

Genetics of Ollier disease and Maffucci syndrome Pansuriya, T.C.

Citation

Pansuriya, T. C. (2012, March 15). *Genetics of Ollier disease and Maffucci syndrome*. Retrieved from https://hdl.handle.net/1887/18591

Version: Corrected Publisher's Version

License: License agreement concerning inclusion of doctoral thesis in the

Institutional Repository of the University of Leiden

Downloaded from: https://hdl.handle.net/1887/18591

Note: To cite this publication please use the final published version (if applicable).

Cover Page



Universiteit Leiden



The handle http://hdl.handle.net/1887/18591 holds various files of this Leiden University dissertation.

Author: Pansuriya, Twinkal C.

Title: Genetics of Ollier disease and Maffucci syndrome

Issue Date: 2012-03-15

Chapter



General Introduction



Genetic disorders of the skeleton known as skeletal dysplasia shows diverse manifestations (1). Skeletal dysplasia is a group of skeletal disorders that result from disturbances in the complex processes of skeletal development and growth which constitute a diagnostic challenge due to the rarity of these diseases (2). Mutations are reported either in regulators of skeletal organogenesis, such as cellular signaling (growth factors and its receptors), or in matrix components that affect cartilage and bone tissues (3). There are two main classes of skeletal dysplasia: osteochondrodysplasia and dysostosis. Osteochondrodysplasia develops due to the abnormal growth and development of bone and/or cartilage. In contrast, dysostosis is a developmental anomaly resulting from localized problems in the migration of mesenchymal cells and their condensation (3). The Nosology and Classification of Genetic Skeletal Disorders provides an overview of different diagnostic entities based on clinical and radiographic features and molecular pathogenesis (2). In 2010, in total 456 conditions were described which were divided into 40 different groups. Presently, for 316 of these 456 conditions, the underlying genetic defects are known (2).

Within this classification (2), enchondromatosis syndrome is recognized as a form of osteochondrodysplasia. Patients with enchondromatosis syndrome have multiple enchondromas (benign cartilage forming tumors in the medulla of bone) in their skeleton. The focus of our study concerns the two main subtypes of enchondromatosis syndrome known as Ollier disease and Maffucci syndrome. In addition to multiple enchondromas present in Ollier disease, soft tissue hemangiomas are present in Maffucci syndrome.

Ollier Disease

Ollier disease (OMIM 166000) is a rare, non-familial skeletal disorder (4). Ollier disease was first described by Louis Ollier, a French surgeon in 1889. The disorder is characterized by the presence of at least three enchondromas with an asymmetric distribution and extreme clinical variability (size, number, location, age of onset and requirement of surgery) (4-6).

Maffucci Syndrome

Maffucci syndrome (OMIM 166000) is characterized by presence of multiple enchondromas, resulting in bone deformities, together with soft tissue haemangiomas especially spindle cell hemangiomas or rarely lymphangiomas (4;7;8).

Chondrosarcomas

Enchondromas in Ollier disease and Maffucci syndrome can undergo malignant transformation towards chondrosarcoma. Chondrosarcomas are defined in the 2002 WHO classification as a "Heterogeneous group of lesions with diverse morphological features and clinical behaviour" (9). Chondrosarcoma is the third most frequent malignant bone tumor, in which the tumor cells deposit a hyaline cartilaginous matrix (9). The incidence of chondrosarcoma is slightly increased in males compared to females (9). The age of onset varies from 30-60 years (9). The pelvis is the most commonly affected site followed by femur, humerus, and ribs. The small bones of hands and feet, spine and craniofacial bones are rarely affected. Symptoms involve pain and swelling. Myxoid changes, calcification or ossification may be present. Radiographically, the development of chondrosarcoma is manifested as a lytic lesion, cortical erosion or destruction, soft tissue extension and irregularity or indistinctness of the surface of tumor (10).



MRI is used to identify soft tissue extension and the extent of tumor while CT scan can be helpful to see calcified matrix. As chondrosarcomas are highly resistant to chemo- and radiotherapy, surgery is the only option to cure the patients so far (11).

Chondrosarcoma subtypes

Chondrosarcomas can be divided mainly into 5 different subtypes including conventional (80-85%), dedifferentiated (6-10%), periosteal (2%), mesenchymal (2%) and clear cell chondrosarcomas (1%). Periosteal chondrosarcomas (juxtacortical chondrosarcomas) are malignant hyaline cartilage forming tumors located at the surface of the bone, arising from perichondrium and not connected to the original bone (9:12). Dedifferentiated chondrosarcoma contains two clearly distinct components: a well-differentiated cartilage tumor (either an enchondroma or a low grade chondrosarcoma), juxtaposed to a high-grade noncartilaginous sarcoma, with a sharp interface between the two components (13). Conventional chondrosarcomas are most frequent and arise de novo (primary) or from a benign precursor (secondary). Conventional chondrosarcomas are divided into two groups based on their anatomical location in the bone: i) secondary peripheral chondrosarcomas (10-15%) and ii) central chondrosarcomas (85-90%). Chondrosarcomas arising in patients with Ollier disease and Maffucci syndrome are classified as the conventional secondary central subtype.

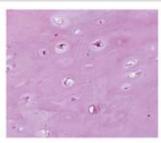
Conventional Chondrosarcomas

- I) Secondary peripheral chondrosarcoma develop within the cartilage cap of a pre-existing osteochondroma. Osteochondroma, a benign tumor, is a cartilage capped bony projection arising at the surface of bone (14). Multiple osteochondromas (MO, also known as hereditary multiple exostoses) is an autosomal dominant skeletal dysplasia caused by mutations in EXT1 or EXT2 (15-18) and characterized by the presence of multiple osteochondromas (19). Although the involvement of EXT1 or EXT2 inactivation in osteochondroma formation is beyond dispute, EXT1 and EXT2 are not involved in the progression towards secondary peripheral chondrosarcoma (20). The most frequent location of secondary peripheral chondrosarcoma is ilium, followed by scapula, tibia, femur, pubic bone and ribs.
- ii) Central chondrosarcoma arises *de novo* in the medulla of the bone (primary) or from a preexisting benign enchondroma (secondary). Most of the central chondrosarcomas are believed to arise primary. Clinical signs suggestive of malignancy are presence of pain, cortical erosion and extension of the tumor into soft tissues (6;21). Central chondrosarcomas can be found in almost all parts of the skeleton which are formed by enchondral ossification. The most preferential sites include femur, followed by ribs and ilium. The distinction between enchondroma and low grade chondrosarcoma is difficult at radiological as well as at the histological level (22) and slightly subjected to interobserver variability (23;24). Criteria to distinguish include mucoid matrix degeneration of more than 20% and/or presence of host bone entrapment (24). In Ollier disease and Maffucci syndrome more cellularity and atypia is tolerated, making the histological distinction even more difficult.

Histologically conventional chondrosarcomas can be divided into three grades (Grade I, II and III) (25) (Figure 1). Grading is so far the most important prognostic predictor for metastasis. The risk of developing metastasis increases with increase in tumor grade. Studies showed that metastases of grade I chondrosarcomas are rare or absent, while 10-33% of grade II and around 70% of grade III chondrosarcomas metastasize (25;26).

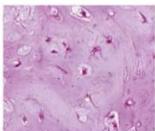


Figure 1 Histology of enchondroma and different grades of chondrosarcomas



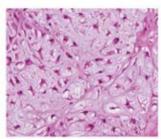
Enchondroma

encasement low cellularity no atypia



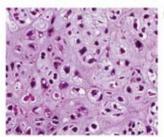
Grade I chondrosarcoma

bone entrapment of preexisting host bone presence of small, densely staining nuclei chondroid or myxoid matrix



Grade II chondrosarcoma

nuclei are of moderate size increased cellularity myxoid matrix presence of mitosis



Grade III chondrosarcoma

pleomorphic nuclei high cellularity myxoid matrix presence of mitosis spindle cell shaped cells at the edge of lobules



Approaches to understand complex diseases

Complex diseases are caused by a combination of genetic and environmental factors. For most of these diseases it is difficult to identify the cause as they do not obey standard Mendelian patterns of inheritance. Genomics is the field of study helping to understand a role of the genome in the development of particular diseases. The Human Genome Project has changed the view of researchers to gain new insights in pathogenesis of diseases. The known role of copy-number alterations in sporadic genomic disorders, combined with emerging information about inherited copynumber variation, indicate the importance of assessing copy-number variants (CNVs), including common copy-number polymorphisms, involved in disease (27). In principle, complex disease can be explained by the presence of particular single nucleotide polymorphisms (SNPs) or CNVs or variation in noncoding sequences associated with diseases. As Ollier disease and Maffucci syndrome are non-inherited disorders, we first started off with a genome-wide analysis to search for candidate regions involved in these disorders. We used high resolution SNP array combined with expression arrays.

Genome-wide approach

Single Nucleotide Polymorphism (SNP) Arrays

SNP arrays offer an opportunity to identify copy number changes together with loss of heterozygosity (LOH) events occurring in the tumor, throughout the genome. SNPs are variable positions in the genome with two different allelic types when the frequency of the minor allele exceeds 1% in at least one population (28;29). SNP arrays are an ideal platform to map somatic and germline genetic alterations (30-33). Only SNPs which are heterozygous in germline are informative as an indication of possible LOH. Paired normal DNA is not always available and therefore, an advantage of SNP arrays are that it provide marker densities that enable the identification of LOH regions, without using germline paired DNA. SNP frequency has an uneven distribution throughout the whole genome. Including copy number variation (CNV) probes to a SNP array platform like Affymetrix SNP 6.0 array can compensate for the unevenness of the SNPs. SNPs are much less frequent inside coding areas including exomic regions, therefore the detection rate of genetic changes that encompass only one or a few exons especially for small genes is rather limited. Also, balanced translocations and point mutations can not be detected by SNP arrays.

Gene Expression Array

Microarrays are used as a routine tool for molecular profiling, identification of new targets and biomarker discoveries in biomedical research (34). Microarray technology has allowed the abundance of thousands of different mRNAs to be measured simultaneously in a given sample using a single hybridization reaction (35;36). Therefore, analysis of individual genes has provided an opportunity to analyze large sets of genes and relationships in their expression (35). Interpretation of the results to gain insight into biological mechanisms is still challenging. There are some critical issues such as correct selection of samples, proper experimental design, sample collection, preparation of targeted RNA, integrity and purity of RNA related to microarrays in order to guarantee the quality and reproducibility of the obtained data (36).

Genomic changes (amplification or deletions) often comprise 10s 100s of genes, therefore integration of copy number changes and expression arrays might be more informative.



Expression changes (up or down regulation) of genes residing in copy number alteration (CNA) regions of the genome might identify genes important in the pathogenesis of tumors.

Methylation array

DNA methylation plays a critical role in regulating gene expression and cellular functions during normal development as well as in carcinogenesis (37). Methylation is largely known as epigenetic modification of DNA (38). Methylation of DNA occurs exclusively in 5-cytosine and in mammals, the majority of cytosine methylation occurs in CpG sites. Non-CpG methylation is rare (39). CpG islands are present in ~70% of human promoters (40). Epigenetic changes (which alter the gene expression) have been recognized as one of the most important molecular signatures of the tumors in recent years. These alterations comprise of hypermethylation of tumor suppressor genes or hypomethylation of oncogenes (38). The exact mechanism of aberrant methylation is still unknown. Methylation profiling helps to understand the nature of gene regulation in cells, and also the epigenetic mechanisms of interactions between cells and environment (37). There are three methods available for DNA methylation profiling which includes I) discrimination of bisulfite induced C to T transition, ii) cleavage of genomic DNA by methylation sensitive restriction enzymes and iii) immunoprecipitation with methyl-binding proteins or antibodies against methylated cytosine (37).

Some of these approaches permit the investigation of the limited number of methylated regions at a time. Whereas, microarray and sequencing based DNA methylation profiling technologies have been developed in order to assess methylation status for a large number of genes or even the entire genome.

Hypothesis driven approach

Based on the literature related to chondrocyte differentiation and enchondral bone formation, one could postulate a number of candidate genes for Ollier disease and/or Maffucci syndrome.

1. NDST1

EXT1 or EXT2 are known to be involved in osteochondroma (14-16). While it is evident that inactivation of EXT1 or EXT2 is the driving force for the development of benign peripheral cartilaginous tumors, they are not involved in central chondrosarcoma and expression of these genes was comparable to the growth plate (41). The EXT proteins are glycosyltransferases responsible for the elongation of heparan sulfate (HS) chains (42;43). HS is a large complex carbohydrate that binds various growth factors and enzymes and its assembly involves three steps I) chain initiation ii) chain elongation iii) chain modification.

I) chain initiation occurs when four sugars are attached to specific serine residue of the core proteins. ii) elongation steps involve N-acetyl glucosamine and glucoronic acids which are alternatively added by copolymerases encoded by EXT1 or EXT2. iii) chain modification involves Ndeacetylation/ N-sulfation (NDST1), C5-epimerization, 2-0 sulfation of uronic acids and 3-0 and 6-0 sulfation of glucosamine residues. Presto et al. proposed a GAGosome model in which cells over-expressing NDST1 and EXT2, NDST1 competes with EXT1 to bind to EXT2 and will form heteroduplex (44). Binding of more NDST1 to EXT2 might alter formation and localization of HS. The role of EXT1, NDST1 and HS is unknown in enchondromas and chondrosarcomas related to Ollier disease and Maffucci syndrome.



2.PTH1R

Enchondroma might arise as a result of abnormal regulation of pathways involved in chondrocyte proliferation and differentiation. One of the most important signaling pathways is the Indian Hedgehog/Parathyroid Hormone Like Hormone IHH/PTHLH negative feed back loop (45). Prehypertrophic chondrocytes secrete IHH which will bind to its receptor Patched (PTCH) which will result in increased secretion of PTHLH. PTHLH will bind to its receptor PTH1R which will inhibit further differentiation of chondrocytes by up-regulating BCL2, resulting in less IHH producing cells (46). PTH1R and IHH pathways are tightly coupled and therefore reduced PTH1R signaling could lead to impaired chondrocyte proliferation and differentiation.

Previously, a R150C PTH1R (3p22 (47)-p21.1) point mutation was reported in two out of six patients with constitutively active IHH signaling (48) but an elaborative study on 28 Ollier patients failed to detect any mutations in PTHR1 by our group (49). G121E PTH1R, A122T PTH1R and R255H PTH1R mutations were subsequently found in 3 out of 14 Ollier patients (50). Two heterozygous mutations, G121E PTH1R and A122T PTH1R were present only in enchondroma from an Ollier patient while R255H PTH1R was present in tumor as well as in leukocyte DNA. All these mutations were claimed to alter the ligand affinity of the receptor as well as its expression at the cell surface and ultimately impaired its function (50). In total four different heterozygous mutations were reported. Thus, heterozygous PTHR1 mutations may contribute or act as a modifier in small subset of Ollier patients (48-50).

3. IDH1 and IDH2

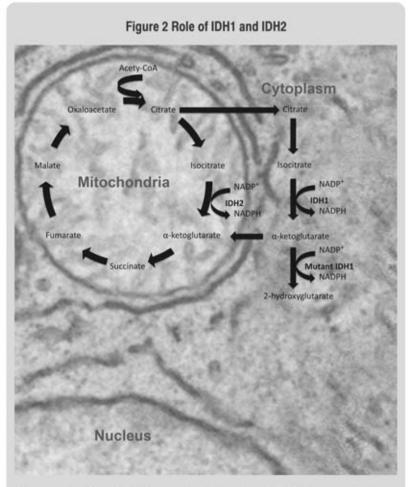
Gliomas are the most frequent non-cartilaginous tumors associated with Ollier disease (51;52). Also, six patients with Maffucci syndrome having a glioma have been reported in the literature (52-57). Glioma is the most common type of primary brain tumor (58). Heterozygous mutations at the R132 codon of isocitrate dehydrogenase 1 (*IDH1*) and R172 codon of isocitrate dehydrogenase 2 (*IDH2*), mutations were described for up to 70% of gliomas (59). Mutations in *IDH1* and *IDH2* are mutually exclusive with rare exceptions, which suggest mutation in either of these two isoforms is sufficient to confer growth advantage and/or cell survival (60). Mutations in *IDH1* or *IDH2* were also reported for solitary central and periosteal chondrosarcomas as well as for few patients with enchondromatosis syndrome (61).

IDH catalyzes the oxidative decarboxylation of isocitrate to α - ketoglutarate (α -KG) and reduce NAD(P+) to NAD(P)H (Figure 2). IDH1 is located in the cytoplasm and in the peroxisome while IDH2 is located in the mitochondria. They are involved in lipid metabolism and in the Krebs cycle (Figure 2). Mutant IDH1 or IDH2 leads to gain of function by producing 2- hydroxyglatarate (2HG), a structural analogue of α -KG (62) and ultimately lead to reduction in α -KG production (Figure 2). Based on the increased incidence of gliomas and *IDH* mutations in solitary central cartilaginous tumors, we hypothesized that *IDH1* and/or *IDH2* mutations may occur in patients with Ollier disease and/or Maffucci syndrome.

4. GNAS

Like gliomas, juvenile ovarian granulosa cell tumors show an increased incidence in patients with Ollier disease and Maffucci syndrome (52;53;63;64).

Mutations in GNAS at R201 were reported in 30% of juvenile ovarian granulosa cell tumors (65). Apart from this, somatic mosaic GNAS mutations are also found in McCune-Albright syndrome, which is a non hereditary disorder characterized by polyostotic fibrous dysplasia combined with endocrinopathies (66). Based on the association of juvenile ovarian granulosa cell tumors with Ollier disease and Maffucci syndrome, we hypothesized that GNAS mutations might be present in these patients.



Enzymes of TCA cycle in the mitochondria and mutated IDH in the cytosol are represented. Mutant IDH1 produces 2HG and reduces the amount of α -KG.



Aim of the investigation and outline of the thesis

The main purpose of the studies described in this thesis is to find the genetic deficit in Ollier disease and Maffucci syndrome and understand their functional consequences. As Ollier disease and Maffucci syndrome are very rare, non-inherited syndromes with a unilateral predominance of the multiple enchondromas, we hypothesized the presence of somatic mosaicism with an early post zygotic mutation resulting in asymmetric involvement of skeletal structures, similar to McCune Albright syndrome caused by somatic mosaic *GNAS* mutations, (67).

We first searched the literature and present a detailed overview of all different subtypes of enchondromatosis syndrome in Chapter 2.

In Chapter 3 and 4, the Affymetrix Genome-Wide Human SNP Array 6.0 platform was used to identify candidate gene/genes for Ollier disease (Chapter 3) and Maffucci syndrome (Chapter 4). The arrays contain 1.8 million reporters, including more than 906,600 SNPs and 946,000 probes for the detection of CNV. We compared genotypes between tumor and paired blood or saliva DNA. We have integrated copy number variation results with Illumina genome-wide expression v3 array and selected few candidate genes for enchondroma development.

In Chapter 5 and 6, a hypothesis driven approach was used to study the five genes as outlined above. As described earlier in detail, NDST1 was selected as a candidate gene and other components of heparan sulfate pathway were analyzed in Chapter 5. Since a small subset of patients with Ollier disease showed mutations in PTH1R, we also performed mutation analysis as described in Chapter 6. Based on the increased incidence of gliomas and juvenile granulosa cell tumors in Ollier disease and Maffucci syndrome, and mutation studies on non-syndromal chondrosarcomas, we evaluated the occurrence of IDH1, IDH2 and GNAS mutations. In addition we performed epigenetic studies to investigate the mechanism of enchondroma development using Illumina HumanMethylation arrays in Ollier disease and Maffucci syndrome as described in Chapter 6. Finally, results are summarized and discussed in Chapter 7.



References

- Superti-Furga A, Bonafe L, Rimoin DL. Molecular-pathogenetic classification of genetic disorders of the skeleton. Am J Med Genet 2001;106(4):282-93.
- Warman ML, Cormier-Daire V, Hall C, Krakow D, Lachman R, Lemerrer M, et al. Nosology and classification of genetic skeletal disorders: 2010 revision. Am J Med Genet A 2011;155A(5):943–68.
- 3. Cotran RS, Robbins St., Pathologic Basis of Disease. 7 ed. Philadelphia: WB Saunders Company; 2010. Chapter 26:1210.
- Spranger J, Kemperdieck H, Bakowski H, Opitz JM. Two peculiar types of enchondromatosis. Pediatr Radiol 1978;7(4):215-9.
- World Health Organization Classification of Tumors. Pathology and Genetics of Tumors of Soft Tissue and Bone. Lyon: IARC Press; 2002.
- Silve C, Juppner H. Ollier disease. Orphanet J Rare Dis 2006;1:37.
- Maffucci A. Di un caso encondroma ed angioma multiplo. Movimento medico-chirurgico, Napoli 1881;3:399-412;565-575.
- Mertens F, Unni KK. Congenital and inherited syndromes associated with bone and soft tissue tumors: Enchondromatosis. In: Fletcher CDM, Unni KK, Mertens F, editors. World Health Organisation classification of tumors. Pathology and genetics of tumors of soft tissue and bone. Lyon: IARC Press; 2002. p. 356-7.
- Bertoni F, Bacchini P, Hogendoorn PCW. Chondrosarcoma. In: Fletcher CDM, Unni KK, Mertens F, editors. World Health Organisation classification of tumors. Pathology and genetics of tumors of soft tissue and bone. Lyon: IARC Press; 2002. p. 247-51.
- Unni KK, Cartilaginous lesions of bone. J Orthop Sci 2001;6(5):457-72.
- Gelderblom H, Hogendoorn PCW, Dijkstra SD, van Rijswijk CS, Krol AD, Taminiau AH, et al. The clinical approach towards chondrosarcoma. Oncologist 2008:13(3):320-9.
- Norman A, Sissons HA. Radiographic hallmarks of peripheral chondrosarcoma. Radiology 1984;151(3):589-96.
- Milchgrub S, Hogendoorn PCW. Dedifferentiated chondrosarcoma. In: Fletcher C.D.M., Unni KK, Mertens F, editors. World health organization classification of tumors. Pathology and genetics. Tumors of soft tissue and bone. 2002. p. 252-4.
- Khurana J, Abdul-Karim F, Bovée JVMG. Osteochondroma. In: Fletcher CDM, Unni KK, Mertens F, editors. World Health Organization classification of tumors. Pathology and genetics of tumors of soft lissue and bone. Lyon (France): IARC Press; 2002. p. 234-6.
- Ahn J, Ludecke H-J, Lindow S, Horton WA, Lee B, Wagner MJ, et al. Cloning of the putative tumor suppressor gene for hereditary multiple exostoses (EXT1). Nature Genet 1995:11:137-43.
- Stickens D, Clines G, Burbee D. Ramos P, Thomas S, Hogue D, et al. The EXT2 multiple exostoses gene defines a family of putative tumor suppressor genes. Nature Genet 1996;14:25-32.
- Bovee JVMG, Cleton-Jansen AM, Taminiau AHM, Hogendoorn PCW. Emerging pathways in the development of chondrosarcoma of bone and implications for targeted treatment. Lancet Oncology 2005;6(8):599-607.
- Bovee JVMG, Hogendoorn PCW, Wunder JS, Alman BA. Cartilage tumors and bone development: molecular pathology and possible therapeutic targets. Nat Rev Cancer 2010;10(7):481-8.
- Wicklund LC, Pauli RM, Johnston D, Hecht JT. Natural history study of hereditary multiple exostoses. Am J Med Genet 1995;55:43-6.
- de Andrea CE, Reijnders CM, Kroon HM, de JD, Hogendoorn PC, Szuhai K, et al. Secondary peripheral chondrosarcoma evolving from osteochondroma as a result of outgrowth of cells with functional EXT. Oncogene doi: 10.1038/onc.2011.311
- Pfleiderer AG, Thomson P, Milroy CM. View from beneath: pathology in focus. ENT presentation of Ollier's disease. J Laryngol Otol 1991;105(2):148-50.
- Mirra JM, Gold R, Downs J, Eckardt JJ. A new histologic approach to the differentiation of enchondroma and chondrosarcoma of the bones. A clinicopathologic analysis of 51 cases. Clin Orthop 1985;214–37.
- Reliability of Histopathologic and Radiologic Grading of Cartilaginous Neoplasms in Long Bones. J Bone Joint Surg Am 2007;89– A(10):2113-23.



- Eefting D, Schrage YM, Geirnaerdt MJ, Le CS, Taminiau AH, Bovee JV, et al. Assessment of interobserver variability and histologic
 parameters to improve reliability in classification and grading of central cartilaginous tumors. Am J Surg Pathol 2009;33(1):50-7.
- Evans HL, Ayala AG, Romsdahl MM. Prognostic factors in chondrosarcoma of bone. A clinicopathologic analysis with emphasis on histologic grading. Cancer 1977;40:818-31.
- Bjørnsson J, McLeod RA, Unni KK, listrup DM, Pritchard DJ. Primary chondrosarcoma of long bones and limb girdles. Cancer 1998;83:2105-19.
- McCarroll SA, Altshuler DM. Copy-number variation and association studies of human disease. Nat Genet 2007;39(7 Suppl):S37-S42.
- 28. Dutt A. Beroukhim R. Single nucleotide polymorphism array analysis of cancer, Curr Opin Oncol 2007;19(1):43-9.
- Botstein D, Risch N. Discovering genotypes underlying human phenotypes: past successes for mendelian disease, future approaches for complex disease. Nat Genet 2003;33 Suppl:228-37.
- Tuefferd M, de BA, Van dW, I, Talloen W, Verbeke T, Carvalho B, et al. Genome-wide copy number alterations detection in fresh frozen and matched FFPE samples using SNP 6.0 arrays. Genes Chromosomes Cancer 2008;47(11):957-64.
- Zhao X, Weir BA, LaFramboise T, Lin M, Beroukhim R, Garraway L, et al. Homozygous deletions and chromosome amplifications in human lung carcinomas revealed by single nucleotide polymorphism array analysis. Cancer Res 2005;65(13):5561-70.
- Beleza-Meireles A, Kockum I, Yuan QP, Picelli S, Wetterberg L, Gustavson KH, et al. Complex aetiology of an apparently Mendelian form of mental retardation. BMC Med Genet 2008:9:6.
- Suzuki M, Kato M, Yuyan C, Takita J, Sanada M, Nannya Y, et al. Whole-genome profiling of chromosomal aberrations in hepatoblastoma using high-density single-nucleotide polymorphism genotyping microarrays. Cancer Sci 2008;99(3):564-70.
- Kauffmann A, Huber W. Microarray data quality control improves the detection of differentially expressed genes. Genomics 2010:95(3):138-42.
- Kuhn K, Baker SC, Chudin E, Lieu MH, Oeser S, Bennett H, et al. A novel, high-performance random array platform for quantitative gene expression profiling. Genome Res 2004;14(11):2347-56.
- Brentani RR, Carraro DM, Verjovski-Almeida S, Reis EM, Neves EJ, de Souza SJ, et al. Gene expression arrays in cancer research; methods and applications. Crit Rev Oncol Hematol 2005;54(2):95-105.
- 37. Bibikova M, Fan JB. Genome-wide DNA methylation profiling. Wiley Interdiscip Rev Syst Biol Med 2010;2(2):210-23.
- Cheung HH, Lee TL. Rennert OM, Chan WY. DNA methylation of cancer genome. Birth Defects Res C Embryo Today 2009;87(4):335-50.
- Ramsahoye BH, Biniszkiewicz D, Lyko F, Clark V, Bird AP, Jaenisch R. Non-CpG methylation is prevalent in embryonic stem cells and may be mediated by DNA methyltransferase 3a. Proc Natl Acad Sci U S A 2000;97(10):5237-42.
- Saxonov S, Berg P, Brutlag DL. A genome-wide analysis of CpG dinucleotides in the human genome distinguishes two distinct classes of promoters. Proc Natl Acad Sci U S A 2006;103(5):1412-7.
- Schrage YM, Hameetman L, Szuhai K, Cleton-Jansen AM, Taminiau AHM, Hogendoorn PCW, et al. Aberrant heparan sulfate proteoglycan localization, despite normal exostosin, in central chondrosarcoma. Am J Pathol 2009;174(3):979-88.
- Lind T, Tufaro F, McCormick C, Lindahl U, Lidholt K. The putative tumor suppressors EXT1 and EXT2 are glycosyltransferases required for the biosynthesis of heparan sulfate. J Biol Chem 1998;273(41):26265-8.
- McCormick C, Duncan G, Goutsos KT, Tufaro F. The putative tumor suppressors EXT1 and EXT2 form a stable complex that accumulates in the golgi apparatus and catalyzes the synthesis of heparan sulfate. Proc Natl Acad Sci USA 2000;97(2):668-73.
- Presto J, Thuveson M, Carlsson P, Busse M, Wilen M, Eriksson I, et al. Heparan sulfate biosynthesis enzymes EXT1 and EXT2 affect NDST1 expression and heparan sulfate sulfation. Proc Natl Acad Sci U S A 2008;105(12):1045-55.
- Van der Eerden BCJ, Karperien M, Gevers EF, Lowik CWGM, Wit JM. Expression of Indian Hedgehog, PTHrP and their receptors in the
 postnatal growth plate of the rat: evidence for a locally acting growth restraining feedback loop after birth. J Bone Miner Res
 2000;15(6):1045-55.
- Bovee JVMG, Van den Broek LJCM, Cleton-Jansen AM, Hogendoorn PCW. Up-regulation of PTHrP and Bcl-2 expression characterizes
 the progression of osteochondroma towards peripheral chondrosarcoma and is a late event in central chondrosarcoma. Lab Invest
 2000:80:1925-33.



- Adams V, Hany MA, Schmid M, Hassam S, Briner J, Niggli FK. Detection of t(11;22)(q24;q12) translocation breakpoint in paraffinembedded tissue of the Ewing's sarcoma family by nested reverse transcription-polymerase chain reaction. Diagn Mol Pathol 1996;5(2):107-13.
- Hopyan S, Gokgoz N, Poon R, Gensure RC, Yu C, Cole WG, et al. A mutant PTH/PTHrP type I receptor in enchondromatosis. Nat Genet 2002;30(3):306-10.
- Rozeman LB, Sangiorgi L, Bruijn IH, Mainil-Varlet P, Bertoni F, Cleton-Jansen AM, et al. Enchondromatosis (Ollier disease, Maffucci syndrome) is not caused by the PTHR1 mutation p.R150C. Hum Mutat 2004;24(6):466-73.
- Couvineau A, Wouters V, Bertrand G, Rouyer C, Gerard B, Boon LM, et al. PTHR1 mutations associated with Ollier disease result in receptor loss of function. Hum Mol Genet 2008;17(18):2766-75.
- Ranger A, Szymczak A. Do intracranial neoplasms differ in Ollier disease and maffucci syndrome? An in-depth analysis of the literature. Neurosurgery 2009;65(6):1106-13.
- 52. Pansuriya TC, Kroon HM, Bovee JVMG. Enchondromatosis: insights on the different subtypes. Int J Clin Exp Pathol 2010;3(6):557-69.
- Schwartz HS, Zimmerman NB, Simon MA, Wroble RR, Millar EA, Bonfiglio M. The malignant potential of enchondromatosis. J Bone Joint Surg Am 1987;69(2):269-74.
- Jirarattanaphochai K, Jitpimolmard S, Jirarattanaphochai K. Maffucci's syndrome with frontal lobe astrocytoma. J Med Assoc Thai 1990;73(5):288-93.
- Goto H, Ito Y, Hirayama A, Sakamoto T, Kowada M. [Maffucci's syndrome associated with primary brain tumor: report of a case]. No Shinkei Geka 1987;15(9):971-5.
- Cremer H, Gullotta F, Wolf L. The Mafucci-Kast Syndrome. Dyschondroplasia with hemangiomas and frontal lobe astrocytoma. J Cancer Res Clin Oncol 1981;101(2):231-7.
- Lewis RJ, Ketcham AS. Maffucci's syndrome: functional and neoplastic significance. Case report and review of the literature. J Bone Joint Surg Am 1973;55(7):1465-79.
- Louis DN, Ohgaki H, Wiestler OD, Cavenee WK, Burger PC, Jouvet A, et al. The 2007 WHO classification of tumors of the central nervous system. Acta Neuropathol 2007;114(2):97-109.
- Yan H, Parsons DW, Jin G, McLendon R, Rasheed BA, Yuan W, et al. IDH1 and IDH2 mutations in gliomas. N Engl J Med 2009;360(8):765-73.
- Dang L, Jin S, Su SM. IDH mutations in glioma and acute myeloid leukemia. Trends Mol Med 2010;16(9):387-97.
- Amary MF, Bacsi K, Maggiani F, Damato S, Halai D, Berisha F, et al. IDH1 and IDH2 mutations are frequent events in central chondrosarcoma and central and periosteal chondromas but not in other mesenchymal tumors. J Pathol 2011;224(3):334-43.
- Dang L, White DW, Gross S, Bennett BD, Bittinger MA, Driggers EM, et al. Cancer-associated IDH1 mutations produce 2hydroxyglutarate. Nature 2009;462(7274):739-44.
- Rietveld L, Nieboer TE, Kluivers KB, Schreuder HW, Bulten J, Massuger LF. First case of juvenile granulosa cell tumor in an adult with Ollier disease. Int J Gynecol Pathol 2009;28(5):464-7.
- Leyva-Carmona M, Vazquez-Lopez MA, Lendinez-Molinos F. Ovarian juvenile granulosa cell tumors in infants. J Pediatr Hematol Oncol 2009;31(4):304-6.
- Kalfa N, Ecochard A, Patte C, Duvillard P, Audran F, Pienkowski C, et al. Activating mutations of the stimulatory g protein in juvenile ovarian granulosa cell tumors: a new prognostic factor? J Clin Endocrinol Metab 2006;91(5):1842-7.
- Lietman SA, Ding C, Levine MA. A highly sensitive polymerase chain reaction method detects activating mutations of the GNAS gene in peripheral blood cells in McCune-Albright syndrome or isolated fibrous dysplasia. J Bone Joint Surg Am 2005;87(11):2489-94.
- Cohen MM, Jr., Siegal GP, McCune-Albright syndrome. In: Fletcher C.D.M, Unni KK, Mertens F, editors. World Health Organization Classification of Tumors. Pathology & Genetics. Tumors of Soft Tissue and Bone Lyon: IARC Press; 2002. p. 357-9.