



Universiteit  
Leiden  
The Netherlands

## **Glycoproteomics characterization of immunoglobulins in health and disease**

Plomp, H.R.

### **Citation**

Plomp, H. R. (2017, May 31). *Glycoproteomics characterization of immunoglobulins in health and disease*. Retrieved from <https://hdl.handle.net/1887/49752>

Version: Not Applicable (or Unknown)

License: [Licence agreement concerning inclusion of doctoral thesis in the Institutional Repository of the University of Leiden](#)

Downloaded from: <https://hdl.handle.net/1887/49752>

**Note:** To cite this publication please use the final published version (if applicable).

Cover Page



Universiteit Leiden



The handle <http://hdl.handle.net/1887/49752> holds various files of this Leiden University dissertation

**Author:** Plomp, H.R.

**Title:** Glycoproteomics characterization of immunoglobulins in health and disease

**Issue Date:** 2017-05-31

---

# Chapter 1: Introduction

---

**Modified from:** *Mol Cell Proteomics* 2016, 15(7), 2217-28

**Authors:** Rosina Plomp<sup>1</sup>, Albert Bondt<sup>1,2</sup>, Noortje de Haan<sup>1</sup>, Yoann Rombouts<sup>1,3</sup>, Manfred Wuhrer<sup>1</sup>

<sup>1</sup>*Center for Proteomics and Metabolomics, Leiden University Medical Center, Leiden, The Netherlands;*

<sup>2</sup>*Department of Rheumatology, Leiden University Medical Center, Leiden, The Netherlands;*

<sup>3</sup>*present address: Institut de Pharmacologie et de Biologie Structurale, Université de Toulouse, CNRS, UPS, France;*

---

**Table of Contents**

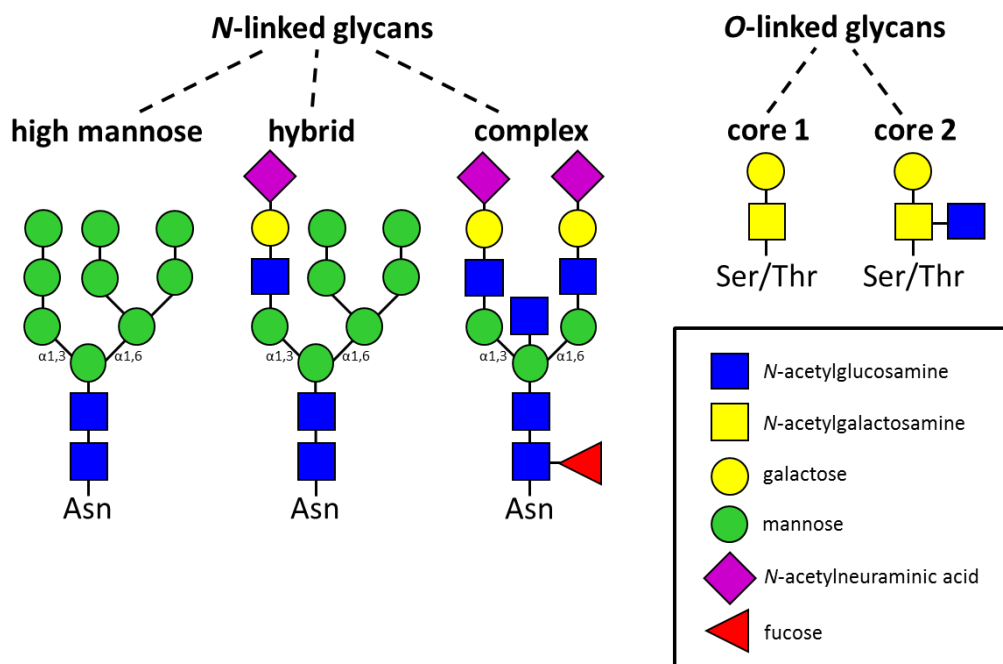
---

1.1: General introduction .....	10
1.1.1: Glycan structure .....	10
1.1.2: <i>N</i> -glycosylation .....	11
1.1.3: <i>O</i> -glycosylation .....	12
1.1.4: Functions of glycosylation .....	13
1.1.5: Glycosylation of immunoglobulins .....	14
1.2: Recent advances in clinical glycoproteomics of immunoglobulins (Igs) .....	16
1.2.1: Introduction.....	16
1.2.2: IgG .....	19
1.2.2.1: Fc glycosylation at Asn180/176/227/177 ( 'Asn297') .....	19
1.2.2.2: Fab glycosylation .....	24
1.2.2.3: Additional <i>N</i> - and <i>O</i> -glycosylation of IgG3 .....	25
1.2.2.4: Antigen-specific IgG.....	26
1.2.3: IgA.....	28
1.2.4: IgM.....	30
1.2.5: IgE.....	31
1.2.6: Conclusions and perspectives .....	31
1.3: Scope of thesis.....	33
References.....	34

## 1.1: General introduction

Carbohydrates, also known as sugars or saccharides, are commonly thought of solely as energy repositories, but they are also a vital part of many other cellular biomolecules. In plants and fungi, respectively cellulose and chitin give the cell wall its strength, and in nucleic acids the sugars deoxyribose and ribose form the backbone involved in storage of the genetic code. A lesser known function of carbohydrates is the alteration of proteins by attachment of an oligosaccharide, called a glycan. It is estimated that half of all mammalian proteins is glycosylated (1), and the purpose of these glycans is manifold: some alter the stability or half-life of proteins, others influence the interaction with ligands, and for many others the function is still unknown.

### 1.1.1: Glycan structure



**Figure 1.1:** N-linked and O-linked glycosylation, and the most common types thereof. Monosaccharides are represented schematically.

Glycans consist of monosaccharides which are linked together by enzymes through glycosidic linkages. The number of monosaccharide building blocks in humans is limited: glycans can consist of hexoses (mannose, galactose, glucose and fucose), N-acetylhexosamines (N-acetylglucosamine and N-acetylgalactosamine), and sialic acid (N-acetylneuraminic acid). Additional types of monosaccharides can be found in other organisms, such as N-glycolylneuraminic acid in other mammals and xylose in plants.

Each glycosylation site is usually occupied by a range of glycan structures, called glycoforms. Glycans are an exceedingly heterogeneous class of polysaccharides. Most are oligosaccharides consisting of less than 20 monosaccharides, but some are large sprawling structures with the total mass of glycans exceeding that of their carrier protein (2). Glycans can be linear, but more often are branched. The reason for this vast diversity of glycan structures lies in their synthesis: this is not a template-driven process as with DNA or proteins, but rather the types of monosaccharides and glycosidic linkages are governed by the activity of glycosyltransferases and glycosidases which assemble and trim the glycan as the glycoprotein travels through various compartments within the cell. Even after secretion, glycans can still be altered by glycosyltransferases and glycosidases which are present in the extracellular matrix (3). Furthermore, the surrounding protein structure influences the structure of the glycan, by stereochemically restricting enzyme access to a site. Moreover, sorting sequences guide the path of proteins through cell compartments, thereby controlling their access to the glycosylation machinery. Hence, two glycosylation sites on the same protein can be occupied by very different glycan structures, exhibiting different functions. There are two main types of protein glycosylation: *N*- and *O*-glycosylation.

### 1.1.2: *N*-glycosylation

The presence of an *N*-glycosylation site in mammalian proteins is relatively easy to predict when the protein sequence is known: *N*-glycans are covalently attached to the nitrogen (N) of the amino acid asparagine (Asn) in the consensus sequence Asn-X-Ser/Thr, with X being any amino acid except for proline. However, even though the consensus sequence is necessary, this alone is not sufficient for *N*-glycosylation, and many potential *N*-glycosylation sites remain unoccupied. All *N*-glycans share a common core which consists of 5 monosaccharides: two *N*-acetylglucosamines (GlcNAcs) and three mannoses (Figure 1.1).

Synthesis is initiated with the assembly of an *N*-glycan precursor onto a lipid dolichol embedded in the membrane of the endoplasmic reticulum (ER). This precursor consists of an *N*-glycan core with 2 additional mannoses, facing the cytoplasm. The orientation of the phospholipid is then reversed by the enzyme flippase, so that the *N*-glycan precursor faces the lumen of the ER, followed by the addition of mannoses to a total of 9, as well as 3 glucose residues. The glycan is then covalently attached to the substrate asparagine, several monosaccharides are trimmed and the glycoprotein is transported to the Golgi system. Here, the *N*-glycan is subjected sequentially to various glycosidases and glycosyltransferases as it

moves through compartments of the Golgi. These glycosidases and glycosyltransferases are generally very specific and can remove or add only one type of monosaccharide with a specific linkage.

In the Golgi, first the glucose residues and outer mannoses are removed. Then, the sequential addition of GlcNAc, galactose and sialic acid leads to the formation of branches, called antennae; most *N*-glycans are diantennary, but tri- and tetra-antennary *N*-glycans are also frequently observed. In addition, a fucose can be added either to the protein-bound GlcNAc (core fucosylation) or in some cases to an antennary GlcNAc (antennary fucosylation). Furthermore, a GlcNAc can be added to the innermost mannose with a  $\beta$ 1-4 linkage (bisecting GlcNAc). This concise summary of *N*-glycan diversity applies to humans as well as some other mammals, whilst other vertebrates have a slightly different set of glycosylation-related enzymes.

*N*-glycans can be subdivided into three types: high mannose (also known as mannosidic), hybrid and complex. High mannose *N*-glycans consist of only the core structure with up to six additional mannoses; these glycans underwent only the first part of the *N*-glycan synthesis pathway. In contrast, complex *N*-glycans have been fully processed in the Golgi and contain at least two antennary GlcNAc residues. Hybrid *N*-glycans fall in between: they have one complex arm bearing a GlcNAc, often elongated with galactose and sialic acid, and one mannosidic arm.

### 1.1.3: *O*-glycosylation

---

Compared to *N*-glycans, *O*-glycans are less predictable both in structure as well as in their site of attachment. *O*-glycans are usually attached to the oxygen in either a serine (Ser) or a threonine (Thr), but tyrosine (Tyr) or modified hydroxyl-amino acids may also serve as attachment site. A range of different types of *O*-glycan classes exist which are distinguished on the basis of the nature of the first sugar which is attached to the protein (4). In the following, mucin-type *O*-glycosylation will be considered, which is the most common form of protein *O*-glycosylation in humans.

For mucin-type *O*-glycosylation, a GalNAc is first added to the protein in the Golgi, then various types of monosaccharides are added sequentially. The two most common core structures can be seen in Figure 1.1, but a variety of different structures have been found. *O*-glycans usually have a more linear structure than *N*-glycans, and they can vary greatly in size: many consist of only a few monosaccharides while others can consist of as many as a

hundred, often organised into tandem repeats. There is no clear consensus sequence for *O*-glycosylation, which is in part due to the fact that humans express 20 different GalNAc transferases initiating the first step of protein *O*-glycosylation. However, mucin-type *O*-glycosylation often occurs in protein regions rich in proline (preferentially at +3 or -1 relative to the *O*-glycosylation site) and alanine residues.

#### 1.1.4: Functions of glycosylation

---

Much like their structures, the functions of glycans are highly diverse as well (5). Firstly, addition of a glycan can alter the folding or basic thermodynamic properties of a protein. In some cases, a protein cannot be produced in its non-glycosylated form, because incorrect folding leads to degradation or otherwise prevents secretion. Glycosylation can also influence the interaction with other proteins, such as receptors. In some cases, interaction with another protein occurs exclusively through the glycan, as is the case for lectins, which recognize specific glycan motifs.

In addition, glycosylation plays a role in the management of protein quality. The chaperone proteins calnexin and calreticulin exist to ensure correct folding of proteins, and incorrectly labelled proteins are tagged by the addition of glucose on their *N*-glycan structures, preventing them from leaving the ER and sending them back to be re-folded (4, 6). Furthermore, asialo receptors capture extracellular non-sialylated glycoproteins and target them for degradation. It is speculated that this pathway clears proteins which have been in the circulation for some time and are thus prone to loss of sialic acids, thereby introducing a kind of expiration date on proteins (4).

Finally, some glycans appear to fulfil a purely structural role. Mucins contain extensively *O*-glycosylated regions which retain water and form a mucous layer, while a separate kind of *O*-glycans, proteoglycans, provide a firm padding in structures like cartilage (7). It should be noted that glycosylation, while it is essential for the functioning of higher organisms, is not necessary for the survival of many single-cell organisms. This is reflected by the fact that bacteria do not produce *N*-glycoproteins and that archaea, which lack a Golgi system, produce only simple precursor-like *N*-glycans. The glycosylation machinery fulfills an important task by managing inter-cell interaction and signalling, and it has evolved to become more complex in multicellular organisms. Lower eukaryotes such as yeast produce solely high mannose *N*-glycans with extensive branching of mannoses. Furthermore, pathogen

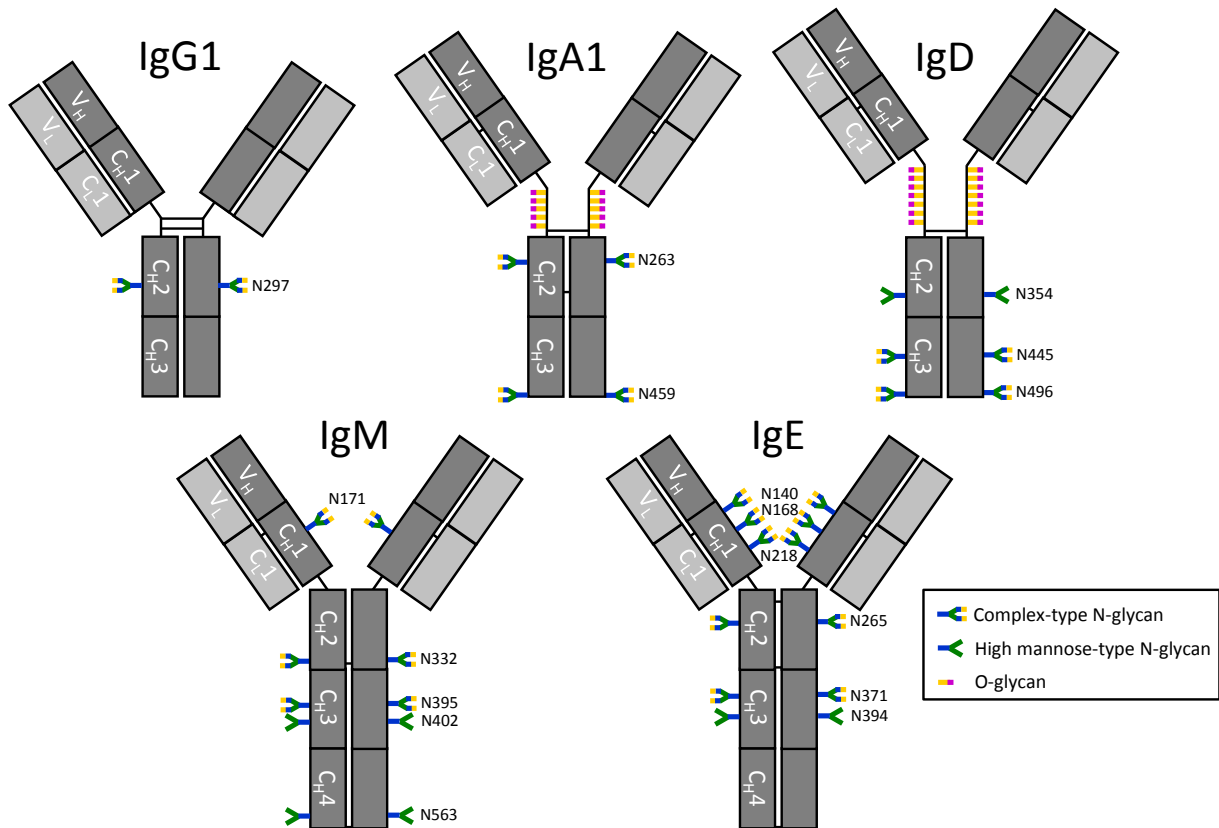


evasion is thought to have provided additional evolutionary pressure in favour of heterogeneity of glycan structures.

### 1.1.5: Glycosylation of immunoglobulins

The importance of immunoglobulins (Igs) in the immune system is well-known: they can function independently to impede pathogens, as well as mount an organized defence reaction by activating the adaptive immune system. They are produced by B-cells and are either secreted or presented on the plasma membrane. Igs are constructed of two heavy chains and two light chains, held together by disulphide bridges. Igs are usually referred to by the name of the heavy chain, of which there are five isotypes: IgG, IgA, IgM, IgD and IgE. The light chains occur in two varieties: lambda and kappa. Each heavy chain is composed of a variable domain ( $V_H$ ) and 3 or 4 constant domains ( $C_H$ ), while a light chain consists of two domains ( $V_L$  and  $C_L$ ). Igs can also be structurally subdivided by function: the antigen-binding Fab portion, which encompasses the variable and the  $C_{H1}$  domain and binds to the antigen, and the fragment crystallizable (Fc) portion, which encompasses  $C_{H2-3/4}$  and binds to Fc receptors and complement to activate the immune system. Within the five antibody isotypes there are still variations in the protein sequences, and based on this IgG and IgA are further categorized into respectively 4 and 2 subclasses. Based on genetic differences between individuals, these can be further subdivided into allotypes. IgG, IgE and IgD exist only as monomers, while IgA and IgM can also be present as secretory protein complexes in dimeric and pentameric form, respectively, bonded together by a J-chain and secretory component (8).

All human antibodies carry one or multiple glycosylation structures, many of which have been shown to influence their properties and function. A single well-characterized complex *N*-glycan is located in the Fc region of IgG, and this glycan has been shown to influence receptor binding (9). In their constant domain, IgA1 and IgA2 carry 2 and 5 *N*-glycans, respectively. IgM carries 5 *N*-glycans, while IgD carries three (9). IgE is the most heavily glycosylated with six *N*-glycans in the conserved protein sequence (10). In addition, IgA1 and IgD exhibit multiple *O*-glycosylation sites in their hinge region (9). The J-chain and secretory component found in multimeric antibody complexes are also glycosylated. Furthermore, the variable domain of both the heavy and light chain, which varies for each antigen-specific type of Ig due to somatic hypermutation, can sometimes contain glycosylation sites (11).



**Figure 1.2: The main immunoglobulin isotypes with their reported *N*- and *O*-glycosylation sites.** Heavy chain domains, conserved (C) and variable (V), are shown in dark grey, while the light chains are shown in light grey.

## **1.2: Recent advances in clinical glycoproteomics of immunoglobulins (Igs)**

---

### **1.2.1: Introduction**

---

Glycosylation of immunoglobulins (Igs) plays a key role in the regulation of immune reactions: glycans located at various sites modulate a diversity of immunoglobulin properties including protein conformation and stability, serum half-life, as well as binding affinities to antigens, receptors and glycan-binding proteins (GBP) (12-14).

The five classes of human antibodies – IgG, IgA, IgM, IgE and IgD – each contain one to six sites for *N*-linked glycosylation within the conserved sequence of each heavy chain (9). IgA1, IgD and IgG3 also carry *O*-linked glycans on their hinge-region (9, 15). In addition, immunoglobulins can be glycosylated in the variable domain of the Fab (antigen-binding fragment) (16-18). Importantly, glycosylation adds a formidable degree of complexity to protein species, since a range of glycan structures is usually present at each glycosylation site.

Studies on the functional consequences of immunoglobulin glycosylation, especially for IgG, have shown that glycans linked to the Fc (fragment crystallizable) part of the antibody influence the interaction with Fc receptors and GBPs, thereby regulating the pro- or anti-inflammatory immune response (12, 19-22). For example, lack of a fucose on the IgG Fc glycan can enact a 100-fold increase in antibody-dependent cellular cytotoxicity (ADCC) (23, 24). Fc-linked glycans may also influence the endocytosis, transcytosis and half-life of some classes of immunoglobulin, such as IgA (25, 26). Next to Fc-linked glycosylation, glycans attached to the Fab region also influence Ig properties and inflammation, especially by modulating antigen recognition and antibody aggregation, as well as through the binding to GBP (11, 17). Importantly, antibody glycosylation has been shown to reflect the physiological and pathological condition of an organism (27-29).

Because of the impact on the immunological response and thus the efficacy of therapeutic antibody treatment, it is crucial to monitor and in some cases alter the glycosylation profile in order to optimize antibody effector functions (19, 23). Glycosylation of antibodies can vary widely depending on the expression system and cell culture conditions during production (23). Because non-human glycan structures can trigger immunogenic responses, therapeutic antibodies are currently produced exclusively in mammalian cell cultures. Due to improvements in glyco-engineering, it is expected that non-mammalian expression systems will soon be applicable as well (23, 30). Robust and high-throughput methods are needed to

monitor the glycosylation of therapeutic antibodies. Additionally, glycosylation analysis should be site-specific since the function of a glycan can depend on its location, as illustrated by the different influence of glycans located at the Fc and at the Fab part of IgG (11, 22).

Glycosylation profiling of antibodies is usually done using one of the following approaches: 1) by releasing glycans from the protein, which is easily done for *N*-glycans by digestion with PNGase F, while *O*-glycans can be released chemically through hydrazinolysis or beta-elimination; 2) by using a proteolytic enzyme to digest the glycoprotein, resulting in glycopeptides; or 3) by analyzing the intact glycoprotein or portions thereof (e.g. Ig heavy and light chains) (31-33). Recent years have seen major methodological advances in all three approaches as detailed in this review. In addition, selected examples are given of antibody glycosylation studies in both biotechnological and biomedical research.

In the field of immunoglobulin (glyco)proteomics, several nomenclatures for the glycosylation sites are used (Table 1.1). The one most commonly used refers to the Asn positions as determined in the old days based on Edman sequencing of both variable and heavy chains (e.g. (9)). Alternatively, the homology-based nomenclature by the international ImMunoGeneTics information system (IMGT) is available for immunoglobulins, which has the advantage of a more intuitive comparison between the different immunoglobulins (e.g. site homology between CH2 84.4 on IgG and IgD, as well as similarity with CH3 84.4 on IgE and IgM). (34). In this review we will use the UniProt based site annotation, since this is more easily integrated with proteomic databases (35).

Analysis of the antibodies themselves is complicated by the variable domain which dictates the specificity of the antigen-binding site. Protein sequencing of monoclonal antibodies or affinity-purified antibodies is done using high resolution liquid chromatography tandem mass spectrometry methods, coupled to DNA sequence information generated by next-generation sequencing of the B-cell antibody repertoire (36, 37). Post-translational modifications, such as glycosylation, further complicate antibody analysis and require specific analysis strategies, as will be detailed in this review.

**Table 1.1: Several different immunoglobulin protein sequence nomenclatures are used in literature.** The nomenclature most frequently used in literature is based on archaic sequencing data of both immunoglobulin variable and constant domains, whereas the UniProt numbering is based on the conserved sequences, and the IMGT nomenclature is based on homology between the immunoglobulins.

<b>Ig</b>	<b>conventional literature<sup>1</sup></b>	<b>UniProt<sup>2</sup></b>	<b>IMGT<sup>3</sup></b>
IgG1	297	180	CH2-84.4
IgG2	297	176	CH2-84.4
IgG3	297	227	CH2-84.4
IgG3	392	322	CH3-79
IgG4	297	177	CH2-84.4
IgA1	263	144	CH2-20
IgA1	459	340	CHS-7
IgA2	166	47	CH1-45.2
IgA2	211	92	CH1-114
IgA2	263	131	CH2-20
IgA2	337	205	CH2-120
IgA2	459	327	CHS-7
IgM	171	46	CH1-45
IgM	332	209	CH2-120
IgM	395	272	CH3-81
IgM	402	279	CH3-84.4
IgM	563	439	CHS-7
IgE	140/145 <sup>4</sup>	21	CH1-15.2
IgE	168/173 <sup>4</sup>	49	CH1-45.2
IgE	218/219 <sup>4</sup>	99	CH1-118
IgE	265	146	CH2-38
IgE	371	252	CH3-38
IgE	394	275	CH3-84.4
IgD	354	225	CH2-84.4
IgD	445	316	CH3-45.4
IgD	496	367	CH3-116

<sup>1</sup> As used in e.g. (9)

<sup>2</sup> (35)

<sup>3</sup> (34)

<sup>4</sup> alternative nomenclature used in (38)

## 1.2.2: IgG

---

### 1.2.2.1: Fc glycosylation at Asn180/176/227/177 (‘Asn297’)

---

The majority of IgG glycosylation analysis has been focused on the Fc glycan because of both the known influence of this glycan on IgG effector functions and the established high-throughput methods which are available to selectively monitor this glycosylation site (22, 32, 33). In human IgG, the conserved *N*-glycosylation site is located at Asn180 (IgG1; UniProt P01857), Asn176 (IgG2; P01859), Asn227 (IgG3; P01860) or Asn177 (IgG4; P01861), alternatively referred to as position CH2-84.4 (34) or Asn297 (e.g. in (9); Table 1.1). In all IgG subclasses, the Fc-glycosylation site has been shown to harbor complex type diantennary *N*-glycans which carry between zero and two galactoses, with the majority carrying a core fucose, and a minority having a bisecting *N*-acetylglucosamine (GlcNAc) and one or two sialic acids (39). The glycan at this site has been shown to influence the inflammatory capacity of IgG through modulation of the binding to Fc-gamma receptors (FcγRs) and C-type lectins: in general, the absence of a core fucose and/or absence of galactoses and sialic acids appear to convey pro-inflammatory properties, while the presence of terminal sialic acids triggers an anti-inflammatory response (12, 22, 40).

Changes in Fc glycosylation, i.e. a decrease in galactosylation and sialylation which contributes to a more inflammatory antibody profile, have been observed in various autoimmune disorders, most recently inflammatory bowel disease (IBD), systemic lupus erythematosus (SLE), multiple sclerosis (MS) and chronic inflammatory demyelinating polyneuropathy (CIDP) (41-44). In addition to autoimmune disorders, IgG glycosylation changes can also occur in infectious diseases, as was shown by recent studies on HIV infection, chronic hepatitis B and the parasitic disease visceral leishmaniasis (45-47). Furthermore, new reports reaffirm the potential role for IgG glycosylation as a biomarker for cancer progression (48, 49). Finally, congenital defects in glycosylation or carbohydrate metabolism also alter IgG glycosylation, as shown recently for Man1B1 deficiency and galactosemia (50, 51).

#### 1.2.2.1.1: Analysis of released glycans

---

The gold standard for studying IgG glycosylation relies on enzymatic *N*-glycan release, subsequent fluorescent labeling by reductive amination and analysis of the labeled glycans by high-performance liquid chromatography (HPLC) using hydrophilic interaction liquid chromatography (HILIC) with fluorescence detection (52). First, this approach has been

further developed by implementing HILIC stationary phases for ultra-performance liquid chromatography (UPLC) instrumentation, thereby improving both throughput and resolution (53). Second, sample preparation has been simplified through the use of 96-well filter plates to increase the throughput of glycan purification (54), as well as by introducing fluorescent tags to label the glycosylamine species released by PNGase F, instead of targeting the aldehyde species that arise from acid-catalyzed hydrolysis of the glycosylamine (55). Of note, the increased throughput capacity allowed for analyses of large sample sets, which could for example show the potential of the IgG *N*-glycans as a marker of chronological and biological age (56). Third, sample preparation has been robotized, resulting in a highly automated, higher-throughput workflow and leading to more robust results (55). However, this method has a disadvantage: because the glycans are released from the IgG, it is impossible to distinguish between Fab and Fc glycans as well as between glycans originating from different IgG subclasses.

Next to HILIC UPLC of fluorescently labeled glycans, various methods for repetitive IgG glycosylation analysis (“profiling”) have reached maturity as evidenced by the high consistency of the results obtained in extensive method comparison studies (31-33). Remarkably, various mass spectrometric methods showed very good performance with respect to resolution, sensitivity and robustness, which opened the way to their broad application in both biotechnological (32, 33) and biomedical applications (31).

#### *1.2.2.1.2: Analysis of glycopeptides*

---

A bottom-up proteomics approach, with trypsin digestion followed by liquid chromatography (LC) coupled to mass spectrometric analysis, is most commonly applied for site-specific analysis of IgG Fc glycosylation (32). Tryptic digestion results in distinct glycopeptides that allow discrimination of the different IgG subclasses – with the peptide moieties EEQYNSTYR for IgG1, EEQFNSTFR for IgG2 and EEQFNSTYR for IgG4. The peptide sequence of the IgG3 glycopeptide shows allotype variation in the amino acid at the position *N*-terminal of the Asn227, causing a mass that is identical to either the IgG2 peptide (EEQFNSTFR; predominant allotype in Caucasian populations) or the IgG4 sequence (EEQYNSTFR; predominant allotype in Asian and African populations) (57). While trypsin digestion forms the gold standard for Fc IgG glycopeptide analysis, we have recently found that incomplete denaturation and digestion of IgG might lead to biases in glycoprofiling (58).

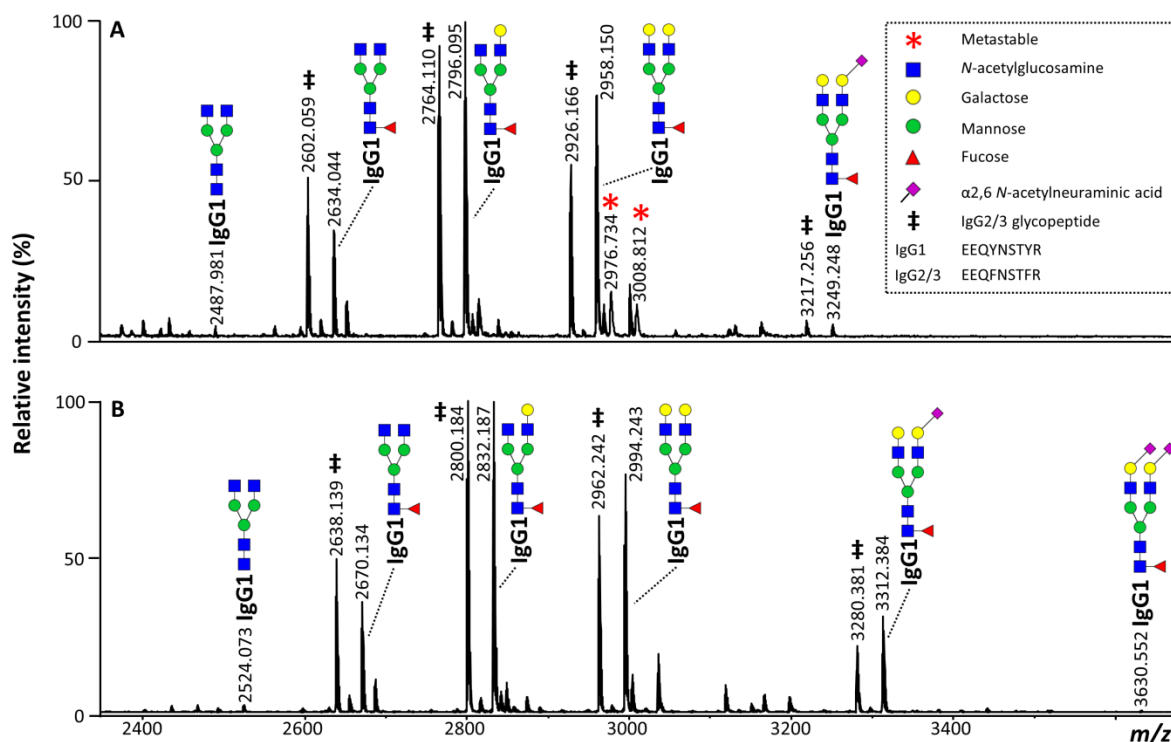
To prevent ion suppression of the glycopeptides by unglycosylated peptides during mass spectrometric analysis, a glycopeptide separation or enrichment step is often applied. This separation is usually performed using either reverse phase LC with C18 or graphitized carbon as a stationary phase, or by HILIC (13). While a variety of stationary phases exist for HILIC, amide is most frequently used, although polysaccharide-based stationary phases may show similar performance. Notably, the affinity of current HILIC materials is often dependent on glycan structure, which can lead to a bias in enrichment (13). The lack of a gold standard method has led to the development of various new HILIC materials for glycopeptide enrichment of tryptic IgG (glyco)peptides, with stationary phases consisting of various polysaccharides (chitosan, dextran, cyclodextrin, maltose) coupled to magnetic particles, silica particles or metallo-organic frameworks (59-62), or functionalized amide polymers embedded in a monolith capillary (63). Furthermore, electrostatic repulsion HILIC (also known as ERLIC) has been successfully employed to enrich IgG glycopeptides, although a thorough analysis of the potential skewing of IgG glycoforms in ERLIC enrichment is still lacking (64). Zwitterionic (ZIC) HILIC has also gained popularity: this technique makes use of highly hydrophilic materials carrying both positive and negative charges and shows a very good performance in glycopeptide enrichment (65). Novel ZIC HILIC materials, consisting of zwitterionic polymers coupled to silica particles or magnetic nanoparticles, have been developed and feature high sensitivity in post-enrichment MALDI-MS measurements (66-68).

Multiple reaction monitoring (MRM) is well-suited for high-throughput quantitative analysis of complex samples and has only recently been applied to glycopeptide Ig analysis (69-71). Hong *et al.* developed a method which simultaneously performs glycoprofiling of IgG and absolute quantitation of IgG and its separate subclasses, which can help determine if a relative change in glycosylation is due to changes in post-translational modification or changes in the level of protein production (71). For the quantification of glycopeptides, MRM was set to specifically detect oxonium ions, fragment ions that originate during the fragmentation of glycopeptides. Similar methods are being developed for IgA and IgM (69). A comparable protocol describing MRM detection of IgG glycoforms was separately developed by Yuan *et al.*, with the added feature of a prior separation of IgG3 so that glycosylation could be observed separately for each of the four IgG subclasses (72).



Capillary electrophoresis (CE) coupled to MS has likewise been applied for tryptic IgG analysis and shows a vastly increased sensitivity compared to the LC-MS approach (73). The sensitivity gain may be largely ascribed to the very low flow rates achieved in CE-MS.

Compared to LC-ESI-MS or CE-MS, MALDI-MS offers higher sample throughput as well as lower data complexity. However, detection of glycopeptides is complicated by the loss of sialic acids through in-source decay. Of note, this can be prevented by neutralizing the charge on the sialic acid by (dimethyl-)amidation (39, 74) or ethyl esterification (75). The derivatization methods target carboxyl groups on the peptide moiety as well as on the glycan, which can provide useful structural information from a combination of positive- and negative-ion MS/MS analyses (74). An additional advantage of these methods is the introduced mass difference between  $\alpha$ 2,3- and  $\alpha$ 2,6-linked sialic acids, caused by the sialic acid linkage-specificity of the reactions. While the ethyl esterification method is highly specific for sialic acid linkages on the level of released glycans (76), the modification of the peptide moiety of glycopeptides was found to be not completely specific, resulting in unwanted byproducts. This issue has been addressed and overcome by a recently published method using dimethylamidation, which provides sialic acid linkage information on a stably modified glycopeptide (Figure 1.3) (39). Currently, the dimethylamidation of sialylated glycopeptides is optimized for IgG Fc-glycopeptides and would need further optimization when used for different glycoproteins, such as other immunoglobulins. MALDI-TOF-MS/MS of sialylated glycopeptides, using laser induced fragmentation, highly benefits from the derivatization, as the loss of the sialic acid is no longer the dominant fragment. In addition to analysis with MALDI-TOF-MS(/MS), the derivatization method has been shown to be applicable for the analysis of IgG Fc-glycopeptides, using LC-MS(/MS), enabling differentiation between differently linked sialic acids, without having a major influence on the fragmentation of the analytes (data not published). Pyrene derivatization is an alternative method for glycopeptide analysis by MALDI-MS and also allows discrimination between  $\alpha$ 2,3- and  $\alpha$ 2,6-linked sialic acids, although its application on immunoglobulins has not been established (77, 78).



**Figure 1.3: MALDI-TOF-MS spectra of human plasma IgG Fc-glycopeptides (A) without derivatization, and (B) after dimethylamidation.** Derivatization results in the stabilization of the sialylated glycopeptides, improving their detection and preventing the formation of metastable signals. With the use of dimethylamidation, no unspecific side reactivity on the peptides was observed and the linkage of the sialic acids could be determined (39).

Furthermore, stable isotope labeling of glycopeptides has recently been achieved: succinic anhydride was used to introduce a mass increment of 8 Da to tryptic IgG1 glycopeptides (79). This method can be used to perform absolute quantification of IgG glycopeptides, or can be used to reduce bias during sample analysis by parallel analysis of two samples. Isobaric labeling using tandem mass tags (TMTs), which lead to different masses upon fragmentation, has also been used for the analysis of pepsin-generated IgG glycopeptides (80).

#### *1.2.2.1.3: Analysis of intact glycoprotein or glycoprotein fragments*

Alternatively, a middle-down or middle-up proteomics approach can be applied for assessing IgG Fc glycosylation. The Fc portion can be cleaved from the Fab portion in whole IgG, or from fused proteins in Fc-fusion proteins, by limited proteolytic digestion with the protease IdeS (FabRICATOR®), followed by either mass spectrometric analysis of the protein fragment or purification of the Fc portion and release of the glycans using PNGase F (16, 81-83). The latter method has recently been applied to a clinical sample set, as is discussed later in this review (16).

Finally, technical advances in the recent years have allowed for top-down mass spectrometric analysis of monoclonal antibodies, allowing the integrated analysis of post-translational modifications (32, 83-86). An extensive review has been recently published by Zhang *et al.*, describing several applications of these techniques, and a comparison between ‘normal’ and native MS, furthermore including ion mobility (87). More recently, ultra-high resolution machinery (e.g. Fourier transform ion cyclotron resonance (FTICR)) allowed for the detection of intact monoclonal antibodies with isotopic resolution, showing several glycoforms (85). Additional top-down MS/MS information was obtained in conjunction with online electrochemical reduction of the antibody (85). Furthermore, isobaric labeling has been applied to intact antibody-drug conjugates (80). Native MS is often applied to analyze intact mAbs (86). The advantage of this approach is the limitation of charge states because of the native 3D protein configuration, causing increased signal-to-noise for the few charge states that do occur. The downside of native MS is a lack of information regarding the glycosylation site(s) or the precise structure of the glycan(s) (86, 87).

#### 1.2.2.2: Fab glycosylation

---

The structural features of IgG Fab glycosylation and its emerging importance in immunity have been recently reviewed (11). It has been estimated that approximately 15-25% of serum IgG of healthy individuals contain *N*-glycosylation sites and carry *N*-glycans (Fab-glycosylation) in their variable domains, in addition to the almost fully occupied IgG Fc *N*-glycosylation site (88, 89). Of note, the percentage of Fab-glycosylation and glycan structures varies during certain pathological and physiological conditions, as shown in RA, lymphoma and pregnancy (16, 17, 90, 91). Since only a few germline-encoded sequences contain an *N*-glycosylation site, the sites present in the variable domains of immunoglobulins are mainly introduced by somatic hypermutation during the process of affinity maturation (18, 92). Within an affinity-purified population of antigen-specific IgG, identification of Fab glycosylation sites has been achieved by labeling the sites with <sup>18</sup>O during deglycosylation with PNGase F. This was followed by mass spectrometry-assisted proteomics analysis which revealed the mass shift denoting the site of glycosylation and the peptide sequence surrounding the site (17). The human immune system comprises of an enormous antibody repertoire, recognizing an estimated billion or more different antigens. Antibody specificity is determined by a unique amino acid sequence in the Fab portion, thus making the analysis of Fab *N*-glycopeptides derived from polyclonal antibodies very difficult, if feasible at all.

Therefore, in order to analyze polyclonal IgG Fab glycosylation, the currently used analytical methods consist of the release of *N*-glycans from purified Fab fragments followed by their analysis using capillary electrophoresis with laser-induced fluorescence detection (CE-LIF), (ultra)high performance liquid chromatography and/or mass spectrometry (16, 17, 93). In order to improve the throughput of IgG Fab glycosylation analysis, we recently set up a new sample preparation method. The method relies on IgG affinity capturing in a 96-well filter plate, on-bead proteolytic release of the Fab portions, and collection of Fab (flow-through) and Fc portions (eluate) followed by enzymatic glycan release. Detailed glycan information was obtained by MALDI-TOF-MS after sialic acid stabilization. The method was applied to study the differences between Fab and Fc glycosylation in young women, and the pregnancy associated changes thereof (16). The levels of galactosylation, sialylation and bisection were significantly higher on the Fab portion compared to the Fc. During pregnancy Fab and Fc glycans showed similar patterns in their changes. Interestingly, the Fab portion was also shown to carry minor amounts of  $\alpha$ 2,3-linked sialic acids. In general, the *N*-glycans on the Fab are more extended, and some species seem to be Fab-specific. Diantennary fucosylated glycans with two sialic acids are hardly present on the Fc, while they are the major species on Fab. Similarly, the presence of a bisecting GlcNAc on glycans with two galactoses appears to be more prominent in Fab versus Fc (16). Of note, these data result from the analysis of solely young women, while characterization of Fab glycosylation in males as well as different age groups is still lacking.

High-sensitivity analysis of released Fab glycans can also be performed by CGE-LIF (93). However, the glycosylation data obtained via these techniques, especially regarding both the levels of *N*-glycan bisection in IgG Fab portions and various glycosylation features of murine IgG, has shown some discrepancies as compared to results obtained with other analytical methods (16, 94, 95). Additional studies are needed to further unravel Fab glycosylation changes with age, sex and diseases.

#### 1.2.2.3: Additional *N*- and *O*-glycosylation of IgG3

In addition to the well-known Fc *N*-glycosylation site, several allotypes of IgG3 possess a second *N*-linked site in the CH3 domain at Asn322 (UniProt P01860; alternatively referred to as CH3-79 (34) or Asn392; Table 1.1) (96). Only 10% of Asn322 was found to be occupied; the *N*-glycans found at this site were mainly complex type diantennary structures, which differ from those at Asn227 in that the majority is afucosylated and contains a bisecting

GlcNAc, and a minority of high mannose type *N*-glycans is also present (96). Since trypsin digestion produces a very large glycopeptide containing this site, the glycan structures may alternatively be examined after digestion with aspecific proteases. Aspecific proteases such as pronase are particularly useful for the study of glycoproteins since they tend to produce small glycopeptides which are well suited for mass spectrometric analysis (97, 98). Sequential chromatography of resulting digests on a C18 reversed phase (RP) column and a porous graphitized carbon column provides the broad retention range necessary to observe all glycopeptides regardless of the heterogeneous retention properties of both the glycan and the peptide moiety. Using collision-induced dissociation with a combination of lower- and enhanced-energy, which produced both glycan and peptide fragmentation, respectively, identification of both the glycan and the peptide moiety in one run could be achieved (96).

Next to *N*-glycosylation, IgG3 may carry up to 3 *O*-glycosylation sites per heavy chain within a triple repeat in the hinge region (15). Proteolytic digestion with trypsin, proteinase K or pronase followed by LC-MS/MS analysis revealed that approximately 10% of each of these sites is occupied, mainly by sialylated core 1 type *O*-glycan structures (15, 96). As both the Fc *N*-glycosylation at Asn322 and the *O*-glycans of IgG3 have been described only recently, their function and clinical relevance remain to be investigated.

#### 1.2.2.4: Antigen-specific IgG

---

The glycoprofiling of antigen-specific antibodies in clinical samples after vaccination or during disease started less than a decade ago. This was made possible by the numerous improvements in sensitivity and throughput of methods for both antibody purification and glycosylation analysis. Antigen-specific antibodies are generally purified by affinity chromatography using antigens coated on 96-well plates or on chromatography beads/columns. Antigens are usually synthetic peptides or recombinant (glyco)proteins. For instance, the high-throughput purification of anti-citrullinated peptide/protein antibodies (ACPA), i.e. autoantibodies specific for rheumatoid arthritis (RA), has been achieved by repeated capturing on 96-well plates coated with a synthetic circular peptide containing citrulline, called CCP2 (cyclic citrullinated peptide 2) (28, 99, 100). Likewise, antibodies directed against multiple HIV and influenza antigens (e.g. HIV gp41 and gp120 or influenza hemagglutinin) have been enriched using amino-link antigen resin or antigen-functionalized streptavidin resins packed into cartridges (46, 101, 102). Alternatively, antigen-specific antibodies can be captured by using viral particles, microorganisms or cells. Thus, Vidarsson

and coworkers have isolated anti-platelet antibodies (causing neonatal alloimmune thrombocytopenia) and anti-red blood cell antibodies (responsible for haemolytic disease of the fetus and newborn) by incubating serum of pregnant women directly on platelets and red blood cells (103-105). Fc- and/or Fab-linked glycosylation of antigen-specific IgG has been analyzed either at the glycopeptide level mainly using LC-MS, or by releasing glycans using a middle-down/middle-up proteomics approach as described above. Of note, unlike glycopeptide detection, analysis of released glycans from antigen-specific IgG requires another purification step prior to or after antigen-specific capturing in order to separate IgG from other serum glycoproteins or other immunoglobulins (101).

Antigen-specific IgG displays different sialylation, galactosylation, fucosylation and/or bisection patterns compared to total IgG isolated from the same individuals. Importantly, these structural differences are clearly associated with clinical and functional consequences including disease outcome, disease severity and/or antiviral control responses. For instance, as compared to total IgG, anti-platelet IgGs found in the serum of pregnant women exhibit an exceptionally low level of fucosylation in their Fc-glycans, which enhances the binding affinity for the Fc $\gamma$ RIIIa/b and the phagocytosis of platelets, and correlates with increased severity of neonatal alloimmune thrombocytopenia (105). Likewise, HIV-specific IgG antibodies isolated from HIV-positive subjects present a higher frequency of afucosylated, agalactosylated, and asialylated *N*-glycans compared to total IgG. Importantly, this glycan difference, especially the greater percentage of agalactosylated glycoforms, is far more pronounced in HIV elite controllers than in (un)treated chronic progressors and is associated with an enhanced capacity to bind to Fc $\gamma$ RIIIa, probably explaining the more potent antibody-dependent cellular viral inhibition activity that characterizes antibody from elite controllers (46). A disruption in the balance between type I (part of the Ig receptor superfamily which includes Fc $\gamma$ RI, II and III) and type II (C-type lectin receptors) Fc receptor signaling also very likely occurs in several autoimmune diseases such as rheumatoid arthritis and granulomatosis with polyangiitis (GPA), in which changes in autoantibody-specific glycosylation have been observed. Thus, the Fc-galactosylation, sialylation, and bisection of anti-proteinase 3 (PR3) antibodies IgG1 are reduced compared to total IgG1 in GPA patients (106). Despite an early study reporting a negative correlation between the level of anti-PR3 specific IgG sialylation and disease activity as measured by the Birmingham Vasculitis Activity Score (BVAS), recent evidence demonstrated that the BVAS is strongly associated with the presence of bisecting GlcNAc on anti-PR3 IgG but not with

galactosylation/sialylation percentages (106, 107). Interestingly, the level of anti-PR3 IgG galactosylation was associated with pro-inflammatory cytokine concentrations and time to remission (106). Similarly, in RA patients, ACPA-IgG autoantibodies exhibit a decrease in Fc galactosylation and sialylation levels that occurs a few months before disease presentation, correlates with disease severity, and potentially determines osteoclast differentiation and bone loss during RA (28, 99, 108). Variations in antigen-specific IgG glycosylation have also been observed following vaccination and, more importantly, can predict the efficacy of vaccination (102, 109). Ravetch and coworkers recently showed that the sialylated Fc glycan abundance on anti-hemagglutinin IgG produced by day 7 following influenza virus vaccination predicts the quality of the vaccine response (102). It was proposed that immune complexes formed with Fc-sialylated IgG signal through the type II FcR CD23 on activated B cells and triggers the expression of Fc $\gamma$ RIIb, thereby driving the selection of higher affinity B cells and the generation of higher affinity and more protective anti-HA IgG (102).

### 1.2.3: IgA

---

There are two subclasses of immunoglobulin A (i.e. IgA1 and IgA2), and two known allotypes for the IgA2 subclass (i.e. A2m(1) and A2m(2)). IgA1 contains a slightly elongated hinge region compared to IgA2. This elongated hinge contains 9 potential *O*-glycosylation sites, of which up to six have been reported to be occupied (110). In addition, IgA1 harbors two *N*-glycosylation sites at Asn144 and Asn340 (UniProt P01876; alternatively referred to as CH2-20 and CHS-7, respectively (34), or Asn263 and Asn459 (e.g. in (9)); Table 1.1), whereas IgA2 harbors four sites at Asn47, Asn131, Asn205, and Asn327 (UniProt P01877; also referred to as CH1-45.2, CH2-20, CH2-120, CHS-7 (34), or Asn166, Asn263, Asn337, and Asn459 (e.g. in (9)); Table 1.1). In the A2m(2) allotype of IgA2 an additional consensus sequence is present due to the replacement of a proline by a serine, thus forming a glycosylation site at Asn92 (CH1-114 / Asn211; Table 1.1). The analysis of (s)IgA glycosylation is generally performed at the level of released glycans (111) or by lectin ELISA, although a few glycopeptide-based LC-MS/MS methods have been described (110, 112-115).

For the analysis of tryptic *O*-glycopeptides, the use of FTICR-MS/MS coupled online to an RP-LC system has been described (110). Electron transfer dissociation, which preferentially fragments the peptide backbone and not the glycan, was used to determine the location of

each glycosylation site. By applying a few additional separation steps using less advanced laboratory techniques, others have achieved similar results by MALDI-TOF/TOF-MS (116).

The IgA *N*-glycosylation is less frequently studied, although it may have important functional consequences as demonstrated by the influence of *N*-glycan sialylation on the transportation of secretory IgA across an *in vitro* model of follicle-associated epithelium via binding to Dectin-1 and Siglec-5 (26).

Nowadays, the use of IgA instead of IgG monoclonal antibodies for biopharmaceutical purposes is being explored, with a focus on anti-HIV drugs (117-119). Therefore, several site-specific glycosylation analysis methods have recently been developed. An LC-ESI-MS/MS method primarily developed for the analysis of HIV gp140 has been adapted and applied to secretory IgA1 produced in plants as well as to human IgA (119, 120). In brief, IgA was digested by sequential application of trypsin and GluC after reduction and alkylation. Next, glycopeptide analysis was performed by first identifying the elution position of deglycosylated peptide moieties. Glycopeptides are known to elute a short time ahead of the deglycosylated variant, with some spread due to the various glycans attached. The addition of a buffered formic acid solution to the flow ascertained very close or even identical elution times for glycosylated peptides bearing sialylated structures. A targeted search for the peptide plus potential glycan *m/z* using selected ion chromatograms completed the analysis. Several high mannose type structures were identified using the applied technique. The analysis did not reveal any hinge region *O*-glycosylation, which could be attributed to the production in plants.

The *N*-glycans of IgA are nevertheless still mainly studied at the level of released glycans. For example, the comparison of released glycans from different IgA constructs obtained from various cell lines showed profound differences, especially regarding the level of sialylation, which correlated with the half-life of these antibodies (121).

Of note, the secreted form of IgA consists of a dimer, which forms a complex with the (also glycosylated) joining (J)-chain and the secretory component. The J-chain harbors a single glycosylation site at Asn71 (UniProt P01591; also referred to as Asn48), which appears to be important for IgA dimerization (122). This site bears mainly highly sialylated diantennary *N*-glycans (111, 114). The secretory component is also highly glycosylated, with seven *N*-glycosylation sites at Asn83, Asn90, Asn135, Asn186, Asn421, Asn469 and Asn499 (UniProt P01833; also referred to as Asn65, Asn72, Asn117, Asn168, Asn403, Asn451, and



Asn481, e.g. in (123); Table 1.1). The protein contains a wide variety of glycan species: di-, tri- and tetraantennary glycans, bearing all Lewis epitopes (111, 114). It was suggested that these glycans are meant to bind to lectins of bacteria (111).

#### 1.2.4: IgM

---

Human serum IgM mainly circulates as a pentamer of 950 kDa consisting of ten light chains, ten heavy chains and one joining chain (J-chain). Each IgM monomer contains five conserved *N*-glycosylation sites at Asn46, Asn209, Asn272, Asn279 and Asn439 (UniProt P01871; also known as CH1-45, CH2-120, CH3-81, CH3-84.4, CHS-7 (34), or Asn171, Asn332, Asn395, Asn402, and Asn563; Table 1.1) located within the constant region of the heavy chain. In addition, the previously mentioned J-chain contains one *N*-glycosylation site at Asn71 (UniProt P01591). In two recent studies, a site-specific *N*-glycosylation mapping of human serum IgM was achieved by analyzing IgM glycopeptides, generated by trypsin or trypsin/GluC digestion, using either the classical LC-ESI-MS method or a nano-LC-microarray-MALDI-MS platform (124, 125). The latter consists of a nano-LC reverse phase separation of IgM (glyco)peptides, including the J-chain glycopeptide, followed by high frequency droplet-based fractionation of the nano-LC outflow on microarray chips. Each spot on the microarray is then analyzed by MALDI-MS, with or without pre-digestion with PNGase F to remove *N*-glycans. Both studies demonstrated that glycans linked to Asn279 and Asn439 are predominantly oligomannose structures, whereas glycans attached to Asn46, Asn209 and Asn272 mainly consist of complex-type structures (124, 125). The glycosylation site Asn71 of the J-chain also exhibits complex-type *N*-glycans. The main complex-type *N*-glycans found in IgM heavy chains are diantennary species carrying one or two sialic acids, bisecting GlcNAc and/or a core fucose. Minor proportions of oligomannosidic and hybrid-type glycans were also detected on Asn46 (124). Likewise, Asn279 carries 10% of hybrid-type structures, which are also present in very low amount on Asn209. Based on computer modeling of the IgM structure, the clear distinction between glycosylation sites carrying oligomannose structures (on Asn279 and Asn439) or complex-type *N*-glycans (on Asn46, Asn209, Asn272) has been proposed to be the consequence of the low accessibility of glycans on Asn279 and Asn439 for the glycosyltransferase/glycosidases within the Golgi (125). Finally, although the functional aspect of IgM glycosylation on immunity has not been examined yet, the recent possibility of producing human-like glycoengineered heteromultimeric IgM in plants may help to provide new insights in this field (125).

### 1.2.5: IgE

---

With six oligosaccharides on each heavy chain at Asn21, Asn49, Asn99, Asn146, Asn252 and Asn275 (10) (UniProt P01854; also referred to as CH1-15.2, CH1-45.2, CH1-118, CH2-38, CH3-38 and CH3-84.4 (34), or Asn140, Asn168, Asn218, Asn265, Asn371 and Asn394 e.g. in (9); Table 1.1), IgE is the most heavily glycosylated of the immunoglobulins. Characterization of the glycan structures on polyclonal IgE was achieved with a combination of proteolytic enzymes and LC-MS/MS analysis (10). Glycosylation sites Asn21, Asn49, Asn99, Asn146 and Asn252 are occupied by complex type *N*-glycans, which are primarily fully galactosylated diantennary structures, containing a core fucose and one or two sialic acids (10, 126). A high mannose type glycan is present at Asn275, the sixth site, which is homologous to the Fc glycosylation site in IgG. Glycosylation at this site has recently been shown to be essential for the binding of IgE to the high affinity receptor FcεRI and initiation of anaphylaxis (127). Individuals with PGM3-related hyper IgE syndrome or with a hyperimmune condition displayed similar IgE glycosylation compared to healthy individuals (10, 126). Glycosylation analysis of IgE is challenging due to the low concentration in biological fluids: at approximately 130-300 ng/mL, the concentration in human serum is roughly 50 000 times lower than that of IgG (128, 129). Due to this limitation, no large-scale glycosylation analysis of IgE in clinical cohorts has been performed as of yet. However, recent advances in LC- and CE-MS sensitivity and robustness may allow for some attempts in the near future.

### 1.2.6: Conclusions and perspectives

---

Thanks to the improvement of sample preparation methods and analytical technologies, recent years have seen an increase in sensitivity, accuracy and robustness of IgG glycosylation analysis. These methodological and technological advances are beneficial for biopharmaceutical companies, allowing a better characterization of antibody-based biopharmaceuticals, biosimilars and bio-betters, but are also crucial tools in both basic and clinical research. Thus, this enables, among others, the characterization of glycosylation of antigen-specific IgG, including autoantibodies, alloantibodies and some anti-pathogen antibodies, which directly impact the immune response and the outcome, progression and/or severity of diseases (28, 46, 99, 101-105, 109). Therefore, methodologies and technologies dedicated to IgG glycosylation analysis have great prospects regarding the early detection and diagnostic of some diseases. Of note, most studies on IgG glycosylation have focused on serum/plasma antibodies, while IgG in other biofluids and tissue remain largely unstudied.

In addition to IgG, the substantial recent advances in purifying and analyzing small amounts of samples have helped to analyze the glycosylation of other immunoglobulin subclasses (i.e. IgA, IgM and IgE) in a more precise and comprehensive manner. Today's technological level allows for the simultaneous analysis of multiple immunoglobulin classes in one run (70).

We expect that in the near future, several hiatuses in immunoglobulin-related glycomics will be covered. Not only by thorough analysis of the glycosylation of all immunoglobulin classes, but additionally by complementary glycoproteomics analysis of many interacting proteins, such as cell surface derived Fc receptors. This may reveal a regulatory role of both antibody and receptor glycomic variation and the interaction thereof in the regulation of antibody effector functions (24).

### 1.3: Scope of thesis

---

The aim of the studies presented in this thesis is to contribute to the knowledge within the field of immunoglobulin glycosylation. The glycosylation of IgG, specifically the *N*-glycan located in the Fc portion, has been extensively studied, but its function as well as its regulation still in part eludes us. Glycosylation of the other antibody isotypes has attracted much less attention from researchers. This can be attributed both to the lower abundance of the other Ig isotypes, which can make analysis challenging, and to the fact that the pharmaceutical industry focusses on IgG for monoclonal antibody therapeutics.

IgE is the antibody which is responsible for allergic reactions: it can trigger the strongest reaction of all of the immunoglobulins, and conversely is found at the lowest concentration in blood. IgE is also the most abundantly glycosylated, with roughly 12% of its mass consisting of saccharides (38). In **chapter 2**, we provide a detailed profile of the *N*-glycan structures present at each of the six *N*-glycosylation sites, in a polyclonal sample from pooled plasma of healthy donors. A seventh site was found to be unoccupied. Since the publication of this paper, compelling evidence has arisen that the *N*-glycan at Asn394 (Asn275 using the Uniprot nomenclature) can influence receptor binding, analogous to the IgG Fc glycan (127).

While research in the past few decades has largely mapped the glycosylation sites on antibodies and is now moving on to functional and clinical studies, structural analysis can still turn up new findings. **Chapter 3** describes a hitherto unreported *O*-glycan found in the hinge region of IgG3. Because only 10% of each of the two to three *O*-glycosylation sites on IgG3 are occupied, this glycan has evaded discovery until recently.

**Chapter 4** describes the application of the same analysis method to a cohort of patients with ANCA-associated vasculitis, an autoimmune disease with an established pathogenic role of autoantibodies (130). Various glycosylation features were found to be associated with a relapse of disease symptoms. This study reinforces the potential of IgG glycosylation as a biomarker to reflect disease severity.

In **chapter 5**, the glycosylation of Fc IgG is examined in a large population cohort of healthy individuals using liquid chromatography mass spectrometry (LC-MS). A clear association was observed between specific glycosylation features and the inflammation-associated proteins C-reactive protein and interleukin-6.

Finally, concluding notes and points of further discussion are presented in **chapter 6**.

## References

1. Apweiler, R., Hermjakob, H., and Sharon, N. (1999) On the frequency of protein glycosylation, as deduced from analysis of the SWISS-PROT database. *Biochim Biophys Acta* 1473, 4-8
2. Chandran, P. L., and Horkay, F. (2012) Aggrecan, an unusual polyelectrolyte: review of solution behavior and physiological implications. *Acta Biomater* 8, 3-12
3. Jones, M. B., Oswald, D. M., Joshi, S., Whiteheart, S. W., Orlando, R., and Cobb, B. A. (2016) B-cell-independent sialylation of IgG. *Proc Natl Acad Sci U S A* 113, 7207-7212
4. Varki, A., Cummings, R. D., Esko, J. D., Stanley, P., Hart, G., Aebi, M., Darvill, A., Kinoshita, T., Packer, N. H., Prestegard, J. J., Schnaar, R. L., and Seeberger, P. H. (2015) *Essentials of Glycobiology*, 3rd Edition Ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor (NY)
5. Varki, A. (1993) Biological roles of oligosaccharides: all of the theories are correct. *Glycobiology* 3, 97-130
6. Roth, J., and Zuber, C. (2016) Quality control of glycoprotein folding and ERAD: the role of N-glycan handling, EDEM1 and OS-9. *Histochem Cell Biol*, R110.003251
7. Vertel, B. M. (1995) The ins and outs of aggrecan. *Trends Cell Biol* 5, 458-464
8. Schroeder, H. W., Jr., and Cavacini, L. (2010) Structure and function of immunoglobulins. *J Allergy Clin Immunol* 125, S41-52
9. Arnold, J. N., Wormald, M. R., Sim, R. B., Rudd, P. M., and Dwek, R. A. (2007) The impact of glycosylation on the biological function and structure of human immunoglobulins. *Annu Rev Immunol* 25, 21-50
10. Plomp, R., Hensbergen, P. J., Rombouts, Y., Zauner, G., Dragan, I., Koeleman, C. A., Deelder, A. M., and Wuhrer, M. (2014) Site-specific N-glycosylation analysis of human immunoglobulin e. *J Proteome Res* 13, 536-546
11. van de Bovenkamp, F. S., Hafkenscheid, L., Rispens, T., and Rombouts, Y. (2016) The Emerging Importance of IgG Fab Glycosylation in Immunity. *J Immunol* 196, 1435-1441
12. Karsten, C. M., Pandey, M. K., Figge, J., Kilchenstein, R., Taylor, P. R., Rosas, M., McDonald, J. U., Orr, S. J., Berger, M., Petzold, D., Blanchard, V., Winkler, A., Hess, C., Reid, D. M., Majoul, I. V., Strait, R. T., Harris, N. L., Kohl, G., Wex, E., Ludwig, R., Zillikens, D., Nimmerjahn, F., Finkelman, F. D., Brown, G. D., Ehlers, M., and Kohl, J. (2012) Anti-inflammatory activity of IgG1 mediated by Fc galactosylation and association of FcγRIIB and dectin-1. *Nat Med* 18, 1401-1406
13. Zauner, G., Selman, M. H., Bondt, A., Rombouts, Y., Blank, D., Deelder, A. M., and Wuhrer, M. (2013) Glycoproteomic analysis of antibodies. *Mol Cell Proteomics* 12, 856-865
14. Schwab, I., Lux, A., and Nimmerjahn, F. (2015) Pathways Responsible for Human Autoantibody and Therapeutic Intravenous IgG Activity in Humanized Mice. *Cell Rep* 13, 610-620
15. Plomp, R., Dekkers, G., Rombouts, Y., Visser, R., Koeleman, C. A., Kammeijer, G. S., Jansen, B. C., Rispens, T., Hensbergen, P. J., Vidarsson, G., and Wuhrer, M. (2015) Hinge-Region O-Glycosylation of Human Immunoglobulin G3 (IgG3). *Mol Cell Proteomics* 14, 1373-1384
16. Bondt, A., Rombouts, Y., Selman, M. H., Hensbergen, P. J., Reiding, K. R., Hazes, J. M., Dolhain, R. J., and Wuhrer, M. (2014) Immunoglobulin G (IgG) Fab glycosylation analysis using a new mass spectrometric high-throughput profiling method reveals pregnancy-associated changes. *Mol Cell Proteomics* 13, 3029-3039
17. Rombouts, Y., Willemze, A., van Beers, J. J., Shi, J., Kerkman, P. F., van Toorn, L., Janssen, G. M., Zaldumbide, A., Hoeben, R. C., Pruijn, G. J., Deelder, A. M., Wolbink, G., Rispens, T., van Veelen, P. A., Huizinga, T. W., Wuhrer, M., Trouw, L. A., Scherer, H. U., and Toes, R. E. (2015) Extensive glycosylation of ACPA-IgG variable domains modulates binding to citrullinated antigens in rheumatoid arthritis. *Ann Rheum Dis* 75, 578-585
18. Dunn-Walters, D., Boursier, L., and Spencer, J. (2000) Effect of somatic hypermutation on potential N-glycosylation sites in human immunoglobulin heavy chain variable regions. *Mol Immunol* 37, 107-113

19. Thomann, M., Schlothauer, T., Dashivets, T., Malik, S., Avenal, C., Bulau, P., Ruger, P., and Reusch, D. (2015) In vitro glycoengineering of IgG1 and its effect on Fc receptor binding and ADCC activity. *PLoS One* 10, e0134949
20. Anthony, R. M., Nimmerjahn, F., Ashline, D. J., Reinhold, V. N., Paulson, J. C., and Ravetch, J. V. (2008) Recapitulation of IVIG anti-inflammatory activity with a recombinant IgG Fc. *Science* 320, 373-376
21. DiLillo, D. J., and Ravetch, J. V. (2015) Fc-Receptor Interactions Regulate Both Cytotoxic and Immunomodulatory Therapeutic Antibody Effector Functions. *Cancer Immunol Res* 3, 704-713
22. Anthony, R. M., and Nimmerjahn, F. (2011) The role of differential IgG glycosylation in the interaction of antibodies with FcγR3 in vivo. *Curr Opin Organ Transplant* 16, 7-14
23. Costa, A. R., Rodrigues, M. E., Henriques, M., Oliveira, R., and Azeredo, J. (2014) Glycosylation: impact, control and improvement during therapeutic protein production. *Crit Rev Biotechnol* 34, 281-299
24. Ferrara, C., Grau, S., Jager, C., Sondermann, P., Brunker, P., Waldhauer, I., Hennig, M., Ruf, A., Rufer, A. C., Stihle, M., Umana, P., and Benz, J. (2011) Unique carbohydrate-carbohydrate interactions are required for high affinity binding between FcγR3 and antibodies lacking core fucose. *Proc Natl Acad Sci U S A* 108, 12669-12674
25. Basset, C., Durand, V., Jamin, C., Clement, J., Pennec, Y., Youinou, P., Dueymes, M., and Roitt, I. M. (2000) Increased N-linked glycosylation leading to oversialylation of monomeric immunoglobulin A1 from patients with Sjogren's syndrome. *Scand J Immunol* 51, 300-306
26. Rochereau, N., Drocourt, D., Perouzel, E., Pavot, V., Redelinguys, P., Brown, G. D., Tiraby, G., Roblin, X., Verrier, B., Genin, C., Corthesy, B., and Paul, S. (2013) Dectin-1 is essential for reverse transcytosis of glycosylated SIgA-antigen complexes by intestinal M cells. *PLoS Biol* 11, e1001658
27. Maverakis, E., Kim, K., Shimoda, M., Gershwin, M. E., Patel, F., Wilken, R., Raychaudhuri, S., Ruhaak, L. R., and Lebrilla, C. B. (2015) Glycans in the immune system and The Altered Glycan Theory of Autoimmunity: a critical review. *J Autoimmun* 57, 1-13
28. Rombouts, Y., Ewing, E., van de Stadt, L. A., Selman, M. H., Trouw, L. A., Deelder, A. M., Huizinga, T. W., Wuhrer, M., van Schaardenburg, D., Toes, R. E., and Scherer, H. U. (2015) Anti-citrullinated protein antibodies acquire a pro-inflammatory Fc glycosylation phenotype prior to the onset of rheumatoid arthritis. *Ann Rheum Dis* 74, 234-241
29. Bondt, A., Selman, M. H., Deelder, A. M., Hazes, J. M., Willemsen, S. P., Wuhrer, M., and Dolhain, R. J. (2013) Association between galactosylation of immunoglobulin G and improvement of rheumatoid arthritis during pregnancy is independent of sialylation. *J Proteome Res* 12, 4522-4531
30. Frenzel, A., Hust, M., and Schirrmann, T. (2013) Expression of recombinant antibodies. *Front Immunol* 4, 217
31. Huffman, J. E., Pucic-Bakovic, M., Klaric, L., Hennig, R., Selman, M. H., Vuckovic, F., Novokmet, M., Kristic, J., Borowiak, M., Muth, T., Polasek, O., Razdorov, G., Gornik, O., Plomp, R., Theodoratou, E., Wright, A. F., Rudan, I., Hayward, C., Campbell, H., Deelder, A. M., Reichl, U., Aulchenko, Y. S., Rapp, E., Wuhrer, M., and Lauc, G. (2014) Comparative performance of four methods for high-throughput glycosylation analysis of immunoglobulin G in genetic and epidemiological research. *Mol Cell Proteomics* 13, 1598-1610
32. Reusch, D., Habberger, M., Falck, D., Peter, B., Maier, B., Gassner, J., Hook, M., Wagner, K., Bonnington, L., Bulau, P., and Wuhrer, M. (2015) Comparison of methods for the analysis of therapeutic immunoglobulin G Fc-glycosylation profiles-Part 2: Mass spectrometric methods. *MAbs* 7, 732-742
33. Reusch, D., Habberger, M., Maier, B., Maier, M., Kloseck, R., Zimmermann, B., Hook, M., Szabo, Z., Tep, S., Wegstein, J., Alt, N., Bulau, P., and Wuhrer, M. (2015) Comparison of methods for the analysis of therapeutic immunoglobulin G Fc-glycosylation profiles--part 1: separation-based methods. *MAbs* 7, 167-179

34. IMGT® (2001) Protein display: Human IGH C-REGIONS. [last updated 20/12/2012; accessed 01/2016]  
[http://www.imgt.org/IMGTrepertoire/Proteins/protein/human/IGH/IGHC/Hu\\_IGHCallgenes.html](http://www.imgt.org/IMGTrepertoire/Proteins/protein/human/IGH/IGHC/Hu_IGHCallgenes.html)
35. (2015) UniProt: a hub for protein information. *Nucleic Acids Res* 43, D204-212
36. Lavinder, J. J., Horton, A. P., Georgiou, G., and Ippolito, G. C. (2015) Next-generation sequencing and protein mass spectrometry for the comprehensive analysis of human cellular and serum antibody repertoires. *Curr Opin Chem Biol* 24, 112-120
37. Cheung, W. C., Beausoleil, S. A., Zhang, X., Sato, S., Schieferl, S. M., Wieler, J. S., Beaudet, J. G., Ramenani, R. K., Popova, L., Comb, M. J., Rush, J., and Polakiewicz, R. D. (2012) A proteomics approach for the identification and cloning of monoclonal antibodies from serum. *Nat Biotechnol* 30, 447-452
38. Dorrington, K. J., and Bennich, H. H. (1978) Structure-function relationships in human immunoglobulin E. *Immunol Rev* 41, 3-25
39. de Haan, N., Reiding, K. R., Habberger, M., Reusch, D., Falck, D., and Wuhler, M. (2015) Linkage-specific sialic acid derivatization for MALDI-TOF-MS profiling of IgG glycopeptides. *Anal Chem* 87, 8284-8291
40. Lin, C. W., Tsai, M. H., Li, S. T., Tsai, T. I., Chu, K. C., Liu, Y. C., Lai, M. Y., Wu, C. Y., Tseng, Y. C., Shivatare, S. S., Wang, C. H., Chao, P., Wang, S. Y., Shih, H. W., Zeng, Y. F., You, T. H., Liao, J. Y., Tu, Y. C., Lin, Y. S., Chuang, H. Y., Chen, C. L., Tsai, C. S., Huang, C. C., Lin, N. H., Ma, C., Wu, C. Y., and Wong, C. H. (2015) A common glycan structure on immunoglobulin G for enhancement of effector functions. *Proc Natl Acad Sci U S A* 112, 10611-10616
41. Akmacic, I. T., Ventham, N. T., Theodoratou, E., Vuckovic, F., Kennedy, N. A., Kristic, J., Nimmo, E. R., Kalla, R., Drummond, H., Stambuk, J., Dunlop, M. G., Novokmet, M., Aulchenko, Y., Gornik, O., Campbell, H., Pucic Bakovic, M., Satsangi, J., and Lauc, G. (2015) Inflammatory bowel disease associates with proinflammatory potential of the immunoglobulin G glycome. *Inflamm Bowel Dis* 21, 1237-1247
42. Wuhler, M., Selman, M. H., McDonnell, L. A., Kumpf, T., Derfuss, T., Khademi, M., Olsson, T., Hohlfeld, R., Meinel, E., and Krumbholz, M. (2015) Pro-inflammatory pattern of IgG1 Fc glycosylation in multiple sclerosis cerebrospinal fluid. *J Neuroinflammation* 12, 235
43. Vuckovic, F., Kristic, J., Gudelj, I., Teruel, M., Keser, T., Pezer, M., Pucic-Bakovic, M., Stambuk, J., Trbojevic-Akmacic, I., Barrios, C., Pavic, T., Menni, C., Wang, Y., Zhou, Y., Cui, L., Song, H., Zeng, Q., Guo, X., Pons-Estel, B. A., McKeigue, P., Leslie Patrick, A., Gornik, O., Spector, T. D., Harjacek, M., Alarcon-Riquelme, M., Molokhia, M., Wang, W., and Lauc, G. (2015) Association of systemic lupus erythematosus with decreased immunosuppressive potential of the IgG glycome. *Arthritis Rheumatol* 67, 2978-2989
44. Wong, A. H., Fukami, Y., Sudo, M., Kokubun, N., Hamada, S., and Yuki, N. (2015) Sialylated IgG-Fc: a novel biomarker of chronic inflammatory demyelinating polyneuropathy. *J Neurol Neurosurg Psychiatry* 87, 275-279
45. Gardinassi, L. G., Dotz, V., Hipgrave Ederveen, A., de Almeida, R. P., Nery Costa, C. H., Costa, D. L., de Jesus, A. R., Mayboroda, O. A., Garcia, G. R., Wuhler, M., and de Miranda Santos, I. K. (2014) Clinical severity of visceral leishmaniasis is associated with changes in immunoglobulin g fc N-glycosylation. *MBio* 5, e01844
46. Ackerman, M. E., Crispin, M., Yu, X., Baruah, K., Boesch, A. W., Harvey, D. J., Dugast, A. S., Heizen, E. L., Ercan, A., Choi, I., Streeck, H., Nigrovic, P. A., Bailey-Kellogg, C., Scanlan, C., and Alter, G. (2013) Natural variation in Fc glycosylation of HIV-specific antibodies impacts antiviral activity. *J Clin Invest* 123, 2183-2192
47. Ho, C. H., Chien, R. N., Cheng, P. N., Liu, J. H., Liu, C. K., Su, C. S., Wu, I. C., Li, I. C., Tsai, H. W., Wu, S. L., Liu, W. C., Chen, S. H., and Chang, T. T. (2015) Aberrant serum immunoglobulin G glycosylation in chronic hepatitis B is associated with histological liver damage and reversible by antiviral therapy. *J Infect Dis* 211, 115-124

48. Chen, G., Wang, Y., Qin, X., Li, H., Guo, Y., Wang, Y., Liu, H., Wang, X., Song, G., Li, F., Li, F., Guo, S., Qiu, L., and Li, Z. (2013) Change in IgG1 Fc N-linked glycosylation in human lung cancer: age- and sex-related diagnostic potential. *Electrophoresis* 34, 2407-2416
49. Kawaguchi-Sakita, N., Kaneshiro-Nakagawa, K., Kawashima, M., Sugimoto, M., Tokiwa, M., Suzuki, E., Kajihara, S., Fujita, Y., Iwamoto, S., Tanaka, K., and Toi, M. (2015) Serum immunoglobulin G Fc region N-glycosylation profiling by matrix-assisted laser desorption/ionization mass spectrometry can distinguish breast cancer patients from cancer-free controls. *Biochem Biophys Res Commun* 469, 1140-1145
50. Saldova, R., Stockmann, H., O'Flaherty, R., Lefeber, D. J., Jaeken, J., and Rudd, P. M. (2015) N-Glycosylation of Serum IgG and Total Glycoproteins in MAN1B1 Deficiency. *J Proteome Res* 14, 4402-4412
51. Maratha, A., Stockmann, H., Coss, K. P., Estela Rubio-Gozalbo, M., Knerr, I., Fitzgibbon, M., McVeigh, T. P., Foley, P., Moss, C., Colhoun, H. O., van Erven, B., Stephens, K., Doran, P., Rudd, P., and Treacy, E. (2016) Classical galactosaemia: novel insights in IgG N-glycosylation and N-glycan biosynthesis. *Eur J Hum Genet*
52. Rudd, P. M., Guile, G. R., Kuster, B., Harvey, D. J., Opdenakker, G., and Dwek, R. A. (1997) Oligosaccharide sequencing technology. *Nature* 388, 205-207
53. Ahn, J., Bones, J., Yu, Y. Q., Rudd, P. M., and Gilar, M. (2010) Separation of 2-aminobenzamide labeled glycans using hydrophilic interaction chromatography columns packed with 1.7 microm sorbent. *J Chromatogr B Analyt Technol Biomed Life Sci* 878, 403-408
54. Akmacic, I. T., Ugrina, I., Stambuk, J., Gudelj, I., Vuckovic, F., Lauc, G., and Pucic-Bakovic, M. (2015) High-Throughput Glycomics: Optimization of Sample Preparation. *Biochemistry (Mosc)* 80, 934-942
55. Stockmann, H., Duke, R. M., Millan Martin, S., and Rudd, P. M. (2015) Ultrahigh throughput, ultrafiltration-based n-glycomics platform for ultraperformance liquid chromatography (ULTRA(3)). *Anal Chem* 87, 8316-8322
56. Kristic, J., Vuckovic, F., Menni, C., Klaric, L., Keser, T., Beceheli, I., Pucic-Bakovic, M., Novokmet, M., Mangino, M., Thaqi, K., Rudan, P., Novokmet, N., Sarac, J., Missoni, S., Kolcic, I., Polasek, O., Rudan, I., Campbell, H., Hayward, C., Aulchenko, Y., Valdes, A., Wilson, J. F., Gornik, O., Primorac, D., Zoldos, V., Spector, T., and Lauc, G. (2014) Glycans are a novel biomarker of chronological and biological ages. *J Gerontol A Biol Sci Med Sci* 69, 779-789
57. Dard, P., Lefranc, M. P., Osipova, L., and Sanchez-Mazas, A. (2001) DNA sequence variability of IGHG3 alleles associated to the main G3m haplotypes in human populations. *Eur J Hum Genet* 9, 765-772
58. Falck, D., Jansen, B. C., Plomp, R., Reusch, D., Habberger, M., and Wührer, M. (2015) Glycoforms of Immunoglobulin G Based Biopharmaceuticals Are Differentially Cleaved by Trypsin Due to the Glycoform Influence on Higher-Order Structure. *J Proteome Res* 14, 4019-4028
59. Fang, C., Xiong, Z., Qin, H., Huang, G., Liu, J., Ye, M., Feng, S., and Zou, H. (2014) One-pot synthesis of magnetic colloidal nanocrystal clusters coated with chitosan for selective enrichment of glycopeptides. *Anal Chim Acta* 841, 99-105
60. Sheng, Q., Su, X., Li, X., Ke, Y., and Liang, X. (2014) A dextran-bonded stationary phase for saccharide separation. *J Chromatogr A* 1345, 57-67
61. Ji, Y., Xiong, Z., Huang, G., Liu, J., Zhang, Z., Liu, Z., Ou, J., Ye, M., and Zou, H. (2014) Efficient enrichment of glycopeptides using metal-organic frameworks by hydrophilic interaction chromatography. *Analyst* 139, 4987-4993
62. Li, S., Wang, L., Zhao, S., Lin, J., Zheng, J., and Lin, Z. (2015) Preparation of phenyl-functionalized magnetic mesoporous silica microspheres for the fast separation and selective enrichment of phenyl-containing peptides. *J Sep Sci* 38, 3954-3960
63. Jiang, H., Yuan, H., Qu, Y., Liang, Y., Jiang, B., Wu, Q., Deng, N., Liang, Z., Zhang, L., and Zhang, Y. (2016) Preparation of hydrophilic monolithic capillary column by in situ photo-



polymerization of N-vinyl-2-pyrrolidinone and acrylamide for highly selective and sensitive enrichment of N-linked glycopeptides. *Talanta* 146, 225-230

64. Cao, L., Yu, L., Guo, Z., Li, X., Xue, X., and Liang, X. (2013) Application of a strong anion exchange material in electrostatic repulsion-hydrophilic interaction chromatography for selective enrichment of glycopeptides. *J Chromatogr A* 1299, 18-24

65. Wohlgemuth, J., Karas, M., Eichhorn, T., Hendriks, R., and Andrecht, S. (2009) Quantitative site-specific analysis of protein glycosylation by LC-MS using different glycopeptide-enrichment strategies. *Anal Biochem* 395, 178-188

66. Shen, A., Guo, Z., Yu, L., Cao, L., and Liang, X. (2011) A novel zwitterionic HILIC stationary phase based on "thiol-ene" click chemistry between cysteine and vinyl silica. *Chem Commun (Camb)* 47, 4550-4552

67. Huang, G., Xiong, Z., Qin, H., Zhu, J., Sun, Z., Zhang, Y., Peng, X., ou, J., and Zou, H. (2014) Synthesis of zwitterionic polymer brushes hybrid silica nanoparticles via controlled polymerization for highly efficient enrichment of glycopeptides. *Anal Chim Acta* 809, 61-68

68. Chen, Y., Xiong, Z., Zhang, L., Zhao, J., Zhang, Q., Peng, L., Zhang, W., Ye, M., and Zou, H. (2015) Facile synthesis of zwitterionic polymer-coated core-shell magnetic nanoparticles for highly specific capture of N-linked glycopeptides. *Nanoscale* 7, 3100-3108

69. Ruhaak, L. R., and Lebrilla, C. B. (2015) Applications of Multiple Reaction Monitoring to Clinical Glycomics. *Chromatographia* 78, 335-342

70. Hong, Q., Ruhaak, L. R., Stroble, C., Parker, E., Huang, J., Maverakis, E., and Lebrilla, C. B. (2015) A Method for Comprehensive Glycosite-Mapping and Direct Quantitation of Serum Glycoproteins. *J Proteome Res* 14, 5179-5192

71. Hong, Q., Lebrilla, C. B., Miyamoto, S., and Ruhaak, L. R. (2013) Absolute quantitation of immunoglobulin G and its glycoforms using multiple reaction monitoring. *Anal Chem* 85, 8585-8593

72. Yuan, W., Sanda, M., Wu, J., Koomen, J., and Goldman, R. (2015) Quantitative analysis of immunoglobulin subclasses and subclass specific glycosylation by LC-MS-MRM in liver disease. *J Proteomics* 116, 24-33

73. Heemskerk, A. A., Wührer, M., Busnel, J. M., Koeleman, C. A., Selman, M. H., Vidarsson, G., Kapur, R., Schoenmaker, B., Derks, R. J., Deelder, A. M., and Mayboroda, O. A. (2013) Coupling porous sheathless interface MS with transient-ITP in neutral capillaries for improved sensitivity in glycopeptide analysis. *Electrophoresis* 34, 383-387

74. Nishikaze, T., Kawabata, S., and Tanaka, K. (2014) In-depth structural characterization of N-linked glycopeptides using complete derivatization for carboxyl groups followed by positive- and negative-ion tandem mass spectrometry. *Anal Chem* 86, 5360-5369

75. Gomes de Oliveira, A. G., Roy, R., Raymond, C., Bodnar, E. D., Tayi, V. S., Butler, M., Durocher, Y., and Perreault, H. (2015) A systematic study of glycopeptide esterification for the semi-quantitative determination of sialylation in antibodies. *Rapid Commun Mass Spectrom* 29, 1817-1826

76. Reiding, K. R., Blank, D., Kuijper, D. M., Deelder, A. M., and Wührer, M. (2014) High-throughput profiling of protein N-glycosylation by MALDI-TOF-MS employing linkage-specific sialic acid esterification. *Anal Chem* 86, 5784-5793

77. Amano, J., Nishikaze, T., Tougasaki, F., Jinmei, H., Sugimoto, I., Sugawara, S., Fujita, M., Osumi, K., and Mizuno, M. (2010) Derivatization with 1-pyrenyldiazomethane enhances ionization of glycopeptides but not peptides in matrix-assisted laser desorption/ionization mass spectrometry. *Anal Chem* 82, 8738-8743

78. Nishikaze, T., Nakamura, T., Jinmei, H., and Amano, J. (2011) Negative-ion MALDI-MS2 for discrimination of alpha2,3- and alpha2,6-sialylation on glycopeptides labeled with a pyrene derivative. *J Chromatogr B Analyt Technol Biomed Life Sci* 879, 1419-1428

79. Pabst, M., Benesova, I., Fagerer, S. R., Jacobsen, M., Eyer, K., Schmidt, G., Steinhoff, R., Krismer, J., Wahl, F., Preisler, J., and Zenobi, R. (2016) Differential Isotope Labeling of Glycopeptides for Accurate Determination of Differences in Site-Specific Glycosylation. *J Proteome Res* 15, 326-331

80. Gautier, V., Boumeester, A. J., Lossl, P., and Heck, A. J. (2015) Lysine conjugation properties in human IgGs studied by integrating high-resolution native mass spectrometry and bottom-up proteomics. *Proteomics* 15, 2756-2765
81. An, Y., Zhang, Y., Mueller, H. M., Shameem, M., and Chen, X. (2014) A new tool for monoclonal antibody analysis: application of IdeS proteolysis in IgG domain-specific characterization. *MAbs* 6, 879-893
82. Lynaugh, H., Li, H., and Gong, B. (2013) Rapid Fc glycosylation analysis of Fc fusions with IdeS and liquid chromatography mass spectrometry. *MAbs* 5, 641-645
83. Tran, B. Q., Barton, C., Feng, J., Sandjong, A., Yoon, S. H., Awasthi, S., Liang, T., Khan, M. M., Kilgour, D. P., Goodlett, D. R., and Goo, Y. A. (2015) Comprehensive glycosylation profiling of IgG and IgG-fusion proteins by top-down MS with multiple fragmentation techniques. *J Proteomics* 134, 93-101
84. Stoll, D. R., Harmes, D. C., Danforth, J., Wagner, E., Guillarme, D., Fekete, S., and Beck, A. (2015) Direct identification of rituximab main isoforms and subunit analysis by online selective comprehensive two-dimensional liquid chromatography-mass spectrometry. *Anal Chem* 87, 8307-8315
85. Nicolardi, S., Deelder, A. M., Palmblad, M., and van der Burgt, Y. E. (2014) Structural analysis of an intact monoclonal antibody by online electrochemical reduction of disulfide bonds and Fourier transform ion cyclotron resonance mass spectrometry. *Anal Chem* 86, 5376-5382
86. Rosati, S., Yang, Y., Barendregt, A., and Heck, A. J. (2014) Detailed mass analysis of structural heterogeneity in monoclonal antibodies using native mass spectrometry. *Nat Protoc* 9, 967-976
87. Zhang, H., Cui, W., and Gross, M. L. (2014) Mass spectrometry for the biophysical characterization of therapeutic monoclonal antibodies. *FEBS Lett* 588, 308-317
88. Holland, M., Yagi, H., Takahashi, N., Kato, K., Savage, C. O., Goodall, D. M., and Jefferis, R. (2006) Differential glycosylation of polyclonal IgG, IgG-Fc and IgG-Fab isolated from the sera of patients with ANCA-associated systemic vasculitis. *Biochim Biophys Acta* 1760, 669-677
89. Stadlmann, J., Pabst, M., and Altmann, F. (2010) Analytical and Functional Aspects of Antibody Sialylation. *J Clin Immunol* 30 Suppl 1, S15-19
90. Radcliffe, C. M., Arnold, J. N., Suter, D. M., Wormald, M. R., Harvey, D. J., Royle, L., Mimura, Y., Kimura, Y., Sim, R. B., Inoges, S., Rodriguez-Calvillo, M., Zabalegui, N., de Cerio, A. L., Potter, K. N., Mockridge, C. I., Dwek, R. A., Bendandi, M., Rudd, P. M., and Stevenson, F. K. (2007) Human follicular lymphoma cells contain oligomannose glycans in the antigen-binding site of the B-cell receptor. *J Biol Chem* 282, 7405-7415
91. Coelho, V., Krysov, S., Ghaemmaghami, A. M., Emara, M., Potter, K. N., Johnson, P., Packham, G., Martinez-Pomares, L., and Stevenson, F. K. (2010) Glycosylation of surface Ig creates a functional bridge between human follicular lymphoma and microenvironmental lectins. *Proc Natl Acad Sci U S A* 107, 18587-18592
92. Sabouri, Z., Schofield, P., Horikawa, K., Spierings, E., Kipling, D., Randall, K. L., Langley, D., Roome, B., Vazquez-Lombardi, R., Rouet, R., Hermes, J., Chan, T. D., Brink, R., Dunn-Walters, D. K., Christ, D., and Goodnow, C. C. (2014) Redemption of autoantibodies on anergic B cells by variable-region glycosylation and mutation away from self-reactivity. *Proc Natl Acad Sci U S A* 111, E2567-2575
93. Mahan, A. E., Tedesco, J., Dionne, K., Baruah, K., Cheng, H. D., De Jager, P. L., Barouch, D. H., Suscovich, T., Ackerman, M., Crispin, M., and Alter, G. (2015) A method for high-throughput, sensitive analysis of IgG Fc and Fab glycosylation by capillary electrophoresis. *J Immunol Methods* 417, 34-44
94. Blomme, B., Van Steenkiste, C., Grassi, P., Haslam, S. M., Dell, A., Callewaert, N., and Van Vlierberghe, H. (2011) Alterations of serum protein N-glycosylation in two mouse models of chronic liver disease are hepatocyte and not B cell driven. *Am J Physiol Gastrointest Liver Physiol* 300, G833-842

95. Raju, T. S., Briggs, J. B., Borge, S. M., and Jones, A. J. (2000) Species-specific variation in glycosylation of IgG: evidence for the species-specific sialylation and branch-specific galactosylation and importance for engineering recombinant glycoprotein therapeutics. *Glycobiology* 10, 477-486
96. Stavenhagen, K., Plomp, R., and Wuhrer, M. (2015) Site-Specific Protein N- and O-Glycosylation Analysis by a C18-Porous Graphitized Carbon-Liquid Chromatography-Electrospray Ionization Mass Spectrometry Approach Using Pronase Treated Glycopeptides. *Anal Chem* 87, 11691-11699
97. Nwosu, C. C., Huang, J., Aldredge, D. L., Strum, J. S., Hua, S., Seipert, R. R., and Lebrilla, C. B. (2013) In-gel nonspecific proteolysis for elucidating glycoproteins: a method for targeted protein-specific glycosylation analysis in complex protein mixtures. *Anal Chem* 85, 956-963
98. Dodds, E. D., Seipert, R. R., Clowers, B. H., German, J. B., and Lebrilla, C. B. (2009) Analytical performance of immobilized pronase for glycopeptide footprinting and implications for surpassing reductionist glycoproteomics. *J Proteome Res* 8, 502-512
99. Scherer, H. U., van der Woude, D., Ioan-Facsinay, A., el Bannoudi, H., Trouw, L. A., Wang, J., Haupl, T., Burmester, G. R., Deelder, A. M., Huizinga, T. W., Wuhrer, M., and Toes, R. E. (2010) Glycan profiling of anti-citrullinated protein antibodies isolated from human serum and synovial fluid. *Arthritis Rheum* 62, 1620-1629
100. Scherer, H. U., Wang, J., Toes, R. E., van der Woude, D., Koeleman, C. A., de Boer, A. R., Huizinga, T. W., Deelder, A. M., and Wuhrer, M. (2009) Immunoglobulin 1 (IgG1) Fc-glycosylation profiling of anti-citrullinated peptide antibodies from human serum. *Proteomics Clin Appl* 3, 106-115
101. Brown, E. P., Normandin, E., Osei-Owusu, N. Y., Mahan, A. E., Chan, Y. N., Lai, J. I., Vaccari, M., Rao, M., Franchini, G., Alter, G., and Ackerman, M. E. (2015) Microscale purification of antigen-specific antibodies. *J Immunol Methods* 425, 27-36
102. Wang, T. T., Maamary, J., Tan, G. S., Bournazos, S., Davis, C. W., Krammer, F., Schlesinger, S. J., Palese, P., Ahmed, R., and Ravetch, J. V. (2015) Anti-HA Glycoforms Drive B Cell Affinity Selection and Determine Influenza Vaccine Efficacy. *Cell* 162, 160-169
103. Kapur, R., Della Valle, L., Verhagen, O. J., Hipgrave Ederveen, A., Ligthart, P., de Haas, M., Kumpel, B., Wuhrer, M., van der Schoot, C. E., and Vidarsson, G. (2015) Prophylactic anti-D preparations display variable decreases in Fc-fucosylation of anti-D. *Transfusion* 55, 553-562
104. Kapur, R., Della Valle, L., Sonneveld, M., Hipgrave Ederveen, A., Visser, R., Ligthart, P., de Haas, M., Wuhrer, M., van der Schoot, C. E., and Vidarsson, G. (2014) Low anti-RhD IgG-Fc-fucosylation in pregnancy: a new variable predicting severity in haemolytic disease of the fetus and newborn. *Br J Haematol* 166, 936-945
105. Kapur, R., Kustiawan, I., Vestrheim, A., Koeleman, C. A., Visser, R., Einarsdottir, H. K., Porcelijn, L., Jackson, D., Kumpel, B., Deelder, A. M., Blank, D., Skogen, B., Killie, M. K., Michaelsen, T. E., de Haas, M., Rispen, T., van der Schoot, C. E., Wuhrer, M., and Vidarsson, G. (2014) A prominent lack of IgG1-Fc fucosylation of platelet alloantibodies in pregnancy. *Blood* 123, 471-480
106. Wuhrer, M., Stavenhagen, K., Koeleman, C. A., Selman, M. H., Harper, L., Jacobs, B. C., Savage, C. O., Jefferis, R., Deelder, A. M., and Morgan, M. (2015) Skewed Fc glycosylation profiles of anti-proteinase 3 immunoglobulin G1 autoantibodies from granulomatosis with polyangiitis patients show low levels of bisection, galactosylation, and sialylation. *J Proteome Res* 14, 1657-1665
107. Espy, C., Morelle, W., Kaviani, N., Grange, P., Goulvestre, C., Viallon, V., Chereau, C., Pagnoux, C., Michalski, J. C., Guillevin, L., Weill, B., Batteux, F., and Guilpain, P. (2011) Sialylation levels of anti-proteinase 3 antibodies are associated with the activity of granulomatosis with polyangiitis (Wegener's). *Arthritis Rheum* 63, 2105-2115
108. Harre, U., Lang, S. C., Pfeifle, R., Rombouts, Y., Fruhbesser, S., Amara, K., Bang, H., Lux, A., Koeleman, C. A., Baum, W., Dietel, K., Grohn, F., Malmstrom, V., Klareskog, L., Kronke, G., Kocijan, R., Nimmerjahn, F., Toes, R. E., Herrmann, M., Scherer, H. U., and Schett, G. (2015) Glycosylation of immunoglobulin G determines osteoclast differentiation and bone loss. *Nat Commun* 6, 6651

109. Selman, M. H., de Jong, S. E., Soonawala, D., Kroon, F. P., Adegnika, A. A., Deelder, A. M., Hokke, C. H., Yazdanbakhsh, M., and Wuhrer, M. (2012) Changes in antigen-specific IgG1 Fc N-glycosylation upon influenza and tetanus vaccination. *Mol Cell Proteomics* 11, M111.014563
110. Takahashi, K., Smith, A. D., Poulsen, K., Kilian, M., Julian, B. A., Mestecky, J., Novak, J., and Renfrow, M. B. (2012) Naturally occurring structural isomers in serum IgA1 o-glycosylation. *J Proteome Res* 11, 692-702
111. Royle, L., Roos, A., Harvey, D. J., Wormald, M. R., van Gijlswijk-Janssen, D., Redwan el, R. M., Wilson, I. A., Daha, M. R., Dwek, R. A., and Rudd, P. M. (2003) Secretory IgA N- and O-glycans provide a link between the innate and adaptive immune systems. *J Biol Chem* 278, 20140-20153
112. Kolka, R., Valdimarsson, H., Bodvarsson, M., Hardarson, S., and Jonsson, T. (2013) Defective immunoglobulin A (IgA) glycosylation and IgA deposits in patients with IgA nephropathy. *Apmis* 121, 890-897
113. Iwatani, H., Inoue, T., Wada, Y., Nagasawa, Y., Yamamoto, R., Iijima, H., Takehara, T., Imai, E., Rakugi, H., and Isaka, Y. (2012) Quantitative change of IgA hinge O-glycan composition is a novel marker of therapeutic responses of IgA nephropathy. *Biochem Biophys Res Commun* 428, 339-342
114. Huang, J., Guerrero, A., Parker, E., Strum, J. S., Smilowitz, J. T., German, J. B., and Lebrilla, C. B. (2015) Site-specific glycosylation of secretory immunoglobulin A from human colostrum. *J Proteome Res* 14, 1335-1349
115. Deshpande, N., Jensen, P. H., Packer, N. H., and Kolarich, D. (2010) GlycoSpectrumScan: fishing glycopeptides from MS spectra of protease digests of human colostrum sIgA. *J Proteome Res* 9, 1063-1075
116. Franc, V., Rehulka, P., Raus, M., Stulik, J., Novak, J., Renfrow, M. B., and Sebela, M. (2013) Elucidating heterogeneity of IgA1 hinge-region O-glycosylation by use of MALDI-TOF/TOF mass spectrometry: role of cysteine alkylation during sample processing. *J Proteomics* 92, 299-312
117. Hur, E. M., Patel, S. N., Shimizu, S., Rao, D. S., Gnanapragasam, P. N., An, D. S., Yang, L., and Baltimore, D. (2012) Inhibitory effect of HIV-specific neutralizing IgA on mucosal transmission of HIV in humanized mice. *Blood* 120, 4571-4582
118. Yu, X., Duval, M., Lewis, C., Gawron, M. A., Wang, R., Posner, M. R., and Cavacini, L. A. (2013) Impact of IgA constant domain on HIV-1 neutralizing function of monoclonal antibody F425A1g8. *J Immunol* 190, 205-210
119. Paul, M., Reljic, R., Klein, K., Drake, P. M., van Dolleweerd, C., Pabst, M., Windwarder, M., Arcalis, E., Stoger, E., Altmann, F., Cosgrove, C., Bartolf, A., Baden, S., and Ma, J. K. (2014) Characterization of a plant-produced recombinant human secretory IgA with broad neutralizing activity against HIV. *MAbs* 6, 1585-1597
120. Pabst, M., Chang, M., Stadlmann, J., and Altmann, F. (2012) Glycan profiles of the 27 N-glycosylation sites of the HIV envelope protein CN54gp140. *Biol Chem* 393, 719-730
121. Rouwendal, G. J., van der Lee, M. M., Meyer, S., Reiding, K. R., Schouten, J., de Roo, G., Egging, D. F., Leusen, J. H., Boross, P., Wuhrer, M., Verheijden, G. F., Dokter, W. H., Timmers, M., and Ubink, R. (2016) A comparison of anti-HER2 IgA and IgG1 in vivo efficacy is facilitated by high N-glycan sialylation of the IgA. *MAbs* 8, 74-86
122. Krugmann, S., Pleass, R. J., Atkin, J. D., and Woof, J. M. (1997) Structural requirements for assembly of dimeric IgA probed by site-directed mutagenesis of J chain and a cysteine residue of the alpha-chain CH2 domain. *J Immunol* 159, 244-249
123. Hughes, G. J., Reason, A. J., Savoy, L., Jatton, J., and Frutiger-Hughes, S. (1999) Carbohydrate moieties in human secretory component. *Biochim Biophys Acta* 1434, 86-93
124. Pabst, M., Kuster, S. K., Wahl, F., Krismer, J., Dittrich, P. S., and Zenobi, R. (2015) A Microarray-Matrix-assisted Laser Desorption/Ionization-Mass Spectrometry Approach for Site-specific Protein N-glycosylation Analysis, as Demonstrated for Human Serum Immunoglobulin M (IgM). *Mol Cell Proteomics* 14, 1645-1656

125. Loos, A., Gruber, C., Altmann, F., Mehofer, U., Hensel, F., Grandits, M., Oostenbrink, C., Stadlmayr, G., Furtmuller, P. G., and Steinkellner, H. (2014) Expression and glycoengineering of functionally active heteromultimeric IgM in plants. *Proc Natl Acad Sci U S A* 111, 6263-6268
126. Wu, G., Hitchen, P. G., Panico, M., North, S. J., Barbouche, M. R., Binet, D., Morris, H. R., Dell, A., and Haslam, S. M. (2015) Glycoproteomic studies of IgE from a novel hyper IgE syndrome linked to PGM3 mutation. *Glycoconj J*
127. Shade, K. T., Platzer, B., Washburn, N., Mani, V., Bartsch, Y. C., Conroy, M., Pagan, J. D., Bosques, C., Mempel, T. R., Fiebiger, E., and Anthony, R. M. (2015) A single glycan on IgE is indispensable for initiation of anaphylaxis. *J Exp Med* 212, 457-467
128. Johansson, S. G. (1967) Raised levels of a new immunoglobulin class (IgND) in asthma. *Lancet* 2, 951-953
129. King, C. L., Poindexter, R. W., Ragunathan, J., Fleisher, T. A., Ottesen, E. A., and Nutman, T. B. (1991) Frequency analysis of IgE-secreting B lymphocytes in persons with normal or elevated serum IgE levels. *J Immunol* 146, 1478-1483
130. Little, M. A., Al-Ani, B., Ren, S., Al-Nuaimi, H., Leite, M., Jr., Alpers, C. E., Savage, C. O., and Duffield, J. S. (2012) Anti-proteinase 3 anti-neutrophil cytoplasm autoantibodies recapitulate systemic vasculitis in mice with a humanized immune system. *PLoS One* 7, e28626
-

