

Optimizing triage and treatment strategies in urinary tract infection Stalenhoef, J.E.

Citation

Stalenhoef, J. E. (2019, May 8). *Optimizing triage and treatment strategies in urinary tract infection*. Retrieved from https://hdl.handle.net/1887/72409

Version: Not Applicable (or Unknown)

License: Leiden University Non-exclusive license

Downloaded from: https://hdl.handle.net/1887/72409

Note: To cite this publication please use the final published version (if applicable).

Cover Page



Universiteit Leiden

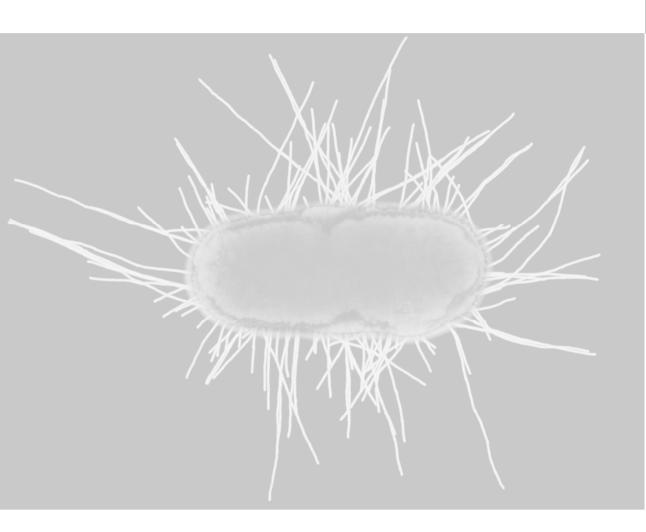


The handle http://hdl.handle.net/1887/72409 holds various files of this Leiden University dissertation.

Author: Stalenhoef, J.E.

Title: Optimizing triage and treatment strategies in urinary tract infection

Issue Date: 2019-05-08



CHAPTER 9

The use of automated urine microscopy analysis in the clinical diagnosis of urinary tract infection; defining an optimal diagnostic score in an academic medical center population

Dimard E. Foudraine, Martijn P. Bauer, Anne Russcher, Elske Kusters, Christa M. Cobbaert, Martha T. van der Beek, Janneke E. Stalenhoef

J Clin Microbiol. 2018 Apr 11. doi: 10.1128/JCM.02030-17

ABSTRACT

A retrospective case record study was conducted that established a scoring tool based on clinical and iQ200 parameters, able to predict or rule out the clinical diagnosis of UTI in the majority of adult patients in an academic hospital.

Automated standardized quantitative urine analysis, such as iQ200 analysis, is on the rise because of its high accuracy and efficiency compared to those of traditional urine analysis. Previous research on automated urinalysis focused mainly on predicting culture results but not on the clinical diagnosis of urinary tract infection (UTI). A retrospective analysis was conducted of consecutive urine samples sent in for culture because of suspected UTI. UTI was defined by expert opinion, based on reported symptoms, conventional urine sediment analysis, and urine cultures. Parameters of iQ200 analysis and clinical symptoms and signs were compared between cases and controls. Optimal cut-off values were determined for iQ200 parameters, and multivariate logistic regression analysis was used to identify the set of variables that best predicts the clinical diagnosis of UTI for development of a scoring tool. A total of 382 patients were included. Optimal cut-off values of iQ200 analysis were 74 white blood cells (WBC)/µl, 6,250 "all small particles" (ASP)/µl, and a bacterial score of 2 on an ordinal scale of 0 to 5. The scoring tool attributed 1 point for frequent micturition or increased urge, 2 points for dysuria, 1 point for a bacterial score of ≥2, 2 points for WBC/µl of ≥50, and an additional point for WBC/µl of ≥150. This score had a sensitivity of 86% and a specificity of 92% when using a threshold of <4 points. The combination of iQ200 analysis and a simple survey could predict or rule out UTIs in a majority of patients in an academic medical center.

INTRODUCTION

Urinary tract infection (UTI) is among the most frequently occurring infections and is the second most frequent clinical indication for empirical antibiotic treatment in primary and secondary care.^{1,2} The gold standard for diagnosis is detection of a pathogen in the urine in the presence of clinical symptoms. Because the result of a traditional urine culture is not readily available, presumptive diagnosis of UTI is based on diagnostic tests such as dipstick or urinary sediment analysis.³

In some populations, the diagnosis of UTI is not as straightforward and should be distinguished from asymptomatic bacteriuria or inflammatory conditions, such as interstitial cystitis.² This is especially the case in a tertiary hospital, where relatively many patients have complex urinary tract problems or kidney transplants or are treated with immunosuppressive medication.

In the past few years, automated, standardized, quantitative urine analysis has been introduced in clinical practice and has shown high efficiency and accuracy compared to traditional sediment analysis.⁴ One of these systems is the IRIS Diagnostics iQ200 Elite (iQ200), currently marketed by Beckman Coulter Inc., which analyzes urinary samples using flow imaging technology and auto particle recognition. The iQ200 classifies and quantifies particles, including bacteria, yeasts, white blood cells (WBC), and squamous epithelial cells, and correlates well with traditional urinary sediment examination with manual cell counts.⁵

Our group and other research groups have so far focused mainly on the use of automated urinalysis as a screening tool to predict negative urine cultures and thus to reduce the culture workload in the laboratory.⁶⁻⁹ For this purpose, a positive culture was used as the "laboratory" gold standard of UTI without taking clinical symptoms into account, therefore predicting the presence of bacteriuria, but not of symptomatic UTI.^{10,11} This distinction is important because it is currently thought that there is no role for treatment of patients with asymptomatic bacteriuria other than for pregnant women and patients undergoing urologic procedures.¹²

The test results of automated urine analysis are, however, subject to different clinical interpretations. This is partly because of unfamiliarity with quantitative results, instead of the semi-quantitative test results that clinicians used before, and the lack of optimal cutoff values for the clinical diagnosis of UTI.

The goal of the current study was to establish cutoff values for parameters of iQ200 analysis, to be used in diagnosing symptomatic UTI in a tertiary hospital population. Subsequently, we aimed to develop a scoring model to predict the clinical diagnosis of urinary tract infection, based on both symptoms and these cutoff values.

MATERIALS AND METHODS

Setting and patient population.

A retrospective study was performed at Leiden University Medical Center, which is an academic tertiary hospital in Leiden, the Netherlands. It has approximately 400 hospital beds and focuses on transplant medicine (solid organ transplants and stem cell transplants), resulting in a large proportion of immunocompromised patients. Samples from inpatients and outpatients of the hospital constitute the majority of samples sent to the clinical chemistry and microbiological laboratories (6). The study was approved by the Ethics Committee.

Urine samples.

Upon receipt at the Department of Medical Microbiology, all urine samples submitted for bacterial culture during a 12-week period from 25 February to 17 May 2013 were divided into two portions under sterile conditions if they had sufficient volume (at least 2 ml for culture and Gram stain and 3 ml for the iQ200 screening). One portion was analyzed by the iQ200 system in the clinical chemistry laboratory within 2 h after receipt from the microbiological laboratory. Results were not reported to the clinician because the iQ200 was still under validation. The other portion was analyzed by the microbiological laboratory. For more detailed information regarding procedures we refer to Russcher et al.⁶

For the purpose of this study, urine samples from children, pregnant women, and patients with an indwelling urinary catheter for more than 24 h, a nephrostomy, or a urostomy were excluded because the diagnosis of UTI is defined differently within these groups. Urine samples from patients without clinical data or clinical suspicion for UTI (e.g., preoperative routine urine controls) were excluded as well. Only the first sample of each patient was included.

Microbiological analysis.

Urine samples were analyzed using local standard microbiological methods for Gram stain and culture. The bacterial load was assessed and scored from <100 CFU/ml (no growth) to \geq 10 5 CFU/ml. The relevance of the urine sample was assessed according to our standard protocol for urine cultures, taking a quality score (the Q score) based on white blood cell (WBC) and squamous epithelial cell (SEC) counts in the Gram stain into account, as previously described (6). In urine samples with a high Q score (\geq 1, corresponding with a high WBC and low SEC count), all growth was identified to the species level. Colonies in samples with a Q score of zero were only identified to the species level if a monoculture with a bacterial load of \geq 10 5 CFU/ml was present. Samples with Q scores of \leq 0 were generally classified as mixed flora.

A positive culture was defined as having $\geq 10^3$ CFU/ml of not more than two different usual uropathogens or as having $\geq 10^5$ CFU/ml of a single unusual urinary pathogen. Common and uncommon pathogens and non-pathogens that were cultured are listed in Table 1.

Table 1. Pathogens isolated from urine cultures of 381 patients with and without UTIs.

Pathogen group or pathogen	No. with UTI ($n = 59$)	No. without UTI ($n = 322$)
Usual urinary pathogens		
Escherichia coli	31	26
Klebsiella spp.	4	6
Enterococcus spp.	2	5
Pseudomonas aeruginosa	2	4
Aerococcus urinae	2	2
Proteus mirabilis	0	3
Unusual urinary pathogens		
Other Enterobacteriaceae	2	4
β-hemolytic streptococci	2	3
Staphylococcus aureus	2	0
Haemophilus parainfluenzae	1	0
Candida spp.	0	2
Nonurinary pathogens		
Staphylococcus haemolyticus	2	0
Other staphylococci	0	3
Gardnerella vaginalis	0	1
Remaining		
Mixed flora	6	186
No growth	3	77

Automated urine microscopic analysis.

All samples derived from the Department of Medical Microbiology were tested by the iQ200° Elite analyzer (Iris Diagnostics, Chatsworth, CA), which is an automated urine microscopy analyzer that uses flow cytometry and digital photography. Automatic particle recognition software categorizes urine particles into 12 groups, including leukocytes, erythrocytes, bacteria, and "all small particles" (ASP). The ASP group consists of unclassified particles of <3 μ m, such as cocci, which are not recognized well by the iQ200, some other bacteria, crystals, and other formed elements. All elements other than bacteria were quantitatively reported (per microliter), and bacteria were reported semi-quantitatively (on a scale from 0 to 5). After automatic classification, a trained technician reviewed all images. Misplaced or unclassified images were placed in the correct categories, and bacterial counts were adjusted in cases when cocci were present.

Conventional urine analysis.

In a vast majority of patients from whom a urine sample was sent in for culture, a different sample was sent to the clinical chemistry for dipstick analysis. If the dipstick tested positive for leukocytes or erythrocytes, sediment analysis was performed using local standard protocol.

The positively tested urine was centrifuged for 5 min at 2,000 rpm. Subsequently, urine was poured off until 0.5 ml supernatant remained. This was shaken, and one drop was analyzed on a slide under a microscope. Observed elements were quantified as the number per high power field and reported qualitatively in the medical record.

Clinical assessment.

Clinical data and characteristics of included patients were obtained from the electronic medical records. Patients were retrospectively classified as either cases having a UTI or controls who did not have a UTI by two infectious diseases specialists using medical chart review. The expert reviewers used data on symptoms, signs, antibiotic (pre)treatment, and outcome, as documented in the electronic patient files. They used data on culture results and conventional urine analysis, which consisted of dipstick and sediment analysis. They also considered whether another diagnosis was more likely or could be the cause of complaints and/or fever. They were blinded to the iQ200 results, which were not reported in the medical records. If they differed in opinion, they reached consensus by means of discussion.

Statistical analysis

Baseline characteristics were compared with $\chi 2$ -tests for dichotomous variables and an unpaired t-test for age. Symptoms and signs were compared with $\chi 2$ -tests. Parameters from the iQ200 analysis were compared using unpaired t-tests and receiver operating characteristic (ROC) curves were plotted. Cutoff values were determined based on the optimal tradeoff between sensitivity and specificity. These cutoffs correspond with coordinates on the ROC curves which are closest to 0.1 (the upper left corner).¹³ Distances for all coordinates on the ROC curves to 0.1 were calculated by the formula: $\sqrt{d} = (1\text{-sensitivity})^2 + (1\text{-Specificity})^2$.

A logistic regression model was established using symptoms and parameters from the iQ200. In the case of information on a specific sign or symptom not being documented in the electronic patient file, that patient was excluded for this specific analysis. Backward selection excluded parameters based on likelihood ratios without significantly changing the fit of the model. The final model retained all variables significantly associated with the presence of UTI at a p<0.05 level. A numerical scoring tool was developed using the model by simplifying β-coefficients of all independent predictor variables. We calculated the area under the receiver operating characteristic curve (AUC) with 95% confidence interval (CI) to assess the scoring tool's discriminatory power to predict or rule out UTI. ROC curves were also plotted for the separate iO200 and clinical variables derived from the model.

Cutoff values were considered based on sensitivity, specificity, positive and negative predictive values (PPV and NPV, respectively). All analyses were performed using SPSS 21.0 (SPPS Inc, Chicago, IL, USA).

RESULTS

Population characteristics

During the study period 1442 urine samples from 1084 unique patients were submitted. The following samples were excluded: 641 samples from patients not suspected of having a UTI, 152 samples from patients with an indwelling urinary catheter >24 hours, 76 samples from pregnant patients, 62 samples of children below the age of 18, 33 samples from a nephrostomy or urostomy drain, 91 subsequent samples of patients already included, and 13 samples lacking data in the corresponding electronic patient files. After exclusion, 381 unique patients and urine samples remained. A total of 29 of 381 urine samples submitted were obtained by one-time catheterization and the rest by midstream clean catch. The prevalence of UTI among the 381 patients according to the expert review was 59. The expert reviewers initially differed in opinion in 30 of 381 patients (7.8%) but reached consensus by means of discussion. Table 2 shows demographic and clinical characteristics of the two patient groups (with and without urinary tract infections). Patients who had a UTI were significantly older (p = 0.041) than those who did not. None of the other characteristics differed significantly between both groups.

Table 2. Baseline characteristics of 381 patients with and without UTIs.

Characteristic	With UTI (n = 59)	Without UTI (n = 322)	P value
Age in yrs (mean [SD]) ^a	61.1 (17.7)	55.8 (18.2)	0.04
Male (no. [%])	33 (56)	152 (47)	0.22
Hospitalized (no. [%])	23 (39)	151 (47)	0.26
Indwelling catheter removed <7 days prior to culture (no. [%])	4 (7)	27 (8)	0.70
Immunosuppressive medication <3 months prior to culture (no. [%])	16 (28)	113 (35)	0.26
Neutropenia (no. [%])	0 (0)	15 (5)	0.09
Antibiotics <48 hours prior to culture (no. [%])	16 (29)	98 (31)	0.74
Renal transplant (no. [%])	6 (10)	39 (12)	0.70
Pancreatic transplant (no. [%])	0 (0)	8 (3)	0.23
Hematopoietic stem cell transplant (no. [%])	2 (3)	14 (4)	0.75
Fever (no. [%]) ^b	14 (25)	100 (31)	0.31

 $^{^{}a}P < 0.05$.

Culture results

Table 1 shows culture results of cases and controls. *Escherichia coli* was the most prevalent pathogen (n = 57). 192 Cultures displayed mixed flora and 80 cultures showed no growth. Patients who were assessed as cases with a UTI while their culture showed no growth were all treated with antibiotics in the 48 hours prior to culture (n = 3).

^b Fever was defined as a temperature higher than 38.1 °C.

Signs and symptoms

The prevalence of signs and symptoms among both patient groups are listed in Table 3. Dysuria, recognition of symptoms from a previous UTI, frequent micturition, and cloudy urine were most strongly associated with UTI. Subgroup analysis was conducted for aggravated lower urinary tract symptoms (LUTS) in male patients, increased cognitive impairment in patients older than 59 years of age and vaginal irritation or changed discharge in women. None of the three symptoms was significantly associated with UTI in their respective subgroups (not shown). The concentration of *C*-reactive protein in serum and leukocyte count in blood did not differ significantly between patients with and without UTIs (p values were 0.95 and 0.69 respectively). The same applied to the proportion of patients with positive blood cultures when comparing both groups (p = 0.31).

Table 3. Signs and symptoms of 381 patients with and without UTIs.

Sign/symptom ^a	With UTI (no. [%])	Without UTI (no. [%])	Odds ratio (CI)	<i>p</i> value ^c
Frequent micturition/increased urgency ^b	27 (54)	56 (19)	5.0 (2.7 – 9.3)	0.00
Dysuria	36 (69)	55 (18)	10.1 (5.2 – 19.5)	0.00
Aggravated LUTS	5 (10)	23 (8)	1.3 (0.5 – 3.6)	0.59
Suprapubic pain ^b	7 (13)	15 (5)	2.9 (1.1 – 7.4)	0.02
Recognition of symptoms from a previous UTI ^b	14 (27)	15 (5)	7.0 (3.1 – 15.6)	0.00
Increased incontinence	7 (13)	21 (7)	2.1 (0.8 – 5.3)	0.10
Macroscopic haematuria	3 (6)	16 (5)	1.1 (0.3 – 4.0)	0.87
Cloudy urine ^b	12 (24)	18 (6)	4.9 (2.2 - 11.0)	0.00
Foul smelling urine ^b	10 (20)	21 (7)	3.3 (1.4 – 7.5)	0.00
Increased cognitive impairment	6 (10)	30 (9)	1.1 (0.4 – 2.8)	0.82
Suprapubic tenderness	5 (18)	16 (9)	2.2 (0.7 - 6.5)	0.16
Costovertebral angle tenderness	3 (19)	9 (16)	1.3 (0.3 – 5.3)	0.76
All signs/symptoms	59 (15)	322 (85)		

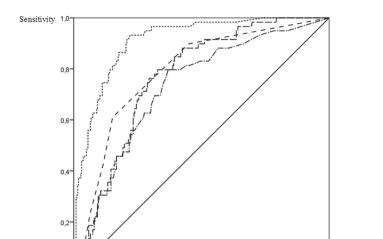
^aAll symptoms and signs as reported in the patient file. LUTS, lower urinary tract symptoms.

iQ200 parameters

The difference in distribution of white blood cells in urine between cases and controls was obvious. Most cases had a count of > 20 leukocytes/ μ l (97%) while most controls had a count \leq 20 leukocytes/ μ l (69%), p < 0.01. A somewhat similar result was found for the concentration of bacteria. The iQ200 reported a bacterial value of 2 or more for 61% of the cases and < 2 for 85% of the controls. ROC curves were plotted for iQ200 parameters and are shown in figure 1. The count of white blood cells per microliter (WBC/ μ L) had the largest area under curve (AUC, 0.91 CI 0.87 – 0.94) and the highest discriminative value when compared to the other parameters. Optimal cutoff values were calculated for WBC/ μ L, Bacteria and ASP/ μ L and are shown in table 4.

 $^{^{}b}P < 0.05$.

[°]CI, confidence interval.



0.4

Figure 1. Receiver operating characteristic curves of different iQ200 parameters predicting UTI.

WBC/μL (AUC 0.91, CI 0.87-0.94)
 Bacteria (AUC 0.79, CI 0.73-0.85)
 ASP/μL (AUC 0.77, CI 0.71-0.82)
 Red blood cells (RBC)/μL (AUC 0.72, CI 0.66-0.79)
 Reference line

0,2

On the Y-axis sensitivity, on the X-axis 1-specificity. AUC, area under curve, CI, 95% confidence interval.

0,6

0,8

1 - Specificity

Because of the high discriminative value of WBC/µL, this parameters was subsequently divided in 3 categories using the cutoffs of 50 and 150. The first cutoff of 50 was selected by prioritizing sensitivity over specificity while maintaining a good tradeoff between both of them (sensitivity 91%, specificity 79%). The second cutoff of 74 was the optimal calculated cutoff (sensitivity 86%, specificity 82%), and the third cutoff of 150 was selected by approximately reducing the amount of false negatives by half (sensitivity 69%, specificity 89%). Cutoffs were rounded to increase clinical applicability.

Table 4. Cut-off values of iQ200 parameters and corresponding sensitivity and specificity.

	Cutoff value	Sensitivity (%)	Specificity (%)
WBC/μL (optimal calculated) WBC/μL (selected for categorization)	<74 <50 <150	86 91 69	82 79 89
Bacteria	<2	61	84
ASP/µLª	<6,250	76	70

^a ASP, all small particles.

Table 5. Variables retained after logistic regression analysis of factors independently associated with UTI and attribution of points based on β coefficients.

Variable	AOR (CI) ^a	β coefficient	Points attributed
Frequent micturition/increased urge	2.8 (1.1 – 7.3)	1.0	1
Bacterial score, ≥2	3.7 (1.3 – 10.2)	1.3	1
Dysuria	12.1 (4.5 – 32.5)	2.5	2
WBC/μL, 50 - 149	15.6 (4.1 – 59.8)	2.8	2
WBC/μL, ≥ 150	44.5 (12.1 - 164.1)	3.8	3

^a AOR, adjusted odds ratio; CI, confidence interval.

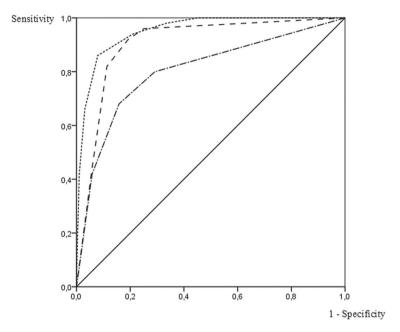
Establishment of a scoring tool

Logistic regression analysis was performed. Symptoms significantly associated with UTI were entered into the model together with the categorized concentration of WBC/ μ L (using the two selected cutoff values from table 4) and the iQ200 parameter 'bacteria'. We did not use ASP/ μ L in our model because we aimed to establish a clinically applicable model and this parameter is nonspecific for measurement of bacteria. A scoring tool was developed to confirm or rule out the diagnosis of UTI (table 5). Points were attributed based on β coefficients with 1 point being given to the parameter with the smallest coefficient. The maximum possible score was 7.

ROC curves of the scoring tool (AUC 0.95; Cl 0.93-0.98) and of its separate components, iQ200 (AUC 0.90, Cl 0.86 – 0.93) and clinical (AUC 0.80, Cl 0.73-0.88) variables, are shown in figure 2. As expected, the scoring tool had a higher sensitivity and specificity in predicting UTl as compared to the separate iQ200 and clinical parameters as derived from the model, as well as the single iQ200 parameters (figure 1).

Different cutoff scores and corresponding characteristics of the scoring tool are displayed in table 6. When using a single threshold, a score of <4 has the optimal tradeoff between sensitivity (86%) and specificity (92%) which are both remarkably high given the complexity of the study population. Using this cutoff 7 patients would incorrectly be scored as negative (14% of cases) while 23 patients would incorrectly be scored as positive (8% of controls). If two cutoffs were to be used, three categories are formed: UTI likely, UTI possible or UTI unlikely. By using one threshold of below 3 and one of 5 or more, both false negatives and false positives are reduced by more than half compared to using a single threshold of 4. However, 19% of all patients would be classified as possibly having a UTI.

Figure 2. Receiver operating characteristic curves of the scoring tool and its separate components (iQ200 and clinical variables) predicting UTI.



On the Y-axis sensitivity, on the X-axis 1-specificity. AUC, area under curve; CI, 95% confidence interval.

---- Combined score (AUC 0.95, CI 0.93-0.98)

--- WBC/μL and bacteria (AUC 0.90, CI 0.86-0.93)

---- Dysuria and frequent micturion/increased urge (AUC 0.80, CI 0.73-0.88)

---- Reference line

DISCUSSION

Our study defined cutoff values for parameters as measured by an automated urine analysis system, the IRIS iQ200, for prediction of the clinical diagnosis of urinary tract infection in a heterogeneous, academic population of adult patients. In contrast to previous research on automated urine analysis by the iQ200,67,10 we did not solely use a positive urine culture as the 'gold' standard, but focused on clinical symptoms and course of disease in combination with culture results. This clinical assessment allowed exclusion of false-positive urine cultures of patients without urinary symptoms and with a diagnosis other than UTI, limiting unnecessary treatment of UTI. Prudent use of antibiotics has become increasingly relevant because of the problem of antibiotic resistance, which currently has become one of the most serious and growing threats to public health.16

We found that urinary white blood cell count had the highest discriminative value for UTI (AUC 0.91) compared to the other individual parameters, bacterial score and ASP/ μ L. The

calculated optimal cutoff for WBC/ μ L was 74, with a sensitivity of 86% and specificity of 82%. For development of the scoring tool we choose to use a lower cutoff of 50 WBC/ μ L with a higher sensitivity of 91% and acceptable specificity of 79% (AUC 0.85), to reduce the amount of false negative results.

The finding that only 3% of the cases had a concentration of ≤ 20 WBC/ μ L in urine corresponds with previous research using conventional urine analysis. The role of the count of ASP/ μ L in UTI diagnosis by iQ200 remains to be determined. One study reported that ASP/ μ L has a better test performance than bacteria at certain cutoffs, but our findings confirm the observation of Parta et al. who did not find ASP count to contribute in ruling out UTIs.

Table 6. Possible thresholds for the scoring tool and corresponding characteristics.

Score threshold	Sensitivity (%)	Specificity (%)	Positive predictive value (%)	Negative predictive value (%)
< 2	98	67	34	100
< 3ª	94	79	44	99
< 4	86	92	65	98
< 5 ^b	66	97	79	94
< 6	42	99	88	91

^aThreshold selected to rule out UTI.

The optimal scoring tool for diagnosis of UTI obtained by multivariable analysis included the iQ200 parameters 'WBC/µL' and 'bacteria' and the clinical symptoms 'dysuria' and 'frequent micturition/ increased urge'. The test characteristics of the scoring tool depend on the chosen threshold(s). Through the selection of different cutoff criteria, the score can be adapted to different clinical situations depending on the relative benefits of maximizing sensitivity or specificity. While a high sensitivity is important to minimize the number of false negatives, specificity might be of equal importance to minimize the number of false positives and limiting inappropriate antibiotic use.

Obviously, the selection of the best cutoff depends on the setting, the clinical condition and individual characteristics of the patient, and the risk of delaying antibiotic treatment. In case of a febrile patient with suspected invasive UTI, a threshold of <2 seems appropriate to rule out UTI and search for an alternative diagnosis, whereas in case of suspected cystitis the threshold of 4 could be used to withhold antibiotics. Therefore, the use of three categories (UTI likely (\ge 5), UTI possible (3 to 4) or UTI unlikely (<3)) is probably most useful for application in patient care, leaving room for interpretation and risk analysis by the clinician.

^bThreshold selected to confirm UTI.

Table 7. Number of patients with and without UTI in each score group and predictive values using two cut-offs.

Score	With UTI (n = 50)	Without UTI (n = 291)	Positive predictive value (%)	Negative predictive value (%)
<3	3	230	1	99
3-4	14	52	21	79
>4	33	9	79	21

Previous research on automated urine analysis showed that its findings correspond well with conventional urine sedimentation. Most research on analysis by the IRIS iQ200 aimed to predict a positive urine culture in order to reduce laboratory work load and associated costs. One of these articles also took clinical data into account, which lead to a reduction of cases considered to be false positive using urine culture only as the gold standard. Since the purpose of this study by Parta was to evaluate the iQ200 as a screening tool to decrease unnecessary urine cultures, a low cutoff for WBC ($\geq 6/\mu L$) was chosen to achieve high sensitivity, resulting in a poor specificity of 67-70%.

Luciano developed a risk score combining both dipstick and iQ200 sediment reading results with age that improved UTI diagnosis in a pediatric population.¹⁹

Similar studies were performed on different commercial systems using flow cytometry, e.g. the Sysmex UF-1000i (Sysmex Japan) to define optimal cutoff points for WBC or bacteria for ruling out bacterial UTI, but these data cannot be extrapolated directly to the iQ200 because both systems work differently. The UF-1000i is laser based and uses fluorescent dye which the iQ200 does not.^{79,20,21}

Strengths of our study are the particular academic population and its reflection of a real life situation which includes a very heterogeneous group of both in- and outpatients of whom some had renal transplants, had a fever, were already treated with antibiotics, had contaminated urine samples or were difficult to classify as either having a UTI or not.

The present study has its limitations. First, data from electronic patient files was obtained retrospectively and might not always have been complete. Second, the entire available dataset was used for the prediction score, which as a result could not be validated in a different patient set. Validation is therefore required before the score could be implemented in clinical use. Third, the diagnosis of UTI lacks a gold standard. However, we feel that assessment by two independent blinded experts who take clinical data, conventional sediment analysis as well as culture results into account is the best reference test currently available. Finally, the study population was too small to distinguish between uncomplicated cystitis and invasive UTIs and to determine if cutoff values of iQ200 parameters would be different for certain subgroups, such as patients with neutropenia or renal transplants.^{2,22}

Further research should prospectively validate the scoring tool for diagnosis of UTI on a new set of data and in different subgroups of patients and demonstrate potential benefits, such as reduction in the unnecessary use of antibiotics.

In conclusion, although the diagnosis of UTI can be challenging in an adult academic patient population, the combination of a simple survey and the results of the iQ200 could rule out infection in the majority of patients and therefore improve antibiotic stewardship in suspected UTI cases.

REFERENCES

- 1. Schappert SM, Rechtsteiner EA. Ambulatory medical care utilization estimates for 2007. Vital Health Stat 13 2011:1-38.
- 2. Wilson ML, Gaido L. Laboratory diagnosis of urinary tract infections in adult patients. Clin Infect Dis 2004;38:1150-8.
- 3. Hooton TM. Clinical practice. Uncomplicated urinary tract infection. N Engl J Med 2012;366:1028-37.
- 4. van den Broek D, Keularts IM, Wielders JP, Kraaijenhagen RJ. Benefits of the iQ200 automated urine microscopy analyser in routine urinalysis. Clin Chem Lab Med 46 2008:1635–1640.
- 5. Wah DT, Wises PK, Butch AW. Analytic performance of the iQ200 automated urine microscopy analyzer and comparison with manual counts using Fuchs-Rosenthal cell chambers. Am J Clin Pathol 2005;123:290-6.
- Russcher A, Kusters E, Wolterbeek R, Kuijper EJ, Cobbaert CM, van der Beek MT. Interlaboratory Collaboration for Optimized Screening for Urinary Tract Infection. J Clin Microbiol 2016;54:93-8.
- 7. Shang YJ, Wang QQ, Zhang JR, et al. Systematic review and meta-analysis of flow cytometry in urinary tract infection screening. Clin Chim Acta 2013;424:90-5.
- 8. Sturenburg E, Kramer J, Schon G, Cachovan G, Sobottka I. Detection of significant bacteriuria by use of the iQ200 automated urine microscope. J Clin Microbiol 2014;52:2855-60.
- 9. Inigo M, Coello A, Fernandez-Rivas G, et al. Evaluation of the SediMax automated microscopy sediment analyzer and the Sysmex UF-1000i flow cytometer as screening tools to rule out negative urinary tract infections. Clin Chim Acta 2016;456:31-5.
- 10. Parta M, Hudson BY, Le TP, Ittmann M, Musher DM, Stager C. IRIS iQ200 workstation as a screen for performing urine culture. Diagn Microbiol Infect Dis 2013;75:5-8.
- 11. European Confederation of Laboratory Medicine. 2000. European urinalysis guidelines. Scand J Clin Lab Invest Suppl 231:1–86.
- 12. Dull RB, Friedman SK, Risoldi ZM, Rice EC, Starlin RC, Destache CJ. Antimicrobial treatment of asymptomatic bacteriuria in noncatheterized adults: a systematic review. Pharmacotherapy 34 2014:941–960.
- 13. Kumar R, Indrayan A. Receiver operating characteristic (ROC) curve for medical researchers. Indian Pediatr 2011;48:277-87.
- 14. Schreiber MP, Chan CM, Shorr AF. Resistant pathogens in nonnosocomial pneumonia and respiratory failure: is it time to refine the definition of health-care-associated pneumonia? Chest 2010;137:1283-8.
- 15. Shorr AF, Myers DE, Huang DB, Nathanson BH, Emons MF, Kollef MH. A risk score for identifying methicillin-resistant Staphylococcus aureus in patients presenting to the hospital with pneumonia. BMC Infect Dis 2013;13:268.
- 16. Sabtu N, Enoch DA, Brown NM. Antibiotic resistance: what, why, where, when and how? Br Med Bull 2015.
- 17. Pappas PG. Laboratory in the diagnosis and management of urinary tract infections. Med Clin North Am 1991;75:313-25.
- 18. Czaja CA, Scholes D, Hooton TM, Stamm WE. Population-based epidemiologic analysis of acute pyelonephritis. Clin Infect Dis 2007;45:273-80.
- 19. Luciano R, Piga S, Federico L, et al. Development of a score based on urinalysis to improve the management of urinary tract infection in children. Clin Chim Acta 2012;413:478-82.
- 20. Le Z, Li F, Fei C, Ye A, Xie X, Zhang J. Performance of the Sysmex UF-1000i urine analyser in the rapid diagnosis of urinary tract infections in hospitalized patients. J Infect Chemother 2016;22:377-82.
- 21. Shayanfar N, Tobler U, von EA, Bestmann L. Automated urinalysis: first experiences and a comparison between the Iris iQ200 urine microscopy system, the Sysmex UF-100 flow cytometer and manual microscopic particle counting. Clin Chem Lab Med 2007;45:1251-6.
- 22. Urabe A. Clinical features of the neutropenic host: definitions and initial evaluation. Clin Infect Dis 2004;39 Suppl 1:S53-S5.