

Clinical aspects of endogenous hypothyroidism and subclinical hyperthyroidism in patients with differentiated thyroid carcinoma Heemstra, K.A.

Citation

Heemstra, K. A. (2009, September 2). *Clinical aspects of endogenous hypothyroidism and subclinical hyperthyroidism in patients with differentiated thyroid carcinoma*. Retrieved from https://hdl.handle.net/1887/13946

Version:	Corrected Publisher's Version
License:	<u>Licence agreement concerning inclusion of doctoral thesis in the</u> <u>Institutional Repository of the University of Leiden</u>
Downloaded from:	https://hdl.handle.net/1887/13946

Note: To cite this publication please use the final published version (if applicable).

General Introduction

Contents

- I. Introduction
- II. Differentiated Thyroid Carcinoma
- III. Thyroid hormones
- IV. Bone metabolism
- V. Glucose metabolism
- VI. Autonomic nervous system
- VII. Quality of Life
- VIII. D2-Thr-92-Ala and thyroxin dose
- IX. Outline of this thesis

I. Introduction

Differentiated thyroid carcinoma (DTC) is a rare disease with an incidence varying from 2-10/100.000 (1-4). The prevalence of DTC is, however, high because of the good prognosis. In general, 80% of the newly diagnosed tumors are differentiated tumors originating from the epithelial follicular cells. Median age at diagnosis is between 45 and 50 year with a female to male predominance of 2:1 (5).

DTC is associated with an excellent prognosis, with reported 10-year survival rates reaching 90% (6). This is because of a combination of the favorable biological behaviour of the tumor as well as the availability of effective therapy, consisting of total thyroidectomy followed by radioiodine ablation. After initial therapy, all patients with DTC are initially treated with high doses of thyroxin aiming at significantly suppressing thyrotropin (TSH) levels, resulting in a subclinical hyperthyroid state. The rationale of this approach is based on the potential harmful effects of TSH on tumor recurrence (7;8). However, long-term TSH suppression may be associated with potential harmful effects on various systems, including bone metabolism (9-11), glucose metabolism (12-14), the autonomic nervous system (15-18) and quality of life (19-23).

According to protocollized follow up, thyroxin replacement therapy can be transiently stopped in these patients to detect residual or recurrent disease by TSH stimulated thyroglobulin levels. As a result of this standardized procedure, patients become overtly hypothyroid within 4-6 weeks. This may reversely affect the systems influenced by subclinical hyperthyroidism, mentioned above.

DTC patients are an unique model to study the metabolic effects of thyroid hormone, both depletion and excess, on physiological systems, because these DTC patients are treated with total thyroidectomy and therefore don't produce any endogenous thyroid hormones. Thyroid hormone levels are well documented in these patients and can be exactly regulated by changing the thyroxin dosages. During clinical follow-up, patients are sometimes withdrawn from thyroxin, which creates a state of controlled hypothyroidism, whereas many patients will be treated with TSH suppressive dosages of thyroxin, thereby creating a state of subclinical hyperthyroidism. Moreover, there is no interfering effect from thyroid disease, like in patients substituted with thyroxin for autoimmune thyroid disease.

In this introductory chapter a general overview of DTC, thyroid hormones and the clinical consequences of exogenous subclinical hyperthyroidism and thyroxin withdrawal will be provided and the questions addressed in this thesis will be introduced.

II. Differentiated thyroid carcinoma

Pathogenesis

Genetic alterations are involved in the pathogenesis of thyroid carcinoma. The analysis of these genetic alterations is important not only for the diagnosis of DTC, but also for the understanding of the pathophysiology of thyroid disorders(24-26). Mutations in one of the three RAS-genes are frequently found in follicular adenomas and carcinomas. Benign hyperfunctioning nodules or adenomas are associated with mutations in the GSP and TSH receptor genes.

The recent identification of mutations in B-RAF, which are present in 40-60 % of papillary thyroid carcinomas (PTC), has improved the understanding of the molecular pathogenesis of PTC. B-RAF is a component of the RET RAS RAF cascade that activates MAP kinase. Almost all patients with PTC have rearrangements and mutations of B-RAF, RAS, RAF and TRK (neutrotrophic tyrosine kinase receptor). Translocations of RET, that are found in DTC, give rise to a chimeric protein consisting of an activated RET tyrosine kinase domain (24;27-

42). Transcriptional and post-transcriptional mechanisms are thought to regulate MET overexpression as a secondary effects (43).

The genetic pathogenesis of follicular thyroid carcinoma (FTC) is less clear. However, it was found that FTC is related with rearrangements in PAX8 and PPAR- γ genes, which are traditionally associated with thyroid development (PAX 8) and cell differentiation and metabolism (PPAR- γ) (44). The chimeric protein acts as a dominant negative competitor for PPAR- γ . A downregulation of the PPAR- γ signaling route has been observed in experimental models of DTC (45).

The genetic alterations that are involved in the pathogenesis of DTC, result in proliferation by multiple pathways and the loss of thyroid specific proteins. Thyroid peroxidase (TPO) is believed to disappear in an early phase, followed by the disappearance of NIS.

Diagnosis

Fine needle aspiration (FNA) is the procedure of choice in patients presenting with thyroid nodules. The sensitivity of FNA is 90-95 %. The specificity of FNA is lower, 60-80%, when all patients with a non-benign FNA are referred for surgery (46). The distinction between benign and malignant follicular tumors is difficult to make by FNA, because the essential criterion for FTC is capsular invasion which can not be determined by cytology. Another problem is the differentiation between follicular adenoma and follicular variant of papillary thyroid carcinoma (FVPTC), because the essential criterion is the aspect of the nuclei. As a consequence, the frequency of FTC in hemi-thyroidectomies performed after suspicious outcome from FNA is only 20-30%.

The Tumor-Node-Metastases classification system is based on pathologic findings. This classification system divides patients into four stages, with progressively poorer survival with increasing stage. Recently, the 6th edition of the TNM system has become available (47). The most important difference with he 5th edition is the fact that the dimension of T1 has been extended to 1.5 cm and that tumors with limited extrathyroidal extension are designated T3 instead of T4, which has implications for the prognosis of DTC (48). Therefore, some experts propagate to continue the use of the 5th edition. In the studies in his thesis the 5th edition of the TNM staging system is used (49).

то	No evidence of primary tumor
T1	Tumor 1 cm or less in greatest dimension
T2	Tumor > 1 cm, but nor more then 4 cm in greatest dimension, limited to the thyroid
ТЗ	Tumor > 4 cm in greatest dimension limited to the thyroid
T4	Tumor of any size, beyond the thyroid capsule
Nx	Regional lymph nodes (cervical and upper mediastinum) cannot be assessed
NO	No regional lymph node metastases
N1	Regional lymph node metastases
N1a	Metastasis in ipsilateral cervical lymph node(s)
N1b	Metastasis in bilateral, midline or contralateral cervical or mediastinal lymph node(s)
Mx	Distant metastasis cannot be assessed
MO	No distant metastasis
M1	Distant metastasis

Figure 1. TNM classification system 5th edition, AJJC, Adapted from (50)

Initial therapy

Initial therapy for DTC consists of near-total thyroidectomy followed by radioiodine ablation. There is still some controversy about the extent of thyroid surgery. However, there are strong arguments in favor of total or near-total thyroidectomy in all patients (51). Only very low-risk patients (T1 (< 1 cm) NOMO (5th edition) DTC, unifocal) may be treated by hemi-thyroidectomy.

In tumor stages of T2 and higher a total thyroidectomy is indicated (52-54). Near-total thyroidectomy results in lower recurrence rates than more limited thyroidectomy because many papillary tumors are multifocal and bilateral (55;56). In addition, total thyroidectomy facilitates total ablation with iodine-131 and reveals a higher specificity of thyroglobulin (Tg) as a tumor marker (52-55). Complications of total thyroidectomy are laryngeal nerves palsy in 2 % of DTC patients and hypoparathyroidism. The latter occurs in 1/3 of patients after total thyroidectomy, but persists longer then 3 months in only 2 % (50).

Controversy also exists about the routine use of iodide-131 ablation of thyroid remnants. However, many clinics give postoperative iodide-131 ablation for three reasons. First, iodide-131 destroys any remaining normal thyroid tissue thereby increasing the specificity of detectable serum Tg levels and positive whole-body scintigraphy indicating persistent or recurrent disease (5;54;57). Second, iodide-131 may destroy occult microscopic carcinomas, thereby decreasing the risk of recurrence thyroid carcinoma (8;54;58;59). Third, the use of large amounts of iodide-131 for therapy permits post ablative scanning to detect recurrent disease (60;61). A meta-analysis showed that the use of iodide-131 to prevent recurrence or death is uncertain (62). A beneficial effect is probably only present in patients with high risk or irradical surgery (8;53;63;64). Many authors are more careful advising I-131 ablation since various papers reported a relation between I-131 therapy and non-thyroid carcinoma (65-67).

In patients with a very low risk of recurrence/mortality (T1 (<1 cm) NOMO unifocal) I-131 is not indicated. I-131 ablation is still the treatment of choice in patients with a high risk of recurrence/mortality 1) T3 or T4, 2) any T N1, and 3) Any T M1, and incomplete tumor resection (68;69). Controversy exists about patients with a low risk (T1 (>1 cm)NOMO, T2NOMO or T1(<1 cm)NOMO multifocal) of recurrence/mortality (50).

After initial therapy, all patients with DTC are treated with high doses of thyroxin aiming at significantly suppressing thyrotropin (TSH<01 mU/L) levels. The rationale of this approach is based on the potential harmful effects of TSH on tumor recurrence (7;8). One study demonstrated a preventive effect of TSH suppression on tumor recurrence or progression only in high risk DTC patients (70). However, long-term TSH suppression may be associated with potential harmful effects on various systems including bone metabolism, glucose metabolism, the autonomic nervous system and quality of life. The recent European Consensus on thyroid cancer (71), recommended that not all patients with DTC should be indiscriminately treated with TSH suppressive therapy because this represents in effect a state of subclinical hyperthyroidism, as defined by suppressed serum TSH levels (below 0.4 mU/I), in the presence of normal serum levels of (free) thyroxin. A recent analysis of our institution showed that TSH levels are positively associated to thyroid carcinoma related death and relapse (72). This effect became apparent at TSH levels above 2 mU/L and is in line with other studies (73).

Follow-up

The purpose of follow-up protocols in DTC is the early detection of tumor recurrence or metastatic disease in order to optimize additional treatment. Most patients during follow up have been cured definitely, and, as a consequence, have a low pre-test probability for recurrent disease. Therefore, the sensitivity of the diagnostic test must be adequate to detect the few patients with evident thyroid carcinoma, whereas specificity must also be high to avoid unnecessary treatments in patients without recurrent disease. In addition, the burden of diagnostic tests for the patient should be kept at a minimum.

a. Thyroglobulin

Thyroglobulin (Tg) is produced by normal or neoplastic thyroid follicular cells and Tg production is stimulated by TSH. In patients treated with a total thyroidectomy and I-131 ablation, Tg should be undetectable. The clinical interpretation of Tg is hampered by the

following analytical problems:

1. lack of universal standardization of the Tg assays, which results in considerable interassay variability (74),

2. a high intra-assay variability, which results in a poor comparability of results obtained within one patient during follow-up,

3. "hook" effects may be present, which affect IMA methods in particular and can lead to inappropriately low- or normal range Tg values in sera with very high serum Tg concentrations, 4. the presence of Tg auto-antibodies that can lead to lower or higher Tg levels.

Despite these analytical problems, Tg measurements are still the basis in the follow-up in DTC. Several studies have been performed on the diagnostic value of Tg measurements. The interpretation of these studies is difficult, because 1. heterogeneous patient groups with respect to initial therapy are included, 2. the time points of Tg measurements after diagnosis are not clearly indicated, and 3. fixed Tg cut-off levels are used, without receiver operator curve (ROC) analyses. The application of ROC data is essential, as a chosen cut-off level is a subjective choice based on the balance between a desired percentage of missed recurrences versus unnecessary therapies. Therefore, in the recent European consensus paper, it was recommended to define institutional Tg cut-off levels (71). In addition, most studies provide data on the diagnostic value of Tg for tumor presence, but do not give data on the *prognostic* significance for recurrence or death. The few studies that were published on the prognostic significance of Tg measurements used fixed cut-off levels, contained selected subgroups of patients, and included either Tg measurements at one time point or at undefined time points (75-79).

We, therefore, performed a study on the diagnostic and prognostic value of Tg in a homogeneous group of DTC patients with respect to initial therapy, using Tg measurements at 5 defined time-points after diagnosis, in combination with ROC analyses (chapter 2).

b. Thyroxin withdrawal versus rhTSH

Serum Tg measurements, I-131 ablation and diagnostic I-131 whole body scans are based on the responsiveness of DTC to TSH (80). TSH stimulated Tg measurements have superior diagnostic value in DTC compared to Tg measurements on thyroxin replacement therapy (81). High serum levels can be achieved by thyroxin withdrawal or injection with recombinant human TSH (rhTSH), which has less impact on quality of life (82). rhTSH is an adequate method to detect recurrence or metastases (78;83-85). A rhTSH stimulated Tg level greater than 2 mg/ml predicts persistent disease (78;83;86), whereas a rhTSH stimulated Tg level lower than 0.5 mg/dl has a 98 % likelihood of detecting patients free of tumor (78). Whole body scans performed after rthTSH-injections have a similar sensitivity and negative predictive value compared to thyroxin withdrawal (83-85). However, more negative whole body scans were found after rhTSH-injections compared to thyroxin withdrawal (83-85). The sensitivity and negative predictive value of Tg values after rhTSH-injections are 96.3 % and 99.5 % respectively by combining these measurements with a neck ultrasound (87).

Several studies have reported that radioiodine ablation of thyroid remnants after rhTSHinjections is as effective as ablation after thyroxin withdrawal (88;89). Radioiodine ablation after rhTSH-injections in patients with recurrence or distant metastases results in a beneficial effect in 75 % of patients (90;91). However, rhTSH has not been approved for this indication.

c. I-131 scintigraphy, Ultrasound, and FDG-PET

The result of iodine-131 whole body scanning depends on the presence and the ability of thyroid-cancer tissue to accumulate iodine-131 in the presence of high serum TSH concentrations. Diagnostic Ral whole body scintigrapies have a much lower sensitivity than ultrasound and Tg measurements. Therefore, the routine use of Ral scintigraphy in

General introduction

01

the diagnostic follow-up of DTC patients is no longer recommended (87;92). Ultrasound combined with FNA had the highest sensitivity (even higher than Tg) for local recurrence and lymph node metastases in recent papers (87;93;94). Thus, ultrasound has an important place in he follow up of DTC. 18-F Fluorodeoxyglucose-positron emission tomography (FDG-PET) may be useful in patients with elevated serum Tg levels, in whom no Ral uptake is observed after diagnostic or post-therapeutic scintigraphy. The sensitivity of FDG-PET is increased with elevating serum Tg levels and after TSH stimulation (95). Robbins *et al* showed that FDG-PET positivity is associated with worse survival (96).

III. Thyroid hormones

The production of thyroid hormones by the thyroid is regulated by the hypothalamus-pituitarythyroid axis. Thyrotropin releasing hormone (TRH), which is produced by the hypothalamus, stimulates the secretion of thyrotropin (TSH) by the anterior pituitary. TSH promotes the thyroid to synthesize the prohormone tetraiodothronine (T4) in the thyroid. Iodide is actively taken up by the thyroid gland by the sodium-iodide-symporter (NIS) at the basolateral plasma membrane. The expression and activity of NIS are controlled by TSH. Thyrogobulin, which is synthesized by the follicular cells, is then iodinated with one or two iodides to form monoiodotyrosine (MIT) or diiodotyrosine (DIT). This process is catalyzed by the enzyme thyroid peroxidase (TPO). Two DIT molecules are then coupled to form T4 and one DIT and one MIT molecule are coupled to form T3. The thyroid secretes approximately 90 % T4, 10 % triiodothyronine (T3) and less then 1 % reverse T3. The T3 molecule is the active form of thyroid hormone. The majority of the active form of thyroid hormone T3 is derived from conversion of T4 to T3 in peripheral tissues, such as the liver (see deiodinases). T4 and T3, in turn, have a negative effect on the TRH secretion by the hypothalamus and TSH secretion by the pituitary. lodide is important for the synthesis of thyroid hormones.

Deiodinases

Peripheral thyroid metabolism is mainly regulated by the iodothyronine deiodinases D1, D2 and D3 (97;98). D1 convertes the prohormone T4 in T3, plays a role in the breakdown of rT3 (97;99) and is expressed in liver, kidney, thyroid and pituitary and at lower levels in other tissues as skeletal muscle, spleen and lung. D2 is essential for the production of T3 through outer ring deiodination of T4. It is present in brain, skeletal muscle, thyroid, pituitary, brown adipose tissue (BAT) and aortic smooth muscle cells (97;100-104). D3 inactivates T3 and prevents T4 activation by innerring deiodination (98) and is present in brain, skin, placenta and fetal tissues (97).

The deiodinases adjust the thyroid hormone levels of individual tissues in response to various conditions. The peripheral conversion of T4 to T3 is increased during *hypothyroidism* (97;105;106). Extrathyroidal T3 production changes from PTU sensitive to PTU insensitive during hypothyroidism in rats, representing an increase in the conversion of T4 to T3 by D2 and a decreased conversion by D1 (107). D1 gene transcription is decreased in liver and kidney during hypothyroidism (108), which is related to the presence of two T3 response elements in the human D1 gene (97;108-110). Thyroid status regulates D2 activity both at the pre- and posttranslational level. D2 activity is increased in different tissues predominantly during hypothyroidism by a decrease in substrate (T4)-induced degradation of D2 protein (97;111-113). Hypothyroidism elevates D2 mRNA in rat brain and BAT (97;100;114;115). D2 mRNA expression and activity were found in skeletal muscle samples from healthy subjects (103;116). This is fascinating, because D2 could therefore play a role in peripheral and intracellular T3 production (103). Maia *et al.* reported that D2 is a major source of T3 during euthyroidism and could therefore play an important role during hypothyroidism

Thyroxine, T4

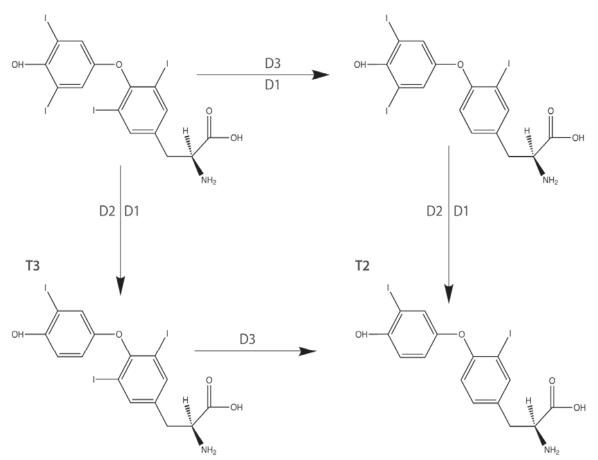


Figure 1. Structure of the iodothyronines and their activation and inactivation by iodothyronine deiodinase.

(117). As patients treated for DTC have no thyroid tissue left, we hypothesized that during hypothyroidism D2 in skeletal muscle could be essential in promoting the conversion of T4 to T3 (chapter 3).

Several polymorphisms in D2 have been described (118-120), with most studies investigating the consequences of the D2-Thr92Ala polymorphism. This D2-Thr92Ala polymorphism has been associated with BMI and insulin resistance in obese subjects and type 2 diabetes mellitus (118;119), although this was not confirmed in another study (121). The maximal velocity of D2 in vitro in thyroid and skeletal muscle of homozygous carriers of the Ala92 allele was decreased by 3–10-fold (118).

IV. Bone metabolism

Thyroid hormone impacts on bone metabolism, ranging from decreased skeletal development in childhood hypothyroidism to an increased risk for osteoporosis in hyperthyroidism (11;122;123). Thyroid hormone indirectly promotes osteoclast formation and activation by inducing the expression of cytokines, prostaglandins and the receptor activator of nuclear factor NF- κ B ligand (RANKL) (124-126). RANKL, the key molecule in osteoclast differentiation, binds to its receptor, RANK, which is expressed on dendritic cells, T cells, osteoclast precursors and mature osteoclasts (127;128). RANKL promotes the survival of RANK positive T cells (127), stimulates osteoclast differentiation (129-133), increases the activity of mature osteoclasts (130;134;135) and stimulates survival of osteoclasts by preventing apoptosis (135). Contact with stromal cells and M-CSF also promotes osteoclast differentiation (136;137). Thyroid hormone inhibits chondrocyte proliferation and promotes hypertrophic differentiation, mineralization, matrix synthesis but also apoptosis of chondrocytes in the growth plate.

Overt hyperthyroidism results in an increased risk for osteoporosis (123), the pathophysiology of which is multifactoral (124), including shortening of the bone remodelling cycle (138) and acceleration of bone turnover (139). The effects of *subclinical hyperthyroidism* on bone metabolism are not clear. Several studies have addressed this issue, but there is no consensus largely because of differences in study design, including patient groups, methodology used, follow-up time and choice of outcome parameters. To study the effects of subclinical hyperthyroidism on bone mineral density, we performed a systematic review including all clinical studies on TSH suppressive thyroxin therapy in thyroid cancer patients (chapter 4).

An interesting development has been the discovery of the *TSH* receptor (TSHR) in bone (140-142). TSHR knockout and haploinsufficient mice with normal thyroid hormone levels have decreased bone mass suggesting that TSH might directly influence bone remodeling (141;143;144). This is intriguing, because effects on bone metabolism that were previously ascribed to high thyroid hormone levels could also be attributed to suppressed TSH levels (143-145). Abe et al. suggested that TSH inhibits osteoclast formation and survival by attenuating JNK/c-jun and NFkB signaling in response to RANK-L and inhibits osteoblast differentiation and type 1 collagen expression as well by downregulating Wnt and VEGF signaling (141). The same group found also that TSH directly inhibits Tumour Necrosis Factor- α (TNF- α) production and that TNF- α is the critical cytokine mediating the downstream antiresorptive effects of TSH on the skeleton (146). Other studies suggest that serum TSH activates the type 2 deiodinase in osteoblasts, thereby linking TSH and increased local thyroid hormone availability (142). Furthermore, in animal studies, low doses of TSH increased bone volume and improved microarchitecture in ovariectomized rats (147), without increasing serum thyroid hormone levels.

It was recently reported that the TSHR-Asp727Glu polymorphism was associated with 2.3% higher BMD in elderly carriers (148). Although the functional consequences of this polymorphism are debated (149), the lower plasma TSH levels in patients carrying the polymorphism could point toward a higher sensitivity of the variant compared to the wild-type TSHR (150;151).

We, therefore, evaluated the independent relation between serum TSH levels and indicators of bone turnover in thyroidectomized patients for differentiated thyroid carcinoma receiving thyroid hormone substitution (chapter 5). In addition, we studied the relationship between the TSHR-Asp727Glu polymorphism and bone as these subjects are not expected to show compensatory lower serum TSH levels if they carry the TSHR-Asp727Glu polymorphism (150;151).

The consequences of *hypothyroidism* on bone metabolism are not clear. Various studies report decreased bone resorption (152-155) or bone formation (152), whereas other studies document no impact on bone turnover (156-158). Furthermore, it is not clear if the effects of hypothyroidism must be attributed to the increased TSH levels or decreased thyroid hormone levels. As mentioned above, TSHR knockout and haploinsufficient mice with normal thyroid hormone levels have decreased bone mass, suggesting that TSH might directly influence bone remodeling (141;143;144). However, other studies question the role of TSH in bone metabolism (159;160). Three studies in humans have investigated the effect of TSH on bone metabolism, but their results were not consistent showing either no impact on bone turnover (161), increased bone formation (162;163) or decreased bone resorption (163).

To document the effects of hypothyroidism on bone metabolism and to discriminate between effects mediated by decreased thyroid hormone levels *versus* those mediated by increased TSH levels, we studied bone metabolism in eleven patients with differentiated thyroid carcinoma (DTC) during short-term thyroxin withdrawal and compared with eleven age-, gender- and BMI-matched DTC patients with increased TSH levels and normal thyroid hormone levels due to rhTSH injections (chapter 6).

Although earlier studies on the role and functional expression of iodothyronine deiodinase enzymes in the skeleton have not revealed unequivocal answers (142;164-167), a recent study reported normal growth in mice with deficiencies in D1 and D2 indicating that D2 may not be critical in skeletal development (168). This was supported by another study, which found that D2 activity is restricted to mature osteoblasts, suggesting a possible role for D2 in mature osteoblast function (169). Because it is difficult to study the role of D2 *per se* on skeletal metabolism in humans, we choose to study the effects of functional D2 polymorphisms on BMD and indicators of bone turnover. Canani *et al.* (118) reported that the maximal velocity of D2 *in vitro* in thyroid and skeletal muscle of homozygous carriers of the Ala⁹² allele was decreased by 3-10-fold. We, therefore, studied the relationship between the functional D2-Thr92Ala polymorphism, BMD and indicators of bone turnover (chapter 7).

V. Glucose metabolism

Thyroid hormone has effects on glucose- and lipid metabolism (13;170). There is a relation between serum thyroid hormone levels and basal and insulin-mediated glucose metabolism in euthyroid subjects with preserved thyroid function (171-173). It has been suggested that T3 regulates insulin response after glucose ingestion in humans (174).

Hyperthyroidism has been associated with impaired glucose tolerance and increased insulin resistance (175-181), predominantly at the level of the liver (182). The pathophysiology has not been completely elucidated, but it has been ascribed to a combination of multiple factors, including diminished pancreatic secretion of insulin (183;184), diminished suppression of glucagon by glucose (185) and increased adrenergic activity (186).

Limited data are available on the consequences of *subclinical hyperthyroidism* on glucoseand lipid metabolism. This issue has been studied only by Yavuz *et al.*, who reported a decreased insulin sensitivity index by oral glucose tolerance test in patients with exogenous subclinical hyperthyroidism compared to values after restoration of euthyroidism and compared to controls (187). Regarding lipid metabolism, most studies report no differences in lipid profiles during subclinical hyperthyroidism (188-190), with the exception of 2 studies, that observed decreased total and LDL cholesterol levels (191;192). Franklyn *et al.* reported decreased total cholesterol concentrations only in patients older than 55 years and LDL cholesterol levels were decreased only in patients older than 65 years (193). We therefore performed a prospective placebo-controlled randomized trial to investigate the effects of restoration of exogenous subclinical hyperthyroidism to euthyroidism on glucose- and lipid metabolism (chapter 8).

VI. Autonomic nervous system

The consequences of *hyperthyroidism* on the heart are profound, including tachycardia and/or arrhythmias, increased systolic pressure, increased systolic function, left ventricular hypertrophy and diastolic dysfunction (194-196). It is suggested that these effects are the

Result of direct effects of thyroid hormone on the cardiovascular system and the interaction of thyroid hormones with the sympathetic nervous system (195;197). Hyperthyroidism is associated with a sympathicovagal imbalance, characterized by increased sympathetic activity in the presence of reduced vagal tone, which corresponds with increased urinary excretion of catecholamines (15;16;198). Therefore, the current consensus is that manifestations of altered autonomic nervous system function play a role in the pathophysiology and clinical presentation of thyrotoxicosis.

During subclinical hyperthyroidism, cardiovascular effects may also occur, but these are less well known and seemingly less severe. Regular findings during subclinical hyperthyroidism include increased heart rate, supraventricular arrhythmias and abnormalities of LV morphology and function (195;199-201). The consequences of subclinical hyperthyroidism on the autonomic nervous system function are less well defined. Several studies, using measures of heart rate variability, found evidence that in patients with endogenous subclinical hyperthyroidism a reduction of cardiac parasympathetic control is present (18), (200), (202). This is supported by findings on heart rate turbulence by Osman *et al* (203). However, in the study of Goichot (18) no differences in the ratio of low frequency power over high frequency power (LF/HF) were reported in these patients. The LF/HF ratio is commonly used to characterize the balance between vagal and sympathetic influences. To further clarify this issue, we performed a prospective, randomized, placebo-controlled study using heart rate variability to assess the autonomic nervous system in patients with DTC with longer than 10 years exogenous subclinical hyperthyroidism and investigated whether restoration to euthyroidism affects autonomic nervous function (chapter 9).

Hypothyroidism is associated with bradycardia, mild diastolic hypertension, increased peripheral cardiovascular resistance (194;204;205), decreased cardiac output and diastolic dysfunction (194;204;206;207). Hypothyroidism also induces sympathovagal imbalance (17;208;209). Nevertheless, current literature shows inconsistent results with either an increased sympathetic activity (17), a decreased sympathethic modulation (208) or an increased vagal tone (209). We therefore investigated the effects of short-term overt hypothyroidism, 4 weeks after thyroxin withdrawal, and restoration to subclinical hyperthyroidism on the autonomic nerves system (chapter 10).

VII. Quality of life

DTC is associated with an excellent prognosis. This may imply that quality of life in cured DTC patients may be quite normal. However, patients are treated long-term with TSH suppressive thyroxin replacement therapy, reflecting in effect a state of *subclinical hyperthyroidism*, which may impact quality of life (210-212).

Quality of life in cured DTC patients is investigated in a few studies (20;21;213-215). However, these studies are limited by small patient numbers(21;213), limited number of quality of life questionnaires (20;215) or the absence of a healthy control group (20;213;214).

Studies reporting the relation between the level of TSH suppression and quality of life in DTC patients are inconclusive because of small patient numbers, selection of patients with symptoms of hyperthyroidism or selection of patients with a long duration of cure (210;216). For that reason, we investigated quality of life in a large cohort of cured DTC patients compared to controls matched for age, gender and socioeconomic status. In addition, the determinants of quality of life, including serum TSH levels were investigated (chapter 11).

Thyroxin withdrawal resulting in overt *hypothyroidism* may also impact quality of life. It results in fatigue, anorexia, constipation, problems with motor skills and fluid retention. Quality of life during thyroxin withdrawal is also affected by a decreased motivation, productivity and quality of work and by interfering with family and social life (22). In addition, a decreased psychomotor function and an increased fear are reported during thyroxin withdrawal (19;217).

VIII. D2-Thr-92-Ala and thyroxin dose

Several polymorphisms in D2 have been described (118;119;218;219). The functional implications of the *D2-Thr92Ala* polymorphism are inconclusive. One in vitro study found an association with a decreased D2 activity (118) whereas another study found no difference (219). So far no associations between the D2-Thr92Ala polymorphism and serum thyroid hormone levels were documented (151;218;220). A study of Torlontano *et al.* documented that homozygous carriers of the D2-Ala92 allele needed higher dosages of thyroxin in thyroidectomized differentiated thyroid carcinoma (DTC) patients, particular in the group with near-suppressed TSH levels (TSH values between 0.1 and 0.5 mU/L)(221). However, this study had limitations, because actual values of serum TSH levels for wild-type and homozygous groups within the near-suppressed TSH group were not given. It is, therefore, unclear whether TSH levels in both groups were indeed identical, which would be a key finding to ascribe the slight differences in thyroxin dose indeed to the polymorphism. The fact that serum T4 and T3 levels did not differ between the wild-type group and D2-Thr92Ala homozygotes is also remarkable. We, therefore, studied the association between the D2-Thr92Ala polymorphism, thyroid hormone levels and thyroxin dosage (chapter 12).

IX. Outline of this thesis

In chapter 2, we describe the diagnostic and prognostic value of thyroglobulin (Tg) in a homogeneous group of differentiated thyroid carcinoma (DTC) patients with respect to initial therapy, using Tg measurements at 5 defined time-points after diagnosis, in combination with ROC analyses.

In the continuation this thesis, questions about the clinical consequences of exogenous subclinical hyperthyroidism and hypothyroidism on bone metabolism, glucose metabolism, the autonomic nervous system and quality of life in patients with DTC are addressed.

Chapter 3 evaluates the D2 activity and expression of deiodinases 1, 2 and 3 in skeletal muscle samples in DTC patients both during hypothyroidism and thyroxin replacement therapy.

Chapter 4 shows the results of a systematic review describing the effects of TSH suppressive thyroxin therapy on bone mineral density in DTC patients.

In chapter 5, we evaluate the independent relation between serum TSH levels and indicators of bone turnover in DTC patients receiving thyroid hormone substitution.

In chapter 6 we describe a prospective study to investigate the effects of hypothyroidism on bone metabolism and to discriminate between potential effects mediated by decreased thyroid hormone levels versus those mediated by increased TSH levels.

Chapter 7 presents the relationship between the functional D2-Thr92Ala polymorphism, BMD and indicators of bone turnover.

In chapter 8, we investigate the effects of restoration of exogenous subclinical hyperthyroidism to euthyroidism on glucose- and lipid metabolism in a prospective, randomised, placebocontrolled trial.

Chapter 9 describes a prospective, randomized, placebo-controlled study to assess autonomic nervous function in patients with DTC with longer than 10 years exogenous subclinical hyperthyroidism and to investigate whether restoration to euthyroidism affects autonomic nervous function.

In chapter 10 we show the effects of short-term overt hypothyroidism, 4 weeks after thyroxin withdrawal, and restoration to subclinical hyperthyroidism on the autonomic nervous system.

Chapter 11 describes quality of life in a large cohort of cured DTC patients compared to

controls matched for age, gender and socioeconomic status. In addition, the determinants of quality of life, including serum TSH levels were investigated.

In chapter 12, we studied the association between the D2-Thr92Ala polymorphism and thyroid hormone levels and thyroxin dosage

References

- 1. Kuijpens JL, Hansen B, Hamming JF, Ribot JG, Haak HR, Coebergh JW. Trends in treatment and longterm survival of thyroid cancer in southeastern Netherlands, 1960-1992. Eur J Cancer 1998; 34(8):1235-1241.
- Smit JW, Schroder-van der Elst JP, Karperien M et al. lodide kinetics and experimental (131)I therapy in a xenotransplanted human sodium-iodide symporter-transfected human follicular thyroid carcinoma cell line. J Clin Endocrinol Metab 2002; 87(3):1247-1253.
- 3. Sakoda LC, Horn-Ross PL. Reproductive and menstrual history and papillary thyroid cancer risk: the San Francisco Bay Area thyroid cancer study. Cancer Epidemiol Biomarkers Prev 2002; 11(1):51-57.
- 4. Burrow GN, Burke WR, Himmelhoch JM, Spencer RP, Hershman JM. Effect of lithium on thyroid function. J Clin Endocrinol Metab 1971; 32(5):647-652.
- 5. Schlumberger MJ. Papillary and follicular thyroid carcinoma. N Engl J Med 1998; 338(5):297-306.
- 6. Hundahl SA, Fleming ID, Fremgen AM, Menck HR. A National Cancer Data Base report on 53,856 cases of thyroid carcinoma treated in the U.S., 1985-1995 . Cancer 1998; 83(12):2638-2648.
- 7. Goretzki PE, Frilling A, Simon D, Roeher HD. Growth regulation of normal thyroids and thyroid tumors in man. Recent Results Cancer Res 1990; 118:48-63.
- 8. Mazzaferri EL, Jhiang SM. Long-term impact of initial surgical and medical therapy on papillary and follicular thyroid cancer. Am J Med 1994; 97(5):418-428.
- 9. Faber J, Galloe AM. Changes in bone mass during prolonged subclinical hyperthyroidism due to Lthyroxine treatment: a meta-analysis. Eur J Endocrinol 1994; 130(4):350-356.
- 10. Giannini S, Nobile M, Sartori L et al. Bone density and mineral metabolism in thyroidectomized patients treated with long-term L-thyroxine. Clin Sci (Lond) 1994; 87(5):593-597.
- 11. Heemstra KA, Hamdy NA, Romijn JA, Smit JW. The effects of thyrotropin-suppressive therapy on bone metabolism in patients with well-differentiated thyroid carcinoma. Thyroid 2006; 16(6):583-591.
- 12. Dimitriadis G, Mitrou P, Lambadiari V et al. Insulin action in adipose tissue and muscle in hypothyroidism. J Clin Endocrinol Metab 2006; 91(12):4930-4937.
- 13. Dimitriadis GD, Raptis SA. Thyroid hormone excess and glucose intolerance. Exp Clin Endocrinol Diabetes 2001; 109 Suppl 2:S225-S239.
- 14. Yavuz DG, Yuksel M, Deyneli O, Ozen Y, Aydin H, Akalin S. Association of serum paraoxonase activity with insulin sensitivity and oxidative stress in hyperthyroid and TSH-suppressed nodular goitre patients. Clin Endocrinol (0xf) 2004; 61(4):515-521.
- 15. Burggraaf J, Tulen JH, Lalezari S et al. Sympathovagal imbalance in hyperthyroidism. Am J Physiol Endocrinol Metab 2001; 281(1):E190-E195.
- 16. Cacciatori V, Bellavere F, Pezzarossa A et al. Power spectral analysis of heart rate in hyperthyroidism. J Clin Endocrinol Metab 1996; 81(8):2828-2835.
- 17. Cacciatori V, Gemma ML, Bellavere F et al. Power spectral analysis of heart rate in hypothyroidism. Eur J Endocrinol 2000; 143(3):327-333.
- 18. Goichot B, Brandenberger G, Vinzio S et al. Sympathovagal response to orthostatism in overt and in subclinical hyperthyroidism. J Endocrinol Invest 2004; 27(4):348-352.
- Botella-Carretero JI, Galan JM, Caballero C, Sancho J, Escobar-Morreale HF. Quality of life and psychometric functionality in patients with differentiated thyroid carcinoma. Endocr Relat Cancer 2003; 10(4):601-610.
- 20. Crevenna R, Zettinig G, Keilani M et al. Quality of life in patients with non-metastatic differentiated thyroid cancer under thyroxine supplementation therapy. Support Care Cancer 2003; 11(9):597-603.
- 21. Dagan T, Bedrin L, Horowitz Z et al. Quality of life of well-differentiated thyroid carcinoma patients. The J Laryngol Otol. 2004; 118(7):537-542.
- 22. Dow KH, Ferrell BR, Anello C. Quality-of-life changes in patients with thyroid cancer after withdrawal of thyroid hormone therapy. Thyroid 1997; 7(4):613-619.
- Duntas LH, Biondi B. Short-term hypothyroidism after Levothyroxine-withdrawal in patients with differentiated thyroid cancer: clinical and quality of life consequences. Eur J Endocrinol 2007; 156(1):13-19.
- 24. Fagin JA. How thyroid tumors start and why it matters: kinase mutants as targets for solid cancer pharmacotherapy. J Endocrinol 2004; 183(2):249-256.
- 25. Soares P, Sobrinho-Simoes M. Recent advances in cytometry, cytogenetics and molecular genetics of

thyroid tumours and tumour-like lesions. Pathol Res Pract 1995; 191(4):304-317.

- 26. Sobrinho-Simoes M, Preto A, Rocha AS et al. Molecular pathology of well-differentiated thyroid carcinomas. Virchows Arch 2005; 447(5):787-793.
- 27. Cinti R, Yin L, Ilc K et al. RET rearrangements in papillary thyroid carcinomas and adenomas detected by interphase FISH. Cytogenet Cell Genet 2000; 88(1-2):56-61.
- 28. Corvi R, Berger N, Balczon R, Romeo G. RET/PCM-1: a novel fusion gene in papillary thyroid carcinoma. Oncogene 2000; 19(37):4236-4242.
- 29. Fukushima T, Suzuki S, Mashiko M et al. BRAF mutations in papillary carcinomas of the thyroid. Oncogene 2003; 22(41):6455-6457.
- Greco A, Pierotti MA, Bongarzone I, Pagliardini S, Lanzi C, Della PG. TRK-T1 is a novel oncogene formed by the fusion of TPR and TRK genes in human papillary thyroid carcinomas. Oncogene 1992; 7(2):237-242.
- 31. Grieco M, Santoro M, Berlingieri MT et al. PTC is a novel rearranged form of the ret proto-oncogene and is frequently detected in vivo in human thyroid papillary carcinomas. Cell 1990; 60(4):557-563.
- 32. Klugbauer S, Jauch A, Lengfelder E, Demidchik E, Rabes HM. A novel type of RET rearrangement (PTC8) in childhood papillary thyroid carcinomas and characterization of the involved gene (RFG8). Cancer Res 2000; 60(24):7028-7032.
- 33. Nikiforov YE, Rowland JM, Bove KE, Monforte-Munoz H, Fagin JA. Distinct pattern of ret oncogene rearrangements in morphological variants of radiation-induced and sporadic thyroid papillary carcinomas in children. Cancer Res 1997; 57(9):1690-1694.
- 34. Puxeddu E, Moretti S, Elisei R et al. BRAF(V599E) mutation is the leading genetic event in adult sporadic papillary thyroid carcinomas. J Clin Endocrinol Metab 2004; 89(5):2414-2420.
- 35. Santoro M, Carlomagno F, Hay ID et al. Ret oncogene activation in human thyroid neoplasms is restricted to the papillary cancer subtype. J Clin Invest 1992; 89(5):1517-1522.
- Santoro M, Dathan NA, Berlingieri MT et al. Molecular characterization of RET/PTC3; a novel rearranged version of the RETproto-oncogene in a human thyroid papillary carcinoma. Oncogene 1994; 9(2):509-516.
- 37. Santoro M, Grieco M, Melillo RM, Fusco A, Vecchio G. Molecular defects in thyroid carcinomas: role of the RET oncogene in thyroid neoplastic transformation. Eur J Endocrinol 1995; 133(5):513-522.
- Santoro M, Chiappetta G, Cerrato A et al. Development of thyroid papillary carcinomas secondary to tissue-specific expression of the RET/PTC1 oncogene in transgenic mice. Oncogene 1996; 12(8):1821-1826.
- 39. Soares P, Trovisco V, Rocha AS et al. BRAF mutations and RET/PTC rearrangements are alternative events in the etiopathogenesis of PTC. Oncogene 2003; 22(29):4578-4580.
- 40. Tallini G. Molecular pathobiology of thyroid neoplasms. Endocr Pathol 2002; 13(4):271-288.
- 41. Viglietto G, Chiappetta G, Martinez-Tello FJ et al. RET/PTC oncogene activation is an early event in thyroid carcinogenesis. Oncogene 1995; 11(6):1207-1210.
- 42. Xing M. BRAF mutation in thyroid cancer. Endocr Relat Cancer 2005; 12(2):245-262.
- 43. Ivan M, Bond JA, Prat M, Comoglio PM, Wynford-Thomas D. Activated ras and ret oncogenes induce over-expression of c-met (hepatocyte growth factor receptor) in human thyroid epithelial cells. Oncogene 1997; 14(20):2417-2423.
- 44. Kroll TG, Sarraf P, Pecciarini L et al. PAX8-PPARgamma1 fusion oncogene in human thyroid carcinoma [corrected] [published erratum appears in Science 2000 Sep 1;289(5484):1474]. Science 2000; 289(5483):1357-1360.
- 45. Ying H, Suzuki H, Furumoto H et al. Alterations in genomic profiles during tumor progression in a mouse model of follicular thyroid carcinoma. Carcinogenesis 2003; 24(9):1467-1479.
- 46. Ravetto C, Colombo L, Dottorini ME. Usefulness of fine-needle aspiration in the diagnosis of thyroid carcinoma: a retrospective study in 37,895 patients. Cancer 2000; 90(6):357-363.
- 47. Sobin LH, Wittekind C. TNM Classification of malignant tumors. 6 ed. Wiley, Hoboken, New Yersey. 2002.
- Kukkonen ST, Haapiainen RK, Franssila KO, Sivula AH. Papillary thyroid carcinoma: the new, age-related TNM classification system in a retrospective analysis of 199 patients. World J Surg 1990; 14(6):837-841.
- 49. Wittekind C, Wagner G. TNM Classification. 5 ed. Springer Berlin. 1997.

- 50. Pacini F, Schlumberger M, Dralle H, Elisei R, Smit JW, Wiersinga W. European consensus for the management of patients with differentiated thyroid carcinoma of the follicular epithelium. Eur J Endocrinol 2006; 154(6):787-803.
- 51. Demeure MJ, Clark OH. Surgery in the treatment of thyroid cancer. Endocrinol Metab Clin North Am 1990; 19(3):663-683.
- 52. Baudin E, Travagli JP, Ropers J et al. Microcarcinoma of the thyroid gland: the Gustave-Roussy Institute experience. Cancer 1998; 83(3):553-559.
- 53. DeGroot LJ, Kaplan EL, McCormick M, Straus FH. Natural history, treatment, and course of papillary thyroid carcinoma. J Clin Endocrinol Metab 1990; 71(2):414-424.
- 54. Mazzaferri EL, Kloos RT. Clinical review 128: Current approaches to primary therapy for papillary and follicular thyroid cancer. J Clin Endocrinol Metab 2001; 86(4):1447-1463.
- 55. Katoh R, Sasaki J, Kurihara H, Suzuki K, Iida Y, Kawaoi A. Multiple thyroid involvement (intraglandular metastasis) in papillary thyroid carcinoma. A clinicopathologic study of 105 consecutive patients. Cancer 1992; 70(6):1585-1590.
- 56. Russell WO, Ibanez ML, Clark RL, White EC, . Thyroid carcinoma. Classification, intraglandular disssemination, and clinicopathological study based upon whole organ sections of 80 glands. Cancer 1963; 16:1425-1460.
- 57. Utiger RD. Follow-up of patients with thyroid carcinoma. N Engl J Med 1997; 337(13):928-930.
- 58. Simpson WJ, Panzarella T, Carruthers JS, Gospodarowicz MK, Sutcliffe SB. Papillary and follicular thyroid cancer: impact of treatment in 1578 patients. Int J Radiat Oncol Biol Phys 1988; 14(6):1063-1075.
- 59. Tubiana M, Schlumberger M, Rougier P et al. Long-term results and prognostic factors in patients with differentiated thyroid carcinoma. Cancer 1985; 55(4):794-804.
- 60. Sherman SI, Tielens ET, Sostre S, Wharam MD, Jr., Ladenson PW. Clinical utility of post treatment radioiodine scans in the management of patients with thyroid carcinoma. J Clin Endocrinol Metab 1994; 78(3):629-634.
- 61. Tenenbaum F, Corone C, Schlumberger M, Parmentier C. Thyroglobulin measurement and postablative iodine-131 total body scan after total thyroidectomy for differentiated thyroid carcinoma in patients with no evidence of disease. Eur J Cancer 1996; 32A(7):1262.
- Sawka AM, Thephamongkhol K, Brouwers M, Thabane L, Browman G, Gerstein HC. Clinical review 170:
 A systematic review and metaanalysis of the effectiveness of radioactive iodine remnant ablation for well-differentiated thyroid cancer. J Clin Endocrinol Metab 2004; 89(8):3668-3676.
- 63. Mazzaferri EL. Thyroid remnant 131I ablation for papillary and follicular thyroid carcinoma. Thyroid 1997; 7(2):265-271.
- 64. Samaan NA, Schultz PN, Hickey RC et al. The results of various modalities of treatment of well differentiated thyroid carcinomas: a retrospective review of 1599 patients. J Clin Endocrinol Metab 1992; 75(3):714-720.
- Brown AP, Chen J, Hitchcock YJ, Szabo A, Shrieve DC, Tward JD. The risk of second primary malignancies up to three decades after the treatment of differentiated thyroid cancer. J Clin Endocrinol Metab 2008; 93(2):504-515.
- 66. De VF, Schlumberger M, Delisle MJ et al. Leukaemias and cancers following iodine-131 administration for thyroid cancer. Br J Cancer 1997; 75(5):734-739.
- 67. Rubino C, De VF, Dottorini ME et al. Second primary malignancies in thyroid cancer patients. Br J Cancer 2003; 89(9):1638-1644.
- 68. Schlumberger M, Challeton C, De VF, Parmentier C. Treatment of distant metastases of differentiated thyroid carcinoma. J Endocrinol Invest 1995; 18(2):170-172.
- 69. Schlumberger M, Challeton C, De VF et al. Radioactive iodine treatment and external radiotherapy for lung and bone metastases from thyroid carcinoma. J Nucl Med 1996; 37(4):598-605.
- 70. Cooper DS, Specker B, Ho M et al. Thyrotropin suppression and disease progression in patients with differentiated thyroid cancer: results from the National Thyroid Cancer Treatment Cooperative Registry. Thyroid 1998; 8(9):737-744.
- 71. Schlumberger M, Pacini F, Wiersinga WM et al. Follow-up and management of differentiated thyroid carcinoma: a European perspective in clinical practice. Eur J Endocrinol 2004; 151(5):539-548.
- 72. Hovens GC, Stokkel MP, Kievit J et al. Associations of serum thyrotropin concentrations with recurrence

and death in differentiated thyroid cancer. J Clin Endocrinol Metab 2007; 92(7):2610-2615.

- Pujol P, Daures JP, Nsakala N, Baldet L, Bringer J, Jaffiol C. Degree of thyrotropin suppression as a prognostic determinant in differentiated thyroid cancer. J Clin Endocrinol Metab 1996; 81(12):4318-4323.
- 74. Spencer CA, Takeuchi M, Kazarosyan M. Current status and performance goals for serum thyroglobulin assays. Clin Chem 1996; 42(1):164-173.
- 75. Baudin E, Do CC, Cailleux AF, Leboulleux S, Travagli JP, Schlumberger M. Positive predictive value of serum thyroglobulin levels, measured during the first year of follow-up after thyroid hormone withdrawal, in thyroid cancer patients. J Clin Endocrinol Metab 2003; 88(3):1107-1111.
- 76. Cailleux AF, Baudin E, Travagli JP, Ricard M, Schlumberger M. Is diagnostic iodine-131 scanning useful after total thyroid ablation for differentiated thyroid cancer? J Clin Endocrinol Metab 2000; 85(1):175-178.
- 77. Kim TY, Kim WB, Kim ES et al. Serum thyroglobulin levels at the time of 131I remnant ablation just after thyroidectomy are useful for early prediction of clinical recurrence in low-risk patients with differentiated thyroid carcinoma. J Clin Endocrinol Metab 2005; 90(3):1440-1445.
- Kloos RT, Mazzaferri EL. A single recombinant human thyrotropin-stimulated serum thyroglobulin measurement predicts differentiated thyroid carcinoma metastases three to five years later. J Clin Endocrinol Metab 2005; 90(9):5047-5057.
- 79. Menendez TE, Lopez Carballo MT, Rodriguez Erdozain RM, Forga LL, Goni Iriarte MJ, Barberia Layana JJ. Prognostic value of thyroglobulin serum levels and 131l whole-body scan after initial treatment of lowrisk differentiated thyroid cancer. Thyroid 2004; 14(4):301-306.
- 80. Brabant G, Maenhaut C, Kohrle J et al. Human thyrotropin receptor gene: expression in thyroid tumors and correlation to markers of thyroid differentiation and dedifferentiation. Mol Cell Endocrinol 1991; 82(1):R7-12.
- Eustatia-Rutten CF, Smit JW, Romijn JA et al. Diagnostic value of serum thyroglobulin measurements in the follow-up of differentiated thyroid carcinoma, a structured meta-analysis. Clin Endocrinol (0xf) 2004; 61(1):61-74.
- 82. Schroeder PR, Haugen BR, Pacini F et al. A comparison of short-term changes in health-related quality of life in thyroid carcinoma patients undergoing diagnostic evaluation with recombinant human thyrotropin compared with thyroid hormone withdrawal. J Clin Endocrinol Metab 2006; 91(3):878-884.
- 83. Haugen BR, Pacini F, Reiners C et al. A comparison of recombinant human thyrotropin and thyroid hormone withdrawal for the detection of thyroid remnant or cancer. J Clin Endocrinol Metab 1999; 84(11):3877-3885.
- 84. Ladenson PW, Braverman LE, Mazzaferri EL et al. Comparison of administration of recombinant human thyrotropin with withdrawal of thyroid hormone for radioactive iodine scanning in patients with thyroid carcinoma. N Engl J Med 1997; 337(13):888-896.
- 85. Robbins RJ, Tuttle RM, Sharaf RN et al. Preparation by recombinant human thyrotropin or thyroid hormone withdrawal are comparable for the detection of residual differentiated thyroid carcinoma. J Clin Endocrinol Metab 2001; 86(2):619-625.
- 86. Mazzaferri EL, Robbins RJ, Spencer CA et al. A consensus report of the role of serum thyroglobulin as a monitoring method for low-risk patients with papillary thyroid carcinoma. J Clin Endocrinol Metab 2003; 88(4):1433-1441.
- 87. Pacini F, Molinaro E, Castagna MG et al. Recombinant human thyrotropin-stimulated serum thyroglobulin combined with neck ultrasonography has the highest sensitivity in monitoring differentiated thyroid carcinoma. J Clin Endocrinol Metab 2003; 88(8):3668-3673.
- 88. Pacini F, Ladenson PW, Schlumberger M et al. Radioiodine ablation of thyroid remnants after preparation with recombinant human thyrotropin in differentiated thyroid carcinoma: results of an international, randomized, controlled study. J Clin Endocrinol Metab 2006; 91(3):926-932.
- 89. Taieb D, Sebag F, Cherenko M et al. Quality of life changes and clinical outcomes in thyroid cancer patients undergoing radioiodine remnant ablation with recombinant human thyrotropin: a randomized controlled study. Clin Endocrinol (0xf) 2008.
- 90. Luster M, Lippi F, Jarzab B et al. rhTSH-aided radioiodine ablation and treatment of differentiated thyroid carcinoma: a comprehensive review. Endocr Relat Cancer 2005; 12(1):49-64.
- 91. Robbins RJ, Driedger A, Magner J. Recombinant human thyrotropin-assisted radioiodine therapy for

patients with metastatic thyroid cancer who could not elevate endogenous thyrotropin or be withdrawn from thyroxine. Thyroid 2006; 16(11):1121-1130.

- 92. Cooper DS, Doherty GM, Haugen BR et al. Management guidelines for patients with thyroid nodules and differentiated thyroid cancer. Thyroid 2006; 16(2):109-142.
- 93. Frasoldati A, Pesenti M, Gallo M, Caroggio A, Salvo D, Valcavi R. Diagnosis of neck recurrences in patients with differentiated thyroid carcinoma. Cancer 2003; 97(1):90-96.
- 94. Torlontano M, Crocetti U, D'Aloiso L et al. Serum thyroglobulin and 131I whole body scan after recombinant human TSH stimulation in the follow-up of low-risk patients with differentiated thyroid cancer. Eur J Endocrinol 2003; 148(1):19-24.
- 95. Schluter B, Bohuslavizki KH, Beyer W, Plotkin M, Buchert R, Clausen M. Impact of FDG PET on patients with differentiated thyroid cancer who present with elevated thyroglobulin and negative 131I scan. J Nucl Med 2001; 42(1):71-76.
- Robbins RJ, Wan Q, Grewal RK et al. Real-time prognosis for metastatic thyroid carcinoma based on 2-[18F]fluoro-2-deoxy-D-glucose-positron emission tomography scanning. J Clin Endocrinol Metab 2006; 91(2):498-505.
- 97. Bianco AC, Salvatore D, Gereben B, Berry MJ, Larsen PR. Biochemistry, cellular and molecular biology, and physiological roles of the iodothyronine selenodeiodinases. Endocr Rev 2002; 23(1):38-89.
- 98. Bianco AC, Kim BW. Deiodinases: implications of the local control of thyroid hormone action. J Clin Invest 2006; 116(10):2571-2579.
- 99. Kohrle J. Local activation and inactivation of thyroid hormones: the deiodinase family. Mol Cell Endocrinol 1999; 151(1-2):103-119.
- 100. Croteau W, Davey JC, Galton VA, St Germain DL. Cloning of the mammalian type II iodothyronine deiodinase. A selenoprotein differentially expressed and regulated in human and rat brain and other tissues. J Clin Invest 1996; 98(2):405-417.
- 101. Maeda A, Toyoda N, Yasuzawa-Amano S, Iwasaka T, Nishikawa M. Type 2 deiodinase expression is stimulated by growth factors in human vascular smooth muscle cells. Mol Cell Endocrinol 2003; 200(1-2):111-117.
- 102. Mizuma H, Murakami M, Mori M. Thyroid hormone activation in human vascular smooth muscle cells: expression of type II iodothyronine deiodinase. Circ Res 2001; 88(3):313-318.
- 103. Salvatore D, Bartha T, Harney JW, Larsen PR. Molecular biological and biochemical characterization of the human type 2 selenodeiodinase. Endocrinology 1996; 137(8):3308-3315.
- 104. Salvatore D, Tu H, Harney JW, Larsen PR. Type 2 iodothyronine deiodinase is highly expressed in human thyroid. J Clin Invest 1996; 98(4):962-968.
- 105. Inada M, Kasagi K, Kurata S et al. Estimation of thyroxine and triiodothyronine distribution and of the conversion rate of thyroxine to triiodothyronine in man. J Clin Invest 1975; 55(6):1337-1348.
- 106. Lum SM, Nicoloff JT, Spencer CA, Kaptein EM. Peripheral tissue mechanism for maintenance of serum triiodothyronine values in a thyroxine-deficient state in man. J Clin Invest 1984; 73(2):570-575.
- 107. Silva JE, Gordon MB, Crantz FR, Leonard JL, Larsen PR. Qualitative and quantitative differences in the pathways of extrathyroidal triiodothyronine generation between euthyroid and hypothyroid rats. J Clin Invest 1984; 73(4):898-907.
- 108. Toyoda N, Zavacki AM, Maia AL, Harney JW, Larsen PR. A novel retinoid X receptor-independent thyroid hormone response element is present in the human type 1 deiodinase gene. Mol Cell Biol 1995; 15(9):5100-5112.
- 109. Jakobs TC, Schmutzler C, Meissner J, Kohrle J. The promoter of the human type I 5'-deiodinase gene--mapping of the transcription start site and identification of a DR+4 thyroid-hormone-responsive element. Eur J Biochem 1997; 247(1):288-297.
- 110. Zhang CY, Kim S, Harney JW, Larsen PR. Further characterization of thyroid hormone response elements in the human type 1 iodothyronine deiodinase gene. Endocrinology 1998; 139(3):1156-1163.
- 111. Silva JE, Larsen PR. Comparison of iodothyronine 5'-deiodinase and other thyroid-hormone-dependent enzyme activities in the cerebral cortex of hypothyroid neonatal rat. Evidence for adaptation to hypothyroidism. J Clin Invest 1982; 70(5):1110-1123.
- 112. St Germain DL. Metabolic effect of 3,3',5'-triiodothyronine in cultured growth hormone-producing rat pituitary tumor cells. Evidence for a unique mechanism of thyroid hormone action. J Clin Invest 1985; 76(2):890-893.
- 113. St Germain DL. The effects and interactions of substrates, inhibitors, and the cellular thiol-disulfide

balance on the regulation of type II iodothyronine 5'-deiodinase. Endocrinology 1988; 122(5):1860-1868.

- 114. Burmeister LA, Pachucki J, St Germain DL. Thyroid hormones inhibit type 2 iodothyronine deiodinase in the rat cerebral cortex by both pre- and posttranslational mechanisms. Endocrinology 1997; 138(12):5231-5237.
- 115. Kaplan MM, Yaskoski KA. Maturational patterns of iodothyronine phenolic and tyrosyl ring deiodinase activities in rat cerebrum, cerebellum, and hypothalamus. J Clin Invest 1981; 67(4):1208-1214.
- 116. Mebis L, Langouche L, Visser TJ, Van den BG. The type II iodothyronine deiodinase is up-regulated in skeletal muscle during prolonged critical illness. J Clin Endocrinol Metab 2007; 92(8):3330-3333.
- 117. Maia AL, Kim BW, Huang SA, Harney JW, Larsen PR. Type 2 iodothyronine deiodinase is the major source of plasma T3 in euthyroid humans. Journal of Clinical Investigation 2005; 115(9):2524-2533.
- 118. Canani LH, Capp C, Dora JM et al. The type 2 deiodinase A/G (Thr92Ala) polymorphism is associated with decreased enzyme velocity and increased insulin resistance in patients with type 2 diabetes mellitus. J Clin Endocrinol Metab 2005; 90(6):3472-3478.
- 119. Mentuccia D, Proietti-Pannunzi L, Tanner K et al. Association between a novel variant of the human type 2 deiodinase gene Thr92Ala and insulin resistance: evidence of interaction with the Trp64Arg variant of the beta-3-adrenergic receptor. Diabetes 2002; 51(3):880-883.
- 120. Peeters RP, van der Deure WM, Visser TJ. Genetic variation in thyroid hormone pathway genes; polymorphisms in the TSH receptor and the iodothyronine deiodinases. Eur J Endocrinol 2006; 155(5):655-662.
- 121. Maia AL, Dupuis J, Manning A et al. The type 2 deiodinase (DIO2) A/G polymorphism is not associated with glycemic traits: the Framingham Heart Study. Thyroid 2007; 17(3):199-202.
- 122. Franklyn JA, Betteridge J, Daykin J et al. Long-term thyroxine treatment and bone mineral density. Lancet 1992; 340(8810):9-13.
- 123. Greenspan SL, Greenspan FS. The effect of thyroid hormone on skeletal integrity. Ann Intern Med 1999; 130(9):750-758.
- 124. Basset P, Okada A, Chenard MP et al. Matrix metalloproteinases as stromal effectors of human carcinoma progression: therapeutic implications. Matrix Biol 1997; 15(8-9):535-541.
- 125. Kanatani M, Sugimoto T, Sowa H, Kobayashi T, Kanzawa M, Chihara K. Thyroid hormone stimulates osteoclast differentiation by a mechanism independent of RANKL-RANK interaction. J Cell Physiol 2004; 201(1):17-25.
- 126. Miura M, Tanaka K, Komatsu Y et al. A novel interaction between thyroid hormones and 1,25(OH)(2) D(3) in osteoclast formation. Biochem Biophys Res Commun 2002; 291(4):987-994.
- 127. Anderson DM, Maraskovsky E, Billingsley WL et al. A homologue of the TNF receptor and its ligand enhance T-cell growth and dendritic-cell function. Nature 1997; 390(6656):175-179.
- 128. Hsu H, Lacey DL, Dunstan CR et al. Tumor necrosis factor receptor family member RANK mediates osteoclast differentiation and activation induced by osteoprotegerin ligand. Proc Natl Acad Sci U S A 1999; 96(7):3540-3545.
- 129. Kong YY, Feige U, Sarosi I et al. Activated T cells regulate bone loss and joint destruction in adjuvant arthritis through osteoprotegerin ligand. Nature 1999; 402(6759):304-309.
- 130. Lacey DL, Timms E, Tan HL et al. Osteoprotegerin ligand is a cytokine that regulates osteoclast differentiation and activation. Cell 1998; 93(2):165-176.
- 131. Matsuzaki K, Udagawa N, Takahashi N et al. Osteoclast differentiation factor (ODF) induces osteoclastlike cell formation in human peripheral blood mononuclear cell cultures. Biochem Biophys Res Commun 1998; 246(1):199-204.
- 132. Quinn JM, Elliott J, Gillespie MT, Martin TJ. A combination of osteoclast differentiation factor and macrophage-colony stimulating factor is sufficient for both human and mouse osteoclast formation invitro. Endocrinology 1998; 139(10):4424-4427.
- 133. Yasuda H, Shima N, Nakagawa N et al. Osteoclast differentiation factor is a ligand for osteoprotegerin/ osteoclastogenesis-inhibitory factor and is identical to TRANCE/RANKL. Proc Natl Acad Sci U S A 1998; 95(7):3597-3602.
- 134. Burgess TL, Qian Y, Kaufman S et al. The ligand for osteoprotegerin (OPGL) directly activates mature osteoclasts. J Cell Biol 1999; 145(3):527-538.
- 135. Fuller K, Wong B, Fox S, Choi Y, Chambers TJ. TRANCE is necessary and sufficient for osteoblast-

mediated activation of bone resorption in osteoclasts. J Exp Med 1998; 188(5):997-1001.

- 136. Hattersley G, Owens J, Flanagan AM, Chambers TJ. Macrophage colony stimulating factor (M-CSF) is essential for osteoclast formation in vitro. Biochem Biophys Res Commun 1991; 177(1):526-531.
- 137. Kodama H, Nose M, Niida S, Yamasaki A. Essential role of macrophage colony-stimulating factor in the osteoclast differentiation supported by stromal cells. J Exp Med 1991; 173(5):1291-1294.
- 138. Eriksen EF, Mosekilde L, Melsen F. Trabecular bone remodeling and bone balance in hyperthyroidism. Bone 1985; 6(6):421-428.
- 139. Mosekilde L, Melsen F, Bagger JP, Myhre-Jensen O, Schwartz SN. Bone changes in hyperthyroidism: interrelationships between bone morphometry, thyroid function and calcium-phosphorus metabolism. Acta Endocrinol (Copenh) 1977; 85(3):515-525.
- 140. Inoue M, Tawata M, Yokomori N, Endo T, Onaya T. Expression of thyrotropin receptor on clonal osteoblastlike rat osteosarcoma cells. Thyroid 1998; 8(11):1059-1064.
- 141. Abe E, Marians RC, Yu W et al. TSH is a negative regulator of skeletal remodeling. Cell 2003; 115(2):151-162.
- 142. Morimura T, Tsunekawa K, Kasahara T et al. Expression of type 2 iodothyronine deiodinase in human osteoblast is stimulated by thyrotropin. Endocrinology 2005; 146(4):2077-2084.
- 143. Galliford TM, Murphy E, Williams AJ, Bassett JH, Williams GR. Effects of thyroid status on bone metabolism: a primary role for thyroid stimulating hormone or thyroid hormone? Minerva Endocrinol 2005; 30(4):237-246.
- 144. Sun L, Davies TF, Blair HC, Abe E, Zaidi M. TSH and bone loss. Ann N Y Acad Sci 2006; 1068:309-318.
- 145. Davies T, Marians R, Latif R. The TSH receptor reveals itself. J Clin Invest 2002; 110(2):161-164.
- 146. Hase H, Ando T, Eldeiry L et al. TNFalpha mediates the skeletal effects of thyroid-stimulating hormone. Proc Natl Acad Sci U S A 2006; 103(34):12849-12854.
- 147. Sampath TK, Simic P, Sendak R et al. Thyroid-stimulating hormone restores bone volume, microarchitecture, and strength in aged ovariectomized rats. J Bone Miner Res 2007; 22(6):849-859.
- 148. van der Deure WM, Uitterlinden AG, Hofman A et al. Effects of serum TSH and FT4 levels and the TSHRAsp727Glu polymorphism on bone: the Rotterdam Study. Clin Endocrinol (Oxf) 2008; 68(2):175-181.
- 149. Haraguchi K, Saito T, Kaneshige M, Endo T, Onaya T. Desensitization and internalization of a thyrotrophin receptor lacking the cytoplasmic carboxy-terminal region. J Mol Endocrinol 1994; 13(3):283-288.
- Hansen PS, van der Deure WM, Peeters RP et al. The impact of a TSH receptor gene polymorphism on thyroid-related phenotypes in a healthy Danish twin population. Clin Endocrinol (Oxf) 2007; 66(6):827-832.
- 151. Peeters RP, van TH, Klootwijk W et al. Polymorphisms in thyroid hormone pathway genes are associated with plasma TSH and iodothyronine levels in healthy subjects. J Clin Endocrinol Metab 2003; 88(6):2880-2888.
- 152. Botella-Carretero JI, varez-Blasco F, San Millan JL, Escobar-Morreale HF. Thyroid hormone deficiency and postmenopausal status independently increase serum osteoprotegerin concentrations in women.Eur J Endocrinol 2007; 156(5):539-545.
- 153. Guang-Da X, Hui-Ling S, Zhi-Song C, Lin-Shuang Z. Changes in plasma concentrations of osteoprotegerin before and after levothyroxine replacement therapy in hypothyroid patients. J Clin Endocrinol Metab 2005; 90(10):5765-5768.
- 154. Nagasaki T, Inaba M, Jono S et al. Increased levels of serum osteoprotegerin in hypothyroid patients and its normalization with restoration of normal thyroid function. Eur J Endocrinol 2005; 152(3):347-353.
- 155. Nakamura H, Mori T, Genma R et al. Urinary excretion of pyridinoline and deoxypyridinoline measured by immunoassay in hypothyroidism. Clin Endocrinol (0xf) 1996; 44(4):447-451.
- 156. Engler H, Oettli RE, Riesen WF. Biochemical markers of bone turnover in patients with thyroid dysfunctions and in euthyroid controls: a cross-sectional study. Clin Chim Acta 1999; 289(1-2):159-172.
- 157. Sabuncu T, Aksoy N, Arikan E, Ugur B, Tasan E, Hatemi H. Early changes in parameters of bone and mineral metabolism during therapy for hyper- and hypothyroidism. Endocr Res 2001; 27(1-2):203-213.
- 158. Sekeroglu MR, Altun ZB, Algun E et al. Serum cytokines and bone metabolism in patients with thyroid dysfunction. Adv Ther 2006; 23(3):475-480.
- 159. Bassett JH, Williams AJ, Murphy E et al. A lack of thyroid hormones rather than excess TSH causes

abnormal skeletal development in hypothyroidism. Mol Endocrinol 2007.

- 160. Bassett JH, O'Shea PJ, Sriskantharajah S et al. Thyroid hormone excess rather than thyrotropin deficiency induces osteoporosis in hyperthyroidism. Mol Endocrinol 2007; 21(5):1095-1107.
- 161. Giusti M, Cecoli F, Ghiara C et al. Recombinant human thyroid stimulating hormone does not acutely change serum osteoprotegerin and soluble receptor activator of nuclear factor-kappa Beta ligand in patients under evaluation for differentiated thyroid carcinoma. Hormones (Athens) 2007; 6(4):304-313.
- 162. Martini G, Gennari L, De P, V et al. The effects of recombinant TSH on bone turnover markers and serum osteoprotegerin and RANKL levels. Thyroid 2008; 18(4):455-460.
- 163. Mazziotti G, Sorvillo F, Piscopo M et al. Recombinant human TSH modulates in vivo C-telopeptides of type-1 collagen and bone alkaline phosphatase, but not osteoprotegerin production in postmenopausal women monitored for differentiated thyroid carcinoma. J Bone Miner Res 2005; 20(3):480-486.
- 164. Gouveia CH, Christoffolete MA, Zaitune CR et al. Type 2 iodothyronine selenodeiodinase is expressed throughout the mouse skeleton and in the MC3T3-E1 mouse osteoblastic cell line during differentiation. Endocrinology 2005; 146(1):195-200.
- 165. LeBron BA, Pekary AE, Mirell C, Hahn TJ, Hershman JM. Thyroid hormone 5'-deiodinase activity, nuclear binding, and effects on mitogenesis in UMR-106 osteoblastic osteosarcoma cells. J Bone Miner Res 1989; 4(2):173-178.
- 166. Miura M, Tanaka K, Komatsu Y et al. Thyroid hormones promote chondrocyte differentiation in mouse ATDC5 cells and stimulate endochondral ossification in fetal mouse tibias through iodothyronine deiodinases in the growth plate. J Bone Miner Res 2002; 17(3):443-454.
- 167. Shen S, Berry W, Jaques S, Pillai S, Zhu J. Differential expression of iodothyronine deiodinase type 2 in growth plates of chickens divergently selected for incidence of tibial dyschondroplasia. Anim Genet 2004; 35(2):114-118.
- 168. Christoffolete MA, Arrojo e Drigo, Gazoni F et al. Mice with impaired extrathyroidal thyroxine to 3,5,3'triiodothyronine conversion maintain normal serum 3,5,3'-triiodothyronine concentrations. Endocrinology 2007; 148(3):954-960.
- Williams AJ, Robson H, Kester MH et al. lodothyronine deiodinase enzyme activities in bone. Bone 2008;
 43(1):126-134.
- 170. Duntas LH. Thyroid disease and lipids. Thyroid 2002; 12(4):287-293.
- 171. Kim BJ, Kim TY, Koh JM et al. Relationship between serum free thyroxine levels and metabolic syndrome and its components in healthy euthyroid subjects. Clin Endocrinol (Oxf) 2008.
- 172. Roos A, Bakker SJ, Links TP, Gans RO, Wolffenbuttel BH. Thyroid function is associated with components of the metabolic syndrome in euthyroid subjects. J Clin Endocrinol Metab 2007; 92(2):491-496.
- 173. Takashima N, Niwa Y, Mannami T, Tomoike H, Iwai N. Characterization of subclinical thyroid dysfunction from cardiovascular and metabolic viewpoints: the Suita study. Circ J 2007; 71(2):191-195.
- 174. Koh H, Fujii S, Nishioheda Y, Tsushima M, Nambu S. 3,5,3'-Triiodothyronine regulates insulin level in the circulation following glucose ingestion in humans. Arzneimittelforschung 1986; 36(2):262-265.
- 175. Cavallo-Perin P, Bruno A, Boine L, Cassader M, Lenti G, Pagano G. Insulin resistance in Graves' disease: a quantitative in-vivo evaluation. Eur J Clin Invest 1988; 18(6):607-613.
- 176. Gimenez-Palop O, Gimenez-Perez G, Mauricio D et al. Circulating ghrelin in thyroid dysfunction is related to insulin resistance and not to hunger, food intake or anthropometric changes. Eur J Endocrinol 2005; 153(1):73-79.
- 177. Iglesias P, Alvarez FP, Codoceo R, Diez JJ. Serum concentrations of adipocytokines in patients with hyperthyroidism and hypothyroidism before and after control of thyroid function. Clin Endocrinol (Oxf) 2003; 59(5):621-629.
- 178. Ikeda T, Fujiyama K, Hoshino T, Takeuchi T, Mashiba H, Tominaga M. Oral and intravenous glucoseinduced insulin secretion in hyperthyroid patients. Metabolism 1990; 39(6):633-637.
- 179. Jenkins RC, Valcavi R, Zini M et al. Association of elevated insulin-like growth factor binding protein-1 with insulin resistance in hyperthyroidism. Clin Endocrinol (Oxf) 2000; 52(2):187-195.
- 180. Tosi F, Moghetti P, Castello R, Negri C, Bonora E, Muggeo M. Early changes in plasma glucagon and growth hormone response to oral glucose in experimental hyperthyroidism. Metabolism 1996; 45(8):1029-1033.
- 181. Yaturu S, Prado S, Grimes SR. Changes in adipocyte hormones leptin, resistin, and adiponectin in thyroid

dysfunction. J Cell Biochem 2004; 93(3):491-496.

- 182. Cavallo-Perin P, Bruno A, Boine L, Cassader M, Lenti G, Pagano G. Insulin resistance in Graves' disease: a quantitative in-vivo evaluation. Eur J Clin Invest 1988; 18(6):607-613.
- 183. Andersen OO, Friis T, Ottesen B. Glucose tolerance and insulin secretion in hyperthyroidism. Acta Endocrinol (Copenh) 1977; 84(3):576-587.
- 184. Malaisse WJ, Malaisse-Lagae F, McCraw EF. Effects of thyroid function upon insulin secretion. Diabetes 1967; 16(9):643-646.
- 185. Kabadi UM, Eisenstein AB. Glucose intolerance in hyperthyroidism: role of glucagon. J Clin Endocrinol Metab 1980; 50(2):392-396.
- 186. Garcia-Sainz JA, Litosch I, Hoffman BB, Lefkowitz RJ, Fain JN. Effect of thyroid status on alpha- and betacatecholamine responsiveness of hamster adipocytes. Biochim Biophys Acta 1981; 678(3):334-341.
- 187. Yavuz DG, Yuksel M, Deyneli O, Ozen Y, Aydin H, Akalin S. Association of serum paraoxonase activity with insulin sensitivity and oxidative stress in hyperthyroid and TSH-suppressed nodular goitre patients. Clin Endocrinol (0xf) 2004; 61(4):515-521.
- 188. Langer P, Kocan A, Tajtakova M et al. Thyroid function and cholesterol level: paradoxical findings in large groups of population with high cholesterol food intake. Endocr Regul 2003; 37(3):175-180.
- 189. Lee WY, Suh JY, Rhee EJ, Park JS, Sung KC, Kim SW. Plasma CRP, apolipoprotein A-1, apolipoprotein B and Lpa levels according to thyroid function status. Arch Med Res 2004; 35(6):540-545.
- 190. Yavuz DG, Yuksel M, Deyneli O, Ozen Y, Aydin H, Akalin S. Association of serum paraoxonase activity with insulin sensitivity and oxidative stress in hyperthyroid and TSH-suppressed nodular goitre patients. Clin Endocrinol (0xf) 2004; 61(4):515-521.
- 191. Franklyn JA, Daykin J, Betteridge J et al. Thyroxine replacement therapy and circulating lipid concentrations. Clin Endocrinol (Oxf) 1993; 38(5):453-459.
- 192. Parle JV, Franklyn JA, Cross KW, Jones SR, Sheppard MC. Circulating lipids and minor abnormalities of thyroid function. Clin Endocrinol (Oxf) 1992; 37(5):411-414.
- 193. Franklyn JA, Daykin J, Betteridge J et al. Thyroxine replacement therapy and circulating lipid concentrations. Clin Endocrinol (Oxf) 1993; 38(5):453-459.
- 194. Fazio S, Palmieri EA, Lombardi G, Biondi B. Effects of thyroid hormone on the cardiovascular system. Recent Prog Horm Res 2004; 59:31-50.
- 195. Klein I. Thyroid hormone and the cardiovascular system. Am J Med 1990; 88(6):631-637.
- 196. Polikar R, Burger AG, Scherrer U, Nicod P. The thyroid and the heart. Circulation 1993; 87(5):1435-1441.
- 197. Levey GS, Klein I. Catecholamine-thyroid hormone interactions and the cardiovascular manifestations of hyperthyroidism. Am J Med 1990; 88(6):642-646.
- 198. Chen JL, Chiu HW, Tseng YJ, Chu WC. Hyperthyroidism is characterized by both increased sympathetic and decreased vagal modulation of heart rate: evidence from spectral analysis of heart rate variability. Clin Endocrinol (Oxf) 2006; 64(6):611-616.
- 199. Biondi B, Palmieri EA, Lombardi G, Fazio S. Effects of subclinical thyroid dysfunction on the heart. Ann Intern Med 2002; 137(11):904-914.
- 200. Petretta M, Bonaduce D, Spinelli L et al. Cardiovascular haemodynamics and cardiac autonomic control in patients with subclinical and overt hyperthyroidism. Eur J Endocrinol 2001; 145(6):691-696.
- 201. Smit JW, Eustatia-Rutten CF, Corssmit EP et al. Reversible diastolic dysfunction after long-term exogenous subclinical hyperthyroidism: a randomized, placebo-controlled study. J Clin Endocrinol Metab 2005; 90(11):6041-6047.
- 202. Portella RB, Pedrosa RC, Coeli CM, Buescu A, Vaisman M. Altered cardiovascular vagal responses in nonelderly female patients with subclinical hyperthyroidism and no apparent cardiovascular disease. Clin Endocrinol (0xf) 2007; 67(2):290-294.
- 203. Osman F, Franklyn JA, Daykin J et al. Heart rate variability and turbulence in hyperthyroidism before, during, and after treatment. Am J Cardiol 2004; 94(4):465-469.
- 204. Klein I, Ojamaa K. Thyroid hormone and the cardiovascular system. N Engl J Med 2001; 344(7):501-509.
- 205. Ladenson PW. Recognition and management of cardiovascular disease related to thyroid dysfunction. Am J Med 1990; 88(6):638-641.
- 206. Osman F, Gammage MD, Franklyn JA. Thyroid disease and its treatment: short-term and long-term

cardiovascular consequences. Curr Opin Pharmacol 2001; 1(6):626-631.

- 207. Tielens ET, Pillay M, Storm C, Berghout A. Cardiac function at rest in hypothyroidism evaluated by equilibrium radionuclide angiography. Clin Endocrinol (Oxf) 1999; 50(4):497-502.
- 208. Galetta F, Franzoni F, Fallahi P et al. Changes in heart rate variability and QT dispersion in patients with overt hypothyroidism. Eur J Endocrinol 2008; 158(1):85-90.
- 209. Xing H, Shen Y, Chen H, Wang Y, Shen W. Heart rate variability and its response to thyroxine replacement therapy in patients with hypothyroidism. Chin Med J (Engl) 2001; 114(9):906-908.
- 210. Biondi B, Fazio S, Carella C et al. Control of adrenergic overactivity by beta-blockade improves the quality of life in patients receiving long term suppressive therapy with levothyroxine. Journal of Clinical Endocrinology and Metabolism 1994; 78(5):1028-1033.
- 211. Biondi B, Palmieri EA, Fazio S et al. Endogenous subclinical hyperthyroidism affects quality of life and cardiac morphology and function in young and middle-aged patients. Journal of Clinical Endocrinology and Metabolism 2000; 85(12):4701-4705.
- 212. Gulseren S, Gulseren L, Hekimsoy Z, Cetinay P, Ozen C, Tokatlioglu B. Depression, anxiety, health-related quality of life, and disability in patients with overt and subclinical thyroid dysfunction. Arch Med Res 2006; 37(1):133-139.
- 213. Giusti M, Sibilla F, Cappi C et al. A case-controlled study on the quality of life in a cohort of patients with history of differentiated thyroid carcinoma. Journal of Endocrinological Investigation 2005; 28(7):599-608.
- 214. Schultz PN, Stava C, Vassilopoulou-Sellin R. Health profiles and quality of life of 518 survivors of thyroid cancer. Head Neck 2003; 25(5):349-356.
- 215. Tan LG, Nan L, Thumboo J, Sundram F, Tan LK. Health-related quality of life in thyroid cancer survivors. Laryngoscope 2007; 117(3):507-510.
- 216. Eustatia-Rutten CF, Corssmit EP, Pereira AM et al. Quality of life in longterm exogenous subclinical hyperthyroidism and the effects of restoration of euthyroidism, a randomized controlled trial. Clinical Endocrinology 2006; 64(3):284-291.
- 217. Tagay S, Herpertz S, Langkafel M et al. Health-related quality of life, anxiety and depression in thyroid cancer patients under short-term hypothyroidism and TSH-suppressive levothyroxine treatment. Eur J Endocrinol 2005; 153(6):755-763.
- 218. Mentuccia D, Thomas MJ, Coppotelli G et al. The Thr92Ala deiodinase type 2 (DIO2) variant is not associated with type 2 diabetes or indices of insulin resistance in the old order of Amish. Thyroid 2005; 15(11):1223-1227.
- 219. Peeters RP, van den Beld AW, Attalki H et al. A new polymorphism in the type II deiodinase gene is associated with circulating thyroid hormone parameters. Am J Physiol Endocrinol Metab 2005; 289(1): E75-E81.
- 220. de Jong FJ, Peeters RP, den HT et al. The association of polymorphisms in the type 1 and 2 deiodinase genes with circulating thyroid hormone parameters and atrophy of the medial temporal lobe. J Clin Endocrinol Metab 2007; 92(2):636-640.
- 221. Torlontano M, Durante C, Torrente I et al. Type 2 deiodinase polymorphism (threonine 92 alanine) predicts L-thyroxine dose to achieve target thyrotropin levels in thyroidectomized patients. J Clin Endocrinol Metab 2008; 93(3):910-913.

General introduction