

**Type 1 diabetes, glucocorticoids and the brain: a sweet connection** Revsin, Y.

# Citation

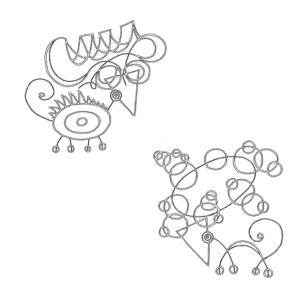
Revsin, Y. (2008, September 17). *Type 1 diabetes, glucocorticoids and the brain: a sweet connection*. Retrieved from https://hdl.handle.net/1887/13211

Version:	Corrected Publisher's Version
License:	<u>Licence agreement concerning inclusion of doctoral thesis in the</u> <u>Institutional Repository of the University of Leiden</u>
Downloaded from:	https://hdl.handle.net/1887/13211

Note: To cite this publication please use the final published version (if applicable).

Chapter 1

# **GENERAL INTRODUCTION**



# Outline

1.	Type 1 diabetes	11
	1.1. Animal Models	12
	1.1.1. Spontaneous models	12
	1.1.1.a. NOD mouse	12
	1.1.2. Experimental models	
	1.1.2.a. STZ mouse	
2.	Hypothalamic-pituitary-adrenal axis and type 1 diabetes	
	2.1.HPA axis	16
	2.1.1. Stress concept	16
	2.2. HPA axis alterations in type 1 diabetes	
3.	Impact of type 1 diabetes	
	3.1. Central nervous system	
	3.2. Behavior	
4.	Scope of the thesis	21
	4.1. Rational and objective	
	4.2. Hypothesis	
	4.3. Questions to address	22
	4.4. Experimental approach	

# 1. Type 1 diabetes

The earliest known record of diabetes dates from 1552 B.C., when the 3<sup>rd</sup> Dynasty Egyptian papyrus by physician Hesy-Ra mentioned polyuria (frequent urination) as a symptom of the disease. Over the years, diabetes was described and studied by Egyptians, Greeks, Chinese, Indians, English, Frenchs, Germans, Czechs, Italians, Canadians, Americans, and others. However, was not until the last century, in 1959, that the two major types of diabetes were recognized: type 1 (insulin-dependent) diabetes and type 2 (non-insulin-dependent) diabetes.

Nowadays diabetes affects over 150 million people worldwide with this number expected to double by 2025; about 90% cases of diabetes are type 2 (Zimmet *et al*, 2001). However, the fraction of type 2 diabetics in different parts of the world varies substantially, almost certainly for environmental and lifestyle reasons, though these are not known in detail. Since type 2 diabetes is not the topic of this thesis, it will not be described at length.

Diabetes mellitus type 2, also called type 2 diabetes, Non Insulin Dependent Diabetes Mellitus (NIDDM) or Adult diabetes, is a metabolic disorder that is primarily characterized by insulin resistance, relative insulin deficiency, and hyperglycemia, and is presently of unknown etiology although there is a strong inheritable genetic connection. About 55% of type 2 are obese (Eberhart *et al*, 2004) -chronic obesity leads to increased insulin resistance that can develop into diabetes, most likely because adipose tissue is a source of chemical signals (hormones and cytokines). Conversely, type 2 diabetes causes obesity (Camastra *et al*, 1999). Additional factors found to increase risk of type 2 diabetes include aging (Jack *et al*, 2004), high-fat diets (Lovejoy, 2002) and a less active lifestyle (Hu, 2003).

Diabetes mellitus type 1, also known as type 1 diabetes (T1D), Insulin Dependent Diabetes Mellitus (IDDM) or Juvenile Diabetes, is an autoimmune disease that results in the permanent destruction or damage of insulin producing beta-cells in the islets of Langerhans of the pancreas. Destruction of these cells leads to insulin deficiency. Therefore, T1D is lethal unless treatment with exogenous insulin via injections replaces the missing hormone. Although, the clinical consequences of the disease have been extensively investigated, the exact cause(s) of T1D are not yet fully understood. Genetic and environmental factors have been suggested to contribute to the etiology of T1D along with other factors (see Box 1 on page 14). So far, the research core on diabetes has focused on the peripheral endocrinology and nervous system. Nowadays, the impact of diabetes on the central nervous system (CNS) is highly recognized but it was not always the case until few decades ago. To study disease initiation, progression, and treatments without exposing humans to unnecessary and potentially unethical risks animal models have been developed. Animal models have contributed important knowledge regarding the study of diabetes. The physiology of mice, rats, and other animals is remarkably conserved in comparison to the human condition. Broad spectrum of animal models of T1D have become available over the last 40 years. They comprise spontaneous models, in which disease develops unprovoked, and experimental models induced by various types of intervention (Table 1, page 15).

## 1.1. Animal models of diabetes

#### 1.1.1. Spontaneous Models

The two major models used are the so-called Bio Breeding (BB) rats and the Non Obese Diabetic (NOD) mice, which develop the disease with similarities to human T1D. These animals derived from inbreeding over many generations by selecting for hyperglycaemia. As a result, many genes and phenotypes have been enriched, but not all will be relevant to the pathophysiology of diabetes, either in rodents or in humans. It is noteworthy that a main advantage of these models is the possibility to study the pre-diabetic state, which is impossible in humans. Other models comprise the Long Evans Tokushima Lean (LETL) rat and the LEW.1AR1/Ztm- iddm rat (Table 1). For sake of clarity, only the model used in the current thesis will be described below.

#### 1.1.1.a. The NOD mouse

The NOD mice were first used in the study of cataract development (i.e. JcI-ICR mouse) (Makino et al, 1980). Insulitis, which is the lymphocytic infiltration of the islets of Langerhans, is present by the time mice reach 4-5 weeks of age. This state is followed by beta-cells destruction and ultimately leads to a drastic decrease in circulating insulin. In the pre-diabetic state (4-5 weeks of age), NOD mice show lower glycemia and higher insulinemia in response to a glucose tolerance test compared with C57Bl/6 control strain (Amrani et al, 1998). Frank diabetes typically begins between 12 and 30 weeks of age. Unlike human T1D, ketoacidosis (metabolic acidosis is caused by high concentrations of ketone bodies and breackdown of fatty acids) is relatively mild and affected animals can survive for weeks without the administration of insulin. In addition and in contrast to the findings of most studies in humans, there is a larger gender difference with 80% of females, but only 50% of males developing diabetes in some colonies (Atkinson and Leiter, 1999). This variation is not surprising knowing that sex steroids are part of the mechanisms underlying the well-recognized immune sexual dimorphism, which is particularly evident in autoimmune diseases (Ansar Ahmed et al, 1985). Moreover, NOD diabetes can be modulated not only by multiple immunotherapeutic agents (Bach 2002), but also by various other factors, including melatonin, insulin growth factor-1 (IGF-1), leptin, insulin and drugs modulating its secretion or sensitivity, and environmental factors such as temperature fluctuations, variations of protein and carbohydrate intake, and stress.

Stressful life events and diabetes onset linkage have been reported in clinical and experimental studies (Homo-Delarche *et al*, 1991; Djarova and; Dube, 1998.). As part of the endocrine response to stress, glucocorticoids exert well-known anti-inflammatory and immunosuppressive actions but also act as counterregulatory hormones inducing hyperglycemia. Therefore, in T1D, glucocorticoids might have both potentially beneficial and deleterious effects.

#### 1.1.2. Experimental models

Most of the experimentally induced models correspond to highly artificial situations far from the conditions in which spontaneous disease develops. However, they have made possible remarkable progresses in understanding the pathogenesis of T1D. Chemically-induce, transgenic, and immunomanipulated mice are among these models. In the following section, the experimental model used in the present thesis is described.

#### 1.1.2.b. Chemically induced T1D: Streptozotocin-induced diabetes mice

Pharmacological methods of inducing T1D by damaging the pancreas also exist. These include the administration of toxins such as streptozotocin (STZ) (Junod et al, 1969) and alloxan. Streptozotocin is a glucosamine-nitrosourea compound isolated from Streptomyces achromogenes with broad-spectrum antibiotic and anti-neoplastic activity (Bono *et al*, 1976). It is a powerful alkylating agent that has been shown to interfere with glucose transport (Wang and Gleichmann, 1998), glucokinase function (Zahner and Malaisse, 1990) and induce multiple DNA strand breaks (Bolzan and Bianchi, 2002). It is taken up into the insulin-producing beta-cells of the islets of Langerhans via the GLUT-2 glucose transporter (Schnedl et al, 1994). The GLUT-2 glucose transporter is absent at the blood-brain barrier (Kumagai, 1999), thus excluding direct effects of STZ on the brain following systemic administration. A single large dose of STZ can produce diabetes in rodents, probably as a result of direct toxic effects. Alternatively, multiple small doses of STZ are used (e.g. 40 mg/kg on five consecutive days) to study the immunological pathways that lead to insulitis and cell death (Mensah-Brown et al, 2002; Holstad and Sandler, 2001). STZ-diabetic rodents are hypoinsulinaemic, but do not require insulin treatment to survive. Blood glucose levels typically are 20-25 mmol/l, which is 5 fold over normal concentration. In rodents, hyperglycemia induces an insulinopenic (lack of insulin) diabetes in which immune destruction plays a role, as in human T1D.

# **Box 1: Etiology of type 1 diabetes**

The origin of the autoimmune process that leads to type 1 diabetes (T1D) involve genetic predisposition (as T1D is known as a hereditary disease on basis of the relatively high rate of familial transmission (1)) and environmental factors, and their interactions, which creates the conditions required for disease onset. The patterns of familial transmission, combined with data from animal models, indicate that the determinism of T1D is polygenic and multifactorial. The search predisposition genes is complex, especially as most if not all predisposition genes appear to be basically "normal" i.e. without mutations or deletions. A fortuitous combination of these genes, together with permissive or triggering environmental factors, provokes the disease. Each of these genes may be present in a large proportion of healthy subjects (notably patient's nondiabetic relatives). the

# *Evidence for the role of environmental factors:*

Several lines of evidence point to a major role of environmental factors in the pathogenesis of T1D. First, more than 60% of identical twins are discordant for the disease, and it is quite unlikely that this is due to differential somatic rearrangement of T cell receptors. Second, disease frequency varies enormously from country to country (2), and these differences cannot simply be explained by ethnic genetic differences since migrants from countries with a low T1D frequency to countries with a high frequency are more susceptible than their compatriots (3). Intriguingly, northern countries are more exposed to the disease than southern countries (2); it will be critical to discover the factor(s) responsible for this

North/South gradient. Third, a number of apparently nonimmunological interventions can increase or decrease the disease rate in animal models: specific diets (low essential fatty acid (4) or protein intake (5, 6)) and several viral infections (7-11) can reduce disease susceptibility in spontaneous models of T1D, the NOD mice and the BB rats, while Kilham's virus (12) and cow's milk (13,14) can increase it in BB rats. These factors, particularly viral infections, probably explain the variations in disease frequency found between NOD colonies (15). Not only do environmental factors seem to influence T1D onset, they can also apparently alter the course of the disease. These factors can be shared by the whole population (climatic factors, hygiene, etc.), or by a given family (e.g. eating habits), or be specific to the individual (e.g. travels and sexual partners). Several studies have focused on many potential environmental factors involved in the etiology of T1D, such as viruses (16-18), bacteria's (19-22), toxic agents (23, 24), food constituents (5, 6, 13, 14), stress (25-28). These factors essentially modulate the expression of predisposing genes, either positively (predisposing factors) or negatively (protective factors). In the case of triggering factors, disease onset is directly related to the encounter with the environmental factor (usually single and limited in time), which can then be considered as the cause of the disease. In the "modulation" hypothesis, the disease can only appear in the fraction of the population at genetic risk and it is on this population that environmental factors (usually multiple and chronic) exert their positive or negative effect. The available data suggests that T1D is of the second type.

# Table 1. Animal models of type 1 diabetes

#### Spontaneous models

- 1. NOD mouse: inbreed strain. Develop Type 1A-Immune Mediated Diabetes. Autoimmune etiology that is heavily influenced by both genetics and environment (1).
- 2. BB rat: inbreed strain. Diabetes in BB rats is also an autoimmune disorder. Substrain BB/Wor has profound T-cell lymphopenia (condition in which there exists an abnormally low number of lymphocytes in the blood) (2).
- 3. Long Evans Tokushima Lean (LETL) Rat: autoimmune T1D (3)
- 4. LEW.1AR1/Ztm- iddm rat: autoimmune T1D, spontaneous mutation within a Mhccongenic LEW.1AR1 colony (4).

## **Experimental models:**

#### Transgenic

- 1. T Cell Receptor (TCR) Tg (transgenic) Mouse: many cell clones isolated from the spleens of diabetic NOD mice, pancreas of pre-diabetic NOD, islet-transplanted diabetic NOD mice, and from islets of NOD mice. Many of these clones have been utilized to produce TCR transgenic (Tg) mice on various backgrounds (5)
- 2. "Humanized" Mice: transgenic expression in mice of human genes (6).

Chemically-induced

- 1. alloxan (7)
- 2. streptozotocin (8)

Immunomanipulation

- 1. thymectomy performed within 2 days after birth can induce a flourishing state of autoimmunity in mice (9).
- 2. adult thymectomy and sublethal irradiation (10, 11).
- 3. athymic rats with transfer of normal spleen cells (12).

# 2. Hypothalamic-pituitary-adrenal axis and type 1 diabetes

## 2.1. HPA axis

The hypothalamic-pituitary-adrenal (HPA) axis refers to a complex set of homeostatic interactions between the hypothalamus (brain area); the pituitary gland (structure located below the hypothalamus), and the adrenal glands (small pair of pyramidal organs located on top of the kidneys). The HPA axis regulates responses to stress and modulates various body processes including growth, metabolism, immune response, mood, reproduction, sexuality, and energy balance. The core of the HPA axis is the paraventricular nucleus of the hypothalamus (PVN). The PVN contains neuroendocrine neurons, the so-called parvocellular neurons, which synthesize and secrete vasopressin (AVP) and corticotropin-releasing hormone (CRH). These two peptides can act in synergy on the anterior lobe of the pituitary gland to stimulate the secretion of the adrenocorticotropic hormone (ACTH) from corticotrope cells. In turn, ACTH enters peripheral circulation where it reaches the adrenal cortex to induce glucocorticoid hormones production (cortisol in humans, corticosterone in rats and mice). Glucocorticoids exert a negative feedback on the PVN and pituitary to suppress CRH and ACTH production, respectively.

Corticosterone is a major stress hormone and has effects on wide arrays of tissues in the body, including the brain. In the brain, corticosterone acts via two types of receptors - mineralocorticoid receptor (MR) and glucocorticoid receptor (GR). These receptors are widely expressed throughout the brain by many different types of cells including neuron and glia. MR and GR have different affinities to glucocorticoids (GCs) with MR showing a greater affinity (10 fold higher) than GR. As a consequence, MR is fully occupied under basal circulating levels whereas GR becomes occupied only when glucocorticoids levels rise above normal. One important target of glucocorticoids is the hippocampus, an area of the limbic system that plays a critical role in memory, learning and spatial navigation. This structure is a major modulator of the HPA axis; hippocampal MR controls the inhibitory tone of this limbic structure on the HPA axis in terms of basal reactivity (Reul et al, 2000). This effect of GCs via MRs is modulated by GRs that become progressively occupied after stress and during the circadian rise of GCs. Therefore, predominant MR activation maintains hippocampal excitability and, through inhibitory projections to the PVN, basal HPA activity. Conversely, with rising GCs concentrations, GR activation suppresses the hippocampal output, resulting in a disinhibition of PVN neurons (de Kloet et al, 1998). In summary, a deficiency in MR is predicted to allow more GC release, thus leading to more pronounced GRmediated effects. Therefore, the functions mediated by both receptor types are linked, and the balance in MR- and GR-mediated effects is important in the HPA regulation.

#### 2.1.1. Stress concept

Stress is the disruption of homeostasis through physical or psychological stimuli. Internal

or external potential disturbances (stressors) activate two systems that serve to normalize the disturbed functions: the rapid sympatho-adrenomedullar system and the slow-acting HPA axis. The activation of the sympathetic branch result in the release of stress hormones including adrenaline from the adrenal medulla. Therefore, activation of noradrenergic neurons leads to temporarily elevated noradrenaline (NA) levels in specific areas of the brain resulting in functional changes of neurons carrying NA receptors. Activation of the HPA system, leads to increase GCs release from the adrenal cortex, which in turn will act in the brain at those sites where its receptors are enriched (Figure 1) (Joëls *et al*, 2007).

Sympathetic nervous output produces the fight-or-flight response, causing the body to divert blood flow to large muscles as the body prepares to run away from or fight something. Lower blood flow is then directed to the digestive system and other organs that do not assist in flying or fighting. Some stressors can cause continual sympathetic nervous system activation with very little opportunity for the parasympathetic nervous system to be activated. The activation of the parasympathetic system stops the fight-flight responses.

Experimental studies have investigated many different types of stressors, and their effects on the HPA axis in many different contexts. Analysis of the literature suggests that different classes of stressor employ different stress circuits. Severe physiologic ("systemic") stress appears to trigger brainstem/circumventricular organ systems that project directly to the PVN. In contrast, stressors requiring interpretation with respect to previous experience ("processive" stressors) reach the PVN by way of multisynaptic limbic pathways. Stressors of the latter category may thus require interaction with homeostatic information prior to promoting an HPA axis response. The HPA stress response thus appears to be a product of both the physiologic importance of the stimulus and the specific pathways a given stimulus excites (Herman *et al*, 1996). The activation of the HPA axis will ultimately trigger GCs secretion. In healthy condition this highly reactive system will turns on and off its responses to stressors. However, if adaptation to stress fails, the stress system responds slowly, or the stress reactions persist, circulating GC levels remain elevated for a prolonged period of time and an enhance vulnerability to disease for which the individual is predisposed may occur (de Kloet and de Rijk, 2004).

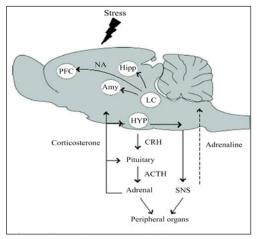


Figure 1: Brain regions activated after stress exposure (Amy: amygdala, Hipp: hippocampus, PFC: prefrontal cortex) and output of these areas through the hypothalamus (HYP). A resulting activation of the fast acting sympatho-adrenomedullar system (right) and the slower acting HPA axis system (left) will affect the function of peripheral organs and feed back to the brain via adrenaline and corticosterone, respectively. Adrenaline will finally rise central release of noradrenaline leves from the locus coeruleous (LC), reaching again the amygdala, prefrontal cortex and hippocampus among other areas. Corticosterone will act on brain areas where its receptors are enriched.

SNS = sympathetic nervous system; ACTH = adrenocorticotropin hormone; CRH = corticotropin releasing hormone.

Reprinted with permission from Joëls and Krugers, Neural Plasticity 2007.

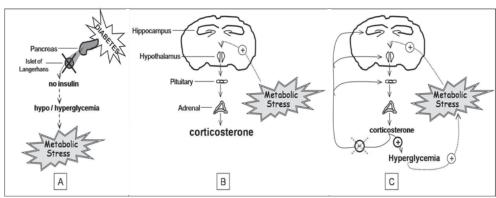
## 2.2. Hypothalamic-pituitary-adrenal axis alterations in type 1 Diabetes

When T1D develops, the insulin producing beta-cells in the islets of Langerhans from the pancreas are destroyed (Figure 2A). Hence, hyperglycemia develops; however, since insulin controls glucose intake, lack of insulin creates of state of cellular starvation. Both hyperglycemia and cellular starvation coexist and, under certain conditions such as T1D, can generate metabolic stress.

The metabolic stress will activate the HPA axis (Figure 2B) in an attempt to restore homoeostasis and recover from metabolic disturbance. The HPA axis activation will ultimately raise basal plasma GCs levels, which will be followed by the shut down of the HPA axis via GCs negative feedback.

However, previous reports in T1D animal models showed GR downregulation in the hippocampus (Tornello *et al*, 1981) and HPA axis hyperactivity (Chan *et al*, 2001 and 2002) (Figure 2C). These results explain, in part, the chronic hypercorticism observed in T1D patients and animals. On the other hand, GCs exert an hyperglycemic effect, inhibiting cellular glucose uptake in the periphery and also in neurons and astrocytes of brain regions such as the hippocampus (Munck *et al*, 1984; Sapolsky, 1992). Therefore, pre-existing hyperglycemia will become chronic state. In this way, continuous metabolic alterations will contribute to the defective shut-off of the stress response. When HPA axis activity is chronically elevated or managed inefficiently, various forms of pathophysiologies are promoted, such as dysfunction of the hippocampus (McEwen 1998; Sapolsky, 1992).

In summary, in agreement with published reports, diabetic animals showed a sustained stimulation of the HPA axis, with elevated basal plasma GC levels. In addition, adrenal hypertrophy, and spleen and thymus atrophy was found; characteristics also of long-lasting exposure to high amounts of GCs (Scribner *et al*, 1993; Oster *et al*, 1988; Bellush, *et al*, 1991). The observation that diabetic animals show a poor shut-off of the stress response (Magarinos and McEwen 2000; Chan *et al*, 2001; McEwen *et al*, 2002) suggests insensitivity to feedback mechanisms and is consistent with the reported resistance to the dexamethasone suppression test (Scribner *et al*, 1993) and downregulation of GR levels in the hippocampus of STZ rats (Tornello *et al*, 1981). In conclusion, processes to adapt to the metabolic imbalance will be created in T1D, involving poor glucose homeostasis, chronically elevated GC levels, increased HPA axis reactivity, and metabolic adjustments. Extreme metabolic adjustments can induce and significantly accelerate hippocampal remodeling, increasing its vulnerability to diabetes, and cognitive dysfunctions.



**Figure 2.** HPA axis alterations in type 1 diabetes. Metabolic stress develops when insulin-producing cells destroy leads to hyperglycemia (A). As a consequence, the activation of the HPA axis triggers an increase in corticosteroids levels (B). The chronic high corticosterone concentration, on one hand exacerbates glucose secretion, and on the other hand acts on the HPA axis in order to shut-off the stress response and restore the homeostasis (C). According to the literature, in type 1 diabetes this response is disrupted.

# 3. Impact of Type 1 Diabetes

## 3.1. Central Nervous System

Diabetic nephropathy, neuropathy, and retinopathy are traditionally considered the late complications of diabetes, whereas CNS was believed to be spared from diabetic complications. However, substantial evidence from clinical and experimental studies demonstrates that diabetes causes primary disease duration-related impairments in CNS function besides secondary sequelae of cerebrovascular events mediated by diabetic macrovascular disease (Li *et al*, 2002; Sima *et al*, 2004). In particular, hyper-and hypoglycaemic episodes may result in acute cerebral dysfunction (Biessels *et al*, 1994; Cryer *et al*, 1994). The consequences of these acute insults to the brain are well recognized. However, there is little knowledge about functional and structural cerebral alterations that develop more insidiously in diabetes. Long-term effects of diabetes on the brain are manifested at the structural, neurophysiological and neuropsychological level. The emerging view is that the diabetic brain features many signs that are best described as accelerated aging.

In diabetic rodents, structural abnormalities including synaptic and neuronal alterations, degeneration, neuronal loss, glycogen accumulation, dilated and fragmented endoplasmic reticulum, increased microtubuli, and irregular nuclei have been demonstrated (Bestetti and Rossi, 1980 and 1982; Garris *et al*, 1982, Luse, 1970, Mukai *et al*, 1980, Magarinos and McEwen 2000, Saravia *et al*, 2001; McEwen 2002). In addition, impaired long-term potentiation in the hippocampus (indicative of pre- and post-synaptic deficits) was reported (Biessels *et al*, 1996). The hippocampus is also vulnerable to damage by stroke and head trauma, susceptible to damage during aging, chronic stress (Sapolsky

1992), and sensitive to the effects of diabetes (Gispen and Biessels, 2000; Magarinos and McEwen 2000; Saravia *et al*, 2002). However, it is also a plastic and adaptable brain region that is capable of considerable structural reorganization. Studies from McEwen (2002) have shown that STZ induction perturbs structural plasticity of the hippocampus and its sensitivity to glucose and oxidative stress. His studies suggest that STZ-diabetes might cause a reduced number of dendrite spines and decreased total length of dendrites of pyramidal neurons of the hippocampus (Magarinos and McEwen, 1995). Other studies by Martinez-Tellez *et al* (2005) showed that dendritic morphological changes also occurs in pyramidal neurons located in structures related to cognitive processes such as, prefrontal cortex, occipital cortex and hippocampus, in the STZ-diabetic rats. The authors suggest that these results together with the available literature, indicate that nitric oxide (NO), GCs, stress, astrogliosis, and glutamate may participate in the dendritic morphological changes.

## 3.2. Behavior

Diabetes-related cognitive dysfunctions were first reported in 1922 (Miles and Root, 1922). Subsequent studies have demonstrated impairments in CNS function. Impairments in learning and memory, problem solving, and intellectual development have been documented in T1D patients (Ryan and Williams, 1993; Ryan *et al*, 1993; Kramer, 1998; Parisi and Uccioli, 2001; McCarthy *et al*, 2002; Schoenle, 2002). Cognitive dysfunction and impaired intellectual development are evident in a duration-related manner in T1D patients independent of hypoglycemic episodes (Kramer, 1998; Schoenle, 2002).

Stress and stress hormones affect different aspects of learning and memory. MR signaling can enhance performance on spatial hippocampal-dependent cognitive tasks (de Kloet *et al*, 1999) and its chronic blockade impairs spatial memory (Douma *et al*, 1998). Decreasing GR signaling attenuates the impairing effects of GR activation on cognition (Nicholas *et al*, 2006). Acute elevation of GCs facilitates the formation of memories of events associated with strong emotions (McGaugh, 2000; Roozendaal, 2000). Chronically, however elevated GC levels contribute to impairment of cognitive function and promote damage to brain structures such as the hippocampus (McEwen and Sapolsky, 1995; Lupien and McEwen, 1997; Sapolsky, 2002). This inverse-U function of GCs is a reflection of the diversity of receptors for GCs in the hippocampus (de kloet *et al*, 1993).

Impaired performances in the Morris water maze are typically observed in STZinduced diabetes rats (Lowy *et al*, 1993; Biessels *et al*, 1996; Lupien and McEwen, 1997) and are associated with impaired LTP in the hippocampus (Biessels *et al*, 1996). It is reported that STZ diabetes does not disturb operant behaviors for food reward (Kaleeswari *et al*, 1986) but facilitates retention of passive avoidance in rats and mice (although not always) (Bellush and Rowland, 1989; Flood *et al*, 1990; Mayer *et al*, 1990). In addition, diabetic rodents consistently displayed performance deficits in more complex learning tasks, such as an active avoidance T-maze, or a Morris water maze depending on the duration of STZ diabetes (Biessels *et al*, 1996 and 1998; Flood *et al*, 1990; Popovic *et al*, 2001). However, discrepancies exist among several behavioral studies and may be partially explained by differences in task complexity, animal models used and duration of diabetes. A key factor, however, appears to be the nature of the stimulus used in behavioral paradigms. There are clear indications that the physiological responses to a novel environment or to stressful stimuli, which are often part of learning paradigms, are larger in STZ-diabetic than in non-diabetic rodents (Bellush *et al*, 1991; Bellush and Rowland, 1989; Flood *et al*, 1990). For example, enhanced retention of simple passive avoidance task in diabetic rodents has been attributed to an increased sensitivity to the foot shock (Bellush and Rowland, 1989, Flood *et al*, 1989, Flood *et al*, 1990).

# 4. Scope of the thesis

## 4.1. Rational and Objective

Peripheral and autonomous neuropathies are well-known and devastating complications of type 1 diabetes. However, T1D can also impact the integrity of the CNS, and the reason why T1D affects CNS integrity remains to be elucidated.

Diabetic animals show high circulating glucocorticoid levels, increased sensitivity to stress, and morphological alteration in various brain areas. How these changes occur is not known, but hypercorticism *per se* can evoke a similar neurodegenerative cascade. The conditions of aberrant GCs levels appear (i) to enhance the vulnerability to metabolic insults of brain areas showing a high degree of plasticity, such as the hippocampus and (ii) may underlie the impairment of cognitive performance.

In T1D, a fundamental question in the central neuropathophysiology is whether GCs aggravate the functional and morphological signs of neurodegeneration and cognitive impairment. Therefore, the objective of the present study is to elucidate the role of GC excess and GC-stress system activity in T1D mice in relation to morphological indices for neuronal viability and cognitive performance.

## 4.2. Hypothesis

We hypothesize that under conditions of T1D, excess GCs and dysregulation of the GCstress system will contribute to cerebral damage by making the brain more vulnerable to metabolic insults and causing concomitant cognitive disturbances. We propose that T1D leads to a more fragile state of the brain in which high levels of GCs may enhance the potential for damage and attenuate protective mechanisms, thus precipitating impairment in cognitive function. We expect to unravel GCs and diabetes interactions and relationships.

#### 4.3. Questions to address

- I. Is there neuronal damage in diabetes? (maybe as a consequence of GCs excess in diabetes?)
- II. What is the pattern of HPA (re)activity in response to diabetes in time?
- III. Do cognitive disturbances parallel these changes?
- IV. Are GCs responsible for brain alterations in diabetes?
- V. Will treatment with anti-glucocorticoids (RU486) prevent and/or restore these alterations?

#### 4.4. Experimental approach

To allow generalization of the results, pharmacological and genetic animal models for diabetes, the STZ-induced and NOD mice, respectively, will be used. For both animal models there are indications of an aberrant functioning GC-stress system (i.e. GC hypersecretion). To test our hypothesis we planned:

- To measure parameters of neuronal damage in NOD pre-diabetic, non-diabetic and diabetic mice and in STZ-diabetic and control mice.
- II. To test HPA axis (re)activity in both models. For this purpose blood samples will be collected to measure basal concentrations of GCs and ACTH. Also central parameters of HPA activity (*in situ* hybridization of mRNA of MR, GR, CRH, AVP) will be measured.
- III. To test cognitive abilities at certain time points of specific HPA (re)activity in the Morris water maze, novel-place recognition, open field, elevated plus maze, forced swimming test. General measures of activity will be recorded as well.
- IV. To use the glucocorticoid antagonist RU486 in order to elucidate the role of GCs and attenuate vulnerability to damage and enhance protection. We discovered that RU486 acts like a double-edged sword: the antagonist blocks the damaging impact of excess GR stimulation while maintaining the beneficial effects of the 'neuroprotective' MR. After four consecutive days of RU486 administration, prevention and/or amelioration of neuropathological signs and restoration of cognitive abilities will be studied.

#### 4.5. Chapters

**Chapter 2** delineates the HPA axis functionality in a genetic model of T1D, the NOD mice. Central parameters of the HPA axis as well as C-peptide and cytokine levels were measured in pre-diabetes and diabetes states. In this model, the results suggest that an enhanced ACTH release may signal the onset of diabetes. **Chapter 3** addresses the underlying mechanism of hypercorticism in STZ mice. Central parameters of the HPA axis and specifically adrenal function were investigated at different time points after STZ-injection. The study demonstrates that adrenal hypersensitivity to ACTH precedes and

maintains the state of chronically elevated glucocorticoids levels. **Chapter 4** describes molecular parameters in the hippocampus of the STZ-diabetic mice that could reflect functional abnormalities of astrocytes and neurons. Parameters of astrocytic and neuronal disturbances such as apolipoprotein E and markers for oxidative stress and early gene expression respectively, were measured between control and diabetic mice. The results showed hippocampal disturbances in this model of T1D, which could be a primary basic mechanism underlying the well-known brain alterations associated with diabetes. **Chapter 5** reveals the role of GCs in STZ-induced diabetic mice, at the molecular and cognitive levels. After the administration of glucocorticoid receptor antagonist (RU486 or mifepristone) for 4 consecutive days to non-diabetic and diabetic mice, the novel placement recognition test was performed and molecular hippocampal parameters reflecting functional abnormalities were measured. Normalization of neuropathological signs and cognitive abilities was found. In **Chapter 6** the experimental data are discussed and placed in a conceptual framework highlighting the central action of glucocorticoids in the onset and progression of diabetes neuropathology.

#### References

Amrani A, Durant S, Throsby M, Coulaud J, Dardenne M, Homo-Delarche F 1998 Glucose homeostasis in the nonobese diabetic mouse at the prediabetic stage. Endocrinology 139:1115-1124

Ansar AS, Penhale WJ, Talal N 1985 Sex hormones, immune responses, and autoimmune diseases. Mechanisms of sex hormone action. Am J Pathol 121:531-551

Atkinson MA, Leiter EH 1999 The NOD mouse model of type 1 diabetes: as good as it gets? Nat Med 5:601-604

**Bach JF** 2002 The effect of infections on susceptibility to autoimmune and allergic diseases. N Engl J Med 347:911-920

**Bellush LL, Rowland NE** 1989 Stress and behavior in streptozotocin diabetic rats: biochemical correlates of passive avoidance learning. Behav Neurosci 103:144-150

**Bellush LL, Reid SG, North D** 1991 The functional significance of biochemical alterations in streptozotocin-induced diabetes. Physiol Behav 50:973-981

**Bestetti G, Rossi GL** 1980 Hypothalamic lesions in rats with long-term streptozotocininduced diabetes mellitus. A semiquantitative light- and electron-microscopic study. Acta Neuropathol (Berl) 52:119-127

**Bestetti G, Rossi GL** 1982 Hypothalamic changes in diabetic Chinese hamsters. A semiquantitative, light and electron microscopic study. Lab Invest 47:516-522

**Biessels GJ, Kappelle AC, Bravenboer B, Erkelens DW, Gispen WH** 1994 Cerebral function in diabetes mellitus. Diabetologia 37:643-650

**Biessels GJ, Kamal A, Ramakers GM, Urban IJ, Spruijt BM, Erkelens DW, Gispen WH** 1996 Place learning and hippocampal synaptic plasticity in streptozotocin-induced diabetic rats. Diabetes 45:1259-1266

**Biessels GJ, Kamal A, Urban IJ, Spruijt BM, Erkelens DW, Gispen WH** 1998 Water maze learning and hippocampal synaptic plasticity in streptozotocin-diabetic rats: effects of insulin treatment. Brain Res 800:125-135

**Bono VH, Jr.** 1976 Review of mechanism of action studies of the nitrosoureas. Cancer Treat Rep 60:699-702

**Camastra S, Bonora E, Del Prato S, Rett K, Weck M, Ferrannini E** 1999 Effect of obesity and insulin resistance on resting and glucose-induced thermogenesis in man. EGIR (European Group for the Study of Insulin Resistance). Int J Obes Relat Metab Disord 23:1307-1313

**Chan O, Chan S, Inouye K, Vranic M, Matthews SG** 2001 Molecular regulation of the hypothalamo-pituitary-adrenal axis in streptozotocin-induced diabetes: effects of insulin treatment. Endocrinology 142:4872-4879

**Chan O, Inouye K, Vranic M, Matthews SG** 2002 Hyperactivation of the hypothalamopituitary-adrenocortical axis in streptozotocin-diabetes is associated with reduced stress responsiveness and decreased pituitary and adrenal sensitivity. Endocrinology 143:1761-1768

**Cryer PE** 1994 Banting Lecture. Hypoglycemia: the limiting factor in the management of IDDM. Diabetes 43:1378-1389

**De Kloet ER, Oitzl MS, Joëls M** 1993 Functional implications of brain corticosteroid receptor diversity. Cell Mol Neurobiol 13:433-455

**De Kloet ER, Vreugdenhil E, Oitzl MS, Joëls M** 1998 Brain corticosteroid receptor balance in health and disease. Endocr Rev 19:269-301

**De Kloet ER, Oitzl MS, Joëls M** 1999 Stress and cognition: are corticosteroids good or bad guys? Trends Neurosci 22:422-426

**De Kloet ER, De Rijk R** 2004 Signaling pathways in brain involved in predisposition and pathogenesis of stress-related disease: genetic and kinetic factors affecting the MR/GR balance. Ann N Y Acad Sci 1032:14-34

**Djarova T, Dube N** 1998 Influence of stressful life events on the onset of Type 1 diabetes in childhood. Afr J Health Sci 5:190-197

**Douma BR, Korte SM, Buwalda B, la Fleur SE, Bohus B, Luiten PG** 1998 Repeated blockade of mineralocorticoid receptors, but not of glucocorticoid receptors impairs food rewarded spatial learning. Psychoneuroendocrinology 23:33-44

**Eberhart M.S, Ogden C, Engelgau M, Cadwell B, Hedley AA, Saydah SH** 2004 "Prevalence of Overweight and Obesity Among Adults with Diagnosed Diabetes --- United States, 1988--1994 and 1999--2002". Morbidity and Mortality Weekly Report 53 (45): 1066-1068.)

**Flood JF, Mooradian AD, Morley JE** 1990 Characteristics of learning and memory in streptozocin-induced diabetic mice. Diabetes 39:1391-1398

Garris DR, Diani AR, Smith C, Gerritsen GC 1982 Depopulation of the ventromedial hypothalamic nucleus in the diabetic Chinese hamster. Acta Neuropathol (Berl) 56:63-66

**Gispen WH, Biessels GJ** 2000 Cognition and synaptic plasticity in diabetes mellitus. Trends Neurosci 23:542-549

Herman JP, Prewitt CM, Cullinan WE 1996 Neuronal circuit regulation of the hypothalamopituitary-adrenocortical stress axis. Crit Rev Neurobiol 10:371-394

Holstad M, Sandler S 2001 A transcriptional inhibitor of TNF-alpha prevents diabetes induced by multiple low-dose streptozotocin injections in mice. J Autoimmun 16:441-447

Homo-Delarche F, Fitzpatrick F, Christeff N, Nunez EA, Bach JF, Dardenne M 1991 Sex steroids, glucocorticoids, stress and autoimmunity. J Steroid Biochem Mol Biol 40:619-637

Hu FB 2003 Sedentary lifestyle and risk of obesity and type 2 diabetes. Lipids 38:103-108

Jack L, Jr., Boseman L, Vinicor F 2004 Aging Americans and diabetes. A public health and clinical response. Geriatrics 59:14-17

Joëls M, Karst H, Krugers HJ, Lucassen PJ 2007 Chronic stress: implications for neuronal morphology, function and neurogenesis. Front Neuroendocrinol 28:72-96

Joëls M, Krugers HJ 2007 LTP after Stress: Up or Down? Neural Plasticity, 2007, Article ID 93202.

Junod A, Lambert AE, Stauffacher W, Renold AE 1969 Diabetogenic action of streptozotocin: relationship of dose to metabolic response. J Clin Invest 48:2129-2139

Kaleeswari R, Thombre DP, Chakrabarty AS 1986 Acute effect of streptozotocin induced

diabetes on bar pressing for food reward in albino rats. Indian J Physiol Pharmacol 30:319-321

Kramer L, Fasching P, Madl C, Schneider B, Damjancic P, Waldhausl W, Irsigler K, Grimm G 1998 Previous episodes of hypoglycemic coma are not associated with permanent cognitive brain dysfunction in IDDM patients on intensive insulin treatment. Diabetes 47:1909-1914

**Kumagai AK** 1999 Glucose transport in brain and retina: implications in the management and complications of diabetes. Diabetes Metab Res Rev 15:261-273

Li ZG, Zhang W, Sima AA 2002 C-peptide prevents hippocampal apoptosis in type 1 diabetes. Int J Exp Diabetes Res 3:241-245

**Lovejoy JC** 2002 The influence of dietary fat on insulin resistance. Curr Diab Rep 2:435-440

Lowy MT, Gault L, Yamamoto BK 1993 Adrenalectomy attenuates stress-induced elevations in extracellular glutamate concentrations in the hippocampus. J Neurochem 61:1957-1960

**Lupien SJ, McEwen BS** 1997 The acute effects of corticosteroids on cognition: integration of animal and human model studies. Brain Res Brain Res Rev 24:1-27

**Luse SA** 1970 The ultrastructure of the brain in the diabetic Chinese hamster with special reference to synaptic abnormalities. Electroencephalogr Clin Neurophysiol 29:410

**Magarinos AM, McEwen BS** 1995 Stress-induced atrophy of apical dendrites of hippocampal CA3c neurons: involvement of glucocorticoid secretion and excitatory amino acid receptors. Neuroscience 69:89-98

Makino S, Kunimoto K, Muraoka Y, Mizushima Y, Katagiri K, Tochino Y 1980 Breeding of a non-obese, diabetic strain of mice. Jikken Dobutsu 29:1-13

**Martinez-Tellez R, Gomez-Villalobos MJ, Flores G** 2005 Alteration in dendritic morphology of cortical neurons in rats with diabetes mellitus induced by streptozotocin. Brain Res 1048:108-115

**Mayer G, Nitsch R, Hoyer S** 1990 Effects of changes in peripheral and cerebral glucose metabolism on locomotor activity, learning and memory in adult male rats. Brain Res 532:95-100

McCarthy AM, Lindgren S, Mengeling MA, Tsalikian E, Engvall JC 2002 Effects of diabetes on learning in children. Pediatrics 109:E9

McEwen BS, Sapolsky RM 1995 Stress and cognitive function. Curr Opin Neurobiol 5:205-216

McEwen BS 1998 Protective and damaging effects of stress mediators. N Engl J Med 338:171-179

McEwen BS, Magarinos AM, Reagan LP 2002 Studies of hormone action in the hippocampal formation: possible relevance to depression and diabetes. J Psychosom Res 53:883-890

McGaugh JL 2000 Memory--a century of consolidation. Science 287:248-251

Mensah-Brown EP, Stosic GS, Maksimovic D, Jasima A, Shahin A, Lukic ML 2002 Downregulation of apoptosis in the target tissue prevents low-dose streptozotocin-induced autoimmune diabetes. Mol Immunol 38:941-946 **Miles WR and Root HF** 1922 Psychologic tests applied to diabetic patients. Archives of Internal Medicine 30: 767–777

Mukai N, Hori S, Pomeroy M 1980 Cerebral lesions in rats with streptozotocin-induced diabetes. Acta Neuropathol (Berl) 51:79-84

**Munck A, Guyre PM, Holbrook NJ** 1984 Physiological functions of glucocorticoids in stress and their relation to pharmacological actions. Endocr Rev 5:25-44

Nicholas A, Munhoz CD, Ferguson D, Campbell L, Sapolsky R 2006 Enhancing cognition after stress with gene therapy. J Neurosci 26:11637-11643

Oster MH, Castonguay TW, Keen CL, Stern JS 1988 Circadian rhythm of corticosterone in diabetic rats. Life Sci 43:1643-1645

**Parisi V, Uccioli L** 2001 Visual electrophysiological responses in persons with type 1 diabetes. Diabetes Metab Res Rev 17:12-18

**Popovic M, Biessels GJ, Isaacson RL, Gispen WH** 2001 Learning and memory in streptozotocin-induced diabetic rats in a novel spatial/object discrimination task. Behav Brain Res 122:201-207

Reul JM, Gesing A, Droste S, Stec IS, Weber A, Bachmann C, Bilang-Bleuel A, Holsboer F, Linthorst AC 2000 The brain mineralocorticoid receptor: greedy for ligand, mysterious in function. Eur J Pharmacol 405:235-249

**Roozendaal B** 2000 1999 Curt P. Richter award. Glucocorticoids and the regulation of memory consolidation. Psychoneuroendocrinology 25:213-238

**Ryan CM, Williams TM, Finegold DN, Orchard TJ** 1993 Cognitive dysfunction in adults with type 1 (insulin-dependent) diabetes mellitus of long duration: effects of recurrent hypoglycaemia and other chronic complications. Diabetologia 36:329-334

**Ryan CM, Williams TM** 1993 Effects of insulin-dependent diabetes on learning and memory efficiency in adults. J Clin Exp Neuropsychol 15:685-700

**Sapolsky RM** 1992 Do glucocorticoid concentrations rise with age in the rat? Neurobiol Aging 13:171-174

Saravia FE, Gonzalez SL, Roig P, Alves V, Homo-Delarche F, De Nicola AF 2001 Diabetes increases the expression of hypothalamic neuropeptides in a spontaneous model of type I diabetes, the nonobese diabetic (NOD) mouse. Cell Mol Neurobiol 21:15-27

Saravia FE, Revsin Y, Gonzalez Deniselle MC, Gonzalez SL, Roig P, Lima A, Homo-Delarche F, De Nicola AF 2002 Increased astrocyte reactivity in the hippocampus of murine models of type 1 diabetes: the nonobese diabetic (NOD) and streptozotocin-treated mice. Brain Res 957:345-353

Schnedl WJ, Ferber S, Johnson JH, Newgard CB 1994 STZ transport and cytotoxicity. Specific enhancement in GLUT2-expressing cells. Diabetes 43:1326-1333

Schoenle EJ, Schoenle D, Molinari L, Largo RH 2002 Impaired intellectual development in children with Type I diabetes: association with HbA(1c), age at diagnosis and sex. Diabetologia 45:108-114

Scribner KA, Akana SF, Walker CD, Dallman MF 1993 Streptozotocin-diabetic rats exhibit facilitated adrenocorticotropin responses to acute stress, but normal sensitivity to feedback by corticosteroids. Endocrinology 133:2667-2674

Sima AA, Kamiya H, Li ZG 2004 Insulin, C-peptide, hyperglycemia, and central nervous system complications in diabetes. Eur J Pharmacol 490:187-197

**Tornello S, Fridman O, Weisenberg L, Coirini H, De Nicola AF** 1981 Differences in corticosterone binding by regions of the central nervous system in normal and diabetic rats. J Steroid Biochem. 14 :77-81.

**Wang Z, Gleichmann H** 1998 GLUT2 in pancreatic islets: crucial target molecule in diabetes induced with multiple low doses of streptozotocin in mice. Diabetes 47:50-56

Zahner D, Malaisse WJ 1990 Kinetic behaviour of liver glucokinase in diabetes. I. Alteration in streptozotocin-diabetic rats. Diabetes Res 14:101-108

Zimmet P, Alberti KG, Shaw J 2001 Global and societal implications of the diabetes epidemic. Nature 414:782-787

#### **References Box 1**

- Rotter JI, Vadheim CM, Rimoin DL 1990 Genetics of diabetes mellitus. In Rifkin H, Porte D (eds) Diabetes Mellitus. Theory and Practice, ed 4. Elsevier, Amsterdam, pp 378-413 Lo SS, Tun RY, Hawa M, Leslie RD 1991 Studies of diabetic
- Diabetes Epidemiology Research International Group 1988 Geographic patterns of childhood insulin-dependent diabeies mellitus. Diabetes 37:1113-1119.
- Patrick SL, Moy CS, Laporte RE 1989 The world of insulin- dependent diabetes mellitus: what international epidemiologic studies reveal about the etiology and natural history of IDDM. Diabetes Metab Rev 5:571-578
- Lefkowith J, Schreiner G, Cormier J, Handler ES, Driscoll HK, Greiner D, Mordes JP, Rossini AA 1990 Prevention of diabetes in the BB rat by essential fatty acid deficiency. Relationship between physiological and biochemical changes. J Exp Med 171: 729-743
- Elliott RB, Reddy SN, Bibby NJ, Kida K 1988 Dietary prevention of diabetes in the nonobese diabetic mouse. Diabetologia 31: 62-64
- Issa-Chergui 8, Guttmann RD, Seemayer TA, Kelley VE, Colle E 1988 The effect of diet on the spontaneous insulin dependent diabetic syndrome in the rat. Diabetes Res 9:81-86
- Oldstone MB 1990 Viruses as therapeutic agents. I. Treatment of nonobese insulindependent diabetes mice with virus prevents insulin-deDendent diabetes mellitus while maintaining general immune com'petence. J Exp Med 171:2077-2089
- Oldstone MB, Ahmed R, Salvato M 1990 Viruses as therapeutic agents. II. Vii& reassortants map prevention of insulin-depe'ndent diabetes mellitus to the small RNA of lymphocytic choriomeningitis virus. J Exp Med 171:2091-2100.
- Takei I, Asaba Y, Kasatani T, Maruvama T, Watanabe K, Yanagawa T, Saruia T, Ishii T 1492 Suppression.of development of diabetes in NOD mice by lactate dehydrogenase virus infection. J Autoimmun 5:665-673.
- Wilberz S, Partke HJ, Dagnaes-Hansen F, Herberg L 1991 Persistent MHV (mouse hepatitis virus) infection reduces the incidence of diabetes mellitus in non-obese diabetic mice. Diabetologia 34:2-5
- Like AA, Guberski DL, Butler L 1991 Influence of environmental viral agents on frequency and tempo of diabetes mellitus in BB/Wor rats. Diabetes 40:259-262

- Guberski DL, Thomas VA, Shek WR, Like AA, Handler ES, Rossini AA, Wallace JE, Welsh RM 1991 Induction of type I diabetes by Kilham's rat virus in diabetes-resistant BB/Wor rats. Science 254:1010-1013
- 13. Elliott RB, Martin JM 1984 Dietary protein: a trigger of insulin-dependent diabetes in the BB rat? Diabetoloeia 26:297-299
- Dineman D, Fishman L, Clarson C, Martin JM 1987 Dietary triggers of insulin-dependent diabetes in the BB rat. Diabetes Res 5:93-97
- 15. Pozzilli P, Signore A, Williams AJ, Beales PE 1993 NOD mouse colonies around the world: recent facts and figures. Immunol Today 14:193-196
- Yoon JW, Ihm SH 1990 Viruses as a triggering factor of autoimmune type I diabetes. In Farid NR, Bona CA (eds) The Molecular Aspects of Autoimmunity. Academic Press, London, pp 231-240
- 17. Yoon JW 1991 Role of viuses in the pathogenesis of IDDM. Ann Med 23:437-445
- MacDonald MJ, Liston L, Carlson I 1987 Seasonal&y in glycosylated hemoglobin in normal subjects. Does seasonal incidence in insulin-dependent diabetes suggest specific etiology? Diabetes 36:265-268
- Sadelain MW, Oin HY, Lauzon 1, Sineh B 1990 Prevention of type I diabetes in NOD mice by idjuv&t immunotherapy. Diabetes 39:583-589
- 20. McInerney MF, Pek SB, Thomas DW 1991 Prevention of insulitis and diabetes onset by treatment with comulete Freund's adjuvant in NOD mice. Diabetes 40:715-725
- Sadelain MW, Qin HY, Sumoski W, Parfrev N, Sinzh B, Rabinovitch A 1990 Prevention of diabetes in the BB rat by early immunotherapy using Freund's adjuvant. J Autoimmun 3: 671-680
- Qin HY, Suarez WL, Parfrey N, Power RF, Rabinovitch A 1992 Mechanisms of complete Freund's adjuvant protection against diabetes in BB rats: induction of non-specific suppressor cells. Autoimmunity 12:193-199
- Renold AE 1985 Possible animal models for diabetes mellitus: syndromes involving toxic or immune etiology. In: Alberti KG, Krall LP (eds) The Diabetes Annual. Elsevier, Amsterdam, DU 492-508
- Karam JH, Lewitt PA, Young CW, Nowlain RE, Frankel BJ, Fujiya H, Freedman ZR, Grodsky GM 1980 Insulinopenic diabetes after rodenticide (Vacor) ingestion: a unique model of acquired diabetes in man. Diabetes 29:971-978
- Surwit RS, Schneider MS, Feinglos MN 1992 Stress and diabetes mellitus. Diabetes Care 15:1413-1422
- Ader DN, Johnson SB, Huang SW, Riley WJ 1991 Group size, cage shelf level, and emotionality in non-obese diabetic mice: impact on onset and incidence of IDDM. Psychosom Med 53:313-321
- Durant S, Coulaud J, Amrani A, El Hasnaoui A, Dardenne M, Homo-Delarche F 1993 Effects of various environmental stress paradigms and adrenalectomy on the expression of autoimmune type 1 diabetes in the nonobese diabetic (NOD) mouse. J Autoimmun 6:735-751
- Williams AJ, Krug J, Lampeter EF, Mansfield K, Beales PE, Signore A, Gale EA, Pozzilli P 1990 Raised temperature reduces the incidence of diabetes in the NOD mouse. Diabetologia 33: 635-637

#### **References Table 1**

- Makino S, Kunimoto K, Muraoka Y, Mizushima Y, Katagiri K, Tochino Y 1980 Breeding of a non-obese, diabetic strain of mice. Exp Anim 29:1-13
- Nakhooda AF, Like AA, Chappel CI, Murray FT, Marliss EB 1977 The spontaneously diabetic Wistar rat. Metabolic and morphologic studies. Diabetes 26:100-112
- Kawano K, Hirashima T, Moris S, Saitoh Y, Kurosumi M, Natori T 1991 New inbred strain of Long-Evans Tokushima lean rats with IDDM without lymphopenia. Diabetes 40(11):1375-1381
- Lenzen S, Tiedge M, Elsner M, Lortz S, Weiss H, Jorns A 2001 The LEW.1AR1/Ztmiddm rat: a new model of spontaneous insulin-dependent diabetes mellitus. Diabetologia 44(9):1189-1196.
- Oldstone MB, Nerenberg M, Southern P, Price J, Lewicki H 1991 Virus infection triggers insulin-dependent diabetes mellitus in a transgenic model: role of anti-self (virus) immune response. Cell 65:319-331
- Fugger L 2000 Human autoimmunity genes in mice. Curr Opin Immunol. 12(6):698-703.
- Malaisse WJ 1982 Alloxan toxicity to the pancreatic B-cell. A new hypothesis. Biochem Pharmacol 31:3527-3534
- Like AA, Rossini AA 1976 Streptozotocin-induced pancreatic insulitis: new model of diabetes mellitus. Science 193:415-417
- Taguchi 0, Takahashi T, Nishizuka Y 1990 Self-tolerance and localized autoimmunity. Curr Opin Immunol 2:576-581
- Fowell D, Mason D 1993 Evidence that the T cell repertoire of normal rats contains cells with the potential to cause diabetes. Characterization of the CD4+ T cell subset that inhibits this autoimmune potential. J Exp Med 177:627-636
- Stumbles PA, Penhale WJ 1993 IDDM in rats induced by thymectomv and irradiation. Diabetes 42:571-578
- McKeever U, Mordes JP, Greiner DL, Appel MC, Razing J, Handler ES, Rossini AA 1990 Adoutive transfer of autoimmune diabetes and thyroiditis to athymic cats. Proc Nat1 Acad Sci USA 87:7618-7622