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## Chapter 3

### **Memory T cell inflation: understanding cause and effect**

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## Abstract

Typically, during viral infections, T cells encounter antigen, undergo proliferative expansion and ultimately contract into a pool of memory cells. However, after infection with cytomegalovirus, a ubiquitous  $\beta$ -herpesvirus, T cell populations specific for certain epitopes do not contract but instead are maintained and/or accumulate at high frequencies with a characteristic effector-memory phenotype. This feature has also been noted after other infections, for example, by parvoviruses. We discuss this so-called memory T cell inflation and the factors involved in this phenomenon. Also, we consider the potential therapeutic use of memory T cell inflation as a vaccine strategy and the associated implications for immune senescence.

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## MEMORY T CELL INFLATION AND CYTOMEGALOVIRUS INFECTION

The Herpesviridae are a large family of DNA viruses characterized by their ability to produce persistent infections. Human cytomegalovirus (HCMV) is a ubiquitous  $\beta$ -herpesvirus and 60–80% of people in the developed world display evidence of infection by the age of 80 years (1). HCMV infection in individuals with a healthy immune system is predominantly asymptomatic but on occasion can cause a mononucleosis-like disease similar to that caused by the  $\gamma$ -herpesvirus Epstein-Barr virus. In neonates and immunocompromised individuals, however, HCMV infection causes significant clinical problems. Currently, no vaccine is available and, although antiviral therapy exists, these drugs are both toxic and difficult to deliver and some viruses are resistant.

Of particular interest to immunologists and vaccinologists is the striking nature of the T cell response to HCMV, which is driven, in part, by the complex long-term, low-level persistence of the virus. Viral persistence and re-emergence are facilitated by the expression of numerous immune evasion genes encoded within the large viral genome. CD8<sup>+</sup> T lymphocytes play an important role in the control of intracellular pathogens, typified by large antigen-specific CD8<sup>+</sup> T cell populations after primary infection, which generally contract to form small pools of central memory T cells (2). Interestingly, comparison of HCMV-specific responses to other virus-specific T cell responses reveals the striking dominant impact of HCMV on the memory T cell compartment; a median of 9.1% and 10.2% of circulating CD4<sup>+</sup> and CD8<sup>+</sup> memory T cells are CMV-specific, respectively, which may increase even further in the elderly (3, 4).

The unusual nature of CMV-specific T cell responses has been dissected predominantly using mouse CMV (MCMV). Gradual accumulation of specific CD8<sup>+</sup> memory T cell populations was first noted in BALB/c mice (5) and subsequently described in longitudinal studies as memory ‘inflation’ (6). Memory T cell inflation is not restricted to CMVs, and arises also after infection with certain other persistent viruses (Box 1). Here, we focus on the factors that govern memory T cell inflation and the potential immune aging effects of memory T cell inflation in the elderly, and discuss recent research suggesting that inflationary T cell responses could be harnessed and used as a vaccine strategy.

## CHARACTERISTICS OF MEMORY T CELL INFLATION

Infection of C57BL/6 mice with MCMV produces a CD8<sup>+</sup> T cell response to at least 24 epitopes derived from 18 viral proteins in the acute phase of infection; however, 50 days post-infection. When viral latency has developed, the immune response is dominated by five singular epitopes that elicit inflationary responses [including two epitopes derived from the immediate early (IE) gene products] (7). The MCMV-specific CD8<sup>+</sup> T cell responses that dominate during the acute phase are typified by classical expansion, contraction and formation of long-term central memory pools (CD27<sup>+</sup>, CD28<sup>+</sup>, CD62L<sup>+</sup>, CD127<sup>+</sup> and IL-2<sup>+</sup>), whereas inflationary responses display an effector-memory phenotype (CD27<sup>-</sup>, CD28<sup>-</sup>, CD62L<sup>-</sup>, CD127<sup>-</sup> and IL-2<sup>+/-</sup>) (7-9) (Figure 1). These inflationary T cells are found in diverse hematopoietic and non-hematopoietic organs but their specificity is restricted to a limited number of viral epitopes.

In humans and in primate models of CMV, inflationary CMV-specific CD8<sup>+</sup> T cells accumulate at high frequencies and have a similar characteristic effector-memory phenotype (e.g. CD27<sup>-</sup>, CD28<sup>-</sup> etc.) (10-12). Inflationary T cells often show restricted T cell receptor (TCR) usage, with many carrying T cell receptors

**Box 1. Other persistent viruses that promote memory T cell inflation.**

Memory inflation is not confined to cytomegaloviruses; it occurs also after systemic, but not corneal, infection with herpes simplex virus-1 (HSV-1), an  $\alpha$ -herpesvirus, during which a subset of CD8<sup>+</sup> T cells specific for a structural glycoprotein B of the virus (gB-8p) gradually accumulates. Inflation begins as early as 3 months post-infection and these cells are maintained until old age. Ex vivo antigen-specific IFN- $\gamma$  production, and in vivo recall response to HSV-1 between adult and 'aged' memory T cells show that inflating cells can maintain their functionality (18,72).

In a murine polyomavirus (PyV) model, a 'memory inflating' immune response can be elicited by a PyV peptide presented by Q9, an MHC class Ib molecule. This peptide, from the VP2 capsid protein, elicits a CD8<sup>+</sup> T cell response that expands over 3 months and then reaches a plateau (73). Inflating CD8<sup>+</sup> T cells in MCMV infection are mostly effector memory cells displaying little or no expression of costimulatory molecules such as CD27 and CD28 or receptors for the homeostatic cytokines IL-7 and IL-15 (8,9). By contrast, CD8<sup>+</sup> T cells specific for PyV reach stable high frequencies, express receptors for both IL-7 and IL-2 and are maintained in the absence of homeostatic proliferation (74). It is not clear if the different phenotypes are related to differences in presentation by MHC 1a and MHC 1b molecules or in host-virus interaction or in different levels of antigen persistence.

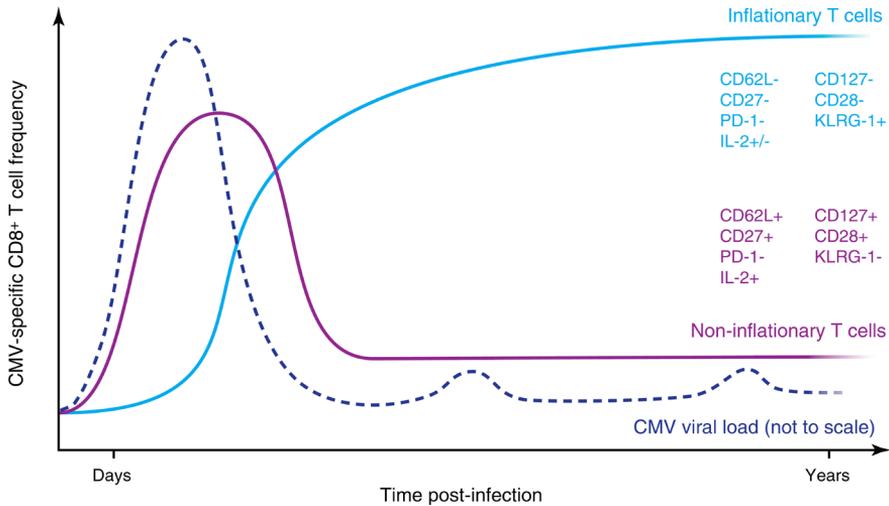
Memory inflation has been noted after acute parvovirus B19 infection in humans, with expansion of effector memory CD8<sup>+</sup> T cells many months after exposure (75,76). This small DNA virus is not a classical latent virus but the T cell responses appear phenotypically very similar to those directed against CMV and similar findings have been made in studies of the related parvovirus PARV4 (77). Thus, persistent viruses from different taxonomic families can elicit inflationary responses, indicating that memory T cell inflation is not a stranger in our midst but a more habitual feature of viral specific T cell responses.

specific for immunodominant CMV antigens (13). Inflation of memory CD4<sup>+</sup> T cells has also been observed but, at least in the mouse model of CMV, is less exaggerated (14) and outside the scope of this review. Despite the persistent nature of CMVs, inflationary T cells remain functional and do not show features of exhaustion as observed with certain chronic viral infections (Box 2) (9,10,15,16).

## FACTORS INVOLVED IN MEMORY T CELL INFLATION

### *Antigenic stimulation: antigen, tissues and cells*

The underlying mechanisms of memory T cell inflation are beginning to be understood (Figure 2). One factor driving this phenomenon must relate to repetitive antigen exposure (17,18). Support for the hypothesis that antigen persistence and re-exposure are required for memory T cell inflation comes from a series of observations and experiments. Firstly, inflationary memory T cells have phenotypically signs of activation such as low expression of CD62L, CD127, CD27 and CD28 (6,8). Moreover, adoptive transfer of inflationary T cells into naive recipients results in a failure to divide and survive, whereas the same cells are maintained when transferred into chronically infected mice (9). Also, when an epitope that, in the context of MCMV or HSV-1 infection, induces T cell inflation is expressed in a recombinant vaccinia virus, memory inflation is not elicited (6). This implies that long-term presentation of viral antigen is required and T cell intrinsic



**Figure 1.** Development of CMV-specific memory T cell populations.

During CMV infection, non-inflationary T cells undergo expansion, contraction and formation of stable memory pools. By contrast, inflationary T cell pools are characterized by lack of contraction and gradually accumulate in frequency over time aided by low levels of viral reactivation. Characteristic phenotypic features of inflationary versus non-inflationary T cells are shown.

factors such as TCR affinity or specificity do not drive memory inflation. In fact, the functional avidity for peptide–MHC complexes does not predict whether epitopes elicit inflationary or non-inflationary T cell responses, nor do they change during the course of a chronic infection (7,19). Furthermore, it was shown recently that an MCMV mutant unable to spread between cells by deletion of the essential glycoprotein L does not impair the development of memory T cell inflation (20). This suggests that once infection is established, systemic viral production is not a prerequisite and merely the presence of the virus in a latent state is sufficient to drive inflation.

The genes containing epitopes that induce inflationary responses can be expressed early or late in CMV infection (6,7). Moreover, both inflationary and non-inflationary epitopes may be found within the same protein, suggesting that differential antigen processing events may be involved as well (7). It is interesting to note that the immunoproteasome, which is constitutively expressed in professional antigen-presenting cells and particularly important for processing of MHC class I proteins, is less important for memory inflating than non-inflating responses (21). Thus, epitopes inducing memory inflation might be selected because of this difference in immunoproteasome dependence.

During acute infection, MCMV replicates in diverse tissues such as spleen, liver and lung, whereas the salivary glands are the primary site where viral replication is detected during the ensuing persistent phase (22,23). However, removal of salivary glands does not influence memory inflation (24), supporting the idea that tissues where the virus becomes latent are important. Although active virus production is not measurable during latency, viral peptide epitopes are most likely to be present. The ability of CMV to sporadically re-activate in certain tissues, such as the lung, and thereby produce relevant viral transcripts possibly contributes (5,25–27). The antiviral CD8<sup>+</sup> T cell pool during memory inflation is also not affected by thymectomy, suggesting that the contribution from the naive pool to ‘top-up’ such large populations is not crucial (24).

**Box 2. Inflation versus exhaustion.**

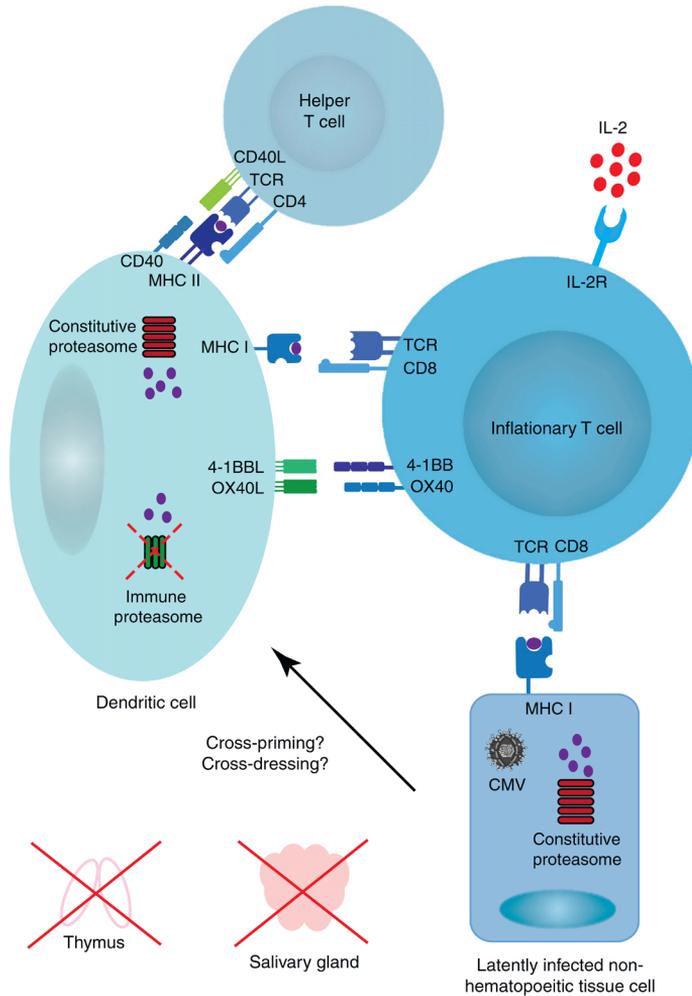
Chronic viral infections can produce a number of outcomes in man and mouse, and much attention has been paid to the impact of high levels of viremia on T cell function. First noted in the LCMV model, one key outcome is loss of T cell functionality, such as autocrine cytokine production, leading ultimately to lack of proliferation and deletion of responses, a process termed exhaustion (78). Related observations have been made in human infections such as HIV, and in both mouse and man T cell exhaustion is associated with high expression of inhibitory receptors, notably PD-1 (15,79). Although CMVs are persistent and the inflationary T cell responses show features of repetitive antigen exposure, they do not show features of exhaustion along the lines of the LCMV model. Instead, inflationary T cells retain their functionality, including proliferative capacity, cytotoxicity and cytokine secretion, and are low in PD-1 expression (9,10,15,16). A number of transcriptional features, such as high expression of T-Bet, EOMES and BLIMP1, have recently been revealed for HCMV-specific CD8<sup>+</sup> T cells (16) and it will be of interest to explore the role of these key transcriptional regulators *in vivo* using experimental CMV models.

It is clear that antigen persistence is important for memory inflation, but it remains unclear which cells present the viral antigens and whether both direct and cross-priming are involved. Despite the induction of CMV-encoded immunomodulatory proteins that interfere with MHC class I presentation, strong CD8<sup>+</sup> T cell responses are elicited, suggesting that cross-presentation plays a dominant role (28-30). Expression of MHC class I molecules by infected antigen-presenting cells may not be inhibited completely, and thus direct presentation cannot be excluded from contributing to priming. In *Batf3*<sup>-/-</sup> mice, which have impaired cross-priming owing to the lack of CD8α<sup>+</sup> and CD103<sup>+</sup> DCs, memory inflation still occurs (31) but cross-presentation by other cells in these mice is still possible (32). Using bone marrow chimera models, it was shown that inflationary response (e.g. to IE-1 protein) were dependent on antigen presentation by non-hematopoietic cells (33,34). However, it was not shown in these models whether direct presentation or cross-presentation (or potentially the recently described cross-dressing) (35) of MHC-loaded viral antigens (derived from infected non-hematopoietic cells) is important to drive memory inflation. Thus, further evidence is required to establish whether both direct and cross-presentation during persistent MCMV infection are operational and contribute to memory inflation.

*Costimulation*

In addition to TCR triggering, costimulatory molecules and inflammatory cytokines provide important signals for programming of effector and memory CD8<sup>+</sup> T cell responses (36). The main costimulatory receptor for T cells, CD28 (binding to CD80 or CD86), plays a dominant role during the acute phase because CD8<sup>+</sup> T cell responses in mice lacking this costimulatory interaction are strongly diminished at early time points. Remarkably, during the persistent phase, the inflationary T cells are able to 'catch up' to wild-type levels, indicating no CD28 requirement (37). By contrast, the costimulatory molecules 4-1BB and OX40, which are upregulated on T cells upon TCR triggering, clearly contribute to the development of inflationary MCMV specific T cell responses because interference with such triggering has a major impact on inflation (38,39).

## Requirements for memory T cell inflation



**Figure 2.** Factors that contribute to memory inflation in CMV infection.

In persistent CMV infection, the virus can sporadically reactivate from latency upon which viral proteins are processed by the constitutive proteasome and viral peptides are presented in MHC class I molecules. Viral antigens can either be presented by latently infected non-hematopoietic tissue cells or by cross-primed/cross-dressed dendritic cells. The presentation of the viral peptides persistently restimulates the inflationary T cell populations. Additional signals via the costimulatory molecules 4-1BB and OX40 contribute to the size of the inflationary T cell pool and IL-2 production is needed for maintenance of the inflationary T cells. CD4<sup>+</sup> T cell help is important for memory inflation but new thymic immigrants and viral replication in the salivary glands are not.

### Cytokines

Using a bone marrow chimeric mouse model with wild-type and *Il2ra*<sup>-/-</sup> cells, it was shown that an inflationary CMV-specific T cell response is dependent on IL-2, whereas a non-inflationary response was not (40). In an acute infectious model, it was shown recently that autocrine but not CD4<sup>+</sup> T cell or DC-derived IL-2 is important for optimal secondary expansion of memory CD8<sup>+</sup> T cell populations (41). Inflationary CD8<sup>+</sup> T cells produce lower levels of IL-2 than those produced by non-inflationary T cells (7,37) thus, the question is whether autocrine IL-2 is similarly crucial for inflationary CD8<sup>+</sup> T cells or if other sources supply this. The

cytokines IL-7 and IL-15, which are important for homeostatic proliferation of memory CD8<sup>+</sup> T cells, have potent proliferative properties for HCMV-specific T cells *in vitro* (42), but their precise role for memory T cell inflation in an *in vivo* setting remains to be elucidated. Another important cytokine is IL-10, which clearly restricts inflation of T cells in the chronic phase of CMV infection by unknown mechanisms (43).

#### *CD4<sup>+</sup> T cell help*

CD4<sup>+</sup> T cells are crucial for controlling MCMV replication (44,45), especially in the salivary gland during chronic infection, and it was shown recently that these cells operate via direct antiviral actions involving IFN- $\gamma$  (46). CD4<sup>+</sup> T cells are generally important for the development of memory CD8<sup>+</sup> T cell responses and even more so for inflationary responses (47,48). The mechanisms for this apparently increased helper-dependence are, however, unidentified.

We conclude that for the maintenance of the effector-memory-like inflationary T cell populations, the presence of latent virus in non-hematopoietic cells is essential and that additional cytokine and costimulatory signals (e.g. IL-2, OX40 and 4-1BB) are required to preserve the high frequency of this population.

### IMPORTANCE AND THERAPEUTIC EXPLOITATION OF MEMORY T CELL INFLATION

The development of CMV-related disease in the immunocompromised, such as post-transplant and AIDS patients, indicates that T cells are important in controlling CMV (49). Severe disease can develop in immunologically immature children after congenital infection or after infection of premature infants (50). The specific importance of T cells is underlined by the successful usage of adoptive immunotherapy, whereby donor T cells are stimulated *in vitro* using viral lysate or CMV-specific peptides and subsequently are transfused into the patient (51). CMV replication has been shown to be controlled in the majority of cases in a clinical trial of bone marrow transplant recipients (52) and there is similar, although more limited evidence for solid organ transplantation (53).

In murine models, depletion of multiple cell subsets (including B cells, CD4<sup>+</sup> T cells and/or NK cells) is required for full viral reactivation to occur (54). However, even in otherwise immunocompetent mice, there is evidence that inflating memory T cell pools specific for an epitope from an IE gene product provide a 'checkpoint' limiting the stochastic viral reactivation that occurs (5).

Experimental T cell-inducing vaccines typically use non-persistent vectors. In the initial phase post-vaccination, when antigen is readily available, an effector memory response develops. However, as the vector is eliminated, the immune response becomes centered on secondary lymphoid tissue and central memory cells become dominant. It is possible to theorize that these central-memory cells, which require re-exposure to antigen to drive proliferation, differentiation and trafficking, may not be the ideal subpopulation for rapid control of all proliferating pathogens in tissues. Instead, effector-memory T cells respond more rapidly and are located in different tissues, including mucosal sites. Thus, placing target antigens in a virus that induces effector-memory T cells could provide the holy grail of vaccinology for certain chronic viral infections and perhaps cancer. The property of CMV to induce such strong effector-memory T cell responses together with the relatively easy transmission (despite pre-existing immunity) and lack of pathogenicity in healthy animals has led to consideration of CMV-based vaccines in both human and animal populations.

*CMV as a vaccine vector*

A recombinant MCMV expressing foreign viral epitopes either from influenza A virus nucleoprotein (NP<sub>366-374</sub>) or lymphocytic choriomeningitis virus (LCMV) glycoprotein (GP<sub>33-41</sub>) induces ‘inflationary’, antigen-specific CD8<sup>+</sup> T cells at a high frequency. These cells protect against subsequent challenge with a recombinant vaccinia virus expressing either the nucleoprotein or the glycoprotein, respectively, as well as LCMV challenge with maintenance of protection up to 200 days post-infection (55). More recently, CMV was assessed as a vaccine against simian immunodeficiency virus (SIV), in rhesus macaques (*Macaca mulatta*) using rhesus CMV (RhCMV) expressing SIV Gag, Rev-Tat-Nef and Env (RhCMV-SIV). Regardless of RhCMV serostatus, macaques could be infected and developed transgene-specific CD4<sup>+</sup> and CD8<sup>+</sup> T cell responses that were predominantly effector-memory in nature and were very long-lived; antigen-specific responses could be detected up to 4 years post-vaccination (56). A subsequent study compared the phenotype of SIV-specific T cell responses in RhCMV seropositive macaques using persistent (RhCMV) vectors and traditional DNA prime /Ad5 boost. Macaques in this study also underwent mucosal SIVMAC239 infection. Vigorous CD4<sup>+</sup> and CD8<sup>+</sup> T cell responses to all vector-encoded SIV antigens were detectable up to 400 days post-vaccination in all groups and, although the kinetics of the SIV-specific T cell responses developed differently between groups, the magnitude of the total SIV-specific T cell response for both CD4<sup>+</sup> and CD8<sup>+</sup> T cells were similar by day 400. As expected, RhCMV induced an effector-memory phenotype (characterized as CD28<sup>-</sup>/CCR7<sup>-</sup>) contrasting with the central-memory phenotype induced by DNA/Ad5. At 400 days after initial vaccination, animals were challenged with SIVMAC239; macaques immunized with RhCMV-SIV displayed rapid complete control of SIV, and ~50% of the animals were protected for >1 year, which correlated with the magnitude of the SIV-specific CD8<sup>+</sup> T cell responses in the vaccine phase. By contrast, all of the DNA/Ad5-vaccinated macaques developed progressive infection, with reduced mean plasma viral load compared to controls (57). The recently reported development of spread-deficient CMV vectors (20) and attenuated CMV vectors expressing activating NK cell ligands (58) allays some of the safety and bioethical concerns about using persistent vectors with potential for person-to-person transmission, which could generate a potentially pathological immune response. Although hurdles still exist, the prospect of using CMV as a vaccine vector for infectious diseases or cancer is great.

**POTENTIAL DOWNSIDES OF MEMORY T CELL INFLATION**

In contrast to the potential utility of a CMV-driven immune response in vaccinology is the consideration that CMV may be deleterious to health. CMV is recognized to be associated with several vascular diseases, including atherosclerosis, restenosis, transplant vascular sclerosis (TVS) and chronic rejection of solid organ transplants (59). Recently, the Sacramento Area Latino Study on Aging showed that individuals with CMV IgG antibody titers in the highest quartile compared with lower quartiles had an all-cause mortality 1.43-fold greater, with a higher risk of cardiovascular mortality (hazard ratio = 1.35) (60).

*CMV and immunosenescence*

Definitions, mechanisms and markers are controversial but, broadly speaking, immunosenescence is used to describe the deleterious phenotypic and functional changes observed in the immune system as it ages. In the mid-1970s it was shown that 80-year-olds with anergic T cells had a 2-year mortality rate of 80%

compared to 35% in non-anegetic age-matched controls (61). In addition, age-associated poor response to T cell mitogens has been associated with increased mortality (62). Two major longitudinal studies, OCTO and NONA, focused on groups of healthy elderly volunteers and yielded a set of immune parameters that were capable of predicting mortality. Measurements such as a reduction in the number of B cells, poor T cell proliferative capacity and increased numbers of CD8<sup>+</sup> CD28<sup>-</sup> T cells, thus altering the CD4:8 ratio, were part of an immune risk profile (IRP) associated with increased all-cause mortality (63,64). Notably, evidence of CMV infection was part of this IRP cluster(65).

Infection with CMV substantially modifies the distribution of cellular subsets in peripheral blood, with greater numbers of CD8<sup>+</sup> T cells because of the increase in memory T cells and consequent reduction in size of the naive T cell pool. These changes are apparent in CMV seropositive individuals at all ages, and this might accelerate immunosenescence (66).

The increasing frequency of CMV-specific cells over a lifetime, especially at extremes of old age, suggests immune control of CMV is of paramount importance. Although some studies have associated CMV infection with increased risk of all-cause mortality, it is unclear at the molecular level why this is so. It has been reported that CMV-seropositive elderly do not respond as well as uninfected age-matched controls to influenza vaccine (67) but others have found no association of CMV with the poor responsiveness to influenza vaccination (68). Likewise, it was reported that CMV seropositivity results in reduced frequency of EBV-specific T cells in the elderly, suggesting CMV infection could affect responses to other viruses (69), but others found that the magnitude and phenotype of EBV- or influenza-specific CD8<sup>+</sup> T cells were not changed by primary CMV infection (70). The latter finding was confirmed in experimental models in which virus-based vaccines elicited an increase in the size of the memory CD8<sup>+</sup> T cell compartment with no impact on other cell populations, suggesting that the immune system retains plasticity and can accommodate development of new memory T cells against other pathogens (71). Overall, we conclude that the accumulation of clonal expansions of CD8<sup>+</sup> T cells specific for CMV antigens, excessive 'memory T cell inflation', in the very elderly might be a characteristic of immunosenescence, but further work is required to define a causal relationship.

## CONCLUDING REMARKS

Memory T cell inflation has emerged over the past few years as a prominent feature of CMV immunobiology. Like the definition of immune exhaustion that was observed initially in the LCMV mouse model and has now proven its importance for active chronic viral infections in humans, memory inflation could have important implications well beyond CMV persistence. In particular, exploiting the influence of the inflated populations to protect against infection and possibly cancer is an exciting prospect. To do this well we will need to understand the molecular signals that govern the maintenance and function of such T cell pools and develop CMV-based vaccine vectors that induce and sustain these. Finding the crucial signals that could underpin safe vaccine development is an important, practical and attainable goal for the future.

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## REFERENCE LIST

1. Cannon, M. J., D. S. Schmid, and T. B. Hyde. 2010. Review of cytomegalovirus seroprevalence and demographic characteristics associated with infection. *Rev. Med. Virol.* 20: 202-213.
2. Halwani, R., M. Doroudchi, B. Yassine-Diab, L. Janbazian, Y. Shi, E. A. Said, E. K. Haddad, and R. P. Sekaly. 2006. Generation and maintenance of human memory cells during viral infection. *Springer Semin. Immunopathol.* 28: 197-208.
3. Sylwester, A. W., B. L. Mitchell, J. B. Edgar, C. Taormina, C. Pelte, F. Ruchti, P. R. Sleath, K. H. Grabstein, N. A. Hosken, F. Kern, J. A. Nelson, and L. J. Picker. 2005. Broadly targeted human cytomegalovirus-specific CD4 and CD8 T cells dominate the memory compartments of exposed subjects. *J. Exp. Med.* 202: 673-685.
4. Komatsu, H., S. Sierro, V. Cuero, and P. Klenerman. 2003. Population analysis of antiviral T cell responses using MHC class I-peptide tetramers. *Clin. Exp. Immunol.* 134: 9-12.
5. Holtappels, R., M. F. Pahl-Seibert, D. Thomas, and M. J. Reddehase. 2000. Enrichment of immediate-early 1 (m123/pp89) peptide-specific CD8 T cells in a pulmonary CD62L(lo) memory-effector cell pool during latent murine cytomegalovirus infection of the lungs. *J. Virol.* 74: 11495-11503.
6. Karrer, U., S. Sierro, M. Wagner, A. Oxenius, H. Hengel, U. H. Koszinowski, R. E. Phillips, and P. Klenerman. 2003. Memory inflation: continuous accumulation of antiviral CD8 T cells over time. *J. Immunol.* 170: 2022-2029.
7. Munks, M. W., K. S. Cho, A. K. Pinto, S. Sierro, P. Klenerman, and A. B. Hill. 2006. Four distinct patterns of memory CD8 T cell responses to chronic murine cytomegalovirus infection. *J. Immunol.* 177: 450-458.
8. Sierro, S., R. Rothkopf, and P. Klenerman. 2005. Evolution of diverse antiviral CD8 T cell populations after murine cytomegalovirus infection. *Eur. J. Immunol.* 35: 1113-1123.
9. Snyder, C. M., K. S. Cho, E. L. Bonnett, D. S. van, G. R. Shellam, and A. B. Hill. 2008. Memory inflation during chronic viral infection is maintained by continuous production of short-lived, functional T cells. *Immunity.* 29: 650-659.
10. Cicin-Sain, L., A. W. Sylwester, S. I. Hagen, D. C. Siess, N. Currier, A. W. Legasse, M. B. Fischer, C. W. Koudelka, M. K. Axthelm, J. Nikolich-Zugich, and L. J. Picker. 2011. Cytomegalovirus-specific T cell immunity is maintained in immunosenescent rhesus macaques. *J. Immunol.* 187: 1722-1732.
11. Gamadia, L. E., E. M. van Leeuwen, E. B. Remmerswaal, S. L. Yong, S. Surachno, P. M. Wertheim-van Dillen, I. J. ten Berge, and R. A. van Lier. 2004. The size and phenotype of virus-specific T cell populations is determined by repetitive antigenic stimulation and environmental cytokines. *J. Immunol.* 172: 6107-6114.
12. Waller, E. C., E. Day, J. G. Sissons, and M. R. Wills. 2008. Dynamics of T cell memory in human cytomegalovirus infection. *Med. Microbiol. Immunol.* 197: 83-96.
13. Ouyang, Q., W. M. Wagner, A. Wikby, S. Walter, G. Aubert, A. I. Dodi, P. Travers, and G. Pawelec. 2003. Large numbers of dysfunctional CD8 T lymphocytes bearing receptors for a single dominant CMV epitope in the very old. *J. Clin. Immunol.* 23: 247-257.
14. Arens, R., P. Wang, J. Sidney, A. Loewendorf, A. Sette, S. P. Schoenberger, B. Peters, and C. A. Benedict. 2008. Cutting edge: murine cytomegalovirus induces a polyfunctional CD4 T cell response. *J. Immunol.* 180: 6472-6476.
15. Day, C. L., D. E. Kaufmann, P. Kiepiela, J. A. Brown, E. S. Moodley, S. Reddy, E. W. Mackey, J. D. Miller, A. J. Leslie, C. DePierres, Z. Mncube, J. Duraiswamy, B. Zhu, Q. Eichbaum, M. Altfeld, E. J. Wherry, H. M. Coovadia, P. J. Goulder, P. Klenerman, R. Ahmed, G. J. Freeman, and B. D. Walker. 2006. PD-1 expression on HIV-specific T cells is associated with T cell exhaustion and disease progression. *Nature* 443: 350-354.
16. Hertoghs, K. M., P. D. Moerland, S. A. van, E. B. Remmerswaal, S. L. Yong, P. J. van de Berg, S. M. van Ham, F. Baas, I. J. ten Berge, and R. A. van Lier. 2010. Molecular profiling of cytomegalovirus-induced human CD8 T cell differentiation. *J. Clin. Invest* 120: 4077-4090.
17. Klenerman, P., and A. Hill. 2005. T cells and viral persistence: lessons from diverse infections. *Nat. Immunol.* 6: 873-879.
18. Lang, A., J. D. Brien, and J. Nikolich-Zugich. 2009. Inflation and long-term maintenance of CD8 T cells responding to a latent herpesvirus depend upon establishment of latency and presence of viral antigens. *J. Immunol.* 183: 8077-8087.
19. Munks, M. W., M. C. Gold, A. L. Zajac, C. M. Doom, C. S. Morello, D. H. Spector, and A. B. Hill. 2006. Genome-wide analysis reveals a highly diverse CD8 T cell response to murine cytomegalovirus. *J. Immunol.* 176: 3760-3766.
20. Snyder, C. M., K. S. Cho, E. L. Bonnett, J. E. Allan, and A. B. Hill. 2011. Sustained CD8 T cell memory inflation after infection with a single-cycle cytomegalovirus. *PLoS. Pathog.* 7: e1002295.
21. Hutchinson, S., S. Sims, G. O'Hara, J. Silk, U. Gileadi, V. Cerundolo, and P. Klenerman. 2011. A dominant role for the immunoproteasome in CD8 T cell responses to murine cytomegalovirus. *PLoS. One.* 6: e14646.
22. Campbell, A. E., V. J. Cavanaugh, and J. S. Slater. 2008. The salivary glands as a privileged site of cytomegalovirus immune

- evasion and persistence. *Med. Microbiol. Immunol.* 197: 205-213.
23. Krmptotic, A., I. Bubic, B. Polic, P. Lucin, and S. Jonjic. 2003. Pathogenesis of murine cytomegalovirus infection. *Microbes Infect.* 5: 1263-1277.
  24. Loewendorf, A. I., R. Arens, J. F. Purton, C. D. Surh, and C. A. Benedict. 2011. Dissecting the Requirements for Maintenance of the CMV-Specific Memory T cell Pool. *Viral Immunol.*
  25. Grzimek, N. K., D. Dreis, S. Schmalz, and M. J. Reddehase. 2001. Random, asynchronous, and asymmetric transcriptional activity of enhancer-flanking major immediate-early genes *ie1/3* and *ie2* during murine cytomegalovirus latency in the lungs. *J. Virol.* 75: 2692-2705.
  26. Kurz, S. K., M. Rapp, H. P. Steffens, N. K. Grzimek, S. Schmalz, and M. J. Reddehase. 1999. Focal transcriptional activity of murine cytomegalovirus during latency in the lungs. *J. Virol.* 73: 482-494.
  27. Simon, C. O., R. Holtappels, H. M. Tervo, V. Bohm, T. Daubner, S. A. Oehrlein-Karpi, B. Kuhnappel, A. Renzaho, D. Strand, J. Podlech, M. J. Reddehase, and N. K. Grzimek. 2006. CD8 T cells control cytomegalovirus latency by epitope-specific sensing of transcriptional reactivation. *J. Virol.* 80: 10436-10456.
  28. Munks, M. W., A. K. Pinto, C. M. Doom, and A. B. Hill. 2007. Viral interference with antigen presentation does not alter acute or chronic CD8 T cell immunodominance in murine cytomegalovirus infection. *J. Immunol.* 178: 7235-7241.
  29. Pinto, A. K., M. W. Munks, U. H. Koszinowski, and A. B. Hill. 2006. Coordinated function of murine cytomegalovirus genes completely inhibits CTL lysis. *J. Immunol.* 177: 3225-3234.
  30. Snyder, C. M., J. E. Allan, E. L. Bonnett, C. M. Doom, and A. B. Hill. 2010. Cross-presentation of a spread-defective MCMV is sufficient to prime the majority of virus-specific CD8 T cells. *PLoS. One.* 5: e9681.
  31. Torti, N., S. M. Walton, K. M. Murphy, and A. Oxenius. 2011. Batf3 transcription factor-dependent DC subsets in murine CMV infection: Differential impact on T cell priming and memory inflation. *Eur. J. Immunol.*
  32. Asano, K., A. Nabeyama, Y. Miyake, C. H. Qiu, A. Kurita, M. Tomura, O. Kanagawa, S. Fujii, and M. Tanaka. 2011. CD169-positive macrophages dominate antitumor immunity by crosspresenting dead cell-associated antigens. *Immunity.* 34: 85-95.
  33. Seckert, C. K., S. I. Schader, S. Ebert, D. Thomas, K. Freitag, A. Renzaho, J. Podlech, M. J. Reddehase, and R. Holtappels. 2011. Antigen-presenting cells of haematopoietic origin prime cytomegalovirus-specific CD8 T cells but are not sufficient for driving memory inflation during viral latency. *J. Gen. Virol.* 92: 1994-2005.
  34. Torti, N., S. M. Walton, T. Brocker, T. Rulicke, and A. Oxenius. 2011. Non-hematopoietic cells in lymph nodes drive memory CD8 T cell inflation during murine cytomegalovirus infection. *PLoS. Pathog.* 7: e1002313.
  35. Wakim, L. M., and M. J. Bevan. 2011. Cross-dressed dendritic cells drive memory CD8 T cell activation after viral infection. *Nature* 471: 629-632.
  36. Arens, R., and S. P. Schoenberger. 2010. Plasticity in programming of effector and memory CD8 T cell formation. *Immunol. Rev.* 235: 190-205.
  37. Arens, R., A. Loewendorf, A. Redeker, S. Sierro, L. Boon, P. Klenerman, C. A. Benedict, and S. P. Schoenberger. 2011. Differential B7-CD28 costimulatory requirements for stable and inflationary mouse cytomegalovirus-specific memory CD8 T cell populations. *J. Immunol.* 186: 3874-3881.
  38. Humphreys, I. R., A. Loewendorf, T. C. De, K. Schneider, C. A. Benedict, M. W. Munks, C. F. Ware, and M. Croft. 2007. OX40 costimulation promotes persistence of cytomegalovirus-specific CD8 T Cells: A CD4-dependent mechanism. *J. Immunol.* 179: 2195-2202.
  39. Humphreys, I. R., S. W. Lee, M. Jones, A. Loewendorf, E. Gostick, D. A. Price, C. A. Benedict, C. F. Ware, and M. Croft. 2010. Biphasic role of 4-1BB in the regulation of mouse cytomegalovirus-specific CD8 T cells. *Eur. J. Immunol.*
  40. Bachmann, M. F., P. Wolint, S. Walton, K. Schwarz, and A. Oxenius. 2007. Differential role of IL-2R signaling for CD8 T cell responses in acute and chronic viral infections. *Eur. J. Immunol.* 37: 1502-1512.
  41. Feau, S., R. Arens, S. Togher, and S. P. Schoenberger. 2011. Autocrine IL-2 is required for secondary population expansion of CD8 memory T cells. *Nat. Immunol.* 12: 908-913.
  42. van Leeuwen, E. M., G. J. de Bree, E. B. Remmerswaal, S. L. Yong, K. Tesselaar, I. J. ten Berge, and R. A. van Lier. 2005. IL-7 receptor alpha chain expression distinguishes functional subsets of virus-specific human CD8 T cells. *Blood* 106: 2091-2098.
  43. Jones, M., K. Ladell, K. K. Wynn, M. A. Stacey, M. F. Quigley, E. Gostick, D. A. Price, and I. R. Humphreys. 2010. IL-10 restricts memory T cell inflation during cytomegalovirus infection. *J. Immunol.* 185: 3583-3592.
  44. Arens, R., A. Loewendorf, M. J. Her, K. Schneider-Ohrum, G. R. Shellam, E. Janssen, C. F. Ware, S. P. Schoenberger, and C. A. Benedict. 2011. B7-mediated costimulation of CD4 T cells constrains cytomegalovirus persistence. *J. Virol.* 85: 390-396.
  45. Jonjic, S., W. Mutter, F. Weiland, M. J. Reddehase, and U. H. Koszinowski. 1989. Site-restricted persistent cytomegalovirus infection after selective long-term depletion of CD4 T lymphocytes. *J. Exp. Med.* 169: 1199-1212.
  46. Walton, S. M., S. Mandarin, N. Torti, A. Zimmermann, H. Hengel, and A. Oxenius. 2011. Absence of cross-presenting cells

- in the salivary gland and viral immune evasion confine cytomegalovirus immune control to effector CD4 T cells. *PLoS Pathog.* 7: e1002214.
47. Snyder, C. M., A. Loewendorf, E. L. Bonnett, M. Croft, C. A. Benedict, and A. B. Hill. 2009. CD4 T cell help has an epitope-dependent impact on CD8 T cell memory inflation during murine cytomegalovirus infection. *J. Immunol.* 183: 3932-3941.
  48. Walton, S. M., N. Torti, S. Mandaric, and A. Oxenius. 2011. T cell help permits memory CD8 T cell inflation during cytomegalovirus latency. *Eur. J. Immunol.* 41: 2248-2259.
  49. Kotton, C. N. 2010. Management of cytomegalovirus infection in solid organ transplantation. *Nat. Rev. Nephrol.* 6: 711-721.
  50. Nigro, G., and S. P. Adler. 2011. Cytomegalovirus infections during pregnancy. *Curr. Opin. Obstet. Gynecol.* 23: 123-128.
  51. Riddell, S. R., and P. D. Greenberg. 1997. T cell therapy of human CMV and EBV infection in immunocompromised hosts. *Rev. Med. Virol.* 7: 181-192.
  52. Einsele, H., E. Roosnek, N. Rufer, C. Sinzger, S. Riegler, J. Loffler, U. Grigoleit, A. Moris, H. G. Rammensee, L. Kanz, A. Kleihauer, F. Frank, G. Jahn, and H. Hebart. 2002. Infusion of cytomegalovirus (CMV)-specific T cells for the treatment of CMV infection not responding to antiviral chemotherapy. *Blood* 99: 3916-3922.
  53. Brestrich, G., S. Zwinger, A. Fischer, M. Schmuck, A. Rohmhild, M. H. Hammer, A. Kurtz, L. Uharek, C. Knosalla, H. Lehmkuhl, H. D. Volk, and P. Reinke. 2009. Adoptive T cell therapy of a lung transplanted patient with severe CMV disease and resistance to antiviral therapy. *Am. J. Transplant.* 9: 1679-1684.
  54. Polic, B., H. Hengel, A. Krmpotic, J. Trgovcich, I. Pavic, P. Luccaronin, S. Jonjic, and U. H. Koszinowski. 1998. Hierarchical and redundant lymphocyte subset control precludes cytomegalovirus replication during latent infection. *J. Exp. Med.* 188: 1047-1054.
  55. Karrer, U., M. Wagner, S. Sierro, A. Oxenius, H. Hengel, T. Dumrese, S. Freigang, U. H. Koszinowski, R. E. Phillips, and P. Klenerman. 2004. Expansion of protective CD8 T cell responses driven by recombinant cytomegaloviruses. *J. Virol.* 78: 2255-2264.
  56. Hansen, S. G., C. Vieville, N. Whizin, L. Coyne-Johnson, D. C. Siess, D. D. Drummond, A. W. Legasse, M. K. Axthelm, K. Oswald, C. M. Trubey, M. Piatak, Jr., J. D. Lifson, J. A. Nelson, M. A. Jarvis, and L. J. Picker. 2009. Effector memory T cell responses are associated with protection of rhesus monkeys from mucosal simian immunodeficiency virus challenge. *Nat. Med.* 15: 293-299.
  57. Hansen, S. G., J. C. Ford, M. S. Lewis, A. B. Ventura, C. M. Hughes, L. Coyne-Johnson, N. Whizin, K. Oswald, R. Shoemaker, T. Swanson, A. W. Legasse, M. J. Chiuchiolo, C. L. Parks, M. K. Axthelm, J. A. Nelson, M. A. Jarvis, M. Piatak, Jr., J. D. Lifson, and L. J. Picker. 2011. Profound early control of highly pathogenic SIV by an effector memory T cell vaccine. *Nature* 473: 523-527.
  58. Slavuljica, I., A. Busche, M. Babic, M. Mitrovic, I. Gasparovic, D. Cekinovic, C. E. Markova, P. E. Pernjak, A. Cikovic, V. J. Lisnic, W. J. Britt, U. Koszinowski, M. Messerle, A. Krmpotic, and S. Jonjic. 2010. Recombinant mouse cytomegalovirus expressing a ligand for the NKG2D receptor is attenuated and has improved vaccine properties. *J. Clin. Invest* 120: 4532-4545.
  59. Streblov, D. N., J. Dumortier, A. V. Moses, S. L. Orloff, and J. A. Nelson. 2008. Mechanisms of cytomegalovirus-accelerated vascular disease: induction of paracrine factors that promote angiogenesis and wound healing. *Curr. Top. Microbiol. Immunol.* 325: 397-415.
  60. Roberts, E. T., M. N. Haan, J. B. Dowd, and A. E. Aiello. 2010. Cytomegalovirus antibody levels, inflammation, and mortality among elderly Latinos over 9 years of follow-up. *Am. J. Epidemiol.* 172: 363-371.
  61. Roberts-Thomson, I. C., S. Whittingham, U. Youngchaiyud, and I. R. Mackay. 1974. Ageing, immune response, and mortality. *Lancet* 2: 368-370.
  62. Murasko, D. M., P. Weiner, and D. Kaye. 1987. Decline in mitogen induced proliferation of lymphocytes with increasing age. *Clin. Exp. Immunol.* 70: 440-448.
  63. Nilsson, B. O., J. Ernerudh, B. Johansson, P. E. Evrin, S. Lofgren, F. G. Ferguson, and A. Wikby. 2003. Morbidity does not influence the T cell immune risk phenotype in the elderly: findings in the Swedish NONA Immune Study using sample selection protocols. *Mech. Ageing Dev.* 124: 469-476.
  64. Wikby, A., P. Maxson, J. Olsson, B. Johansson, and F. G. Ferguson. 1998. Changes in CD8 and CD4 lymphocyte subsets, T cell proliferation responses and non-survival in the very old: the Swedish longitudinal OCTO-immune study. *Mech. Ageing Dev.* 102: 187-198.
  65. Olsson, J., A. Wikby, B. Johansson, S. Lofgren, B. O. Nilsson, and F. G. Ferguson. 2000. Age-related change in peripheral blood T-lymphocyte subpopulations and cytomegalovirus infection in the very old: the Swedish longitudinal OCTO immune study. *Mech. Ageing Dev.* 121: 187-201.
  66. Chidrawar, S., N. Khan, W. Wei, A. McLarnon, N. Smith, L. Nayak, and P. Moss. 2009. Cytomegalovirus-seropositivity has a profound influence on the magnitude of major lymphoid subsets within healthy individuals. *Clin. Exp. Immunol.* 155: 423-432.
  67. Trzonkowski, P., J. Mysliwska, E. Szmit, J. Wieckiewicz, K. Lukaszuk, L. B. Brydak, M. Machala, and A. Mysliwski. 2003.

- Association between cytomegalovirus infection, enhanced proinflammatory response and low level of anti-hemagglutinins during the anti-influenza vaccination--an impact of immunosenescence. *Vaccine* 21: 3826-3836.
68. den Elzen, W. P., A. C. Vossen, H. J. Cools, R. G. Westendorp, A. C. Kroes, and J. Gussekloo. 2011. Cytomegalovirus infection and responsiveness to influenza vaccination in elderly residents of long-term care facilities. *Vaccine* 29: 4869-4874.
  69. Khan, N., A. Hislop, N. Gudgeon, M. Cobbold, R. Khanna, L. Nayak, A. B. Rickinson, and P. A. Moss. 2004. Herpesvirus-specific CD8 T cell immunity in old age: cytomegalovirus impairs the response to a coresident EBV infection. *J. Immunol.* 173: 7481-7489.
  70. van Leeuwen, E. M., J. J. Koning, E. B. Remmerswaal, B. D. van, R. A. van Lier, and I. J. ten Berge. 2006. Differential usage of cellular niches by cytomegalovirus versus EBV- and influenza virus-specific CD8 T cells. *J. Immunol.* 177: 4998-5005.
  71. Vezys, V., A. Yates, K. A. Casey, G. Lanier, R. Ahmed, R. Antia, and D. Masopust. 2009. Memory CD8 T cell compartment grows in size with immunological experience. *Nature* 457: 196-199.
  72. Lang, A., and J. Nikolich-Zugich. 2011. Functional CD8 T cell memory responding to persistent latent infection is maintained for life. *J. Immunol.* 187: 3759-3768.
  73. Swanson, P. A., A. R. Hofstetter, J. J. Wilson, and A. E. Lukacher. 2009. Cutting edge: shift in antigen dependence by an antiviral MHC class Ib-restricted CD8 T cell response during persistent viral infection. *J. Immunol.* 182: 5198-5202.
  74. Swanson, P. A., C. D. Pack, A. Hadley, C. R. Wang, I. Stroynowski, P. E. Jensen, and A. E. Lukacher. 2008. An MHC class Ib-restricted CD8 T cell response confers antiviral immunity. *J. Exp. Med.* 205: 1647-1657.
  75. Isa, A., V. Kasprovicz, O. Norbeck, A. Loughry, K. Jeffery, K. Broliden, P. Klenerman, T. Tolfvenstam, and P. Bowness. 2005. Prolonged activation of virus-specific CD8T cells after acute B19 infection. *PLoS. Med.* 2: e343.
  76. Norbeck, O., A. Isa, C. Pohlmann, K. Broliden, V. Kasprovicz, P. Bowness, P. Klenerman, and T. Tolfvenstam. 2005. Sustained CD8 T cell responses induced after acute parvovirus B19 infection in humans. *J. Virol.* 79: 12117-12121.
  77. Simmons, R., C. Sharp, S. Sims, H. Kloverpris, P. Goulder, P. Simmonds, P. Bowness, and P. Klenerman. 2011. High frequency, sustained T cell responses to PARV4 suggest viral persistence in vivo. *J. Infect. Dis.* 203: 1378-1387.
  78. Wherry, E. J. 2011. T cell exhaustion. *Nat. Immunol.* 12: 492-499.
  79. Barber, D. L., E. J. Wherry, D. Masopust, B. Zhu, J. P. Allison, A. H. Sharpe, G. J. Freeman, and R. Ahmed. 2006. Restoring function in exhausted CD8 T cells during chronic viral infection. *Nature* 439: 682-687.