA, van der Aa MA, de Hullu JA. Trends in incidence and survival of Dutch women with vulvar squamous cell carcinoma. Eur J Cancer 2013 Dec;49(18):3872-80.
- (5) van de Nieuwenhof HP, Massuger LF, van der Avoort IA, Bekkers RL, Casparie M, Abma W, et al. Vulvar squamous cell carcinoma development after diagnosis of VIN increases with age. Eur J Cancer 2009 Mar;45(5):851-6.
- (6) Dittmer C, Katalinic A, Mundhenke C, Thill M, Fischer D. Epidemiology of vulvar and vaginal cancer in Germany. Arch Gynecol Obstet 2011 Jul;284(1):169-74.
- (7) Sankaranarayanan R, Ferlay J. Worldwide burden of gynaecological cancer: the size of the problem. Best Pract Res Clin Obstet Gynaecol 2006 Apr;20(2):207-25.
- (8) Gonzalez BJ, Magrina JF, Gaffey TA, Hernandez JL, Webb MJ, Cliby WA, et al. Long-term survival and disease recurrence in patients with primary squamous cell carcinoma of the vulva. Gynecol Oncol 2005 Jun;97(3):828-33.
- (9) Coulter J, Gleeson N. Local and regional recurrence of vulval cancer: management dilemmas. Best Pract Res Clin Obstet Gynaecol 2003 Aug;17(4):663-81.
- (10) de Hullu JA, van der Zee AG. Surgery and radiotherapy in vulvar cancer. Crit Rev Oncol Hematol 2006 Oct;60(1):38-58.
- (11) Gaarenstroom KN, Kenter GG, Trimbos JB, Agous I, Amant F, Peters AA, et al. Posto-perative complications after vulvectomy and inguinofemoral lymphadenectomy using separate aroin incisions. Int J Gynecol Cancer 2003 Jul;13(4):522-7.
- (12) Ansink A, van der Velde. Surgical interventions for early squamous cell carcinoma of the vulva. Cochrane Database Syst Rev 2000;(2):CD002036.
- (13) Pilotti S, D'Amato L, Della TG, Donghi R, Longoni A, Giarola M, et al. Papillomavirus, p53 alteration, and primary carcinoma of the vulva. Diagn Mol Pathol 1995 Dec;4(4):239-48.
- (14) Carless MA, Griffiths LR. Cytogenetics of melanoma and nonmelanoma skin cancer. Adv Exp Med Biol 2008;624:227-40.
- (15) McCluggage WG. Premalignant lesions of the lower female genital tract: cervix, vagina and vulva. Pathology 2013 Apr;45(3):214-28.
- (16) Hantschmann P, Sterzer S, Jeschke U, Friese K. P53 expression in vulvar carcinoma, vulvar intraepithelial neoplasia, squamous cell hyperplasia and lichen sclerosus. Anticancer Res 2005 May;25(3A):1739-45.





- (17) Kokka F, Singh N, Faruqi A, Gibbon K, Rosenthal AN. Is differentiated vulval intraepithe-lial neoplasia the precursor lesion of human papillomavirus-negative vulval squamous cell carcinoma? Int J Gynecol Cancer 2011 Oct;21(7):1297-305.
- (18) Hoang LN, Park KJ, Soslow RA, Murali R. Squamous precursor lesions of the vulva: current classification and diagnostic challenges. Pathology 2016 Jun;48(4):291-302.
- (19) Knopp S, Trope C, Nesland JM, Holm R. A review of molecular pathological markers in vulvar carcinoma: lack of application in clinical practice. J Clin Pathol 2009 Mar;62(3):212-8.
- (20) Oonk MH, Hollema H, de Hullu JA, van der Zee AG. Prediction of lymph node metastases in vulvar cancer: a review. Int J Gynecol Cancer 2006 May;16(3):963-71.
- (21) Pecorelli S. Revised FIGO staging for carcinoma of the vulva, cervix, and endometrium. Int J Gynaecol Obstet 2009 May;105(2):103-4.
- (22) Beller U, Quinn MA, Benedet JL, Creasman WT, Ngan HY, Maisonneuve P, et al. Carcinoma of the vulva. FIGO 6th Annual Report on the Results of Treatment in Gynecological Cancer. Int J Gynaecol Obstet 2006 Nov;95 Suppl 1:S7-27.
- (23) van der Zee AG, Oonk MH, de Hullu JA, Ansink AC, Vergote I, Verheijen RH, et al. Sentinel node dissection is safe in the treatment of early-stage vulvar cancer. J Clin Oncol 2008 Feb 20;26(6):884-9.
- (24) Erickson BK, Alvarez RD, Huh WK. Human papillomavirus: what every provider should know. Am J Obstet Gynecol 2013 Mar;208(3):169-75.
- (25) Raspollini MR, Asirelli G, Moncini D, Taddei GL. A comparative analysis of lichen sclerosus of the vulva and lichen sclerosus that evolves to vulvar squamous cell carcinoma. Am J Obstet Gynecol 2007 Dec;197(6):592-5.
- (26) Sigal A, Rotter V. Oncogenic mutations of the p53 tumor suppressor: the demons of the guardian of the genome. Cancer Res 2000 Dec 15;60(24):6788-93.
- (27) Growdon WB, Boisvert SL, Akhavanfard S, Oliva E, Dias-Santagata DC, Kojiro S, et al. Decreased survival in EGFR gene amplified vulvar carcinoma. Gynecol Oncol 2008 Nov;111(2):289-97.
- (28) Holway AH, Rieger-Christ KM, Miner WR, Cain JW, Dugan JM, Pezza JA, et al. Somatic mutation of PTEN in vulvar cancer. Clin Cancer Res 2000 Aug;6(8):3228-35.
- (29) Gasco M, Sullivan A, Repellin C, Brooks L, Farrell PJ, Tidy JA, et al. Coincident inactivation of 14-3-3sigma and p16lNK4a is an early event in vulval squamous neoplasia. Oncogene 2002 Mar 14;21(12):1876-81.
- (30) O'Nions J, Brooks LA, Sullivan A, Bell A, Dunne B, Rozycka M, et al. p73 is over-expressed in vulval cancer principally as the Delta 2 isoform. Br J Cancer 2001 Nov 16;85(10):1551-6.
- (31) Soufir N, Queille S, Liboutet M, Thibaudeau O, Bachelier F, Delestaing G, et al. Inactivation of the CDKN2A and the p53 tumour suppressor genes in external genital carcinomas and their precursors. Br J Dermatol 2007 Mar;156(3):448-53.
- (32) Romagosa C, Simonetti S, Lopez-Vicente L, Mazo A, Lleonart ME, Castellvi J, et al. p16(Ink4a) overexpression in cancer: a tumor suppressor gene associated with senescence and high-grade tumors. Oncogene 2011 May 5;30(18):2087-97.

- (33) Shi Y, Paluch BE, Wang X, Jiang X. PTEN at a glance. J Cell Sci 2012 Oct 15;125(Pt;%20):4687-92.
- (34) Chapman PB, Hauschild A, Robert C, Haanen JB, Ascierto P, Larkin J, et al. Improved survival with vemurafenib in melanoma with BRAF V600E mutation. N Engl J Med 2011 Jun 30;364(26):2507-16.
- (35) De Roock W, Claes B, Bernasconi D, De Schutter J., Biesmans B, Fountzilas G, et al. Effects of KRAS, BRAF, NRAS, and PIK3CA mutations on the efficacy of cetuximab plus chemotherapy in chemotherapy-refractory metastatic colorectal cancer: a retrospective consortium analysis. Lancet Oncol 2010 Aug;11(8):753-62.
- (36) Lynch TJ, Bell DW, Sordella R, Gurubhagavatula S, Okimoto RA, Brannigan BW, et al. Activating mutations in the epidermal growth factor receptor underlying responsiveness of non-small-cell lung cancer to gefitinib. N Engl J Med 2004 May;%20;350(21):2129-39.
- (37) Santarpia L, Qi Y, Stemke-Hale K, Wang B, Young EJ, Booser DJ, et al. Mutation profiling identifies numerous rare drug targets and distinct mutation patterns in different clinical subtypes of breast cancers. Breast Cancer Res Treat 2012 Jul;134(1):333-43.
- (38) Aide S, Lattario FR, Almeida G, do Val IC, Carvalho MG. Promoter hypermethylation of death-associated protein kinase and p16 genes in vulvar lichen sclerosus. J Low Genit Tract Dis 2012 Apr;16(2):133-9.
- (39) Gasche JA, Goel A. Epigenetic mechanisms in oral carcinogenesis. Future Oncol 2012 Nov;8(11):1407-25.
- (40) Guerrero D, Guarch R, Ojer A, Casas JM, Mendez-Meca C, Esteller M, et al. Differential hypermethylation of genes in vulvar cancer and lichen sclerosus coexisting or not with vulvar cancer. Int J Cancer 2011 Jun 15;128(12):2853-64.
- (41) Kelemen LE, Kobel M, Chan A, Taghaddos S, Dinu I. Differentially Methylated Loci Distinguish Ovarian Carcinoma Histological Types: Evaluation of a DNA Methylation Assay in FFPE Tissue. Biomed Res Int 2013;2013:815894.
- (42) Smith ZD, Chan MM, Mikkelsen TS, Gu H, Gnirke A, Regev A, et al. A unique regulatory phase of DNA methylation in the early mammalian embryo. Nature 2012 Apr 19;484(7394):339-44.
- (43) Worsham MJ, Chen KM, Meduri V, Nygren AO, Errami A, Schouten JP, et al. Epigenetic events of disease progression in head and neck squamous cell carcinoma. Arch Otolaryngol Head Neck Surg 2006 Jun;132(6):668-77.
- (44) Rodrigues IS, Lavorato-Rocha AM, de MM, Stiepcich MM, de Carvalho FM, Baiocchi G, et al. Epithelial-mesenchymal transition-like events in vulvar cancer and its relation with HPV. Br J Cancer 2013 Jul 9;109(1):184-94.
- (45) Ahluwalia H, Gupta SC, Gupta SC. Pathology in focus. Spindle-cell carcinoma of the nasal septum. J Laryngol Otol 1996 Mar;110(3):284-7.
- (46) Minami SB, Shinden S, Yamashita T. Spindle cell carcinoma of the palatine tonsil: report of a diagnostic pitfall and literature review. Am J Otolaryngol 2008 Mar;29(2):123-5.
- (47) Pelosi G, Sonzogni A, De PT, Galetta D, Veronesi G, Spaggiari L, et al. Review article: pulmonary sarcomatoid carcinomas: a practical overview. Int J Surg Pathol 2010 Apr;18(2):103-20.





- (48) Santeusanio G, Schiaroli S, Anemona L, Sesti F, Valli E, Piccione E, et al. Carcinoma of the vulva with sarcomatoid features: a case report with immunohistochemical study. Gynecol Oncol 1991 Feb;40(2):160-3.
- (49) Travis WD. Sarcomatoid neoplasms of the lung and pleura. Arch Pathol Lab Med 2010 Nov;134(11):1645-58.
- (50) Kalluri R, Weinberg RA. The basics of epithelial-mesenchymal transition. J Clin Invest 2009 Jun;119(6):1420-8.
- (51) Acloque H, Adams MS, Fishwick K, Bronner-Fraser M, Nieto MA. Epithelial-mesenchymal transitions: the importance of changing cell state in development and disease. J Clin Invest 2009 Jun;119(6):1438-49.
- (52) Hawinkels LJ, Paauwe M, Verspaget HW, Wiercinska E, van der Zon JM, van der Ploeg K, et al. Interaction with colon cancer cells hyperactivates TGF-beta signaling in cancerassociated fibroblasts. Oncogene 2014 Jan 2;33(1):97-107.
- (53) Massague J. TGFbeta signalling in context. Nat Rev Mol Cell Biol 2012 Oct;13(10):616-30.
- (54) Gavert N, Ben-Shmuel A, Raveh S, Ben-Ze'ev A. L1-CAM in cancerous tissues. Expert Opin Biol Ther 2008 Nov;8(11):1749-57.

