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CHAPTER 4

CD226 (DNAM-1) IS ASSOCIATED WITH SUSCEPTIBILITY TO JUVENILE IDIOPATHIC ARTHRITIS

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ABSTRACT

Objectives

Juvenile idiopathic arthritis (JIA) is considered a complex genetic autoimmune disease. We investigated the association of genetic variants previously implicated in JIA, autoimmunity and/or immunoregulation, with susceptibility to JIA.

Methods

A genetic association study was performed in 639 JIA patients and 1613 healthy controls of North-West European descent. Ninety-three single nucleotide polymorphisms (SNPs) were genotyped in a candidate gene approach. Results of the entire JIA patient group (all subtypes) were compared to results obtained, alternatively, with a clinically homogeneous patient group including only oligoarticular and rheumatoid factor (RF) negative polyarticular JIA patients ($n=493$). Meta-analyses were performed for all SNPs that have been typed in other Caucasian JIA cohorts before.

Results

SNPs in or near *PTPN22*, *VTCN1*, the *IL2-IL21* region, *ANKRD55*, and *TNFA* were confirmed to be associated with JIA ($p<0.05$), strengthening the evidence for involvement of these genes in JIA. In the majority of these replicated SNPs, effect sizes were larger when analysing a homogeneous patient cohort than when analysing all subtypes. We identified two novel associations with oligoarticular and RF negative polyarticular JIA: *CD226 rs763361* (OR 1.30, 95%-CI 1.12-1.51, $p=0.0006$) and *CD28 rs1980422* (OR 1.29, 95%-CI 1.07-1.55, $p=0.008$). Meta-analyses including reported studies confirmed the association of both SNPs with susceptibility to JIA (OR 1.16, $p=0.001$ and OR 1.18, $p=0.001$, for rs763361 and rs1980422 respectively).

Conclusions

The *CD226* gene has been identified as novel association with JIA, and a SNP near *CD28* as a suggestive association. Both genes are probable candidate risk factors since they are involved in costimulation of T cells.

INTRODUCTION

Juvenile idiopathic arthritis (JIA) is the most common chronic rheumatic disease in childhood. Prevalence numbers vary from 4 to 400 per 100,000 children.¹ JIA comprises a heterogeneous group of conditions that share chronic arthritis with onset before the age of sixteen. Seven distinct subtypes have been defined by the International League of Associations of Rheumatologists (ILAR) based on clinical characteristics and laboratory parameters.² However, phenotypic overlap between subtypes does exist, particularly between oligoarthritis (persistent and extended) and rheumatoid factor (RF) negative polyarthritis. These subtypes are only distinguished on the basis of the number of affected joints at onset and during the course of the disease.³ A proportion of these patients have circulating antinuclear antibodies (ANA) and are specifically at risk for developing JIA-associated uveitis.³

The pathogenesis of JIA is not well understood. It is considered an autoimmune disease in which a deregulated T cell response towards an, as yet unidentified, self-antigen causes joint inflammation.⁴ In most subtypes, synovial inflammation, which eventually leads to bone erosion, is associated with an overproduction of pro-inflammatory cytokines, such as TNF- α and IL-17.⁵⁻⁹ JIA is a complex trait in which both genetic and environmental factors seem to be involved. Ethnic differences in epidemiologic studies,^{1,10} as well as an increased risk of JIA for relatives of patients (sibling recurrence risk ratio λ_S of 12),¹¹⁻¹³ form evidence for genetic contribution to the risk of JIA.

It has become increasingly clear that autoimmune diseases cluster in individuals and families.^{14,15} In line with this, genetic variations have been identified that are associated with more than one autoimmune disease, like rheumatoid arthritis (RA), type 1 diabetes mellitus, autoimmune thyroid disease, inflammatory bowel disease, systemic lupus erythematosus, multiple sclerosis, and also JIA.¹⁶⁻²⁰ These results imply the existence of general genetic susceptibility to autoimmune diseases.

Identification of genetic risk variants could contribute to understanding of disease pathways, improve diagnosis of (subtypes of) JIA, and ultimately improve prognosis by providing new targets for therapy. Both candidate gene and genome-wide association studies (GWAS) have been performed to elucidate the genetic basis of JIA. Compared to more common autoimmune diseases, until recently JIA cohorts were small and heterogeneous. Only few genetic associations had been replicated, such as *PTPN22* and the major histocompatibility complex (MHC) region.^{21,22} Other (suggestive) JIA susceptibility loci have been reported, but not confirmed, such as *ANKRD55* on 5q11.²² Nevertheless, replication of these loci is essential to exclude false positive associations. Therefore we investigated in a Caucasian JIA cohort

genetic loci previously implicated in JIA. Additionally, we investigated the association of genetic loci implicated in autoimmunity and/or immunoregulation with JIA. Analyses were performed in both a large but relatively heterogeneous JIA patient cohort (including all subtypes), and a smaller but phenotypically more homogeneous patient group (including only the persistent and extended oligoarticular and RF negative polyarticular subtypes).

METHODS

Subjects

DNA was available from 639 JIA patients with all subtypes, recruited through seven collaborating paediatric rheumatology referral centres in The Netherlands, Belgium, Germany, and Switzerland. All patients were of self-reported or parent-reported North-West European Caucasian descent. JIA cases were classified according to the revised ILAR criteria.² The patient group contained 263 patients with persistent oligoarthritis, 88 with extended oligoarthritis, and 142 with RF negative polyarthritis, resulting in a homogeneous group of 493 patients (Table 1).

DNA samples from healthy Caucasian controls were collected from three sources. See online supplementary methods for a detailed description of the control panels. Of the 93 markers that were successfully typed in the JIA patients, 40 were typed in 869 controls and 53 in 1319 controls (supplementary Tables S1, S2).

All patients and controls provided informed consent. The institutional review boards of all participating centres approved this study.

Genotyping

We genotyped single nucleotide polymorphisms (SNPs) adopting a candidate gene approach. The choice for specific genes and/or SNPs was based on previous reports suggesting involvement in JIA, other autoimmune diseases and/or immunoregulation. 93 SNPs located in 57 genes/loci passed quality control, listed in supplementary Table S2. See online supplementary methods for a detailed description of genotyping methods.

Statistical analysis

Allele frequencies were compared between JIA cases and controls. Allelic odds ratios (OR) and 95% confidence intervals (CI) were calculated using the allelic case-control association test in PLINK.²³ These ORs correspond to the genotypic ORs of an additive model. We differentiated between loci that have been associated with JIA before ('JIA replication loci'; 36 of 93 successfully typed SNPs) and loci that have not been impli-

Table 1
Patient characteristics of the JIA cohort

	n	(%)
Total cohort	639	
Gender		
Female	439	(69)
Male	200	(31)
Origin		
The Netherlands	324	(51)
Belgium	94	(15)
Germany	93	(15)
Switzerland	128	(20)
Subtype		
Persistent oligoarthritis	263	(41)
Extended oligoarthritis	88	(14)
RF negative polyarthritis	142	(22)
RF positive polyarthritis	22	(3)
Systemic JIA	73	(11)
Psoriatic arthritis	4	(<1)
Enthesitis related arthritis	3	(<1)
Undifferentiated JIA	44	(7)
ANA status		
Positive	280	(44)
Negative	229	(36)
Inconclusive/unknown	130	(20)
Family history ^a		
AID in 1 st degree relative	69	(16)
AID in 1 st or 2 nd degree relative	169	(38)

RF: rheumatoid factor; ANA: antinuclear antibodies; AID: autoimmune disease

a) Family history known of 442 patients

cated in JIA before (57 SNPs) (supplementary Table S2). For analysis of 'replication loci' we included patients with all JIA subtypes, to conform to reported studies that revealed these JIA loci. Because there was a prior probability that these loci would be associated with JIA in our study too, a p value < 0.05 was considered significant for these SNPs. We also performed the association analyses including only the most homogeneous JIA subtypes (persistent and extended oligoarthritis and RF negative polyarthritis, n=493). For the other 57 SNPs we analysed these 493 homogeneous JIA patients by comparing them as a group to controls. To adjust for multiple testing, a Bonferroni correction should be applied to the results for these 57 SNPs, leading to a significance threshold

of $p < 0.001$. Additionally, we performed ILAR subtype-specific case-control analyses (within the homogeneous patient group) for all 93 SNPs, and also compared all ANA positive patients within this group to controls (supplementary Table S3).

Meta-analyses were performed for all SNPs that have been investigated in JIA before (supplementary Tables S4, S5). We included reported genetic association studies in Caucasian case-control cohorts (including a mixed set of JIA subtypes), in which the same SNPs have been typed. We excluded studies for which data necessary to calculate allelic ORs were not available. Not all JIA replication loci were included for meta-analyses because of these inclusion and exclusion criteria. In case of overlapping individuals in reported studies, we only included the study with the largest JIA cohort. For meta-analyses of rs1980422 near *CD28* and rs763361 in *CD226*, we performed analyses in homogeneous (oligoarticular and RF negative polyarticular) JIA patients.^{18,24,25} Because of the small number of studies included in the meta-analyses, a fixed-effects model was used in a Mantel-Haenszel test.

All statistical analyses were performed with use of the software PLINK v1.07 (<http://pngu.mgh.harvard.edu/purcell/plink/>).²³

RESULTS

Replication of JIA loci

Thirty-six SNPs that have been previously reported to be associated with JIA were investigated in our entire JIA cohort consisting of 639 patients with all seven different subtypes. To conform to reported studies, all ILAR subtypes were included. Nine of 36 reported SNPs were confirmed to be associated ($p < 0.05$): *VTCN1* rs10923217, *KIAA1109* rs4505848, *IL21* rs1398553, *ANKRD55* rs6859219, *TNFA* rs1799724, rs1800750, rs361525, and rs1800610, and *MIF* rs755622 (Table 2). Because we propose to perform association studies only in a patient cohort as homogeneous as possible, additional analyses were limited to only oligoarthritis and RF negative polyarthritis (n=493). Although limiting the patient group led to fewer cases, all associations were still significant (Table 2). By performing analyses in the smaller, more homogeneous JIA cohort, the reported JIA SNPs *PTPN22* rs2476601 and *TNFA* rs1800629 were additionally confirmed. In the majority of these replicated associations, analysis in the more homogeneous patient cohort led to larger effect sizes. For all SNPs that had been investigated in Caucasian JIA cohorts before, a meta-analysis was performed. For 23 of 36 replication loci, appropriate data for meta-analyses were available, of which *PTPN22* rs2476601, *VTCN1* rs10923217, rs6669320, rs10923223 and rs12046117, *PTPRC* rs10919563, *AFF3* rs1160542, *CCR5* rs333, *TNFA* rs1799724,

Table 2. Association analyses of reported JIA loci

Chr	Position ^a	Gene/ region	SNP	Minor allele	All JIA subtypes			Only homogeneous JIA subtypes ^b			Reference
					MAF controls	MAF cases	OR (95% CI)	P (allelic)	MAF cases	OR (95% CI)	
1	114377568	PTPN22	rs2476601	A	0.10	0.12	1.26 (1.00-1.59)	0.051	0.13	1.32 (1.03-1.69)	0.02702
1	117685992	VTCN1	rs6673837	A	0.20	0.21	1.04 (0.86-1.24)	0.7101	0.20	0.99 (0.82-1.21)	0.9474
1	117690758	VTCN1	rs2358817	T	0.08	0.07	0.83 (0.63-1.10)	0.1933	0.07	0.88 (0.65-1.19)	0.4164
1	117711911	VTCN1	rs2358820	A	0.07	0.06	0.89 (0.66-1.20)	0.4467	0.07	0.92 (0.67-1.26)	0.602
1	117730048	VTCN1	rs10923217	C	0.48	0.52	1.20 (1.04-1.40)	0.01358	0.52	1.21 (1.03-1.42)	0.02079
1	117730623	VTCN1	rs6669320	A	0.15	0.13	0.89 (0.72-1.11)	0.3032	0.13	0.89 (0.70-1.12)	0.3117
1	117746573	VTCN1	rs10923223	C	0.15	0.16	1.16 (0.95-1.42)	0.1571	0.16	1.09 (0.88-1.36)	0.4383
1	117751365	VTCN1	rs12046117	T	0.13	0.14	1.11 (0.89-1.37)	0.3522	0.13	1.04 (0.82-1.32)	0.7252
1	198700442	PTPRC	rs10919563	A	0.13	0.12	0.91 (0.74-1.13)	0.3853	0.11	0.86 (0.68-1.08)	0.195
2	100832155	AFF3	rs1160542	G	0.45	0.46	1.05 (0.92-1.21)	0.4699	0.46	1.05 (0.90-1.22)	0.5177
2	100835734	AFF3	rs10865035	A	0.46	0.47	1.05 (0.91-1.23)	0.4924	0.47	1.05 (0.89-1.24)	0.5762
2	113590390	IL1B	rs1143634	A	0.25	0.24	0.93 (0.79-1.11)	0.4424	0.24	0.95 (0.79-1.14)	0.5542
2	204732714	CTLA4	rs231775	G	0.37	0.35	0.91 (0.78-1.06)	0.2112	0.35	0.92 (0.78-1.09)	0.3251
3	46414947	CCR5	rs333	del	0.10	0.09	0.83 (0.65-1.05)	0.1155	0.08	0.81 (0.62-1.06)	0.1168
4	123132492	KIAA1109	rs4505348	G	0.36	0.40	1.23 (1.07-1.42)	0.004019	0.40	1.20 (1.03-1.41)	0.01901
4	123348345	ADAD1	rs11732095	G	0.08	0.07	0.90 (0.68-1.17)	0.4229	0.07	0.91 (0.68-1.23)	0.5508
4	123514528	IL2/IL21	rs4492018	A	0.24	0.21	0.85 (0.72-1.00)	0.05315	0.21	0.88 (0.75-1.05)	0.1495
4	123548068	IL21	rs1398553	T	0.33	0.38	1.24 (1.07-1.43)	0.003359	0.38	1.22 (1.05-1.43)	0.0109
5	55438580	ANKRDS5	rs6859219	A	0.22	0.18	0.77 (0.64-0.92)	0.003175	0.17	0.74 (0.61-0.90)	0.002953
6	31542482	TNFA	rs1799724	T	0.10	0.13	1.37 (1.09-1.73)	0.007318	0.13	1.40 (1.09-1.79)	0.007489
6	31542963	TNFA	rs1800750	A	0.02	0.01	0.37 (0.17-0.80)	0.008935	0.01	0.36 (0.15-0.86)	0.01635
6	31543031	TNFA	rs1800629	A	0.17	0.15	0.87 (0.71-1.06)	0.172	0.14	0.79 (0.63-0.99)	0.04234

Table 2. Association analyses of reported JIA loci (continued)

Chr	Position ^a	Gene/ region	SNP	All JIA subtypes				Only homogeneous JIA subtypes ^b			
				Minor allele controls	MAF cases	OR (95% CI)	P (allelic)	MAF cases	OR (95% CI)	P (allelic)	Reference
6	31543101	TNFA	rs301525	A	0.05	0.03	0.56 (0.37-0.83)	0.003987	0.03	0.55 (0.35-0.86)	0.007526
6	31543827	TNFA	rs1800610	A	0.10	0.13	1.34 (1.06-1.69)	0.01332	0.13	1.38 (1.08-1.77)	0.009952
6	31544189	TNFA	rs3093662	G	0.06	0.04	0.75 (0.53-1.05)	0.09238	0.04	0.75 (0.52-1.09)	0.1328
6	138006504	TNFAIP3	rs6920220	A	0.21	0.20	0.98 (0.82-1.18)	0.8275	0.20	0.98 (0.80-1.19)	0.821
7	128594183	TNPO3	rs10488631	C	0.10	0.10	1.00 (0.79-1.26)	0.9903	0.10	1.03 (0.80-1.33)	0.8078
10	6053163	IL2RA	rs12722605	T	0.14	0.15	1.03 (0.85-1.25)	0.7694	0.15	1.05 (0.85-1.30)	0.6622
10	6099045	IL2RA	rs2104286	G	0.25	0.25	0.97 (0.82-1.15)	0.7435	0.23	0.91 (0.76-1.10)	0.3382
10	6114660	IL2RA	rs41295061	A	0.09	0.09	1.02 (0.81-1.30)	0.8551	0.09	1.02 (0.79-1.32)	0.8834
16	11179873	CLEC16A	rs12708716	G	0.35	0.34	0.94 (0.82-1.09)	0.4245	0.33	0.92 (0.78-1.07)	0.2729
16	11249329	CLEC16A	rs6498169	G	0.34	0.36	1.08 (0.93-1.24)	0.3049	0.37	1.09 (0.93-1.28)	0.2653
22	24236392	MIF	rs755622	C	0.21	0.16	0.69 (0.57-0.84)	0.0002126	0.15	0.67 (0.54-0.83)	0.0002084
22	37544245	IL2RB	rs3218258	T	0.28	0.28	0.99 (0.85-1.15)	0.8765	0.27	0.97 (0.82-1.15)	0.7135
22	375444810	IL2RB	rs3218253	T	0.28	0.28	1.01 (0.87-1.18)	0.8677	0.28	0.99 (0.84-1.17)	0.9164
22	37551607	IL2RB	rs743777	G	0.34	0.33	0.97 (0.84-1.12)	0.6348	0.33	0.95 (0.81-1.12)	0.5386

Chr: chromosome; MAF: minor allele frequency; OR: odds ratio; CI: confidence interval; LD: linkage disequilibrium with reported JIA locus

a) Base-pair position is based on NCBI dbSNP build 136

b) Patient group limited to oligoarticular (persistent and extended) and RF negative polyarticular JIA patients

TNFAIP3 rs6920220, *TNPO3* rs10488631, *IL2RA* rs2104286, and *CLEC16A* rs6498169 were significant (supplementary Table S4).

Novel JIA loci

An additional 57 SNPs, not reported to be associated with JIA before, located in or near autoimmune loci, were investigated in the homogeneous patient cohort. One SNP was strongly associated with JIA, *CD226* rs763361 (OR 1.30, 95% CI 1.12-1.51, p=0.0006). This SNP has not been reported to be associated with JIA before and represents a novel association (Table 3). The effect is particularly prominent in the persistent oligoarthritis patients (OR 1.39, p=0.0008) (supplementary Table S3). *CD226* rs763361 has been investigated before in two other Caucasian JIA cohorts and a non-significant trend of this SNP towards association with JIA (all subtypes) was reported (p=0.13¹⁸; p=0.059²⁵). We performed a meta-analysis combining the results from our study and the published data. To limit clinical heterogeneity between study cohorts, only patients with oligoarticular and RF negative polyarticular JIA were included in this meta-analysis. The meta-analysis revealed a combined association of this *CD226* variant with JIA, OR 1.16, p=0.001 (supplementary Table S5, Figure S1).

Rs1980422 near *CD28* was revealed as a suggestive association (OR 1.29, 95% CI 1.07-1.55, p=0.008) which was not significant after correction for multiple testing (Table 3). The effect of this SNP is particularly prominent in ANA-positive patients (OR 1.52, p=0.0004) (supplementary Table S3). This SNP was investigated before in a GWAS in Caucasian (oligoarticular and RF-negative polyarticular) JIA patients and a trend towards association was reported (p=0.04).²⁴ Also for this SNP we performed a meta-analysis combining the results of the present and the reported study, which resulted in a combined association of rs1980422 with JIA, OR 1.18, p=0.001 (supplementary Table S5, Figure S1).

Table 3

Polymorphisms in immune related genes associated with homogeneous subtypes of JIA^a

Chr	Position ^b	Gene/ region	SNP	Minor allele	MAF controls	MAF cases	OR (95% CI)	P (allelic)
2	204610396	<i>CD28</i>	rs1980422	C	0.22	0.27	1.29 (1.07-1.55)	0.008079
18	67531642	<i>CD226</i>	rs763361	T	0.47	0.54	1.30 (1.12-1.51)	0.0006295

Chr: chromosome; MAF: minor allele frequency; OR: odds ratio; CI: confidence interval

^aHomogeneous subtypes include oligoarticular (persistent and extended) and RF negative polyarticular JIA patients. Only SNPs that are (suggestively) associated with p<0.05 are listed.

^bBase-pair position is based on NCBI dbSNP build 136

Pooling our results with published non-significant associations (24 of 57 SNPs) revealed a potential novel association of *PRKCQ* rs4750316 with JIA (OR 0.90, p=0.01) (supplementary Table S5).

DISCUSSION

We investigated the role of 93 genetic markers previously associated with JIA or involved in autoimmunity or immunoregulation in a large Caucasian JIA cohort. We identified the *CD226* gene as a novel association with JIA, and a SNP near *CD28* as a suggestive association.

While this study was powered ($>=80\%$) to detect associations with an effect size (OR) $>= 1.4$, for SNPs with a minor allele frequency $>= 0.10$ at an alpha of 0.05, replication is the golden standard in genetic association studies and essential to exclude false-positive results. Collecting a large, homogeneous patient cohort which generates sufficient power is challenging in JIA, given the relatively low prevalence. A recent large study in JIA with dense genotyping of immune-related disease loci, which was published whilst this study was in progress, has also shown that increasing sample size improves power to detect true JIA loci.²² Although meta-analyses have limitations in case of publication bias and clinical heterogeneity between cohorts, they are also valuable for evaluating potentially false-positive or false-negative associations. Pooling of our and published results provides additional evidence for associations of 13 SNPs in 9 loci previously implicated in JIA. Furthermore, pooling of non-significant results from several studies suggests association of rs4750316 near *PRKCQ* with JIA, which is also an RA-locus.^{26,27} However, heterogeneity within a cohort (e.g. by including clinically different JIA subtypes) can be a pitfall leading to false-negative results.

We compared results obtained by analysing the entire JIA patient group in a case-control study to, alternatively, only JIA patients with the two clinically most similar ILAR subtypes (persistent and extended) oligoarthritis and RF negative polyarthritis). Although limiting the inclusion to two JIA subtypes resulted in smaller patient numbers, the study had enough power to confirm well-established JIA loci, e.g. *PTPN22* and *TNFA*. In the majority of replicated SNPs, effect sizes were larger when analysing only the homogeneous cohort. This argues in favour of restricting analyses to a homogeneous patient cohort, as has also been performed in two recent large studies in JIA.^{22,24}

The associated loci *PTPN22* (rs2476601), *VTCN1* (rs10923217), *KIAA1109* (rs4505848), *IL21* (rs1398553), *ANKRD55* (rs6859219), and *TNFA* (rs1799724,

rs1800750, rs1800629, rs361525, rs1800610) were replicated in this study strengthening their involvement in JIA. Four of these were confirmed in a meta-analysis: *PTPN22* rs2476601, *VTCN1* rs10923217, and *TNFA* rs1799724 and rs361525. An additional nine JIA replication SNPs, not confirmed by this study, were significantly associated in the meta-analysis. All of these genes are also associated with RA and/or other autoimmune diseases and are probably involved in the regulation of the immune response. All genes except *ANKRD55* have obvious roles in immune processes. It should be noted that the C allele of rs755622 in the promoter region of *MIF* (encoding the proinflammatory cytokine macrophage migration inhibitory factor) is associated with protection to JIA in our study, but this conflicts with results from other JIA cohorts that suggest association of the C allele with susceptibility.²⁸⁻³¹ The reason for these opposing results is not clear, but the minor allele frequency (MAF) of this SNP in our control cohort is comparable to the MAF in other North-West European control cohorts.^{29,32-34} Genetic studies of *MIF* in RA and inflammatory bowel disease have yielded similarly opposing results,^{29,35-40} indicating that the role of this SNP in JIA and other autoimmune diseases is still unclear. The other investigated JIA loci, of which *IL2RA* and *IL2RB* reached genome-wide significance in a previous report, were not confirmed in our cohort, which might be due to insufficient power to detect modest risk loci.²² A SNP in *IL2RA* was significant in the meta-analysis, underlining the importance of combining data from different JIA cohorts.

One of the strongest replicated associations with JIA is rs6859219, located at locus 5q11, in *ANKRD55*, an ankyrin repeat domain-containing gene with unknown function. This region has been recently identified by dense genotyping of immune-related disease loci in JIA patients.²² Although nearby genes *IL6ST* and *IL31RA* encode proteins involved in immunity, SNPs in these genes are not in linkage disequilibrium with the associated SNP in *ANKRD55*. Ankyrin repeat domains are common protein structures and mediate protein-protein interactions. Although the precise function of *ANKRD55* is not known, it is specifically expressed in resting CD4+ T cells (<http://www.amazonia.transcriptome.eu>), which is interesting when a role in autoimmunity is presumed. Rs6859219 was previously found to be associated with RA in a GWAS and with multiple sclerosis.⁴¹⁻⁴³

In addition to the confirmation of previously identified associations, two novel susceptibility loci for the most common JIA subtypes were discovered. *CD28* rs1980422 is only weakly associated, but is interesting because of its location between two immune related genes: *CD28* (approximately 10 kb away) and *CTLA4* or cytotoxic T-lymphocyte antigen-4 (approximately 129 kb away), a gene that is associated with multiple autoimmune diseases including RA, with conflicting results in JIA.^{18,21,29,44,45} There is minimal linkage disequilibrium between rs1980422 and

several SNPs in *CTLA4*. We also investigated the well-established general autoimmunity SNP rs231775 in *CTLA4*, but this SNP was not associated with JIA in this study. Both gene products have opposing roles in T cell activation. CD28 is expressed on the T cell surface and involved in costimulation of T cells. CTLA4 is expressed by T cells upon activation by antigen presenting cells. It functions as an attenuator of T cell activation by competing with CD28 for shared ligands (CD80 and CD86). Rs1980422 near *CD28* was associated with RA in a GWAS.⁴⁶ In a recent GWAS in JIA, this SNP was tested, but not significantly associated with JIA after correction for multiple testing.²⁴ Combining these results with ours in a meta-analysis resulted in a significant association. The *CD28* region has not been identified as a significant JIA susceptibility locus in a recent, large association study in which the ImmunoChip was used, but interestingly, this ImmunoChip study as well as the JIA GWAS revealed a strong association of SNPs in the region of *CD80*, coding for a ligand of CD28, with JIA.^{22,24} The implication of three components of this costimulatory pathway, CD80, CTLA4 and CD28, in JIA and other autoimmune diseases is supportive for a role in autoimmune pathogenesis.

The most strongly associated novel SNP rs763361 is located in *CD226*, which encodes CD226 or DNAX accessory molecule 1 (DNAM-1). DNAM-1 is a type 1 membrane protein belonging to the Ig-supergene family. It is mainly expressed on T and NK cells and is involved in the adhesion and costimulation of these cells via its ligands CD112 and CD155.^{47,48} The *CD226* region has not been identified as a (genome-wide significant) JIA susceptibility locus in the large association study with use of the ImmunoChip, which also captures *CD226*.²² Nevertheless, a meta-analysis of our study with two previous candidate gene studies confirmed the association with JIA.^{18,25} Furthermore, recent genetic studies have reported an association of this non-synonymous SNP (Gly307Ser) with susceptibility to multiple autoimmune diseases, as type 1 diabetes mellitus, autoimmune thyroid disease, multiple sclerosis, rheumatoid arthritis, and systemic lupus erythematosus, denoting it as a general autoimmunity locus.^{49,50} A fine-mapping study of the 18q22 region, in which the SNP lies, was performed in type 1 diabetes mellitus and multiple sclerosis patients by exonic resequencing and tag SNP mapping.⁴⁹ This study pointed out the SNP as a probable causal variant. However, this study cannot exclude other (rare) variants in linkage disequilibrium with rs763361 being the true causal variant. If rs763361 would be correlated with altered expression and/or signaling, this could explain the contribution of this variant to autoimmunity. DNAM-1 deficient mice show impaired control of viral infections and less cytotoxic activity against tumors compared to wild type mice, suggesting a form of immunodeficiency when *CD226* function is impaired.^{51,52} In another report in which the role of *CD226* in experimental autoimmune

encephalomyelitis (EAE, a model for multiple sclerosis) was studied, application of a monoclonal antibody against CD226 led to delayed onset and reduced severity of EAE.⁵³ This is suggestive for a role of CD226 in the development of autoimmune disease. By contrast, it is apparently contradictory that individuals that were homozygous for a *CD226* haplotype associated with susceptibility to systemic lupus erythematosus expressed lower *CD226* transcript levels and lower surface proteins on T cells and NK cells.⁵⁰ Replication in independent, homogeneous cohorts and additional fine-mapping and functional studies are needed to clarify the pathogenic implications of variation in these loci.

In summary, our data generate renewed interest for a role in JIA of two biological pathways that aid priming of T cells: the costimulatory mechanism involving CD28, CTLA4 and CD80/CD86, and the *CD226* gene, encoding the accessory molecule DNAM-1, which is a novel JIA susceptibility locus. This does not only contribute to knowledge of JIA pathogenesis, but targeting these T cell stimulating processes might also be of therapeutic interest.

REFERENCES

- (1) Thierry S, Fautrel B, Lemelle I et al. Prevalence and incidence of juvenile idiopathic arthritis: A systematic review. *Joint Bone Spine* 2013
- (2) Petty RE, Southwood TR, Manners P et al. International League of Associations for Rheumatology classification of juvenile idiopathic arthritis: second revision, Edmonton, 2001. *J Rheumatol* 2004; 31(2):390-392.
- (3) Ravelli A, Martini A. Juvenile idiopathic arthritis. *Lancet* 2007; 369(9563):767-778.
- (4) Prakken B, Albani S, Martini A. Juvenile idiopathic arthritis. *Lancet* 2011; 377(9783):2138-2149.
- (5) Lepore L, Pennesi M, Saletta S et al. Study of IL-2, IL-6, TNF alpha, IFN gamma and beta in the serum and synovial fluid of patients with juvenile chronic arthritis. *Clin Exp Rheumatol* 1994; 12(5):561-565.
- (6) Mangge H, Kenzian H, Gallistl S et al. Serum cytokines in juvenile rheumatoid arthritis. Correlation with conventional inflammation parameters and clinical subtypes. *Arthritis Rheum* 1995; 38(2):211-220.
- (7) Grom AA, Murray KJ, Luyrink L et al. Patterns of expression of tumor necrosis factor alpha, tumor necrosis factor beta, and their receptors in synovia of patients with juvenile rheumatoid arthritis and juvenile spondylarthropathy. *Arthritis Rheum* 1996; 39(10):1703-1710.
- (8) Nistala K, Moncrieffe H, Newton KR et al. Interleukin-17-producing T cells are enriched in the joints of children with arthritis, but have a reciprocal relationship to regulatory T cell numbers. *Arthritis Rheum* 2008; 58(3):875-887.
- (9) Agarwal S, Misra R, Aggarwal A. Interleukin 17 levels are increased in juvenile idiopathic arthritis synovial fluid and induce synovial fibroblasts to produce proinflammatory cytokines and matrix metalloproteinases. *J Rheumatol* 2008; 35(3):515-519.
- (10) Berkun Y, Padeh S. Environmental factors and the geoepidemiology of juvenile idiopathic arthritis. *Autoimmun Rev* 2010; 9(5):A319-A324.
- (11) Prahalad S, Shear ES, Thompson SD et al. Increased prevalence of familial autoimmunity in simplex and multiplex families with juvenile rheumatoid arthritis. *Arthritis Rheum* 2002; 46(7):1851-1856.
- (12) Prahalad S, O'brien E, Fraser AM et al. Familial aggregation of juvenile idiopathic arthritis. *Arthritis Rheum* 2004; 50(12):4022-4027.
- (13) Prahalad S, Zeft AS, Pimentel R et al. Quantification of the familial contribution to juvenile idiopathic arthritis. *Arthritis Rheum* 2010; 62(8):2525-2529.
- (14) Pohjankoski H, Kautiainen H, Kotaniemi K et al. Diabetes, coeliac disease, multiple sclerosis and chronic arthritis in first-degree relatives of patients with juvenile idiopathic arthritis. *Acta Paediatr* 2012; 101(7):767-771.
- (15) Cardenas-Roldan J, Rojas-Villarraga A, Anaya JM. How do autoimmune diseases cluster in families? A systematic review and meta-analysis. *BMC Med* 2013; 11:73.

- (16) Prahalad S, Hansen S, Whiting A et al. Variants in TNFAIP3, STAT4, and C12orf30 loci associated with multiple autoimmune diseases are also associated with juvenile idiopathic arthritis. *Arthritis Rheum* 2009; 60(7):2124-2130.
- (17) Hinks A, Eyre S, Ke X et al. Overlap of disease susceptibility loci for rheumatoid arthritis and juvenile idiopathic arthritis. *Ann Rheum Dis* 2010; 69(6):1049-1053.
- (18) Hinks A, Eyre S, Ke X et al. Association of the AFF3 gene and IL2/IL21 gene region with juvenile idiopathic arthritis. *Genes Immun* 2010; 11(2):194-198.
- (19) Thompson SD, Sudman M, Ramos PS et al. The susceptibility loci juvenile idiopathic arthritis shares with other autoimmune diseases extend to PTPN2, COG6, and ANGPT1. *Arthritis Rheum* 2010; 62(11):3265-3276.
- (20) Hinks A, Cobb J, Sudman M et al. Investigation of rheumatoid arthritis susceptibility loci in juvenile idiopathic arthritis confirms high degree of overlap. *Ann Rheum Dis* 2012; 71(7):1117-1121.
- (21) Prahalad S, Glass DN. A comprehensive review of the genetics of juvenile idiopathic arthritis. *Pediatr Rheumatol Online J* 2008; 6:11.
- (22) Hinks A, Cobb J, Marion MC et al. Dense genotyping of immune-related disease regions identifies 14 new susceptibility loci for juvenile idiopathic arthritis. *Nat Genet* 2013; 45(6):664-669.
- (23) Purcell S, Neale B, Todd-Brown K et al. PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am J Hum Genet* 2007; 81(3):559-575.
- (24) Thompson SD, Marion MC, Sudman M et al. Genome-wide association analysis of juvenile idiopathic arthritis identifies a new susceptibility locus at chromosomal region 3q13. *Arthritis Rheum* 2012; 64(8):2781-2791.
- (25) Ellis JA, Chavez RA, Pezic A et al. Independent replication analysis of genetic loci with previous evidence of association with juvenile idiopathic arthritis. *Pediatr Rheumatol Online J* 2013; 11(1):12.
- (26) Barton A, Thomson W, Ke X et al. Rheumatoid arthritis susceptibility loci at chromosomes 10p15, 12q13 and 22q13. *Nat Genet* 2008; 40(10):1156-1159.
- (27) Raychaudhuri S, Remmers EF, Lee AT et al. Common variants at CD40 and other loci confer risk of rheumatoid arthritis. *Nat Genet* 2008; 40(10):1216-1223.
- (28) Donn R, Alourfi Z, De BF et al. Mutation screening of the macrophage migration inhibitory factor gene: positive association of a functional polymorphism of macrophage migration inhibitory factor with juvenile idiopathic arthritis. *Arthritis Rheum* 2002; 46(9):2402-2409.
- (29) Mitterski B, Drynda S, Boschow G et al. Complex genetic predisposition in adult and juvenile rheumatoid arthritis. *BMC Genet* 2004; 5:2.
- (30) Berdelli A, Ozyurek AR, Ulger Z et al. Association of macrophage migration inhibitory factor gene -173 G/C polymorphism with prognosis in Turkish children with juvenile rheumatoid arthritis. *Rheumatol Int* 2006; 26(8):726-731.

- (31) Kaalla MJ, Broadway KA, Rohani-Pichavant M et al. Meta-analysis confirms association between TNFA-G238A variant and JIA, and between PTPN22-C1858T variant and oligoarticular, RF-polyarticular and RF-positive polyarticular JIA. *Pediatr Rheumatol Online J* 2013; 11(1):40.
- (32) Lehmann LE, Schroeder S, Hartmann W et al. A single nucleotide polymorphism of macrophage migration inhibitory factor is related to inflammatory response in coronary bypass surgery using cardiopulmonary bypass. *Eur J Cardiothorac Surg* 2006; 30(1):59-63.
- (33) Dambacher J, Staudinger T, Seiderer J et al. Macrophage migration inhibitory factor (MIF)-173G/C promoter polymorphism influences upper gastrointestinal tract involvement and disease activity in patients with Crohn's disease. *Inflamm Bowel Dis* 2007; 13(1):71-82.
- (34) Grigorenko EL, Han SS, Yrigollen CM et al. Macrophage migration inhibitory factor and autism spectrum disorders. *Pediatrics* 2008; 122(2):e438-e445.
- (35) Radstake TR, Sweep FC, Welsing P et al. Correlation of rheumatoid arthritis severity with the genetic functional variants and circulating levels of macrophage migration inhibitory factor. *Arthritis Rheum* 2005; 52(10):3020-3029.
- (36) de la Fontaine L, Schwarz MJ, Riedel M et al. Investigating disease susceptibility and the negative correlation of schizophrenia and rheumatoid arthritis focusing on MIF and CD14 gene polymorphisms. *Psychiatry Res* 2006; 144(1):39-47.
- (37) Martinez A, Orozco G, Varade J et al. Macrophage migration inhibitory factor gene: influence on rheumatoid arthritis susceptibility. *Hum Immunol* 2007; 68(9):744-747.
- (38) Palomino-Morales R, Gonzalez-Juanatey C, Vazquez-Rodriguez TR et al. Lack of association between macrophage migration inhibitory factor-173 gene polymorphism with disease susceptibility and cardiovascular risk in rheumatoid arthritis patients from northwestern Spain. *Clin Exp Rheumatol* 2010; 28(1):68-72.
- (39) Liu R, Xu N, Wang X et al. Influence of MIF, CD40, and CD226 polymorphisms on risk of rheumatoid arthritis. *Mol Biol Rep* 2012; 39(6):6915-6922.
- (40) Falvey JD, Bentley RW, Merriman TR et al. Macrophage migration inhibitory factor gene polymorphisms in inflammatory bowel disease: An association study in New Zealand Caucasians and meta-analysis. *World J Gastroenterol* 2013; 19(39):6656-6664.
- (41) Stahl EA, Raychaudhuri S, Remmers EF et al. Genome-wide association study meta-analysis identifies seven new rheumatoid arthritis risk loci. *Nat Genet* 2010; 42(6):508-514.
- (42) Alloza I, Otaegui D, de Lapuente AL et al. ANKRD55 and DHCR7 are novel multiple sclerosis risk loci. *Genes Immun* 2012; 13(3):253-257.
- (43) Lill CM, Schjeide BM, Graetz C et al. Genome-wide significant association of ANKRD55 rs6859219 and multiple sclerosis risk. *J Med Genet* 2013; 50(3):140-143.
- (44) Ueda H, Howson JM, Esposito L et al. Association of the T-cell regulatory gene CTLA4 with susceptibility to autoimmune disease. *Nature* 2003; 423(6939):506-511.
- (45) Plenge RM, Padyukov L, Remmers EF et al. Replication of putative candidate-gene associations with rheumatoid arthritis in >4,000 samples from North America and Sweden:

- association of susceptibility with PTPN22, CTLA4, and PADI4. *Am J Hum Genet* 2005; 77(6):1044-1060.
- (46) Raychaudhuri S, Thomson BP, Remmers EF et al. Genetic variants at CD28, PRDM1 and CD2/CD58 are associated with rheumatoid arthritis risk. *Nat Genet* 2009; 41(12):1313-1318.
- (47) Shibuya A, Campbell D, Hannum C et al. DNAM-1, a novel adhesion molecule involved in the cytolytic function of T lymphocytes. *Immunity* 1996; 4(6):573-581.
- (48) Xu Z, Jin B. A novel interface consisting of homologous immunoglobulin superfamily members with multiple functions. *Cell Mol Immunol* 2010; 7(1):11-19.
- (49) Hafler JP, Maier LM, Cooper JD et al. CD226 Gly307Ser association with multiple autoimmune diseases. *Genes Immun* 2009; 10(1):5-10.
- (50) Lofgren SE, Delgado-Vega AM, Gallant CJ et al. A 3'-untranslated region variant is associated with impaired expression of CD226 in T and natural killer T cells and is associated with susceptibility to systemic lupus erythematosus. *Arthritis Rheum* 2010; 62(11):3404-3414.
- (51) Welch MJ, Teijaro JR, Lewicki HA et al. CD8 T cell defect of TNF-alpha and IL-2 in DNAM-1 deficient mice delays clearance in vivo of a persistent virus infection. *Virology* 2012; 429(2):163-170.
- (52) Iguchi-Manaka A, Kai H, Yamashita Y et al. Accelerated tumor growth in mice deficient in DNAM-1 receptor. *J Exp Med* 2008; 205(13):2959-2964.
- (53) Dardalhon V, Schubart AS, Reddy J et al. CD226 is specifically expressed on the surface of Th1 cells and regulates their expansion and effector functions. *J Immunol* 2005; 175(3):1558-1565.
- (54) Hinks A, Barton A, John S et al. Association between the PTPN22 gene and rheumatoid arthritis and juvenile idiopathic arthritis in a UK population: further support that PTPN22 is an autoimmunity gene. *Arthritis Rheum* 2005; 52(6):1694-1699.
- (55) Hinks A, Barton A, Shephard N et al. Identification of a novel susceptibility locus for juvenile idiopathic arthritis by genome-wide association analysis. *Arthritis Rheum* 2009; 60(1):258-263.
- (56) Cimaz R, Cazalis MA, Reynaud C et al. IL1 and TNF gene polymorphisms in patients with juvenile idiopathic arthritis treated with TNF inhibitors. *Ann Rheum Dis* 2007; 66(7):900-904.
- (57) Prahalad S, Bohnsack JF, Jorde LB et al. Association of two functional polymorphisms in the CCR5 gene with juvenile rheumatoid arthritis. *Genes Immun* 2006; 7(6):468-475.
- (58) Albers HM, Kurreeman FA, Stoeken-Rijsbergen G et al. Association of the autoimmunity locus 4q27 with juvenile idiopathic arthritis. *Arthritis Rheum* 2009; 60(3):901-904.
- (59) Ozen S, Alikasifoglu M, Bakkaloglu A et al. Tumour necrosis factor alpha G-->A -238 and G-->A -308 polymorphisms in juvenile idiopathic arthritis. *Rheumatology (Oxford)* 2002; 41(2):223-227.

- (60) Zeggini E, Thomson W, Kwiatkowski D et al. Linkage and association studies of single-nucleotide polymorphism-tagged tumor necrosis factor haplotypes in juvenile oligoarthritis. *Arthritis Rheum* 2002; 46(12):3304-3311.
- (61) Hinks A, Ke X, Barton A et al. Association of the IL2RA/CD25 gene with juvenile idiopathic arthritis. *Arthritis Rheum* 2009; 60(1):251-257.
- (62) Skinningsrud B, Lie BA, Husebye ES et al. A CLEC16A variant confers risk for juvenile idiopathic arthritis and anti-cyclic citrullinated peptide antibody negative rheumatoid arthritis. *Ann Rheum Dis* 2010; 69(8):1471-1474.

SUPPLEMENTARY DATA

Supplementary methods

Control subjects

1130 controls were healthy blood bank donors (758 randomly selected by the Immunogenetics and Transplantation Immunology (ITI) section of the Department of Immunohematology and Bloodtransfusion, and 372 by the Laboratory for Diagnostic Genome Analyses (LDGA) at Leiden University Medical Center, all from the region of Leiden, The Netherlands), 372 controls were anonymized individuals that requested genetic counselling at the LDGA regarding a monogenic disease in their families, but tested negative for this single genetic defect, and 111 controls were recruited via participating patients and their families (but unrelated) (supplementary Table S1). Due to limited availability of DNA, genetic markers were genotyped in different control cohorts. Of the 93 markers that were successfully typed in the JIA patients, 40 were typed in 869 controls and 53 in 1319 controls (supplementary Tables S1, S2).

DNA and genotyping

DNA was isolated from buccal swabs or a blood sample. We performed genotyping of 112 SNPs in 65 genes/loci by iPLEX MassARRAY according to the manufacturer's recommendations (Sequenom, San Diego, California, USA). Only SNPs exceeding a 90% call rate and no evidence for deviation from Hardy-Weinberg equilibrium in the control population ($p > 0.005$) were used for further analysis. 93 of 112 SNPs (83%) located in 57 genes/loci passed quality control, listed in supplementary Table S2. SNP call rates per individual exceeded 90%.

Supplementary Table S1

Control subjects

Set	Source	n	Type of control subjects
1	ITI	464	Healthy blood donors
2	ITI	294	Healthy blood donors
3	LDGA	372	Healthy blood donors
4	LDGA	372	Healthy relatives of monogenic disease patients
5	JIA families	111	Healthy unrelated acquaintances of JIA patients

ITI: Immunogenetics and Transplantation Immunology section; LDGA: Laboratory for Diagnostic Genome Analyses

Supplementary Table S2 Allele and genotype frequencies in oligoarticular (persistent and extended) and RF negative polyarticular JIA patients versus controls

Chr	Position ^a	Gene/ region	SNP	Minor allele	MAF controls	MAF cases	OR (95% CI)	P (allelic)	Genotype counts	Genotype counts	Typed in control sets ^c	Reported JIA locus ^d
1	2553624	TNFRSF14-MMEL1	rs3890745	G	0.32	0.30	0.90 (0.77-1.06)	0.2137	126 / 568	570 47 / 187 / 231	1, 3, 4, 5 (n = 1319)	no
1	12091210	TNFRSF8-MMP	rs946461	T	0.26	0.29	1.15 (0.98-1.36)	0.09358	84 / 480	668 41 / 196 / 240	1, 3, 4, 5 (n = 1319)	no
1	12252955	TNFRSF1B	rs1061622	G	0.23	0.24	1.03 (0.85-1.24)	0.7678	44 / 309	492 34 / 160 / 281	1, 2, 5 (n = 869)	no
1	32729702	LCK	rs1004420	T	0.17	0.17	0.98 (0.80-1.19)	0.8087	46 / 332	839 11 / 135 / 314	1, 3, 4, 5 (n = 1319)	no
1	32743866	LCK	rs695161	C	0.48	0.47	0.97 (0.83-1.13)	0.6721	295 / 592	345 108 / 233 / 135	1, 3, 4, 5 (n = 1319)	no
1	114377568	PTPN22	rs2476601	A	0.10	0.13	1.32 (1.03-1.69)	0.02702	10 / 150	689 9 / 104 / 363	1, 2, 5 (n = 869)	yes
1	117635992	VTCN1	rs6673837	A	0.20	0.20	0.99 (0.82-1.21)	0.9474	40 / 265	544 19 / 154 / 302	1, 2, 5 (n = 869)	yes
1	117690758	VTCN1	rs2358817	T	0.08	0.07	0.88 (0.65-1.19)	0.4164	6 / 125	699 0 / 69 / 400	1, 2, 5 (n = 869)	yes
1	117711911	VTCN1	rs2558820	A	0.07	0.07	0.92 (0.67-1.26)	0.602	4 / 112	731 1 / 59 / 406	1, 2, 5 (n = 869)	yes
1	117730048	VTCN1	rs10923217	C	0.48	0.52	1.21 (1.03-1.42)	0.02079	201 / 397	129 132 / 232 / 109	1, 2, 5 (n = 869)	yes
1	117730623	VTCN1	rs6669320	A	0.15	0.13	0.89 (0.70-1.12)	0.3117	27 / 194	625 12 / 94 / 341	1, 2, 5 (n = 869)	yes
1	117746573	VTCN1	rs10923223	C	0.15	0.16	1.09 (0.88-1.36)	0.4383	13 / 221	614 15 / 119 / 341	1, 2, 5 (n = 869)	yes
1	117751365	VTCN1	rs12046117	T	0.13	0.13	1.04 (0.82-1.32)	0.7252	11 / 197	638 10 / 10 / 356	1, 2, 5 (n = 869)	yes
1	198700442	PTPRC	rs10919563	A	0.13	0.11	0.86 (0.68-1.08)	0.195	23 / 268	935 5 / 95 / 370	1, 3, 4, 5 (n = 1319)	yes
1	206946897	IL10	rs1800896	C	0.50	0.47	0.90 (0.77-1.06)	0.2119	219 / 408	219 92 / 238 / 115	1, 2, 5 (n = 869)	no
1	20705957	IL19	rs2243191	T	0.20	0.23	1.17 (0.96-1.43)	0.1209	27 / 283	556 21 / 155 / 261	1, 2, 5 (n = 869)	no
1	207038686	IL20	rs1400986	T	0.16	0.14	0.86 (0.69-1.08)	0.1975	23 / 230	595 7 / 122 / 344	1, 2, 5 (n = 869)	no
2	100832155	AFF3	rs1160542	G	0.45	0.46	1.05 (0.90-1.22)	0.5177	255 / 629	384 95 / 241 / 131	1, 3, 4, 5 (n = 1319)	yes
2	100835734	AFF3	rs10865035	A	0.46	0.47	1.05 (0.89-1.24)	0.5762	269 / 631	367 88 / 174 / 108	1, 3, 4, 5 (n = 1319)	yes
2	103070568	IL18RαP	rs917997	T	0.22	0.24	1.14 (0.94-1.37)	0.1782	45 / 283	519 29 / 172 / 272	1, 2, 5 (n = 869)	no
2	113537223	IL1A	rs17561	A	0.30	0.29	0.92 (0.77-1.09)	0.3289	74 / 366	406 32 / 194 / 226	1, 2, 5 (n = 869)	no
2	113542960	IL1A	rs1800587	A	0.31	0.28	0.89 (0.74-1.07)	0.2285	74 / 369	404 29 / 169 / 205	1, 2, 5 (n = 869)	no
2	113590390	IL1B	rs1143634	A	0.25	0.24	0.95 (0.79-1.14)	0.5542	57 / 306	485 20 / 185 / 269	1, 2, 5 (n = 869)	yes
2	113594867	IL1B	rs16944	A	0.33	0.35	1.11 (0.94-1.31)	0.2315	79 / 395	370 61 / 211 / 203	1, 2, 5 (n = 869)	no

Supplementary Table S2 Allele and genotype frequencies in oligoarticular (persistent and extended) and RF negative polyarticular JIA patients versus controls

Chr	Position ^a	Gene/ region	SNP	Minor allele	MAF controls	MAF cases	OR (95% CI)	P (allelic)	Genotype counts	Genotype controls	Genotype cases	Typed in control sets ^c	Reported JIA locus ^d	
2	162856148	DPP4	rs2268894	C	0.45	0.47	1.07 (0.91-1.25)	0.4192	164 / 437	164 / 247	99 / 246	1, 2, 5 (n = 869)	no	
2	191835596	STAT1	rs3771300	C	0.48	0.50	1.10 (0.94-1.27)	0.2321	281 / 608	337	118 / 238	1, 3, 4, 5 (n = 1319)	no	
2	191843445	STAT1	rs12010343	A	0.14	0.13	0.96 (0.77-1.20)	0.7449	29 / 287	924	10 / 107	354	1, 3, 4, 5 (n = 1319)	no
2	191845725	STAT1	rs1547550	C	0.35	0.34	0.95 (0.82-1.12)	0.5685	158 / 536	519	62 / 197	212	1, 3, 4, 5 (n = 1319)	no
2	191855521	STAT1	rs7562024	T	0.40	0.39	0.95 (0.82-1.11)	0.5502	203 / 579	447	71 / 221	174	1, 3, 4, 5 (n = 1319)	no
2	204610396	CD28	rs1980422	C	0.22	0.27	1.29 (1.07-1.55)	0.008079	65 / 433	759	30 / 144	203	1, 3, 4, 5 (n = 1319)	no
2	204732714	CTLA4	rs231775	G	0.37	0.35	0.92 (0.78-1.09)	0.3251	126 / 375	347	44 / 240	184	1, 2, 5 (n = 869)	yes
3	46414947	CCRS	rs333	del	0.10	0.08	0.81 (0.62-1.06)	0.1168	17 / 224	1028	5 / 68	392	1, 3, 4, 5 (n = 1319)	yes
3	58556841	FAM107A	rs13215591	C	0.07	0.08	1.13 (0.86-1.50)	0.3795	5 / 170	1092	2 / 71	397	1, 3, 4, 5 (n = 1319)	no
4	261083197	4p15	rs874040	G	0.31	0.29	0.93 (0.79-1.09)	0.3556	112 / 534	594	43 / 189	243	1, 3, 4, 5 (n = 1319)	no
4	123133492	KIAA1109	rs4505848	G	0.36	0.40	1.20 (1.03-1.41)	0.01901	170 / 567	538	73 / 222	166	1, 3, 4, 5 (n = 1319)	yes
4	12338345	ADA1	rs11732095	G	0.08	0.07	0.91 (0.68-1.23)	0.5508	4 / 182	1057	3 / 57	388	1, 3, 4, 5 (n = 1319)	yes
4	12354528	IL2-IL21	rs4492018	A	0.24	0.21	0.88 (0.73-1.05)	0.1495	72 / 464	740	19 / 161	283	1, 3, 4, 5 (n = 1319)	yes
4	123548068	IL21	rs1598553	T	0.33	0.38	1.22 (1.05-1.43)	0.0109	148 / 527	556	69 / 225	183	1, 3, 4, 5 (n = 1319)	yes
5	55438380	ANKR/D55	rs6859219	A	0.22	0.17	0.74 (0.61-0.90)	0.02953	55 / 441	774	14 / 131	320	1, 3, 4, 5 (n = 1319)	yes
5	96124330	ERAP1	rs30187	T	0.32	0.35	1.14 (0.97-1.34)	0.1018	139 / 510	574	55 / 224	196	1, 3, 4, 5 (n = 1319)	no
5	135287029	LECT2	rs351517	A	0.36	0.37	1.03 (0.88-1.20)	0.7415	166 / 561	514	63 / 223	191	1, 3, 4, 5 (n = 1319)	no
6	31540141	LTA	rs2239704	A	0.39	0.36	0.89 (0.75-1.05)	0.1671	131 / 386	324	58 / 222	192	1, 2, 5 (n = 869)	no
6	31540313	LTA	rs909253	G	0.34	0.35	1.07 (0.90-1.26)	0.4406	103 / 365	378	62 / 210	202	1, 2, 5 (n = 869)	no
6	31540784	LTA	rs1041981	A	0.33	0.35	1.09 (0.93-1.29)	0.2906	101 / 361	384	61 / 212	200	1, 2, 5 (n = 869)	no
6	31542482	TNFA	rs1799724	T	0.10	0.13	1.40 (1.09-1.79)	0.007489	11 / 145	691	10 / 105	356	1, 2, 5 (n = 869)	yes
6	31542963	TNFA	rs1800750	A	0.02	0.01	0.36 (0.15-0.86)	0.01635	0 / 30	818	0 / 6	465	1, 2, 5 (n = 869)	yes
6	31543031	TNFA	rs1800629	A	0.17	0.14	0.79 (0.63-0.99)	0.04234	29 / 228	591	9 / 114	353	1, 2, 5 (n = 869)	yes
6	31543101	TNFA	rs361525	A	0.05	0.03	0.55 (0.35-0.86)	0.007526	3 / 77	769	0 / 26	449	1, 2, 5 (n = 869)	yes
6	31543327	TNFA	rs1800610	A	0.10	0.13	1.38 (1.08-1.77)	0.009952	11 / 145	693	8 / 108	357	1, 2, 5 (n = 869)	yes

Supplementary Table S2 Allele and genotype frequencies in oligoarticular (persistent and extended) and RF negative polyarticular JIA patients versus controls

Chr	Position ^a	Gene/ region	SNP	Minor allele	MAF controls	OR (95% CI)	P (allelic)	Genotype counts	Genotype counts control cases	Typed in control sets ^c	Reported JIA locus ^d
6	31544189	TNFA	rs3093662	G	0.06	0.04	0.75 (0.52-1.09)	0.1328	4 / 91 / 752	2 / 38 / 430	1, 2, 5 (n = 869)
6	57012930	ZNF451	rs3734738	A	0.21	0.21	0.99 (0.82-1.19)	0.9212	53 / 428 / 772	20 / 158 / 290	1, 3, 4, 5 (n = 1319)
6	106568034	PRDM1	rs548234	C	0.32	0.35	1.13 (0.96-1.33)	0.1366	112 / 574 / 548	54 / 213 / 191	1, 3, 4, 5 (n = 1319)
6	138006504	TNFAIP3	rs6920220	A	0.21	0.20	0.98 (0.80-1.19)	0.821	41 / 269 / 538	25 / 139 / 301	1, 2, 5 (n = 869)
6	159482521	TACAP	rs394581	C	0.28	0.27	0.97 (0.82-1.15)	0.7544	103 / 500 / 670	33 / 188 / 246	1, 3, 4, 5 (n = 1319)
6	167534290	CCR6	rs3093023	A	0.44	0.46	1.07 (0.91-1.26)	0.3993	244 / 624 / 400	82 / 186 / 116	1, 3, 4, 5 (n = 1319)
7	75442759	CCL24	rs2302005	T	0.22	0.21	0.93 (0.78-1.12)	0.4639	55 / 400 / 703	14 / 171 / 292	1, 3, 4, 5 (n = 1319)
7	75442855	CCL24	rs2302004	C	0.44	0.42	0.94 (0.81-1.09)	0.4046	245 / 592 / 405	82 / 234 / 158	1, 3, 4, 5 (n = 1319)
7	92246744	CDK6	rs42041	G	0.26	0.25	0.95 (0.80-1.13)	0.5815	96 / 464 / 708	28 / 177 / 262	1, 3, 4, 5 (n = 1319)
7	128594183	TNPO3	rs10488631	C	0.10	0.10	1.03 (0.80-1.33)	0.8078	7 / 228 / 1023	5 / 83 / 382	1, 3, 4, 5 (n = 1319)
9	34743681	CCL21	rs951005	C	0.15	0.16	1.08 (0.88-1.34)	0.4482	24 / 295 / 849	7 / 136 / 354	1, 3, 4, 5 (n = 1319)
9	139775146	TRAF2	rs7048473	C	0.26	0.25	0.95 (0.80-1.13)	0.5385	78 / 487 / 691	30 / 171 / 269	1, 3, 4, 5 (n = 1319)
9	139787453	TRAF2	rs2811761	G	0.21	0.21	0.99 (0.83-1.20)	0.9556	59 / 405 / 774	22 / 155 / 296	1, 3, 4, 5 (n = 1319)
9	139815053	TRAF2	rs10781522	G	0.38	0.38	1.00 (0.86-1.17)	0.9959	176 / 602 / 488	77 / 198 / 192	1, 3, 4, 5 (n = 1319)
9	139821068	TRAF2	rs3750512	C	0.38	0.37	0.96 (0.82-1.12)	0.6014	180 / 570 / 479	74 / 203 / 199	1, 3, 4, 5 (n = 1319)
10	60533163	IL2RA	rs12722605	T	0.14	0.15	1.05 (0.85-1.30)	0.6622	32 / 295 / 916	12 / 111 / 340	1, 3, 4, 5 (n = 1319)
10	6099045	IL2RA	rs2104286	G	0.25	0.23	0.91 (0.76-1.10)	0.3382	76 / 472 / 695	29 / 139 / 252	1, 3, 4, 5 (n = 1319)
10	6114660	IL2RA	rs41295061	A	0.09	0.09	1.02 (0.79-1.32)	0.8834	6 / 219 / 1049	4 / 78 / 384	1, 3, 4, 5 (n = 1319)
10	6393260	PRKQ	rs4750316	C	0.19	0.19	0.99 (0.82-1.20)	0.9552	39 / 394 / 794	16 / 148 / 306	1, 3, 4, 5 (n = 1319)
11	36525293	TRAF6	rs540386	T	0.13	0.11	0.83 (0.66-1.05)	0.1194	26 / 285 / 941	4 / 99 / 364	1, 3, 4, 5 (n = 1319)
11	71709272	IL18BP	rs3814721	C	0.06	0.07	1.18 (0.85-1.64)	0.3216	1 / 95 / 751	0 / 63 / 408	1, 2, 5 (n = 869)
11	71710478	IL18BP	rs2298455	C	0.12	0.12	0.96 (0.75-1.24)	0.7774	11 / 179 / 656	7 / 90 / 355	1, 2, 5 (n = 869)
11	71714078	IL18BP	rs1541304	T	0.03	0.02	0.87 (0.52-1.46)	0.5935	1 / 43 / 805	0 / 22 / 454	1, 2, 5 (n = 869)
11	112035458	IL18	rs1946518	T	0.40	0.40	1.00 (0.85-1.18)	0.9851	137 / 395 / 312	64 / 233 / 158	1, 2, 5 (n = 869)
11	117869670	IL10RA	rs2229113	A	0.31	0.30	0.97 (0.81-1.16)	0.7345	85 / 327 / 394	40 / 205 / 227	1, 2, 5 (n = 869)

Supplementary Table S2 Allele and genotype frequencies in oligoarticular (persistent and extended) and RF negative polyarticular JIA patients versus controls

Chr	Position ^a	Gene/ region	SNP	Minor allele	MAF controls	OR (95% CI)	P (allelic)	Genotype counts	Genotype counts	Typed in control sets ^c	Reported JIA locus ^d	
12	6450945	TNFRSF1A	rs767455	C	0.42	0.40	0.92 (0.79-1.09)	0.3416	161 / 389 / 295	80 / 220 / 173	1, 2, 5 (n = 869)	no
12	6451590	TNFRSF1A	rs4149570	A	0.40	0.42	1.06 (0.90-1.25)	0.4766	151 / 377 / 315	87 / 213 / 164	1, 2, 5 (n = 869)	no
12	57968715	KIF5A	rs1678542	C	0.37	0.37	1.02 (0.87-1.19)	0.8226	135 / 599 / 442	66 / 217 / 184	1, 3, 4, 5 (n = 1319)	no
16	11179873	CLEC16A	rs12708716	G	0.35	0.33	0.92 (0.78-1.07)	0.2729	148 / 594 / 515	61 / 192 / 217	1, 3, 4, 5 (n = 1319)	yes
16	11243529	CLEC16A	rs6408169	G	0.34	0.37	1.09 (0.93-1.28)	0.2653	145 / 585 / 539	68 / 205 / 194	1, 3, 4, 5 (n = 1319)	yes
16	27448401	IL21R	rs3093341	G	0.10	0.08	0.80 (0.62-1.05)	0.1069	12 / 227 / 1005	1 / 77 / 399	1, 3, 4, 5 (n = 1319)	no
16	67189486	TRADD	rs11574518	T	0	0	not polymorphic	0 / 0 / 1143	0 / 0 / 450	1, 3, 4, 5 (n = 1319)	no	
17	32594568	CCl2-CCl7	rs8079244	C	0	0	not polymorphic	0 / 0 / 1236	0 / 0 / 473	1, 3, 4, 5 (n = 1319)	no	
17	40447401	STAT5A	rs7217728	C	0.31	0.29	0.91 (0.77-1.07)	0.2662	102 / 581 / 570	51 / 174 / 245	1, 3, 4, 5 (n = 1319)	no
17	40461003	STAT5A	rs2293154	A	0.18	0.18	1.05 (0.86-1.27)	0.6606	32 / 378 / 849	23 / 120 / 313	1, 3, 4, 5 (n = 1319)	no
18	67531642	CD226	rs763361	T	0.47	0.54	1.30 (1.12-1.51)	0.0006295	290 / 623 / 361	132 / 237 / 97	1, 3, 4, 5 (n = 1319)	no
19	44515514	ZNF230	rs12753	A	0.14	0.15	1.05 (0.85-1.30)	0.6291	22 / 304 / 905	10 / 121 / 346	1, 3, 4, 5 (n = 1319)	no
20	43280231	ADA	rs6031698	A	0	0	not polymorphic	0 / 0 / 844	0 / 0 / 472	1, 2, 5 (n = 869)	no	
20	44746982	CD40	rs1883832	T	0.25	0.25	0.99 (0.82-1.19)	0.9027	57 / 306 / 484	31 / 172 / 273	1, 2, 5 (n = 869)	no
21	34640788	IL10RB	rs2834167	G	0.24	0.27	1.14 (0.95-1.37)	0.1566	61 / 293 / 494	39 / 174 / 254	1, 2, 5 (n = 869)	no
22	24226392	MIF	rs755622	C	0.21	0.15	0.67 (0.54-0.83)	0.0002084	37 / 283 / 526	10 / 121 / 334	1, 2, 5 (n = 869)	yes
22	37544245	IL2RB	rs3218258	T	0.28	0.27	0.97 (0.82-1.15)	0.7135	99 / 514 / 655	31 / 196 / 243	1, 3, 4, 5 (n = 1319)	yes
22	37544810	IL2RB	rs3218253	T	0.28	0.28	0.99 (0.84-1.17)	0.9164	98 / 506 / 652	33 / 195 / 242	1, 3, 4, 5 (n = 1319)	yes
22	37551607	IL2RB	rs743777	G	0.34	0.33	0.95 (0.81-1.12)	0.5386	143 / 552 / 543	43 / 225 / 207	1, 3, 4, 5 (n = 1319)	yes

Chr: chromosome; MAF: minor allele frequency; OR: odds ratio; CI: confidence interval

a) Base-pair position is based on NCBI dbSNP build 136

b) 11: homozygous for the minor allele; 12: heterozygous; 22: homozygous for the major allele

c) See Supplementary Table S1 for details of control sets

d) See Table 2 for references

All presented results are derived from analyses with the homogeneous patient group (persistent or extended oligoarticular and RF negative polyarticular JIA) compared to controls.

Supplementary Table S3 Allele frequencies and associations per JIA subgroup versus controls

Gene/region	SNP	Minor allele	Controls n = 869 / 1319 ^a	JIA homogeneous ^b (n = 493) vs controls			Persistent oligoarthritis (n = 263) vs controls			Extended oligoarthritis (n = 88) vs controls			RF negative polyarthritis (n = 142) vs controls			ANA positive JIA ^c (n = 254) vs controls		
TNFRSF14- MME1	rs3890745	G	0.32	0.30	0.90	0.2137	0.31	0.92	0.4347	0.29	0.85	0.347	0.30	0.90	0.4611	0.28	0.83	0.08291
TNFRSF8-MIP	rs946461	T	0.26	0.29	1.15	0.09358	0.26	0.99	0.9563	0.33	1.40	0.0428	0.32	1.32	0.04465	0.30	1.20	0.09404
TNFRSF1B	rs1061622	G	0.23	0.24	1.03	0.7678	0.23	1.00	0.9709	0.22	0.91	0.6237	0.26	1.17	0.2849	0.26	1.16	0.21331
LCK	rs1004420	T	0.17	0.17	0.98	0.8087	0.18	1.03	0.8286	0.17	0.98	0.9097	0.16	0.88	0.4721	0.17	0.96	0.7445
LCK	rs695161	C	0.48	0.47	0.97	0.6721	0.46	0.94	0.5339	0.50	1.09	0.6046	0.47	0.95	0.6838	0.45	0.90	0.2833
PTPN22	rs2476601	A	0.10	0.13	1.32	0.02702	0.11	1.08	0.6668	0.14	1.51	0.07322	0.16	1.67	0.00493	0.13	1.39	0.03457
VTCN1	rs6673837	A	0.20	0.20	0.99	0.9474	0.21	1.07	0.5881	0.19	0.90	0.5935	0.19	0.92	0.6078	0.21	1.03	0.8411
VTCN1	rs2358817	T	0.08	0.07	0.88	0.4164	0.08	1.00	0.9886	0.08	1.00	0.9936	0.05	0.61	0.0817	0.08	0.94	0.7407
VTCN1	rs2358820	A	0.07	0.07	0.92	0.602	0.08	1.07	0.7114	0.07	0.96	0.8891	0.04	0.61	0.1135	0.07	1.06	0.7725
VTCN1	rs10923217	C	0.48	0.52	1.21	0.02079	0.51	1.12	0.2585	0.49	1.07	0.6787	0.58	1.50	0.002179	0.52	1.20	0.07851
VTCN1	rs6669320	A	0.15	0.13	0.89	0.3117	0.13	0.88	0.4457	0.13	0.83	0.4585	0.14	0.92	0.6767	0.14	0.94	0.6633
VTCN1	rs10923223	C	0.15	0.16	1.09	0.4383	0.17	1.21	0.1692	0.16	1.08	0.7348	0.13	0.89	0.5622	0.18	1.28	0.07157
VTCN1	rs12046117	T	0.13	0.13	1.04	0.7252	0.14	1.11	0.473	0.14	1.07	0.7856	0.12	0.90	0.6181	0.16	1.26	0.1072
PTPRC	rs10919563	A	0.13	0.11	0.86	0.195	0.12	0.96	0.7804	0.10	0.76	0.2868	0.10	0.73	0.14446	0.12	0.94	0.6871
IL10	rs1800896	C	0.50	0.47	0.90	0.2119	0.47	0.88	0.2208	0.45	0.82	0.2266	0.50	1.00	1	0.49	0.94	0.5658
IL19	rs2243191	T	0.20	0.23	1.17	0.1209	0.21	1.10	0.4737	0.25	1.36	0.1071	0.23	1.19	0.2811	0.22	1.15	0.2677
IL20	rs1400986	T	0.16	0.14	0.86	0.1975	0.16	0.98	0.857	0.17	1.04	0.8427	0.10	0.57	0.007092	0.15	0.90	0.4437
AFF3	rs1160542	G	0.45	0.46	1.05	0.5177	0.44	0.95	0.6405	0.52	1.35	0.06018	0.47	1.07	0.582	0.46	1.06	0.5746
AFF3	rs10865055	A	0.46	0.47	1.05	0.5762	0.45	0.97	0.7889	0.54	1.37	0.08522	0.47	1.03	0.8378	0.47	1.04	0.7041
IL18RAP	rs91797	T	0.22	0.24	1.14	0.1782	0.23	1.07	0.5657	0.27	1.34	0.1124	0.24	1.15	0.3743	0.25	1.17	0.1966
IL1A	rs17561	A	0.30	0.29	0.92	0.3289	0.29	0.91	0.4294	0.29	0.94	0.7177	0.28	0.90	0.4967	0.28	0.90	0.3559

Supplementary Table S3 Allele frequencies and associations per JIA subgroup versus controls (continued)

Gene/region	SNP	JIA homogenous ^b (n = 493) vs controls				Persistent oligoarthritis (n = 263) vs controls				Exerted oligoarthritis (n = 88) vs controls				RF negative polyarthritis (n = 142) vs controls				ANA positive JIA ^c (n = 254) vs controls			
		Minor allele	Controls n = 869 / 1319 ^a	MAF	OR	p	MAF	OR	p	MAF	OR	p	MAF	OR	p	MAF	OR	p	MAF	OR	p
<i>IL1A</i>	rs1800587	A	0.31	0.28	0.89	0.22285	0.29	0.95	0.6497	0.25	0.74	0.1427	0.28	0.89	0.4431	0.27	0.84	0.1471			
<i>IL1B</i>	rs1143634	A	0.25	0.24	0.95	0.5542	0.23	0.92	0.5044	0.23	0.89	0.5441	0.25	1.02	0.8818	0.23	0.91	0.4422			
<i>IL1B</i>	rs16944	A	0.33	0.35	1.11	0.2315	0.33	1.01	0.9639	0.39	1.32	0.09227	0.37	1.18	0.2235	0.35	1.12	0.3072			
<i>DPP4</i>	rs2268894	C	0.45	0.47	1.07	0.4192	0.47	1.06	0.5513	0.41	0.86	0.3464	0.50	1.24	0.105	0.46	1.04	0.6856			
<i>STAT1</i>	rs3771300	C	0.48	0.50	1.10	0.2321	0.47	0.96	0.7153	0.51	1.15	0.3818	0.55	1.35	0.01999	0.48	1.02	0.8538			
<i>STAT1</i>	rs13010345	A	0.14	0.13	0.96	0.7449	0.14	0.99	0.9477	0.12	0.86	0.5314	0.14	0.98	0.9254	0.15	1.09	0.5533			
<i>STAT1</i>	rs1547550	C	0.35	0.34	0.95	0.5685	0.35	0.98	0.8245	0.34	0.93	0.6744	0.33	0.93	0.5853	0.34	0.97	0.744			
<i>STAT1</i>	rs7562024	T	0.40	0.39	0.95	0.5502	0.39	0.95	0.6424	0.43	1.14	0.4157	0.36	0.85	0.2286	0.40	1.01	0.9559			
<i>CD28</i>	rs1980422	C	0.22	0.27	1.29	0.008079	0.26	1.23	0.09935	0.27	1.30	0.1918	0.29	1.39	0.03501	0.30	1.52	0.004481			
<i>CTLA4</i>	rs231775	G	0.37	0.35	0.92	0.3251	0.35	0.90	0.3164	0.36	0.94	0.716	0.36	0.95	0.6931	0.35	0.92	0.4572			
<i>CCR5</i>	rs333	del	0.10	0.08	0.81	0.1168	0.09	0.83	0.2842	0.06	0.57	0.08363	0.09	0.92	0.721	0.09	0.86	0.3884			
<i>FAM107A</i>	rs13315591	C	0.07	0.08	1.13	0.3795	0.08	1.15	0.4292	0.06	0.90	0.7552	0.09	1.25	0.3376	0.09	1.30	0.1396			
4p15	rs874040	G	0.31	0.29	0.93	0.3556	0.27	0.84	0.09887	0.36	1.28	0.1325	0.28	0.90	0.4424	0.30	0.95	0.6689			
<i>KIAA1109</i>	rs4505848	G	0.36	0.40	1.20	0.01901	0.38	1.13	0.2286	0.41	1.26	0.1603	0.42	1.31	0.03709	0.40	1.20	0.07744			
<i>ADAD1</i>	rs11732095	C	0.08	0.07	0.91	0.5508	0.07	0.90	0.6068	0.04	0.48	0.07923	0.09	1.20	0.4245	0.07	0.96	0.8327			
<i>IL2-IL21</i>	rs4492018	A	0.24	0.21	0.88	0.1495	0.20	0.81	0.08452	0.25	1.05	0.8018	0.22	0.89	0.4681	0.22	0.92	0.4964			
<i>IL21</i>	rs1398553	T	0.33	0.38	1.22	0.0109	0.38	1.21	0.05882	0.40	1.31	0.0934	0.38	1.20	0.1178	0.38	1.25	0.03146			
<i>ANKRD55</i>	rs6859219	A	0.22	0.17	0.74	0.002953	0.18	0.81	0.09507	0.19	0.84	0.3994	0.14	0.57	0.002153	0.16	0.69	0.005596			
<i>ERAP1</i>	rs30187	T	0.32	0.35	1.14	0.1018	0.34	1.09	0.4044	0.36	1.19	0.2774	0.36	1.20	0.163	0.35	1.12	0.2729			
<i>LECT2</i>	rs31517	A	0.36	0.37	1.03	0.7415	0.37	1.05	0.6601	0.34	0.93	0.6575	0.37	1.06	0.6833	0.36	1.02	0.8649			
<i>LTA</i>	rs2229704	A	0.39	0.36	0.89	0.1671	0.38	0.98	0.8285	0.30	0.68	0.02545	0.35	0.88	0.3353	0.37	0.94	0.5656			

Supplementary Table S3 Allele frequencies and associations per JIA subgroup versus controls (continued)

Gene/region	SNP	Minor allele	Controls n = 869 / 1319 ^a	JIA homogeneous ^b (n = 493) vs controls				Persistent oligoarthritis (n = 233) vs controls				Exerted oligoarthritis (n = 88) vs controls				RF negative polyarthritis (n = 142) vs controls				RF positive JIA ^c (n = 254) vs controls			
				MAF	OR	P	MAF	OR	P	MAF	OR	P	MAF	OR	P	MAF	OR	P	MAF	OR	P		
<i>LTA</i>	rs909253	G	0.34	0.35	1.07	0.4406	0.34	1.01	0.9397	0.39	1.25	0.1703	0.35	1.07	0.617	0.37	1.13	0.2434					
<i>LTA</i>	rs1041981	A	0.33	0.35	1.09	0.2906	0.34	1.04	0.7436	0.40	1.33	0.08439	0.35	1.07	0.6184	0.37	1.16	0.1618					
<i>TNFA</i>	rs1799724	T	0.10	0.13	1.40	0.007489	0.13	1.39	0.03609	0.13	1.39	0.1626	0.14	1.43	0.06627	0.15	1.58	0.002279					
<i>TNFA</i>	rs1800750	A	0.02	0.01	0.36	0.016335	0.01	0.34	0.06219	0.01	0.65	0.54285	0.00	0.20	0.08317	0.00	0.23	0.02768					
<i>TNFA</i>	rs1800629	A	0.17	0.14	0.79	0.04234	0.12	0.68	0.01005	0.15	0.87	0.5175	0.16	0.97	0.8566	0.14	0.80	0.1244					
<i>TNFA</i>	rs361525	A	0.05	0.03	0.55	0.007526	0.03	0.52	0.02582	0.04	0.83	0.6328	0.02	0.44	0.04588	0.02	0.36	0.002901					
<i>TNFA</i>	rs1800610	A	0.10	0.13	1.38	0.009952	0.13	1.42	0.02286	0.13	1.31	0.2737	0.13	1.37	0.1157	0.15	1.61	0.001418					
<i>TNFA</i>	rs3093662	G	0.06	0.04	0.75	0.1328	0.04	0.75	0.2442	0.05	0.88	0.7177	0.04	0.67	0.2213	0.03	0.55	0.02543					
<i>ZNF451</i>	rs3734738	A	0.21	0.21	0.99	0.9212	0.21	0.97	0.8117	0.23	1.12	0.5601	0.20	0.95	0.7468	0.22	1.06	0.622					
<i>PRDM1</i>	rs546234	C	0.32	0.35	1.13	0.1366	0.33	1.05	0.644	0.38	1.28	0.1353	0.36	1.19	0.1948	0.36	1.20	0.0787					
<i>TNFAIP3</i>	rs6920220	A	0.21	0.20	0.98	0.821	0.19	0.92	0.5275	0.22	1.08	0.7053	0.21	1.02	0.8938	0.20	0.98	0.8617					
<i>TAGAP</i>	rs394581	C	0.28	0.27	0.97	0.7544	0.28	1.00	0.9951	0.28	1.00	0.9958	0.26	0.91	0.5282	0.27	0.97	0.7971					
<i>CCR6</i>	rs393023	A	0.44	0.46	1.07	0.3993	0.47	1.12	0.3127	0.44	1.01	0.9509	0.45	1.04	0.7957	0.43	0.98	0.8225					
<i>CCL24</i>	rs2502005	T	0.22	0.21	0.93	0.4639	0.19	0.86	0.2088	0.23	1.06	0.7666	0.22	1.00	0.9886	0.21	0.93	0.5674					
<i>CCL24</i>	rs2502004	C	0.44	0.42	0.94	0.4046	0.38	0.81	0.03051	0.49	1.27	0.1343	0.44	1.02	0.8712	0.42	0.95	0.6442					
<i>CDK6</i>	rs42041	G	0.26	0.25	0.95	0.5815	0.26	1.02	0.8385	0.26	1.00	0.9918	0.22	0.80	0.15	0.25	0.95	0.6539					
<i>TNPO3</i>	rs10488631	C	0.10	0.10	1.03	0.8078	0.11	1.15	0.3883	0.09	0.98	0.9295	0.08	0.85	0.498	0.10	1.10	0.5706					
<i>CCL21</i>	rs95105	C	0.15	0.16	1.08	0.4482	0.17	1.15	0.2895	0.15	1.02	0.9258	0.15	1.00	0.992	0.17	1.17	0.2316					
<i>TRAF2</i>	rs7048473	C	0.26	0.25	0.95	0.5385	0.25	0.97	0.8145	0.24	0.89	0.5493	0.24	0.93	0.6308	0.25	0.95	0.6402					
<i>TRAF2</i>	rs2811761	G	0.21	0.21	0.99	0.9556	0.21	0.98	0.8389	0.20	0.93	0.7283	0.22	1.07	0.6615	0.23	1.11	0.3989					
<i>TRAF2</i>	rs10781522	G	0.38	0.38	1.00	0.9959	0.38	1.03	0.7762	0.37	0.99	0.9326	0.37	0.96	0.7444	0.39	1.06	0.5554					

Supplementary Table S3 Allele frequencies and associations per JIA subgroup versus controls (continued)

Gene/region	SNP	Minor allele		JIA homogeneous ^b (n = 493) vs controls		Persistent oligoarthritis (n = 233) vs controls		Eroded oligoarthritis (n = 88) vs controls		RF negative polyarthritis (n = 142) vs controls		ANA positive JIA ^c (n = 254) vs controls						
		Controls n = 869 / 1319 ^a	MAF	MAF	OR	p	MAF	OR	p	MAF	OR	p	MAF	OR	p			
<i>TRAF2</i>	rs3750512	C	0.38	0.37	0.96	0.6014	0.37	0.96	0.6644	0.37	0.96	0.7817	0.37	0.97	0.7972	0.38	1.03	0.7936
<i>IL2RA</i>	rs12722605	T	0.14	0.15	1.05	0.6622	0.16	1.11	0.4539	0.15	1.07	0.7598	0.14	0.93	0.6884	0.15	1.06	0.6882
<i>IL2RA</i>	rs2104286	G	0.25	0.23	0.91	0.3582	0.24	0.94	0.582	0.19	0.71	0.09939	0.25	1.02	0.9141	0.23	0.92	0.4751
<i>IL2RA</i>	rs41295061	A	0.09	0.09	1.02	0.8834	0.09	0.98	0.8898	0.10	1.14	0.6107	0.09	1.02	0.9163	0.09	1.00	0.993
<i>PRKQ</i>	rs4750316	C	0.19	0.19	0.99	0.9552	0.18	0.93	0.5832	0.21	1.13	0.5353	0.20	1.03	0.8562	0.22	1.15	0.2461
<i>TRAF6</i>	rs540386	T	0.13	0.11	0.83	0.1194	0.12	0.84	0.2268	0.11	0.83	0.4601	0.11	0.82	0.3398	0.11	0.76	0.08008
<i>IL18BP</i>	rs3814721	C	0.06	0.07	1.18	0.3216	0.07	1.18	0.4299	0.07	1.33	0.3517	0.06	1.09	0.7532	0.07	1.31	0.1792
<i>IL18BP</i>	rs2228455	C	0.12	0.12	0.96	0.7774	0.12	1.03	0.856	0.13	1.06	0.817	0.10	0.80	0.3006	0.13	1.07	0.6699
<i>IL18BP</i>	rs1541304	T	0.03	0.02	0.87	0.5935	0.02	0.74	0.4001	0.02	0.86	0.7822	0.03	1.11	0.798	0.02	0.76	0.4337
<i>IL18</i>	rs1946518	T	0.40	0.40	1.00	0.9851	0.42	1.11	0.3339	0.38	0.94	0.7224	0.36	0.86	0.2847	0.39	0.95	0.6597
<i>IL10RA</i>	rs2229113	A	0.31	0.30	0.97	0.7345	0.28	0.89	0.2876	0.34	1.17	0.353	0.31	1.01	0.9266	0.32	1.03	0.7612
<i>TNFRSF1A</i>	rs767455	C	0.42	0.40	0.92	0.3416	0.42	0.99	0.9141	0.37	0.82	0.2178	0.39	0.88	0.3469	0.39	0.89	0.2598
<i>TNFRSF1A</i>	rs4149570	A	0.40	0.42	1.06	0.4766	0.43	1.12	0.2806	0.43	1.12	0.484	0.39	0.93	0.5849	0.41	1.05	0.6677
<i>KIF5A</i>	rs1678542	C	0.37	0.37	1.02	0.8226	0.35	0.92	0.4395	0.42	1.25	0.1668	0.39	1.07	0.6106	0.37	0.99	0.8935
<i>CLEC16A</i>	rs12708716	G	0.35	0.33	0.92	0.2729	0.36	1.01	0.9414	0.36	1.02	0.8991	0.28	0.70	0.011183	0.31	0.82	0.05822
<i>CLEC16A</i>	rs6498169	G	0.34	0.37	1.09	0.2653	0.35	1.01	0.9101	0.37	1.10	0.5513	0.40	1.25	0.09136	0.38	1.18	0.1082
<i>IL21R</i>	rs3693341	G	0.10	0.08	0.80	0.1069	0.07	0.72	0.0596	0.10	1.04	0.8741	0.08	0.82	0.3732	0.08	0.77	0.1394
<i>TRADD</i>	rs11574518	T	0	0	Not polymorphic													
<i>CCL2-CCl7</i>	rs8079244	C	0	0	Not polymorphic													
<i>STAT5A</i>	rs7217728	C	0.31	0.29	0.91	0.2662	0.27	0.83	0.08635	0.28	0.86	0.3998	0.34	1.12	0.4273	0.30	0.94	0.5591
<i>STAT5A</i>	rs22933154	A	0.18	0.18	1.05	0.6606	0.18	1.00	0.9989	0.21	1.23	0.302	0.18	1.02	0.9117	0.19	1.11	0.4299

Supplementary Table S3 Allele frequencies and associations per JIA subgroup versus controls (continued)

Gene/region	SNP	Minor allele	JIA homogeneous ^b			Persistent oligoarthritis (n = 233) vs controls			Extended oligoarthritis (n = 88) vs controls			RF negative polyarthritis (n = 142) vs controls			ANA positive JIA ^c (n = 254) vs controls			
			Controls n = 869 / 1319 ^a	MAF	OR	P	MAF	OR	P	MAF	OR	P	MAF	OR	P	MAF	OR	P
CD226	rs763361	T	0.47	0.54	1.30	0.0006295 0.55	1.39	0.00008022 0.52	1.23	0.194	0.51	1.19	0.1818	0.53	1.27	0.01752		
ZNF230	rs12753	A	0.14	0.15	1.05	0.6291	0.15	1.09	0.549	0.20	1.53	0.03052	0.11	0.73	0.1149	0.17	1.26	0.07806
ADA	rs6031698	A	0	0	Not polymorphic													
CD40	rs1883832	T	0.25	0.99	0.9027	0.24	0.95	0.6524	0.22	0.88	0.4875	0.27	1.14	0.3614	0.24	0.97	0.8199	
IL10RB	rs2834167	G	0.24	0.27	1.14	0.1566	0.28	1.21	0.09756	0.21	0.83	0.339	0.28	1.23	0.1563	0.27	1.12	0.3515
MIF	rs755622	C	0.21	0.15	0.67	0.0002084 0.18	0.82	0.1387	0.11	0.47	0.002125	0.13	0.53	0.0009787 0.15	0.66	0.66	0.002712	
IL2RB	rs3218258	T	0.28	0.27	0.97	0.7135	0.26	0.92	0.4651	0.23	0.76	0.1478	0.32	1.22	0.1579	0.25	0.84	0.1144
IL2RB	rs3218253	T	0.28	0.28	0.99	0.9164	0.27	0.96	0.69	0.23	0.77	0.1579	0.32	1.22	0.1449	0.25	0.85	0.1534
IL2RB	rs743777	G	0.34	0.33	0.95	0.5386	0.32	0.91	0.3628	0.30	0.82	0.2601	0.37	1.12	0.3797	0.30	0.85	0.1558

MAF: minor allele frequency; RF: rheumatoid factor; ANA: antinuclear antibodies

a) Typed control sets per SNP are listed in Supplementary Table S2

b) Including oligoarthritis (persistent and extended) and RF negative polyarthritis patients

c) Including persistent oligoarthritis (n=129), extended oligoarthritis (n=60), and RF negative polyarthritis (n=65) patients

Supplementary Table S4
Meta-analyses of reported JIA loci^z

SNP (minor allele)	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
<i>PTPN22 rs2476601 (1858) (A)</i>							
Viken 2005	Norway	320	555				
Hinks 2005	UK	661	595	17%	1.53	(1.2 - 2)	
Seldin 2005	Finland	230	1400	5%	1.17	(0.9 - 1.5)	
Cinek 2006	Czech Republic	130	400	7%	2.35	(1.61 - 3.42)	
Pazár 2008	Hungary	150	200	3%	1.13	(0.66 - 1.95)	
Thompson 2010	USA, Germany, Czech Republic	949	1214	27%	1.67	(1.38 - 2.01)	
Ellis 2013	Australia	200	341	5%	1.45	(0.93 - 2.27)	
Dimopoulou 2013	Greece	128	221	2%	2.23	(1.03 - 4.84)	
Kaalla 2013	USA (overlap not reported)	636	733	18%	1.29	(1.02 - 1.62)	
Reinards et al.	present study			16%	1.32	(1.03 - 1.69)	
pooled					1.50		8.95E-16
<i>VTCN1 rs6672837 (A)</i>							
Hinks 2009 II	UK	654	1847	35%	1.16	(1 - 1.37)	
Thompson 2012	USA	810	3040	43%	0.95	(0.82 - 1.09)	
Reinards et al.	present study			22%	0.99	(0.82 - 1.21)	
pooled					1.03		0.5485
<i>VTCN1 rs2358817 (I)</i>							
Hinks 2009 II	UK	654	1847	27%	0.68	(0.52 - 0.89)	
Thompson 2012	USA	812	3056	51%	1.12	(0.92 - 1.36)	
Reinards et al.	present study			22%	0.88	(0.65 - 1.19)	
pooled					0.93		0.3035

Supplementary Table S4 (continued)
Meta-analyses of reported JIA loci^a

SNP (minor allele)	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
VTCN1 rs2358820 (A)	UK discovery	249	184	10%	0.40	(0.23 - 0.68)	
Hinks 2009	UK validation	321	2024	9%	0.45	(0.26 - 0.78)	
Thompson 2012	USA	814	3058	58%	1.10	(0.9 - 1.36)	
Reinards et al.	present study			24%	0.92	(0.67 - 1.26)	
pooled				0.88			0.1227
VTCN1 rs10923217 (C)	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
Hinks 2009	UK	654	1847	59%	1.17b	(1.02 - 1.33)	
Reinards et al.	present study			41%	1.21	(1.03 - 1.42)	
pooled				1.19			0.001113
VTCN1 rs6669320 (A)	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
Hinks 2009	UK	654	1847	62%	0.80	(0.67 - 0.97)	
Reinards et al.	present study			38%	0.89	(0.7 - 1.12)	
pooled				0.83			0.01299
VTCN1 rs10923223 (C)	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
Hinks 2009	UK	654	1847	58%	1.45	(1.2 - 1.75)	
Reinards et al.	present study			42%	1.09	(0.88 - 1.36)	
pooled				1.29			0.0005872
VTCN1 rs12046117 (T)	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
Hinks 2009	UK	654	1847	48%	1.58	(1.29 - 1.94)	
Ellis 2013	Australia	200	341	16%	1.15	(0.81 - 1.64)	
Reinards et al.	present study			36%	1.04	(0.82 - 1.32)	
pooled				1.29			0.0003681

Supplementary Table S4 (continued)
Meta-analyses of reported JIA loci^a

SNP (minor allele)	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
<i>PTPRC rs10919563 (A)</i>	UK, USA	1611 ^a	12719 ^a	80%	0.88	(0.78 - 0.99)	
Hinks 2012	USA						
Thompson 2012							
Reinards et al.	present study			20%	0.86	(0.68 - 1.08)	
pooled				0.88			0.01388
<i>AFF3 rs1160542 (G)</i>	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
Hinks 2010 II	UK	915	2967	4.0%	1.25	(1.13 - 1.39)	
Thompson 2012	USA	810	3054	3.4%	1.11	(0.99 - 1.24)	
Ellis 2013	Australia	200	341	7%	1.16	(0.91 - 1.49)	
Reinards et al.	present study			19%	1.05	(0.9 - 1.22)	
pooled				1.16			1.31E-05
<i>CTLA4 rs231775 (+49) (G)</i>	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
Miterski 2004	Germany	197	362	13%	0.92	(0.71 - 1.19)	
Suppiah 2005	UK (Northern Ireland)	72	475	7%	0.76	(0.53 - 1.09)	
Prahalaad 2008	USA (>90% of Northern European ancestry)	650	345	25%	0.95	(0.79 - 1.15)	
Thompson 2010	USA, UK	809 ^a	3521 ^a	22%	0.98	(0.84 - 1.15)	
Reinards et al.	present study			32%	0.92	(0.78 - 1.09)	
pooled				0.93			0.1224
<i>CCR5 rs333 (delta32) (del)</i>	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
Lindner 2007	Norway	515	645	22%	0.82	(0.63 - 1.07)	
Hinks 2010 III	UK	983	3121	56%	0.79	(0.66 - 0.94)	
Reinards et al.	present study			22%	0.81	(0.62 - 1.06)	
pooled				0.80			0.0005273

Supplementary Table S4 (continued)
Meta-analyses of reported IIA loci^{†*}

SNP (minor allele)	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
TNFA rs1799724 (-857) (T)							
Zeggini 2002 II	UK	170	415	28%	1.10	(0.74 - 1.63)	
Miterski 2004	Germany						
Reinards et al.	present study						
pooled				72%	1.40	(1.09 - 1.79)	0.01207
TNFA rs1800629 (-308) (A)							
Ozen 2002	Czech Republic	159	100	8%	1.48	(0.94 - 2.33)	
Zeggini 2002 II	UK	138	75	5%	2.12	(1.2 - 3.7)	
Miterski 2004	Germany	122	312	10%	1.13	(0.76 - 1.68)	
Schmeling 2006	Germany (overlap not reported)	228	196	11%	0.78	(0.53 - 1.13)	
Cimaz 2007	France (cases and controls), Italy (cases)	107	630				
Mourão 2009	Portugal	114	117				
Kaalla 2013	USA	628	729	35%	0.82	(0.66 - 1.01)	
Reinards et al.	present study						
pooled				32%	0.79	(0.63 - 0.99)	0.1609
TNFA rs361525 (-238) (A)							
Ozen 2002	Czech Republic	159	100	22%	2.02	(1.28 - 3.19)	
Zeggini 2002 II	UK	137	76	6%	0.41	(0.17 - 0.95)	
Miterski 2004	Germany	130	375	6%	0.71	(0.29 - 1.77)	
Schmeling 2006	Germany (overlap not reported)	228	196	7%	1.30	(0.58 - 2.93)	
Cimaz 2007	France (cases and controls), Italy (cases)	107	630				
Kaalla 2013	USA	638	749	35%	0.66	(0.46 - 0.95)	
Reinards et al.	present study						
pooled				23%	0.55	(0.35 - 0.86)	0.08965

Supplementary Table S4 (continued)
Meta-analyses of reported JIA loci^a

SNP (minor allele)	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
<i>TNFAIP3 rs6920220 (A)</i>	USA (>90% of Northern European ancestry)	441	619	15%	1.30	(1.05 - 1.61)	
Prahalaad 2009	UK	873	3644	42%	1.16	(1.02 - 1.31)	
Hinks 2010	USA (overlap not reported)	809 ^a	531 ^a	19%	0.94	(0.78 - 1.13)	
Thompson 2010	USA	<i>overlap with Thompson 2010</i>					
Thompson 2012	Australia	200	341	7%	1.04	(0.77 - 1.41)	
Ellis 2013	<i>present study</i>			17%	0.98	(0.8 - 1.19)	
Reinards et al. pooled				1.09			0.03309
<i>TNPO3 rs10488631 (C)</i>	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
Hinks 2012	UK	1179	5176	78%	1.20	(1.05 - 1.37)	
Reinards et al. pooled	<i>present study</i>			22%	1.03	(0.8 - 1.33)	
				1.16			0.01277
<i>IL2RA rs2104286 (G)</i>	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
Hinks 2009	UK	593	>3000a	20%	0.76	(0.66 - 0.88)	
Prahalaad 2009	USA (>90% of Northern European ancestry)	438	634	11%	1.04	(0.85 - 1.27)	
Thompson 2010 initial cohort	USA, UK (overlap not reported)	809a	3521a	25%	0.76	(0.66 - 0.86)	
Thompson 2010 replication cohort	USA, Germany, Czech Republic (overlap not reported)	1015a	1569a	26%	0.96	(0.84 - 1.1)	
Thompson 2012	USA	<i>overlap with Thompson 2010</i>					
Ellis 2013	Australia	200	341	5%	0.87	(0.64 - 1.18)	
Reinards et al. pooled	<i>present study</i>			13%	0.91	(0.76 - 1.1)	
					0.86		8.21E-06

Supplementary Table S4 (continued)
Meta-analyses of reported JIA loci^a

SNP (minor allele)	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
<i>IL2RA</i> rs41295061 (A)	UK	619	3614	58%	0.80	(0.63 - 1)	
Hinks 2009	present study			4.2%	1.02	(0.79 - 1.32)	
Reinards et al. pooled				0.89			0.1646
<i>CLEC16A</i> rs12708716 (G)	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
Skinningsrud 2010	Norway	507	2109	36%	0.89	(0.77 - 1.03)	
Thompson 2010	USA	809 ^a	531 ^a	31%	0.98	(0.84 - 1.16)	
Thompson 2012	USA						
Reinards et al. pooled	present study			33%	0.92	(0.78 - 1.07)	
				0.93			0.09463
<i>CLEC16A</i> rs6498169 (G)	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
Skinningsrud 2010	Norway	498	2110	28%	1.19	(1.03 - 1.37)	
Thompson 2012	USA	814	3056	41%	1.11	(0.99 - 1.25)	
Ellis 2013	Australia	200	341	8%	1.03	(0.79 - 1.33)	
Reinards et al. pooled	present study			23%	1.09	(0.93 - 1.28)	
				1.12			0.002768
<i>MIF</i> rs755622 (-173) (C)	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
Donn 2002	UK	526	259	15%	1.86	(1.36 - 2.55)	
Miterski 2004	Germany	150	390	14%	1.21	(0.88 - 1.66)	
Kaalla 2013	USA	638	742	39%	1.06	(0.88 - 1.29)	
Reinards et al. pooled	present study			32%	0.67	(0.54 - 0.83)	
				1.01			0.8222

Supplementary Table S4 (continued)
Meta-analyses of reported JIA loci*

SNP (minor allele)			nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
<i>IL2RB rs743777 (G)</i>	origin							
Thompson 2012	USA		812	3051	67%	1.02	(0.91 - 1.14)	
Reinards et al. pooled	present study				33%	0.95	(0.81 - 1.12)	
<i>IL1B rs1143634 (3954) (A)</i>	origin							
Cimaz 2007	France (cases+controls), Italy (cases)		107	630				
Reinards et al. <i>no meta-analysis performed</i>	present study							
<i>TNFA rs1800750 (-376) (A)</i>	origin							
Zeggini 2002 II	UK							
Reinards et al. <i>no meta-analysis performed</i>	present study							

OR: odds ratio; CI: confidence interval

Patient group of the present study limited to oligoarticular (persistent and extended) and RF negative JIA patients. References are listed in Supplementary Table S6.s

- a) Exact number of successfully typed individuals not known
- b) Minor allele unclear

Supplementary Table S5
Meta-analyses of loci previously tested in JIA (but not significantly associated*)

SNP (minor allele)	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
CD28 rs1980422 (C) ^b	origin						
Thompson 2012	USA	814	3058	69%	1.14	(1 - 1.29)	
Reinards et al.	present study			31%	1.29	(1.07 - 1.55)	
pooled				1.18			0.001411
CD226 rs763361 (T) ^b	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
Hinks 2010 II	UK	600	3494	55%	1.07	(0.94 - 1.21)	
Ellis 2013	Australia	158	341	11%	1.22	(0.93 - 1.59)	
Reinards et al.	present study			36%	1.30	(1.12-1.51)	
pooled				1.16			0.001115
TNFRSF14-MME1 rs3890745 (G)	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
Thompson 2010	USA, UK	809 ^a	7521 ^a	65%	0.92	(0.81 - 1.04)	
Thompson 2012	USA		overlap with Thompson 2010				
Reinards et al.	present study			37%	0.90	(0.77 - 1.06)	
pooled				0.91			0.07282
TNFRSF1B rs1061622 (G)	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
Zeggini 2002	UK	435	261	35%	1.25	(0.97 - 1.61)	
Reinards et al.	present study			65%	1.03	(0.85 - 1.24)	
pooled				1.10			0.2057
LCR rs695161 (C)	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
Thompson 2012	USA	813	3057	64%	0.98	(0.88 - 1.1)	
Reinards et al.	present study			36%	0.97	(0.83 - 1.15)	
pooled				0.98			0.5911

Supplementary Table S5 (continued)
Meta-analyses of loci previously tested in JIA (but not significantly associated**)

SNP (minor allele)	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
<i>IL10</i> rs1800896 (-1082) (C)	UK	435	274	28%	1.07	(0.86 - 1.32)	
Donn 2001	UK (overlap not reported)	348	239	24%	0.83	(0.66 - 1.05)	
Reinards et al.	present study			48%	0.90	(0.77 - 1.06)	
pooled				0.93		0.1908	
<i>IL18RAP</i> rs917997 (T)	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
Hinks 2010 JV	UK	987	2931	7.1%	1.00	(0.89 - 1.13)	
Reinards et al.	present study			29%	1.14	(0.94 - 1.37)	
pooled				1.04		0.4693	
<i>IL1A</i> rs1800587 (-889) (A)	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
McDowell 1995	Norway	269	99	6%	1.34	(0.94 - 1.9)	
Donn 2001	UK	330	236	11%	0.75	(0.57 - 0.97)	
Cinek 2004	Czech Republic	130	102	4%	1.19	(0.78 - 1.81)	
Thompson 2012	USA	814	2058	55%	1.04	(0.93 - 1.18)	
Reinards et al.	present study			23%	0.89	(0.74 - 1.07)	
pooled				0.99		0.825	
<i>IL1B</i> rs16944 (-511) (A)	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
Cinek 2004	Czech Republic	130	103	14%	1.29	(0.86 - 1.95)	
Reinards et al.	present study			86%	1.11	(0.94 - 1.31)	
pooled				1.13		0.116	

Supplementary Table S5 (continued)

Meta-analyses of loci previously tested in JIA (but not significantly associated*)

SNP (minor allele)	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
<i>STAT1 rs7562024 (T)</i>	USA	814	3058	67%	1.00	(0.9 - 1.12)	
Reinards et al.	<i>present study</i>			35%	0.95	(0.82 - 1.11)	
pooled				0.98			0.7298
<i>FAM107A rs13315591 (C)</i>	USA	814	3056	64%	0.92	(0.75 - 1.14)	
Thompson 2012	<i>present study</i>			35%	1.13	(0.86 - 1.5)	
Reinards et al.				0.99			0.9204
pooled							
<i>4p15 rs874040 (G)</i>	USA	811	3057	67%	1.03	(0.92 - 1.16)	
Thompson 2012	<i>present study</i>			33%	0.93	(0.79 - 1.09)	
Reinards et al.				0.99			0.9008
pooled							
<i>ERAP1 rs30187 (T)</i>	UK	1054 ^a	5200 ^a	71%	1.02	(0.92 - 1.15)	
Hinks 2011	Australia	200	741	8%	1.06	(0.82 - 1.37)	
Ellis 2013	<i>present study</i>			21%	1.14	(0.97 - 1.34)	
Reinards et al.				1.05			0.2089
pooled							
<i>LECT2 rs31517 (T)</i>	USA	814	3057	65%	0.92	(0.82 - 1.03)	
Thompson 2012	<i>present study</i>			35%	1.03	(0.88 - 1.2)	
Reinards et al.				0.96			0.3354
pooled							

Supplementary Table S5 (continued)
Meta-analyses of loci previously tested in JA (but not significantly associated^a)

SNP (minor allele)	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
<i>PRDM1 rs548234 (C)</i>							
Hinks 2012	UK	1170	8001	43%	0.96	(0.88 - 1.06)	
Thompson 2012	USA	813	3056	37%	1.03	(0.92 - 1.16)	
Reinards et al.	present study			20%	1.13	(0.96 - 1.33)	
pooled				1.02		0.6309	
<i>TAGAP rs394581 (C)</i>	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
Hinks 2012	UK	1175	2617	70%	1.00	(0.9 - 1.12)	
Reinards et al.	present study			30%	0.97	(0.82 - 1.15)	
pooled				0.99		0.8646	
<i>CCR6 rs3093023 (A)</i>	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
Hinks 2012	UK	1002	8128	49%	0.96	(0.87 - 1.05)	
Thompson 2012	USA	806	3047	35%	1.08	(0.97 - 1.21)	
Reinards et al.	present study			16%	1.07	(0.91 - 1.26)	
pooled				1.02		0.5788	
<i>CDK6 rs42041 (G)</i>	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
Hinks 2010	UK	926	2962	53%	1.03	(0.92 - 1.16)	
Thompson 2010	USA, UK (overlap not reported)	809 ^a	3521 ^a	27%	0.99	(0.87 - 1.13)	
Thompson 2012	USA	overlap with Thompson 2010		20%	0.95	(0.8 - 1.13)	
Reinards et al.	present study				1.00		0.9404
pooled							

Supplementary Table S5 (continued)

Meta-analyses of loci previously tested in JIA (but not significantly associated*)

SNP (minor allele)	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
<i>CCL21rs951005 (C)</i>	UK	999	7947	72%	0.91	(0.8 - 1.04)	
Hinks 2012				28%	1.08	(0.88 - 1.34)	
Reinards et al.							
pooled					0.96		0.43
<i>PRKQrs4750316 (C)</i>	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
Hinks 2010	UK	943	7500	42%	0.88	(0.77 - 1)	
Thompson 2012	USA	814	3058	33%	0.87	(0.75 - 1)	
Ellis 2013	Australia	200	341	6%	0.88	(0.63 - 1.23)	
Reinards et al.				19%	0.99	(0.82 - 1.2)	
pooled					0.90		0.01012
<i>TRAF6rs540386 (T)</i>	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
Thompson 2012	USA	814	3051	69%	1.10	(0.94 - 1.28)	
Reinards et al.				31%	0.83	(0.66 - 1.05)	
pooled					1.01		0.8841
<i>TNFRSF1A rs767455(C)</i>	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
Hinks 2013	UK	987	5194	74%	0.98	(0.89 - 1.08)	
Reinards et al.				26%	0.92	(0.79 - 1.09)	
pooled					0.97		0.4013
<i>TNFRSF1A rs4149570 (A)</i>	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p meta-analysis
Thompson 2012	USA	812	3055	37%	1.02	(0.91 - 1.14)	
Hinks 2013	UK	929	5191	46%	0.94	(0.85 - 1.04)	
Reinards et al.				18%	1.06	(0.9 - 1.25)	
pooled					0.99		0.7597

Supplementary Table S5 (continued)Meta-analyses of loci previously tested in JIA (but not significantly associated^{a)*})

SNP (minor allele)	origin	nr of cases	nr of controls	weight	OR	(95% CI)	p_{meta-analysis}
<i>KIF5A</i> rs1678542 (C)							
Hinks 2010	UK	941	3530	42%	0.91	(0.82 - 1.01)	
Thompson 2010	USA, UK	809 ^{b)}	5521 ^{a)}	33%	1.00	(0.89 - 1.13)	
Thompson 2012	USA	overlap with Thompson 2010					
Ellis 2013	Australia	200	341	7%	0.96	(0.74 - 1.24)	
Reinards et al.	present study			19%	1.02	(0.87 - 1.19)	
pooled				0.96			0.2652
<i>IL21R</i> rs3093341 (G)							
Thompson 2012	USA	814	3057	66%	0.99	(0.82 - 1.2)	
Reinards et al.	present study			34%	0.80	(0.62 - 1.05)	
pooled				0.92			0.3058
<i>STAT5A</i> rs7217728 (C)							
Thompson 2012	USA	812	3058	65%	0.96	(0.85 - 1.08)	
Reinards et al.	present study			35%	0.91	(0.77 - 1.07)	
pooled				0.94			0.2319

OR: odds ratio; CI: confidence interval

* Patient group of the present study limited to oligoarticular (persistent and extended) and RF-negative JIA patients References are listed in Supplementary Table S6

- a) Exact number of successfully typed individuals not known
- b) Patient group of all included studies limited to oligoarticular (persistent and extended) and RF-negative JIA patients

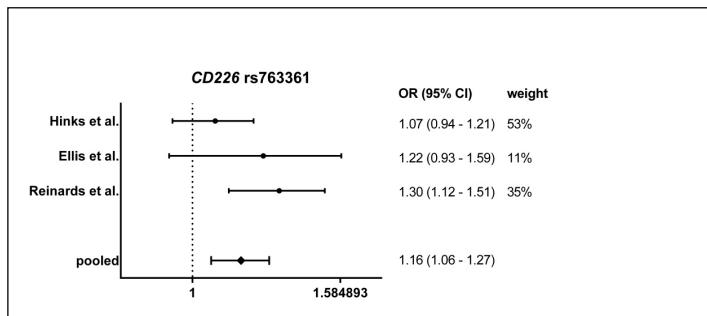
SupplementaryTable S6. References of studies analysed for meta-analysis (supplementary Tables S4 and S5)

Cimaz 2007 ¹	Cimaz R, Cazalis MA, Reynaud C et al. IL1 and TNF gene polymorphisms in patients with juvenile idiopathic arthritis treated with TNF inhibitors. <i>Ann Rheum Dis</i> 2007; 66(7):900-904.
Cinek 2004 ²	Cinek O, Vavrinova P, Striz I et al. Association of single nucleotide polymorphisms within cytokine genes with juvenile idiopathic arthritis in the Czech population. <i>J Rheumatol</i> 2004; 31(6):1206-1210.
Cinek 2006 ³	Cinek O, Hradsky O, Ahmedov G et al. No independent role of the -1123 G>C and +2740 A>G variants in the association of PTPN22 with type 1 diabetes and juvenile idiopathic arthritis in two Caucasian populations. <i>Diabetes Res Clin Pract</i> 2007; 76(2):297-303.
Crawley 1999 ⁴	Crawley E, Kay R, Sillibourne J et al. Polymorphic haplotypes of the interleukin-10 5' flanking region determine variable interleukin-10 transcription and are associated with particular phenotypes of juvenile rheumatoid arthritis. <i>Arthritis Rheum</i> 1999; 42(6):1101-1108.
Dimopoulou 2013 ⁵	Dimopoulou DG, Zervou MI, Trachana M et al. Investigation of juvenile idiopathic arthritis susceptibility loci: results from a Greek population. <i>Hum Immunol</i> 2013; 74(9):1194-1198.
Donn 2001 ⁶	Donn RP, Barrett JH, Farhan A et al. Cytokine gene polymorphisms and susceptibility to juvenile idiopathic arthritis. British Paediatric Rheumatology Study Group. <i>Arthritis Rheum</i> 2001; 44(4):802-810.
Donn 2002 ⁷	Donn R, Alourfi Z, De BF et al. Mutation screening of the macrophage migration inhibitory factor gene: positive association of a functional polymorphism of macrophage migration inhibitory factor with juvenile idiopathic arthritis. <i>Arthritis Rheum</i> 2002; 46(9):2402-2409.
Ellis 2013 ⁸	Ellis JA, Chavez RA, Pezic A et al. Independent replication analysis of genetic loci with previous evidence of association with juvenile idiopathic arthritis. <i>Pediatr Rheumatol Online J</i> 2013; 11(1):12.
Hinks 2005 ⁹	Hinks A, Barton A, John S et al. Association between the PTPN22 gene and rheumatoid arthritis and juvenile idiopathic arthritis in a UK population: further support that PTPN22 is an autoimmunity gene. <i>Arthritis Rheum</i> 2005; 52(6):1694-1699.
Hinks 2009 ¹⁰	Hinks A, Ke X, Barton A et al. Association of the IL2RA/CD25 gene with juvenile idiopathic arthritis. <i>Arthritis Rheum</i> 2009; 60(1):251-257.
Hinks 2009 II ¹¹	Hinks A, Barton A, Shephard N et al. Identification of a novel susceptibility locus for juvenile idiopathic arthritis by genome-wide association analysis. <i>Arthritis Rheum</i> 2009; 60(1):258-263.
Hinks 2010 ¹²	Hinks A, Eyre S, Ke X et al. Overlap of disease susceptibility loci for rheumatoid arthritis and juvenile idiopathic arthritis. <i>Ann Rheum Dis</i> 2010; 69(6):1049-1053.
Hinks 2010 II ¹³	Hinks A, Eyre S, Ke X et al. Association of the AFF3 gene and IL2/IL21 gene region with juvenile idiopathic arthritis. <i>Genes Immun</i> 2010; 11(2):194-198.
Hinks 2010 III ¹⁴	Hinks A, Martin P, Flynn E et al. Association of the CCR5 gene with juvenile idiopathic arthritis. <i>Genes Immun</i> 2010; 11(7):584-589.
Hinks 2010 IV ¹⁵	Hinks A, Martin P, Flynn E et al. Investigation of type 1 diabetes and coeliac disease susceptibility loci for association with juvenile idiopathic arthritis. <i>Ann Rheum Dis</i> 2010; 69(12):2169-2172.
Hinks 2011 ¹⁶	Hinks A, Martin P, Flynn E et al. Subtype specific genetic associations for juvenile idiopathic arthritis: ERAP1 with the enthesitis related arthritis subtype and IL23R with juvenile psoriatic arthritis. <i>Arthritis Res Ther</i> 2011; 13(1):R12.
Hinks 2012 ¹⁷	Hinks A, Cobb J, Sudman M et al. Investigation of rheumatoid arthritis susceptibility loci in juvenile idiopathic arthritis confirms high degree of overlap. <i>Ann Rheum Dis</i> 2012; 71(7):1117-1121.
Hinks 2013 ¹⁸	Hinks A, Martin P, Thompson SD et al. Autoinflammatory gene polymorphisms and susceptibility to UK juvenile idiopathic arthritis. <i>Pediatr Rheumatol Online J</i> 2013; 11(1):14.

Kaalla 2013 ¹⁹	Kaalla MJ, Broadaway KA, Rohani-Pichavant M et al. Meta-analysis confirms association between TNFA-G238A variant and JIA, and between PTPN22-C1858T variant and oligoarticular, RF-polyarticular and RF-positive polyarticular JIA. <i>Pediatr Rheumatol Online J</i> 2013; 11(1):40.
Lindner 2007 ²⁰	Lindner E, Nordang GB, Melum E et al. Lack of association between the chemokine receptor 5 polymorphism CCR5delta32 in rheumatoid arthritis and juvenile idiopathic arthritis. <i>BMC Med Genet</i> 2007; 8:33.
McDowell 1995 ²¹	McDowell TL, Symons JA, Ploski R et al. A genetic association between juvenile rheumatoid arthritis and a novel interleukin-1 alpha polymorphism. <i>Arthritis Rheum</i> 1995; 38(2):221-228.
Miterski 2004 ²²	Miterski B, Drynda S, Boschow G et al. Complex genetic predisposition in adult and juvenile rheumatoid arthritis. <i>BMC Genet</i> 2004; 5:2.
Mourão 2009 ²³	Mourao AF, Caetano-Lopes J, Costa P et al. Tumor necrosis factor-alpha -308 genotypes influence inflammatory activity and TNF-alpha serum concentrations in children with juvenile idiopathic arthritis. <i>J Rheumatol</i> 2009; 36(4):837-842.
Ozen 2002 ²⁴	Ozen S, Alikasifoglu M, Bakkaloglu A et al. Tumour necrosis factor alpha G-->A -238 and G-->A -308 polymorphisms in juvenile idiopathic arthritis. <i>Rheumatology (Oxford)</i> 2002; 41(2):223-227.
Pazár 2008 ²⁵	Pazar B, Gergely P Jr, Nagy ZB et al. Role of HLA-DRB1 and PTPN22 genes in susceptibility to juvenile idiopathic arthritis in Hungarian patients. <i>Clin Exp Rheumatol</i> 2008; 26(6):1146-1152.
Prahalaad 2008 ²⁶	Prahalaad S, Bohnsack JF, Whiting A et al. Lack of association of functional CTLA4 polymorphisms with juvenile idiopathic arthritis. <i>Arthritis Rheum</i> 2008; 58(7):2147-2152.
Prahalaad 2009 ²⁷	Prahalaad S, Hansen S, Whiting A et al. Variants in TNFAIP3, STAT4, and C12orf30 loci associated with multiple autoimmune diseases are also associated with juvenile idiopathic arthritis. <i>Arthritis Rheum</i> 2009; 60(7):2124-2130.
Schmeling 2006 ²⁸	Schmeling H, Wagner U, Peterson A et al. Tumor necrosis factor alpha promoter polymorphisms in patients with juvenile idiopathic arthritis. <i>Clin Exp Rheumatol</i> 2006; 24(1):103-108.
Seldin 2005 ²⁹	Seldin MF, Shigeta R, Laiho K et al. Finnish case-control and family studies support PTPN22 R620W polymorphism as a risk factor in rheumatoid arthritis, but suggest only minimal or no effect in juvenile idiopathic arthritis. <i>Genes Immun</i> 2005; 6(8):720-722.
Skinningsrud 2010 ³⁰	Skinningsrud B, Lie BA, Husebye ES et al. A CLEC16A variant confers risk for juvenile idiopathic arthritis and anti-cyclic citrullinated peptide antibody negative rheumatoid arthritis. <i>Ann Rheum Dis</i> 2010; 69(8):1471-1474.
Suppiah 2005 ³¹	Suppiah V, O'doherty C, Heggarty S et al. The CTLA4+49A/G and CT60 polymorphisms and chronic inflammatory arthropathies in Northern Ireland. <i>Exp Mol Pathol</i> 2006; 80(2):141-146.
Thompson 2010 ³²	Thompson SD, Sudman M, Ramos PS et al. The susceptibility loci juvenile idiopathic arthritis shares with other autoimmune diseases extend to PTPN2, COG6, and ANGPT1. <i>Arthritis Rheum</i> 2010; 62(11):3265-3276.
Thompson 2012 ³³	Thompson SD, Marion MC, Sudman M et al. Genome-wide association analysis of juvenile idiopathic arthritis identifies a new susceptibility locus at chromosomal region 3q13. <i>Arthritis Rheum</i> 2012; 64(8):2781-2791.
Viken 2005 ³⁴	Viken MK, Amundsen SS, Kvien TK et al. Association analysis of the 1858C>T polymorphism in the PTPN22 gene in juvenile idiopathic arthritis and other autoimmune diseases. <i>Genes Immun</i> 2005; 6(3):271-273.
Zeggerini 2002 ³⁵	Zeggerini E, Thomson W, Alansari A et al. Tumour necrosis factor receptor II polymorphism and juvenile idiopathic arthritis. <i>Rheumatology (Oxford)</i> 2002; 41(4):462-465.
Zeggerini 2002 II ³⁶	Zeggerini E, Thomson W, Kwiatkowski D et al. Linkage and association studies of single-nucleotide polymorphism-tagged tumor necrosis factor haplotypes in juvenile oligoarthritis. <i>Arthritis Rheum</i> 2002; 46(12):3304-3311.

Supplementary Figure S1 Meta-analysis of novel JIA susceptibility loci

a. Meta-analysis of *CD226 rs763361*, restricted to oligoarticular and RF negative JIA patients of Caucasian origin



b. Meta-analysis of *CD28 rs1980422*, restricted to oligoarticular and RF negative JIA patients of Caucasian origin

