

## Genetically modified (GM) corn in the Philippines : Ecological impacts on agroecosystems, effects on the economic status and farmers' experiences

Mabutol-Afidchao, M.B.

### Citation

Mabutol-Afidchao, M. B. (2013, November 20). *Genetically modified (GM) corn in the Philippines : Ecological impacts on agroecosystems, effects on the economic status and farmers' experiences*. Retrieved from https://hdl.handle.net/1887/22273

Version:	Corrected Publisher's Version
License:	<u>Licence agreement concerning inclusion of doctoral thesis in the</u> <u>Institutional Repository of the University of Leiden</u>
Downloaded from:	https://hdl.handle.net/1887/22273

Note: To cite this publication please use the final published version (if applicable).

Cover Page



# Universiteit Leiden



The handle <u>http://hdl.handle.net/1887/22273</u> holds various files of this Leiden University dissertation

Author: Mabutol-Afidchao, Miladis B.

**Title:** Genetically modified (GM) corn in the Philippines : ecological impacts on agroecosystems, effects on the economic status and farmers' experiences **Issue Date:** 2013-11-20



Invertebrate abundance and species richness in transgenic and non-transgenic cornfields in the Philippines

Miladis M. Afidchao, C.J.M. Musters, Mercedes D. Masipiqueňa, Denyse J. Snelder and Geert R. de Snoo

To be submitted

#### Abstract

This study examined the effects of large-scale and long-term use of transgenic corn varieties on the abundance, species richness, and guilds of non-target invertebrates in a wet tropical environment. The study was conducted in 30 fields, including non-transgenic cornfields for comparison, and distributed over three lowland sites in the Philippines. The transgenic corn varieties Bt (Bacillus thuringiensis) and BtHT (Bt Roundup Ready) in this study were introduced to the area in 2002 and 2005. Information on aerial, surface, and soil-dwelling invertebrates were gathered during the mature stage of corn development using sticky-trap, pitfall-trap, and soil-core sampling technologies along 100 m transect lines laid out in the middle of 1.4 to 3.8 hectare fields. A total of 21,639 non-target invertebrates representing 129 different species were identified at the three sites, including surface (69%), aerial (26%), and soil (5%) dwellers. The non-Bt cornfields had significantly higher abundance and species richness of all non-target invertebrates than the Bt and BtHT fields (p-values, 0.001 and 0.020 respectively). Likewise, the abundance and species richness of aerial (p-values: 0.010 and 0.009, respectively) and abundance of soil (p-value: 0.03) dwelling non-target invertebrates were notably higher in the non-Bt cornfields. Cornfields' soil chemical properties such as pH, potassium and nitrogen content also influenced the abundance of invertebrates; however corn varieties indicated a stronger influence. Most importantly, the effects of these confounding variables did not take away the effect of corn varieties.

#### Introduction

One promising way of increasing profits in crop production is the use of genetically modified crop varieties, which possess novel genetic characteristics introduced to protect them against pest infestation or herbicides. Varieties available for corn (*Zea mays* L.) include HT (Herbicide tolerant), *Bt* (*Bacillus thuringiensis*), and *Bt*HT (*Bt* Herbicide tolerant). The first of these is a variety modified to be resistant to the glyphosate-containing herbicides used to control weeds, while the second is a variety able to produce a bacterial toxin with proven efficacy against the Asian Corn Borer (ACB; *Ostrinia furnacalis* Guenée), and the third is a combination of the two. In the Philippines, the Department of Agriculture approved *Bt* corn for direct use in December 2002, making the country the first in Asia to commercialize a transgenic food crop. This was followed by the approval of the *Bt*HT corn variety for commercial technology demonstration in the 2005 dry season planting (Gonzales *et al.,* 2009).

Transgenic corn varieties are believed to offer a number of advantages compared to non-transgenic varieties. They produce higher yields and allow for effective utilization of scarce land because of improved pest resistance and nutrient utilization. Specifically, *Bt* corn has been shown to produce higher yields than a near-isogenic non-*Bt* variety (Qaim and Zilberman, 2003; Dilehay *et al.*, 2004; Rice, 2004; Stanger and Lauer, 2006) and to significantly reduce pesticide use (Yorobe & Quicoy, 2006; Dillehay *et al.*, 2004). *Bt* corn has delivered important improvements in grain quality through significant reductions in the levels of mycotoxins found in the grain (Hammond *et al.*, 2004; Wu, 2007; Folcher *et al.*, 2010). Furthermore, *Bt*HT corn is suitable for no-tillage agriculture, a system of planting crops without plowing in order to reduce soil erosion and nutrient loss. Benefits like these directly or indirectly contribute to livelihood improvement and poverty alleviation among farmers.

James (2010) reported a significant increase to one billion hectares in transgenic crop production in 2010. The Philippines was the first Asian country among the 29 mega transgenic crop adopting countries. The adoption of transgenic corn in the Philippines has increased remarkably since its first field testing in 2001, as potential economic benefits made it an attractive alternative to conventional corn varieties. To date, the country ranked 13<sup>th</sup>, with more than 500,000 hectares planted with transgenic corn by about 270,000 small farmers (James, 2011). Such large-scale use of transgenic corn varieties, however, may considerably change agroecosystems, raising the question of long-term sustainability.

Transgenic crops may affect non-target organisms. For example, *Bt* corn can have adverse effects on non-target invertebrates like Common Swallowtail, *Papilio machaon* L. (Lang and Vojtech, 2006), the larvae of the herbivorous African cotton leaf worm *Spodoptera littoralis* (Meissle *et al.*, 2005a), and the generalist predatory ground beetle *Poecilus cupreus* L. (Meissle *et al.*, 2005b), and can exert a sublethal behavioral effect on honey bees (Romero *et al.*, 2008). Romero *et al.* (2007) confirmed that the *Bt* Cry1Ab protein has negative effects on non-target insects, including the parasitoids *Cotesia marginiventris* (Hymenoptera). A 200-day feeding experiment by Zwahlen *et al.* (2003) showed that *Bt* corn litter reduced the weight of adult earthworms by 18%, compared

to a 4% gain when fed with non-*Bt* corn. Further, reduced application of insecticides in areas under *Bt* cotton cultivation may cause an outbreak of non-targeted pests, as happened with the Mirid bug *Creontiades biseratense* in China (Lu *et al.*, 2010).

All the above-mentioned studies refer to impacts of *Bt* corn under temperate conditions. To our knowledge, none of the peer-reviewed articles reflects the potential impact of transgenic corn in a wet tropical environment like the Philippines. Furthermore, quantitative and evidence-based risk analyses are still needed to settle ongoing controversies over the ecological impact of transgenic crops. A meta-analysis of 42 field experiments involving *Bt* corn and insecticide application (Marvier *et al.*, 2007) indicated that non-target organisms are more abundant in *Bt* cornfields without insecticide application; but if comparison is made with non-*Bt* cornfields without insecticides, some non-target groups prove significantly less abundant. This means that *Bt* corn always requires insecticides, while *Bt* corn never does. The effects of transgenic corn requires further study in tropical agro-ecosystems, considering that the meta-analysis referred to above was based on controlled field experiments in temperate agroecosystems. The Philippines, with its relatively long-term practice of large-scale transgenic corn cultivation, provides an excellent environment for such studies.

There have been only a few, non-reviewed field studies on the impacts of transgenic corn on nontarget organisms in the Philippines (Alcantara, 2004; Javier *et al.*, 2004; Alcantara *et al.*, 2008). While these studies showed no negative effects of *Bt* corn, they mainly focused on aboveground arthropods, particularly the aerial fauna, and not on surface- and soil-dwelling invertebrates. As such, it is imperative to conduct studies covering invertebrate-corn interactions in all layers of the agroecosystem, to improve our ecological understanding of the impact of large-scale application of transgenic corn.

This study aimed to generate solid data on the effects of transgenic corn on invertebrates in a Philippine setting, representing a wet tropical environment. Specifically, this study assessed the impact of long-term cultivation of transgenic corn (*Bt* and *Bt*HT) on non-target organisms in terms of changes in abundance, species richness, and guilds of invertebrates found in different ecosystem layers, including aerial-, surface-, and soil-dwelling species.

#### Study area

The study was undertaken in the province of Isabela (17°20'N; 121°53'E) in northeast Luzon, the Philippines. Isabela is among the top six yellow corn producing provinces in the country, with a total area of 101,901 hectares used for corn cultivation, comprising 31,190 hectares of river floodplains (frequently flooded), 23,276 hectares of broad plains (occasionally flooded), and 47,436 hectares of hilly land (Anonymous, 2006).

The selected research sites cover the lower floodplain areas along the Cagayan River in the municipalities of Cabagan (near Pilig Abajo village), Tumauini (near Tunggui village), and Ilagan (near Angassian village). All sites have been major corn areas for more than 50 years. White corn was the most cultivated variety up to the mid-1980s, when yellow corn became economically viable due to the rapid increase in demand for animal feed. In recent years, transgenic corn varieties have become widely cultivated in the area.

The Cabagan and Ilagan sites are classified as first-class cornfields because of the favorable moisture and nutrient content of the clayey alluvial soils. While monocropping is the basic practice in Tumauini and Ilagan, multiple cropping with tobacco, legumes or vegetables is common practice in Cabagan (Fig. 1). The corn-based cultivation systems are all rainfed, with yields during both the dry (November to April) and wet (June to October) seasons. Corn growth and development normally take 110-120 days.



Figure 1. Study sites showing examples of mono-cropping system at the Ilagan and Tumauini sites (left photo) and a multiple-cropping system at the Cabagan site, where legumes are planted beside conventional corn (center photo) and tobacco planted near conventional and Bt corn (right photo)

#### Sampling

The Philippines

The research, which was conducted during the dry cropping season from January to May 2008, started with a study of the differences between the various corn growth stages, i.e. the vegetative, tasselling, and maturity stages, from January to March.

Five fields with transgenic corn and five with non-transgenic corn were selected at three sites, totaling 30 cornfields. Since isolated and small sized fields could affect the abundance of aerial invertebrates (Prasifka et al., 2005), which might bias the results, the study included only large fields (ranging from 1.4 to 3.79 hectares, Table 1) that were not isolated. The selected fields were located almost at the center of a large area of continuous corn landscape, so surrounding riparian habitats were assumed to have negligible effects on infield biodiversity.

Sampling of invertebrates from different habitats within the cornfields was accomplished with the same techniques used in a previous study (Afidchao et al., Chapter 3). Yellow sticky cards, pitfall traps and soil cores were used for aerial species, aboveground dwellers and belowground dwellers, respectively. Yellow sticky cards were used thrice per cropping season (i.e. during the vegetative, tasselling and maturity stages of the corn). Four sticky traps were placed within each sub-plot and left in the fields for two nights. Pitfall trap samplings were done twice per major corn stage per cropping season. Soil cores were taken with an improvised metal core during the maturity stage of the corn, to prevent damage to the standing crop.

Table 1. Average (±se) field size and soil physico-chemical characteristics of the 30 cornfields surveyed in Isabela province,

	Physical Characterist	ics	Chemical Ch	naracteristics				
Corn variety	Field Size (hectare)	Soil Texture	Soil pH	% N	P (ppm Olsen's Mtd)	K (ppm H2SO4 Etxn)	%OC	%OM
Bt	3.79 <u>+</u> 3.39	Sandy to silt Ioam	5.76 <u>+</u> 0.73	0.76 <u>+</u> 0.17	10.19 <u>+</u> 8.91	92.78 <u>+</u> 51.63	0.49 <u>+</u> 0.25	0.85 <u>+</u> 0.43
<i>Bt</i> HT	1.86 <u>+</u> 1.60	Clay to silt clay	5.69 <u>+</u> 0.68	0.71 <u>+</u> 0.20	11.79 <u>+</u> 8.23	80.42 <u>+</u> 18.30	0.50 <u>+</u> 0.19	0.86 <u>+</u> 0.33
non- <i>Bt</i>	1.40 <u>+</u> 0.92	Sandy to silt Ioam	5.79 <u>+</u> 0.63	0.75 <u>+</u> 0.25	11.27 <u>+</u> 8.69	106.83 <u>+</u> 61.96	0.52 <u>+</u> 0.17	0.90 <u>+</u> 0.29

The main physical and chemical characteristics of the soils in the different cornfields are presented in Table 1. All Bt fields surveyed had been cultivated with Bt corn for a minimum of two consecutive years. Although the Bt and BtHT corn varieties in this study were from genetically different corn lines, they were presumed to be similar, both containing the Cry1Ab protein only (Rauschen et al., 2009; Rauschen et al., 2010). More than half (18 out of 30) of the fields surveyed had been subject to pesticides, viz. insecticides (trade names: Furadan or Cymbush) or herbicides (Roundup Ready or Gramoxone), using a lever-operated knapsack sprayer (Table 2).

Table 2. Pesticides used by the farmer respondents (N = 30) on the cornfields surveyed (N = 30) during the first croppingseason of 2008 in Isabela Province, The Philippines

Corn variety	Farmers	Insecti- cides	Active Ingredients	Farmers using the insec- ticide	Herbicide <sup>4</sup>	Active Ingredients	Farmers using the herbicide
Bt	9	Furadan Cymbush	Carbofuran <sup>1</sup> Cypermethrin <sup>1</sup>	2 1	HT	Glyphosate <sup>2</sup>	4
<i>Bt</i> HT	6	Cymbush	Cypermethrin <sup>1</sup>	1	HT	Glyphosate <sup>2</sup>	4
non-Bt	15	Furadan Cymbush	Carbofuran <sup>1</sup> Cypermethrin <sup>1</sup>	3 3	HT Gramoxone	Glyphosate <sup>2</sup> Paraquat <sup>3</sup>	1 1

<sup>1</sup>Source: Snelder *et al*. (2008)

<sup>2</sup>Source: Williams et al. (2000)

<sup>3</sup>Source: www.syngenta.com/en/products\_brands/gramoxone

<sup>4</sup>Round-up Ready

#### Confounding variables

Information on confounding variables (i.e., cornfields' size, elevation and location, chemical inputs, plant height, percent weed cover, and soil physical and chemical properties) was gathered and considered in the analysis. Pesticide input was estimated by interviewing 30 farm-owner respondents about the use of pesticides in their fields. Plant height measurements were taken at the cob development stage. Soil analysis used four samples from each cornfield, weighing a minimum of one kilogram per sample. Samples were analyzed at the Regional Soil and Water Laboratory of the Department of Agriculture in Tuguegarao City. Soil texture was determined using the "Texture-by-Feel Method" (Franzmeier & Owens, 2008). Soil chemical properties were analyzed following standard methods, i.e. a 1:5 soil-water ratio to determine soil pH (Mahaney *et al.*, 2007), the Walkley-Black method for organic matter (OM), the Kjeldhal method for total nitrogen (N), the Olsen/Bray method for available soil phosphorus (P), and the flame photometric method for exchangeable soil potassium (K).

#### Statistical design and analysis

Statistical analyses used the Restricted Maximum Likelihood (REML) and Generalized Linear Mixed Model (GLMM). These models were used to be able to include both fixed (corn stage, corn variety, isolines, cornfields physical characteristics, soil physico-chemical properties and chemical applications) and random factors (site, field within site, sampling method) in the analyses (Quinn and Keough, 2007).

For each response variable, we firstly employed a general model including corn stage and corn variety and all the chosen confounding variables, including isoline, as fixed factors. Also, the random factors were included. The general model was then simplified using a stepwise regression

analyses. Model simplification involved gradual elimination of the confounding variables. For that a Maximum Likelihood fitting of the models was applied and subsequent models were compared based on AIC. The final model has the lowest AIC value, but may retain confounding variables with low p-values. We will call this the 'best fitting model'. In order to meet the model assumptions, natural log transformations were applied after adding 1 to the abundance or species number. The analysis was performed in R Statistical software version 2.7.3. Bonferroni correction was done by dividing the significance level by the number of models within the specific analysis.

#### Results

#### Growth stages

The sampling of non-target invertebrates during three different corn growth stages produced 31,171 individuals, including 8,557 aerial and 22,614 surface dwellers, belonging to 63 aerial and 56 surface species. Invertebrate abundance and species richness were significantly different between corn stages, with aerial dwellers being most abundant during the vegetative stage, while surface dwellers dominated the maturity stage (Fig. 2). However, no significant interaction was found between corn stage and corn variety (*Bt* versus non-*Bt*), neither in terms of abundance nor of species richness (Table 3). These results justify the focus of the study on one growth stage, that of maturity, in the next sections.

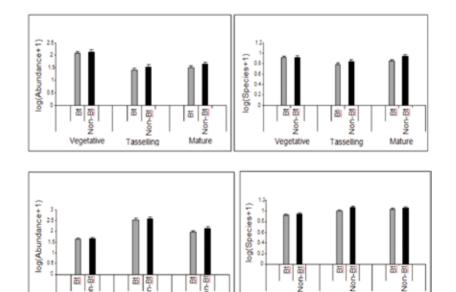


Figure 2. Estimated means (±se) of abundance (left graphs) and species richness (right graphs) of non-target aerial- (upper graphs) and surface- (lower graphs) dwelling invertebrates per corn stage of each corn variety, from the mixed regression analyses (REML, table 3)

Mature

Tasseiiing

Vegetative

Vegetative

Tasselling

Mature

**Table 3**. Results of mixed regression analyses (REML) of abundance and species richness of surface-dwelling and aerial invertebrates, with growth stages and corn variety as fixed factors and field as random factor. (n.d.f.: numerator of degrees of freedom; d.d.f.: denominator of degrees of freedom; Sign B: significance after Bonferroni correction: \*\* = equivalent to p<0.01; \* = equivalent to p<0.05; (\*) = equivalent to p<0.10).

	Wald stat.	n.d.f.	F	d.d.f.	р	Sign B
Abundance of aerial spe	ecies					
Growth stage	244.22	2	122.11	106.0	<0.001	*
Corn variety	1.10	1	1.10	8.0	0.325	ns
Stage*Variety	1.01	2	0.51	106.0	0.604	ns
Abundance of surface s	pecies					
Growth stage	412.27	2	206.14	106.0	<0.001	*
Corn variety	1.02	1	1.02	8.0	0.341	ns
Stage*Variety	3.66	2	1.83	106.0	0.166	ns
Species richness of aeria	al species					
Growth stage	20.15	2	10.08	106.0	<0.001	*
Corn variety	2.57	1	2.57	8.0	0.147	ns
Stage*Variety	3.84	2	1.92	106.0	0.152	ns
Species richness of surfa	ace species					
Growth stage	39.46	2	19.73	106.0	<0.001	*
Corn variety	2.70	1	2.70	8.0	0.139	ns
Stage*Variety	0.85	2	0.43	106.0	0.654	ns

Varieties: transgenic vs. non-transgenic corn

At the three research sites (Table 4), 21,639 individual non-target invertebrates were recorded, belonging to 129 different species. Sixty-nine percent or 14,995 individuals were surface dwellers, 26 percent or 5,585 individuals were aerial dwellers and only five percent or 1,059 individuals were soil dwellers.

**Table 4**. Total counts of Invertebrate assemblage collected in transgenic and non-transgenic corn fields in Isabela Province,Philippines during the dry cropping season 2008. FG= Functional guild (H=herbivore, O=omnivore, Pa=parasitoid,Pr=predator, De=detritivore); ID= Types of invertebrate dwellers (AD= aerial dweller, SF = soil fauna, SD = surface dweller)

Species	FG	ID	Abundance
Empoasca fabae	Н	AD	2171
Drosophila melanogaster	De	AD	844
Sciara sp.	De	AD	615
Circulife tenellus	Н	AD	358
Phaenicia sericata	De	AD	276
Micraspis discolor Fabr.	Pr	AD	262
Aphidalestes sp.	Н	AD	179
Dicampus sp.	Ра	AD	113
Heppelates sp.	Н	AD	77
Allonemobius fasciatus	0	AD	57
Pnyxia sp.	De	AD	45
Tetragnatha mandibulata	Pr	AD	44
Cheilomenes sexmaculatus	Pr	AD	40
Ephydra sp.	Н	AD	31
Rhysella nitida	Ра	AD	30
Draeculacephala mollipes	Н	AD	28
Erythroneura viridis	Н	AD	28
Phytodictus vulgaris	Ра	AD	27
Solenopsis invicta	0	AD	27
Z-Species D	Pr	AD	26
Fannia sp.	De	AD	23
Aulacophora sp.	Н	AD	22
Kestocephalus sp.	Н	AD	20
Salticus sp.	Pr	AD	18
Tettigella viridis	Н	AD	17
Orchesia sp.	_	AD	16
Oxyopes sp.	Pr	AD	15
Myrmarachne maxillosa	Pr	AD	14
Chelonus sp.	Pa	AD	12
Olibrus sp.	De	AD	10
Felleogryllus sp.	0	AD	10
sdromus sp.	Pa	AD	9
Dolichopus sp.	Pr	AD	7
Tetragnatha sp.	Pr	AD	7
Diatraes sp.	Pa	AD	6
Andrena sp.	Pr	AD	5
Phormia regina	De	AD	5
Rhagoletis pomonella	Н	AD	5
Scirtes sp.	H	AD	5
Cocinella sp.	Pr	AD	4
Gastrolinoides sp.	Н	AD	4
Heterothrips sp.	H	AD	4
Phobocampe disparis	Pa	AD	4
Amerimicromus sp.	Pr	AD	3
Chortoicetus sp.	Н	AD	3 3
Labidura truncate	Н	AD	
Solenopsis globularia	0	AD	3
Syrphus ribesii	0	AD	3 3
ZZ-Species A	0	AD	3
ZZ-Species B	De	AD	
Balaurara sp. Conocontralus en	0	AD	2
Conocephalus sp. Euroma basaba	Н	AD AD	2 2
Eurema hecabe Geocoris sp	H Pr	AD AD	2
Geocoris sp.	Pr Pr	AD AD	2
Hesperus sp. Labidura sp.			2
Labidura sp. Liadantamarus sp	H	AD	
Liodontomerus sp.	Pa	AD	2
Oncocephalus cenfusus	Pr	AD	2 2
Pholcus phalangoides	Pr	AD	2
Solenopsis pergundei	0	AD	2
Adelphocoris ropidus	Н	AD	1

Species	FG	ID	Abundance	
Agrypnus sp.	Н	AD	1	
Apion sp.	Н	AD	1	
Aufidus sp.	Н	AD	1	
Cercion calamorum	Pr	AD	1	
Cylas formicarius elegantulus	н	AD	1	
Draeculacephala sp. Euphorocera sp.	H Pa	AD AD	1	
Euphoroceru sp. Euxoa excellens	Pa H	AD AD	1	
Fornax sp.	Н	AD AD	1	
Goelerucella maculicollis	H	AD	1	
Hemiptarsemus sp.	Pa	AD	1	
Iphiaulax sp.	Pa	AD	1	
Lucilia ilustris	De	AD	1	
Meteosus sp.	Ра	AD	1	
Metoponium sp	Н	AD	1	
Ostrinia furnacalis	Н	AD	1	
Perkinsiella sp.	Н	AD	1	
Scaphisoma sp.	Pr	AD	1	
Sericesthis geminate	Pr	AD	1	
Syrphus torus	0	AD	1	
Tachys sp.	Pr	AD	1	
Trichiohelcon sp.	Pa	AD	1	
ZZ-Species C	H	AD		
Total # of AD Individuals			5,5	
Total # of AD Species				84
Hesperus sp.	Pr	SD	5499	
Pselaphus sp.	Pr	SD	2637	
Myara sp.	0	SD	1949	
Sippuna sp.	Pr	SD	1172	
Oxidus gracilis	De	SD	701	
Drosophila melanogaster	De	SD	667	
Albonemobius fasciatus	0	SD	599	
Adelocera sp	н	SD	365	
Solenopsis invicta	0	SD	307	
Solenopsis globularia	0	SD	197	
Cocinella sp.	Pr	SD	137	
Labidura truncate	н	SD	131	
Cymindis sp.	Pr	SD	93	
	De	SD	93	
Sciara sp.				
Hister sp.	Pr	SD	78	
Micraspis discolor Fabr.	Pr	SD	70	
Lumbricoides sp.	De	SD	46	
Monomorium minimum	0	SD	43	
Platyzosteria nitidella	0	SD	39	
Callibaetis sp.	0	SD	35	
Phaenicia sericata	De	SD	35	
Adrisa sp.	Pr	SD	19	
Draeculacephala mollipes	Н	SD	17	
Heppelates sp.	н	SD	15	
	Н	SD	15	
Ctenicera recolendens		30		
		50	0	
Brachymeria sp.	Ра	SD	8	
Brachymeria sp. Euxoa excellens	Pa H	SD	8	
Brachymeria sp. Euxoa excellens Thiara sp.	Pa H De	SD SD	8 7	
Brachymeria sp. Euxoa excellens Thiara sp.	Pa H	SD	8	
Brachymeria sp. Euxoa excellens Thiara sp. Chortoicetus sp.	Pa H De	SD SD	8 7	
Brachymeria sp. Euxoa excellens Thiara sp. Chortoicetus sp. Musca domestica	Pa H De O	SD SD SD	8 7 6	
Ctenicera resplendens Brachymeria sp. Euxoa excellens Thiara sp. Chortoicetus sp. Musca domestica Anoplolepis longipes Onthophagus sp.	Pa H De O De	SD SD SD SD	8 7 6 4	

Species	FG	ID	Abundance
Pachycondyla sp.	0	SD	1
Ploiaria regina	Pr	SD	1
Tetragnatha mandibulata	Pr	SD	1
white grub	н	SD	1
Total # of SD Indiv	viduals		14,995
Total # of SD_S	36		
Anomalomyrma taylori	0	SF	292
Lumbricoides sp.	De	SF	222
Solenopsis globularia	0	SF	67
Nabis ferus	Pr	SF	60
Thiara sp.	De	SF	53
Oxidus gracilis	De	SF	47
Adrisa sp.	Pr	SF	42
Oligomyrmex nitidulus	0	SF	37
Anoplolepis longipes	0	SF	36
white grub	Ĥ	SF	31
Xylion sp.	Pr	SF	31
Cerapachys augustae	0	SF	25
Pheidole megacephala	õ	SF	16
Monomorium minimum	0	SF	12
Oxyopes sp.	Pr	SF	12
Sericesthis geminate	Pr	SF	12
Titanolabis colossea(Dohrn)	Pr	SF SF	12
Paratrechina longicormis	0	SF	 
Cylisticus convexus	De	SF SF	6
Blatella sp.	0	SF SF	5
Nomadacris gutturosa	U H	SF SF	5
		SF SF	2
Pselaphus sp.	Pr		2
Geophilomorpha sp.	Pr Pr	SF	3
Hesperus sp.		SF	3
Platyzosteria nitidella	0	SF	3
Coniontis sp.	Н	SF	2
Dermestes sp.	Н	SF	2
Geophilus electricus	Pr	SF	2
Pinophilus sp.	Pr	SF	2
Cryptopone mjobergi	0	SF	1
Cymindis sp.	Pr	SF	1
Nomius pygmaeus	Pr	SF	1
Onthophagus declivis	Н	SF	1
Stegobium panicerum	Pr	SF	1
Talia sp.	0	SF	1
Trogoderma sp.	H	SF	1
Total # of SF Indiv	viduals		1,059
Total # of SF S	pecies		36

GRAND TOTAL OF INVERBRATE INDIVIDUALS ACCOUNTED=21,639

The non-*Bt* corn plots had the highest non-target invertebrate abundance and species richness, while the *Bt* corn plots had the lowest abundance and species richness (Table 5). This same pattern emerged for the aerial, surface, and soil dwellers. The apparent significance of the difference for the aerial dwellers did not hold under Bonferroni correction for multiple models (Table 6), but may indicate relatively large differences in this group. The lowest abundance of aerial dwellers was found for *Bt*HT corn (Table 6).

Our best fitting models show that soil pH, potassium and nitrogen have significant influence on invertebrate abundance and that nitrogen also has a slight effect on species richness. However, the effects of soil chemical characteristics did not take away the effect of corn variety. Non-*Bt* corn has highest abundance and species richness (Table 5).

Regarding the effects of confounding variables on different invertebrate dwellers (Table 6), the best fitting models show a high effect of soil pH and a slight effect of soil organic matter content on the aerial abundance but greater effect was manifested of corn variety. For aerial species richness, herbicides manifested a slight influence but greater effect is manifested of corn variety.

The pesticide application rate was highest in *Bt* cornfields, with herbicides applied most frequently in *Bt*HT and insecticides most frequently in non-*Bt* cornfields. However, application of pesticides did not differ significantly between the corn varieties (Table 7, Fig. 3).

**Table 5**. Results of mixed regression analyses (REML) of abundance and species richness of all invertebrates, with corn variety, soil pH, soil nitrogen (N), and soil potassium (K) contents, as confounding variable/fixed factors and field within site within sampling method as random factor. Mean abundance per dweller was ln(x+1) transformed. SD = standard deviation. P-values in italics are of contrasts. Sign B: significance after Bonferroni correction: \*\* = equivalent to p<0.01; \* = equivalent to p<0.05; (\*) = equivalent to p<0.10).

	Mean± SD	F value	p-value	Sign B
I. Total Abundance				
Variate				
Corn variety		7.384	0.0011	*
- Non-Bt corn (Intercept)	3.490 ± 1.405			
- Bt corn	$3.146 \pm 1.491$		0.0019	*
- BtHT corn	3.162 ± 1.382		0.3553	ns
Covariates				
Soil pH		6.311	0.0126	*
Ν		6.080	0.0143	*
InK		7.333	0.0072	*
II. Total Species Richness				
Variate				
Corn variety		4.129	0.0197	*
- Non-Bt corn (Intercept)	7.511 ± 4.129			
- Bt corn	6.824 ± 3.266		0.0069	*
- BtHT corn	7.028 ± 3.113		0.2455	ns
Covariates				
Ν		4.616	0.0326	(*)

#### Table 6. next Page (page 85):

**Table 6**. Results of mixed regression analyses (REML) of abundance and species richness of aerial, surface and soil invertebrates, with corn variety, corn isolines, weed cover (WC), plant height (PH), soil texture, insecticide, herbicide, ln(x+1) field size (lnAr), ln(x+1) field elevation (lnElev), field longitude (Longi) and latitude (Lat), soil organic matter (OM), soil pH, soil nitrogen (N), soil phosphorus (P) and soil potassium (K) contents as confounding variable/fixed factors and field within site as random factor. Only the best fitted models are given. Mean abundance per dweller was ln(x+1) transformed. SD = standard deviation. P-values in italics are of contrasts. Sign B: significance after Bonferroni correction: \*\* = equivalent to p<0.01; \* = equivalent to p<0.05; (\*) = equivalent to p<0.10).

	Mean± SD	F-value	p-value	Sign B
I. ABUNDANCE				-
1. Aerial Dweller				
Variate				
Corn variety		5.635	0.0099	*
- Non-Bt corn (Intercept)	3.885 ± 0.568			
- Bt corn	3.500 ± 0.540		0.0050	*
- BtHT corn	$3.360 \pm 0.380$		0.0412	ns
Covariates	0.000 - 0.000		0.0.112	
OM		4.132	0.0533	ns
pH		5.796	0.0181	(*)
		5.750	0.0101	
2. Surface Dweller				
Variate				
Corn variety		0.931	0.4123	ns
- Non-Bt corn (Intercept)	4.688 ± 0.625			
- Bt corn	4.619 ± 0.515		0.2635	ns
- <i>Bt</i> HT corn	4.500 ± 0.486		0.0614	ns
Covariates				
Isolines		3.648	0.0126	(*)
рН		8.573	0.0043	*
InElev		12.594	0.0023	*
Longi		6.622	0.0191	(*)
3. Soil Dweller		0.011	0.0101	
Variate				
Corn variety		4.161	0.0275	ns
- Non-Bt corn (Intercept)	$1.897 \pm 1.035$			
- Bt corn	$1.320 \pm 0.672$		0.0365	ns
- BtHT corn	$1.625 \pm 1.068$		0.0144	(*)
Covariates				
InK		6.237	0.0144	(*)
II. SPECIES RICHNESS				
1. Aerial Dweller				
Variate				
Corn variety		5.787	0.0089	*
- Non-Bt corn (Intercept)	7.967 ± 1.965			
- Bt corn	6.556 ± 2.335		0.3061	ns
- BtHT corn	7.250 ± 1.700		0.6839	ns
Covariates				
Herbicides		7.512	0.0114	(*)
2. Surface Dweller				
Variate				
		1 266	0 2021	ns
Corn variety	10 450 / 2 424	1.366	0.2921	
- Non-Bt corn (Intercept)	10.450 ± 2.134		0.2025	25
- Bt corn	10.528 ± 2.360		0.2635	ns
- BtHT corn	9.792 ± 2.519		0.0614	ns
Covariates				
Isolines		5.324	0.0058	*
ST		4.520	0.0242	(*)
рН		11.716	0.0009	*
InElev		3.049	0.1063	ns
Longi		2.767	0.1221	ns
Lat		9.519	0.0094	(*)
3. Soil Dweller				
Variate			0.4704	20
Corn variety		2.034	0.1731	ns
- Non-Bt corn (Intercept)	4.117 ± 1.795			
- Bt corn	3.389 ± 1.572		0.4525	ns
- BtHT corn	4.042 ± 1.829		0.7277	ns
Covariates				
Isolines		2.194	0.0981	ns
		2.034	0.1731	ns
PH				
PH N				ns
PH N Insecticides		1.295 1.310	0.2582 0.2692	ns ns

**Table 7**. Results of mixed logistic regression analyses (GLMM) of pesticide use, with corn variety as fixed factor and site as random factor. (n.d.f.: numerator of degrees of freedom; d.d.f.: denominator of degrees of freedom.

Pesticide use models	Wald stat.	n.d.f.	F	d.d.f.	р	
All pesticides ~ Corn variety	2.07	2	1.03	27.0	0.369	
Herbicides ~ Corn variety	6.26	2	3.11	24.1	0.063	
Insecticides ~ Corn variety	0.89	2	0.44	27.0	0.646	

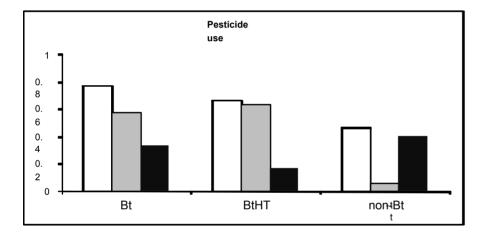


Figure 3. Estimated frequency of pesticide use per corn variety (GLMM, table 6). Open bars: all pesticides; grey bars: herbicides; black bars: insecticides

#### Functional guilds

For all functional guilds except parasitoids species, the highest abundance and species richness were recorded in non-*Bt* cornfields. Parasitoids were found to have higher species richness in *Bt* corn though not significantly different from other corn variety. Omnivores and detritivores were shown to have relatively large differences, although the differences were not statistically significant under Bonferroni correction for multiple testing (Table 8).

**Table 8.** Results of mixed regression analyses (REML) of abundance and species richness of all herbivore, detrivore, omnivore, predator, and parasitic invertebrates, with corn variety, corn isolines, weed cover (WC), plant height (PH), soil texture, herbicide, ln(x+1) field elevation (InElev), soil organic matter (OM), soil pH, soil nitrogen (N), soil phosphorus (P) and soil potassium (K) contents, as confounding variable/fixed factors and field within site within sampling method as random factor. P values: \*\*\* = p<0.001, \*\* = p<0.01, \* = <0.05, (\*) = <0.10. Abundance and species richness of all the functional guilds were ln(x+1) transformed. SD = standard deviation. P-values in italics are of contrasts. Sign B: significance after Bonferroni correction: \*\* = equivalent to p<0.01; \* = equivalent to p<0.05; (\*) = equivalent to p<0.10).

	Mean± SD	F value	p-value	Sign B
. ABUNDANCE				
1. Predator				
Variate				
Corn variety		1.182	0.3126	ns
- Non-Bt corn (Intercept)	1.725 ± 1.322			
- Bt corn	$1.566 \pm 1.354$		0.0671	ns
- <i>Bt</i> HT corn	$1.601 \pm 1.367$		0.8956	ns
Covariates				
Isolines		2.264	0.0468	ns
PH		3.882	0.0527	ns
InElev		2.863	0.0950	ns
2. Herbivores				
Variate				
Corn variety		1.636	0.2013	ns
Non-Bt corn (Intercept)	$1.400 \pm 1.181$			
- Bt corn	$1.255 \pm 1.322$		0.0796	ns
- <i>Bt</i> HT corn	$1.175 \pm 1.104$		0.9088	ns
Covariates				
рН		2.102	0.1483	ns
InP		2.422	0.1208	ns
3. Omnivores				
Variate				
Corn variety		3.170	0.0478	ns
- Non-Bt corn (Intercept)	$1.426 \pm 1.376$			
- Bt corn	$1.198 \pm 1.209$		0.0026	(*)
- BtHT corn	$1.152 \pm 1.192$		0.2586	ns
Covariates				
ST		2.911	0.0400	ns
OM		2.634	0.1088	ns
рН		4.019	0.0460	ns
InK		6.417	0.0119	ns
Herbicide		9.639	0.0027	*
4. Detrivores				
Variate				
Corn variety		2.494	0.0892	ns
- Non-Bt corn (Intercept)	1.982 ± 1.256			
- Bt corn	1.786 ± 1.194		0.0336	ns
- <i>Bt</i> HT corn	1.725 ± 1.187		0.1064	ns
Covariates				
InElev		3.687	0.0585	ns
5. Parasitoids				
Variate				
Corn variety		1.327	0.2711	ns
Non-Bt corn (Intercept)	$0.114 \pm 0.348$			
- Bt corn	0.102 ± 0.296		0.5362	ns
- <i>Bt</i> HT corn	$0.044 \pm 0.188$		0.0818	ns
Covariates				
Ν		4.335	0.0383	ns
InK		2.613	0.1072	ns

84

	Mean± SD	F value	p-value	Sign B
II. SPECIES RICHNESS				
1. Predator				
Variate				
Corn variety		1.811	0.1708	ns
- Non-Bt corn (Intercept)	$0.960 \pm 0.580$			
- Bt corn	$0.879 \pm 0.681$		0.4368	ns
- BtHT corn	0.874 ± 0.597		0.6065	ns
Covariates				
Isolines		2.111	0.0622	ns
2. Herbivores				
Variate				
Corn variety		2.713	0.0725	ns
- Non-Bt corn (Intercept)	$0.750 \pm 0.513$			
- <i>Bt</i> corn	0.632 ± 0.509		0.0105	ns
- <i>Bt</i> HT corn	0.703 ± 0.540		0.6550	ns
Covariates				
рН		8.047	0.0049	*
Ν		2.432	0.1201	ns
3. Omnivores				
Variate				
Corn variety		2.043	0.1365	ns
- Non-Bt corn (Intercept)	0.695 ± 0.621			
- Bt corn	$0.616 \pm 0.616$		0.0147	ns
- <i>Bt</i> HT corn	0.661 ± 0.604		0.3720	ns
Covariates				
Herbicide		2.272	0.1358	ns
4. Detritivores				
Variate				
Corn variety		3.020	0.0545	ns
- Non-Bt corn (Intercept)	1.076 ± 0.458			
- Bt corn	$0.961 \pm 0.478$		0.0375	ns
- <i>Bt</i> HT corn	$0.961 \pm 0.538$		0.0663	ns
5. Parasitoids				
Variate				
Corn variety		1.419	0.2480	ns
- Non-Bt corn (Intercept)	0.085 ± 0.239			
- Bt corn	0.087 ± 0.240		0.8632	ns
- BtHT corn	0.038 ± 0.160		0.1061	ns
Covariates				
N		4.857	0.0284	ns

#### Discussion

#### Growth stages

The high abundance of non-target aerial invertebrates in the vegetative stage and the low abundance in the tasselling stage can be attributed to plant height and density of canopy. These corn plant characteristics limit the flight of aerial dwellers only in the inner parts of the cornfields, whereas aerial dwellers can move freely in a field with an open and lower density of canopy field. This result supports the study by Alston *et al.* (1991) which found larger Corn Earworm, *Helicoverpa zea* (Boddi) and larval populations in more open canopies. Similarly, the within-plant distribution of Fall Armyworm, *Spodoptera frugiperda* (J.E. Smith) was highest in the pre-tasselling stage, mostly in the wrapped leaves of the whorl (Labate 1993). The findings of the survey by Hagen *et al.* (2010) using fly traps also coincides with our result, with more abundant aerial arthropods recorded in both native and invaded forests with lower canopies.

#### Abundance and species richness

This study demonstrates that the abundance and species richness of all non-target invertebrates are slightly lower in Bt cornfields than in non-Bt cornfields. Pesticide input did not confound these results, i.e., the abundance and species richness remained lowest in Bt cornfields, whether or not pesticides were used. These results confirm the study by Marvier et al. (2007), albeit that in their case the adverse effects on non-target invertebrates were only demonstrated for experimental Bt fields under equal management as the non-Bt control fields, i.e. involving no pesticide use. It should be noted however that the adverse effects of *Bt* corn in our study were weak. In addition, increasing or decreasing pH and nitrogen compositions of the soil directly or indirectly favored invertebrate abundance which is probably due to the response of species to the presence of these chemicals or pH state supportive to their physiological needs. A study by Fischer and Führer (1990) showed that soil acidity plays a major role in the nematode's ability to parasitize Cephalcia nymphs and soil with pH levels below 4.0 may limit the nematode's host-finding. Moreover, invertebrates can tolerate soil acidity at different ranges like termites which are most tolerant to acidity with maximum abundance at pH 4 to 6 and coleopteran larvae are abundant only in soils with high pH whilst ants are not affected by soil acidity (Lavelle et al., 1995). Also, as stated in Lavelle et al. (1995), invertebrates are abundant and active population may exist in soil with pH 3.8 to 4.0.

#### On invertebrate dwellers

Abundance and species richness of non-target surface-dwelling invertebrates were not affected by transgenic corn varieties. This finding supports previous studies (Toschki *et al.*, 2007; Peterson *et al.*, 2011) in temperate regions, which concluded that some non-target species such as carabid beetles and spiders were not affected by *Bt* corn. The results of the current study further indicate that aerial and soil-dwelling species are more susceptible to *Bt* corn. The transgenic *Bt* and *Bt*HT corn had lower abundance and species richness of aerial species, and we also found a nearly significantly lower abundance of soil-dwelling species, though significance was lost after Bonferroni correction. The unintended effects of *Bt* toxin on the abundance and species richness of the aerial- and soil dwelling invertebrates other than the target pest species (ACB) may be caused by numerous factors. One conceivable reason is the accumulation of *Bt* pollen in the axils of the leaves and its deposition within the cornfield (Hansen-Jesse and Obrycki, 2000; Li *et al.*, 2005). Furthermore, the ability of aerial dwellers to move from one plant to another may expose them to high concentration of *Bt* toxin in wind-dispersed pollen even outside fields (Koziel *et al.*, 1993; Fearing *et al.*, 1997). Although Obrist *et al.* (2006) found no or negligible amounts of Cry1Ab protein in some predators (hemerobiids, *Nabis sp.*, *Hippodamia sp.*, *Demetrias sp.*), the dilution of protein in the animal body may vary between species and groups of organisms. *Bt* toxin has been detected in aphids, a herbivore, with a mean concentration of 2.0±0.8 ppb (Burgio *et al.*, 2007).

Cry1Ab delta-endotoxin, the active component of *Bt* corn seeds in the Philippines, persists in the soil, and can still be detected after 240 days in tillage and 200 days in no tillage cornfields (Zwahlen *et al.*, 2003). After four consecutive years of *Bt* corn cultivation, the Cry1Ab protein can be detected even in the rhizosphere soil (Icoz *et al.*, 2008). Soil properties of cornfields may also play a critical role in Cry1Ab protein absorption. An increase in the amount of surface clay particles and a decrease in organic carbon content will lead to an increased absorption capacity (Nguyen and Jehle, 2007). The fields we surveyed consist of clayey soils, and the organic carbon contents in the *Bt*HT and *Bt* cornfields are lower than in those in the non-*Bt* cornfields (Table 2). The persistence and absorption of Cry1Ab protein in the field soils (Saxena and Stotzky, 2000; Saxena *et al.*, 2002) may have been one of the factors causing the decreased abundance of non-target aerial- and soil- dwelling invertebrates in the *Bt*HT and *Bt* cornfields.

Lastly, as noted from our analysis on confounding variables, some soil chemical properties of the surveyed cornfield could potentially affect the aerial or foliage dwelling invertebrates. Since foliage dwellers, mostly herbivores, are not feeding on the soil but feed on plant parts, hence the effect is considered to be indirect. Our result was supported by previous studies done by Prudic *et al.* (2005) and Kinney *et al.* (1997). Their studies showed that alteration of soil chemical properties can have indirect effects to the insect's performance. The alteration of soil chemistry can modify plant chemistry as well as the performances of insects particularly the herbivory insects (Prudic *et al.*, 2005; Kinney *et al.*, 1997). In particular, Prudic *et al.* (2005) showed that fertilized fields increased the availability of soil nutrients which in turn caused the host-plant's foliar nitrogen to increase and its chemical defense against pests to decrease. Also, Kinney *et al.* (1997) found out that the feeding performance of the Penultimate gypsy moth larvae (*Lymantria dispar*) can be affected by elevated  $CO_2$  and  $NO_3$  in the soil. The increased soil  $CO_2$  and/or low  $NO_3$  caused the plants to increase the concentrations of starch, condensed tannins and ellagitannins increased which can affect larvae feeding performance.

#### Guilds

Among the functional guilds recorded in the *Bt* cornfields, the omnivores seem to be affected the most in terms of abundance and species richness. This may be explained by the way *Bt* toxin spreads through the food web. Groot and Dicke (2002), for example, refer to direct effects of *Bt* corn when non-target invertebrates feed on plant parts containing the toxin, or indirect effects

when they prey on herbivores containing the toxin. These effects also vary between species, depending on differences in Bt toxin ingestion (Head et al., 2001; Raps et al., 2001; Obrist et al., 2005). In addition, ingestion is not the only way in which non-target species can be affected. Experiments by Prasifka et al. (2007), for instance, attributed the decreased weight and feeding habits of monarch butterfly larvae to the avoidance behavior of larvae when exposed to Bt expressing anthers. In the meta-analysis by Wolfenbarger et al. (2008), omnivores were more abundant in insecticide-sprayed non-Bt corn than in non-sprayed Bt corn, and the high abundance of omnivores, mostly soil-dwelling, was associated with a reduction in the population of predators, greatly affected by insecticide spraying. Overall, they found that the pesticide effect was stronger than the Bt corn effect. In the current study, we found no difference in the influence of pesticides between corn types. Hence, the low abundance of omnivores in Bt corn indicates that other causal factors must be taken into account. Omnivores are phytophagous as well as entomophagous invertebrates. Their ability to change prey and feed on plant materials allows them to survive in an environment inimical to specialized entomophagous invertebrates (Coll and Guershon, 2002). However, this also makes them more susceptible to toxin exposure. Nonetheless, further research is needed to uncover the mechanisms.

#### Conclusion

The study shows that long-term and continuous cultivation of transgenic corn has an impact on humid tropical corn-based agro-ecosystems, in terms of reducing the abundance and species richness of non-target invertebrates. Our results seem to contradict earlier studies in temperate regions, where endotoxin from *Bt* and *Bt*HT corn affected only the targeted pest species (ACB) (Candolfi *et al.*, 2004). As large-scale monocropping of transgenic corn is currently highly prevalent in the Philippines, precautionary measures or effective refuge strategies should be considered to abate serious implications for the biodiversity and sustainability of corn agro-ecosystems. Nonetheless, this study suggests that more research is needed to enable continuous monitoring and to address some emanating ecological issues about recently released *Bt*, *Bt*HT and HT corn lines.

#### Acknowledgements

We would like to thank the Louwes scholarship program of the Oxford and Leiden Universities, through the Cagayan Valley Program for Environment and Development (CVPED), for financially assisting this research. We are grateful to the former CVPED Coordinators Dr. A.B. Masipiqueña and Drs. M. van Weerd for their field assistance and technical advice. We also owe a debt of gratitude to Mr. V. Samaritan of the Entomology Division of the National Museum of the Philippines for the identification of invertebrate species, to Mr. H. van Mill for his assistance with the statistical analysis and to Mr. E. Gertenaar (Institute of Environmental Sciences) for facilitating the access to sticky traps. Finally, we would like to thank Dr. WL.M. B. Tamis for his suggestions about sampling techniques for surface-dwelling invertebrates.

#### References

- Afidchao, M.M., Musters, C.J.M., Masipiqueňa, M.D. and de Snoo, G.R., Chapter 3. Field assessment of the impact of genetically modified (GM) corn cultivation and its associated agricultural practices on in-field invertebrate populations in the Philippines.
- Alcantara, E., 2004. Monitoring insect abundance and diversity in *Bt* corn. *In:* Impact assessment of *Bt* corn in the Philippines. Terminal Report. International Service for the Acquisition of Agribiotech Applications (ISAAA), UPLB, Philippines.
- Alcantara, E.P., Mostoles, M.D.J., Caoili, B.L. and Javier, P.J., 2008 Arthropod abundance and diversity in commercial *Bt* corn farms and adjacent riparian areas in the Philippines. Poster for XXIII International Congress of Entomology, Durban, South Arica. July 6-12.
- Alston, D.G., Bradley, J.R., Schmitt, D.P. and Coble, H.D., 1991. Response of *Helicoverpa zea* (lepidoptera, noctuidae) populations to canopy development in soybean as influenced by *Heterodera glycines* (nematoda, heteroderidae) and annual weed population-densities. Journal of Economic Entomology 84(1): 267-276.
- Anonymous, 2006. DA-CVIARC development zone. Updated physical corn area per province. Summary report. Provincial Agricultural Statistics (PAS), Isabela, Philippines.
- Burgio, G., Lanzoni, A., Accinelli, G., Dinelli, G., Bonetti, A., Marotti, I. and Ramilli, F., 2007. Evaluation of *Bt*-toxin uptake by the non-target herbivore, *Myzus persicae* (Hemiptera: Aphididae), feeding on transgenic oilseed rape. Bulletin of Entomological Research 97: 211-215.
- Candolfi, M.P., Brown, K., Grimm, C., Reber, B. and Schmidli, H., 2004. A faunistic approach to assess potential side-effects of genetically modified *Bt*-Corn on non-target arthropods under field conditions. Biocontrol Science and Technology 14(2): 129-170.
- Coll, M. and Guershon, M., 2002. Omnivory in terrestrial arthropods: mixing plant and prey diets. Annual Review of Entomology 47: 267-297.
- Dillehay, B.L., Roth, G.W., Calvin, D.D., Kratochvil, R.J., Kuldau, G.A. and Hyde, J.A., 2004. Performance of *Bt* corn hybrids, their near isolines, and leading corn hybrids in Pennsylvania and Maryland. Agronomy Journal 96: 818-824.
- Fearing, P.L., Brown, D., Vlachos, D., Meghji, M. and Privalle, L., 1997. Quantitative analysis of CryIAb expression in *Bt* maize plants, tissues, and silage and stability of expression over successive generations. Molecular Breeding 3: 169-176.
- Fischer, P. and Führer, E., 1990. Effect of soil acidity on the entomophilic nematode *Steinernema kraussei* Steiner. Biology and Fertility of Soils 9(2): 174-177.
- Folcher, L., Delos, M., Marengue, E., Jarry, M., Weissenberger, A., Eychenne, N., Regnault-Roger, C., 2010. Lower mycotoxin levels in *Bt* maize grain. Agronomy for Sustainable Development 30(4): 711-719.
- Franzmeier, D.P. and Owens, P.R., 2008. Soil texture estimates: A tool to compare texture-by-feel and lab data. Journal of Natural Resources and Life Sciences Education 37: 111-116.
- Gonzales, L.A., Javier, E.Q., Ramirez, D.A., Cariño, F.A. and Baria, F.A., 2009. Modern Biotechnology and Agriculture: A history of the commercialization of biotech maize in the Philippines. Book Publication of STRIVE Foundation. ISBN 978-971-91904-8-6.
- Groot, A.T. and Dicke, M., 2002. Insect-resistant transgenic plants in a multi-trophic context. Plant Journal 31: 387-406.
- Hagen, E.N., Bakker, J.D. and Gara, R.I., 2010. Aerial arthropod communities of native and invaded forests, Robinson Crusoe Island, Chile. Environmental Entomology 39(4): 1159-1164.

- Hammond, B.G., Campbell, K.W., Pilcher, K.W., Degooyer, T.A., Robinson, A.E., McMillen, B.L., Spangler, S.M., Riordan, S.G., Rice, L.G. and Richard, J.L., 2004. Lower fumonisin mycotoxin levels in the grain of *Bt* corn grown in the United States in 2000-2002. Journal of Agricultural and Food Chemistry 52(5): 1390-1397.
- Hansen-Jesse, L., and Obrycki, J.J., 2000. Field deposition of *Bt* transgenic corn pollen: lethal effects on the monarch butterfly. Oecologia 125(2): 241-248.
- Head, G., Brown, C.R., Groth, M.E. and Duan, J.J., 2001. Cry1Ab protein levels in phytophagous insects feeding on transgenic corn: Implications for secondary exposure risk assessment. Entomologia experimentalis et applicata 99: 37-45.
- Icoz, I. and Stotzky, G., 2008. Cry3Bb1 protein from *Bacillus thuringiensis* in root exudates and biomass of transgenic corn does not persist in soil. Transgenic Research 17(4): 609-620.
- James, C., 2011. Global status of commercialized biotech/GM crops: ISAAA Brief No. 43-2011, Ithaca, NY.
- James, C., 2010. Global status of commercialized biotech/GM crops: ISAAA Brief No. 420, Ithaca, NY.
- Javier, P.A., 2004. Bt corn is safe to beneficial arthropods. AgriNotes College of Agriculture, UPLB, Philippines.
- Kinney, K.K., Lindroth, R.L., Jung, S.M. and Nordheim, E.V., 1997. Effects of CO<sub>2</sub> and NO<sub>3</sub> availability on deciduous trees: phytochemistry and insect performance. Ecology 78: 215–230.
- Koziel, M.G., Beland, G.L., Bowman, C., Carozzi, N.B., Crenshaw, R., Crossland, L., Dawson, J., Desai, N., Hill, M. and Kadwell, S., 1993. Field performance of elite transgenic maize plants expressing an insecticidal protein derived from *Bacillus thuringiensis*. Nature Biotechnology 11: 194-200.
- Labate, J.M., 1993. Within-plant distribution of Fall Armyworm (Lepidoptera: Noctuidae) larvae on corn during whorl-stage infestation. Florida Entomologist 76(3): 437-447.
- Lang, A. and Vojtech, E., 2006. The effects of pollen consumption of transgenic *Bt* maize on the Common Swallowtail, *Papilio machaon* L. (Lepidoptera, Papilionidae). Basic and Applied Ecology 7(4): 296-306.
- Lavelle, P., Chauvel, A. and Fragoso, C., 1995. Faunal activity in acid soils. *In*: R.A. Date et al. (eds). Plant Soil Interactions at Low pH. 201-211. 1995 Kluwer Academic Publishers Printed in Netherlands.
- Li, W.D., Wu, K.M., Wang, X.Q., Wang, G.R. and Guo, Y.Y., 2005. Impact of pollen grains from *Bt* transgenic corn on the growth and development of Chinese tussah silkworm, *Antheraea pernyi* (Lepidoptera: Saturniidae). Environmental Entomology 34(4): 922-928.
- Lu, Y., Wu, K., Jiang, Y., Xia, B., Li, P., Feng, H., Wyckhuys, K. and Guo, Y., 2010. Mirid bugs outbreaks in multiple crops correlated with wide-scale adoption of *Bt* cotton in China. Science 328: 1151-1154.
- Mahaney, W.C., Milner, M.W., Sanmugadas, K., Hancock, R.G.V., Aufreiter, S., Wrangham, R.W. and Pier, H.W., 1997. Analysis of geophagy soils in Kibale forest, Uganda. Primates 38: 159-176.
- Marvier, M., McCreedy, C., Regetz, J. and Kareiva, P., 2007. A meta-analysis of effects of *Bt* cotton and maize on nontarget invertebrates. Science 316(5830): 1475-1477.
- Meissle, M., Vojtech, E. and Poppy, G.M., 2005a. Effects of *Bt* maize on the herbivore *Spodoptera littoralis* (Lepidoptera: Noctuidae) and the parasitoid *Cotesia marginiventris* (Hymenoptera: Braconidae). Transgenic Research 14(2): 133-144.
- Meissle, M., Vojtech, E. and Poppy, G.M., 2005b. Effects of *Bt* maize-fed prey on the generalist predator *Poecilus cupretis* L. (Coleoptera: Carabidae). Transgenic Research 14(2): 23-132.
- Nguyen, H.T. and Jehle, J.A., 2007. Quantitative analysis of the seasonal and tissue specific expression of Cry1Ab in transgenic maize Mon 810. Journal of Plant Diseases and Protection 114(2): 82-87.
- Obrist, L.B., Dutton, A., Albajes, R. and Bigler, F., 2006. Exposure of arthropod predators to Cry1Ab toxin in *Bt* maize fields. Ecological Entomology 31(2): 143-154.

- Obrist, L.B., Klein, H., Dutton, A. and Bigler, F., 2005. Effects of *Bt* maize on *Frankliniella tenuicornis* and exposure to prey-mediated *Bt* toxin. Entomologia experimentalis et applicata 115: 409-416.
- Peterson, J.A., Lundgren, J.G. and Harwood, J.D., 2011. Interactions of transgenic *Bacillus thuringiensis* insecticidal crops with spiders (Araneae). Journal of Arachnology 39(1): 1-21.
- Prasifka, P.L., Hellmich, R.L., Prasifka, J.R. and Lewis, L.C., 2007. Effects of cry1Ab-expressing corn anthers on the movement of monarch butterfly larvae. Environmental Entomology 36(1): 228-233.
- Prasifka, J.R., Hellmich, R.L., Dively, G.P. and Lewis, L.C., 2005. Assessing the effects of pest management on non-target arthropods: The influence of plot size and isolation. Environmental Entomology 34(5): 1181-1192.
- Prudic, K.L., Oliver, J.C. and Bowers, M.D., 2005. Soil nutrient effects on oviposition preference, larval performance, and chemical defense of a specialist insect herbivore. Oecologia, 143 (4): 578-587.
- Qaim, M. and Zilberman, D., 2003. Yield effects of genetically modified crops in developing countries. Science 299(5608): 900-902.
- Quinn, G.P. and Keough, M.J., 2007. Experimental design and data analysis for biologists. Book CambridgeUniversity Press, Cambridge, UK. ISBN 0521811287.
- Raps, A., Kehr, J., Gugerli, P., Moar, W.J. and Bigler, F., 2001. Detection of cry 1Ab in phloem sap of *Bacillus thuringiensis* corn and in the selected herbivores *Rhopalosiphum padi* (Homoptera: Aphidae) and *Spodoptera littoralis* (Lepidoptera: Noctuidae). Molecular Ecology 10: 525-533.
- Rauschen, S., Schultheis, E., Hunfeld, H., Schaarschmidt, F., Schuphan, I. and Eber, S., 2010. Diabroticaresistant *Bt*-maize DKc5143 event MON88017 has no impact on the field densities of the leafhopper *Zyginidia scutellaris*. Environmental Biosafety Research 9: 87-99.
- Rauschen, S., Schultheis, E., Wieder, S.P., Schuphan, I. and Eber, S., 2009. Impact of *Bt*-corn MON88017 in comparison to three conventional lines on *Trigonotylus caelestialium* (Kirkaldy) (Heteroptera: Miridae) field densities. Transgenic Research 18: 203-214.
- Rice, M.E., 2004. Transgenic rootworm corn: Assessing potential agronomic, economic, and environmental benefits. Online. *Plant Health Progress* doi:10.1094/PHP-2004-0301-01-RV.
- Romero, R.R., Desneux, N., Decourtye, A., Chaffiol and Pham-Delègue, M.H., 2008. Does cry1Ab protein affect learning performances of the honey bee *Apis mellifera* L. (Hymenoptera, Apidae)? Ecotoxicology and Environmental Safety 70(2): 327-333.
- Romero, R., Bernal, J.S., Chaufaux, J. and Kaiser, L., 2007. Impact assessment of *Bt*-maize on a moth parasitoid, *Cotesia marginiventris* (Hymenoptera: Braconidae), via host exposure to purified Cry1Ab protein or *Bt*-plants. Crop Protection 26: 953-962.
- Saxena, D., Flores, S. and Stotzky, G., 2002. *Bt* toxin is released in root exudates from 12 transgenic corn hybrids representing three transformation events. Soil Biology & Biochemistry 34: 133-137.
- Saxena, D. and Stotzky, G., 2000. Insecticidal toxin from *Bacillus thuringiensis* is released from roots of transgenic *Bt* corn in vitro and in situ. Federation of European Microbiological Societies Microbiology Ecology 33: 35-39.
- Snelder, D.J., Masipiqueña, M.D. and Snoo de, G.R., 2008. Risk assessment of pesticide usage by smallholder farmers in the Cagayan Valley (Philippines). Crop Protection 27: 747-762.
- Stanger, T.F., and Lauer, J.G., 2006. Optimum plant population of *Bt* and non-*Bt* corn in Wisconsin. Agronomy Journal 98: 914-921.
- Toschki, A., Hothorn, L.A. and Ross-Nickoll, M., 2007. Effects of cultivation of genetically modified *Bt* maize on epigeic arthropods (Araneae; Carabidae). Environmental Entomology 36: 967-981.

- Wolfenbarger, L.L., Naranjo, S.E., Lundgren, J.G., Bitzer, R.J., Watrud, L.S., 2008. *Bt* crop effects on functional guilds of non-target arthropods: A meta-analysis. PLoS ONE 3(5):e2118.
- Williams, G.M., Kroes, R. and Munro, I.C., 2000. Safety evaluation and risk assessment of the herbicide Roundup and its active ingredient, Glyphosate, for humans. Regulatory Toxicology and Pharmacology 31(21): 117-165.
- Wu, F., 2007. *Bt* corn and impact on mycotoxins. CAB Reviews: Perspectives in Agriculture, Veterinary Science, Nutrition and Natural Resources 2(60): 1-6.
- Yorobe, J.M. Jr. and Quicoy, C.B., 2006. Economic impact of *Bt* corn in the Philippines. The Philippine Agricultural Scientist 89(3): 258-267.
- Zwahlen, C., Hilbeck, A., Howald, R. and Nentwig, W., 2003. Effects of transgenic *Bt* corn litter on the earthworm *Lumbricus terrestris*. Molecular Ecology 12(4): 1077-1086.