

 $\label{lem:hypopituitarism:clinical assessment in different conditions} \\ \text{Kokshoorn, N.E.}$

Citation

Kokshoorn, N. E. (2011, December 7). *Hypopituitarism : clinical assessment in different conditions*. Retrieved from https://hdl.handle.net/1887/18194

Version: Corrected Publisher's Version

License: License agreement concerning inclusion of doctoral thesis in the

Institutional Repository of the University of Leiden

Downloaded from: https://hdl.handle.net/1887/18194

Note: To cite this publication please use the final published version (if applicable).

Chapter 1

General introduction and outline of this thesis

Contents

I.	General introduction		11
II.	The pituitary gland		12
	Anatomy and Physiology		12
	1.	Regulation and secretion of growth hormone and IGF-I	13
	2.	Regulation of ACTH and cortisol secretion	16
	3.	Regulation and secretion of thyroid hormone,	
		gonadotropins and prolactin	17
III.	II. Pituitary insufficiency		
	Growth hormone deficiency		19
	Corticotropin deficiency		21
	Thyrotropin deficiency		23
	Gonadotropin deficiency		23
	Prola	ctin deficiency	24
IV.	Outli	ine of this thesis	25
	The evaluation of pituitary function in patients after traumation brain injury Dynamic tests of pituitary function in other pituitary disease Treatment of GH deficiency		
			25
			26
			27

I. General introduction

The pituitary gland is the master regulator of the endocrine system. Different pathophysiological conditions can affect the function of the pituitary gland and, consequently, endocrine homeostasis. The evaluation of pituitary function is therefore complex and the different tools that have become available to evaluate pituitary function only provide limited information of different aspects of hormone secretion. In this thesis, several difficulties encountered in establishing a diagnosis of pituitary insufficiency are studied in different pathophysiological conditions.

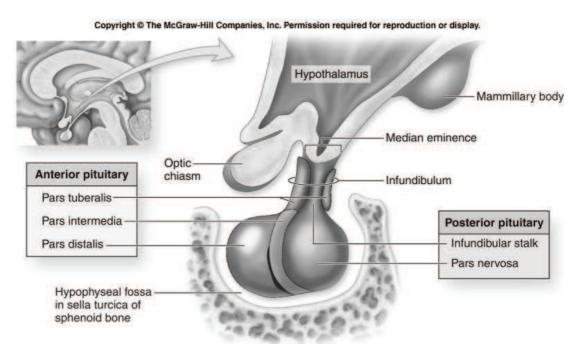


Figure 1. The pituitary gland

II. The pituitary gland

Anatomy and Physiology

The pituitary gland is a small gland located at the base of the skull in a socket of sphenoid bone, called the sella turcica. The gland consists of two lobes, the anterior lobe (or adenohypophysis; 80%), and a posterior lobe (neurohypophysis; 20%) (Figure 1).

Together with the hypothalamus, the pituitary controls the function of different endocrine glands (*i.e.* thyroid, adrenal and reproductive glands) (1). The hypothalamus receives signals from upper corticol inputs and the environment (such as light and temperature) and, in turn, delivers signals to the pituitary gland (*i.e.* regulating the endocrine system). Hormones released by the pituitary gland influence the endocrine systems in the body and also have a feedback on the hypothalamus.

The communication between the hypothalamus and anterior pituitary is via the portal system that runs through the pituitary stalk. Hormones released by the hypothalamus are delivered to the anterior pituitary through these vessels and reach the anterior lobe through a dense capillary network. The communication with the posterior gland is via axons and nerve terminals of larger neurons that originate from within the hypothalamus. The hormones produced in these neurons, arginine-vasopressin and oxytocin, are released directly from the posterior pituitary into the systemic circulation.

The anterior pituitary is controlled by specific hypothalamic hormones: thyrotropin releasing hormone (TRH), gonadotropin releasing hormone (GnRH), corticotropin releasing hormone (CRH), growth hormone releasing hormone (GHRH), and somatostatin, that bind specific transmembrane receptors expressed in different anterior pituitary cells. These anterior pituitary cells are classified by their specific secretory products: somatotrophs (GH-secreting cells, expressing the GHRH and somatostatin receptor; 50%), lactotrophs (PRL-secreting cells, expressing the prolactin receptor; 10–25%), corticotrophs (cells secreting ACTH,

expressing the CRH receptor; 15–20%), thyrotrophs (cells secreting TSH, expressing the TRH receptor; 10%), and gonadotrophs (LH and FSH secreting cells, expressing the GnRH receptor; 10–15%) (2).

1. Regulation and secretion of growth hormone and IGF-I

The regulation of growth hormone (GH) secretion is complex and involves many stimulatory and inhibitory hypothalamic peptides. However, the two most important components are growth hormone-releasing hormone (GHRH), which stimulates the somatotrophic cells, and somatostatin (SST) which inhibits GH release (2). The secretion of GH is also affected by factors such as nutrition (increased in fasting, stimulated by high protein meals and inhibit by hyperglycemia and leptin), other hormones (stimulated by estrogens and inhibited by glucocorticoid excess), neuropeptides, neurotransmitters and opiates (2–4).

The secretion of GH is pulsatile with undetectable serum GH levels between the pulses. In normal subjects the 24-hour profile of plasma GH levels consists of stable low levels interrupted by bursts of secretion (Figure 2 and 3). The major determinant of GH secretion in humans is sleep. GH secretion is lower in elderly and obese subjects and there are sex-specific differences in GH pulse amplitude and mass (5;6). The age-associated changes in the GH profile include a reduction in GH secretory burst frequency, the half life of endogenous GH and the daily secretory rate (7). In obese subjects decreased GH concentrations result from both diminished pulsatile GH secretion and accelerated metabolic clearance (4;8).

In the liver, GH stimulates the production of insulin-like growth factor (IGF-I). The primary function of GH is promotion of linear growth in children by acting directly and indirectly (via the synthesis of IGF-I which mediates most of the peripheral actions of GH) on the epiphyseal plates of long bones. Whereas GH and IGF-I have synergistic effects on linear and organ growth by their control of mitogenesis and apoptosis and on glomerular filtration rate, their metabolic actions are opposing: GH stimulates lipolysis and reduces insulin sensitivity, IGF-I is anti-lipolytic and ameliorates insulin sensitivity (9;10).

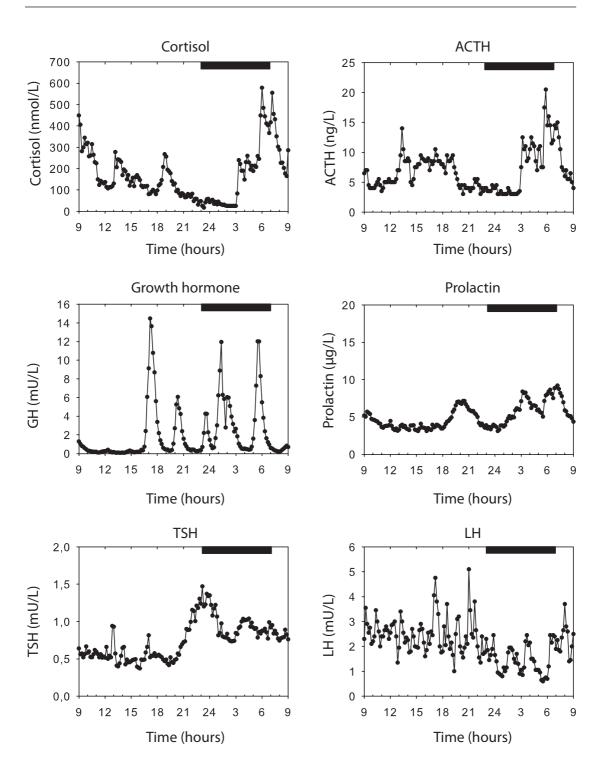


Figure 2. Plasma hormone concentration profiles of a 33 year-old healthy female volunteer. Blood samples were taken at 10 min intervals during 24 hours. The black bar in the top of the panels indicate the period with lights off. Note the diurnal and the pulsatile characteristics of each hormone. (*The figure was provided by Dr. F. Roelfsema, Leiden University.*)

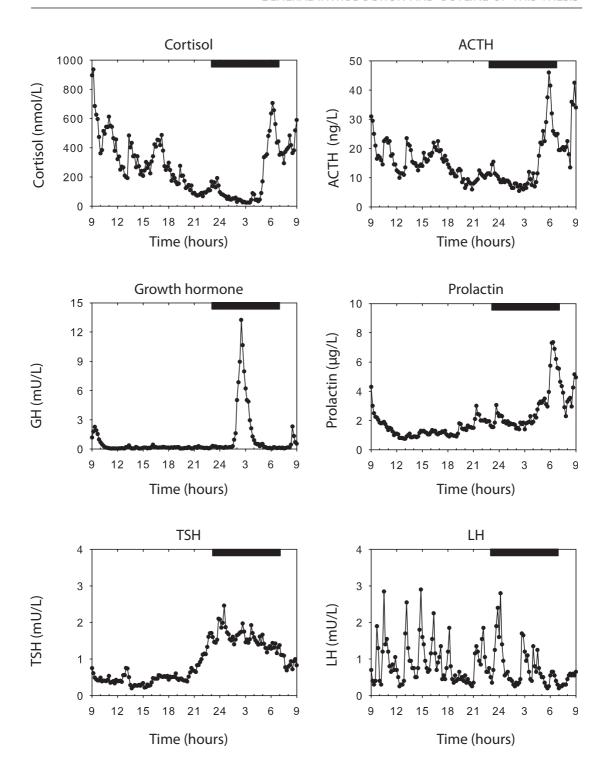


Figure 3. Plasma hormone concentration profiles of a 37 year-old healthy male volunteer. Blood samples were taken at 10 min intervals during 24 hours. The black bar in the top of the panels indicate the period with lights off. Note the diurnal and the pulsatile characteristics of each hormone. (*The figure was provided by Dr. F. Roelfsema, Leiden University.*)

2. Regulation of ACTH and cortisol secretion

The secretion of corticotropin (ACTH) and cortisol is regulated by hormonal interactions between the hypothalamus, pituitary, and adrenal glands. The secretion of hypothalamic corticotropin-releasing hormone (CRH) is regulated mainly by hippocampal neurons that express both receptors for cortisol, the mineralocorticoid- and glucocorticoid receptor. In addition the secretion is influenced by the circadian pacemaker and stress (2;11). CRH regulates the secretion of ACTH by the pituitary gland, which is potentiated by arginine-vasopressin. Subsequently ACTH binds to its receptor on the adrenal cortex to stimulate the secretion of cortisol and other steroids. The negative feedback loop is completed by the inhibitory effect of cortisol on CRH and ACTH synthesis and secretion (11).

Pulsatile secretion and circadian rhythm

ACTH is secreted in brief episodic bursts resulting in a diurnal rhythm of ACTH secretion with a concordant diurnal secretion of cortisol from the adrenal cortex (12;13). Plasma ACTH and serum cortisol concentrations are highest early in the morning at time of awakening. During the day plasma cortisol levels fall resulting in low levels in the late afternoon and evening with a nadir one or two hours after sleep onset (Figure 2 and 3) (11;14;15).

Stress-induced secretion

The HPA axis is activated both by physical and psychological stressors, resulting in increased plasma ACTH and cortisol concentrations. Physical stressors include severe trauma, like burns (16;17), or illnesses, major surgery (18;19), but also hypoglycemia (20;21), hypotension, exercise (22), and cold exposure (23).

Negative feedback inhibition by glucocorticoids

Both endogenous and exogenous glucocorticoids have a negative feedback on ACTH secretion which occurs at both the hypothalamic (CRH suppression) and pituitary (ACTH suppression) levels. This leads to atrophy of the adrenal glands resulting in loss of cortisol secretory capacity (24). The degree of probably depends upon the dose, potency and duration of action of the glucocorticoid, and the time of its administration (25–29). The shorter the interval between the

administration of glucocorticoid and the normal early morning peak of ACTH secretion, the greater the suppressive effect of the glucocorticoid. The duration of suppression is increased by higher doses and longeracting glucocorticoids. After withdrawal of chronic administration of high doses of glucocorticoid, suppression of the hypothalamic-pituitary-adrenal axis may persist for weeks but may even persist for many years.

3. Regulation and secretion of thyroid hormone, gonadotropins and prolactin

The hypothalamus-pituitary thyroidal axis regulates the production of thyroid hormone by the thyroid gland. The hypothalamus produces thyroptropin releasing hormone (TRH) which stimulates the pituitary gland to secrete thyrotropin (TSH). TSH stimulates the synthesis of the thyroid hormones (thyroxine (T_4) and triiodothyronine (T_3)) by binding to the TSH receptors on the thyroid cells. The response of TSH to TRH is, in turn, modulated by the circulating concentrations of T_3 and T_4 . High serum levels of T_3 and T_4 inhibit and low levels stimulate TSH synthesis (2).

The reproductive axis is controlled by periodic pulsatile release of the hypothalamic gonadotropin-releasing hormone (GnRH). GnRH stimulates the pituitary to secrete the gonadotropins, luteinizing hormone (LH) and follicle-stimulating hormone (FSH). The production of steroid hormones (including estradiol, progesterone and testosterone), as well as other factors such as inhibin, activin and insulin-like growth factor-I (IGF-I) are induced by the gonadotropins. The circulating sex steroids have a positive as well as negative feedback on GnRH and thereby also influence LH and FSH concentrations.

The function of LH in men is to stimulate testosterone production from interstitial cells of the testes (Leydig cells) and FSH is required for spermatogenesis. In women, LH is critical for ovulation and maintenance of the corpus luteum, whereas FSH promotes follicular development (2).

The main physiological role of prolactin (PRL) is for nursing. The hypothalamic control of PRL secretion is predominantly inhibitory, and dopamine is the most important inhibitory factor. TRH is a potent prolactin-releasing factor (2).

III. Pituitary insufficiency

Hypopituitarism refers to decreased secretion of pituitary hormones, which can result from diseases of the pituitary gland and/or the hypothalamus, which cause diminished secretion of hypothalamic releasing hormones, thereby reducing secretion of the corresponding pituitary hormones. Pituitary insufficiency can be congenital (which will not be further addressed in this thesis) or acquired.

Pituitary adenomas and its treatment

A common cause of pituitary dysfunction is the presence of a pituitary adenoma. Macro-adenomas (> 10 mm) can be associated with pituitary insufficiency, with one or more anterior pituitary hormone deficiencies (30–32). In the presence of macroadenomas, hypopituitarism may result from compression of the rest of the pituitary and/or compression of the portal vessels in the pituitary stalk, secondary to either the expanding tumor mass or directly by increased intra-sellar pressure (30). Conversely, reduction of tumor mass by surgery and/or medication relieves the pressure and may restore pituitary function.

In pituitary surgery the surgeon attempts to preserve the adjacent normal pituitary tissue. However, if the surgeon is not able to visually distinguish the normal pituitary tissue from the adenoma, the normal tissue may be damaged, resulting in pituitary deficiency (33;34).

Radiation of pituitary adenomas, usually to prevent regrowth of residual tissue after surgery or to control excessive GH or ACTH secretion, also exposes the nonadenomatous pituitary and the hypothalamus to irradiation resulting in pituitary insufficiency (35;36). Not only patients with pituitary tumors but also patients treated with radiotherapy for suprasellar lesions, primary brain tumors, nasopharyngeal tumors, head and neck tumors, or hematological malignancies (*i.e.* acute lymphoblastic leukemia (ALL)) are at risk for developing pituitary hormone deficiencies if the hypothalamus and/or the pituitary have been exposed to radiation (37–39).

Traumatic brain injury

In recent years, several studies have reported a high prevalence of pituitary insufficiency ranging from 15–90% in patients who experienced traumatic brain injury (TBI) (40–51). Large neuropathological series demonstrate pituitary as well as hypothalamic lesions after TBI (52). Infarction is believed to be the cause of posttraumatic hypopituitarism, found at post mortem in 26% to 86% of patients who died after TBI. Possible mechanisms for post-traumatic infarction include compression of the pituitary gland caused by changes in intracranial pressure resulting from cerebral edema, hemorrhage or skull fracture, hypoxia, or direct damage to the gland itself (53;54).

The diagnosis of hypopituitarism, defined as deficient secretion of one or more pituitary hormones secondary to pituitary or hypothalamic disease, is made by documenting subnormal secretion of these pituitary hormones under defined (*i.e.* controlled) circumstances. Since there is a variable pattern of hormone deficiencies among patients with hypopituitarism, each pituitary hormone must be tested separately. For the evaluation of each axis basal serum hormones levels, but also dynamic testing is available.

Growth hormone deficiency

Clinical consequences

Growth hormone deficiency (GHD) in adults is characterized by increased body fat and decreased lean body mass, decreased bone mass and increased fracture rate, impaired cardiac function and reduced muscle strength (55–57). Adult patients with GHD also share a number of characteristics of the metabolic syndrome, including hypertension, abdominal obesity, insulin resistance and dyslipidemia (58). In addition, quality of life is impaired, with reduction in physical and mental energy, increased anxiety, and dissatisfaction with body image and poor memory (2;57;59;60). Replacement therapy with growth hormone (rhGH) was associated with apparent benefits, particularly in terms of body composition, bone mass, muscle strength, cardiac function and quality of life (61).

Diagnosis

Because of the pulsatile nature of GH secretion, basal serum GH levels are not useful to assess the GH-IGF-I axis, although basal serum IGF-I levels below the reference ranges are indicative for GHD in the presence of two or more other insufficiencies (62;63). Normal IGF-I concentrations, however, do not exclude the diagnosis of GHD, as IGF-I levels are within the normal reference range in about one third of patients with GHD, especially in elderly subjects (64–66). Therefore, the use of dynamic testing is mandatory for the evaluation of GH secretory reserve.

Different stimulation tests are available (i.e. insulin tolerance test (ITT), and stimulation tests with glucagon, GHRH, GHRH-arginine, or GHRH-GHRP6). However the preferred test for evaluation of this axis still remains the insulin tolerance test (63;67). With the administration of insulin a hypoglycemia is induced which is a very strong physiological stimulator of the stress response. Hypoglycemia activates the hypothalamus to secrete GHRH resulting in stimulation of GH by the somatotropic cells of the pituitary gland. A peak GH response below 3 µg/L indicates a severe GHD (63;67). During ITT simultaneous assessment of the hypothalamus-pituitary-adrenal (HPA) axis is possible. Important contra-indications to perform the ITT are coronary insufficiency and/or epilepsy. To assess the GH axis in these patients, alternative provocative tests for GH secretion must be used with adapted appropriate cut-offs. The combined administration of arginine and GHRH is the most frequently used alternative GH stimulation test (66;68;69). GHRH and arginine both have a stimulatory effect on the pituitary gland (70;71). When given simultaneously they enhance their effect resulting in a secretion of GH. A bolus dose of GHRH (1 µg/kg body weight) is given intravenously at baseline, immediately followed by an intravenous infusion of arginine (0.5 gr/kg body weight (to a maximum of 30 gr)) for 30 minutes. Measurements of GH are done 30, 45, 60 and 90 minutes after infusion. Recently, cut-off values adjusted for both body mass index (BMI) and age have been published (72).

Pitfalls

Several factors play a role when testing the GH secretion reserve, such as age, gender, BMI, other hormones and insulin sensitivity. Obese subjects have a blunted GH response to any provocative stimulus (8;73;74). There is an estrogen-related difference in GH axis activity: GH secreted per burst

greater and 24-hour GH release pattern is less orderly in women than men (75). Finally, GH secretion decreases with increasing age. Therefore, these factors should be considered when defining the diagnostic cut-off points in the assessment of GHD (72;76).

Corticotropin deficiency

Clinical consequences

ACTH deficiency leads to adrenocortical insufficiency, characterized by decreased secretion of cortisol. Normal corticotroph function is mandatory for adequate increase of cortisol concentrations in case of stress. However, to maintain sufficient cortisol concentrations, normal basal secretion of ACTH is necessary.

Hypocortisolism can be secondary to either adrenal gland destruction (primary adrenal insufficiency, mostly auto-immune adrenalitis or tuberculous adrenalitis) or to ACTH deficiency (secondary or central adrenal insufficiency) (77).

Diagnosis

Similar to the secretion of GH, the secretion of ACTH is pulsatile with circadian variation resulting in a circadian rhythm of cortisol secretion. Therefore, it is necessary to evaluate basal serum cortisol secretion in the early morning, during fasting. When cortisol concentrations are lower than, or exceed, a certain threshold (< 100 nmol/L or > 500 nmol/L) the likelihood of the presence or absence of adrenal insufficiency is very high, or negligible, respectively. In these cases stimulatory tests are not necessary (78). In all other condition, a dynamic test is mandatory.

The initial and most convenient test to evaluate the function of the HPA axis is the plasma cortisol response to synthetic ACTH (Synacthen test) (79;80). The test is performed by administering a bolus of 1 or 250 μ g of cosyntropin intramuscularly or intravenously with measurements of serum cortisol 30 and 60 minutes thereafter. A serum cortisol concentration of $\geq 500-550$ nmol/L is considered a normal response. However, this test does not discriminate between the different causes of adrenal insufficiency, and a normal test response does not exclude mild secondary forms of adrenal insufficiency (81–84).

Other tests, that directly evaluate pituitary reserve are also available (insulin induced hypoglycemia, metyrapone administration, or CRH stimulation). Also in the assessment of the HPA axis (similar to the GH-IGF-I axis) the ITT still remains the golden standard (85;86). Insulin induced hypoglycemia results in stress which actives the entire HPA axis providing proof for adequate hypothalamic (CRH) and pituitary (ACTH) function. In healthy subjects serum cortisol levels will increase above 550 nmol/L if adequate hypoglycemia is achieved (glucose 2.2 mmol/L or lower). Stimulation with metyrapone is an alternative test to assess the HPA axis. The adrenal enzyme 11-β-hydroxylase (CYP11B1) that catalyzes the conversion of 11-deoxycortisol to cortisol, is inhibited by metyrapone, resulting in a reduction of cortisol secretion. Administration of metyrapone will thus result in activation of the HPA axis, an increase in ACTH secretion and consequently an increase in adrenal steroidogenesis up to 11-deoxycortisol. An 11-deoxycortisol concentration above 200 nmol/L in the presence of suppressed cortisol levels (below 100 nmol/L) is then indicative for central adrenal insufficiency (87–89). This test can be performed as a prolonged and short overnight version, depending on the number of dosages of metyrapone given. It appears, however, that the ACTH stimulus of a single dose of metyrapone is comparable to that of an insulin tolerance test (89).

Since the 1980's ovine CRH is used for the evaluation of the HPA axis, mainly to discriminate between pituitary or adrenal causes of Cushing's syndrome (90-94). However, in recent years the CRH test is more often used to assess secondary adrenal insufficiency (95;96). Administration of an intravenous bolus of ovine CRH results in pituitary ACTH secretion resulting in cortisol secretion by the adrenal glands. In healthy subjects a 1 μ g/kg i.v. CRH bolus results in a peak ACTH response within 15 min and a peak cortisol response within 30–60 min. A peak cortisol of 550 nmol/L or higher is considered to be a sufficient reaction. The CRH test however, is inferior to the ITT and metyrapone test (97).

Pitfalls

The use of exogenous corticosteroids can suppress the HPA axis. Therefore, in case of exogenous glucocorticoid use, a reliable evaluation of the HPA axis can not be performed within 6 weeks after withdrawal of the steroid but might even be disturbed many months thereafter. Contraceptives in females should also be stopped for at least 6 weeks

because of the effects of hormonal agents on cortisol binding globulin (CBG) levels (98;99).

Thyrotropin deficiency

Clinical manifestation

The symptoms and signs associated with thyrotropin (TSH) deficiency are similar to those of primary hypothyroidism but usually are less severe, as there often is some residual thyrotropin secretion. In addition, TSH deficiency is almost always part of complete anterior pituitary hormone deficiency because thyreotroph secretion is the most resistant to insufficiency. Tiredness, cold intolerance, weight gain, constipation, dry skin, and hair loss are common features.

Diagnosis

TSH deficiency is diagnosed by low or normal serum TSH concentrations in the presence of low serum free thyroxine (fT_4) level. Measurement of serum fT_3 is not of additional value but may be low or normal. Thyrotropin-releasing hormone (TRH) can be used to assess TSH secretion. However, the response to TRH varies widely among individuals. Therefore it is not possible to discriminate between a normal and abnormal response in the majority of cases and TRH has not been incorporated into routine clinical practice of the evaluation of TSH deficiency (100).

Gonadotropin deficiency

Clinical manifestations

The clinical features of gonadotropin deficiency are determined by gender and the age of development. The physical examination in men with recent onset hypogonadism will usually be normal. However, in longstanding hypogonadism diminished facial and body hair, gynaecomastia, and small, weak testes can be present. Libido may be reduced and the ability to achieve and maintain an erection may be compromised. Patients can also complain of nonspecific symptoms, such as tiredness, reduced muscle strength, reduced exercise capacity, but also emotional lability and depression. The symptoms in men are nonspecific and therefore

may not become evident for many years, particularly if fertility is not an issue (77).

In women gonadotropin deficiency leads to menstrual disturbances (*i.e.* oligomenorrhea or amenorrhea) and therefore often earlier diagnosed compared to men (2).

Diagnosis

The diagnosis in women is straightforward. In premenopausal women secondary amenorrhea with low levels of estradiol and low or nomal levels of gonadotropins will confirm the diagnosis. Whereas, in postmenopausal women FSH and LH will be (undetectably) low.

Low or normal gonadotropin levels combined with serum testosterone levels below the reference range, corrected for age are sufficient to confirm the diagnosis. Because of the great circadian variation randomly found decreased testosterone levels should be repeated in the early morning (between 8–9:00 AM).

Prolactin deficiency

Clinical manifestations

Mild hyperprolactinaemia (up to 5 times the upper limit of normal) is common in patients with hypopituitarism. A pituitary mass with supra-sellar extension may compress the stalk resulting in decreased dopaminergic inhibition of prolactin secretion.

Raised prolactine levels effects pulsatile secretion of gonadtropins resulting in hypogonadism. Galacthorrhea can also be present. Prolactin deficiency almost invariably results from lactotroph deficiency secondary to hypothalamic damage as a result of irradiation and or surgery.

Diagnosis

The diagnosis of prolactin deficiency is straightforward using commercially available assays with gender adjusted reference ranges for the determination of prolactin concentrations. However, unless it is in the postpartum period, there are no clinical implications.

IV. Outline of this thesis

Pituitary insufficiency in the presence of a pituitary macroadenoma or after pituitary irradiation is frequently reported. In addition, pituitary insufficiency is increasingly reported after traumatic head injuries. The correct evaluation and interpretation, however, of the pituitary axes, and consequently, the potential therapeutical consequences are a matter of controversies. The studies reported in this thesis aim to provide better insight into the complexity of different endocrine tests used for the evaluation of possible pituitary insufficiency and in the treatment of patients with pituitary insufficiency.

The evaluation of pituitary function in patients after traumatic brain injury

Traumatic brain injury (TBI) has emerged as an important cause of hypopituitarism. However, considerable variations in the prevalence of hypopituitarism are reported. These variations can partly be explained by the severity of trauma and timing of hormonal evaluation, but may also be dependent on endocrine tests and criteria used for diagnosis of hypopituitarism. Therefore, in **chapter 2**, we performed a systematic review of the literature to critically compare pituitary function tests, and definitions of hypopituitarism in studies that assessed the long-term outcome of TBI on pituitary function.

Because of the great variation in prevalence rates reported and the great variation in endocrine assessments used, we decided to perform a cross-sectional study in the Netherlands of a large cohort of 112 TBI patients evaluated after long-term follow-up. We assessed the prevalence of pituitary insufficiency in our own large cohort of TBI patients using a standardized endocrine evaluation, described in **chapter 3**. In these patients, we also evaluated quality of life (QoL) using different QoL questionnaires.

Dynamic tests of pituitary function in other pituitary diseases

Pituitary adenomas and their treatment (*i.e.* surgery and/or radiotherapy) are also causes for pituitary insufficiency. Pituitary insufficiency is a complication that can be attributed to the tumor itself (compression) but also to the surgical approach and/or subsequent radiotherapeutical intervention. Therefore, accurate assessment of pituitary function is critical for appropriate management of patients with pituitary adenoma after surgery with or without irradiation. For many years, all patients in our hospital underwent a CRH stimulation test for the evaluation of the HPA axis shortly after pituitary surgery. In **chapter 4**, we describe a retrospective study that evaluated the clinical applicability of the CRH test directly after TS in our center.

The ITT, however, is considered the golden standard test for the evaluation of the HPA axis. In **chapter 5**, we describe a study on the long-term prevalence of adrenal insufficiency after transsphenoidal surgery for growth-hormone secreting pituitary adenomas using the ITT and CRH test in the majority of the patients. The reason for this evaluation was a recently published study that reported a remarkably high prevalence of adrenal insufficiency after surgical and/or medical treatment without postoperative radiotherapy in patients treated for acromegaly. Therefore, in our study, we evaluated the prevalence and incidence rate of adrenal insufficiency in 91 consecutive patients during long-term follow-up after successful transsphenoidal surgery for acromegaly.

In addition to patients with pituitary tumors, patients with nonpituitary intracranial and or nasopharyngeal tumors are treated by radiotherapy, in which the pituitary gland is involved in the radiation field. These patients are also at risk for pituitary insufficiency. To assess the prevalence of possible pituitary insufficiencies we performed a systemic literature search and meta-analysis focusing on the prevalence of pituitary dysfunction in adult patients treated with radiotherapy for nonpituitary tumors, which is described in **chapter 6**.

Treatment of GH deficiency

When growth hormone deficiency is diagnosed, the therapeutical consequences should be carefully evaluated, especially in certain conditions like obesity and during senescence where GH secretion overlaps with a GH deficient state. With increasing age, but also increasing BMI, GH secretion decreases. Therefore, the effects of treatment with rhGH in obesity and in the elderly diagnosed with GHD might be different.

Therefore, in **chapter 7,** we performed a structured review, to critically assess the available literature in order to evaluate the available evidence for treatments of elderly patients with GHD.

References

- 1. Berne R, Levy NM, Koeppen BM, Stanton A. Physiology. 5th ed. 2011
- 2. Greenspan FS, Strewler GJ. Basic & Clinical Endocrinology. 5th ed. 1997
- 3. Giustina A, Veldhuis JD 1998 Pathophysiology of the neuroregulation of growth hormone secretion in experimental animals and the human. Endocr Rev 19:717–797
- 4. Hartman ML, Veldhuis JD, Thorner MO 1993 Normal control of growth hormone secretion. Horm Res 40:37–47
- 5. Pincus SM, Gevers EF, Robinson IC, van den BG, Roelfsema F, Hartman ML, Veldhuis JD 1996 Females secrete growth hormone with more process irregularity than males in both humans and rats. Am J Physiol 270:E107–E115
- 6. Veldhuis JD, Liem AY, South S, Weltman A, Weltman J, Clemmons DA, Abbott R, Mulligan T, Johnson ML, Pincus S, . 1995 Differential impact of age, sex steroid hormones, and obesity on basal versus pulsatile growth hormone secretion in men as assessed in an ultrasensitive chemiluminescence assay. J Clin Endocrinol Metab 80:3209–3222
- 7. Iranmanesh A, Lizarralde G, Veldhuis JD 1991 Age and relative adiposity are specific negative determinants of the frequency and amplitude of growth hormone (GH) secretory bursts and the half-life of endogenous GH in healthy men. J Clin Endocrinol Metab 73:1081–1088
- 8. Veldhuis JD, Iranmanesh A, Ho KK, Waters MJ, Johnson ML, Lizarralde G 1991 Dual defects in pulsatile growth hormone secretion and clearance subserve the hyposomatotropism of obesity in man. J Clin Endocrinol Metab 72:51–59
- 9. Rosenfeld RG, Roberts CT. The IGF System. 1999
- 10. Le Roith D, Bondy C, Yakar S, Liu JL, Butler A 2001 The somatomedin hypothesis: 2001. Endocr Rev 22:53–74
- 11. Gwinup G, Johnson B 1975 Clinical testing of the hypothalamic-pituitary-adrenocortical system in states of hypo- and hypercortisolism. Metabolism 24:777–791
- 12. Debono M, Ghobadi C, Rostami-Hodjegan A, Huatan H, Campbell MJ, Newell-Price J, Darzy K, Merke DP, Arlt W, Ross RJ 2009 Modified-release hydrocortisone to provide circadian cortisol profiles. J Clin Endocrinol Metab 94:1548–1554
- Weitzman ED, Fukushima D, Nogeire C, Roffwarg H, Gallagher TF, Hellman L 1971 Twenty-four hour pattern of the episodic secretion of cortisol in normal subjects. J Clin Endocrinol Metab 33:14–22

- 14. Gallagher TF, Yoshida K, Roffwarg HD, Fukushima DK, Weitzman ED, Hellman L 1973 ACTH and cortisol secretory patterns in man. J Clin Endocrinol Metab 36:1058–1068
- 15. Veldhuis JD, Iranmanesh A, Johnson ML, Lizarralde G 1990 Twenty-four-hour rhythms in plasma concentrations of adenohypophyseal hormones are generated by distinct amplitude and/or frequency modulation of underlying pituitary secretory bursts. J Clin Endocrinol Metab 71:1616–1623
- 16. Vaughan GM, Becker RA, Allen JP, Goodwin CW, Jr., Pruitt BA, Jr., Mason AD, Jr. 1982 Cortisol and corticotrophin in burned patients. J Trauma 22:263–273
- 17. Jeffries MK, Vance ML 1992 Growth hormone and cortisol secretion in patients with burn injury. J Burn Care Rehabil 13:391–395
- 18. Udelsman R, Norton JA, Jelenich SE, Goldstein DS, Linehan WM, Loriaux DL, Chrousos GP 1987 Responses of the hypothalamic-pituitary-adrenal and renin-angiotensin axes and the sympathetic system during controlled surgical and anesthetic stress. J Clin Endocrinol Metab 64:986–994
- Calogero AE, Norton JA, Sheppard BC, Listwak SJ, Cromack DT, Wall R, Jensen RT, Chrousos GP 1992 Pulsatile activation of the hypothalamic-pituitary-adrenal axis during major surgery. Metabolism 41:839–845
- Watabe T, Tanaka K, Kumagae M, Itoh S, Takeda F, Morio K, Hasegawa M, Horiuchi T, Miyabe S, Shimizu N 1987 Hormonal responses to insulin-induced hypoglycemia in man. J Clin Endocrinol Metab 65:1187–1191
- 21. Fish HR, Chernow B, O'Brian JT 1986 Endocrine and neurophysiologic responses of the pituitary to insulin-induced hypoglycemia: a review. Metabolism 35:763–780
- 22. Luger A, Deuster PA, Kyle SB, Gallucci WT, Montgomery LC, Gold PW, Loriaux DL, Chrousos GP 1987 Acute hypothalamic-pituitary-adrenal responses to the stress of treadmill exercise. Physiologic adaptations to physical training. N Engl J Med 316:1309–1315
- 23. Streeten DH, Anderson GH, Jr., Dalakos TG, Seeley D, Mallov JS, Eusebio R, Sunderlin FS, Badawy SZ, King RB 1984 Normal and abnormal function of the hypothalamic-pituitary-adrenocortical system in man. Endocr Rev 5:371–394
- 24. Keller-Wood M, Shinsako J, Dallman MF 1984 Interaction between stimulus intensity and corticosteroid feedback in control of ACTH. Am J Physiol 247:E489–E494
- 25. Schlaghecke R, Kornely E, Santen RT, Ridderskamp P 1992 The effect of long-term glucocorticoid therapy on pituitary-adrenal responses to exogenous corticotropin-releasing hormone. N Engl J Med 326:226–230
- 26. Livanou T, Ferriman D, James VH 1967 Recovery of hypothalamo-pituitary-adrenal function after corticosteroid therapy. Lancet 2:856–859
- 27. Harrison BD, Rees LH, Cayton RM, Nabarro JD 1982 Recovery of hypothalamo-pituitary-adrenal function in asthmatics whose oral steroids have been stopped or reduced. Clin Endocrinol (Oxf) 17:109–118

- 28. Streck WF, Lockwood DH 1979 Pituitary adrenal recovery following short-term suppression with corticosteroids. Am J Med 66:910–914
- 29. Melby JC 1974 Drug spotlight program: systemic corticosteroid therapy: pharmacology and endocrinologic considerations. Ann Intern Med 81:505–512
- 30. Arafah BM, Prunty D, Ybarra J, Hlavin ML, Selman WR 2000 The dominant role of increased intrasellar pressure in the pathogenesis of hypopituitarism, hyperprolactinemia, and headaches in patients with pituitary adenomas. J Clin Endocrinol Metab 85:1789–1793
- 31. Fainstein DP, Guitelman M, Artese R, Fiszledjer L, Chervin A, Vitale NM, Stalldecker G, De M, V, Cornalo D, Alfieri A, Susana M, Gil M 2004 Retrospective multicentric study of pituitary incidentalomas. Pituitary 7:145–148
- 32. Feldkamp J, Santen R, Harms E, Aulich A, Modder U, Scherbaum WA 1999 Incidentally discovered pituitary lesions: high frequency of macroadenomas and hormone-secreting adenomas results of a prospective study. Clin Endocrinol (Oxf) 51:109–113
- 33. Gondim JA, Almeida JP, Albuquerque LA, Schops M, Gomes E, Ferraz T, Sobreira W, Kretzmann MT Endoscopic endonasal approach for pituitary adenoma: surgical complications in 301 patients. Pituitary 2011 Jun;14(2):174-83
- 34. Tabaee A, Anand VK, Barron Y, Hiltzik DH, Brown SM, Kacker A, Mazumdar M, Schwartz TH 2009 Endoscopic pituitary surgery: a systematic review and meta-analysis. J Neurosurg 111:545–554
- 35. Ayuk J, Stewart PM 2009 Mortality following pituitary radiotherapy. Pituitary 12:35–39
- 36. Fernandez A, Brada M, Zabuliene L, Karavitaki N, Wass JA 2009 Radiation-induced hypopituitarism. Endocr Relat Cancer 16:733–772
- 37. Bhandare N, Kennedy L, Morris CG, Malyapa R, Mendenhall WM 2006 1099: Hypopituitarism After Radiation Therapy for Extracranial Head and Neck Cancers. International Journal of Radiation Oncology Biology Physics 66:S187–S188
- 38. Darzy KH, Shalet SM 2009 Hypopituitarism following Radiotherapy Revisited. Endocr Dev 15:1–24
- 39. Shalet SM 1993 Radiation and pituitary dysfunction. N Engl J Med 328:131–133
- 40. Agha A, Rogers B, Sherlock M, O'Kelly P, Tormey W, Phillips J, Thompson CJ 2004 Anterior pituitary dysfunction in survivors of traumatic brain injury. J Clin Endocrinol Metab 89:4929–4936
- 41. Bondanelli M, Ambrosio MR, Cavazzini L, Bertocchi A, Zatelli MC, Carli A, Valle D, Basaglia N, Uberti EC 2007 Anterior pituitary function may predict functional and cognitive outcome in patients with traumatic brain injury undergoing rehabilitation. J Neurotrauma 24:1687–1697
- 42. Bushnik T, Englander J, Katznelson L 2007 Fatigue after TBI: association with neuroendocrine abnormalities. Brain Inj 21:559–566

- 43. Herrmann BL, Rehder J, Kahlke S, Wiedemayer H, Doerfler A, Ischebeck W, Laumer R, Forsting M, Stolke D, Mann K 2006 Hypopituitarism following severe traumatic brain injury. Exp Clin Endocrinol Diabetes 114:316–321
- 44. Kelly DF, Gonzalo IT, Cohan P, Berman N, Swerdloff R, Wang C 2000 Hypopituitarism following traumatic brain injury and aneurysmal subarachnoid hemorrhage: a preliminary report.

 J Neurosurg 93:743–752
- 45. Klose M, Juul A, Poulsgaard L, Kosteljanetz M, Brennum J, Feldt-Rasmussen U 2007 Prevalence and predictive factors of post-traumatic hypopituitarism. Clin Endocrinol (Oxf) 67:193–201
- 46. Leal-Cerro A, Flores JM, Rincon M, Murillo F, Pujol M, Garcia-Pesquera F, Dieguez C, Casanueva FF 2005 Prevalence of hypopituitarism and growth hormone deficiency in adults long-term after severe traumatic brain injury. Clin Endocrinol (Oxf) 62:525–532
- 47. Lieberman SA, Oberoi AL, Gilkison CR, Masel BE, Urban RJ 2001 Prevalence of neuroendocrine dysfunction in patients recovering from traumatic brain injury. J Clin Endocrinol Metab 86:2752–2756
- 48. Popovic V, Pekic S, Pavlovic D, Maric N, Jasovic-Gasic M, Djurovic B, Medic SM, Zivkovic V, Stojanovic M, Doknic M, Milic N, Djurovic M, Dieguez C, Casanueva FF 2004 Hypopituitarism as a consequence of traumatic brain injury (TBI) and its possible relation with cognitive disabilities and mental distress. J Endocrinol Invest 27:1048–1054
- 49. Schneider HJ, Schneider M, Saller B, Petersenn S, Uhr M, Husemann B, von RF, Stalla GK 2006
 Prevalence of anterior pituitary insufficiency 3 and 12 months after traumatic brain injury.
 Eur J Endocrinol 154:259–265
- 50. Tanriverdi F, Senyurek H, Unluhizarci K, Selcuklu A, Casanueva FF, Kelestimur F 2006 High risk of hypopituitarism after traumatic brain injury: a prospective investigation of anterior pituitary function in the acute phase and 12 months after trauma. J Clin Endocrinol Metab 91:2105–2111
- 51. Wachter D, Gundling K, Oertel MF, Stracke H, Boker DK 2009 Pituitary insufficiency after traumatic brain injury. J Clin Neurosci 16:202–208
- 52. Schneider HJ, Kreitschmann-Andermahr I, Ghigo E, Stalla GK, Agha A 2007 Hypothalamopituitary dysfunction following traumatic brain injury and aneurysmal subarachnoid hemorrhage: a systematic review. JAMA 298:1429–1438
- 53. Urban RJ, Harris P, Masel B 2005 Anterior hypopituitarism following traumatic brain injury.

 Brain Inj 19:349–358
- 54. Agha A, Thompson CJ 2006 Anterior pituitary dysfunction following traumatic brain injury (TBI). Clin Endocrinol (Oxf) 64:481–488
- 55. de Boer H, Blok GJ, Van der Veen, V 1995 Clinical aspects of growth hormone deficiency in adults. Endocr Rev 16:63–86

- 56. Brabant G, Poll EM, Jonsson P, Polydorou D, Kreitschmann-Andermahr I 2009 Etiology, baseline characteristics, and biochemical diagnosis of GH deficiency in the adult: are there regional variations? Eur J Endocrinol 161 Suppl 1:S25–S31
- 57. Carroll PV, Christ ER, Bengtsson BA, Carlsson L, Christiansen JS, Clemmons D, Hintz R, Ho K, Laron Z, Sizonenko P, Sonksen PH, Tanaka T, Thorne M 1998 Growth hormone deficiency in adulthood and the effects of growth hormone replacement: a review. Growth Hormone Research Society Scientific Committee. J Clin Endocrinol Metab 83:382–395
- 58. Attanasio AF, Mo D, Erfurth EM, Tan M, Ho KY, Kleinberg D, Zimmermann AG, Chanson P 2010 Prevalence of metabolic syndrome in adult hypopituitary growth hormone (GH)-deficient patients before and after GH replacement. J Clin Endocrinol Metab 95:74–81
- 59. Burman P, Deijen JB 1998 Quality of life and cognitive function in patients with pituitary insufficiency. Psychother Psychosom 67:154–167
- 60. Rosen T, Wiren L, Wilhelmsen L, Wiklund I, Bengtsson BA 1994 Decreased psychological well-being in adult patients with growth hormone deficiency. Clin Endocrinol (Oxf) 40:111–116
- 61. Abs R, Mattsson AF, Bengtsson BA, Feldt-Rasmussen U, Goth MI, Koltowska-Haggstrom M, Monson JP, Verhelst J, Wilton P 2005 Isolated growth hormone (GH) deficiency in adult patients: baseline clinical characteristics and responses to GH replacement in comparison with hypopituitary patients. A sub-analysis of the KIMS database. Growth Horm IGF Res 15:349–359
- 62. Aimaretti G, Corneli G, Rovere S, Granata R, Baldelli R, Grottoli S, Ghigo E 2004 Insulin-like growth factor I levels and the diagnosis of adult growth hormone deficiency. Horm Res 62 Suppl 1:26–33
- 63. Ghigo E, Aimaretti G, Corneli G 2008 Diagnosis of adult GH deficiency. Growth Horm IGF Res 18:1–16
- 64. Aimaretti G, Corneli G, Baldelli R, Di SC, Gasco V, Durante C, Ausiello L, Rovere S, Grottoli S, Tamburrano G, Ghigo E 2003 Diagnostic reliability of a single IGF-I measurement in 237 adults with total anterior hypopituitarism and severe GH deficiency. Clin Endocrinol (Oxf) 59:56–61
- 65. Aimaretti G, Corneli G, Di SC, Baldelli R, Gasco V, Rovere S, Migliaretti G, Colao A, Tamburrano G, Lombardi G, Ghigo E, Camanni F 2005 Different degrees of GH deficiency evidenced by GHRH-arginine test and IGF-I levels in adults with pituritary disease. J Endocrinol Invest 28:247–252
- 66. Ghigo E, Aimaretti G, Arvat E, Camanni F 2001 Growth hormone-releasing hormone combined with arginine or growth hormone secretagogues for the diagnosis of growth hormone deficiency in adults. Endocrine 15:29–38
- 67. Consensus guidelines for the diagnosis and treatment of adults with growth hormone deficiency: summary statement of the Growth Hormone Research Society Workshop on Adult Growth Hormone Deficiency 1998 J Clin Endocrinol Metab 83:379–381

- 68. Aimaretti G, Corneli G, Razzore P, Bellone S, Baffoni C, Arvat E, Camanni F, Ghigo E 1998 Comparison between insulin-induced hypoglycemia and growth hormone (GH)-releasing hormone + arginine as provocative tests for the diagnosis of GH deficiency in adults. J Clin Endocrinol Metab 83:1615–1618
- 69. Biller BM, Samuels MH, Zagar A, Cook DM, Arafah BM, Bonert V, Stavrou S, Kleinberg DL, Chipman JJ, Hartman ML 2002 Sensitivity and specificity of six tests for the diagnosis of adult GH deficiency. J Clin Endocrinol Metab 87:2067–2079
- 70. Alba-Roth J, Muller OA, Schopohl J, von WK 1988 Arginine stimulates growth hormone secretion by suppressing endogenous somatostatin secretion. J Clin Endocrinol Metab 67:1186–1189
- 71. Shibasaki T, Hotta M, Masuda A, Imaki T, Obara N, Demura H, Ling N, Shizume K 1985 Plasma GH responses to GHRH and insulin-induced hypoglycemia in man. J Clin Endocrinol Metab 60:1265–1267
- 72. Colao A, Di SC, Savastano S, Rota F, Savanelli MC, Aimaretti G, Lombardi G 2009 A reappraisal of diagnosing GH deficiency in adults: role of gender, age, waist circumference, and body mass index. J Clin Endocrinol Metab 94:4414–4422
- 73. Kopelman PG, Noonan K, Goulton R, Forrest AJ 1985 Impaired growth hormone response to growth hormone releasing factor and insulin-hypoglycaemia in obesity. Clin Endocrinol (Oxf) 23:87–94
- 74. Scacchi M, Pincelli Al, Cavagnini F 1999 Growth hormone in obesity. Int J Obes Relat Metab Disord 23:260–271
- 75. Veldhuis JD, Roelfsema F, Keenan DM, Pincus S 2011 Gender, age, body mass index, and IGF-I individually and jointly determine distinct GH dynamics: analyses in one hundred healthy adults. J Clin Endocrinol Metab 96:115–121
- 76. Qu XD, Gaw G, I, Al Sayed MY, Cohan P, Christenson PD, Swerdloff RS, Kelly DF, Wang C 2005 Influence of body mass index and gender on growth hormone (GH) responses to GH-releasing hormone plus arginine and insulin tolerance tests. J Clin Endocrinol Metab 90:1563–1569
- 77. van Aken MO, Lamberts SW 2005 Diagnosis and treatment of hypopituitarism: an update. Pituitary 8:183–191
- 78. Arlt W, Allolio B 2003 Adrenal insufficiency. Lancet 361:1881–1893
- 79. Clayton RN 1996 Short Synacthen test versus insulin stress test for assessment of the hypothalamo [correction of hypothalmo]-pituitary--adrenal axis: controversy revisited. Clin Endocrinol (Oxf) 44:147–149
- 80. Grinspoon SK, Biller BM 1994 Clinical review 62: Laboratory assessment of adrenal insufficiency.

 J Clin Endocrinol Metab 79:923–931
- 81. Streeten DH 1999 What test for hypothalamic-pituitary-adrenocortical insufficiency? Lancet 354:179–180

- 82. Nye EJ, Grice JE, Hockings GI, Strakosch CR, Crosbie GV, Walters MM, Jackson RV 1999 Comparison of adrenocorticotropin (ACTH) stimulation tests and insulin hypoglycemia in normal humans: low dose, standard high dose, and 8-hour ACTH-(1–24) infusion tests. J Clin Endocrinol Metab 84:3648–3655
- 83. Mayenknecht J, Diederich S, Bahr V, Plockinger U, Oelkers W 1998 Comparison of low and high dose corticotropin stimulation tests in patients with pituitary disease. J Clin Endocrinol Metab 83:1558–1562
- 84. Abdu TA, Elhadd TA, Neary R, Clayton RN 1999 Comparison of the low dose short synacthen test (1 microg), the conventional dose short synacthen test (250 microg), and the insulin tolerance test for assessment of the hypothalamo-pituitary-adrenal axis in patients with pituitary disease. J Clin Endocrinol Metab 84:838–843
- 85. Stewart PM, Clark PM, Sheppard MC 1998 Comparison of the short ACTH stimulation test with the insulin tolerance/glucagon test. Clin Endocrinol (Oxf) 48:124–126
- 86. Mukherjee JJ, de Castro JJ, Kaltsas G, Afshar F, Grossman AB, Wass JA, Besser GM 1997 A comparison of the insulin tolerance/glucagon test with the short ACTH stimulation test in the assessment of the hypothalamo-pituitary-adrenal axis in the early post-operative period after hypophysectomy. Clin Endocrinol (Oxf) 47:51–60
- 87. Fiad TM, Kirby JM, Cunningham SK, McKenna TJ 1994 The overnight single-dose metyrapone test is a simple and reliable index of the hypothalamic-pituitary-adrenal axis. Clin Endocrinol (Oxf) 40:603–609
- 88. Courtney CH, McAllister AS, McCance DR, Hadden DR, Leslie H, Sheridan B, Atkinson AB 2000 The insulin hypoglycaemia and overnight metyrapone tests in the assessment of the hypothalamic-pituitary-adrenal axis following pituitary surgery. Clin Endocrinol (Oxf) 53:309–312
- 89. Staub JJ, Noelpp B, Girard J, Baumann JB, Graf S, Ratcliffe JG 1979 The short metyrapone test: comparison of the plasma ACTH response to metyrapone and insulin-induced hypoglycaemia. Clin Endocrinol (Oxf) 10:595–601
- 90. de Lange WE, Sluiter WJ, Pratt JJ, Wolf RF, Doorenbos H 1988 Is the ovine CRF test an aid in the evaluation of patients with secondary adrenal insufficiency? Neth J Med 32:72–78
- 91. Hermus AR, Pieters GF, Smals AG, Kloppenborg PW 1985 Corticotropin-releasing factor, a new aid in the diagnosis of pituitary-adrenal disorders? Neth J Med 28:2–5
- 92. Pieters GF, Hermus AR, Smals AG, Bartelink AK, Benraad TJ, Kloppenborg PW 1983 Responsiveness of the hypophyseal-adrenocortical axis to corticotropin-releasing factor in pituitary-dependent Cushing's disease. J Clin Endocrinol Metab 57:513–516
- 93. Schulte HM, Chrousos GP, Avgerinos P, Oldfield EH, Gold PW, Cutler GB, Jr., Loriaux DL 1984 The corticotropin-releasing hormone stimulation test: a possible aid in the evaluation of patients with adrenal insufficiency. J Clin Endocrinol Metab 58:1064–1067

- 94. Stalla GK, Losa M, Oeckler R, Muller OA, von WK 1988 Insulin hypoglycemia test and releasing hormone (corticotropin-releasing hormone and growth hormone-releasing hormone) stimulation in patients with pituitary failure of different origin. Horm Res 29:191–196
- 95. Dullaart RP, Pasterkamp SH, Beentjes JA, Sluiter WJ 1999 Evaluation of adrenal function in patients with hypothalamic and pituitary disorders: comparison of serum cortisol, urinary free cortisol and the human-corticotrophin releasing hormone test with the insulin tolerance test. Clin Endocrinol (Oxf) 50:465–471
- 96. Tsukada T, Nakai Y, Koh T, Tsujii S, Inada M, Nishikawa M, Shinoda H, Kawai I, Takezawa N, Imura H 1984 Plasma adrenocorticotropin and cortisol responses to ovine corticotropin-releasing factor in patients with adrenocortical insufficiency due to hypothalamic and pituitary disorders. J Clin Endocrinol Metab 58:758–760
- 97. Dickstein G 2003 The assessment of the hypothalamo-pituitary-adrenal axis in pituitary disease: are there short cuts? J Endocrinol Invest 26:25–30
- 98. Kirschbaum C, Kudielka BM, Gaab J, Schommer NC, Hellhammer DH 1999 Impact of gender, menstrual cycle phase, and oral contraceptives on the activity of the hypothalamus-pituitary-adrenal axis. Psychosom Med 61:154–162
- 99. Kumsta R, Entringer S, Hellhammer DH, Wust S 2007 Cortisol and ACTH responses to psychosocial stress are modulated by corticosteroid binding globulin levels. Psychoneuroendocrinology 32:1153–1157
- 100. Faglia G 1998 The clinical impact of the thyrotropin-releasing hormone test. Thyroid 8:903–908