

Epidemiological transition in Indonesia : impact of helminths and urbanization on the development of Type 2 diabetes

Tahapary, D.L.; Tahapary D.L.

Citation

Tahapary, D. L. (2017, September 19). *Epidemiological transition in Indonesia : impact of helminths and urbanization on the development of Type 2 diabetes*. Retrieved from https://hdl.handle.net/1887/52966

Version: Not Applicable (or Unknown)

License: License agreement concerning inclusion of doctoral thesis in the

Institutional Repository of the University of Leiden

Downloaded from: https://hdl.handle.net/1887/52966

Note: To cite this publication please use the final published version (if applicable).

Cover Page



Universiteit Leiden



The handle http://hdl.handle.net/1887/52966 holds various files of this Leiden University dissertation.

Author: Tahapary, D.L.

Title: Epidemiological transition in Indonesia: impact of helminths and urbanization on

the development of Type 2 diabetes

Issue Date: 2017-09-19



Chapter 4

EFFECT OF ANTHELMINTIC TREATMENT ON LEPTIN, ADIPONECTIN, AND LEPTIN TO ADIPONECTIN RATIO: A Randomized Controlled Trial

Dicky L. Tahapary^{1,2,3,4}, Karin de Ruiter², Ivonne Martin^{5,6}, Eric A.T. Brienen², Lisette van Lieshout², Yenny Djuardi^{3,7,} Clara C. Djimandjaja³, Jeanine J Houwing-Duistermaat^{5,8}, Pradana Soewondo^{1,4}, Erliyani Sartono², Taniawati Supali^{3,7#}, Johannes W.A. Smit^{9,10#}, Maria Yazdanbakhsh^{2#\$}

(Manuscript in submission)

ABSTRACT

Aims/hypothesis: We aimed to assess the role of adipokines in mediating the effect of helminths on insulin resistance. We hypothesized that the increase in IR after anthelmintic treatment in helminth-infected subjects is mediated by a shift in leptin to adiponectin ratio driving a more proinflammatory state.

Methods: Serum samples were obtained from a randomized-controlled trial of anthelmintic treatment in an area endemic for soil-transmitted helminths (STH), Flores Island, Indonesia. All subjects in the study area received albendazole or matching placebo for three consecutive days. This three-monthly treatment regimen was given for four rounds. We measured leptin, adiponectin, and resistin changes in those >16 years old. STH infections were assessed by microscopy and PCR.

Results: In STH-infected subjects, anthelmintic treatment significantly increased the ratio of leptin to adiponectin [treatment effect factor (95% CI), p-value for interaction: $1.20 \ (1.06 - 1.35)$, p=0.010], which largely stemmed from a significant reduction in adiponectin [0.91 (0.85 - 0.98), p=0.020] and a trend for an increase in leptin level [1.10 (1.00 - 1.21)], p=0.119]. No significant effect on resistin level was observed. This increase in leptin to adiponectin ratio seemed to contribute to the observed effect of deworming on increased IR as adjustment for leptin to adiponectin ratio attenuated the effect on IR from 1.07 (1.01 – 1.14, p=0.023) to 1.05 (0.99 – 1.11, p=0.075).

Conclusions: Anthelmintic treatment in STH-infected subjects increases leptin to adiponectin ratio which may in small part contribute to the modest increase in IR. Further studies will be needed to assess the effect of the changes in adipokine levels on the host immune response and metabolism.

Keywords: Helminths. Adipokines. Leptin. Adiponectin. Leptin to adiponectin ratio. Insulin resistance

Abbreviations: AT: adipose tissue, L/A: leptin to adiponectin, STH: soil-transmitted helminth, T2D: type 2 diabetes.

INTRODUCTION

Emerging evidence suggests that helminths might confer protection against the development of type 2 diabetes (T2D),[1-5] presumably by modulating the host immune responses.[6-8] Thus, in addition to the more established risk factors, such as sedentary lifestyle and high-energy foods, current deworming programs in parallel with rapid socioeconomic development might potentially contribute to the development of T2D in many low and middle-income countries.[6] In line with this, we have recently reported that removal of helminth infections increases insulin resistance (IR),[9] which is mainly mediated by the increase in adiposity,[9] suggesting a central role of adipose tissue (AT).[10-13]

Human AT secretes various adipokines, most notably leptin and adiponectin, affecting metabolic homeostasis and immune regulation.[14] Leptin and adiponectin have been consistently shown to be positively and negatively associated with IR, respectively.[14] Whereas leptin promotes pro-inflammatory immune responses and inhibits the proliferation of regulatory T-cells, adiponectin induces the secretion of anti-inflammatory cytokines.[15] The imbalance between those two adipokines, leptin to adiponectin (L/A) ratio, has been reported to be associated with pro-inflammatory conditions and IR.[16, 17]

Assessment of adipokines might provide a valuable insight into the role of human AT in mediating the helminths effect on metabolic homeostasis. To our knowledge, no studies have been published so far on the association between helminth infections and adipokines, except for resistin.[18] Therefore, we measured leptin, adiponectin, and resistin in serum samples obtained from a randomized-controlled trial of anthelmintic treatment in an area endemic for soil-transmitted helminth (STH).[19] We hypothesized that the increase in IR after anthelmintic treatment in helminth-infected subjects might be mediated by a shift in L/A ratio towards a more pro-inflammatory state.

METHODS

This present study is part of a household-based cluster-randomized double-blind placebo-controlled anthelmintic trial (The Sugarspin study), conducted in Nangapanda, Flores, an endemic area for soil-transmitted helminth (STH). [19] The primary outcome of the Sugarspin study is changes in insulin resistance (IR), as assessed using the homeostatic model assessment of IR (HOMA-IR), after anthelmintic treatment, which has been published recently.[9] Written informed

consent was obtained from all participants. The study was approved by the ethics committee of Faculty of Medicine, Universitas Indonesia (FKUI) (ref: 549/H2·F1/ETIK/2013), and filed by the ethics committee of Leiden University Medical Center (LUMC). The trial is registered as a clinical trial (http://www.isrctn.com/ISRCTN75636394).

The population was randomised by blocks at household level. After randomisation, all subjects in the study area, except children <2 years old and pregnant women, received a single tablet of albendazole (400mg) or matching placebo for three consecutive days with direct supervision. This three-monthly treatment regimen was given for four rounds. All measurements and sample collections were performed at baseline and 6 weeks after the end of the fourth treatment round (follow-up).[19] All subjects without sufficient sera samples, and/or incomplete data on body mass index, and soil-transmitted helminth (STH) infection status at baseline were excluded from the present study. Subjects receiving active treatment for diabetes were also excluded from analysis.

All subjects ≥16 years old were invited to undergo clinical measurements and blood drawing after an overnight fast.[19] Body weight and height were measured, and body mass index (BMI) was calculated as weight (kg) divided by square of height (m). Adipokines (leptin, adiponectin and resistin) were measured by ELISA using commercial reagents (DuoSet ELISA R&D System Europe Ltd, Abingdon, UK), according to the manufacturer's protocol. Leptin to adiponectin (L/A) ratio was calculated by L/A= leptin level (ng/ml) / adiponectin level (ug/ml).[17] Soil-transmitted helminth infection status was assessed using both microscopy (Kato Katz) and PCR, which was further stratified by the number of species a subject was infected with at baseline (no infection, single infection, multiple infection).[9]

Statistical Analysis

Leptin, adiponectin, L/A ratio, and resistin were log-transformed (log10) for analysis and summarized as geometric mean [95% confidence interval (CI)]. The effect of anthelmintic treatment on adipokine was assessed using mixed models to account for the correlation within households, as described previously.[9] The treatment effect estimates were the regression coefficient obtained from mixed models (β) indicating changes in log10 (leptin, adiponectin, L/A ratio, resistin) of subjects using albendazole compared to placebo. The treatment effect factors (10 β) are multiplicative instead of additive. Thus treatment effect factors indicate the proportional change for each

variable (leptin, adiponectin, resistin, L/A ratio), in comparison to the placebo. All models were fitted using the lme4 package (R software).

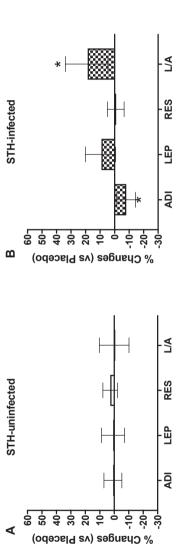
RESULTS

At baseline, the prevalence of STH infection was 42.0% (503/1195) and 54.1% (760/1405), as assessed by microscopy and PCR respectively. Serum leptin, adiponectin, L/A ratio, and resistin levels were similar in both treatment arms (**Table 1**). The consort diagram of the present study is shown in **figure S1**.

Similar to the main study, anthelmintic treatment significantly reduced the prevalence of STH infections, as assessed by microscopy or PCR (**Table S1**). In comparison to placebo, albendazole treatment had no effect on adipokine levels in subjects without STH infections (**Figure 1A**). In STH-infected subjects, as assessed by microscopy, albendazole treatment increased L/A ratio [treatment effect factor (95% CI), p-value for interaction: 1.20 (1.06 – 1.35), p=0.010], which was mostly derived from a significant reduction in adiponectin level [0.91 (0.85 – 0.98), p=0.020] and a trend for an increase in leptin level [1.10 (1.00 – 1.21)], p=0.119] (**Figure 1B**). No significant treatment effect on resistin level was observed [1.00 (0.94 – 1.05), p=0.363] (**Figure 1B**).

Table 1. Study Population

	Placebo N=807	Albendazole N=750
Age (in years, mean, SD)	41.9 (15.4)	42.6 (15.5)
Sex (female %, n/N)	62.0 (500/807)	59.9 (449/750)
Body Mass Index (kg/m², mean, SD)	22.5 (4.0)	22.5 (4.0)
Leptin to adiponectin ratio [geomean (95% CI)]	1.38 (1.25 – 1.53)	1.35 (1.21 – 1.51)
Leptin (ng/ml) [geomean (95% CI)]	7.1 (6.5 – 7.7)	6.7 (6.1 – 7.4)
Adiponectin (ug/ml) [geomean (95% CI)]	5.1 (4.9 – 5.4)	5.0 (4.7 – 5.3)
Resistin (ng/ml) [geomean (95% CI)]	15.6 (15.0 – 16.2)	15.7 (15.1 – 16.4)
Helminth-infected by microscopy (%, n/N)	43.5 (270/620)	40.5 (233/575)
- Single species	28.2 (175/620)	26.4 (152/575)
- Multiple species	15.3 (95/620)	14.1 (81/575)
Helminth-infected by PCR (%, n/N)	53.8 (392/729)	54.4 (368/676)
- Single species	31.7 (231/729)	35.2 (238/676)
- Multiple species	22.1 (161/729)	19.2 (130/676)



5TH-uninfected and (B) STH-infected subjects, as assessed by microscopy, are presented as proportion of changes (95% CI) between pre and post treatment Analysis was performed on 1183 subjects, after excluding 12 subjects with diabetes. Treatment effect estimates were the regression coefficient (β) obtained from mixed models indicating changes in \log (ADI or LEP or RES or L/A); the treatment effect factors (10 β) are proportional instead of additive. Thus, treatment Figure 1. Effect of anthelmintic treatment on adiponectin, leptin, resistin, and leptin to adiponectin ratio in soil-transmitted helminth (STH)-infected and uninfected subjects. The effect of anthelmintic treatment on adiponectin (ADI), leptin (LEP), resistin (RES), and leptin to adiponectin ratio (L/A) in (A) in the albendazole group compared to the placebo group which is set to zero. Adiponectin, leptin, resistin, and L/A ratio were log-transformed for analysis. effect factors indicate the proportional change in each variable in comparison to the placebo group. *p<0.05

Pathway analysis showed that adjustment for changes in BMI partly attenuated the treatment effect on adiponectin level [to 0.92 (0.86-0.99), p=0.030] and L/A ratio [to 1.13 (1.02-1.26), p=0.040]. Analysis of the Sugarspin trial primary outcome revealed that the increased IR after anthelmintic treatment in infected subjects might be due to an increased BMI and a reduced eosinophil counts.[9] Therefore we also assessed whether the increase in L/A ratio contributes to the increased IR after treatment in helminth-infected subjects. This analysis showed that adjustment for changes in L/A ratio, attenuated the treatment effect on IR from 1.07 (1.01-1.14, p=0.023) to 1.05 (0.99-1.11, p=0.075), even more than adjustment for changes in BMI [to 1.06 (1.00-1.12), p=0.048].

When light infections were also considered by using PCR, albendazole treatment did not significantly increase L/A ratio [1.10 (1.00 – 1.22), p=0.31], despite a significantly reduced adiponectin level [0.94 (0.88 – 0.99), p=0.048]. No significant treatment effect was observed on the level of leptin, nor resistin (**Figure S2**). Next, we further stratified STH-infected subjects based on the number of STH species a subject was infected with at baseline. In subjects with multiple STH infections, albendazole significantly increased L/A ratio [1.25 (1.06 – 1.48), p=0.041] which derived from a significant reduction in adiponectin level [0.88 (0.80 – 0.97), p=0.013] and a non-significant increase in leptin level [1.10 (0.97 – 1.25), p=0.47]. (**Figure S3**) Using microscopy, a more pronounced reduction in adiponectin [0.88 (0.78 – 0.99), p=0.049] was observed in subjects infected with multiple STH species. The treatment effect on L/A ratio [1.18 (0.96 – 1.45), p=0.157] and leptin level [1.04 (0.89 - 1.22), p=0.71] in subjects infected with multiple species did not reach statistical significance (**Figure S4**).

DISCUSSION

Our study is the first to report the effect of anthelmintic treatment on serum adipokine levels. In STH-infected subjects, treatment significantly increased L/A ratio, which has been reported to be associated with low-grade inflammation [16] and IR.[16, 17] The increased L/A ratio was derived by the significant reduction in adiponectin level, and to a lesser extent, a trend of increase in leptin level. As adiponectin induces the secretion of anti-inflammatory cytokines,[15] while leptin increases Th1, suppresses Th2, and can act as a negative signal for the proliferation of human T regulatory cells [20], these changes may reverse the helminth-associated type 2 and regulatory immune responses, and presumably contribute to the development of IR. Indeed, adjustment for the increase in L/A ratio attenuated

the treatment-associated increase in IR, observed in the main trial,[9] even more than adjustment for increase in BMI. This suggests that adipokines play a relatively more important role than the adiposity in the mediation of helminth-associated beneficial effect on IR.

Using PCR, a more sensitive method, able to detect non-clinically relevant STH infections, the treatment effects were less in magnitude, as it significantly reduced adiponectin level only, but to a lesser extent. In line with this, in subjects with multiple STH infections, associated with a higher infection intensity,[9] treatment resulted in more pronounced effects, namely a significant reduction in adiponectin level, a trend for increase in leptin level, as well as a significant increase in L/A ratio. Except for the effect on adiponectin, these pronounced treatment effects were not observed when infection was assessed by microscopy, which might be due to the lower number of subjects who were found to be infected with multiple species, when using microscopy.

Despite having an ideal study design to study the causal relationship between helminth infections and adipokine levels, and to assess the contribution of adipokine levels to the increased IR after anthelmintic treatment, our study would have been more complete if we would have assessed food intake, appetite, and physical activity. In addition, measurements of other hormones that influence metabolism, such as ghrelin and cortisol, as well as analysis of AT biopsies and gut microbiome, could provide a more complete overview on how helminths may modulate human metabolism.

In conclusion, anthelmintic treatment in STH-infected subjects increases L/A ratio which may in small part contribute to the increased IR. Further studies will be needed to assess the effect of these changes in adipokine levels on the host metabolism and modulation of the host immune responses.

Acknowledgements

The authors would like first to thank all study participants in Nangapanda, Ende, Flores, Indonesia. The authors would also like to thank The Indonesian Directorate General of Higher Education for providing scholarship to two PhD candidates involved in this project. The authors would like to thank all local government and health officers in Nangapanda who supported this project, and also all field workers from Universitas Indonesia and Nangapanda. The authors would like to thank Itziar Munos Pagazaurtundua for measuring the adipokines. The authors would also like to thank all colleagues at Department of Parasitology FKUI and LUMC for their technical supports.

Data Availability

The datasets generated during and/or analysed during the current study are available from the corresponding author on reasonable request.

Funding

This study was funded by The Royal Netherlands Academy of Arts and Science (KNAW), Ref 57-SPIN3-JRP and Universitas Indonesia (Research Grant BOPTN 2742/H2.R12/HKP.05.00/2013.).

Duality of interests

All authors declare no competing interests. The funder of the study had no role in study design, data collection, data analysis, data interpretation, or writing of the report. The senior authors TS, JWAS, MY had final responsibility for the decision to submit for publication.

Contribution statement

D.L.T. is a medical doctor in charge of the field study, involved in setting up the study, supervising gathering of data, treatment, clinical care, follow up of the study population, analyzed the data and wrote the manuscript. K.R. is a medical biologist in charge of the field study, involved in setting up the laboratory in the study area, performing the immunological analysis, supervising the data cleaning, and the follow up of the study population. I.M. is a mathematician who is developing methods to analyze the complex data generated during the lifetime of the project and was involved in the randomization and data analysis. L.v.L. is a parasitologist who was involved in the performance and analysis of diagnostic assays for the detection of helminths in stool samples. E.A.T.B. is a technician who develop, optimized and performed multiplex real time PCR for detection of helminth infections. P.S. is an endocrinologist who advised on the metabolic aspects of the study. Y.D. is a medical doctor who was involved in coordinating the field study and advised on the immunological and parasitological aspects of the study. C.C.D. is a nurse who was involved in the field study, especially gathering of data, supervising anthelmintic treatment, and follow up the study population. J.J.H. is a biostatistician who developed the study, and was involved in supervising sample size calculation, randomization and statistical analysis. E.S. is an immunoparasitologist who was involved in coordinating the study and advising on parasitological and immunological aspects of the study and supervised the writing of the manuscript. T.S. is a parasitologist who developed the study and is the Indonesian coordinator of the SUGARSPIN program. J.W.A.S. is an endocrinologist who developed the study, supervised the writing of the manuscript, and is the Dutch coordinator of the SUGARSPIN program. M.Y. is an immunologist who developed the study, supervised the writing of the manuscript and is the scientific coordinator of the SUGARSPIN program. All authors read and approved the final manuscript.

Author Details

*These authors have contributed equally; \$Corresponding author.¹Department of Internal Medicine, Division of Endocrinology, Faculty of Medicine Universitas Indonesia, Dr. Cipto Mangunkusumo National General Hospital, Jakarta, Indonesia; ²Department of Parasitology, Leiden University Medical Center, Leiden, The Netherlands. ³Nangapanda Community Research Cluster, The Indonesian Medical Education and Research Institute, Universitas Indonesia, Jakarta, Indonesia; ⁴Metabolic, Cardiovascular, and Aging Research Cluster, The Indonesian Medical Education and Research Institute, Universitas Indonesia, Jakarta, Indonesia; ⁵Department of Medical Statistics and Bioinformatics, Leiden University Medical Center, Leiden, The Netherlands; ôDepartment of Mathematics, Parahyangan Catholic University, Bandung, Indonesia; ∂Department of Parasitology, Faculty of Medicine Universitas Indonesia, Jakarta, Indonesia; ôDepartment of Statistics, University of Leeds, Leeds, United Kingdom; ôDepartment of Internal Medicine, Radboud University Medical Center, Nijmegen, The Netherlands; ¹Department of Internal Medicine, Leiden University Medical Center, Leiden, The Netherlands.

REFERENCES

- Tracey EF, McDermott RA, McDonald MI.
 Do worms protect against the metabolic
 syndrome? A systematic review and metaanalysis. Diabetes Res Clin Pract 2016; 120:
 209-20.
- Wiria AE, Hamid F, Wammes LJ, et al. Infection with Soil-Transmitted Helminths Is Associated with Increased Insulin Sensitivity. PLoS One 2015; 10(6): e0127746.
- Chen Y, Lu J, Huang Y, et al. Association of previous schistosome infection with diabetes and metabolic syndrome: a cross-sectional study in rural China. J Clin Endocrinol Metab 2013; 98(2): E283-7.
- Shen SW, Lu Y, Li F, et al. The potential long-term effect of previous schistosome infection reduces the risk of metabolic syndrome among Chinese men. Parasite Immunol 2015; 37(7): 333-9.
- Hays R, Esterman A, Giacomin P, Loukas A, McDermott R. Does Strongyloides stercoralis infection protect against type 2 diabetes in humans? Evidence from Australian Aboriginal adults. Diabetes Res Clin Pract 2015; 107(3): 355-61.
- Wammes LJ, Mpairwe H, Elliott AM, Yazdanbakhsh M. Helminth therapy or elimination: epidemiological, immunological, and clinical considerations. The Lancet Infectious diseases 2014; 14(11): 1150-62.
- de Ruiter K, Tahapary DL, Sartono E, et al. Helminths, hygiene hypothesis and type 2 diabetes. Parasite Immunol 2016.
- Berbudi A, Ajendra J, Wardani AP, Hoerauf A, Hubner MP. Parasitic helminths and their beneficial impact on type 1 and type 2 diabetes. Diabetes Metab Res Rev 2015.
- Tahapary DL, de Ruiter K, Martin I, et al. Effect of Anthelmintic Treatment on Insulin Resistance: A Cluster-Randomized Placebo-Controlled Trial in Indonesia. Clinical infectious diseases: an official publication of the Infectious Diseases Society of America 2017.
- Hussaarts L, Garcia-Tardon N, van Beek L, et al. Chronic helminth infection and helminth-derived egg antigens promote adipose tissue M2 macrophages and

- improve insulin sensitivity in obese mice. FASEB J **2015**; 29(7): 3027-39.
- Yang Z, Grinchuk V, Smith A, et al. Parasitic nematode-induced modulation of body weight and associated metabolic dysfunction in mouse models of obesity. Infect Immun 2013; 81(6): 1905-14.
- Wu D, Molofsky AB, Liang HE, et al. Eosinophils sustain adipose alternatively activated macrophages associated with glucose homeostasis. Science 2011; 332(6026): 243-7.
- Berbudi A, Surendar J, Ajendra J, et al. Filarial Infection or Antigen Administration Improves Glucose Tolerance in Diet-Induced Obese Mice. J Innate Immun 2016; 8(6).
- Ouchi N, Parker JL, Lugus JJ, Walsh K. Adipokines in inflammation and metabolic disease. Nat Rev Immunol 2011; 11(2): 85-97.
- Carbone F, La Rocca C, Matarese G. Immunological functions of leptin and adiponectin. Biochimie 2012; 94(10): 2082-8.
- 16. Chou HH, Hsu LA, Wu S, Teng MS, Sun YC, Ko YL. Leptin-to-Adiponectin Ratio is Related to Low Grade Inflammation and Insulin Resistance Independent of Obesity in Non-Diabetic Taiwanese: A Cross-Sectional Cohort Study. Acta Cardiol Sin 2014; 30(3): 204-14.
- Finucane FM, Luan J, Wareham NJ, et al. Correlation of the leptin:adiponectin ratio with measures of insulin resistance in nondiabetic individuals. Diabetologia 2009; 52(11): 2345-9.
- Jang JC, Chen G, Wang SH, et al. Macrophage-derived human resistin is induced in multiple helminth infections and promotes inflammatory monocytes and increased parasite burden. PLoS Pathog 2015; 11(1): e1004579.
- Tahapary DL, de Ruiter K, Martin I, et al. Helminth infections and type 2 diabetes: a cluster-randomized placebo controlled SUGARSPIN trial in Nangapanda, Flores, Indonesia. BMC Infect Dis 2015; 15: 133.
- De Rosa V, Procaccini C, Cali G, et al. A key role of leptin in the control of regulatory T cell proliferation. Immunity 2007; 26(2): 241-55.

SUPPLEMENTARY MATERIALS

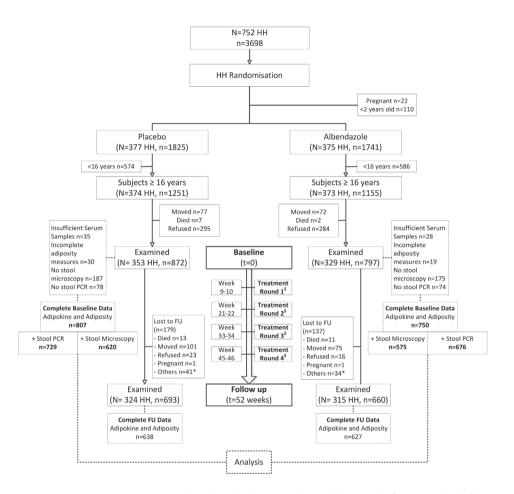
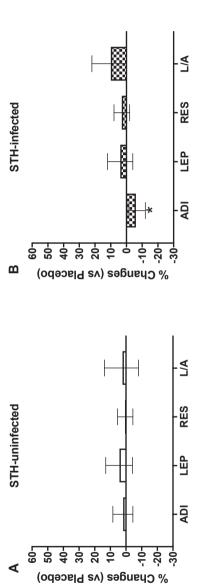
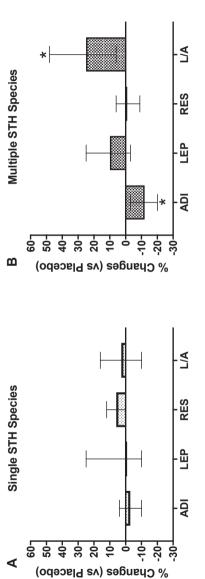


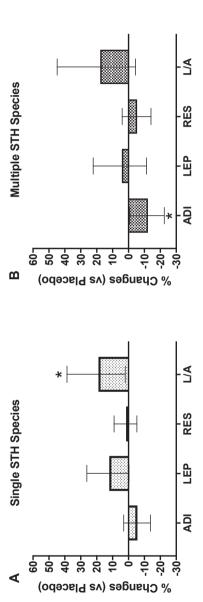
Figure S1. Consort Diagram. Baseline data (t=0) were collected during the first 8 weeks before the start of the drug administration. ^{\$}Single dose of albendazole or matching placebo was given for three consecutive days to all household members, except children below 2 years of age and pregnant women. *Other reasons of lost to follow-up were harvesting crops, working on funeral ceremonies, severely ill, hospitalized, nursing mother. HH: Household, FU: Follow Up.



infected and uninfected subjects, as assessed by PCR. The effects of anthelmintic treatment on adiponectin (ADD), leptin (LEP), resistin (RES), and leptin to adiponectin ratio (L/A) are presented as proportion of changes between pre and post treatment in the albendazole group compared to the placebo by PCR. Adiponectin, leptin, resistin, and L/A ratio were log-transformed for analysis. Analysis was performed on 1387 subjects, after excluding 14 subjects L/A); the treatment effect factors (108) are proportional instead of additive. Thus, treatment effect factors indicate the proportional change in each variable group which is set to zero. The effects of treatment are presented for each group of subjects: (A) STH-uninfected and (B) STH-infected subjects, as assessed with diabetes. Treatment effect estimates were the regression coefficient (eta) obtained from mixed models indicating changes in log (ADI or LEP or RES or Figure S2. Effect of anthelmintic treatment on adiponectin, leptin, resistin, and leptin to adiponectin ratio in soil-transmitted helminth (STH)in comparison to the placebo group. *p<0.05.



and leptin to adiponectin ratio (L/A) are presented as proportion of changes between pre and post treatment in the albendazole group compared to the placebo group which is set to zero. The effects of treatment are presented for each group of STH-infected subjects with: (A) single STH species, (B) multiple Figure S3. Effect of anthelmintic treatment on adiponectin, leptin, resistin, and leptin to adiponectin ratio stratified by number of helminth species 5TH species. Adiponectin, leptin, resistin, and L/A ratio were log-transformed for analysis. Analysis was performed on 1387 subjects, after excluding 14 subjects with diabetes. Treatment effect estimates were the regression coefficient (B) obtained from mixed models indicating changes in log (ADI or LEP or RES or L/A); the treatment effect factors (10 \(\)) are proportional instead of additive. Thus, treatment effect factors indicate the proportional change in each a subject was infected with at baseline, as assessed by PCR. The effects of anthelmintic treatment on adiponectin (ADI), leptin (LEP), resistin (RES), variable in comparison to the placebo group. *p<0.05.



(B) multiple STH species. Adiponectin, leptin, resistin, and leptin to adiponectin ratio were log-transformed for analysis. Analysis was performed on 1183 Figure S4. Effect of anthelmintic treatment on adiponectin, leptin, resistin, and leptin to adiponectin ratio stratified by number of helminth species a subject was infected with at baseline, as assessed by microscopy. The effects of anthelmintic treatment on adiponectin (ADI), leptin (LEP), resistin subjects, after excluding 12 subjects with diabetes. Treatment effect estimates were the regression coefficient (β) obtained from mixed models indicating changes in log (ADI or LEP or RES or L/A); the treatment effect factors (10 B) are proportional instead of additive. Thus, treatment effect factors indicate the (RES), and leptin to adiponectin ratio (L/A) are presented as proportion of changes between pre and post treatment in the albendazole group compared to the placebo group which is set to zero. The effects of treatment are presented for each group of STH-infected subjects with: (A) single STH species, proportional change in each variable in comparison to the placebo group. *p<0.05

Table S1. Effect of Anthelmintic Treatment on Soil-transmitted Helminth Prevalence

	Placebo		Albendazole		
Method	Baseline	Follow-up	Baseline	Follow-up	p-value*
Microscopy (%, n/N)	43.5% (270/620)	26.8% (166/497)	40.5% (233/575)	5.2% (24/466)	<0.0001
PCR (%, n/N)	53.8% (392/729)	45.0% (250/555)	54.4% (368/676)	10.4% (55/529)	<0.0001

^{*}Analyzed using logistic model (Ime4 package R software) with random household effects and random subject effects

Table S2. Pathway analysis on the role of leptin to adiponectin ratio in the increased insulin resistance after anthelmintic treatment

	Crude	L/A Ratio	BMI	L/A Ratio + BMI
HOMA-IR*	1.07 (1.01 – 1.14)	1.05 (0.99 – 1.11)	1.06 (1.00 – 1.12)	1.05 (0.99 – 1.11)
	p=0.023	p=0.075	p=0.048	p=0.075

^{*}Analyses were performed using linear mixed model in unadjusted model (crude) and adjusted for leptin to adiponectin (L/A) ratio, BMI, or both.