



Universiteit
Leiden
The Netherlands

The interplay between cholesterol and inflammation in the evolution of atherosclerosis

Verschuren, L.

Citation

Verschuren, L. (2009, January 22). *The interplay between cholesterol and inflammation in the evolution of atherosclerosis*. Retrieved from <https://hdl.handle.net/1887/13415>

Version: Corrected Publisher's Version

License: [Licence agreement concerning inclusion of doctoral thesis in the Institutional Repository of the University of Leiden](#)

Downloaded from: <https://hdl.handle.net/1887/13415>

Note: To cite this publication please use the final published version (if applicable).

Chapter 1
General Introduction

1.1	Introduction	10
1.2	Role of cholesterol and lipoproteins in atherogenesis	11
1.3	Role of inflammation in atherogenesis	12
1.4	Selected targets for treatment of atherosclerosis	13
	1.4.1 HMG-CoA-reductase	
	1.4.2 Nuclear hormone receptors	
	Peroxisome proliferator activated receptor (PPAR)	
	Liver-X-Receptor (LXR)	
	1.4.3 Macrophage Migration Inhibitory Factor (MIF)	
1.5	Mouse models for inflammation and atherosclerosis	17
	1.5.1 Human C-reactive protein (huCRP) transgenic mouse model	
	1.5.2 ApoE*3Leiden (E3L) transgenic mouse model	
	1.5.3 LDL receptor-deficient (Ldlr ^{-/-}) mouse model	
1.6	Scope, aims and outline of thesis	19

1.1 Introduction

Despite significant advances in treatment and in understanding of its biology, coronary atherosclerosis remains the leading cause of morbidity and mortality of men and women in industrialized societies. Hypercholesterolemia, particularly of low-density lipoprotein (LDL) cholesterol and very low-density lipoprotein (VLDL) cholesterol, is a well-established risk factor of atherosclerosis and its pathologic complications, myocardial infarction and stroke. For the past 20 years, treatment of atherosclerosis was mainly directed at normalizing plasma cholesterol levels and the statin class of cholesterol-lowering drugs has been the mainstay for the treatment of hypercholesterolemia (¹, and references therein). However, despite effectively lowering cholesterol levels and reducing cardiovascular causes of death, two thirds of the statin-treated patients still experience adverse coronary events.² Consequently, in atherosclerosis research there is increasing attention to risk factors other than hypercholesterolemia. Also, atherosclerosis is now recognized as a multifactorial, multistep disease with numerous etiologies which have to act in concert to initiate and promote the atherosclerotic process.³

In addition to “classical” risk factors discovered by classical epidemiology, including hypercholesterolemia, hypertriglyceridemia, low high-density lipoprotein (HDL), hypertension, insulin resistance and type-2 diabetes, inflammation has been accepted as a major driving force in atherosclerotic lesion development.⁴ There is increasing experimental evidence that (chronic) inflammation plays a causal role in atherosclerosis. Many studies have shown that raised levels of circulating inflammatory markers, and in particular C-reactive protein (CRP), are prospectively associated with increased cardiovascular disease risk.^{5,6} As in many human inflammatory diseases, the inciting stimulus in each case remains uncertain. Also, the intricate interaction between cholesterol and inflammation in the pathogenesis of atherosclerosis is not exactly understood.

The general theme of this thesis is to further delineate the role of inflammation in the atherosclerotic process and to elucidate the interplay between cholesterol and inflammation in atherosclerotic lesion formation and progression. In this general introduction, first some background information is provided regarding the relevance of cholesterol and inflammation in atherogenesis. Next, some potential anti-atherosclerotic targets are described that have been studied in this thesis with respect to their effect on plasma cholesterol levels, systemic inflammation, and/or the local (vascular) inflammatory state. Subsequently, mouse models for inflammatory processes and atherosclerosis used in our studies are introduced. Finally, the scope, aims and outline of the thesis are given.

1.2 Role of cholesterol and lipoproteins in atherogenesis

Cholesterol and triglycerides are important lipids in human physiology and required for many physiological processes. For example, cholesterol plays an important role in the regulation of the fluidity and barrier function of cell membranes and is a precursor of bile acids and steroid hormones. The most important physiological role of triglycerides is to transport and store energy. Triglycerides can either be stored in adipose tissue (storage) or can be used to generate ATP by tissues that rely on lipids as energy source (e.g. cardiac and skeletal muscle). The uptake and transport of the lipids cholesterol and triglycerides is coupled as discussed briefly in the following.

characteristic	chylomicron	very-low-density lipoproteins	intermediate-density lipoproteins	low-density lipoproteins	high-density lipoproteins
abbreviation		VLDL	IDL	LDL	HDL
density (g/ml)	<0.95	0.95 - 1.006	1.006 - 1.019	1.019 - 1.063	1.063 - 1.210
diameter (nm)	75-1200	30-80	25-35	18-25	5-12
composition (% dry wt)					
protein	2	8	19	22	47
triglycerides	86	55	23	6	4
cholesterol	5	19	38	50	19
phospholipid	7	18	20	22	30
apolipoproteins	A1, A2 B-48 C1, C2, C3 E	B-100 C1, C2, C3 E	B-100 C1, C2, C3 E	B-100	A1, A2 C1, C2, C3 E

Table 1. Physical properties and composition of human plasma lipoproteins

Most of the endogenous pool of cholesterol/cholesterol esters and triglycerides is taken up in the intestine. Since cholesterol and triglycerides are hydrophobic molecules they have to be transported in the form of soluble lipoproteins which can differ with respect to density, size, apolipoprotein composition and electrical charge (Table 1): chylomicrons, very low density lipoproteins (VLDL), low density lipoproteins (LDL), intermediate density lipoproteins (IDL), and high density lipoproteins (HDL).^{7,8}

In the intestine, dietary cholesterol and triglycerides are packed into chylomicrons which enter the circulation via the lymph. Chylomicrons are the main transporters of dietary lipids towards the liver, while VLDL, IDL, LDL and HDL function to transport endogenous lipids throughout the body.⁹ The liver plays an important role in the homeostasis of plasma lipoproteins and is the major assembling site of VLDL. Based on lipoprotein composition, lipoproteins can be divided into an apoB-containing lipoprotein group (VLDL, IDL, and

LDL) and a non-apoB-containing group (HDL). In particular, the apoB-containing lipoproteins are important for the delivery of cholesterol from the liver to peripheral tissues, whereas HDL appears to mediate the reverse process of movement of cholesterol from tissues back to the liver.¹⁰

Epidemiological studies have established that elevated levels of apoB-containing lipoproteins in humans are associated with an increase in the incidence of cardiovascular disease.^{11, 12} Enzymatic modifications, enhanced retention and accumulation of these lipoproteins in the arterial wall are generally considered to be the first steps in the development of atherosclerosis. The end-product of enzymatic modification of LDL, oxLDL, acts as a potent pro-inflammatory chemoattractant for macrophages and T lymphocytes in the vasculature. Furthermore, oxLDL is cytotoxic for endothelial cells and stimulates the release of soluble inflammatory molecules, thus creating an inflamed environment in the human vasculature.¹³

1.3 Role of inflammation in atherogenesis

It is now well-accepted that atherosclerosis is not only a lipid disorder but also an inflammatory disease. Inflammatory processes have been shown to play important roles at all stages of the disease process. In fact, as depicted in Figure 1, inflammatory processes play a role in all stages of atherogenesis, from early endothelial activation to progression and, eventually, plaque rupture.^{4,14} In early atherogenesis for example, adhesion molecules are upregulated on endothelial cells in response to inflammatory cytokines. Circulating monocytes can then attach to the activated endothelium cell layer and can migrate into the vessel wall.

Within the vessel wall, transformation of monocytes to macrophages takes place, followed by engulfment of oxidized LDL, resulting in foam cells. The lipid-laden foam cells further promote inflammation and atherosclerotic plaque progression via secretion of inflammatory mediators that attract additional leukocytes including T-cells. The newly attracted and immigrated T-cells secrete cytokines which subsequently further amplify the inflammatory response and promote the migration and proliferation of intimal smooth muscle cells. This self-perpetuating cycle further amplifies inflammation thereby accelerating the atherogenic process.¹⁶ Later in the atherogenic process, inflammation-induced proteases can weaken the protective fibrous cap of the atheroma, ultimately leading to plaque rupture, thrombosis and the occurrence of acute coronary syndromes such as unstable angina pectoris and myocardial infarction (MI).

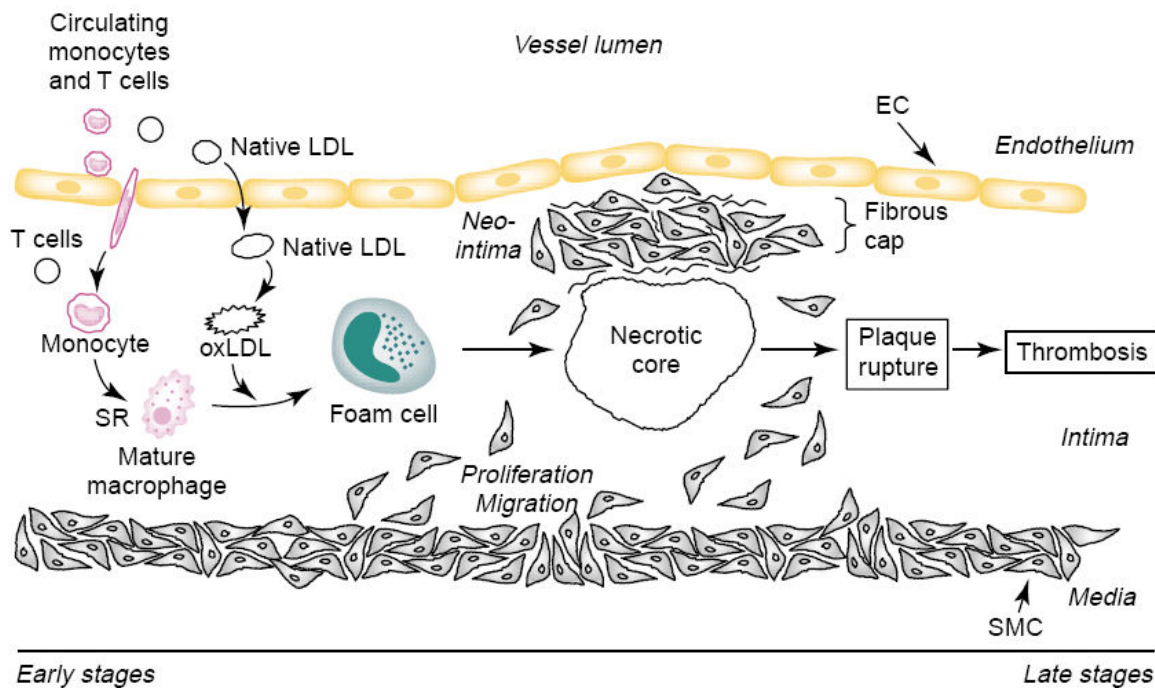


Figure 1. Schematic overview of inflammatory processes in various stages of atherosclerosis. ¹⁵

In the past several years it has become clear that the inflammatory component of atherogenesis is not restricted to local inflammation in the vascular wall but that inflammation originating from other organs such as the liver and the adipose tissue can contribute significantly to progression of atherosclerotic disease.¹⁷ This is also underlined by data describing increased incidence of cardiovascular disease in patients suffering from chronic inflammatory diseases such as systemic lupus erythematosus¹⁸ and rheumatoid arthritis.^{19,20}

1.4 Selected targets for treatment of atherosclerosis

1.4.1 HMG-CoA-reductase

Most circulating cholesterol is synthesized in the body, primarily from acetyl-CoA through the 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase pathway. Statins form a class of hypolipidemic drugs used to lower cholesterol levels in humans with or at risk of cardiovascular disease.²¹ They lower cholesterol by inhibiting the enzyme HMG-CoA reductase, which catalyzes -what is thought to be- the rate-limiting step in cholesterol biosynthesis, the formation of mevalonate.^{22,23} Reduction of mevalonate synthesis leads to reduction in the regulatory sterol pool, which in turn causes up-regulation of HMG-CoA reductase²⁴ and other enzymes of the cholesterol biosynthesis pathway^{25, 26}, and, most

importantly, the LDL receptor.^{27,28} Although not the original target, the induction of the LDL receptor expression is crucial for the effectiveness of statins in lowering cholesterol.

In the past several years, very powerful new statins have been developed, such as rosuvastatin and atorvastatin. These statins have a higher affinity for HMG-CoA reductase and – because of different chemical properties – a different bioavailability than the first generation statins. Molecular differences among the statins also give rise to some important differences in their properties, including their anti-atherogenic and anti-inflammatory actions (among the so-called pleiotropic effects).

1.4.2 Nuclear hormone receptors

The nuclear hormone receptor superfamily of ligand-activated transcription factors regulates gene expression in such diverse processes as metabolism, development, and reproduction. The family has 48 members in humans, and includes, for example, retinoid, steroid, and thyroid hormone receptors.

The structure of nuclear hormone receptors (see Figure 2) consists of a variable NH2 terminal region and a highly conserved DNA-binding domain (DBD).^{29,30} The DBD region contains a short motif responsible for DNA binding-specificity and is involved in dimerization of nuclear receptors. A linker region is situated between the DBD and the ligand binding domain (LBD). This region functions as a flexible hinge and contains the nuclear localization signal. The LBD is involved in the binding of ligand. This domain also contains a ligand-regulated transcriptional activation function-2 (AF-2) necessary for binding transcriptional co-activators which interact, among others, with the general transcriptional activation machinery. Most nuclear hormone receptors contain in the N-terminal variable domain a transcriptional activation function-1 (AF-1) that mediates ligand-independent transcriptional activation.

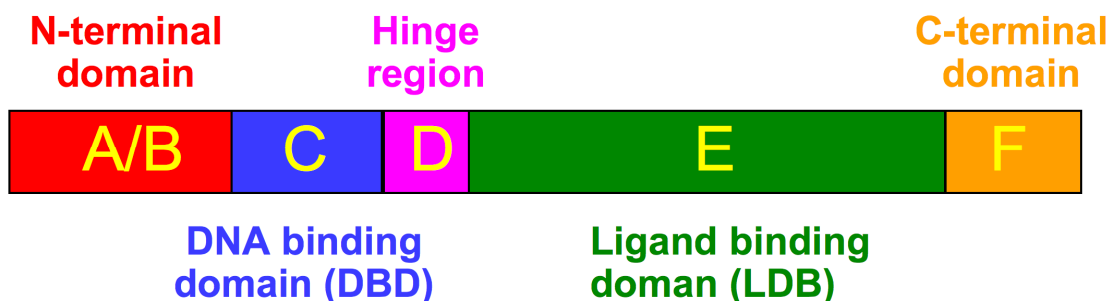


Figure 2: Schematic 1D amino acid sequence of a nuclear receptor.

The subfamilies known as peroxisome proliferator-activated receptor (PPAR) and liver-X-receptor (LXR) have emerged as dominant regulators of processes that influence cardiovascular risk, namely various aspects of lipid and glucose metabolism, insulin sensitivity, as well as inflammation.³¹⁻³⁶

Notably, PPAR and LXR not only show these effects at the systemic level but also regulate lipid homeostasis and inflammation in macrophages, endothelial cells, and smooth muscle cells within the vessel wall. Agonists that specifically activate these receptors may therefore retard the development of atherosclerosis at several levels.³⁷

Peroxisome Proliferator-Activated Receptor (PPAR)

PPARs are transcriptional factors belonging to the ligand-activated nuclear receptor superfamily.³⁸ There are three distinct PPAR subtypes, PPAR α , PPAR β/δ , and PPAR γ . Although there is overlap in natural ligands (fatty acids, eicosanoids) that are capable of activating the three PPARs, each receptor subtype has a tissue-specific expression pattern and exhibits both overlapping and distinct biological activities.^{1,39}

In rodents and humans, PPAR α is expressed in tissues involved in fatty acid oxidation, including liver, kidney, heart, skeletal muscle and brown fat⁴⁰ and in a range of vascular cells such as endothelial cells⁴¹, vascular smooth muscle cells (VSMCs)⁴² and monocytes/macrophages.⁴³

When activated by its ligand, PPAR α positively regulates the transcription of target genes through binding to specific gene promoter response elements (Figure 3). This mode of action is particularly important for the regulation of genes that control lipid and lipoprotein metabolism.⁴⁴ Nowadays, synthetic PPAR α activators such as fibrates are clinically used to lower plasma lipid levels and treat hypertriglyceridemia and mixed dyslipidemia.⁴⁵ There is a large body of clinical evidence that fibrates lower plasma triglycerides and LDL cholesterol, and increase HDL cholesterol by up-regulating the hepatic gene expression and synthesis of ApoA1, the major apolipoprotein of HDL.⁴⁵ Independent of this mechanism, activation of PPAR α can also act as a negative regulator of pro-inflammatory genes via antagonizing the activity of inflammatory transcription factors or co-activators (Figure 3).⁴⁶

Liver X Receptor (LXR)

The liver X receptors are members of the nuclear receptor family of proteins that are critical for the control of lipid homeostasis in virtually all vertebrates.^{47,48} Two LXR isoforms

have been described so far, LXR α and LXR β . LXR β has a ubiquitous tissue distribution, whereas LXR α predominates in liver, adipose tissue, and intestinal tissue, as well as macrophages. Both LXR α and LXR β are activated by physiological concentrations of sterol metabolites such as 22(R)-hydroxycholesterol, 24(S)-hydroxycholesterol, 27-hydroxycholesterol, and 24(S), 25-epoxycholesterol.⁴⁹⁻⁵¹ Both isoforms appear to respond to the same natural and synthetic ligands. When activated by its ligand, LXR heterodimerize with RXR and positively regulates the transcription of target genes through binding to specific gene promoter response elements.

LXR activation has several reported beneficial effects on the expression of genes that are involved in cholesterol efflux in macrophages⁵², hepatic bile acid synthesis⁴⁸, intestinal cholesterol absorption⁵³, inflammation⁵⁴, and glucose tolerance⁵⁵. These features make LXRs attractive targets for the development of drugs for treatment of cardiovascular, metabolic and/or inflammatory diseases.

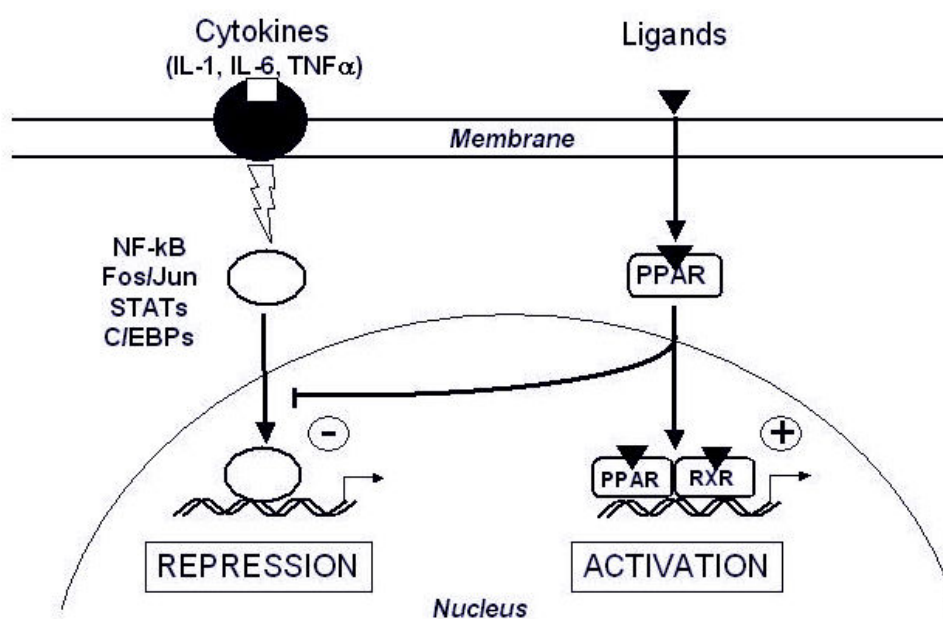


Figure 3: Mechanisms of transcriptional regulation by PPARs. Following activation, PPARs heterodimerize with retinoic X receptor (RXR) and bind to response elements in the promoter region of target genes, thereby activating their transcription. PPARs also repress gene transcription in a DNA-binding independent fashion by interfering directly with transcription factors or co-activators.

1.4.3 Macrophage Migration Inhibitory Factor (MIF)

MIF was discovered over 40 years ago and constitutes one of the first cytokines described in literature. In these early studies, MIF was found to inhibit the random migration of peritoneal macrophages which gave rise to its name.⁵⁶ In 1993, the protein was rediscovered as a pituitary-derived factor with hormone-like properties and a mediator of

endotoxic shock.⁵⁷ Today, MIF is known as a pleiotropic macrophage and T-cell cytokine⁵⁸ and endocrine factor⁵⁹ that is involved in the regulation of a great number of inflammatory processes^{57, 60} and inflammatory diseases.⁶¹ Recent studies have demonstrated that MIF expression is up-regulated in atherosclerotic lesions in humans⁶² and animals.⁶³ In the vessel wall, increased MIF expression stimulates cells to produce pro-inflammatory cytokines such as $\text{TNF}\alpha$ and $\text{IL-1}\beta$.^{58, 60} Increased $\text{TNF}\alpha$ and $\text{IL-1}\beta$ levels in turn stimulate vascular cells to produce substantial amounts of MIF which is a good example for a self-perpetuating cycle in the vasculature that accelerates the atherogenic process.¹⁶ Interference in this cycle by inhibition or blockage of MIF activity could retard the progression of atherosclerotic disease, but also other diseases in which this inflammatory cycle is of relevance (e.g. insulin resistance).

1.5 Mouse models

Detailed analyses of processes that contribute to human disease development are often hampered by the complex interactions and variation in environmental and genetic factors in humans. Therefore, mice with homogenous genetic background (inbred strains) that are housed in a standardized way (comparable environmental factors) have been proven to be a useful tool. Since regular wild-type mice such as C57BL/6 mice differ from humans with respect to lipid metabolism and inflammation (i.e. the two major risk factors of cardiovascular disease), transgenic mice have been developed that allowed us to better study the disease process and the risk factors. In the following, three models will be discussed in more detail: 1) Human CRP transgenic mice as a model to study the risk factor inflammation; 2) ApoE*3Leiden transgenic mice and 3) *Ldlr*^{-/-} mice to study atherogenesis (lipid metabolism and inflammation).

1.5.1 Human C - reactive protein transgenic mice

Several clinical studies have shown that C-reactive protein (CRP), a liver-derived inflammation marker, is the strongest independent predictor of future cardiovascular events (summarized by Libby et al.⁴ and Ridker et al.⁶⁴). In response to infection or inflammation, CRP is highly induced in man but not in wild-type mice, where its serum levels never raise above 2-3mg/l.⁶⁵ To overcome this difference in response, transgenic mice have been generated that carry the full human CRP gene. This so called “human-CRP transgenic (huCRP_{tg})” mouse carries a 31-kb human DNA fragment containing the human *CRP* gene including all known *cis*-acting regulatory elements including the entire

human CRP promoter. Unlike its wild-type mouse counterpart, human CRP behaves as a major acute phase reactant when introduced into the mouse genome.⁶⁶ The expression of huCRP is highly inducible and tissue specific. The generation of this mouse model enabled us to monitor inflammatory responses in mice.

1.5.2 ApoE*3Leiden (E3L) transgenic mice

The APOE*3Leiden mutation is a rare, dominant negative mutation in the human *APOE3* gene. It is characterized by a tandem duplication of codons 120-126 and associated with familial dysbetalipoproteinemia in humans. ApoE*3Leiden transgenic (E3L) mice have been generated by introducing a human *APOE*3Leiden* gene construct into C57Bl/6 mice. Besides the *APOE*3Leiden* gene, this construct consists of the *APOC1* gene and a promoter element that regulates the expression of *APOE* and *APOC1* genes (^{67,68}, and references therein). Although E3L mice still express endogenous ApoE protein, the clearance of ApoE-containing lipoproteins is impaired, albeit less dramatically than in ApoE^{-/-} mice. The introduction of the *APOC1* gene may further increase plasma lipid levels by diminished VLDL uptake through the LDL receptor and LRP.

E3L mice show significant elevations of plasma cholesterol and triglycerides on a regular chow diet and are, in contrast to wild-type mice, highly responsive to fat-, sugar-, and cholesterol-containing diets, resulting in strongly elevated plasma cholesterol and triglyceride levels, with a prominent increase in VLDL- and LDL-sized lipoprotein fractions.⁶⁷ Plasma lipid levels can easily be adjusted to a desired concentration by titrating the amount of cholesterol and sugar in the diet. As compared to ApoE^{-/-} and Ldlr^{-/-} mice, E3L mice represent a moderate mouse model for hyperlipidemia (cholesterol levels on chow are about 2 mM and do not exceed 25 mM on a high-cholesterol diet). In addition, the plasma cholesterol and triglyceride levels respond strongly to changes in hepatic VLDL production. Therefore, drugs and diets influencing the chylomicron and VLDL production show parallel effects on plasma cholesterol and triglyceride levels. In this respect, E3L mice are more sensitive than ApoE^{-/-} and Ldlr^{-/-} mice and respond to hypolipidemic compounds with cholesterol-lowering.

E3L mice develop atherosclerotic lesions with all the characteristics of human vascular pathology, varying from fatty streak to mild, moderate and severe plaques. Atherosclerosis development starts at the aortic root and progresses along the entire arterial tree in a time-dependent fashion.⁶⁹

1.5.3 LDL receptor-deficient (*Ldlr*^{-/-}) mice

In humans, mutations in the gene for the LDL-receptor (LDLr) cause familial hypercholesterolemia. Mice lacking the gene for LDLr display a modestly elevated plasma cholesterol level when maintained on a regular chow diet (about 5 mmol/l versus 2 mmol/l in wild-type animals), and they develop atherosclerosis only slowly. On HC diet feeding, *Ldlr*^{-/-} mice show strongly elevated plasma cholesterol (>25mmol/l) and rapid development of atherosclerosis.⁷⁰ The plasma lipoprotein profile of *Ldlr*^{-/-} mice resembles that of humans, with the cholesterol being confined mainly to the LDL fraction.

The morphology of the lesions in *Ldlr*^{-/-} mice is comparable to that in *ApoE*^{-/-} and *ApoE**3Leiden mice, with the plaques developing in a time-dependent manner, starting from the proximal aorta. Quantification of atherosclerosis in *Ldlr*^{-/-} mice shows mild to moderate atherosclerotic lesions in the aortic root and coronary arteries of mice fed a mild atherogenic diet lacking excessive amounts of cholesterol and cholate.⁷¹

Most interestingly, and in contrast with the *ApoE**3Leiden transgenic mouse model, the *Ldlr*^{-/-} mouse model does also develop diet-induced obesity, hypertriglyceridemia and insulin resistance, in parallel with the development of atherosclerotic lesion⁷².

1.6 Scope, aims and outline of this thesis.

Atherosclerosis is a multifactorial disease of the large arteries and the leading cause of morbidity and mortality in industrialized countries.⁷³ There is ample evidence that hypercholesterolemia (i.e. elevated plasma levels of VLDL and LDL) is a major causative factor in atherogenesis.^{74,75} It is equally clear that atherogenesis has an inflammatory component which is thought to drive the progression of the disease.^{76,77} However, while the lipid component in atherosclerosis development is relatively well-understood, the origin and exact contribution of the inflammatory component remains largely unknown.

The aim of this thesis is to further define and delineate the contribution of the inflammatory component to the atherosclerotic process and to elucidate the link between cholesterol and inflammation in atherosclerotic lesion formation and progression. Since inflammatory markers associated with elevated cardiovascular risk are increased by cholesterol feeding in animal models, particular emphasis is devoted to cholesterol as an inflammation-evoking factor.

In **Chapter 2** we elucidated the link between dietary cholesterol intake and the onset of inflammation in the context of early atherosclerotic lesion formation using *ApoE**3Leiden (E3L) mice. Since cholesterol feeding is associated with elevations in liver-derived

inflammation markers such as CRP in humans, we studied the effect of increasing dietary cholesterol concentrations on liver gene expression (genome-wide transcriptome) and the liver lipid profile (lipidome). Functional systems biology was applied to unravel key inflammatory pathways activated by cholesterol and to identify the molecules that link cholesterol metabolism to inflammation.

To further delineate the relative contribution of lipids versus inflammation to atherogenesis, different approaches have been applied in the following chapters.

First, in **Chapter 3** we investigated compounds with well-established lipid-modulating (anti-atherogenic) effects on their putative anti-inflammatory activity, *viz.* a statin, a PPAR α agonist and an LXR-agonist. To characterize their lipid-independent anti-inflammatory capacity, we have evaluated these compounds in a mouse model for inflammation, huCRP transgenic mice.

Since additional anti-inflammatory effects were observed for all drug classes, one compound per class was further analyzed in the context of atherosclerosis. Dietary experimental conditions were established in the E3L model that enabled us to delineate the contribution of the lipid component and the inflammatory component to the overall atherosclerotic process. In **Chapter 4** we evaluated the pleiotropic anti-inflammatory effects of fenofibrate, a PPAR α activator. The additional contribution of the inflammatory component to the atherosclerotic process could be deduced by comparing of the anti-atherogenic activity of fenofibrate with a dietary cholesterol-matched control group. In **Chapter 5** we analyzed the pleiotropic effects of atorvastatin, a HMG-CoA reductase inhibitor, in the context of lesion progression (*i.e.* the treatment of already established lesions). In **Chapter 6** we studied the anti-inflammatory effects of T0901317, an LXR-agonist in longitudinal studies of lesion progression and regression. Since T0901317 had no plasma cholesterol-lowering effect in E3L mice, this study in particular provided insight into the importance of (cellular) inflammation in atherogenesis.

In **Chapter 7** we used a different approach (genetic deletion of the pro-inflammatory factor Macrophage Migration Inhibitory Factor (MIF)), to study the role of inflammation on atherogenesis. The *Ldlr*^{-/-} model not only enabled us to find further evidence for a role of inflammation alone in atherogenesis, it also allows to extent our findings to another disease area and combine atherosclerosis and insulin resistance. The relevance of MIF in atherosclerotic disease in the human situation was confirmed in **Chapter 8**, in which the involvement of MIF in the occurrence of an atherosclerosis-related disease, abdominal

aortic aneurysm, was evaluated. The general conclusions from the studies performed in this thesis and future perspectives are described in **Chapter 9**.

Reference List

- (1) Li AC, Palinski W. Peroxisome proliferator-activated receptors: how their effects on macrophages can lead to the development of a new drug therapy against atherosclerosis. *Annu Rev Pharmacol Toxicol* 2006;46:1-39.
- (2) Libby P, Aikawa M. Stabilization of atherosclerotic plaques: new mechanisms and clinical targets. *Nat Med* 2002 November;8(11):1257-62.
- (3) Ross R. Atherosclerosis--an inflammatory disease. *N Engl J Med* 1999 January 14;340(2):115-26.
- (4) Libby P, Ridker PM, Maseri A. Inflammation and atherosclerosis. *Circulation* 2002 March 5;105(9):1135-43.
- (5) Wu JT, Wu LL. Linking inflammation and atherogenesis: Soluble markers identified for the detection of risk factors and for early risk assessment. *Clin Chim Acta* 2006 April;366(1-2):74-80.
- (6) Ridker PM, Hennekens CH, Buring JE, Rifai N. C-reactive protein and other markers of inflammation in the prediction of cardiovascular disease in women. *N Engl J Med* 2000 March 23;342(12):836-43.
- (7) Mahley RW, Innerarity TL, Rall SC, Jr., Weisgraber KH. Plasma lipoproteins: apolipoprotein structure and function. *J Lipid Res* 1984 December 1;25(12):1277-94.
- (8) Wasan KM, Cassidy SM. Role of plasma lipoproteins in modifying the biological activity of hydrophobic drugs. *J Pharm Sci* 1998 April;87(4):411-24.
- (9) Beisiegel U. Lipoprotein metabolism. *Eur Heart J* 1998 February;19 Suppl A:A20-A23.
- (10) Cuchel M, Rader DJ. Macrophage reverse cholesterol transport: key to the regression of atherosclerosis. *Circulation* 2006 May 30;113(21):2548-55.
- (11) Proctor SD, Vine DF, Mamo JC. Arterial retention of apolipoprotein B(48)- and B(100)-containing lipoproteins in atherogenesis. *Curr Opin Lipidol* 2002 October;13(5):461-70.
- (12) Phillips NR, Waters D, Havel RJ. Plasma lipoproteins and progression of coronary artery disease evaluated by angiography and clinical events. *Circulation* 1993 December;88(6):2762-70.
- (13) Hurt-Camejo E, Paredes S, Masana L, Camejo G, Sartipy P, Rosengren B, Pedreno J, Vallve JC, Benito P, Wiklund O. Elevated levels of small, low-density lipoprotein with high affinity for arterial matrix components in patients with rheumatoid arthritis: possible contribution of phospholipase A2 to this atherogenic profile. *Arthritis Rheum* 2001 December;44(12):2761-7.
- (14) Lusis AJ. Atherosclerosis. *Nature* 2000 September 14;407(6801):233-41.
- (15) Duval C, Chinetti G, Trottein F, Fruchart JC, Staels B. The role of PPARs in atherosclerosis. *Trends Mol Med* 2002 September;8(9):422-30.
- (16) Raines EW, Ferri N. Thematic review series: The immune system and atherogenesis. Cytokines affecting endothelial and smooth muscle cells in vascular disease. *J Lipid Res* 2005 June;46(6):1081-92.

- (17) Subramanian S, Han CY, Chiba T, McMillen TS, Wang SA, Haw A, III, Kirk EA, O'Brien KD, Chait A. Dietary Cholesterol Worsens Adipose Tissue Macrophage Accumulation and Atherosclerosis in Obese LDL Receptor-Deficient Mice. *Arterioscler Thromb Vasc Biol* 2008 January 31.
- (18) Zampieri S, Iaccarino L, Ghirardello A, Tarricone E, Arienti S, Sarzi-Puttini P, Gambari P, Doria A. Systemic lupus erythematosus, atherosclerosis, and autoantibodies. *Ann N Y Acad Sci* 2005 June;1051:351-61.
- (19) Hahn BH, Grossman J, Chen W, McMahon M. The pathogenesis of atherosclerosis in autoimmune rheumatic diseases: Roles of inflammation and dyslipidemia. *J Autoimmun* 2007 March;28(2-3):69-75.
- (20) Frostegard J. Atherosclerosis in patients with autoimmune disorders. *Arterioscler Thromb Vasc Biol* 2005 September;25(9):1776-85.
- (21) Stroes E. Statins and LDL-cholesterol lowering: an overview. *Curr Med Res Opin* 2005;21 Suppl 6:S9-16.
- (22) Naoumova RP, Marais AD, Mountney J, Firth JC, Rendell NB, Taylor GW, Thompson GR. Plasma mevalonic acid, an index of cholesterol synthesis in vivo, and responsiveness to HMG-CoA reductase inhibitors in familial hypercholesterolaemia. *Atherosclerosis* 1996 January 26;119(2):203-13.
- (23) Pappu AS, Illingworth DR. Contrasting effects of lovastatin and cholestyramine on low-density lipoprotein cholesterol and 24-hour urinary mevalonate excretion in patients with heterozygous familial hypercholesterolemia. *J Lab Clin Med* 1989 November;114(5):554-62.
- (24) Brown MS, Goldstein JL. Multivalent feedback regulation of HMG CoA reductase, a control mechanism coordinating isoprenoid synthesis and cell growth. *J Lipid Res* 1980 July;21(5):505-17.
- (25) Bergstrom JD, Wong GA, Edwards PA, Edmond J. The regulation of acetoacetyl-CoA synthetase activity by modulators of cholesterol synthesis in vivo and the utilization of acetoacetate for cholesterol synthesis. *J Biol Chem* 1984 December 10;259(23):14548-53.
- (26) Balasubramanian S, Goldstein JL, Brown MS. Regulation of cholesterol synthesis in rat adrenal gland through coordinate control of 3-hydroxy-3-methylglutaryl coenzyme A synthase and reductase activities. *Proc Natl Acad Sci U S A* 1977 April;74(4):1421-5.
- (27) Ma PT, Gil G, Sudhof TC, Bilheimer DW, Goldstein JL, Brown MS. Mevinolin, an inhibitor of cholesterol synthesis, induces mRNA for low density lipoprotein receptor in livers of hamsters and rabbits. *Proc Natl Acad Sci U S A* 1986 November;83(21):8370-4.
- (28) Kovanen PT, Bilheimer DW, Goldstein JL, Jaramillo JJ, Brown MS. Regulatory role for hepatic low density lipoprotein receptors in vivo in the dog. *Proc Natl Acad Sci U S A* 1981 February;78(2):1194-8.
- (29) Krust A, Green S, Argos P, Kumar V, Walter P, Bornert JM, Chambon P. The chicken oestrogen receptor sequence: homology with v-erbA and the human oestrogen and glucocorticoid receptors. *EMBO J* 1986 May;5(5):891-7.
- (30) Desvergne B, Wahli W. Peroxisome proliferator-activated receptors: nuclear control of metabolism. *Endocr Rev* 1999 October;20(5):649-88.
- (31) Li AC, Glass CK. 1. *J Lipid Res* 2004 December;45(12):2161-73.
- (32) Barish GD. Peroxisome proliferator-activated receptors and liver X receptors in atherosclerosis and immunity. *J Nutr* 2006 March;136(3):690-4.

- (33) Chen J, Li D, Schaefer RF, Mehta JL. Inhibitory effect of candesartan and rosuvastatin on CD40 and MMPs expression in apo-E knockout mice: novel insights into the role of RAS and dyslipidemia in atherogenesis. *J Cardiovasc Pharmacol* 2004 October;44(4):446-52.
- (34) Zelcer N, Tontonoz P. Liver X receptors as integrators of metabolic and inflammatory signaling. *J Clin Invest* 2006 March;116(3):607-14.
- (35) Geyeregger R, Zeyda M, Stulnig TM. Liver X receptors in cardiovascular and metabolic disease. *Cell Mol Life Sci* 2006 March;63(5):524-39.
- (36) Barish GD, Evans RM. PPARs and LXRs: atherosclerosis goes nuclear. *Trends Endocrinol Metab* 2004 May;15(4):158-65.
- (37) Li AC, Glass CK. PPAR- and LXR-dependent pathways controlling lipid metabolism and the development of atherosclerosis. *J Lipid Res* 2004 December;45(12):2161-73.
- (38) Rizzo G, Fiorucci S. PPARs and other nuclear receptors in inflammation. *Curr Opin Pharmacol* 2006 August;6(4):421-7.
- (39) Li CC, Dai RM, Chen E, Longo DL. Phosphorylation of NF-KB1-p50 is involved in NF-kappa B activation and stable DNA binding. *J Biol Chem* 1994 December 2;269(48):30089-92.
- (40) Ziouzenkova O, Perrey S, Asatryan L, Hwang J, MacNaul KL, Moller DE, Rader DJ, Sevanian A, Zechner R, Hoefler G, Plutzky J. Lipolysis of triglyceride-rich lipoproteins generates PPAR ligands: evidence for an antiinflammatory role for lipoprotein lipase. *Proc Natl Acad Sci U S A* 2003 March 4;100(5):2730-5.
- (41) Braissant O, Fougere F, Scotto C, Dauca M, Wahli W. Differential expression of peroxisome proliferator-activated receptors (PPARs): tissue distribution of PPAR-alpha, -beta, and -gamma in the adult rat. *Endocrinology* 1996 January;137(1):354-66.
- (42) Staels B, Koenig W, Habib A, Merval R, Lebret M, Torra IP, Delerive P, Fadel A, Chinetti G, Fruchart JC, Najib J, Maclouf J, Tedgui A. Activation of human aortic smooth-muscle cells is inhibited by PPARalpha but not by PPARgamma activators. *Nature JID - 0410462* 1998 June 25;393(6687):790-3.
- (43) Inoue I, Shino K, Noji S, Awata T, Katayama S. Expression of peroxisome proliferator-activated receptor alpha (PPAR alpha) in primary cultures of human vascular endothelial cells. *Biochem Biophys Res Commun* 1998 May 19;246(2):370-4.
- (44) Pineda T, I, Gervois P, Staels B. Peroxisome proliferator-activated receptor alpha in metabolic disease, inflammation, atherosclerosis and aging. *Curr Opin Lipidol* 1999 April;10(2):151-9.
- (45) Despres JP, Lemieux I, Robins SJ. Role of fibric acid derivatives in the management of risk factors for coronary heart disease. *Drugs* 2004;64(19):2177-98.
- (46) Kleemann R, Gervois PP, Verschuren L, Staels B, Princen HM, Kooistra T. Fibrates down-regulate IL-1-stimulated C-reactive protein gene expression in hepatocytes by reducing nuclear p50-NFkappa B-C/EBP-beta complex formation. *Blood* 2003 January 15;101(2):545-51.
- (47) Tontonoz P, Mangelsdorf DJ. Liver X receptor signaling pathways in cardiovascular disease. *Mol Endocrinol* 2003 June;17(6):985-93.
- (48) Peet DJ, Turley SD, Ma W, Janowski BA, Lobaccaro JM, Hammer RE, Mangelsdorf DJ. Cholesterol and bile acid metabolism are impaired in mice lacking the nuclear oxysterol receptor LXR alpha. *Cell* 1998 May 29;93(5):693-704.
- (49) Fu X, Menke JG, Chen Y, Zhou G, MacNaul KL, Wright SD, Sparrow CP, Lund EG. 27-hydroxycholesterol is an endogenous ligand for liver X receptor in cholesterol-loaded cells. *J Biol Chem* 2001 October 19;276(42):38378-87.

- (50) Janowski BA, Willy PJ, Devi TR, Falck JR, Mangelsdorf DJ. An oxysterol signalling pathway mediated by the nuclear receptor LXR alpha. *Nature* 1996 October 24;383(6602):728-31.
- (51) Lehmann JM, Kliewer SA, Moore LB, Smith-Oliver TA, Oliver BB, Su JL, Sundseth SS, Winegar DA, Blanchard DE, Spencer TA, Willson TM. Activation of the nuclear receptor LXR by oxysterols defines a new hormone response pathway. *J Biol Chem* 1997 February 7;272(6):3137-40.
- (52) Naik SU, Wang X, Da Silva JS, Jaye M, Macphee CH, Reilly MP, Billheimer JT, Rothblat GH, Rader DJ. Pharmacological activation of liver X receptors promotes reverse cholesterol transport in vivo. *Circulation* 2006 January 3;113(1):90-7.
- (53) Duval C, Touche V, Tailleux A, Fruchart JC, Fievet C, Clavey V, Staels B, Lestavel S. Niemann-Pick C1 like 1 gene expression is down-regulated by LXR activators in the intestine. *Biochem Biophys Res Commun* 2006 February 24;340(4):1259-63.
- (54) Joseph SB, Castrillo A, Laffitte BA, Mangelsdorf DJ, Tontonoz P. Reciprocal regulation of inflammation and lipid metabolism by liver X receptors. *Nat Med* 2003 February;9(2):213-9.
- (55) Laffitte BA, Chao LC, Li J, Walczak R, Hummasti S, Joseph SB, Castrillo A, Wilpitz DC, Mangelsdorf DJ, Collins JL, Saez E, Tontonoz P. Activation of liver X receptor improves glucose tolerance through coordinate regulation of glucose metabolism in liver and adipose tissue. *Proc Natl Acad Sci U S A* 2003 April 29;100(9):5419-24.
- (56) Bloom BR, Bennett B. Mechanism of a reaction in vitro associated with delayed-type hypersensitivity. *Science* 1966 July 1;153(731):80-2.
- (57) Bernhagen J, Calandra T, Mitchell RA, Martin SB, Tracey KJ, Voelter W, Manogue KR, Cerami A, Bucala R. MIF is a pituitary-derived cytokine that potentiates lethal endotoxaemia. *Nature* 1993 October 21;365(6448):756-9.
- (58) Calandra T, Roger T. Macrophage migration inhibitory factor: a regulator of innate immunity. *Nat Rev Immunol* 2003 October;3(10):791-800.
- (59) Calandra T, Bernhagen J, Metz CN, Spiegel LA, Bacher M, Donnelly T, Cerami A, Bucala R. MIF as a glucocorticoid-induced modulator of cytokine production. *Nature* 1995 September 7;377(6544):68-71.
- (60) Calandra T, Bernhagen J, Mitchell RA, Bucala R. The macrophage is an important and previously unrecognized source of macrophage migration inhibitory factor. *J Exp Med* 1994 June 1;179(6):1895-902.
- (61) Lue H, Kleemann R, Calandra T, Roger T, Bernhagen J. Macrophage migration inhibitory factor (MIF): mechanisms of action and role in disease. *Microbes Infect* 2002 April;4(4):449-60.
- (62) Burger-Kentischer A, Goebel H, Seiler R, Fraedrich G, Schaefer HE, Dimmeler S, Kleemann R, Bernhagen J, Ihling C. Expression of macrophage migration inhibitory factor in different stages of human atherosclerosis. *Circulation* 2002 April 2;105(13):1561-6.
- (63) Lin SG, Yu XY, Chen YX, Huang XR, Metz C, Bucala R, Lau CP, Lan HY. De novo expression of macrophage migration inhibitory factor in atherogenesis in rabbits. *Circ Res* 2000 December 8;87(12):1202-8.
- (64) Ridker PM. Clinical application of C-reactive protein for cardiovascular disease detection and prevention. *Circulation* 2003 January 28;107(3):363-9.
- (65) Pepys MB. Isolation of serum amyloid P-component (protein SAP) in the mouse. *Immunology* 1979 July;37(3):637-41.
- (66) Ciliberto G, Arcone R, Wagner EF, Ruther U. Inducible and tissue-specific expression of human C-reactive protein in transgenic mice. *EMBO J* 1987 December 20;6(13):4017-22.

- (67) van Vlijmen BJ, van den Maagdenberg AM, Gijbels MJ, Van Der BH, HogenEsch H, Frants RR, Hofker MH, Havekes LM. Diet-induced hyperlipoproteinemia and atherosclerosis in apolipoprotein E3-Leiden transgenic mice. *J Clin Invest* 1994 April;93(4):1403-10.
- (68) Jong MC, Hofker MH, Havekes LM. Role of ApoCs in lipoprotein metabolism: functional differences between ApoC1, ApoC2, and ApoC3. *Arterioscler Thromb Vasc Biol* 1999 March;19(3):472-84.
- (69) Lutgens E, Daemen M, Kockx M, Doevendans P, Hofker M, Havekes L, Wellens H, de Muinck ED. Atherosclerosis in APOE*3-Leiden transgenic mice: from proliferative to atheromatous stage. *Circulation* 1999 January 19;99(2):276-83.
- (70) Knowles JW, Maeda N. Genetic modifiers of atherosclerosis in mice. *Arterioscler Thromb Vasc Biol* 2000 November;20(11):2336-45.
- (71) Paigen B, Morrow A, Brandon C, Mitchell D, Holmes P. Variation in susceptibility to atherosclerosis among inbred strains of mice. *Atherosclerosis* 1985 October;57(1):65-73.
- (72) Schreyer SA, Vick C, Lystig TC, Mystkowski P, Leboeuf RC. LDL receptor but not apolipoprotein E deficiency increases diet-induced obesity and diabetes in mice. *Am J Physiol Endocrinol Metab* 2002 January;282(1):E207-E214.
- (73) Braunwald E. Shattuck lecture--cardiovascular medicine at the turn of the millennium: triumphs, concerns, and opportunities. *N Engl J Med* 1997 November 6;337(19):1360-9.
- (74) Blum CB, Levy RI. Role of dietary intervention in the primary prevention of coronary heart disease. Individuals with high-normal or elevated serum cholesterol levels should be placed on cholesterol-lowering diets. *Cardiology* 1987;74(1):2-21.
- (75) Steinberg D. Hypercholesterolemia and inflammation in atherogenesis: two sides of the same coin. *Mol Nutr Food Res* 2005 November;49(11):995-8.
- (76) Steinberg D. Atherogenesis in perspective: Hypercholesterolemia and inflammation as partners in crime. *Nat Med* 2002 November;8(11):1211-7.
- (77) Willerson JT, Ridker PM. Inflammation as a cardiovascular risk factor. *Circulation* 2004 June 1;109(21 Suppl 1):II2-10.

