

The course of clinically suspect arthralgia and early rheumatoid arthritis : clinical features, imaging and genetics

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SPP1 rs9138 variant contributes to the severity of radiological damage in anti-citrullinated peptide autoantibody-negative rheumatoid arthritis

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ABSTRACT

Objective

We recently reported an association of the *SPP1* rs9138 and rs11439060 functional variants with the risk of rheumatoid arthritis (RA), the association being greater in anti-citrullinated peptide autoantibody (ACPA)-negative patients. We hypothesised that *SPP1* may contribute to the severity of joint destruction in RA, specifically in the ACPA-negative population.

Methods

Patients with RA in the ESPOIR cohort underwent genotyping for *SPP1* rs9138 and rs11439060. Radiographs of the hands and feet were obtained at the first visit and at 1- and 2 year follow-up. Association analyses were performed by ACPA-status. A replication study of the relevant subset of the Leiden Early Arthritis Clinic (EAC) cohort was performed.

Results

In the ESPOIR cohort (652 patients), rs9138 was significantly associated with radiological progression of joint destruction at 2 years, the association being restricted to 358 ACPAnegative patients (p=0.034). In the replication study within the Leiden EAC cohort (273 ACPA-negative patients), rs4754, which is in complete linkage disequilibrium with rs9138, was significantly associated with joint damage progression in ACPA-negative patients at 2 and 7 year follow-up (p=0.019 and p=0.005, respectively). Combined analysis of the two cohorts revealed a 0.95 fold rate of joint destruction per year per minor allele (p=0.022).

Conclusions

The SPP1 rs9138 variant contributes to joint damage progression in ACPA-negative RA.

INTRODUCTION

Rheumatoid arthritis (RA) is a systemic, inflammatory, autoimmune disease characterised by peripheral synovial joint inflammation which can lead to joint destruction. Approximately two-thirds of RA cases are seropositive for rheumatoid factor or anti-citrullinated peptide autoantibodies (ACPA) ¹. The heritability of ACPA-positive and ACPA-negative disease is comparable ², and recent association studies have provided further support for distinct genetic aetiologies of ACPA-positive and ACPA-negative RA subsets ^{3,4}. ACPA-positive RA patients are particularly characterised by progressive joint destruction ⁵. However, little information is available on joint destruction processes in the ACPA-negative subset. Currently, we cannot offer personalised medicine for patients with RA because we cannot identify those who will have the most severe disease course, nor do we understand the pathogenesis underlying these interindividual variances. To improve this situation, identification of risk factors for joint destruction is required.

Genetic variants are estimated to contribute to 58% of the total variance in RA joint destruction, with clinical and serological risk factors explaining only about one-third of the total phenotypic variation ⁶. Most risk alleles for RA joint destruction have been identified in ACPA-positive patients or in pooled ACPA-negative and ACPA-positive patients, but we lack information about the genetic contribution to ACPA-negative RA severity ^{7–9}. Identifying individual genetic risk factors would increase our understanding of the mechanisms underlying variation in severity of joint destruction, particularly in ACPA-negative disease.

Recently, through a large case-control association study, our group reported a significant contribution of the combination of the *SPP1* rs11439060 and rs9138 frequent alleles to risk of RA, the magnitude of the association being greater in ACPA-negative patients ³. These patients fulfilled the 1987 American College of Rheumatology (ACR) revised criteria for RA, which include radiographic changes typical of RA ¹⁰; so ACPA-negative individuals may have had joint destruction to be classified as having RA. *SPP1* encodes osteopontin (OPN), an extracellular-matrix glycosylated phosphoprotein with multiple functions including bone formation and remodelling ¹¹. Consequently, we hypothesised that *SPP1* variants may contribute to the severity of joint destruction in RA, specifically in ACPA-negative patients.

PATIENTS AND METHODS

Study population

The exploratory study included 652 patients with RA from the ESPOIR cohort who were positive or negative for ACPA and were included in the large case-control association study previously reported ^{3,12}. The replication study included 273 ACPA-negative RA cases from the Leiden Early Arthritis Clinic (EAC) cohort (table 1) ¹³. All patients fulfilled the 1987 ACR revised criteria for RA ¹⁰. They all provided informed written consent as approved by the

Table 1. Characteristics of the ESPOIR and Leiden EAC cohorts of patients with RA genotyped for *SPP1* rs9138 or rs4574

Cohort	No of patients	No of sets of radiographs of hands and feet	Year of diagnosis	Follow- up (years)	Method of scoring	ICC	ACPA- negative (%)	Age (years), mean±SD	Female patients, n (%)
ESPOIR	652	1,768	2002- 2005	2	SHS	0.97	54.9	47.9±12.2	500 (76.7)
Leiden EAC:	273	1,316	1993- 2006	7	SHS	0.91	100	58.9±16.3	187 (68.5)
ACPA- negative									
population									

recruiting site review board at each of the affiliated institutions.

Genotyping

In the exploratory study with the ESPOIR cohort, *SPP1* rs11439060 and rs9138 variants were genotyped by use of a competitive allele-specific PCR system (Kaspar genotyping; Kbioscience, Hoddeston, UK) ³. In the Leiden EAC cohort, *SPP1* rs4754, which is in complete linkage disequilibrium with rs9138 ³, was genotyped using Illumina Human CytoSNP-12V2.

Radiographic joint destruction

In both the ESPOIR and Leiden EAC cohorts, all radiographs of hands and feet were scored by the Sharp-van der Heijde score (SHS) by one experienced reader per cohort who was blinded to clinical, biological and genetic data ¹⁴. The intraclass correlation coefficient was 0.97 and 0.91, respectively.

Statistical analysis

A multivariate regression analysis (MRA) was used, with radiographic damage as the response variable (see online supplementary text for a detailed description of the MRA). The analyses were performed with the genetic variable and its interaction with time in the model, reflecting a constant and a time-dependent effect of progression of joint damage, respectively ¹⁵.

To validate our a priori hypothesis (ie, the contribution of *SPP1* rs11439060, rs9138 and the rs11439060-rs9138 risk allele combination in the ACPA-negative population), we selected the best-fit model at the exploratory stage, as previously described, which was then replicated in the Leiden EAC cohort ³. MRA of both ACPA-negative populations was used to assess the magnitude of the *SPP1* effect on radiographic joint destruction in early RA. Analyses used SPSS V.20.0.

RESULTS

Exploratory study

We analysed data for 652 patients with RA and 1768 radiographs. For a complete overview of the MRA of *SPP1* rs11439060, rs9138 and risk allele combination, see online supplementary table S1. Briefly, the best-fit model involved rs9138 (see online supplementary table S2). In agreement with our a priori hypothesis, rs9138 was significantly associated with radiographic progression over a 2 year follow-up in ACPA-negative patients, as seen by an additive model with a 0.93 fold rate of joint destruction per year per minor allele as compared with the wild-type (p=0.034) (figure 1A-C).

Replication study

We analysed data for 273 ACPA-negative patients in the Leiden EAC cohort and 704 radiographs over a 2 year follow-up. We replicate the contribution of rs9138, as MRA revealed that rs4754 was associated with radiographic progression over the 2 year follow-up with a 0.81 fold rate of joint destruction for each minor allele at any time compared with the wild-type (p=0.019). Analysis of 1,316 radiographs over 7 years of follow-up revealed a persistent effect of rs4754, with a 0.78 fold rate of joint destruction for each minor allele at any time compared with the wild-type (p=0.005; figure 2A).

Combined analysis of early RA during the 2 year follow-up

Data for 631 ACPA-negative patients and 1,664 radiographs were available for the combined analysis of the ESPOIR and Leiden EAC cohorts. MRA over the 2 year follow-up revealed a 0.95 fold rate of joint destruction per year per minor allele compared with the wild-type (p=0.022; figure 2B).

DISCUSSION

The rate of progression of joint damage in RA is highly variable and is associated with the

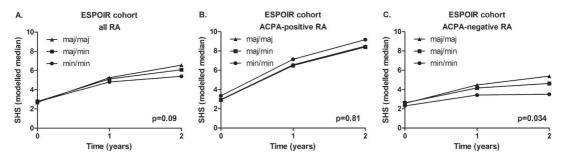


Figure 1. Association of *SPP1* rs9138 and joint damage progression in RA patients in the exploratory study. Multivariate regression analysis modelled median Sharp-van der Heijde scores (SHS) in patients with RA in the ESPOIR cohort (exploratory study) at 1 and 2 year follow-up. (A) Overall RA patients. (B) ACPA-positive RA patients. (C) ACPA-negative RA patients. Maj=major allele; min=minor allele.

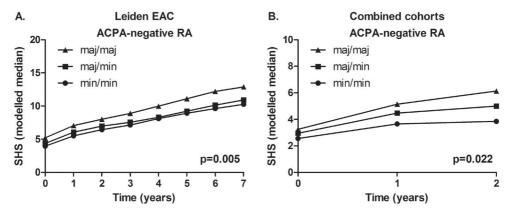


Figure 2. Association of *SPP1* rs4754 and joint damage progression in RA patients in the replication study and combined analysis at 2 year follow-up. (A) Multivariate regression analysis modelled median Sharp-van der Heijde scores (SHS) in patients with RA in the Leiden EAC ACPA-negative cohort during a 7 year follow-up (B) and combined analysis of the ESPOIR and Leiden EAC ACPA-negative cohorts at a 2 year follow-up. Maj=major allele; min=minor allele.

severity of the disease. Genetic variants are estimated to contribute to most of the local variance in RA joint destruction. Several data have suggested that OPN, encoded by *SPP1*, may be involved in bone erosion. In addition, we recently identified *SPP1* as a new RA susceptibility gene, the magnitude of the association being greater in ACPA-negative disease ³. Because ACPA-negative patients fulfilled the ACR modified criteria, *SPP1* may contribute to the variation in joint destruction in this particular subset of the disease.

For the two *SPP1* rs11439060 and rs9138/rs4754 RA risk variants investigated, the rs9138/rs4754 common allele contributed to joint destruction of RA - that is, the minor allele had a protective effect. This finding is in agreement with our a priori hypothesis because the rs9138 common A allele has been identified as an ACPA-negative RA risk allele ³. The replication study, including analyses at the same time of follow-up (2 years) and also after a longer period (7 years), provided evidence that the *SPP1* rs9138 variant contributes to the severity of radiographic damage in ACPA-negative RA in both the early and intermediate course of the disease. Analyses of the combined sets revealed an interaction between *SPP1* rs9138/rs4754 and time at the 2 year follow-up, which suggests a strong effect of *SPP1* on radiographic damage at the early stage of the disease. A recent study of the ESPOIR cohort found that the first-year radiographic progression was a predictor of further progression in early RA, which suggests that, after the early period of the disease, time has a constant effect

Complex diseases, such as RA, invariably involve multiple genes and often exhibit variable symptom profiles. The extent to which disease symptoms, course and severity differ between affected patients may result from underlying genetic heterogeneity. Genes with modifier effects may or may not also influence disease susceptibility. Indeed, *SPP1* seems to act as a susceptibility and a modifier gene. The effect of the rs11439060 variant differed

in this study compared with our previous case-control study ³, the rs11439060-rs9138 risk allele combination not being identified as the best-fit model. Nonetheless, a contribution of rs11439060 could not be definitely excluded; the sample size required to detect such association with a power of 80% would be 1,000 ACPA-negative patients with early RA.

To our knowledge, this is the first report of the identification of a genetic variant associated with joint damage progression in ACPA-negative RA. We took advantage of two cohorts including sequential radiographs of hands and feet, which strengthened the evidence of the contribution of *SPP1* rs9138. Several studies have reported an association of the rs9138 A risk allele with low serum levels of soluble OPN ^{3,17}. However, to date, the exact role of OPN in RA joint damage is controversial: distinct murine models of RA have shown conflicting results on the relevance of OPN in bone erosion pathogenesis ^{18,19}, and, more importantly, OPN blockade was found to be unlikely to induce robust clinical improvement in patients with RA ²⁰.

In conclusion, we have identified and replicated a genetic *SPP1* variant predisposing to joint damage progression in ACPA-negative RA. Further studies of OPN at the protein level are required to better understand the role of this variant in the pathogenesis of the progression of radiographic damage in ACPA-negative RA.

SUPPLEMENTARY DATA

Supplementary data are published on the website of the *Annals of the Rheumatic Diseases*.

REFERENCES

- Klareskog L, Catrina AI, Paget S. Rheumatoid arthritis. The Lancet 2009;373:659–72.
- Van der Woude D, Houwing-Duistermaat JJ,
 Toes RE, et al. Quantitative heritability of anti citrullinated protein antibody-positive and anti citrullinated protein antibody-negative rheumatoid 13.
 arthritis. Arthritis Rheum 2009:60:916–23.
- Gazal S, Sacre K, Allanore Y, et al. Identification of secreted phosphoprotein 1 gene as a new rheumatoid arthritis susceptibility gene. Ann Rheum Dis 2015;74:e19–e19.
- Padyukov L, Seielstad M, Ong RT, et al. A genomewide association study suggests contrasting associations in ACPA-positive versus ACPAnegative rheumatoid arthritis. Ann Rheum Dis 2011;70:259–65.
- Van der Helm-van Mil AH, Huizinga TW, de Vries RR, et al. Emerging patterns of risk factor make-up 16. enable subclassification of rheumatoid arthritis. Arthritis Rheum 2007;56:1728–35.
- Knevel R, Gröndal G, Huizinga TW, et al. Genetic predisposition of the severity of joint destruction in rheumatoid arthritis: a population-based study. Ann Rheum Dis 2012;71:707–9.
- De Rooy DP, Yeremenko NG, Wilson AG, et al. Genetic studies on components of the Wnt signalling pathway and the severity of joint destruction in rheumatoid arthritis. Ann Rheum Dis 2013;72:769–75.
- 8. De Rooy DP, Zhernakova A, Tsonaka R, et al.
 A genetic variant in the region of MMP-9 is
 associated with serum levels and progression of
 joint damage in rheumatoid arthritis. Ann Rheum
 Dis 2014;73:1163–9.
- Knevel R, de Rooy DP, Zhernakova A, et al.
 Association of Variants in IL2RA With Progression of Joint Destruction in Rheumatoid Arthritis.

 Arthritis Rheum 2013;65:1684–93.
- Arnett FC, Edworthy SM, Bloch DA, et al. The American Rheumatism Association 1987 revised criteria for the classification of rheumatoid arthritis. Arthritis Rheum 1988;31:315–24.
- Denhardt DT, Noda M. Osteopontin expression and function: role in bone remodeling. J Cell Biochem Suppl 1998;30-31:92–102.

- Combe B, Benessiano J, Berenbaum F, et al. The ESPOIR cohort: A ten-year follow-up of early arthritis in France: Methodology and baseline characteristics of the 813 included patients. Joint Bone Spine 2007;74:440–5.
- De Rooy DP, van der Linden MP, Knevel R, et al. Predicting arthritis outcomes—what can be learned from the Leiden Early Arthritis Clinic? Rheumatology 2011;50:93–100.
- Van der Heijde D, Boers M, Lassere M.
 Methodological issues in radiographic scoring methods in rheumatoid arthritis. J Rheumatol 1999;26:726–30.
- Knevel R, Tsonaka R, le Cessie S, et al. Comparison of methodologies for analysing the progression of joint destruction in rheumatoid arthritis. Scand J Rheumatol 2013:42:182–9.
- Tobón G, Saraux A, Lukas C, et al. First-Year Radiographic Progression as a Predictor of Further Progression in Early Arthritis: Results of a Large National French Cohort. Arthritis Care Res 2013;65:1907–15.
- 17. D'Alfonso S, Barizzone N, Giordano M, et al. Two single-nucleotide polymorphisms in the 5' and 3' ends of the osteopontin gene contribute to susceptibility to systemic lupus erythematosus. Arthritis Rheum 2005;52:539–47.
- Yumoto K, Ishijima M, Rittling SR, et al.
 Osteopontin deficiency protects joints against destruction in anti-type II collagen antibody-induced arthritis in mice. Proc Natl Acad Sci 2002;99:4556–61.
- Jacobs JP, Pettit AR, Shinohara ML, et al. Lack of requirement of osteopontin for inflammation, bone erosion, and cartilage damage in the K/ BxN model of autoantibody-mediated arthritis. Arthritis Rheum 2004;50:2685–94.
- Boumans MJ, Houbiers JG, Verschueren P, et al. Safety, tolerability, pharmacokinetics, pharmacodynamics and efficacy of the monoclonal antibody ASK8007 blocking osteopontin in patients with rheumatoid arthritis: a randomised, placebo controlled, proof-of-concept study. Ann Rheum Dis 2012;71:180-5.