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Reference List

1. Northrup, S. H.; Erickson, H. P. *Proc. Natl. Acad. Sci. U. S. A* **1992**, *89*, 3338-3342.
2. Shapiro, R.; Vallee, B. L. *Biochemistry* **1991**, *30*, 2246-2255.
3. Albeck, S.; Schreiber, G. *Biochemistry* **1999**, *38*, 11-21.
4. Crowley, P. B.; Ubbink, M. *Acc. Chem. Res.* **2003**, *36*, 723-730.
5. Kang, C. H.; Ferguson-Miller, S.; Margoliash, E. *J. Biol. Chem.* **1977**, *252*, 919-926.
6. Setif, P. Q.; Bottin, H. *Biochemistry* **1995**, *34*, 9059-9070.
7. Schlarb-Ridley, B. G.; Bendall, D. S.; Howe, C. J. *Biochemistry* **2002**, *41*, 3279-3285.
8. Albarran, C.; Navarro, J. A.; Molina-Heredia, F. P.; Murdoch, P. S.; De la Rosa, M. A.; Hervas, M. *Biochemistry* **2005**, *44*, 11601-11607.
9. Prudencio, M.; Ubbink, M. *J. Mol. Recognit.* **2004**, *17*, 524-539.
10. Schreiber, G.; Fersht, A. R. *J. Mol. Biol.* **1995**, *248*, 478-486.
11. Clackson, T.; Wells, J. A. *Science* **1995**, *267*, 383-386.
12. Moreira, I. S.; Fernandes, P. A.; Ramos, M. J. *Proteins* **2007**, *68*, 803-812.
13. Crowley, P. B.; Carrondo, M. A. *Proteins* **2004**, *55*, 603-612.
14. Kiel, C.; Selzer, T.; Shaul, Y.; Schreiber, G.; Herrmann, C. *Proc Natl Acad Sci USA* **2004**, *101*, 9223-9228.
15. Volkov, A. N.; Bashir, Q.; Worrall, J. A. R.; Ubbink, M. *J. Mol. Biol.* **2009**, *385*, 1003-1013.
16. Schreiber, G.; Keating, A. E. *Curr. Opin. Struct. Biol.* **2011**, *21*, 50-61.
17. Williams, P. A.; Fulop, V.; Leung, Y. C.; Chan, C.; Moir, J. W.; Howlett, G.; Ferguson, S. J.; Radford, S. E.; Hajdu, J. *Nat. Struct. Biol.* **1995**, *2*, 975-982.
18. De Rienzo, F.; Gabdoulhine, R. R.; Menziani, M. C.; Wade, R. C. *Protein Sci.* **2000**, *9*, 1439-1454.
19. Nersissian, A. M.; Shipp, E. L. *Adv. Protein Chem.* **2002**, *60*, 271-340.
20. Bashir, Q.; Scanu, S.; Ubbink, M. *FEBS J.* **2011**, *278*, 1391-1400.
21. Ubbink, M. *FEBS Lett.* **2009**, *583*, 1060-1066.
22. Schreiber, G. *Curr. Opin. Struct. Biol.* **2002**, *12*, 41-47.
23. Schreiber, G.; Fersht, A. R. *Nat. Struct. Biol.* **1996**, *3*, 427-431.
24. Harel, M.; Spaar, A.; Schreiber, G. *Biophys. J.* **2009**, *96*, 4237-4248.

25. Keizers, P. H. J.; Ubbink, M. *Prog. Nucl. Mag. Res. Sp.* **2011**, *58*, 88-96.
26. Volkov, A. N.; Worrall, J. A. R.; Holtzmann, E.; Ubbink, M. *Proc Natl Acad Sci USA* **2006**, *103*, 18945-18950.
27. Kim, Y. C.; Tang, C.; Clore, G. M.; Hummer, G. *Proc. Natl. Acad. Sci. U. S. A* **2008**, *105*, 12855-12860.
28. Bashir, Q.; Volkov, A. N.; Ullmann, G. M.; Ubbink, M. *J. Am. Chem. Soc.* **2010**, *132*, 241-247.
29. Scanu, S.; Forster, J.; Finiguerra, M. G.; Shabestari, M. H.; Huber, M.; Ubbink, M. *Chembiochem* **2012**.
30. Xu, X. F.; Reinle, W. G.; Hannemann, F.; Konarev, P. V.; Svergun, D. I.; Bernhardt, R.; Ubbink, M. *J. Am. Chem. Soc.* **2008**, *130*, 6395-6403.
31. Volkov, A. N.; Bashir, Q.; Worrall, J. A. R.; Ullmann, G. M.; Ubbink, M. *J. Am. Chem. Soc.* **2010**, *132*, 11487-11495.
32. Hulsker, R.; Baranova, M. V.; Bullerjahn, G. S.; Ubbink, M. *J. Am. Chem. Soc.* **2008**, *130*, 1985-1991.
33. Mitchell, P. *FEBS Lett.* **1977**, *78*, 1-20.
34. Hervas, M.; Navarro, J. A.; Diaz, A.; Bottin, H.; De la Rosa, M. A. *Biochemistry* **1995**, *34*, 11321-11326.
35. Mason, J. M.; Bendall, D. S.; Howe, C. J.; Worrall, J. A. *Biochim. Biophys. Acta* **2012**, *1824*, 311-318.
36. Worrall, J. A.; Luisi, B. F.; Schlarb-Ridley, B. G.; Bendall, D. S.; Howe, C. J. *Biochem. Soc. Trans.* **2008**, *36*, 1175-1179.
37. Howe, C. J.; Schlarb-Ridley, B. G.; Wastl, J.; Purton, S.; Bendall, D. S. *J. Exp. Bot.* **2006**, *57*, 13-22.
38. Katoh, S. *Nature* **1960**, *186*, 533-534.
39. Gorman, D. S.; Levine, R. P. *Proc. Natl. Acad. Sci. U. S. A* **1965**, *54*, 1665-1669.
40. Canters, G. W.; Gilardi, G. *FEBS Lett.* **1993**, *325*, 39-48.
41. Gray, H. B.; Malmstrom, B. G.; Williams, R. J. *J. Biol. Inorg. Chem.* **2000**, *5*, 551-559.
42. Schmidt, L.; Christensen, H. E.; Harris, P. *Acta Crystallogr. , Sect. D: Biol. Crystallogr.* **2006**, *62*, 1022-1029.
43. Bertini, I.; Ciurli, S.; Dikiy, A.; Fernández C.O.; Luchinat, C.; Safarov, N.; Shumilin, S.; Vila, A.J. *J. Am. Chem. Soc.* **2001**, *123*, 2405-2413.
44. Adman, E. T. *Adv. Protein Chem.* **1991**, *42*, 145-197.
45. Bertini, I.; Bryant, D.A.; Ciurli, S.; Dikiy, A.; Fernández C.O.; Luchinat, C.; Safarov, N.; Vila, A.J.; Zhao J. *J. Biol. Chem.* **2001**, *276*, 47217-47226.

46. Ubbink, M.; Ejdeback, M.; Karlsson, B. G.; Bendall, D. S. *Structure* **1998**, *6*, 323-335.
47. Ubbink, M.; Ejdeback, M.; Crowley, P. B.; Schlarb, B. G.; Howe, C. J.; Karlsson, B. G.; Bendall, D. S.; Canters, G. W. *J. Inorg. Biochem.* **1999**, *74*, 321.
48. Ejdeback, M.; Bergkvist, A.; Karlsson, B. G.; Ubbink, M. *Biochemistry* **2000**, *39*, 5022-5027.
49. Diaz-Moreno, I.; Diaz-Quintana, A.; De la Rosa, M. A.; Crowley, P. B.; Ubbink, M. *Biochemistry* **2005**, *44*, 3176-3183.
50. Diaz-Moreno, I.; Diaz-Quintana, A.; De la Rosa, M. A.; Ubbink, M. *J. Biol. Chem.* **2005**, *280*, 18908-18915.
51. Cramer, W. A.; Soriano, G. M.; Ponomarev, M.; Huang, D.; Zhang, H.; Martinez, S. E.; Smith, J. L. *Annu. Rev. Plant Physiol Plant Mol. Biol.* **1996**, *47*, 477-508.
52. Stroebel, D.; Choquet, Y.; Popot, J. L.; Picot, D. *Nature* **2003**, *426*, 413-418.
53. Martinez, S. E.; Huang, D.; Szczepaniak, A.; Cramer, W. A.; Smith, J. L. *Structure* **1994**, *2*, 95-105.
54. Crowley, P. B.; Otting, G.; Schlarb-Ridley, B. G.; Canters, G. W.; Ubbink, M. *J. Am. Chem. Soc.* **2001**, *123*, 10444-10453.
55. Cruz-Gallardo, I.; Diaz-Moreno, I.; Diaz-Quintana, A.; De la Rosa, M. A. *FEBS Lett.* **2012**, *586*, 646-652.
56. Pearson, D. C., Jr.; Gross, E. L.; David, E. S. *Biophys. J.* **1996**, *71*, 64-76.
57. Ullmann, G. M.; Knapp, E. W.; Kostic, N. M. *J. Am. Chem. Soc.* **1997**, *119*, 42-52.
58. Pearson, D. C., Jr.; Gross, E. L. *Biophys. J.* **1998**, *75*, 2698-2711.
59. Kannt, A.; Young, S.; Bendall, D. S. *Biochim. Biophys. Acta* **1996**, *1277*, 115-126.
60. Gong, X. S.; Wen, J. Q.; Fisher, N. E.; Young, S.; Howe, C. J.; Bendall, D. S.; Gray, J. C. *Eur. J. Biochem.* **2000**, *267*, 3461-3468.
61. De Rienzo, F.; Gabdoulline, R. R.; Menziani, M. C.; De Benedetti, P. G.; Wade, R. C. *Biophys. J.* **2001**, *81*, 3090-3104.
62. Lange, C.; Cornvik, T.; Diaz-Moreno, I.; Ubbink, M. *Biochim. Biophys. Acta* **2005**, *1707*, 179-188.
63. Soriano, G. M.; Ponamarev, M. V.; Tae, G. S.; Cramer, W. A. *Biochemistry* **1996**, *35*, 14590-14598.
64. Soriano, G. M.; Ponamarev, M. V.; Piskowski, R. A.; Cramer, W. A. *Biochemistry* **1998**, *37*, 15120-15128.
65. Meyer, T. E.; Zhao, Z. G.; Cusanovich, M. A.; Tollin, G. *Biochemistry* **1993**, *32*, 4552-4559.
66. Albarran, C.; Navarro, J.; De la Rosa, M. A.; Hervas, M. *Biochemistry* **2007**, *46*, 997-1003.
67. Schlarb-Ridley, B. G.; Navarro, J. A.; Spencer, M.; Bendall, D. S.; Hervas, M.; Howe, C. J.; De la Rosa, M. A. *Eur. J. Biochem.* **2002**, *269*, 5893-5902.

68. Gross, E. L. *Biophys. J.* **2004**, *87*, 2043-2059.
69. Hart, S. E.; Schlarb-Ridley, B. G.; Delon, C.; Bendall, D. S.; Howe, C. J. *Biochemistry* **2003**, *42*, 4829-4836.
70. Schlarb-Ridley, B. G.; Bendall, D. S.; Howe, C. J. *Biochemistry* **2003**, *42*, 4057-4063.
71. Gabdoulline, R. R.; Wade, R. C. *J. Am. Chem. Soc.* **2009**, *131*, 9230-9238.
72. Worrall, J. A. R.; Liu, Y. J.; Crowley, P. B.; Nocek, J. M.; Hoffman, B. M.; Ubbink, M. *Biochemistry* **2002**, *41*, 11721-11730.
73. Worrall, J. A. R.; Reinle, W. G.; Bernhardt, R.; Ubbink, M. *Biochemistry* **2003**, *42*, 7068-7076.
74. Worrall, J. A. R.; Kolczak, U.; Canters, G. W.; Ubbink, M. *Biochemistry* **2001**, *40*, 7069-7076.
75. Iwahara, J.; Anderson, D. E.; Murphy, E. C.; Clore, G. M. *J. Am. Chem. Soc.* **2003**, *125*, 6634-6635.
76. Prudencio, M.; Rohovec, J.; Peters, J. A.; Tocheva, E.; Boulanger, M. J.; Murphy, M. E. P.; Hupkes, H. J.; Kusters, W.; Impagliazzo, A.; Ubbink, M. *Chemistry-A European Journal* **2004**, *10*, 3252-3260.
77. Iwahara, J.; Schwieters, C. D.; Clore, G. M. *J. Am. Chem. Soc.* **2004**, *126*, 12800-12808.
78. Vlasie, M. D.; Comuzzi, C.; van den Nieuwendijk, A. M. C. H.; Prudencio, M.; Overhand, M.; Ubbink, M. *Chem- Eur. J.* **2007**, *13*, 1715-1723.
79. Keizers, P. H. J.; Desreux, J. F.; Overhand, M.; Ubbink, M. *J. Am. Chem. Soc.* **2007**, *129*, 9292-9293.
80. Gillespie, J. R.; Shortle, D. *J. Mol. Biol.* **1997**, *268*, 158-169.
81. Battiste, J. L.; Wagner, G. *Biochemistry* **2000**, *39*, 5355-5365.
82. Dedmon, M. M.; Lindorff-Larsen, K.; Christodoulou, J.; Vendruscolo, M.; Dobson, C. M. *J. Am. Chem. Soc.* **2005**, *127*, 476-477.
83. Liang, B.; Bushweller, J. H.; Tamm, L. K. *J. Am. Chem. Soc.* **2006**, *128*, 4389-4397.
84. Clore, G. M.; Iwahara, J. *Chem. Rev.* **2009**, *109*, 4108-4139.
85. Otting, G. *Annu. Rev. Biophys.* **2010**, *39*, 387-405.
86. Otting, G. *J. Biomol. NMR* **2008**, *42*, 1-9.
87. Keizers, P. H. J.; Mersinli, B.; Reinle, W. G.; Donauer, J.; Hiruma, Y.; Hannemann, F.; Overhand, M.; Bernhardt, R.; Ubbink, M. *Biochemistry* **2010**, *49*, 6846-6855.
88. Koehler, J.; Meiler, J. *Prog. Nucl. Magn. Res. Sp.* **2011**, *59*, 360-389.
89. Schmitz, C.; Vernon, R.; Otting, G.; Baker, D.; Huber, T. *J. Mol. Biol.* **2012**, *416*, 668-677.
90. Pintacuda, G.; Park, A. Y.; Keniry, M. A.; Dixon, N. E.; Otting, G. *J. Am. Chem. Soc.* **2006**, *128*, 3696-3702.

91. Schmitz, C.; Stanton-Cook, M. J.; Su, X. C.; Otting, G.; Huber, T. *J. Biomol. NMR* **2008**, *41*, 179-189.
92. Tang, C.; Iwahara, J.; Clore, G. M. *Nature* **2006**, *444*, 383-386.
93. Tang, C.; Ghirlando, R.; Clore, G. M. *J. Am. Chem. Soc.* **2008**, *130*, 4048-4056.
94. Ahmad, A.; Bhattacharya, A.; McDonald, R. A.; Cordes, M.; Ellington, B.; Bertelsen, E. B.; Zuiderweg, E. R. *Proc. Natl. Acad. Sci. U. S. A* **2011**, *108*, 18966-18971.
95. Zhuang, T.; Vishnivetskiy, S. A.; Gurevich, V. V.; Sanders, C. R. *Biochemistry* **2010**, *49*, 10473-10485.
96. Iwahara, J.; Clore, G. M. *Nature* **2006**, *440*, 1227-1230.
97. Iwahara, J.; Clore, G. M. *Nature* **2006**, *440*, 1227-1230.
98. Diaz-Moreno, I.; Diaz-Quintana, A.; De la Rosa, M. A.; Ubbink, M. *J. Biol. Chem.* **2005**, *280*, 35784.
99. Gabdouliline, R. R.; Wade, R. C. *J. Mol. Recognit.* **1999**, *12*, 226-234.
100. Spaar, A.; Dammer, C.; Gabdouliline, R. R.; Wade, R. C.; Helms, V. *Biophys. J.* **2006**, *90*, 1913-1924.
101. Anthi, N. J.; Doucleff, M.; Clore, G. M. *J. Am. Chem. Soc.* **2011**, *133*, 18966-18974.
102. Tang, C.; Schwieters, C. D.; Clore, G. M. *Nature* **2007**, *449*, 1078-1082.
103. Takayama, Y.; Clore, G. M. *Proc. Natl. Acad. Sci. U. S. A* **2011**, *108*, E169-E176.
104. Takayama, Y.; Clore, G. M. *J. Biol. Chem.* **2012**, *287*, 14349-14363.
105. Suh, J. Y.; Tang, C.; Clore, G. M. *J. Am. Chem. Soc.* **2007**, *129*, 12954-12955.
106. Gross, E. L.; Pearson, D. C., Jr. *Biophys. J.* **2003**, *85*, 2055-2068.
107. Gross, E. L.; Rosenberg, I. *Biophys. J.* **2006**, *90*, 366-380.
108. Gross, E. L. *Photosynth. Res.* **2007**, *94*, 411-422.
109. Northrup, S. H.; Boles, J. O.; Reynolds, J. C. *Science* **1988**, *241*, 67-70.
110. Pelletier, H.; Kraut, J. *Science* **1992**, *258*, 1748-1755.
111. Schreiber, G.; Haran, G.; Zhou, H. X. *Chem Rev* **2009**, *109*, 839-860.
112. Nocek, J. M.; Knutson, A. K.; Xiong, P.; Co, N. P.; Hoffman, B. M. *J. Am. Chem. Soc.* **2010**, *132*, 6165-6175.
113. Clore, G. M. *Protein Sci.* **2011**, *20*, 229-246.
114. Volkov, A. N.; Worrall, J. A. R.; Holtzmann, E.; Ubbink, M. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 18945-18950.

115. Ubbink, M.; Ejdebäck, M.; Karlsson, B. G.; Bendall, D. S. *Structure* **1998**, *6*, 323-335.
116. Lange, C.; Cornvik, T.; Diaz-Moreno, I.; Ubbink, M. *Biochim. Biophys. Acta-Bioenerg.* **2005**, *1707*, 179-188.
117. Sato, K.; Kohzuma, T.; Dennison, C. *J. Am. Chem. Soc.* **2003**, *125*, 2101-2112.
118. Schlarb-Ridley, B. G.; Bendall, D. S.; Howe, C. J. *Biochemistry* **2002**, *41*, 3279-3285.
119. Hart, S. E.; Schlarb-Ridley, B. G.; Delon, C.; Bendall, D. S.; Howe, C. J. *Biochemistry* **2003**, *42*, 4829-4836.
120. Diaz-Moreno, I.; Diaz-Quintana, A.; De la Rosa, M. A.; Ubbink, M. *J. Biol. Chem.* **2005**, *280*, 18908-18915.
121. Albarran, C.; Navarro, J. A.; Molina-Heredia, F. P.; Murdoch, P. S.; De la Rosa, M. A.; Hervas, M. *Biochemistry* **2005**, *44*, 11601-11607.
122. Albarran, C.; Navarro, J. A.; De la Rosa, M. A.; Hervas, M. *Biochemistry* **2007**, *46*, 997-1003.
123. Milikisyants, S.; Scarpelli, F.; Finiguerra, M. G.; Ubbink, M.; Huber, M. *J. Magn. Reson.* **2009**, *201*, 48-56.
124. Arslan, E.; Schulz, H.; Zufferey, R.; Kunzler, P.; Thony-Meyer, L. *Biochem. Biophys. Res. Commun.* **1998**, *251*, 744-747.
125. Delaglio, F.; Grzesiek, S.; Vuister, G. W.; Zhu, G.; Pfeifer, J.; Bax, A. *J. Biomol. NMR* **1995**, *6*, 277-293.
126. Vranken, W. F.; Boucher, W.; Stevens, T. J.; Fogh, R. H.; Pajon, A.; Llinas, M.; Ulrich, E. L.; Markley, J. L.; Ionides, J.; Laue, E. D. *Proteins* **2005**, *59*, 687-696.
127. Kannt, A.; Young, S.; Bendall, D. S. *Biochim. Biophys. Acta-Bioenerg.* **1996**, *1277*, 115-126.
128. Battiste, J. L.; Wagner, G. *Biochemistry* **2000**, *39*, 5355-5365.
129. de la Torre, J. G.; Huertas, M. L.; Carrasco, B. *J. Magn. Reson.* **2000**, *147*, 138-146.
130. Schwieters, C. D.; Kuszewski, J. J.; Tjandra, N.; Clore, G. M. *J. Magn. Reson.* **2003**, *160*, 65-73.
131. Baniulis, D.; Yamashita, E.; Whitelegge, J. P.; Zatsman, A. I.; Hendrich, M. P.; Hasan, S. S.; Ryan, C. M.; Cramer, W. A. *J. Biol. Chem.* **2009**, *284*, 9861-9869.
132. Iwahara, J.; Schwieters, C. D.; Clore, G. M. *J. Am. Chem. Soc.* **2004**, *126*, 5879-5896.
133. Diaz-Moreno, I.; Diaz-Quintana, A.; De la Rosa, M. A.; Ubbink, M. *J. Biol. Chem.* **2005**, *280*, 18908-18915.
134. Ubbink, M.; Lian, L. Y.; Modi, S.; Evans, P. A.; Bendall, D. S. *Eur. J. Biochem.* **1996**, *242*, 132-147.
135. Diaz-Moreno, I.; Diaz-Quintana, A.; De la Rosa, M. A.; Crowley, P. B.; Ubbink, M. *Biochemistry* **2005**, *44*, 3176-3183.
136. Fawzi, N. L.; Doucleff, M.; Suh, J. Y.; Clore, G. M. *Proc. Natl. Acad. Sci. U. S. A* **2010**, *107*, 1379-1384.

137. Gabdouliline, R. R.; Wade, R. C. *Biophys. J.* **1997**, *72*, 1917-1929.
138. Camacho, C. J.; Weng, Z.; Vajda, S.; DeLisi, C. *Biophys. J.* **1999**, *76*, 1166-1178.
139. Camacho, C. J.; Vajda, S. *Proc. Natl. Acad. Sci. U. S. A* **2001**, *98*, 10636-10641.
140. Sugase, K.; Dyson, H. J.; Wright, P. E. *Nature* **2007**, *447*, 1021-1025.
141. Tang, C.; Iwahara, J.; Clore, G. M. *Nature* **2006**, *444*, 383-386.
142. Suh, J. Y.; Tang, C.; Clore, G. M. *J. Am. Chem. Soc.* **2007**, *129*, 12954-12955.
143. Milikisyants, S.; Scarpelli, F.; Finiguerra, M. G.; Ubbink, M.; Huber, M. *J. Magn. Reson.* **2009**, *201*, 48-56.
144. Arslan, E.; Schulz, H.; Zufferey, R.; Kunzler, P.; Thony-Meyer, L. *Biochem. Biophys. Res. Commun.* **1998**, *251*, 744-747.
145. Delaglio, F.; Grzesiek, S.; Vuister, G. W.; Zhu, G.; Pfeifer, J.; Bax, A. *J. Biomol. NMR* **1995**, *6*, 277-293.
146. Vranken, W. F.; Boucher, W.; Stevens, T. J.; Fogh, R. H.; Pajon, A.; Llinas, M.; Ulrich, E. L.; Markley, J. L.; Ionides, J.; Laue, E. D. *Proteins* **2005**, *59*, 687-696.
147. Baniulis, D.; Yamashita, E.; Whitelegge, J. P.; Zatsman, A. I.; Hendrich, M. P.; Hasan, S. S.; Ryan, C. M.; Cramer, W. A. *J. Biol. Chem.* **2009**, *284*, 9861-9869.
148. Brunger, A. T.; Karplus, M. *Proteins* **1988**, *4*, 148-156.
149. Brooks, B. R.; Bruccoleri, R. E.; Olafson, B. D.; States, D. J.; Swaminathan, S.; Karplus, M. *J. Comput. Chem.* **1983**, *4*, 187-217.
150. MacKerell, A. D.; Bashford, D.; Bellott, Dunbrack, R. L.; Evanseck, J. D.; Field, M. J.; Fischer, S.; Gao, J.; Guo, H.; Ha, S.; Joseph-McCarthy, D.; Kuchnir, L.; Kuczera, K.; Lau, F. T. K.; Mattos, C.; Michnick, S.; Ngo, T.; Nguyen, D. T.; Prodhom, B.; Reiher, W. E.; Roux, B.; Schlenkrich, M.; Smith, J. C.; Stote, R.; Straub, J.; Watanabe, M.; Wiorkiewicz-Kuczera, J.; Yin, D.; Karplus, M. *J. Phys. Chem. B* **1998**, *102*, 3586-3616.
151. Baker, N. A.; Sept, D.; Joseph, S.; Holst, M. J.; McCammon, J. A. *Proc. Natl. Acad. Sci. U. S. A* **2001**, *98*, 10037-10041.
152. Metropolis, N.; Ulam S. *J. Am. Stat. Assoc.* **1949**, *44*, 335-341.
153. García de la Torre, J.; Huertas, M. L.; Carrasco, B. *J. Magn. Reson.* **2000**, *147*, 138-146.
154. Schwieters, C. D.; Kuszewski, J. J.; Tjandra, N.; Clore, G. M. *J. Magn Reson.* **2003**, *160*, 65-73.
155. Ubbink, M.; Lian, L. Y.; Modi, S.; Evans, P. A.; Bendall, D. S. *Eur. J. Biochem.* **1996**, *242*, 132-147.
156. Tang, C.; Schwieters, C. D.; Clore, G. M. *Nature* **2007**, *449*, 1078-1082.
157. Tang, C.; Ghirlando, R.; Clore, G. M. *J. Am. Chem. Soc.* **2008**, *130*, 4048-4056.
158. Tang, C.; Louis, J. M.; Aniana, A.; Suh, J. Y.; Clore, G. M. *Nature* **2008**, *455*, 693-696.

159. Fawzi, N. L.; Doucleff, M.; Suh, J. Y.; Clore, G. M. *Proc. Natl. Acad. Sci. U. S. A* **2010**, *107*, 1379-1384.
160. Anthis, N. J.; Doucleff, M.; Clore, G. M. *J. Am. Chem. Soc.* **2011**, *133*, 18966-18974.
161. Volkov, A. N.; Ubbink, M.; van Nuland, N. A. J. *J. Biomol. NMR* **2010**, *48*, 225-236.
162. Moser, C. C.; Keske, J. M.; Warncke, K.; Farid, R. S.; Dutton, P. L. *Nature* **1992**, *355*, 796-802.
163. Gabdouliline, R. R.; Wade, R. C. *J. Mol. Biol.* **2001**, *306*, 1139-1155.
164. Hasan, S. S.; Cramer, W. A. *Phys. Chem. Chem. Phys.* **2012**, *14*, 13853-13860.
165. Harel, M.; Cohen, M.; Schreiber, G. *J. Mol. Biol.* **2007**, *371*, 180-196.
166. Liang, Z. X.; Nocek, J. M.; Huang, K.; Hayes, R. T.; Kurnikov, I. V.; Beratan, D. N.; Hoffman, B. M. *J. Am. Chem. Soc.* **2002**, *124*, 6849-6859.
167. Hope, A. B. *Biochim. Biophys. Acta* **2000**, *1456*, 5-26.
168. Diaz-Quintana, A.; Hervás, M.; Navarro, J.A.; De la Rosa, M. A. *Photosynthetic protein complexes: A structural approach.* (Fromme P. Ed.) Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim. **2008**, 181-200.
169. Camacho, C. J.; Weng, Z.; Vajda, S.; DeLisi, C. *Biophys J* **1999**, *76*, 1166-1178.
170. Camacho, C. J.; Kimura, S. R.; DeLisi, C.; Vajda, S. *Biophys. J.* **2000**, *78*, 1094-1105.
171. Kim, Y. C.; Tang, C.; Clore, G. M.; Hummer, G. *Proc. Natl. Acad. Sci. U. S. A* **2008**, *105*, 12855-12860.
172. Musiani, F.; Dikiy, A.; Semenov, A.Y.; Ciurli, S. *J. Biol. Chem.* **2005**, *280*, 18833-18841.
173. Schlarb, B. G.; Wagner, M. J.; Vijgenboom, E.; Ubbink, M.; Bendall, D. S.; Howe, C. J. *Gene* **1999**, *234*, 275-283.
174. Banci, L.; Bertini, I.; Cavallaro, G.; Giachetti, A.; Luchinat, C.; Parigi, G. *J. Biomol. NMR* **2004**, *28*, 249-261.
175. Schwieters, C. D.; Kuszewski, J. J.; Tjandra, N.; Clore, G. M. *J. Magn Reson.* **2003**, *160*, 65-73.
176. Kasha, M. *J. Chem. Phys.* **1952**, *20*, 71-74.
177. Marcus, R. A. *J. Chem. Phys.* **1956**, *24*, 966-978.
178. Marcus, R. A.; Sutin, N. *Biochim. Biophys. Acta* **1985**, *811*, 265-322.
179. Northrup, S. H.; Boles, J. O.; Reynolds, J. C. L. *J. Phys. Chem.* **1987**, *91*, 5991-5998.
180. Bertini, I.; Luchinat, C.; Parigi, G.; Pierattelli, R. *Chembiochem.* **2005**, *6*, 1536-1549.
181. Clore, G. M.; Iwahara, J. *Chem. Rev.* **2009**, *109*, 4108-4139.
182. Clore, G. M.; Schwieters, C. D. *Curr. Opin. Struct. Biol.* **2002**, *12*, 146-153.

183. Gabdoulline, R. R.; Wade, R. C. *Methods* **1998**, *14*, 329-341.
184. Gabdoulline, R. R.; Wade, R. C. *Curr. Opin. Struct. Biol.* **2002**, *12*, 204-213.
185. Smith, G. R.; Sternberg, M. J. *Curr. Opin. Struct. Biol.* **2002**, *12*, 28-35.
186. Dominguez, C.; Boelens, R.; Bonvin, A. M. J. J. *J. Am. Chem. Soc.* **2003**, *125*, 1731-1737.
187. Wider, G. *Nuclear Magnetic Resonance of Biological Macromolecules, Part C* **2005**, 394, 382-398.
188. Kay, L. E. *J. Magn Reson.* **2011**, *210*, 159-170.
189. Janin, J.; Henrick, K.; Moulf, J.; Eyck, L. T.; Sternberg, M. J.; Vajda, S.; Vakser, I.; Wodak, S. J. *Proteins: Struct. Funct. Genet.* **2003**, *52*, 2-9.
190. Comeau, S. R.; Gatchell, D. W.; Vajda, S.; Camacho, C. J. *Bioinformatics.* **2004**, *20*, 45-50.

Appendices

Appendix 1

Input file for rigid-body docking of Cyt *f*-Pc using PRE restraints for Xplor-NIH 2.9.9

```
parameter
  @nostoc_cytf_SL_pc.par
end
structure
  @nostoc_cytf_SL_pc.psf
end
eval ($infile="nostoc_cytf_SL_pc.pdb")
coordinates @$infile
@learn.pcf.par
constraints fix (segid="CYTF" or resn SL) end
eval ($a05 = 6000)           !total nr cycles / $a53
eval ($a53 = 1000)         !nr of steps
eval ($u2 = 30)             !nr of cycles per docking approach
eval ($a54 = 0.01)         !fimestep in ps
eval ($a61 = $a54*$a53)    !time per cycle (ps)
eval ($a80 = $a61*$a05)    !total time (ps)
eval ($a58=30.0)           !velocity factor
eval ($a56=1.5)            !fbeta
eval ($a55=300.0)          !TBATH
eval ($a59=45.0)           !.pdb writing threshold
eval ($van=100.0)          ! vdw writing threshold
eval ($a59a=0.01*$a59)     !vx-reset minimum
eval ($a60=1.0)            !.pdb lag factor           15
eval ($a301=1.0)           !VDW repel scale factor
eval ($a93=1)              !time-spent factor: 0<$a93<1 (fraction bound)
eval ($g1a = "N71C")       !names of the spin label 1 used in this calculation
eval ($g1b = "Q104C")     !names of the spin label 2 used in this calculation
eval ($g1c = "S192C")     !names of the spin label 3 used in this calculation
eval ($g2a = 13)           !residue number of the 1st SL conformation of SL 1
eval ($g2b = 17)           !residue number of the 1st SL conformation of SL 2
eval ($g2c = 21)           !residue number of the 1st SL conformation of SL 3
eval ($field=600.1328E6)   !field strength in Hz
eval ($frl=1)              !fraction of proteins with spin label
eval ($tau_c=30.0E-9)      !tau c for the complex in sec
eval ($a9=0.01)           !general scaling PRE (NOE) term
eval ($a10=1.0)            !scale factor for CL1 (peaks disappeared)
eval ($a11=1.0)            !scale factor for CL2 (peaks unaffected)
eval ($a12=1.0)            !scale factor for CL3 (peaks reduced)
eval ($low1=14.0)          !lower(d_minus) limit (SLA restraints upper limit only)
eval ($up1=4.0)           !upper(d_plus) limit (restraints with upper limit only)
eval ($low2=4.0)          !lower(d_minus) limit (restraints with lower limit only)
eval ($up2=100.0)         !upper(d_plus) limit (restraints with lower limit only)
eval ($low3=4.0)          !lower(d_minus) limit (restraints with both limits)
eval ($up3=4.0)           !upper(d_plus) limit (restraints with both limits)
eval ($a01 = 1)           !cycle counter
eval ($a48=$cpu*1e4)
set seed=$a48 end
eval ($a14=0)
eval ($a18=1)
eval ($nout1=0)           ! Number of structures output per run
eval ($min1=9999.0)
eval ($min2=9999.0)
eval ($ref=0)
set display=coord.dat end !write parameters
display xx -----PARAMETERS-----
display xx startdate:    $DATE
display xx starttime:    $TIME
display xx ini.file:     $infile
display xx timestep (ps): $a54
```

```

display xx nr of steps:   $a53           time /cycle (ps): $a61
display xx nr of cycles: $a05           total time (ps): $a80
display xx time-spent fact: $a93
display xx general scale: $a9
display xx TBATH:       $a55  velocity factor: $a56  fbeta: $a58
display xx .pdb threshold $a59
display xx .pdb lag factor $a60
display xx Local mimim. impulse after 10 cycles with 'constant' Etot > $a59a
display xx -----
display
set display=OUTPUT end
flag exclude elec bond angl dihe impr include vdw noe end
vector do (fbeta=$a58) (segid="PC")
vector do (vx=$a56) (segid="PC")
vector do (vy=$a56) (segid="PC")
vector do (vz=$a56) (segid="PC")
set disp=ener.dat end
display Energies for the output structures
display file          Etot      VDW      NOE
display -----
set disp=OUTPUT end
@restraints.xpl
eval ($a213=100*(rand()-0.5) ) ! push away PC
eval ($a214=100*(rand()-0.5) )
eval ($a215=100*(rand()-0.5) )
vector do (x=x+$a213) (segid="PC")
vector do (y=y+$a214) (segid="PC")
vector do (z=z+$a215) (segid="PC")
parameter
  @nbfix.4sl_expl.xpl
  nbonds
    cutnb=8.5
    inhi=0.25
    ctofnb=7.5
    ctonnb=6.5
    repe1=0.6 !0.6
    NBXMod=-2
    rexp=2
    irex=2
    rcon=$a301
    wmin=1.5
  end
end
constraints
  interactions (segid="CYTF")(segid="PC")
end
energy end
while ($a01 LE $a05) loop calc
display cycle $a01
dynamics rigid
  dt=$a54
  group=(segid="PC")
  dynmode=TCOU
  tbath=$a55
  nprint=500
  nstep=$a53
  NTRFRQ=0 !new for XPLOR vs 3.8
end
eval ($a14=$ENER)
if ($a14 < $min2) then
  coor copy end
  eval ($min2 = $a14)
  eval ($abc=$VDW)
  eval ($a15=$NOE)

```

```

eval ($a16=$a01)
eval ($a17=$a18)
end if
if ($a18 = $u2) then           !counting number of dockings
if ($min2 < $a59) then
if ($abc < $van) then
    coor swap end
    eval ($nout1=$nout1+1)
    eval ($pdb="structure_"+encode($nout1)+".pdb")
    write coord output=$pdb end
    if ($a14<$min1) then
        eval ($ref=$nout1)
    end if
    eval ($min1 = $a14)
end if
pick bond (segid="CYTF" and name FE) (segid="PC" and name CU) geom
eval ($a50=$RESULT)
set disp=ener.dat end
display $nout1 $min2 $abc $a15
set disp=OUTPUT end
set disp=coor.dat end
display Cycle: $a16 Dock step: $a17 File#: $nout1 Fe-Cu: $a50 temp: $TEMP
display Etot: $min2 Evdw: $abc Enoe: $a15
display -----
set disp=OUTPUT end
!violation analysis
eval ($b3="structure_"+encode($nout1)+".viol.dat")
set display=$b3 end
display Violation analysis PRE data
display res# PRE(meas) PRE(calc) Dist(calc)
display
for $g2 in ($g2a $g2b $g2c)
loop spinlabel
eval ($g2d=$g2)
eval ($g2e=$g2+1)
eval ($g2f=$g2+2)
eval ($g2g=$g2+3)
if ($g2=$g2a) then
eval ($g1h = $g1a)
display Spin Label $g1h
for
    $c02 in ID (store1)
loop C3
vector show elem (resi) (ID $c02)
eval ($c04=$RESULT) !residue nr
vector show elem (store1) (ID $c02)
eval ($c08=$RESULT)
pick bond (resn SL and resi $g2d and name O) (ID $c02) geom
eval ($ca1=$RESULT)
pick bond (resn SL and resi $g2e and name O) (ID $c02) geom
eval ($ca2=$RESULT)
pick bond (resn SL and resi $g2f and name O) (ID $c02) geom
eval ($ca3=$RESULT)
pick bond (resn SL and resi $g2g and name O) (ID $c02) geom
eval ($ca4=$RESULT)
eval ($c03=($ca1^(-6)+$ca2^(-6)+$ca3^(-6)+$ca4^(-6))/4)
!r-6 average distance over 4 positions
eval ($c05=(1.23E16*(4*$tau_c+(3*$tau_c/(1+($field*2*3.14*$tau_c)^2)))*$frl*$a93*$c03) )
!back calculated PRE
eval ($c10=$c03^(-1/6))
display $c04 $c08 $c05 $c10
end loop C3
display
end if
if ($g2=$g2b) then

```



```

eval ($g1h = $g1b)
display Spin Label $g1h
for
    $c02 in ID (store2)
loop C3
    vector show elem (resi) (ID $c02)
    eval ($c04=$RESULT) !residue nr
    vector show elem (store2) (ID $c02)
    eval ($c08=$RESULT)
    pick bond (resn SL and resi $g2d and name O) (ID $c02) geom
    eval ($ca1=$RESULT)
    pick bond (resn SL and resi $g2e and name O) (ID $c02) geom
    eval ($ca2=$RESULT)
    pick bond (resn SL and resi $g2f and name O) (ID $c02) geom
    eval ($ca3=$RESULT)
    pick bond (resn SL and resi $g2g and name O) (ID $c02) geom
    eval ($ca4=$RESULT)
    eval ($c03=($ca1^(-6)+$ca2^(-6)+$ca3^(-6)+$ca4^(-6))/4)
    !r-6 average distance over 4 positions
    eval ($c05=(1.23E16*(4*$tau_c+(3*$tau_c/(1+($field*2*3.14*$tau_c)^2)))*$fri*$a93*$c03) )
    !back calculated PRE
    eval ($c10=$c03^(-1/6))
    display $c04 $c08 $c05 $c10
end loop C3
display
end if
if ($g2=$g2c) then
    eval ($g1h = $g1c)
    display Spin Label $g1h
    for
        $c02 in ID (store3)
    loop C3
        vector show elem (resi) (ID $c02)
        eval ($c04=$RESULT) !residue nr
        vector show elem (store3) (ID $c02)
        eval ($c08=$RESULT)
        pick bond (resn SL and resi $g2d and name O) (ID $c02) geom
        eval ($ca1=$RESULT)
        pick bond (resn SL and resi $g2e and name O) (ID $c02) geom
        eval ($ca2=$RESULT)
        pick bond (resn SL and resi $g2f and name O) (ID $c02) geom
        eval ($ca3=$RESULT)
        pick bond (resn SL and resi $g2g and name O) (ID $c02) geom
        eval ($ca4=$RESULT)
        eval ($c03=($ca1^(-6)+$ca2^(-6)+$ca3^(-6)+$ca4^(-6))/4)
        !r-6 average distance over 4 positions
        eval ($c05=(1.23E16*(4*$tau_c+(3*$tau_c/(1+($field*2*3.14*$tau_c)^2)))*$fri*$a93*$c03) )
        !back calculated PRE
        eval ($c10=$c03^(-1/6))
        display $c04 $c08 $c05 $c10
    end loop C3
    display
end if
end loop spinlabel
close $b3 end
set display=OUTPUT end
!end violation analysis
coor swap end
end if
end if
!impulse to escape local minimum
eval ($a213=100*(rand()-0.5) )
eval ($a214=100*(rand()-0.5) )
eval ($a215=100*(rand()-0.5) )

```

```

vector do (x=x+$a213) (segid="PC")
vector do (y=y+$a214) (segid="PC")
vector do (z=z+$a215) (segid="PC")
vector do (vx=50) (segid="PC")
vector do (vy=50) (segid="PC")
vector do (vz=50) (segid="PC")
set disp=coor.dat end
display cycle $a01: impulse: $a213, $a214, $a215 to x,y,z; v=50
set disp=OUTPUT end
eval ($a18=0)
eval ($min2=9999.0)
end if
eval ($a18=$a18+1)
if ($TEMP>200000.0) then                !correct excessive temperature
  eval ($a45=RAND()+0.01)
  vector do (vx=$a56*$a45*rand()) (segid="PC")
  vector do (vy=$a56*$a45*rand()) (segid="PC")
  vector do (vz=$a56*$a45*rand()) (segid="PC")
end if
eval ($a01 = $a01 + 1)
end loop calc
if ($ref > 0) then
vector idend ( store9 ) ( name ca or name n or name c )      !backbone selection
eval ($ref_file="structure_"+encode($ref)+".pdb")
set display=rms.dat end
display Backbone pairwise RMSD from the lowest energy strucutre ($ref_file)
display file      rmsd
display
set disp=OUTPUT end
coor swap end
coor init end
coor swap end
coor disp=comp @@$ref_file
eval ($count1=0)
while ($count1 < $nout1) loop fill
evaluate ($count1=$count1+1)
  evaluate ($file="structure_"+encode($count1)+".pdb")
  coor init end
  coor @@$file
  coor sele=(recall 9) fit end
  coor sele=(recall 9) rms end
  eval ($b1 = $result)
set display=rms.dat end
display $file      $b1
end loop fill
end if
set display=OUTPUT end
set echo=true end
stop

```

Appendix 2

Input file to determine distances from Cyt *f*-SL to Pc amide protons for Xplor-NIH 2.9.9

```
parameter
  @nostoc_cytf_SL_pc.par
end
structure
  @nostoc_cytf_SL_pc.psf
end
eval ($inifile="nostoc_cytf_SL_pc.pdb")
coordinates @$inifile
@learn.pcf.par
constraints fix (segid="CYTF" or resn SL) end
eval ($a05 = 6000)                !total nr cycles / $a53
eval ($a53 = 1000)                !nr of steps
eval ($u2 = 30)                   !nr of cycles per docking approach
eval ($a54 = 0.01)                !timestep in ps
eval ($a61 = $a54*$a53)           !time per cycle (ps)
eval ($a80 = $a61*$a05)           !total time (ps)
eval ($a58=30.0)                  !velocity factor
eval ($a56=1.5)                   !fbeta
eval ($a55=300.0)                 !TBATH
eval ($a59=45.0)                  !.pdb writing threshold
eval ($van=100.0)                 ! vdw writing threshold
eval ($a59a=0.01*$a59)            !vx-reset minimum
eval ($a60=1.0)                   !.pdb lag factor
eval ($a301=1.0)                  !VDW repel scale factor
eval ($a93=1)                     !time-spent factor: 0<$a93<1 (fraction bound)
eval ($g1a = "N71C")              !names of the spin label 1 used in calculation
eval ($g1b = "Q104C")             !names of the spin label 2 used in calculation
eval ($g1c = "S192C")             !names of the spin label 3 used in calculation
eval ($g2a = 13)                  !residue number of the 1st SL conformation of SL 1
eval ($g2b = 17)                  !residue number of the 1st SL conformation of SL 2
eval ($g2c = 21)                  !residue number of the 1st SL conformation of SL 3
eval ($field=600.1328E6)           !field strength in Hz
eval ($frl=1)                     !fraction of proteins with spin label
eval ($tau_c=30.0E-9)              !tau c for the complex in sec
eval ($a9=0.01)                   !general scaling PRE (NOE) term
eval ($a10=1.0)                   !scale factor for CL1 (peaks disappeared)
eval ($a11=1.0)                   !scale factor for CL2 (peaks unaffected)
eval ($a12=1.0)                   !scale factor for CL3 (peaks reduced)
eval ($low1=14.0)                 !lower(d_minus) limit
eval ($up1=4.0)                   !upper(d_plus) limit
eval ($low2=4.0)                  !lower(d_minus) limit
eval ($up2=100.0)                 !upper(d_plus) limit
eval ($low3=4.0)                  !lower(d_minus) limit
eval ($up3=4.0)                   !upper(d_plus) limit
eval ($a01 = 1)                   !cycle counter
eval ($a48=$cpu*1e4)
set seed=$a48 end
eval ($a14=0)
eval ($a18=1)
eval ($nout1=0)                   ! Number of structures output per run
eval ($min1=9999.0)
eval ($min2=9999.0)
eval ($ref=0)
set display=coord.dat end         !write parameters
display xx -----PARAMETERS-----
display xx startdate:            $DATE
```

```

display xx starttime:      $TIME
display xx ini.file:      $inifile
display xx timestep (ps): $a54
display xx nr of steps:   $a53           time /cycle (ps): $a61
display xx nr of cycles:  $a05           total time (ps): $a80
display xx time-spent fact: $a93
display xx general scale:  $a9
display xx TBATH:         $a55 velocity factor: $a56 fbeta: $a58
display xx .pdb threshold $a59
display xx .pdb lag factor $a60
display xx Local mimim. impulse after 10 cycles with 'constant' Etot > $a59a
display xx -----
display
set display=OUTPUT end
flag exclude elec bond angl dihe impr include vdw noe end
vector do (fbeta=$a58) (segid="PC")
vector do (vx=$a56) (segid="PC")
vector do (vy=$a56) (segid="PC")
vector do (vz=$a56) (segid="PC")
set disp=ener.dat end
display Energies for the output structures
display file      Etot      VDW      NOE
display -----
set disp=OUTPUT end
@restraints.xpl
eval ($a213=100*(rand()-0.5) ) ! push away PC
eval ($a214=100*(rand()-0.5) )
eval ($a215=100*(rand()-0.5) )
vector do (x=x+$a213) (segid="PC")
vector do (y=y+$a214) (segid="PC")
vector do (z=z+$a215) (segid="PC")
!write coor OUTPUT="test1.pdb" end
parameter
  @nbfix.4sl_expl.xpl
  nbonds
    cutnb=8.5
    inhi=0.25
    ctofnb=7.5
    ctonnb=6.5
    repe1=0.6 !0.6
                NBXMod=-2
    rexp=2
    irex=2
    rcon=$a301
    wmin=1.5
  end
end
constraints
  interactions (segid="CYTF")(segid="PC")
end
energy end
while ($a01 LE $a05) loop calc
display cycle $a01
dynamics rigid
  dt=$a54
  group=(segid="PC")
  dynmode=TCOU
  tbath=$a55
  nprint=500
  nstep=$a53
  NTRFRQ=0 !new for XPLOr vs 3.8
end
eval ($a14=$ENER)
if ($a14 < $min2) then

```

```

    coor copy end
    eval ($min2 = $a14)
    eval ($abc=$VDW)
    eval ($a15=$NOE)
    eval ($a16=$a01)
    eval ($a17=$a18)
end if
if ($a18 = $u2) then           !counting number of dockings
if ($min2 < $a59) then
if ($abc < $van) then
    coor swap end
    eval ($nout1=$nout1+1)
    eval ($pdb="structure_"+encode($nout1)+".pdb")
    write coord output=$pdb end
    if ($a14<$min1) then
        eval ($ref=$nout1)
    eval ($min1 = $a14)
    end if
    pick bond (segid="CYTF" and name FE) (segid="PC" and name CU) geom
    eval ($a50=$RESULT)
    set disp=ener.dat end
    display $nout1 $min2      $abc      $a15
    set disp=OUTPUT end
    set disp=coor.dat end
    display Cycle: $a16      Dock step: $a17      File#: $nout1      Fe-Cu: $a50      temp: $TEMP
    display Etot:  $min2      Evdw: $abc      Enoe: $a15
    display -----
        set disp=OUTPUT end
!violation analysis
    eval ($b3="structure_"+encode($nout1)+".viol.dat")
    set display=$b3 end
    display Violation analysis PRE data
    display res#  PRE(meas) PRE(calc) Dist(calc)
    display
    for $g2 in ($g2a $g2b $g2c)
    loop spinlabel
        eval ($g2d=$g2)
        eval ($g2e=$g2+1)
        eval ($g2f=$g2+2)
        eval ($g2g=$g2+3)
        if ($g2=$g2a) then
            eval ($g1h = $g1a)
            display Spin Label $g1h
            for
                $c02 in ID (store1)
            loop C3
                vector show elem (resi) (ID $c02)
                eval ($c04=$RESULT)           !residue nr
                vector show elem (store1) (ID $c02)
                eval ($c08=$RESULT)
                pick bond (resn SL and resi $g2d and name O) (ID $c02) geom
                eval ($ca1=$RESULT)
                pick bond (resn SL and resi $g2e and name O) (ID $c02) geom
                eval ($ca2=$RESULT)
                pick bond (resn SL and resi $g2f and name O) (ID $c02) geom
                eval ($ca3=$RESULT)
                pick bond (resn SL and resi $g2g and name O) (ID $c02) geom
                eval ($ca4=$RESULT)
                eval ($c03=($ca1^(-6)+$ca2^(-6)+$ca3^(-6)+$ca4^(-6))/4)
                    !r-6 average distance over 4 positions
                eval ($c05=(1.23E16*(4*$tau_c+(3*$tau_c/(1+($field*2*3.14*$tau_c)^2)))*$fri*$a93*$c03) )
                    !back calculated PRE
                eval ($c10=$c03^(-1/6))
                display $c04      $c08      $c05      $c10
            end loop
        end for
    end for
end if
end if
end if

```

```

end loop C3
display
end if

if ($g2=$g2b) then
eval ($g1h = $g1b)
display Spin Label $g1h
for
    $c02 in ID (store2)
loop C3
vector show elem (resi) (ID $c02)
eval ($c04=$RESULT) !residue nr
vector show elem (store2) (ID $c02)
eval ($c08=$RESULT)
pick bond (resn SL and resi $g2d and name O) (ID $c02) geom
eval ($ca1=$RESULT)
pick bond (resn SL and resi $g2e and name O) (ID $c02) geom
eval ($ca2=$RESULT)
pick bond (resn SL and resi $g2f and name O) (ID $c02) geom
eval ($ca3=$RESULT)
pick bond (resn SL and resi $g2g and name O) (ID $c02) geom
eval ($ca4=$RESULT)
eval ($c03=($ca1^(-6)+$ca2^(-6)+$ca3^(-6)+$ca4^(-6))/4)
!r-6 average distance over 4 positions
eval ($c05=(1.23E16*(4*$tau_c+(3*$tau_c/(1+($field*2*3.14*$tau_c)^2)))*$fri*$a93*$c03) )
!back calculated PRE
eval ($c10=$c03^(-1/6))
display $c04 $c08 $c05 $c10
end loop C3
display
end if

if ($g2=$g2c) then
eval ($g1h = $g1c)
display Spin Label $g1h
for
    $c02 in ID (store3)
loop C3
vector show elem (resi) (ID $c02)
eval ($c04=$RESULT) !residue nr
vector show elem (store3) (ID $c02)
eval ($c08=$RESULT)
pick bond (resn SL and resi $g2d and name O) (ID $c02) geom
eval ($ca1=$RESULT)
pick bond (resn SL and resi $g2e and name O) (ID $c02) geom
eval ($ca2=$RESULT)
pick bond (resn SL and resi $g2f and name O) (ID $c02) geom
eval ($ca3=$RESULT)
pick bond (resn SL and resi $g2g and name O) (ID $c02) geom
eval ($ca4=$RESULT)
eval ($c03=($ca1^(-6)+$ca2^(-6)+$ca3^(-6)+$ca4^(-6))/4)
!r-6 average distance over 4 positions
eval ($c05=(1.23E16*(4*$tau_c+(3*$tau_c/(1+($field*2*3.14*$tau_c)^2)))*$fri*$a93*$c03) )
!back calculated PRE
eval ($c10=$c03^(-1/6))
display $c04 $c08 $c05 $c10
end loop C3
display
end if
end loop spinlabel
close $b3 end
set display=OUTPUT end
!end violation analysis
coor swap end
end if

```

```

end if
                                !impulse to escape local minimum
eval ($a213=100*(rand()-0.5) )
eval ($a214=100*(rand()-0.5) )
eval ($a215=100*(rand()-0.5) )
vector do (x=x+$a213) (segid="PC")
vector do (y=y+$a214) (segid="PC")
vector do (z=z+$a215) (segid="PC")
vector do (vx=50) (segid="PC")
vector do (vy=50) (segid="PC")
vector do (vz=50) (segid="PC")
set disp=coord.dat end
display cycle $a01: impulse: $a213, $a214, $a215 to x,y,z; v=50
set disp=OUTPUT end
eval ($a18=0)
eval ($min2=9999.0)
end if
eval ($a18=$a18+1)
if ($TEMP>200000.0) then          !correct excessive temperature
  eval ($a45=RAND()+0.01)
  vector do (vx=$a56*$a45*rand()) (segid="PC")
  vector do (vy=$a56*$a45*rand()) (segid="PC")
  vector do (vz=$a56*$a45*rand()) (segid="PC")
end if
eval ($a01 = $a01 + 1)
end loop calc
if ($ref > 0) then
vector idend ( store9 ) ( name ca or name n or name c )      !backbone selection
eval ($ref_file="structure_"+encode($ref)+".pdb")
set display=rms.dat end
display Backbone pairwise RMSD from the lowest energy strucutre ($ref_file)
display file      rmsd
display -----
set disp=OUTPUT end
  coor swap end
  coor init end
  coor swap end
  coor disp=comp @@$ref_file
eval ($count1=0)
while ($count1 < $nout1) loop fill
  evaluate ($count1=$count1+1)
    evaluate ($file="structure_"+encode($count1)+".pdb")
    coor init end
    coor @$file
    coor sele=(recall 9) fit end
    coor sele=(recall 9) rms end
    eval ($b1 = $result)
  set display=rms.dat end
  display $file      $b1
end loop fill
end if
set display=OUTPUT end
set echo=true end
stop

```

Appendix 3

Input file for ensemble docking of Cyt *f*-Pc using PRE restraints for Xplor-NIH 2.9.9

```
parameter
  @nostoc_cytf_SL_pc.par
end
structure
  @nostoc_cytf_SL_pc.psf
end
eval ($infile="nostoc_cytf_SL_pc.pdb")
coordinates @$infile
@learn.pcf.par
constraints fix (segid="CYTF" or resn SL) end
eval ($a05 = 6000)           !total nr cycles / $a53
eval ($a53 = 1000)         !nr of steps
eval ($u2 = 30)             !nr of cycles per docking approach
eval ($a54 = 0.01)         !fimestep in ps
eval ($a61 = $a54*$a53)    !time per cycle (ps)
eval ($a80 = $a61*$a05)    !total time (ps)
eval ($a58=30.0)           !velocity factor
eval ($a56=1.5)            !fbeta
eval ($a55=300.0)          !TBATH
eval ($a59=45.0)           !.pdb writing threshold
eval ($van=100.0)          ! vdW writing threshold
eval ($a59a=0.01*$a59)     !vx-reset minimum
eval ($a60=1.0)            !.pdb lag factor
eval ($a301=1.0)           !VDW repel scale factor
eval ($a93=1)              !time-spent factor: 0<$a93<1 (fraction bound)
eval ($g1a = "N71C")       !names of the spin label 1
eval ($g1b = "Q104C")      !names of the spin label 2
eval ($g1c = "S192C")      !names of the spin label 3
eval ($g2a = 13)           !residue number of the 1stSL conformation of SL 1
eval ($g2b = 17)           !residue number of the 1st SL conformation of SL 2
eval ($g2c = 21)           !residue number of the 1st SL conformation of SL 3
eval ($field=600.1328E6)   !field strength in Hz
eval ($frr=1)              !fraction of proteins with spin label
eval ($tau_c=30.0E-9)      !tau c for the complex in sec
eval ($a9=0.01)           !general scaling PRE (NOE) term
eval ($a10=1.0)            !scale factor for CL1 (peaks disappeared)
eval ($a11=1.0)            !scale factor for CL2 (peaks unaffected)
eval ($a12=1.0)            !scale factor for CL3 (peaks reduced)
eval ($low1=14.0)          !lower(d_minus) limit (restraints upper limit only)
eval ($up1=4.0)           !upper(d_plus) limit (restraints upper limit only)
eval ($low2=4.0)           !lower(d_minus) limit (restraints lower limit only)
eval ($up2=100.0)          !upper(d_plus) limit (restraints lower limit only)
eval ($low3=4.0)           !lower(d_minus) limit (restraints with both limits)
eval ($up3=4.0)           !upper(d_plus) limit (restraints with both limits)
eval ($a01 = 1)           !cycle counter
eval ($a48=$cpu*1e4)
set seed=$a48 end
eval ($a14=0)
eval ($a18=1)
eval ($nout1=0)           ! Number of structures output per run
eval ($min1=9999.0)
eval ($min2=9999.0)
eval ($ref=0)
set display=coord.dat end           !write parameters
display xx -----PARAMETERS-----
display xx startdate:   $DATE
display xx starttime:   $TIME
```



```

display xx ini.file:      $inifile
display xx timestep (ps): $a54
display xx nr of steps:  $a53          time /cycle (ps): $a61
display xx nr of cycles:  $a05          total time (ps): $a80
display xx time-spent fact: $a93
display xx general scale:  $a9
display xx TBATH:        $a55          velocity factor: $a56  fbeta: $a58
display xx .pdb threshold $a59
display xx .pdb lag factor $a60
display xx Local mimim. impulse after 10 cycles with 'constant' Etot > $a59a
display xx -----
display
set display=OUTPUT end
flag exclude elec bond angl dihe impr include vdw noe end
vector do (fbeta=$a58) (segid="PC")
vector do (vx=$a56) (segid="PC")
vector do (vy=$a56) (segid="PC")
vector do (vz=$a56) (segid="PC")
set disp=ener.dat end
display Energies for the output structures
display file      Etot      VDW      NOE
display -----
set disp=OUTPUT end
@restraints.xpl
eval ($a213=100*(rand()-0.5) ) ! push away PC
eval ($a214=100*(rand()-0.5) )
eval ($a215=100*(rand()-0.5) )
vector do (x=x+$a213) (segid="PC")
vector do (y=y+$a214) (segid="PC")
vector do (z=z+$a215) (segid="PC")
!write coor OUTPUT="test1.pdb" end
parameter
  @nbfix.4sl_expl.xpl
  nbonds
    cutnb=8.5
    inhi=0.25
    ctofnb=7.5
    ctonnb=6.5
    repe1=0.6 !0.6
              NBXMod=-2
    rexp=2
    irex=2
    rcon=$a301
    wmin=1.5
  end
end
constraints
  interactions (segid="CYTF")(segid="PC")
end
energy end
while ($a01 LE $a05) loop calc
display cycle $a01
dynamics rigid
  dt=$a54
  group=(segid="PC")
  dynmode=TCOU
  tbath=$a55
  nprint=500
  nstep=$a53
  NTRFRQ=0 !new for XPLOR vs 3.8
end
eval ($a14=$ENER)
if ($a14 < $min2) then
  coor copy end

```

```

eval ($min2 = $a14)
eval ($abc=$VDW)
eval ($a15=$NOE)
eval ($a16=$a01)
eval ($a17=$a18)
end if
if ($a18 = $u2) then           !counting number of dockings
if ($min2 < $a59) then
if ($abc < $van) then
    coor swap end
    eval ($nout1=$nout1+1)
    eval ($pdb="structure_"+encode($nout1)+".pdb")
    write coord output=$pdb end
    if ($a14<$min1) then
        eval ($ref=$nout1)
    eval ($min1 = $a14)
    end if
    pick bond (segid="CYTF" and name FE) (segid="PC" and name CU) geom
    eval ($a50=$RESULT)
    set disp=ener.dat end
    display $nout1 $min2      $abc      $a15
    set disp=OUTPUT end
    set disp=coord.dat end
    display Cycle: $a16      Dock step: $a17      File#: $nout1      Fe-Cu: $a50      temp: $TEMP
    display Etot:  $min2      Evdw: $abc      Enoe: $a15
    display -----
        set disp=OUTPUT end
!violation analysis
    eval ($b3="structure_"+encode($nout1)+".viol.dat")
    set display=$b3 end
    display Violation analysis PRE data
    display res#  PRE(meas) PRE(calc) Dist(calc)
    display
    for $g2 in ($g2a $g2b $g2c)
    loop spinlabel
        eval ($g2d=$g2)
        eval ($g2e=$g2+1)
        eval ($g2f=$g2+2)
        eval ($g2g=$g2+3)
        if ($g2=$g2a) then
            eval ($g1h = $g1a)
            display Spin Label $g1h
            for
                $c02 in ID (store1)
            loop C3
                vector show elem (resi) (ID $c02)
                eval ($c04=$RESULT)                                !residue nr
                vector show elem (store1) (ID $c02)
                eval ($c08=$RESULT)
                pick bond (resn SL and resi $g2d and name O) (ID $c02) geom
                eval ($ca1=$RESULT)
                pick bond (resn SL and resi $g2e and name O) (ID $c02) geom
                eval ($ca2=$RESULT)
                pick bond (resn SL and resi $g2f and name O) (ID $c02) geom
                eval ($ca3=$RESULT)
                pick bond (resn SL and resi $g2g and name O) (ID $c02) geom
                eval ($ca4=$RESULT)
                eval ($c03=($ca1^(-6)+$ca2^(-6)+$ca3^(-6)+$ca4^(-6))/4)
                    !r-6 average distance over 4 positions
                eval ($c05=(1.23E16*(4*$tau_c+(3*$tau_c/(1+($field*2*3.14*$tau_c)^2)))*frit*$a93*$c03) )
                    !back calculated PRE
                eval ($c10=$c03^(-1/6))
                display $c04      $c08      $c05      $c10
            end loop C3
        end for
    end for
end if
end if
end if

```

```

display
end if
if ($g2=$g2b) then
eval ($g1h = $g1b)
display Spin Label $g1h
for
    $c02 in ID (store2)
loop C3
vector show elem (resi) (ID $c02)
eval ($c04=$RESULT) !residue nr
vector show elem (store2) (ID $c02)
eval ($c08=$RESULT)
pick bond (resn SL and resi $g2d and name O) (ID $c02) geom
eval ($ca1=$RESULT)
pick bond (resn SL and resi $g2e and name O) (ID $c02) geom
eval ($ca2=$RESULT)
pick bond (resn SL and resi $g2f and name O) (ID $c02) geom
eval ($ca3=$RESULT)
pick bond (resn SL and resi $g2g and name O) (ID $c02) geom
eval ($ca4=$RESULT)
eval ($c03=($ca1^(-6)+$ca2^(-6)+$ca3^(-6)+$ca4^(-6))/4)
!r-6 average distance over 4 positions
eval ($c05=(1.23E16*(4*$tau_c+(3*$tau_c/(1+($field*2*3.14*$tau_c)^2)))*$frl*$a93*$c03) )
!back calculated PRE
eval ($c10=$c03^(-1/6))
display $c04 $c08 $c05 $c10
end loop C3
display
end if
if ($g2=$g2c) then
eval ($g1h = $g1c)
display Spin Label $g1h
for
    $c02 in ID (store3)
loop C3
vector show elem (resi) (ID $c02)
eval ($c04=$RESULT) !residue nr
vector show elem (store3) (ID $c02)
eval ($c08=$RESULT)
pick bond (resn SL and resi $g2d and name O) (ID $c02) geom
eval ($ca1=$RESULT)
pick bond (resn SL and resi $g2e and name O) (ID $c02) geom
eval ($ca2=$RESULT)
pick bond (resn SL and resi $g2f and name O) (ID $c02) geom
eval ($ca3=$RESULT)
pick bond (resn SL and resi $g2g and name O) (ID $c02) geom
eval ($ca4=$RESULT)
eval ($c03=($ca1^(-6)+$ca2^(-6)+$ca3^(-6)+$ca4^(-6))/4)
!r-6 average distance over 4 positions
eval ($c05=(1.23E16*(4*$tau_c+(3*$tau_c/(1+($field*2*3.14*$tau_c)^2)))*$frl*$a93*$c03) )
!back calculated PRE
eval ($c10=$c03^(-1/6))
display $c04 $c08 $c05 $c10
end loop C3
display
end if
end loop spinlabel
close $b3 end
set display=OUTPUT end
!end violation analysis
coor swap end
end if
end if
!impulse to escape local minimum

```

```

eval ($a213=100*(rand()-0.5) )
eval ($a214=100*(rand()-0.5) )
eval ($a215=100*(rand()-0.5) )
vector do (x=x+$a213) (segid="PC")
vector do (y=y+$a214) (segid="PC")
vector do (z=z+$a215) (segid="PC")
vector do (vx=50) (segid="PC")
vector do (vy=50) (segid="PC")
vector do (vz=50) (segid="PC")
set disp=coor.dat end
display cycle $a01: impulse: $a213, $a214, $a215 to x,y,z; v=50
set disp=OUTPUT end
eval ($a18=0)
eval ($min2=9999.0)
end if
eval ($a18=$a18+1)
if ($TEMP>200000.0) then                !correct excessive temperature
  eval ($a45=RAND()+0.01)
  vector do (vx=$a56*$a45*rand()) (segid="PC")
  vector do (vy=$a56*$a45*rand()) (segid="PC")
  vector do (vz=$a56*$a45*rand()) (segid="PC")
end if
eval ($a01 = $a01 + 1)
end loop calc
if ($ref > 0) then
vector idend ( store9 ) ( name ca or name n or name c )      !backbone selection
eval ($ref_file="structure_"+encode($ref)+".pdb")
set display=rms.dat end
display Backbone pairwise RMSD from the lowest energy strucutre ($ref_file)
display file      rmsd
display -----
set disp=OUTPUT end
  coor swap end
  coor init end
  coor swap end
  coor disp=comp @@$ref_file
eval ($count1=0)
while ($count1 < $nout1) loop fill
  evaluate ($count1=$count1+1)
  evaluate ($file="structure_"+encode($count1)+".pdb")
  coor init end
  coor @@$file
  coor sele=(recall 9) fit end
  coor sele=(recall 9) rms end
  eval ($b1 = $result)
  set display=rms.dat end
  display $file      $b1
end loop fill
end if
set display=OUTPUT end
set echo=true end
stop

```

Appendix 4

Input file for rigid-body docking of Cyt *f*-Pc using PCS restraints for Xplor-NIH 2.9.9

```
parameter
  @nostoc_cytf_SL_pc.par
end
structure
  @nostoc_cytf_SL_pc.psf
end
eval ($infile="nostoc_cytf_SL_pc.pdb")
coordinates @$infile
@learn.pcf.par
constraints fix (segid="CYTF" or resn SL) end
eval ($a05 = 6000)           !total nr cycles / $a53
eval ($a53 = 1000)         !nr of steps
eval ($u2 = 30)             !nr of cycles per docking approach
eval ($a54 = 0.01)         !fimestep in ps
eval ($a61 = $a54*$a53)    !time per cycle (ps)
eval ($a80 = $a61*$a05)    !total time (ps)
eval ($a58=30.0)           !velocity factor
eval ($a56=1.5)            !fbeta
eval ($a55=300.0)          !TBATH
eval ($a59=45.0)           !.pdb writing threshold
eval ($van=100.0)          ! vdW writing threshold
eval ($a59a=0.01*$a59)     !vx-reset minimum
eval ($a60=1.0)            !.pdb lag factor
eval ($a301=1.0)           !VDW repel scale factor
eval ($a93=1)              !time-spent factor: 0<$a93<1 (fraction bound)
eval ($g1a = "N71C")       !names of the spin label 1 used in calculation
eval ($g1b = "Q104C")      !names of the spin label 2 used in calculation
eval ($g1c = "S192C")      !names of the spin label 3 used in calculation
eval ($g2a = 13)           !residue number of the 1stSL conformation of SL 1
eval ($g2b = 17)           !residue number of the 1st SL conformation of SL 2
eval ($g2c = 21)           !residue number of the 1st SL conformation of SL 3
eval ($field=600.1328E6)   !field strength in Hz
eval ($frr=1)              !fraction of proteins with spin label
eval ($tau_c=30.0E-9)      !tau c for the complex in sec
eval ($a9=0.01)           !general scaling PRE (NOE) term
eval ($a10=1.0)            !scale factor for CL1 (peaks disappeared)
eval ($a11=1.0)            !scale factor for CL2 (peaks unaffected)
eval ($a12=1.0)            !scale factor for CL3 (peaks reduced)
eval ($low1=14.0) !lower(d_minus) limit (restraints upper limit only)
eval ($up1=4.0) !upper(d_plus) limit (restraints upper limit only)
eval ($low2=4.0) !lower(d_minus) limit (restraints lower limit only)
eval ($up2=100.0) !upper(d_plus) limit (restraints lower limit only)
eval ($low3=4.0) !lower(d_minus) limit (restraints with both limits)
eval ($up3=4.0) !upper(d_plus) limit (restraints with both limits)
eval ($a01 = 1)           !cycle counter
eval ($a48=$cpu*1e4)
set seed=$a48 end
eval ($a14=0)
eval ($a18=1)
eval ($nout1=0) ! Number of structures output per run
eval ($min1=9999.0)
eval ($min2=9999.0)
eval ($ref=0)
set display=coord.dat end !write parameters
display xx -----PARAMETERS-----
display xx startdate: $DATE
display xx starttime: $TIME
```

```

display xx ini.file:      $inifile
display xx timestep (ps): $a54
display xx nr of steps:  $a53          time /cycle (ps): $a61
display xx nr of cycles:  $a05          total time (ps): $a80
display xx time-spent fact: $a93
display xx general scale:  $a9
display xx TBATH:         $a55 velocity factor: $a56 fbeta: $a58
display xx .pdb threshold $a59
display xx .pdb lag factor $a60
display xx Local mimim. impulse after 10 cycles with 'constant' Etot > $a59a
display xx -----
display
set display=OUTPUT end
flag exclude elec bond angl dihe impr include vdw noe end
vector do (fbeta=$a58) (segid="PC")
vector do (vx=$a56) (segid="PC")
vector do (vy=$a56) (segid="PC")
vector do (vz=$a56) (segid="PC")
set disp=ener.dat end
display Energies for the output structures
display file      Etot      VDW      NOE
display -----
set disp=OUTPUT end
@restraints.xpl
eval ($a213=100*(rand()-0.5) ) ! push away PC
eval ($a214=100*(rand()-0.5) )
eval ($a215=100*(rand()-0.5) )
vector do (x=x+$a213) (segid="PC")
vector do (y=y+$a214) (segid="PC")
vector do (z=z+$a215) (segid="PC")
!write coor OUTPUT="test1.pdb" end
parameter
  @nbfix.4sl_expl.xpl
  nbonds
    cutnb=8.5
    inhi=0.25
    ctofnb=7.5
    ctonnb=6.5
    repe1=0.6 !0.6
              NBXMod=-2
    rexp=2
    irex=2
    rcon=$a301
    wmin=1.5
  end
end
constraints
  interactions (segid="CYTF")(segid="PC")
end
energy end
while ($a01 LE $a05) loop calc
display cycle $a01
dynamics rigid
  dt=$a54
  group=(segid="PC")
  dynmode=TCOU
  fbath=$a55
  nprint=500
  nstep=$a53
  NTRFRQ=0 !new for XPLOR vs 3.8
end
eval ($a14=$ENER)
if ($a14 < $min2) then
  coor copy end

```

```

eval ($min2 = $a14)
eval ($abc=$VDW)
eval ($a15=$NOE)
eval ($a16=$a01)
eval ($a17=$a18)
end if
if ($a18 = $u2) then           !counting number of dockings
if ($min2 < $a59) then
if ($abc < $van) then
    coor swap end
    eval ($nout1=$nout1+1)
    eval ($pdb="structure_"+encode($nout1)+".pdb")
    write coord output=$pdb end
    if ($a14<$min1) then
        eval ($ref=$nout1)
    eval ($min1 = $a14)
    end if
    pick bond (segid="CYTF" and name FE) (segid="PC" and name CU) geom
    eval ($a50=$RESULT)
    set disp=ener.dat end
    display $nout1 $min2      $abc      $a15
    set disp=OUTPUT end
    set disp=coord.dat end
    display Cycle: $a16      Dock step: $a17      File#: $nout1      Fe-Cu: $a50      temp: $TEMP
    display Etot:  $min2      Evdw: $abc      Enoe: $a15
    display -----
        set disp=OUTPUT end
!violation analysis
    eval ($b3="structure_"+encode($nout1)+".viol.dat")
    set display=$b3 end
    display Violation analysis PRE data
    display res#  PRE(meas) PRE(calc) Dist(calc)
    display
    for $g2 in ($g2a $g2b $g2c)
    loop spinlabel
        eval ($g2d=$g2)
        eval ($g2e=$g2+1)
        eval ($g2f=$g2+2)
        eval ($g2g=$g2+3)
        if ($g2=$g2a) then
            eval ($g1h = $g1a)
            display Spin Label $g1h
            for
                $c02 in ID (store1)
            loop C3
                vector show elem (resi) (ID $c02)
                eval ($c04=$RESULT)                                !residue nr
                vector show elem (store1) (ID $c02)
                eval ($c08=$RESULT)
                pick bond (resn SL and resi $g2d and name O) (ID $c02) geom
                eval ($ca1=$RESULT)
                pick bond (resn SL and resi $g2e and name O) (ID $c02) geom
                eval ($ca2=$RESULT)
                pick bond (resn SL and resi $g2f and name O) (ID $c02) geom
                eval ($ca3=$RESULT)
                pick bond (resn SL and resi $g2g and name O) (ID $c02) geom
                eval ($ca4=$RESULT)
                eval ($c03=($ca1^(-6)+$ca2^(-6)+$ca3^(-6)+$ca4^(-6))/4)
                    !r-6 average distance over 4 positions
                eval ($c05=(1.23E16*(4*$tau_c+(3*$tau_c/(1+($field*2*3.14*$tau_c)^2)))*frit*$a93*$c03) )
                    !back calculated PRE
                eval ($c10=$c03^(-1/6))
                display $c04      $c08      $c05      $c10
            end loop C3
        end for
    end for
end if
end if
end if

```

```

display
end if
if ($g2=$g2b) then
eval ($g1h = $g1b)
display Spin Label $g1h
for
    $c02 in ID (store2)
loop C3
vector show elem (resi) (ID $c02)
eval ($c04=$RESULT) !residue nr
vector show elem (store2) (ID $c02)
eval ($c08=$RESULT)
pick bond (resn SL and resi $g2d and name O) (ID $c02) geom
eval ($ca1=$RESULT)
pick bond (resn SL and resi $g2e and name O) (ID $c02) geom
eval ($ca2=$RESULT)
pick bond (resn SL and resi $g2f and name O) (ID $c02) geom
eval ($ca3=$RESULT)
pick bond (resn SL and resi $g2g and name O) (ID $c02) geom
eval ($ca4=$RESULT)
eval ($c03=($ca1^(-6)+$ca2^(-6)+$ca3^(-6)+$ca4^(-6))/4)
!r-6 average distance over 4 positions
eval ($c05=(1.23E16*(4*$tau_c+(3*$tau_c/(1+($field*2*3.14*$tau_c)^2)))*$frl*$a93*$c03) )
!back calculated PRE
eval ($c10=$c03^(-1/6))
display $c04 $c08 $c05 $c10
end loop C3
display
end if
if ($g2=$g2c) then
eval ($g1h = $g1c)
display Spin Label $g1h
for
    $c02 in ID (store3)
loop C3
vector show elem (resi) (ID $c02)
eval ($c04=$RESULT) !residue nr
vector show elem (store3) (ID $c02)
eval ($c08=$RESULT)
pick bond (resn SL and resi $g2d and name O) (ID $c02) geom
eval ($ca1=$RESULT)
pick bond (resn SL and resi $g2e and name O) (ID $c02) geom
eval ($ca2=$RESULT)
pick bond (resn SL and resi $g2f and name O) (ID $c02) geom
eval ($ca3=$RESULT)
pick bond (resn SL and resi $g2g and name O) (ID $c02) geom
eval ($ca4=$RESULT)
eval ($c03=($ca1^(-6)+$ca2^(-6)+$ca3^(-6)+$ca4^(-6))/4)
!r-6 average distance over 4 positions
eval ($c05=(1.23E16*(4*$tau_c+(3*$tau_c/(1+($field*2*3.14*$tau_c)^2)))*$frl*$a93*$c03) )
!back calculated PRE
eval ($c10=$c03^(-1/6))
display $c04 $c08 $c05 $c10
end loop C3
display
end if
end loop spinlabel
close $b3 end
set display=OUTPUT end
!end violation analysis
coor swap end
end if
end if
eval ($a213=100*(rand()-0.5) ) !impulse to escape local minimum

```



```

eval ($a214=100*(rand()-0.5) )
eval ($a215=100*(rand()-0.5) )
vector do (x=x+$a213) (segid="PC")
vector do (y=y+$a214) (segid="PC")
vector do (z=z+$a215) (segid="PC")
vector do (vx=50) (segid="PC")
vector do (vy=50) (segid="PC")
vector do (vz=50) (segid="PC")
set disp=coor.dat end
display cycle $a01: impulse: $a213, $a214, $a215 to x,y,z; v=50
set disp=OUTPUT end
eval ($a18=0)
eval ($min2=9999.0)
end if
eval ($a18=$a18+1)
if ($TEMP>200000.0) then                !correct excessive temperature
  eval ($a45=RAND()+0.01)
  vector do (vx=$a56*$a45*rand()) (segid="PC")
  vector do (vy=$a56*$a45*rand()) (segid="PC")
  vector do (vz=$a56*$a45*rand()) (segid="PC")
end if
eval ($a01 = $a01 + 1)
end loop calc
if ($ref > 0) then
vector idend ( store9 ) ( name ca or name n or name c )      !backbone selection
eval ($ref_file="structure_"+encode($ref)+".pdb")
set display=rms.dat end
display Backbone pairwise RMSD from the lowest energy strucutre ($ref_file)
display file      rmsd
display -----
set disp=OUTPUT end
  coor swap end
  coor init end
  coor swap end
  coor disp=comp @@$ref_file
eval ($count1=0)
while ($count1 < $nout1) loop fill
  evaluate ($count1=$count1+1)
    evaluate ($file="structure_"+encode($count1)+".pdb")
    coor init end
    coor @$file
    coor sele=(recall 9) fit end
    coor sele=(recall 9) rms end
    eval ($b1 = $result)
  set display=rms.dat end
  display $file      $b1
end loop fill
end if
set display=OUTPUT end
set echo=true end
stop

```

Appendix 5

Input file to backcalculate PCS from Cyt *f* Fe to Pc amide protons for Xplor-NIH 2.9.9

```
parameter
  @nostoc_cytf_SL_pc.par
end
structure
  @nostoc_cytf_SL_pc.psf
end
eval ($infile="nostoc_cytf_SL_pc.pdb")
coordinates @$infile
@learn.pcf.par
constraints fix (segid="CYTF" or resn SL) end
eval ($a05 = 6000)           !total nr cycles / $a53
eval ($a53 = 1000)          !nr of steps
eval ($u2 = 30)             !nr of cycles per docking approach
eval ($a54 = 0.01)          !timestep in ps
eval ($a61 = $a54*$a53)     !time per cycle (ps)
eval ($a80 = $a61*$a05)     !total time (ps)
eval ($a58=30.0)           !velocity factor
eval ($a56=1.5)             !fbeta
eval ($a55=300.0)          !TBATH
eval ($a59=45.0)           !.pdb writing threshold
eval ($van=100.0)          ! vdw writing threshold
eval ($a59a=0.01*$a59)     !vx-reset minimum
eval ($a60=1.0)            !.pdb lag factor
eval ($a301=1.0)           !VDW repel scale factor
eval ($a93=1)              !time-spent factor: 0<$a93<1 (fraction bound)
eval ($g1a = "N71C")       !names of the spin label 1 used in calculation
eval ($g1b = "Q104C")     !names of the spin label 2 used in calculation
eval ($g1c = "S192C")     !names of the spin label 3 used in calculation
eval ($g2a = 13)           !residue number of the 1stSL conformation of SL 1
eval ($g2b = 17)           !residue number of the 1st SL conformation of SL 2
eval ($g2c = 21)           !residue number of the 1st SL conformation of SL 3
eval ($field=600.1328E6)   !field strength in Hz
eval ($frl=1)              !fraction of proteins with spin label
eval ($tau_c=30.0E-9)      !tau c for the complex in sec
eval ($a9=0.01)           !general scaling PRE (NOE) term
eval ($a10=1.0)            !scale factor for CL1 (peaks disappeared)
eval ($a11=1.0)            !scale factor for CL2 (peaks unaffected)
eval ($a12=1.0)            !scale factor for CL3 (peaks reduced)
eval ($low1=14.0)          !lower(d_minus) limit (restraints upper limit only)
eval ($up1=4.0)           !upper(d_plus) limit (restraints upper limit only)
eval ($low2=4.0)          !lower(d_minus) limit (restraints lower limit only)
eval ($up2=100.0)         !upper(d_plus) limit (restraints lower limit only)
eval ($low3=4.0)          !lower(d_minus) limit (restraints both limits)
eval ($up3=4.0)           !upper(d_plus) limit (restraints both limits)
eval ($a01 = 1)           !cycle counter
eval ($a48=$cpu*1e4)
set seed=$a48 end
eval ($a14=0)
eval ($a18=1)
eval ($nout1=0)           ! Number of structures output per run
eval ($min1=9999.0)
eval ($min2=9999.0)
eval ($ref=0)
set display=coord.dat end !write parameters
display xx -----PARAMETERS-----
display xx startdate:    $DATE
```

```

display xx starttime:      $TIME
display xx ini.file:      $inifile
display xx timestep (ps): $a54
display xx nr of steps:   $a53           time /cycle (ps): $a61
display xx nr of cycles:  $a05           total time (ps): $a80
display xx time-spent fact: $a93
display xx general scale:  $a9
display xx TBATH:         $a55 velocity factor: $a56 fbeta: $a58
display xx .pdb threshold $a59
display xx .pdb lag factor $a60
display xx Local mimim. impulse after 10 cycles with 'constant' Etot > $a59a
display xx -----
display
set display=OUTPUT end
flag exclude elec bond angl dihe impr include vdw noe end
vector do (fbeta=$a58) (segid="PC")
vector do (vx=$a56) (segid="PC")
vector do (vy=$a56) (segid="PC")
vector do (vz=$a56) (segid="PC")
set disp=ener.dat end
display Energies for the output structures
display file      Etot      VDW      NOE
display -----
set disp=OUTPUT end
@restraints.xpl
eval ($a213=100*(rand()-0.5) ) ! push away PC
eval ($a214=100*(rand()-0.5) )
eval ($a215=100*(rand()-0.5) )
vector do (x=x+$a213) (segid="PC")
vector do (y=y+$a214) (segid="PC")
vector do (z=z+$a215) (segid="PC")
!write coor OUTPUT="test1.pdb" end
parameter
  @nbfix.4sl_expl.xpl
  nbonds
    cutnb=8.5
    inhi=0.25
    ctofnb=7.5
    ctonnb=6.5
    repe1=0.6 !0.6
                NBXMod=-2
    rexp=2
    irex=2
    rcon=$a301
    wmin=1.5
  end
end
constraints
  interactions (segid="CYTF")(segid="PC")
end
energy end
while ($a01 LE $a05) loop calc
display cycle $a01
dynamics rigid
  dt=$a54
  group=(segid="PC")
  dynmode=TCOU
  tbath=$a55
  nprint=500
  nstep=$a53
  NTRFRQ=0 !new for XPLOr vs 3.8
end
eval ($a14=$ENER)
if ($a14 < $min2) then

```

```

    coor copy end
    eval ($min2 = $a14)
    eval ($abc=$VDW)
    eval ($a15=$NOE)
    eval ($a16=$a01)
    eval ($a17=$a18)
end if
if ($a18 = $u2) then           !counting number of dockings
if ($min2 < $a59) then
if ($abc < $van) then
    coor swap end
    eval ($nout1=$nout1+1)
    eval ($pdb="structure_"+encode($nout1)+".pdb")
    write coord output=$pdb end
    if ($a14<$min1) then
        eval ($ref=$nout1)
    eval ($min1 = $a14)
    end if
    pick bond (segid="CYTF" and name FE) (segid="PC" and name CU) geom
    eval ($a50=$RESULT)
    set disp=ener.dat end
    display $nout1 $min2      $abc      $a15
    set disp=OUTPUT end
    set disp=coor.dat end
    display Cycle: $a16      Dock step: $a17      File#: $nout1      Fe-Cu: $a50      temp: $TEMP
    display Etot:  $min2      Evdw: $abc      Enoe: $a15
    display -----
        set disp=OUTPUT end
!violation analysis
    eval ($b3="structure_"+encode($nout1)+".viol.dat")
    set display=$b3 end
    display Violation analysis PRE data
    display res#  PRE(meas) PRE(calc) Dist(calc)
    display
    for $g2 in ($g2a $g2b $g2c)
    loop spinlabel
        eval ($g2d=$g2)
        eval ($g2e=$g2+1)
        eval ($g2f=$g2+2)
        eval ($g2g=$g2+3)
        if ($g2=$g2a) then
            eval ($g1h = $g1a)
            display Spin Label $g1h
            for
                $c02 in ID (store1)
            loop C3
                vector show elem (resi) (ID $c02)
                eval ($c04=$RESULT)           !residue nr
                vector show elem (store1) (ID $c02)
                eval ($c08=$RESULT)
                pick bond (resn SL and resi $g2d and name O) (ID $c02) geom
                eval ($ca1=$RESULT)
                pick bond (resn SL and resi $g2e and name O) (ID $c02) geom
                eval ($ca2=$RESULT)
                pick bond (resn SL and resi $g2f and name O) (ID $c02) geom
                eval ($ca3=$RESULT)
                pick bond (resn SL and resi $g2g and name O) (ID $c02) geom
                eval ($ca4=$RESULT)
                eval ($c03=($ca1^(-6)+$ca2^(-6)+$ca3^(-6)+$ca4^(-6))/4)
                !r-6 average distance over 4 positions
                eval ($c05=(1.23E16*(4*$tau_c+(3*$tau_c/(1+($field*2*3.14*$tau_c)^2)))*$fri*$a93*$c03) )
                !back calculated PRE
                eval ($c10=$c03^(-1/6))
                display $c04      $c08      $c05      $c10
            end loop
        end for
    end for
end if
end if
end if

```

```

end loop C3
display
end if
if ($g2=$g2b) then
eval ($g1h = $g1b)
display Spin Label $g1h
for
    $c02 in ID (store2)
loop C3
vector show elem (resi) (ID $c02)
eval ($c04=$RESULT)
!residue nr
vector show elem (store2) (ID $c02)
eval ($c08=$RESULT)
pick bond (resn SL and resi $g2d and name O) (ID $c02) geom
eval ($ca1=$RESULT)
pick bond (resn SL and resi $g2e and name O) (ID $c02) geom
eval ($ca2=$RESULT)
pick bond (resn SL and resi $g2f and name O) (ID $c02) geom
eval ($ca3=$RESULT)
pick bond (resn SL and resi $g2g and name O) (ID $c02) geom
eval ($ca4=$RESULT)
eval ($c03=($ca1^(-6)+$ca2^(-6)+$ca3^(-6)+$ca4^(-6))/4)
!r-6 average distance over 4 positions
eval ($c05=(1.23E16*(4*$tau_c+(3*$tau_c/(1+($field*2*3.14*$tau_c)^2)))*$fri*$a93*$c03) )
!back calculated PRE
eval ($c10=$c03^(-1/6))
display $c04 $c08 $c05 $c10
end loop C3
display
end if
if ($g2=$g2c) then
eval ($g1h = $g1c)
display Spin Label $g1h
for
    $c02 in ID (store3)
loop C3
vector show elem (resi) (ID $c02)
eval ($c04=$RESULT)
!residue nr
vector show elem (store3) (ID $c02)
eval ($c08=$RESULT)
pick bond (resn SL and resi $g2d and name O) (ID $c02) geom
eval ($ca1=$RESULT)
pick bond (resn SL and resi $g2e and name O) (ID $c02) geom
eval ($ca2=$RESULT)
pick bond (resn SL and resi $g2f and name O) (ID $c02) geom
eval ($ca3=$RESULT)
pick bond (resn SL and resi $g2g and name O) (ID $c02) geom
eval ($ca4=$RESULT)
eval ($c03=($ca1^(-6)+$ca2^(-6)+$ca3^(-6)+$ca4^(-6))/4)
!r-6 average distance over 4 positions
eval ($c05=(1.23E16*(4*$tau_c+(3*$tau_c/(1+($field*2*3.14*$tau_c)^2)))*$fri*$a93*$c03) )
!back calculated PRE
eval ($c10=$c03^(-1/6))
display $c04 $c08 $c05 $c10
end loop C3
display
end if
end loop spinlabel
close $b3 end
set display=OUTPUT end
!end violation analysis
!coor swap end
end if
end if

```

```

                                !impulse to escape local minimum
eval ($a213=100*(rand()-0.5) )
eval ($a214=100*(rand()-0.5) )
eval ($a215=100*(rand()-0.5) )
vector do (x=x+$a213) (segid="PC")
vector do (y=y+$a214) (segid="PC")
vector do (z=z+$a215) (segid="PC")
vector do (vx=50) (segid="PC")
vector do (vy=50) (segid="PC")
vector do (vz=50) (segid="PC")
set disp=coord.dat end
display cycle $a01: impulse: $a213, $a214, $a215 to x,y,z; v=50
set disp=OUTPUT end
eval ($a18=0)
eval ($min2=9999.0)
end if
eval ($a18=$a18+1)

if ($TEMP>200000.0) then                !correct excessive temperature
  eval ($a45=RAND()+0.01)
  vector do (vx=$a56*$a45*rand()) (segid="PC")
  vector do (vy=$a56*$a45*rand()) (segid="PC")
  vector do (vz=$a56*$a45*rand()) (segid="PC")
end if
eval ($a01 = $a01 + 1)
end loop calc
if ($ref > 0) then
  vector idend ( store? ) ( name ca or name n or name c )      !backbone selection
  eval ($ref_file="structure_"+encode($ref)+".pdb")
  set display=rms.dat end
  display Backbone pairwise RMSD from the lowest energy strucutre ($ref_file)
  display file      rmsd
  display -----
  set disp=OUTPUT end
  coor swap end
  coor init end
  coor swap end
  coor disp=comp @@$ref_file
  eval ($count1=0)
  while ($count1 < $nout1) loop fill
    evaluate ($count1=$count1+1)
    evaluate ($file="structure_"+encode($count1)+".pdb")
    coor init end
    coor @@$file
    coor sele=(recall 9) fit end
    coor sele=(recall 9) rms end
    eval ($b1 = $result)
  set display=rms.dat end
  display $file      $b1
  end loop fill
end if
set display=OUTPUT end
set echo=true end
stop

```

List of publications

Scanu S., Förster J.M., Timmer M., Ullmann G.M. and Ubbink M. (2013) Loss of electrostatic interactions causes increase of dynamics within the plastocyanin-cytochrome *f* complex. *Manuscript under revision*.

Scanu S., Förster J.M., Ullmann G.M. and Ubbink M. (2013) Role of hydrophobic interactions in the encounter complex formation of plastocyanin and cytochrome *f* complex revealed by paramagnetic NMR spectroscopy. *J. Am. Chem. Soc.* **135**, 7681-7692.

Scanu S., Förster J., Finiguerra M.G., Shabestari M.H., Huber M. and Ubbink M. (2012) The complex of cytochrome *f* and plastocyanin from *Nostoc* sp. PCC 7119 is highly dynamic. *ChemBiochem.* **13**, 1312-1318

Bashir Q., **Scanu S.**, and Ubbink M. (2011) Dynamics in electron transfer protein complexes. *FEBS J.* **278**, 1391-1400

Xu X., **Scanu S.**, Chung J.S., Hirasawa M., Knaff D.B. and Ubbink M. (2010) Structural and functional characterization of the Ga-substituted ferredoxin from *Synechocystis* sp. PCC6803, a mimic of the native protein. *Biochemistry*, **49**, 7790-7797

Barbiroli, A., Beringhelli, T., Bonomi, F., Donghi, D., Ferranti, P., Galliano, M., Iametti, S., Maggioni, D., Rasmussen, P., **Scanu, S.** and Vilaro, M. C. (2010) Bovine β -lactoglobulin acts as an acid-resistant drug carrier by exploiting its diverse binding regions. *Biological Chemistry*. **391**, 21-32

Curriculum vitae

Sandra Scanu was born on August 27th 1981 in Pattada, a small village located in the heart of Sardinia. She spent a light-hearted childhood in contact with wild and beautiful nature, surrounded by the love of family and friends. Since she was a child, she has showed her curiosity for the world by asking unexpected and, sometimes, surprising questions about life and people. After obtaining her diploma in the scientific high school A. Segni in 2000, she decided to move to Milan to pursue her dream of becoming a scientist. Here, she enrolled in Medical Biotechnology and she obtained her Bachelor degree in 2004, after attending a three months internship in the lab of Prof. G.P. Comi at the Policlinico of Milan. After one year of Chemistry courses, she started her studies in Pharmaceutical Biotechnology, earning a master's degree in 2008. From 2006 to 2008 she worked in the lab of Prof. T. Beringhelli, first as a master's student then as a post-graduate fellow. During these years she has remained fascinated by the endless applications of NMR spectroscopy. This experience led her to Leiden in 2008, where she became a PhD student in the ProtChem group under the supervision of Prof. M. Ubbink. Here, she studied the principles of protein interactions using paramagnetic NMR spectroscopy. Once she overcame the initial troubles with English, she managed to settle down perfectly in the amazing city that accommodated her for four years. Soon she met friends from all over the world, with whom she organized many gatherings and dinner parties. Beside her passion for science, Sandra is in fact successfully dedicated to another art: the good cuisine.

She is now about to begin another journey to Munich, where she will start a post-doctorate position in the group of Biomolecular NMR spectroscopy under the supervision of Dr. T. Madl.

By Giannina Scanu