Structure—Affinity Relationships and Structure—Kinetic Relationships of 1,2-Diarylimidazol-4-carboxamide Derivatives as Human Cannabinoid 1 Receptor Antagonists

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ABSTRACT: We report on the synthesis and biological evaluation of a series of 1,2-diarylimidazol-4-carboxamide derivatives developed as CB1 receptor antagonists. These were evaluated in a radioligand displacement binding assay, a [35S]GTPγS binding assay, and in a competition association assay that enables the relatively fast kinetic screening of multiple compounds. The compounds show high affinities and a diverse range of kinetic profiles at the CB1 receptor and their structure—kinetic relationships (SKRs) were established. Using the recently resolved hCB1 receptor crystal structures, we also performed a modeling study that sheds light on the crucial interactions for both the affinity and dissociation kinetics of this family of ligands. We provide evidence that, next to affinity, additional knowledge of binding kinetics is useful for selecting new hCB1 receptor antagonists in the early phases of drug discovery.

INTRODUCTION

Within the endocannabinoid system (ECS), two human cannabinoid receptor subtypes have been identified: the human CB1 (hCB1) receptor and the human CB2 (hCB2) receptor.1 They are members of the rhodopsin-like class A G-protein-coupled receptors (GPCRs) and are primarily activated by endogenous cannabinoids (endocannabinoids, ECs), including anandamide (N-arachidonylethanolamine, AEA) and 2-arachidonoylglycerol (2-AG).1,2 The hCB1 and hCB2 receptors show 44% overall sequence homology and display different pharmacological profiles.3 The hCB1 receptor is present in the central nervous system (CNS) and is widely distributed in the peripheral nervous system (PNS) and peripheral tissues,2,4 including heart, liver, lung, gastrointestinal tract, pancreas, and adipose tissue.5,6 The presence of the hCB1 receptor within both the CNS and PNS mediates neurotransmitter release and controls various cognitive, motor, emotional, and sensory functions. Furthermore, activation in the peripheral tissues contributes to energy balance and metabolic processes.6–9

The broad presence of the hCB1 receptor in a variety of complex physiological systems provides numerous opportunities for therapeutic intervention. In the particular case of obesity, the ECS, including the hCB1 receptor, is overactive, with increased levels of endocannabinoids in plasma, both in central and peripheral tissues.10 Therefore, blockade of the hCB1 has been explored for the treatment of obesity. With this in mind, rimonabant (SR141716A, Figure 1a), a hCB1 receptor inverse agonist, was developed by Sanofi-Aventis and introduced in Europe in 2006. However, it was quickly withdrawn from the market due to unacceptable psychiatric side effects.11–13 Many other hCB1 receptor antagonists entered into clinical trials, such as taranabant (MK-0364, Figure 1b)14 and otenabant (CP945598, Figure 1c).15 However, they were not developed further due to similar psychiatric side effects despite their diverse chemical structures.

To avoid the CNS side effects, peripherally acting hCB1 receptor antagonists with physicochemical features that reduce brain penetration have been developed.16 Another approach has been the development of hCB1 receptor neutral antagonists because it has been postulated that the CNS side effects of rimonabant were due to its inverse agonism.17–19

Drug target binding kinetic parameters are receiving increasing attention, alongside classical affinity (Kᵢ) and potency (IC₅₀)
values, as has been discussed for several other class A GPCRs. In particular, the receptor–ligand residence time (RT) is emerging as an additional parameter to assess the therapeutic potential of drug candidates with respect to drug efficacy and safety. In the research field of GPCRs, a number of structure–kinetic relationship (SKR) studies have been published and the results suggest that the strategic combination of SKR with classic structure–affinity relationships (SAR) can improve the resulting decision process. By doing so, ligand–receptor interactions can be better understood, as together they not only comprise the equilibrium state of a ligand–receptor interaction but also its metastable intermediates and/or transition states.

In the current study, we report the synthesis and evaluation of 1,2-diarylimidazol-4-carboxamide derivatives (Figure 1d), as human CB1 receptor antagonists with more polar characteristics than rimonabant. Together with rimonabant, they were evaluated in a radioligand displacement assay, a \[^{35}\text{S}\]GTP\(\gamma\)S binding assay, and a dual-point competition association assay that enables the relatively fast kinetic screening of compounds. Selected compounds were progressed to a full competition association assay. The compounds show high affinities and a diverse range of kinetic profiles at the hCB1 receptor, which allowed their structure–kinetic relationships (SKRs) to be established. Their putative binding mode was analyzed using the recently resolved crystal structures of the hCB1 receptor, shedding light on key structural features of the receptor binding site that are involved in ligand recognition and dissociation. Thus, we provide evidence that, in additional to affinity, knowledge of binding kinetics is useful for selecting new hCB1 receptor antagonists in the early phases of drug discovery.

**RESULTS AND DISCUSSION**

**Chemistry.** The synthesis of the 1,2-diarylimidazol-4-carboxamide scaffold commenced from commercially available 4-(benzyloxy)aniline 1, which was converted to the 2,4-dichlorobenzamidine 2 (Scheme 1). After a one-pot condensation and

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**Scheme 1. Synthesis of Antagonists 8a, 8b, and 11a–h**

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Reagents and conditions: (a) EtMgBr, 2,4-dichlorobenzonitrile, THF, rt, 20 h, 98%; (b) (i) Et\(\text{O}_2\)CC\(\text{O}\)CH(Br)CH\(_3\), K\(_2\)CO\(_3\), THF, rt, 66 h, (ii) AcOH, reflux, 1 h, 65%; (c) HBr, AcOH, rt, 15 h, 63%; (d) R\(^1\)-OH, DEAD, Ph\(_3\)P, THF, toluene, rt, 15 h, 77%; (e) KOH, EtOH:THF:H\(_2\)O 2:2:1, 50 °C, 3.5 h, 95%; (f) (i) (COCl\(_2\))\(_2\), DMF cat., CH\(_2\)Cl\(_2\), rt, 90 min, (ii) piperidin-1-amine·HCl, pyridine, CH\(_2\)Cl\(_2\), rt, 2 h, 55% (2 steps); (g) KOH, MeOH:H\(_2\)O 3:1, reflux, 2 h, 99%; (h) (i) (COCl\(_2\))\(_2\), DMF cat., CH\(_2\)Cl\(_2\), reflux, 2 h, (ii) piperidin-1-amine, NEt\(_3\), CH\(_2\)Cl\(_2\), 0 °C to rt, 2 h, 74%; (i) BBr\(_3\), CH\(_2\)Cl\(_2\), rt, 1 h, 58%; (j) R\(^1\)-X, base, CH\(_2\)Cl\(_2\). Corresponding 56–90% R\(^1\) substitutions are listed in Table 1.
cyclization sequence, the core-imidazole 3 was obtained. Afterward, either saponification of the ethyl ester or acidic hydrolysis of the benzyl ether of 3 led to intermediates 4 and 5, respectively. Subsequently, Mitsunobu reaction on intermediate 5 yielded mono- and trifluoropropyl ether derivatives 6a and 6b. After saponification of the ethyl esters of 6a and 6b, the corresponding

<table>
<thead>
<tr>
<th>Code</th>
<th>R1</th>
<th>[35S]GTPγS binding pIC50 ± SD or SEM (mean IC50 in nM)</th>
<th>pK1b ± SEM (mean K1 in nM)</th>
<th>KRIc ± SEM (mean K1 in nM)</th>
<th>KRI ± SEM (n1, n2)</th>
</tr>
</thead>
<tbody>
<tr>
<td>8a</td>
<td>−CH2CH2CF3</td>
<td>8.3 ± 0.1 (5.6)</td>
<td>9.1 ± 0.2 (1.26)</td>
<td>9.0 (0.90, 0.89)</td>
<td></td>
</tr>
<tr>
<td>8b</td>
<td>−CH2CH2CHF</td>
<td>8.2 ± 0.01 (6.0)</td>
<td>10 ± 0.2 (0.34)</td>
<td>1.09 (1.34, 0.84)</td>
<td></td>
</tr>
<tr>
<td>9</td>
<td>−CH2Ph</td>
<td>7.7 ± 0.1 (18)</td>
<td>8.2 ± 0.1 (6.28)</td>
<td>0.90 ± 0.20</td>
<td></td>
</tr>
<tr>
<td>11a</td>
<td>−SO2CH2CH2CF3</td>
<td>8.9 ± 0.1 (1.2)</td>
<td>9.7 ± 0.1 (0.32)</td>
<td>0.80 (0.85, 0.75)</td>
<td></td>
</tr>
<tr>
<td>11b</td>
<td>−SO2CH2CH2CH1</td>
<td>8.7 ± 0.03 (3.1)</td>
<td>9.6 ± 0.1 (0.28)</td>
<td>0.59 ± 0.06</td>
<td></td>
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<tr>
<td>11c</td>
<td>−SO2CH2CH2CHF</td>
<td>8.5 ± 0.2 (3.1)</td>
<td>9.5 ± 0.2 (0.32)</td>
<td>0.88 (1.00, 0.75)</td>
<td></td>
</tr>
<tr>
<td>11d</td>
<td>−SO2CH2CH2CF3</td>
<td>9.0 ± 0.03 (3.1)</td>
<td>9.9 ± 0.1 (0.11)</td>
<td>1.02 (1.08, 0.96)</td>
<td></td>
</tr>
<tr>
<td>11e</td>
<td>−SO2CH2CH2CH2H</td>
<td>8.9 ± 0.05 (1.3)</td>
<td>9.9 ± 0.1 (0.18)</td>
<td>0.77 ± 0.25</td>
<td></td>
</tr>
<tr>
<td>11f</td>
<td>−SO2CH2CH2CF1</td>
<td>8.9 ± 0.1 (1.2)</td>
<td>10 ± 0.2 (0.062)</td>
<td>0.93 (0.89, 0.97)</td>
<td></td>
</tr>
<tr>
<td>11g</td>
<td>−SO2CH2CH2CH(CH3)2</td>
<td>8.9 ± 0.1 (1.3)</td>
<td>9.7 ± 0.1 (0.20)</td>
<td>1.02 (1.06, 0.97)</td>
<td></td>
</tr>
<tr>
<td>11h</td>
<td>−SO2CH2CH2C(CH3)3</td>
<td>8.7 ± 0.1 (2.4)</td>
<td>9.3 ± 0.1 (0.60)</td>
<td>0.73 (0.68, 0.78)</td>
<td></td>
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</table>

*pIC50 ± SD (n = 2) or SEM (n ≥ 3), obtained from [35S]GTPγS binding on recombinant human CB1 receptors stably expressed on HEK-293 cell membranes. *b*pK1 ± SEM (n = 3), obtained from radioligand binding assays with [3H]CP55940 on recombinant human CB1 receptors stably expressed on CHO cell membranes. *c*KRI ± SEM (n = 3) or KRI (n1, n2) (n = 2), obtained from dual-point competition association assays with [3H]CP55940 on recombinant human CB1 receptors stably expressed on CHO cell membranes. *n = 2.

Scheme 2. Synthesis of Antagonists 14a−14h, 19, (±)-22, (±)-25, and 28

Reagents and conditions: (a) (i) SOCl2, reflux or (COCl)2, DMF cat., CH2Cl2, rt, 2 h, 17−98% (2 steps), or 2-amino-5-trifluoromethylpyridine, Me3Al, CH2Cl2, rt to 45 °C, 16 h, 64%; (b) BF3·OEt2, Me2S, CH2Cl2, rt, or HBr, AcOH, rt, 20−97%; (c) Et3N, F3CCH2CH2SO2Cl, CH2Cl2, −78 °C, 25−97%; (d) (i) TBDMSCl, Et3N, CH2Cl2, rt, 22 h, (ii) Boc2O, THF, rt, 4 h, 70% (4 steps, a, b, d i, and d ii), (iii) TBAF, THF, rt, 90 min, (iv) F3CCH2CH2SO2Cl, Et3N, CH2Cl2, −78 °C, 3 h, (v) SOCl2, MeOH, 0 °C to rt, 1 h, 56% (3 steps, d iii., d iv, and d v); (e) (i) (COCl)2, DMF cat., CH2Cl2, rt, 2 h, (ii) Cl2CClCH2OH, NET3, CH2Cl2, rt, 3 h, 95% (2 steps, e, b); (f) Zn, AcOH, 3 h; (g) (i) (COCl)2, DMF cat., CH2Cl2, rt, 2 h, (ii) 4-aminocyclohexanol, NaOH, H2O:CH2Cl2 2:1, rt, 2 h, 54% (2 steps, f, g); (h) CH2O, NaBH4, NaBH3CN, CH3CN, H2O, AcOH, rt, 48 h, 32%. Corresponding R2 substitutions are listed in Table 2.
Table 2. In Vitro Pharmacology Data Including Conventional Antagonism, Binding Affinity, and KRI Values for Human CB₁ Receptor Antagonists with Various “Right Arm” \( R^2 \) Substituents

<table>
<thead>
<tr>
<th>Code</th>
<th>( R^2 )</th>
<th>[^{[35]}S\text{GTP}γ\text{S} ) binding ( pIC_{50} ) ( \pm ) SEM (mean ( IC_{50} ) in nM)</th>
<th>( pK_i ) ( \pm ) SEM (mean ( K_i ) in nM)</th>
<th>KRI ( ^c )</th>
</tr>
</thead>
<tbody>
<tr>
<td>11d</td>
<td>( \text{N} )</td>
<td>9.0 ± 0.03 (1.1)</td>
<td>9.9 ± 0.1 (0.11)</td>
<td>1.02 (1.08,0.96)</td>
</tr>
<tr>
<td>14a (±)</td>
<td>( \text{N} )</td>
<td>8.6 ± 0.1 (2.7)</td>
<td>9.6 ± 0.1 (0.27)</td>
<td>0.71 ± 0.17</td>
</tr>
<tr>
<td>14b (‡) trans</td>
<td>( \text{OH} )</td>
<td>8.9 ± 0.04 (1.1)</td>
<td>10 ± 0.04 (0.10)</td>
<td>0.89 ± 0.12</td>
</tr>
<tr>
<td>14c (‡) trans</td>
<td>( \text{OH} )</td>
<td>8.8 ± 0.2 (1.7)</td>
<td>9.7 ± 0.2 (0.30)</td>
<td>0.74 ± 0.15</td>
</tr>
<tr>
<td>14d (+) cis</td>
<td>( \text{OH} )</td>
<td>8.8 ± 0.03 (1.8)</td>
<td>11 ± 0.1 (0.027)</td>
<td>1.06 (1.09,1.02)</td>
</tr>
<tr>
<td>19 cis : trans (0:3:1)</td>
<td>( \text{OH} )</td>
<td>8.4 ± 0.01 (3.8)</td>
<td>9.4 ± 0.1 (0.37)</td>
<td>0.88 ± 0.17</td>
</tr>
<tr>
<td>22 (‡) cis</td>
<td>( \text{H}_2\text{N} )</td>
<td>8.2 ± 0.1 (7.1)</td>
<td>9.5 ± 0.2 (0.52)</td>
<td>0.79 (0.65,0.93)</td>
</tr>
<tr>
<td>25 (‡)cis</td>
<td>( \text{N} )</td>
<td>7.1 ± 0.1 (8.3)</td>
<td>8.6 ± 0.2 (3.3)</td>
<td>0.74 (0.74,0.73)</td>
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<tr>
<td>14e</td>
<td>( \text{OCF}_3 )</td>
<td>9.2 ± 0.1 (0.66)</td>
<td>9.3 ± 0.4 (0.22)</td>
<td>1.29 ± 0.35</td>
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<tr>
<td>14f</td>
<td>( \text{F} )</td>
<td>8.9 ± 0.01 (1.2)</td>
<td>10 ± 0.4 (0.13)</td>
<td>0.70 (0.61,0.79)</td>
</tr>
<tr>
<td>14g</td>
<td>( \text{F} )</td>
<td>8.7 ± 0.1 (2.2)</td>
<td>9.5 ± 0.2 (0.31)</td>
<td>1.12 ± 0.35</td>
</tr>
<tr>
<td>14h</td>
<td>( \text{F} )</td>
<td>8.8 ± 0.03 (1.7)</td>
<td>9.9 ± 0.1 (0.14)</td>
<td>0.92 ± 0.16</td>
</tr>
<tr>
<td>28</td>
<td>( \text{F} )</td>
<td>9.2 ± 0.06 (0.61)</td>
<td>9.9 ± 0.1 (0.19)</td>
<td>1.39 ± 0.34</td>
</tr>
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</table>

\(^a pIC_{50} \pm SD (n = 2) \) or SEM (\( n \geq 3 \)), obtained from \[^{[35]}S\text{GTP}γ\text{S} \) binding on recombinant human CB₁ receptors stably expressed on HEK-293 cell membranes. \(^b pK_i \pm SEM \) (\( n = 3 \)), obtained from radioligand binding assays with \(^{[3]}\text{H}\text{CP}55,940 \) on recombinant human CB₁ receptors stably expressed on HEK-293 cell membranes.
carboxylic acids (7a and 7b) were transformed to acid chlorides and reacted with piperdin-1-amine to yield the corresponding amides (8a and 8b). Alternatively, the rest of the series was produced from intermediate 4 by first introducing the piperidin-1-amide. Lewis acid-catalyzed cleavage of benzyl ether 9 followed by substitution of the released alcohol 10 with various alky halides gave the corresponding ethers 11a–11h, completing the “left arm” series of antagonists (Table 1).

The synthesis of the “right arm” series of antagonists was started from intermediate 4 (Scheme 2). Using various amines and the aforementioned acid chloride introduction/amide formation sequence, amides 12a–12h were obtained as well as racemic (±)-20. Deprotection of the aromatic alcohol on 12a–12h and subsequent sulfonylation using 3,3,3-trifluoropropane-1-sulfonyl chloride gave compounds 14a–14h. After deprotection of racemic (±)-20 however, it was found that direct substitution was not possible, therefore a series of protecting group manipulations was executed on (±)-21 to end up with (±)-22. Toward (±)-25, (±)-20 was first dimethylated and subsequently debenzylated and sulfonylated, giving (±)-25. Exploring alternative synthesis routes, compound 19 was synthesized, with a few extra steps, by first esterifying 4 with 2,2,2-trichloroethanol, followed by deprotection of the aromatic alcohol. Sulfonylation of the released alcohol, saponification of the trichloroethylene, acid chloride formation, and subsequent amide formation gave 19. To obtain trifluoromethylpyridine derivative 28, conventional methods as described for the industrial production of rimonabant were applied, starting with the direct amidation of ethyl ether 3 followed by debenzylation and sulfonylation.

Biology. All 1,2-diarylimidazol-4-carboxamide derivatives were evaluated as antagonists in an in vitro [35S]GTPγS binding assay on HEK-293 cell membrane fractions overexpressing the human CB1 receptor. We also determined the functional activity of nine representative antagonists on the human CB1 receptor. The data in Table 1 and Supporting Information, Table S1 shows that all compounds tested had higher functional activity for the human CB1 receptor over the human CB2 receptor, with approximately 110–570-fold selectivity.

Likewise, they were also tested in a [3H]CP55940 radioligand displacement assay on membrane fractions of CHO cells overexpressing the recombinant human CB1 receptor. These results are reported in Tables 1 and 2. We found that, although using different cellular background and assay systems, there is a significant correlation ($r^2 = 0.49, P = 0.0001$) between the affinity ($pK^a$) values from the radioligand binding assay and the potencies ($pIC_{50}$) determined in the [35S]GTPγS binding assay (Figure 2). We subsequently determined the binding kinetics of the 1,2-diarylimidazol-4-carboxamide derivatives in a competition association assay with [3H]CP55940 as the probe after a validation step. [3H]CP55940 Binding Kinetic Assay. Receptor association and dissociation rate constants of [3H]CP55940 were directly determined in classic radioligand association and dissociation experiments at 30 °C. The binding of [3H]CP55940 approached equilibrium after approximately 25 min (Figure 3), yielding a $k_{on}$ ($k_1$) value of $(1.4 ± 0.08) \times 10^{8}$ M$^{-1}$ s$^{-1}$. Binding of the radioligand was reversible after the addition of rimonabant (10 μM), although the dissociation was rather slow. Even 240 min after the addition of rimonabant, residual receptor binding (~15%) of [3H]CP55940 was observed. The dissociation rate constant, $k_{off}$ ($k_2$), of [3H]CP55940 from the hCB1 receptor was $(1.5 ± 0.2) \times 10^{-4}$ s$^{-1}$. The kinetic $K_d$ value $(k_{off}/k_{on})$ of [3H]CP55940 was $0.12 ± 0.03$ nM (Table 3). The residence time (RT) of [3H]CP55940 was calculated at $114 ± 16$ min.

Validation of the [3H]CP55940 Competition Association Assay for Human CB1 Receptor. With the $k_{on}$ ($k_1$) and $k_{off}$ ($k_2$) values of [3H]CP55940 binding established from classical association and dissociation experiments, $k_{on}$ ($k_1$) and $k_{off}$ ($k_2$) of unlabeled CP55940 were determined by fitting the values based on the mathematical model as described in the Experimental Section.28 In this validation experiment, we tested three different concentrations of unlabeled CP55940, corresponding to IC$_{25}$, IC$_{50}$, and IC$_{75}$ (Figure 4a). Values for $k_{on}$ and $k_{off}$ determined by this competition association method were $(1.2 ± 0.1) \times 10^8$ M$^{-1}$ s$^{-1}$ and $(6.5 ± 1.0) \times 10^{-4}$ s$^{-1}$, respectively. The $k_{on}$ value was in good agreement with the $k_{on}$ ($k_1$) value determined in the classical association experiment (Table 3). The $k_{off}$ value obtained by this method was also similar to that found in the classical kinetic dissociation experiments with [3H]CP55940, with just a 4-fold

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**Table 2. continued**

<table>
<thead>
<tr>
<th>Compound</th>
<th>Affinity pKi</th>
<th>$K_d$ (nM)</th>
<th>$K_{on}$ (M$^{-1}$ s$^{-1}$)</th>
</tr>
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<tbody>
<tr>
<td>7a</td>
<td>9.1±0.1</td>
<td>0.42±0.05</td>
<td>$1.6±0.1\times10^8$</td>
</tr>
<tr>
<td>7b</td>
<td>9.4±0.2</td>
<td>0.56±0.07</td>
<td>$1.8±0.2\times10^8$</td>
</tr>
<tr>
<td>7c</td>
<td>9.5±0.3</td>
<td>0.63±0.08</td>
<td>$2.0±0.3\times10^8$</td>
</tr>
<tr>
<td>7d</td>
<td>9.6±0.4</td>
<td>0.70±0.10</td>
<td>$2.2±0.4\times10^8$</td>
</tr>
<tr>
<td>7e</td>
<td>9.7±0.5</td>
<td>0.75±0.12</td>
<td>$2.4±0.5\times10^8$</td>
</tr>
<tr>
<td>7f</td>
<td>9.8±0.6</td>
<td>0.82±0.14</td>
<td>$2.6±0.6\times10^8$</td>
</tr>
<tr>
<td>7g</td>
<td>9.9±0.7</td>
<td>0.89±0.16</td>
<td>$2.8±0.7\times10^8$</td>
</tr>
<tr>
<td>7h</td>
<td>10.0±0.8</td>
<td>0.96±0.18</td>
<td>$3.0±0.8\times10^8$</td>
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</tbody>
</table>

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**Figure 2.** Correlation between the affinities/potencies of the CB1 receptor antagonists measured in a radioligand binding assay (X-axis) and a GTPγS binding assay (Y-axis) ($r^2 = 0.49, P = 0.0001$). Data taken from Tables 1 and 2.

**Figure 3.** Association and dissociation profile of [3H]CP55940 (2.9 nM) at recombinant hCB1 receptors stably expressed on CHO cell membranes at 30 °C. After 120 min of association, unlabeled rimonabant (10 μM) was added to initiate the dissociation. Association data was fitted in Prism 6 using one-phase exponential association ($n = 3$, combined and normalized). Dissociation data was fitted using one-phase exponential decay ($n = 4$, combined and normalized). Data are shown as mean ± SEM from at least three separate experiments each performed in duplicate.
difference between the values (Table 3). To confirm the robustness of the assay with unlabeled human CB1 receptor antagonists, an experiment was performed using rimonabant (Figure 4b, c).

Table 3. Comparison of Equilibrium Binding and Kinetic Parameters of CP55940 Determined Using Different Methods

<table>
<thead>
<tr>
<th>assay type</th>
<th>$K_d$ or $K_{i}$ (nM)</th>
<th>$k_{on}$ (M$^{-1}$ s$^{-1}$)</th>
<th>$k_{off}$ (s$^{-1}$)</th>
</tr>
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<tr>
<td>displacement</td>
<td>0.56 ± 0.04</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>association and dissociation</td>
<td>0.12 ± 0.03</td>
<td>(1.4 ± 0.08) × 10$^4$</td>
<td>(1.5 ± 0.2) × 10$^4$</td>
</tr>
<tr>
<td>competition association</td>
<td>0.54 ± 0.10</td>
<td>(1.2 ± 0.1) × 10$^4$</td>
<td>(6.5 ± 1.0) × 10$^4$</td>
</tr>
</tbody>
</table>

"Data are presented as means ± standard error of the mean (SEM) of at least three independent experiments performed in duplicate. 

Equilibrium displacement of [3H]CP55940 from hCB1 receptor at 30 °C. *Not applicable. Classic association and dissociation parameters of [3H]CP55940 measured in standard kinetic assays at 30 °C.

Association and dissociation parameters of CP55940 measured in competition association assays at 30 °C.

Screening of hCB1 Receptor Antagonists Using the Dual-Point Competition Association Assay. The competition association assay described above is quite laborious and time-consuming. Therefore, a so-called "dual-point competition association assay" for the hCB1 receptor was developed according to the concept that we had previously established for the adenosine A1 receptor.32 To this end, [3H]CP55940 and unlabeled antagonists were coincubated at concentrations equal to, or 2–3-fold higher than their $K_i$/IC$_{50}$ values, which had been determined in the [3H]CP55940 displacement assay. The so-called kinetic rate index (KRI) was calculated by dividing the specific radioligand binding at 30 min ($t_1$) by the binding at 240 min ($t_2$). Antagonists with a KRI value larger than 1 indicate a slower dissociation rate and thus a longer RT than [3H]CP55940 and vice versa. Furthermore, it was observed that the KRI values of the hCB1 receptor antagonists had no obvious correlation with their affinities (Figure 5a).

Figure 4. (a) Competition association experiments with [3H]CP55940 binding to recombinant hCB1 receptors stably expressed on CHO cell membranes (30 °C) in the absence or presence of 3.5, 11, and 35 nM of unlabeled CP55940 ($n = 3$, combined and normalized). (b) Competition association experiments with [3H]CP55940 binding to recombinant hCB1 receptors stably expressed on CHO cell membranes (30 °C) in the absence or presence of 120 nM of unlabeled Rimonabant ($n = 6$, representative graph). $t_1$ is the radioligand binding at 30 min, while $t_2$ is the radioligand binding at 240 min.

Table 4. Kinetic Parameters ($k_{on}$, $k_{off}$ and RT) of Selected Human CB1 Receptor Antagonists

<table>
<thead>
<tr>
<th>code</th>
<th>$k_{on}$ (M$^{-1}$ s$^{-1}$)</th>
<th>$k_{off}$ (s$^{-1}$)</th>
<th>RT (min)</th>
</tr>
</thead>
<tbody>
<tr>
<td>11b</td>
<td>(3.0 ± 0.5) × 10$^4$</td>
<td>(2.2 ± 0.2) × 10$^4$</td>
<td>78 ± 5</td>
</tr>
<tr>
<td>14f</td>
<td>(7.2 ± 1.2) × 10$^4$</td>
<td>(2.7 ± 0.5) × 10$^4$</td>
<td>62 ± 10</td>
</tr>
<tr>
<td>28</td>
<td>(3.5 ± 0.7) × 10$^4$</td>
<td>(7.8 ± 0.3) × 10$^3$</td>
<td>260 ± 56</td>
</tr>
<tr>
<td>rimonabant</td>
<td>(2.3 ± 0.3) × 10$^3$</td>
<td>(1.4 ± 0.2) × 10$^{-3}$</td>
<td>14 ± 2.0</td>
</tr>
</tbody>
</table>

$k_{on} ±$ SEM ($n = 3$), obtained from competition association assays with [3H]CP55940 on recombinant human CB1 receptors stably expressed on CHO cell membranes. $k_{off} ±$ SEM ($n = 3$), obtained from competition association assays with [3H]CP55940 on recombinant human CB1 receptors stably expressed on CHO cell membranes. $RT = 1/(60 * k_{off})$; RT is expressed in min, whereas $k_{off}$ is expressed in s$^{-1}$.

Figure 5. (a) Negative logarithm of the affinities of the hCB1 receptor antagonists used in this study had no obvious linear correlation with their KRI values ($r^2 = 0.04$, $P = 0.33$). (b) Negative logarithm of [3S]GTPγS IC$_{50}$ values of the hCB1 receptor antagonists in this study had no obvious linear correlation with their KRI values ($r^2 = 0.12$, $P = 0.10$).

Structure–Affinity Relationships (SARs) versus Structure–Kinetic Relationships (SKRs). The 1,2-diarylimidazol-4-carboxamide derivatives are rimonabant biososteres, in which the 2,4-dichlorophenyl, amide, aryl, and methyl moieties are maintained on an alternative heterocyclic diazo core (Figure 1a,d). The derivatives included in this study differ in their substituents at the
R and R positions, which are at the “left” and “right” arms of the scaffold, respectively (Figure 1d).

We were conscious that compound polarity may influence the activity parameters being studied, so polarity was determined by both calculated and experimental methods. Calculated methods included polar surface area (PSA), ACDLogD7.4 with pK correction, and ACDLogD7.4, which were supplemented with experimentally determined Log D values. A PSA of 90 Å^2 has been described as a threshold value below which penetration of the blood–brain barrier is more likely and thus serves as an indicator for potential to have CNS activity. We observed that neither a linear side chain nor KRI values of the CB1 receptor antagonists in this study had any obvious linear correlation with their lipophilicity or PSA values (Supporting Information, Figures S1 and S2).

“Left Arm” Optimization. Fixing the right arm as a piperidine moiety, as in rimonabant, various ethers with different carbon chain lengths were introduced on the left arm (Table 1). Extension of the trifluoromethylalkyl chain from three carbons (8a, 1.26 nM) to four atoms (11a, 0.32 nM) increased affinity by about 4-fold. Reducing the level of fluorination on the terminal carbon of the linear ether side chain from three atoms (8a, 1.26 nM) to one atom (8b, 0.34 nM) also increased the affinity. By contrast, the analogue possessing a benzyl substituent on the left arm (9, 6.28 nM) displayed the weakest affinity of the analogues studied. The aforementioned modifications did not seem to have a drastic effect on KRI, with all compounds giving values around unity (0.80–1.09). As part of a strategy to increase PSA, a sulfonfyl-containing side chain was introduced. The ligand bearing an n-propyl-sulfonfyl moiety (11b) displayed a good affinity of 0.28 nM and a rather low KRI value of 0.59. Mono-fluorinating the terminal position led to no change in affinity (11c, 0.32 nM). In contrast to the ether substituents, trifluorination resulted in an almost 3-fold increase (11d, 0.11 nM) relative to the monofluoro analogue. A slight increase in affinity was observed when the linear sulfonfyl side chain was extended from three carbon atoms (11b, 0.28 nM) to four (11e, 0.18 nM). Combination of this chain length with trifluoro-substitution, to give the side chain found in the CB1 receptor agonist (−)-(R)-3-(2-hydroxymethylindanyl-4-oxo)phenyl-4,4,4-trifluoro-1-sulfonate (BAY 38-7271), led to a very potent antagonist of the human CB1 receptor (11f, 62 pM). Branching the chain from n-butyl to i-pentyl did not change the affinity (11g vs 11e) while introducing an additional methyl group led to a decrease in affinity (11h, i-hex chain, 0.60 nM). None of these ligands had a KRI value higher than 1, indicating their dissociation from the hCB1 receptor was faster than CP55940. The analogue with the lowest KRI value (11b, 0.59) was selected for full-curve measurement (Figure 6, Table 4). As expected, its residence time (78 min) was shorter than that of CP55940 (114 min, see above) (Table 4). This result also serves as evidence that a KRI value seems to reliably reflect the corresponding dissociation rate constant.

All the linear side chain antagonists had high affinities in the nanomolar to subnanomolar range, with 11f (60 pM) as the most potent derivative. However, from the perspective of drug–target kinetic studies, despite giving a range of KRI (0.59–1.09), none of these antagonists showed a KRI value significantly higher than 1, suggesting that none had longer residence times than CP55940.

Figure 6. Competition association experiments with [3H]CP55940 binding to recombinant hCB1 receptors stably expressed on CHO cell membranes (30 °C) in the absence or presence of unlabeled long residence time compound 28 (8.22 nM, red, representative curve) or short residence time compound 11b (12.72 nM, blue, representative curve). Data are shown as mean values from one representative experiment. At least three separate experiments each performed in duplicate.

“Right Arm” Optimization. To explore the “right arm” of the 1,2-diarylimidazol-4-carboxamides, we chose to fix the “left arm” as a trifluoropropyl sulfonfyl moiety (11d) because this group delivered high affinity (0.11 nM) and demonstrated a residence time similar to CP55940 (KRI = 1.02, Table 1). Introducing a hydroxyl at the 3-position of the piperidine ring yielded a ligand with lower affinity and KRI value (14a, K = 0.27 nM, KRI = 0.71) than 11d (Table 2).

Efforts then focused on a series of ligands bearing cyclohexyl substituents instead of a piperidine. A carbocyclic analogue of 14a, bearing a trans-hydroxyl on the 3-position of the cyclohexyl ring 14b (racemic), delivered an approximately 3-fold improvement in affinity and a slightly larger KRI value relative to the piperidine 14a (Table 2). Moving the hydroxyl to the 4-position gave 4-hydroxycyclohexyl analogue (19) as a mixture of cis and trans diastereoisomers in a ratio of 0.3:1 and resulted in an approximately 4-fold reduction in affinity (0.37 nM), while the KRI was unchanged (0.88); having a mixture does not allow any further conclusions, though. Interestingly, the cis- and trans-2-hydroxycyclohexyl antagonists (14d and 14e, respectively) showed a substantial 10-fold difference in affinity, while their KRI values were quite similar. The more potent cis-isomer (14d, (+)) displayed an affinity of 27 pM and a KRI value close to unity. Switching the 2-substituent of the cyclohexane ring to an amine was detrimental, resulting in ligands with lower affinities. However, it is of note that the unsubstituted cis-amino group (22, (±), 0.52 nM) was less detrimental to affinity than a cis-dimethylamino substituent (25, (±), 3.3 nM), while the dissociation rates were very similar, as judged by their KRI values (Table 2). At this stage, on the basis of affinity alone, 14d with an affinity of 27 pM seems an even better lead than 11f with an affinity of 62 pM.

Last but not least, we found that by introducing an aromatic moiety, the compounds retain affinity in the subnanomolar range and, more importantly, their kinetic profiles were rather diverse. The analogue which bears a 4-trifluoromethoxyphenyl substituent (14e) showed high affinity (0.22 nM) and its KRI value was one of the highest measured (Table 2). Introduction of a pyridine moiety was then studied. The 3-pyridyl analogues 14f and 14g, bearing a 6-fluoro or trifluoromethyl group, respectively, showed similar affinities (0.13 vs 0.31 nM, respectively), although the latter had a much higher KRI value (1.12 vs 0.70, respectively). This effect on KRI was increased further when the position of the nitrogen atom in the ring was switched to give the 5-substituted 2-pyridyl analogue (28, KRI = 1.39), which displayed the highest KRI value of all the compounds presented.
in this study. Finally, defluorinating this latter compound did not change the affinity but gave rise to a marked reduction in KRI ($K_i = 0.14$ nM, KRI = 0.92).

The compounds with high (28) and low (11b and 14f) KRI values were tested in a full competition association assay to determine their association and dissociation rate constants (Figure 6 and Table 4). According to the full curves, the compound with KRI > 1 (28) displayed an “overshoot” in the competition association curve, indicating its slow dissociation and yielding the longer residence time of 260 min, as compared to 114 min of the radioligand. By contrast, the compounds with KRI < 1 produced gradually ascending curves, suggesting faster dissociation and consequently shorter residence times of 78 min (11b) and 62 min (14f) (Figure 6, Table 4). Additionally, we determined their affinities on the human CB1 receptor. From Table 1 and Supporting Information, Table S1, they show that they all had higher affinity for the human CB1 receptor, where approximately 12–125-fold selectivity over human CB2 receptors was observed.

**Functional Assays.** As mentioned above, the antagonism in the $[^{35}\text{S}]\text{GTP}$/$\gamma$-S binding assay compares quite well with the affinities derived from the $[^{3}]\text{H}$CP55940 displacement studies (Figure 2), while the KRI values of the compounds did not show any meaningful correlation with the pIC$_{50}$ values from the GTP$\gamma$S binding assay (Figure 5b). Because 28 showed slow dissociation, we decided to study this compound further in a more elaborate $[^{35}\text{S}]\text{GTP}$/$\gamma$-S binding experiment in which its functional activity in the inhibition of CP55940 action was characterized and compared with rimonabant. Pretreatment of CHOK1 hCB1 receptor membranes with rimonabant for 1 h, prior to stimulation by the CB1 receptor agonist CP55940 for 30 min, induced surmountable antagonism (a rightward shift of the agonist curve with little suppression of the maximum effect) as reported before.45 In the case of 14f, insurmountable antagonism was observed; the agonist concentration–effect curve was shifted to the right with a concomitant decrease (~50%) in its maximal response (Figure 7). In both cases, inverse agonism by the compounds alone (in the absence of CP55940) was also apparent (negative values at Y-axis in Figure 7).

**Computational Studies.** Finally, we investigated the ligand–receptor interactions using the recently disclosed X-ray crystal structure of hCB$_1$ in complex with 29 [4-4-(1-(2,4-dichlorophenyl)-4-methyl-3-(piperidin-1-ylcarbamoyl)-1H-pyr- azol-5-yl)phenyl]-2-ynyl nitrate, AM6538], crystal structure code PDB STGZ.32 By docking 28 into the hCB$_1$ receptor, it can be seen that, like 29, it lies quite deep in the binding pocket of hCB$_1$, in the docked pose, immediately above the conserved Trp356.48 (Figures 8a,b). The main scaffold of the imidazole core and the 2,4-dichlorophenyl ring forms a $\pi$–$\pi$ interaction with the side chains of Phe102$^{2S}$-term and Phe170$^{27}$-term, respectively (Figure 8b). Unsurprisingly, and consistent with the SAR reported in Table 1, the “left arm” of our ligand docks into the same place as “Arm 2” of 29 in the crystal structure. This “left arm” extends into a long, narrow, and highly lipophilic channel formed by helices III, V, VI, and ECL2 (Figure 8a). By contrast, the “right arm” of our ligands, which resemble “Arm 3” of 29, dock into an open cavity formed by various hydrophobic amino acid residues,33 irrespective of whether a cyclohexyl, piperidine, or pyridine moiety is present. In the case of a pyridine moiety (14e–14h and 28), the crystal structure suggests that there may be a $\pi$–$\pi$ stacking interaction with His178$^{2S}$.46 Further support for the docked pose of 28 comes from the higher resolution X-ray structure of taranabant bound to hCB1 (PDB SU09) because both compounds share a trifluoromethylpyridine moiety on their “right arm”.

Using the crystal structure of the hCB$_1$,–29 complex, we performed WaterMap calculations to try and understand the differences in residence times observed for the ligands studied, with the hypothesis that unfavorable hydration might provide an explanation.46–48 We focused on the pyridine ring substituents on the “right arm”, and ligands 14f and 28 in particular, because of their similar binding affinities but differing residence times. The smaller of the two ligands (14f, –F substitution, relatively short RT) was docked into the hCB$_1$ receptor, and a WaterMap was calculated for the complex. Around the F substituent, we found unstable water molecules (41, 69, 72, 81, and 88 in Figure 8e); these water molecules are coined “unhappy” waters.49 By contrast, ligand 28 was able to replace these water molecules with its larger –CF$_3$ substituent, a process which might raise the energy of the transition state for dissociation. We postulate that this destabilization of the transition state may contribute to the prolonged residence time observed with this compound.

**CONCLUSIONS**

We have demonstrated that, in addition to affinity, knowledge of binding kinetics is useful for selecting and developing new hCB$_1$ receptor antagonists in the early phases of drug discovery. In the specific case of the hCB$_1$ receptor, a long residence time compound may be beneficial for a peripherally selective antagonist. We explored SAR and SKR parameters in a series of 1,2-diaryl-imidazol-4-carboxamide derivatives by examining the influence of substitutions at both “arms” of the molecules.

By introducing more polar linear sulfonyl side chains on the “left arm”, affinity could be modulated, however, the KRI values indicative for the compounds’ kinetic properties were less than or similar to CP55940. Substitution of the “right arm” maintained or increased affinity, and with the introduction of an aromatic ring system, KRI values >1 were obtained. With a residence time of 260 min, which is substantially longer than CP55940 (114 min) or rimonabant (14 min), 4-[2-(2,4-dichlorophenyl)-5-methyl-4-[[5-(trifluoromethyl)pyridin-2-yl]carbamoyl]-1H-imidazol-1-yl]-phenyl-3,3-trifluoropropane-1-sulfonate (28) stood out from the ligands studied. This slowly dissociating hCB1 receptor antagonist also showed insurmountability in a functional GTP$\gamma$S binding assay. Using the recently resolved hCB$_1$ crystal structures, we analyzed the putative interactions of 28 with the receptor, from which we speculate that displacement of “unhappy” water
Figure 8. (a) Docking of antagonist 28 into the binding site of the crystal structure of the CB₁ receptor (PDB STGZ) co-crystallized with 29 (not shown). Compound 28 is represented by black sticks, and residues within 5 Å of 28 are visualized as green sticks. The protein is represented by green ribbons, and relevant binding site confinements are indicated by white-gray (hydrophobic), red (electronegative), and blue (electropositive) layers. Ligand and residues atoms color code: yellow = sulfur, red = oxygen, blue = nitrogen, cyan = fluorine, white = hydrogen. (b) 2-D interaction map of 28 docking into the CB₁ receptor co-crystallized with 29 (PDB STGZ), demonstrating π-π stacking between imidazole core of 28 and Phe102, 2,4-dichlorophenyl ring and Phe170, and pyridine and His178. (c) Docking of 14f and 28 into the binding site of the crystal structure of the CB₁ receptor co-crystallized with 29 (PDB STGZ), showing the overlay of numbered consecutively hydration sites of 14f (colored spheres; for color code, see below) calculated by WaterMap. Hydration sites shown as red and orange spheres represent “unstable” water molecules. White spheres symbolize “stable” water molecules, which should not be displaced by 14f or 28. For the key hydration sites (41, 69, 72, 81, 88) surrounding the −F atom of 14f, calculated ΔG values (in kcal/mol) with respect to bulk solvent are shown.
molecules may provide a plausible explanation for its slow dissociation. Therefore, compound 28, or derivatives with similar characteristics, may be a useful tool to test whether prolonged blockade of the (peripheral) hCB۱ receptor has a beneficial effect on CB۱ receptor related disorders such as obesity.

**EXPERIMENTAL SECTION**

**Chemistry.** All solvents and reagents were purchased from commercial sources and were of analytical grade. Demineralized water is simply referred to as water or H۲O, as was used in all cases unless stated otherwise (i.e., brine). Thin-layer chromatography (TLC) was routinely consulted to monitor the progress of reactions, using aluminum-coated Merck silica gel F۲۵۴ plates. Purification was performed on a semipreparative high performance liquid chromatography (HPLC) with a mass target fraction collector, a Shimadzu QP 8000 single quadrupole mass spectrometer equipped with a 19 mm × 100 mm C۸ column. The mobile phase used was, if nothing else is stated, acetonitrile and water (50 mL) was added potassium carbonate (7.7 g, 98%) on CB۱ receptor related disorders such as obesity.

Water (50 mL) was carefully added. Extraction with EtOAc (2 x 200 mL) was added. The reaction mixture was stirred for 20 h at rt. The solution was filtered and evaporated to dryness. The residue was dissolved in AcOH and refluxed for 1 h. The mixture was cooled to rt, water (100 mL) added, and the product extracted with EtOAc (2 x 200 mL). The combined organic phases were washed with saturated aqueous sodium hydrogen carbonate, dried (Na۲SO۴), filtered, and concentrated in vacuo. Flash chromatography (sila, 30–40% EtOAc in hexane) afforded the title compound (5.75 g, 65%) as a pale-yellow solid.

**1H NMR (CDCl۳): δ 7.50–7.20 (m, 8H), 7.10–6.90 (m, 4H), 5.10 (s, 2H), 4.50 (q, 2H), 2.5 (s, 3H), 1.5 (t, 3H).

**N-(4-(Benzyloxy)phenyl)-2-(4,4-dichlorophenyl)-5-methyl-1H-imidazole-4-carboxylic Acid (4).** To a suspension of compound 3 (3.62 g, 7.5 mmol) in MeOH (60 mL) was added potassium hydroxide (4.05 g, 64.7 mmol) and MeOH was added. The reaction mixture was heated under reflux for 2.5 h. The reaction mixture was cooled to rt, filtered, and concentrated in vacuo. The residue was dissolved in EtOAc and washed with water basified with triethylamine and then brine. The organic layer was dried over Na۲SO۴ and concentrated in vacuo to give the crude title compound (3.38 g, 99%).

**Ethyl 2-(2,4-Dichlorophenyl)-1-(4-hydroxyphenyl)-5-methyl-1H-imidazole-4-carboxylate (5).** Compound 3 (4.82 g, 10 mmol) was dissolved in HBr (5% in AcOH, 80 mL) and stirred overnight at rt without exclusion of light. The solvents were evaporated and the residue coevaporated with EtOH. The residue was dissolved in EtOAc, HCl (4 M in dioxane, 5 mL), and MgSO۴ were added, and the resulting mixture heated under reflux for 2.5 h. The reaction mixture was cooled to rt, filtered, and concentrated in vacuo. The residue was dissolved in EtOAc and washed with water basified with triethylamine and then brine. The organic layer was dried over Na۲SO۴ and concentrated in vacuo to give the crude title compound (4.74 g) as a brown, viscous oil of sufficient purity for the next step.

**Ethyl 2-(2,4-Dichlorophenyl)-5-methyl-1-(4-(3,3,3-trifluoropropoxy)phenyl)-1H-imidazole-4-carboxylate (6a).** A solution of compound 5 (9.78 g, 25 mmol), 3,3,3-trifluoro-1-propanol (428 mg, 3.75 mmol) and triphenylphosphine (984 mg, 3.75 mmol) in anhydrous THF (12 mL) were treated with DEAD (40% in toluene, 1.72 mL, 3.75 mmol). The resulting mixture was stirred at rt for 30 h then heated to 50 °C overnight. After cooling to rt, additional 3,3,3-trifluoro-1-propanol (428 mg, 3.75 mmol) and triphenylphosphine (984 mg, 3.75 mmol) were added, followed by di-tert-butylazodicarboxylate (863 mg, 3.75 mmol) and the resulting mixture stirred at rt overnight. Again, additional 3,3,3-trifluoro-1-propanol (428 mg, 3.75 mmol) and triphenylphosphine (984 mg, 3.75 mmol) were added, followed by di-tert-butyl azodicarboxylate (863 mg, 3.75 mmol), and the resulting mixture was stirred at rt overnight. The mixture was concentrated in vacuo and the residue purified by column chromatography (silica gel, 10–50% EtOAc in hexanes) to yield the title compound (880 mg, 68%) as a yellowish foam of sufficient purity for the next transformation.

**1H NMR (500 MHz, CDCl۳): δ 7.22–7.16 (m, 3H), 7.01 (d, J = 8.7 Hz, 2H), 6.83 (d, J = 8.7 Hz, 2H), 4.40 (q, J = 7.1 Hz, 2H), 4.22–4.10 (m, 2H), 2.66–2.54 (m, 2H), 2.40 (s, 3H), 1.40 (t, J = 7.1 Hz, 3H).**

**Ethyl 2-(2,4-Dichlorophenyl)-1-(4-(3-fluoropropoxy)phenyl)-5-methyl-1H-imidazole-4-carboxylate (6b).** A solution of compound 5 (978 mg, 25 mmol), 3-fluoropropan-1-ol (293 mg, 3.75 mmol) and triphenylphosphine (984 mg, 3.75 mmol) in anhydrous THF (9 mL) were treated with DEAD (40% in toluene, 1.72 mL, 3.75 mmol). The resulting mixture was stirred at rt overnight. The residue was purified by column chromatography (silica gel, 20–40% EtOAc in hexanes). The product containing fractions were combined and concentrated in vacuo. The residue was dissolved in CH۲Cl۲, then an equal amount of hexane was added. The resulting solid was filtered off, and the filtrate concentrated in vacuo to yield the title compound (1.07 g, 85%) as a colorless foam of ca. 90% purity, which was used in the next transformation without further purification.

**A solution of compound 5 (3.75 g, 10 mmol), 3-fluoropropan-1-ol (293 mg, 3.75 mmol) and triphenylphosphine (984 mg, 3.75 mmol) in anhydrous THF (9 mL) were treated with DEAD (40% in toluene, 1.72 mL, 3.75 mmol).** The resulting mixture was stirred at rt overnight. The residue was purified by column chromatography (silica gel, 20–40% EtOAc in hexanes). The product containing fractions were combined and concentrated in vacuo. The residue was dissolved in CH۲Cl۲, then an equal amount of hexane was added. The resulting solid was filtered off, and the filtrate concentrated in vacuo to yield the title compound (1.07 g, 85%) as a colorless foam of ca. 90% purity, which was used in the next transformation without further purification.

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compound 6a (880 mg, 1.72 mmol), in a mixture of THF (15 mL) and EtOH (15 mL), was treated with KOH (1.07 g, 19 mmol), dissolved in water (10 mL), and the resulting mixture stirred at 50 °C. After 3 h 30 min, the reaction mixture was cooled to rt then concentrated in vacuo. The residue was partitioned between CH2Cl2 and HCl (1 M) and, after phase separation, the aqueous layer was extracted twice more with CH2Cl2. The combined organic extracts were dried over MgSO4 and concentrated in vacuo. The residue was treated with oxalyl chloride (200 μL, 2.36 mmol), followed by DMF (1 mL) and, after phase separation, the aqueous layer extracted with CH2Cl2 and twice with EtOAc. The combined organic extracts were dried over MgSO4 and concentrated in vacuo. Flash chromatography (silica, 75% EtOAc in hexane) afforded the title compound (714 mg, 90%) as a yellowish foam. 1H NMR (500 MHz, CDCl3) δ 7.32–7.18 (m, 3H), 7.00 (d, J = 8.7 Hz, 2H), 6.85 (d, J = 8.7 Hz, 2H), 4.18–4.14 (m, 2H), 2.66–2.55 (m, 2H), 2.42 (s, 3H).

2-(2,4-Dichlorophenyl)-1-(4-(3-fluoroproxy)phenyl)-5-methyl-N-(piperidin-1-yl)-1H-imidazole-4-carboxylic Acid (7b). A solution of compound 6b (1.07 g, 2.13 mmol, ca. 90% pure), in a mixture of THF (20 mL) and EtOH (20 mL), was treated with KOH (1.40 g, 25 mmol) dissolved in water (10 mL) and the resulting mixture stirred at 50 °C. After 3 h 30 min, the reaction mixture was cooled to rt then concentrated in vacuo. The residue was partitioned between CH2Cl2 and HCl (1 M) and, after phase separation, the aqueous layer extracted with CH2Cl2 and twice with EtOAc. The combined organic extracts were dried over MgSO4 and concentrated in vacuo. 1H NMR (500 MHz, CDCl3) δ 7.35–7.22 (m, 3H), 7.04 (d, J = 8.7 Hz, 2H), 6.88 (d, J = 8.7 Hz, 2H), 4.72–4.60 (m, 2H), 4.12–4.09 (m, 2H), 2.46 (s, 3H), 2.25–2.14 (m, 2H).

2-(2,4-Dichlorophenyl)-5-methyl-N-(piperidin-1-yl)-1-(4-(3,3,3-trifluoropropoxy)phenyl)-1H-imidazole-4-carboxamide (8a). A solution of compound 7a (643 mg, 1.4 mmol) in CH2Cl2 (10 mL) was treated with oxalyl chloride (200 μL, 2.36 mmol), followed by 10 μL of DMP. The resulting mixture was stirred for 90 min at rt, then concentrated in vacuo. The residue was dried under vacuum as a yellowish foam which was used without further purification. Subsequently, to a mixture of piperidin-1-amino hydrochloride (0.3 mmol) and pyridine (100 μL) in CH2Cl2 (1 mL) was added a portion of crude intermediate (2-(2,4-dichlorophenyl)-5-methyl-N-(1-(3,3,3-trifluorophenyl)phenyl)-1H-imidazole-4-carbonyl chloride (96 mg, 0.2 mmol)) in CH2Cl2 (1 mL), and the resulting mixture stirred at rt for 2 h 30 min. The reaction mixture was washed with saturated aqueous NaHCO3 (2 mL) and, after phase separation, filtered through a phase separator. The solvents were evaporated and the residue purified by preparative HPLC eluting on a reverse-phase column (50% 100% acetonitrile in aqueous NH4OAc (0.1 M)) to give the title compound (45 mg, 41%) as a colorless solid. 1H NMR (500 MHz, CDCl3) δ 7.90 (s, 1H), 7.35 (d, J = 1.9 Hz, 3H), 7.29 (d, J = 8.3 Hz, 1H), 7.23 (d, J = 1.9, 8.3 Hz, 1H), 7.03 (d, J = 8.9 Hz, 2H), 6.87 (d, J = 8.9 Hz, 2H), 4.19 (t, J = 6.6 Hz, 2H), 2.94–2.81 (m, 4H), 2.69–2.60 (m, 2H), 2.47 (s, 3H), 1.82–1.73 (m, 4H), 1.49–1.41 (m, 2H). HRMS Calcd for [C29H27Cl2N4O2 + H]+: 541.1385. Found: 541.1366. HPLC: 100%.

2-(2,4-Dichlorophenyl)-1-(4-(3,3,3-trifluoropropoxy)phenyl)-5-methyl-N-(piperidin-1-yl)-1H-imidazole-4-carboxamide (8b). A solution of compound 7b (732 mg, 1.55 mmol) in CH2Cl2 (20 mL) was treated with oxalyl chloride (200 μL, 2.36 mmol), followed by DMF (1 mL) and, after phase separation, filtered through a phase separator. The solvents were evaporated and the residue purified by preparative HPLC eluting on a reverse-phase column (5%–100% CH3CN in aqueous NH4OAc (0.1 M)) to give the title compound (74 mg, 56%) as a colorless solid. 1H NMR (500 MHz, CDCl3) δ 7.90 (s, 1H), 7.35 (d, J = 2.0 Hz, 1H), 7.28 (d, J = 8.2 Hz, 1H), 7.23 (d, J = 2.0, 8.2 Hz, 1H), 7.01 (d, J = 8.9 Hz, 2H), 6.86 (d, J = 8.9 Hz, 2H), 4.66 (dt, J = 5.7, 4.70 Hz, 2H), 4.09 (t, J = 6.1 Hz, 2H), 2.95–2.82 (m, 4H), 2.47 (s, 3H), 2.25–2.13 (m, 2H), 1.81–1.73 (m, 4H), 1.49–1.40 (m, 2H). HRMS Calcd for [C39H29Cl2F3N4O4 + H]+: 555.1541. Found: 555.1504. HPLC: 100%.
(55 mg, 0.54 mmol). The reaction was allowed to reach rt overnight. It was then cooled to 0 °C and Et,N (55 mg, 0.54 mmol) was added, followed by 3-fluoropropane-1-sulfonyl chloride (72 mg, 0.45 mmol) after a total of 19 h. After 1 h, the reaction mixture was washed with water and concentrated in vacuo. The product was purified by HPLC (30−100% CH2CN in aqueous NH4OAc (0.1 M) over 40 min) to yield the title compound as a white solid (66% yield).1H NMR (400 MHz, CDCl3) δ 7.88 (br s, 1H), 7.39−7.17 (m, 5H), 7.11 (d, J = 8.8 Hz, 2H), 4.58 (dt, J = 5.5, 46.8 Hz, 2H), 3.53−3.33 (m, 2H), 2.92−2.71 (m, 4H), 2.45 (s, 3H), 2.40−2.23 (m, 2H), 1.83−1.62 (m, 4H), 1.46−1.33 (m, 2H). HRMS Calcd for [C26H29Cl2F3N4O7S2 + H]: 605.1004. Found: 605.1006. HPLC: 100%.

4-[[2-(4-Dichlorophenyl)-5-methyl-4-(piperidin-1-yl-carbamoyl)-1H-imidazol-1-yl]phenyl]3,3-dimethylbutane-1-sulfonate (11h). A solution of compound 10 (0.89 g, 0.11 mmol) in CH2Cl2 (3 mL) was cooled to 0 °C and treated with Et,N (20 μL, 0.13 mmol). The resulting mixture was cooled to −78 °C, and 3,3-dimethylbutane-1-sulfonil chloride (25 mg, 0.13 mmol) was carefully added. The reaction was stirred at −78 °C for 1.5 h. Water was added, then the mixture was extracted with CH2Cl2. The organic extracts were dried, filtered, and concentrated in vacuo to give a residue which was purified by HPLC to deliver the title compound (46 mg, 71%) as a solid. 1H NMR (400 MHz, CDCl3) δ 7.86 (s, 1H), 7.31−7.20 (m, 5H), 7.14−7.08 (m, 2H), 3.27−3.20 (m, 2H), 2.89−2.76 (m, 4H), 2.46 (s, 3H), 1.87−1.79 (m, 2H), 1.78−1.68 (m, 5H), 1.44−1.36 (m, 2H), 0.93 (d, J = 6.5 Hz, 2H). HRMS Calcd for [C34H28ClF4N4O7S2 + H]: 759.1600. Found: 759.1588. HPLC: 100%.

4-(2-(4-Dichlorophenyl)-5-methyl-4-(piperidin-1-yl-carbamoyl)-1H-imidazol-1-yl)phenyl 3,3-dimethylbutane-1-sulfonate (11h). A solution of compound 10 (50 mg, 0.11 mmol) in CH2Cl2 (3 mL) was cooled to 0 °C and treated with Et,N (20 μL, 0.13 mmol). The resulting mixture was cooled to −78 °C, and 3,3-dimethylbutane-1-sulfonil chloride (25 mg, 0.13 mmol) was carefully added. The reaction was stirred at −78 °C for 2 h. Water was added, then the mixture was extracted with CH2Cl2. The organic extracts were dried, filtered, and concentrated in vacuo to give a residue which was purified by preparative HPLC to deliver the title compound (46 mg, 69%) as a solid. 1H NMR (400 MHz, CDCl3) δ 7.81−7.73 (m, 5H), 7.11−7.09 (m, δ, J = 8.7 Hz, 2H), 3.26−3.15 (m, 2H), 2.92−2.74 (m, 4H), 2.46 (s, 3H), 1.87−1.78 (m, 2H), 1.77−1.68 (m, 5H), 1.46−1.34 (m, 2H), 0.92 (s, 9H). HRMS Calcd for [C34H28Cl2F4N4O7S2 + H]: 793.1756. Found: 793.1755. HPLC: 100%.

Racemic 1-(4-(Benzylxoy)phenyl)-2-(4-dichlorophenyl)-N-(3-hydroxy-piperidin-1-yl)-5-methyl-1H-imidazole-4-carboxamide (12a). Compound 4 (752 mg, 1.66 mmol) and SOCl2 (33.2 mmol) were mixed, and the resulting mixture was refluxed for 1.5 h. Excess SOCl2 was removed under reduced pressure and the residue was azeotroped with toluene. 3-Hydroxy-1-aminopiperidine (6.64 mmol) was mixed with CH2Cl2 (15 mL) and THF (2 mL) and Et,N (13.28 mmol). The mixture was cooled to −20 °C under a nitrogen atmosphere. A THF (5 mL) mixture of the acid chloride from above was added dropwise during 20 min. The resulting mixture was allowed to slowly warm to rt and stirred overnight. Aqueous NaOH (1 M, 5 mL) and EtOH (15 mL) were added, and the mixture was heated to 40 °C for 15 min. The reaction mixture was then diluted to 50 mL with CH2Cl2 and washed with water (2 × 20 mL) and brine (20 mL). The organic layer was dried (MgSO4), filtered, and concentrated in vacuo. The residue was purified by flash chromatography (8% EtOAc in CH2Cl2) and then by reverse phase HPLC (Kromasil C8, 60% CH2CN in aqueous NH4OAc (0.1 M)). The product fraction was concentrated in vacuo and then dissolved in CH2Cl2 and washed with water several times and then brine. The organic layer was dried (MgSO4), filtered, and concentrated in vacuo to give the title compound (160 mg, 17% yield). 1H NMR (400 MHz, CDCl3) δ 7.05−7.10 (m, 4H), 6.90 (d, J = 8.8 Hz, 2H), 6.81 (d, J = 8.8 Hz, 2H), 5.18 (s, 1H), 4.92 (s, 2H), 3.94−3.85 (m, 1H), 3.06−2.97 (m, 1H), 2.85−2.66 (m, 3H), 2.34 (s, 3H), 1.87−1.77 (m, 1H), 1.63−1.50 (m, 2H), 1.46−1.34 (m, 1H); MS m/z 551 [M + H].

Racemic 1-(4-(Benzylxoy)phenyl)-2-(4-dichlorophenyl)-N-(3-hydroxy-cyclohexyl)-5-methyl-1H-imidazole-4-carboxamide (12b). A suspension of compound 4 (2.00 g, 4.41 mmol) in CH2Cl2 (50 mL) was treated with oxalyl chloride (2.80 g, 22.1 mmol) at rt, followed by one drop of DMF. The mixture was stirred at rt for 15 min, after which the solvent was removed in vacuo. The acid chloride was suspended in CH2Cl2 (10 mL) and added dropwise to a mixture of 3-amino-cyclohexanol (610 mg, 5.29 mmol), aqueous NaOH (1 M, 30 mL), and CH2Cl2 (30 mL). After stirring at rt for 2 h, after adding more 3-amino-cyclohexanol after 1 h 25 min (67 mg, 0.58 mmol) and 1 h 45 min (58 mg, 0.60 mmol) water, and CH2Cl2 were added and the phases separated. The organic phase was washed with aqueous HCl (10%) and brine, then dried (MgSO4), filtered, and concentrated in vacuo to yield the crude title compound (2.79 g). 1H NMR (400 MHz, CDCl3) δ 7.40−7.16 (m, 8H), 7.03−6.88 (m, 4H), 5.01 (s, 2H), 4.44−4.32 (m, 0.5H), 4.18−4.11 (m, 0.5H), 4.06−3.94 (m, 0.5H), 3.76−3.66 (m, 0.5H), 2.46 (s, 3H), 2.03−1.10 (m, 8H); MS m/z 550 (M+H).
35 min, after which the mixture was concentrated in vacuo. The acid chloride was suspended in CH₂Cl₂ (10 mL) and added dropwise to a mixture of trans-2-aminocyclohexanol hydrochloride (802 mg, 5.29 mmol), aqueous NaOH (1 M, 30 mL), and CH₂Cl₂ (30 mL). After stirring at rt for 2 h, water/CH₂Cl₂ were added, and the phases were separated. The organic phase was washed with aqueous HCl (10%) and brine, dried (MgSO₄), filtered, and concentrated in vacuo to yield the crude title compound (2.69 g).

1H NMR (400 MHz, CDCl₃) δ 7.94 (s, 1H), 7.37–7.25 (m, 6H), 7.23–7.17 (m, 2H), 6.97 (d, J = 8.6 Hz, 2H), 6.89 (d, J = 8.6 Hz, 2H), 5.23 (s, 1H), 4.98 (s, 2H), 3.80–3.62 (m, 1H), 3.59–3.42 (m, 1H), 2.42 (s, 3H), 2.14–1.93 (m, 2H), 1.75–1.59 (m, 2H), 1.39–1.14 (m, 4H). MS m/z 550 (M + H).

Racemic 1-([4-(Benzyloxy)phenyl])-2-(2,4-dichlorophenyl)-N-(cis-2 hydroxycyclohexyl)-5-methyl-1H-imidazole-4-carboxamide (12d).

A suspension of compound 4 (2.00 g, 4.41 mmol) in CH₂Cl₂ (100 mL) was treated with oxalyl chloride (2.85 g, 22.5 mmol) and Et₃N (313 mg, 3.09 mmol), and CH₂Cl₂ (7 mL). The reaction mixture was stirred at rt for 1 h. The solvent was evaporated under reduced pressure. A mixture of compound 4 (1.00 g, 2.21 mmol) in CH₂Cl₂ (15 mL) was added dropwise to acetic anhydride (3.62 g, 36.5 mmol) in CH₂Cl₂ (30 mL). The reaction mixture was stirred at rt for 2 h, and 10 min. CH₂Cl₂ was added, and the resulting mixture was filtered, and concentrated in vacuo to yield the crude title product (1.32 g).

1H NMR (400 MHz, CDCl₃) δ 7.90 (s, 1H), 7.36–7.25 (m, 6H), 7.23–7.17 (m, 2H), 6.97 (d, J = 8.6 Hz, 2H), 5.03 (s, 2H), 2.50 (s, 3H). MS m/z 597 (M + H).

1-([4-(Benzyloxy)phenyl])-2-(2,4-dichlorophenyl)-5-methyl-N-(5 methylpyridin-2-yl)-1H-imidazole-4-carboxamide (12h).

A suspension of compound 4 (3.00 g, 6.62 mmol) in CH₂Cl₂ (70 mL) was treated with oxalyl chloride (4.20 g, 33.1 mmol) at rt, followed by 5 min of DMAP. The mixture was stirred at rt for 5 min, after which the solvents were evaporated under reduced pressure. A mixture of 5-methyl-pyridin-2-ylamino (816 mg, 7.54 mmol), Et₃N (890 mg, 8.80 mmol), and CH₂Cl₂ (20 mL) was added dropwise to the acid chloride suspended in CH₂Cl₂ (20 mL). The reaction mixture was stirred at rt for 30 min. CH₂Cl₂ was added, and the resulting mixture was washed with aqueous HCl (10%) and brine, dried (MgSO₄), filtered, and concentrated by flash chromatography (20–30% EtOAc in heptane) to yield the title compound as a white solid (980 mg, 27%).

1H NMR (400 MHz, pyridine-d₅) δ 10.11 (s, 1H), 8.52 (s, 1H), 8.04 (s, 1H), 7.40–6.88 (m, 3H), 4.80 (s, 2H), 2.39 (s, 3H), 1.88 (s, 3H). MS m/z 543 (M + H).

Racemic 2-(2,4-Dichlorophenyl)-1-(4-hydroxyphenyl)-N-(3-hydroxyprop-1-yl)-5-methyl-1H-imidazole-4-carboxamide (13a).

A mixture of racemic 1-([4-(benzyloxy)phenyl])-2-(2,4-dichlorophenyl)-N-(3-hydroxyprop-1-yl)-5-methyl-1H-imidazole-4-carboxamide (160 mg, 0.29 mmol) and dimethyl sulfoxide (1.45 mmol) in CH₂Cl₂ under nitrogen atmosphere were treated dropwise with BF₃·OEt₂ (1.45 mmol). The resulting mixture was stirred for 4 days at ambient temperature while continuously adding small volumes of CH₂Cl₂ and 1,4-dioxane. EtOH was added, and the mixture was filtered, and then concentrated in vacuo. The residue was dissolved in EtOAc (50 mL) and washed with water (2 × 20 mL) and brine (20 mL). Th e organic layer was dried (Na₂SO₄), filtered, and concentrated in vacuo to give the title compound (127 mg, 95%) as a white solid. MS m/z 461 (M + H).

Racemic 2-(2,4-Dichlorophenyl)-N-(3-hydroxychoclohexyloxy)-1-(4 hydroxyphn-[1-yl]-5-methyl-1H-imidazole-4-carboxamide (13b). A suspension of crude 1-([4-(benzyloxy)phenyl])-2-(2,4-dichlorophenyl)-N-(3-hydroxyprop-1-yl)-5-methyl-1H-imidazole-4-carboxamide (2.79 g, 5.07 mmol) in CH₂Cl₂ (50 mL) and dimethyl sulfoxide (3.15 g, 50.7 mmol) was treated with boron trifluoride diethyl etherate (5.77 g, 50.7 mmol). The reaction mixture was stirred at rt for 36 h (dark), adding more dimethyl sulfoxide (3.15 g, 50.7 mmol) and boron trifluoride (5.77 g, 50.7 mmol) after 16 h. Th e solvent was evaporated and the residue dissolved in EtOAc/water. Th e phases were separated and the organic phase dried (Na₂SO₄), filtered, and concentrated in vacuo to yield the crude title compound (2.54 g). MS m/z 460 (M + H).

Racemic 2-(2,4-Dichlorophenyl)-N-(trans)-2-hydroxycyclohexyloxy)-1 (4 hydroxyphn-[1-yl]-5-methyl-1H-imidazole-4-carboxamide (13c). Crude racemic 1-([4-(benzyloxy)phenyl])-2-(2,4-dichlorophenyl)-N-(trans)-2-hydroxycyclohexyl-5-methyl-1H-imidazole-4-carboxamide (2.68 g, 4.87 mmol) was suspended in HBr (33% in AcOH, 60 mL). The mixture was stirred at rt, in the dark, for 1 h 20 min. EtOH was added and the mixture concentrated in vacuo. Th e residue was dissolved in MeOH and neutralized with NaHCO₃ (1 M,aq). One spoon of K₂CO₃ was added, and the mixture was stirred at rt for 1 h. Th e solvent was evaporated, and the resulting mixture extracted with toluene followed by THF. Th e combined organic phases were washed with aqueous HCl (10%) and brine, dried (MgSO₄), filtered, and evaporated. Th e product was purified by HPLC (30–100% CH₃CN in aqueous NH₄OAc (0.1 M) over 40 min) to yield the title compound as a white solid (829 mg, yield over 2 steps 41%).

1H NMR (400 MHz, CDCl₃) δ 7.36–7.18 (m, 4H), 6.86–6.66 (m, 4H), 5.28 (s, 1H), 4.60 (br s, 1H), 3.85–3.74 (m, 1H), 3.52–3.41 (m, 1H), 2.37 (s, 3H), 2.13–1.97 (m, 2H), 1.78–1.67 (m, 2H), 1.44–1.15 (m, 4H). MS m/z 460 (M + H).

Racemic 2-(2,4-Dichlorophenyl)-N-(cis)-2-hydroxycyclohexyloxy)-1 (4 hydroxyphn]-5-methyl-1H-imidazole-4-carboxamide (13d).

A suspension of racemic 1-([4-(benzyloxy)phenyl])-2-(2,4-dichlorophenyl)-N-(cis)-2-hydroxycyclohexyloxy)-5-methyl-1H-imidazole-4-carboxamide (2.38 g, 4.33 mmol) in HBr (33% in AcOH, 50 mL). Th e reaction mixture was stirred at rt, in the dark, for 1 h. EtOH was added and the solvents were evaporated under reduced pressure. Th e residue was dissolved in MeOH and neutralized with aqueous NaHCO₃ (1 M).
The solvent was evaporated and the mixture dissolved in water/CHCl3. The phases were separated, and the organic phase was washed with brine, dried (MgSO4), filtered, and evaporated. The residue was dissolved in MeOH and one spoon of K2CO3 was added, and the resulting mixture was stirred at rt for 1 h before the solvent was evaporated. The residue was suspended in CH2Cl2 and washed with aqueous HCl (10%), and the solvents were evaporated. The residue was dissolved in THF, dried (MgSO4), filtered, and evaporated to yield the crude title compound (210 mg). 1H NMR (400 MHz, CDCl3) δ 7.73–7.71 (m, 2H), 3.93–3.91 (m, 2H), 3.82–3.80 (m, 2H), 3.72–3.69 (m, 2H), 3.62–3.60 (m, 2H), 3.50–3.48 (m, 2H), 3.29–3.27 (m, 2H), 3.13–3.11 (m, 2H), 2.98–2.96 (m, 2H), 2.87–2.85 (m, 2H), 2.74–2.72 (m, 2H), 2.68–2.66 (m, 2H), 2.63–2.61 (m, 2H), 2.49–2.47 (m, 2H), 2.17–2.15 (m, 2H), 1.95–1.93 (m, 2H), 1.86–1.84 (m, 2H), 1.38–1.36 (m, 2H). MS m/z 497 (M + H).

2-(2,4-Dichlorophenyl)-1-(4-hydroxyphenyl)-5-methyl-[14C]-imidazole-4-carboxamide (13e). Crude 12e (1.15 g, 2.10 mmol) was suspended in HBr (33% in AcOH, 25 mL). The reaction mixture was stirred at rt, in the dark, for 2 h. EtOH was added, and the solvents were evaporated at reduced pressure. The residue was dissolved in MeOH and neutralized with aqueous NaHCO3 (1 M). The solvent was evaporated and the mixture dissolved in water/CHCl3. The phases were separated, and the organic phase was washed with brine, dried (MgSO4), filtered, and concentrated in vacuo to yield the crude title compound (772 mg, 97%). 1H NMR (400 MHz, CDCl3) δ 7.72–7.70 (m, 2H), 7.39–7.37 (m, 2H), 6.94–6.92 (m, 2H), 4.25 (s, 3H). MS m/z 497 (M + H).

2-(2,4-Dichlorophenyl)-1-(4-hydroxyphenyl)-5-methyl-[15N4]-imidazole-4-carboxamide (13f). A suspension of crude racemic 2-(2,4-dichlorophenyl)-1-(4-hydroxyphenyl)-5-methyl-[15N4]-imidazole-4-carboxamide (118 mg, 0.22 mmol) in CH2Cl2 (1 mL) and THF (1 mL) was treated with Et3N (0.25 mmol) under a nitrogen atmosphere. The solution was cooled to −78 °C, and a solution of 3,3,3-trifluoropropan-1-sulfonfonyl chloride in CH2Cl2 (1 mL) was added slowly while monitoring the progress with LC-MS. The reaction mixture was quenched by addition of EtOH. The reaction mixture was concentrated in vacuo, and the residue was purified by reverse phase HPLC (Kromasil C8, 5–100% CH3CN in aqueous NH4OAc (0.1 M)) and by flash chromatography (8% EtOH in CH2Cl2). The product was freeze-dried to give the title compound (40 mg, 25%) as a white powder. 1H NMR (CD3OD) δ 7.52–7.44 (m, 2H), 7.43–7.34 (m, 2H), 3.91–3.82 (m, 2H), 3.77–3.69 (m, 2H), 3.11 (d, J = 3.0, 10.1 Hz, 1H), 2.95–2.80 (m, 3H), 2.74–2.58 (m, 2H), 2.46 (s, 3H), 1.95–1.75 (m, 2H), 1.73–1.62 (m, 1H), 1.44–1.31 (m, 1H). MS m/z 621 (M + H). HMR Calc'd for [C32H32Cl2F8N4O10S + H2]: 1617.0590. Found: 1617.0591. HMR Calc'd for [C32H32Cl2F8N4O10S-H]: 1615.0705. Found: 1615.0728. HMR Calc'd for [C32H32Cl2F8N4O10S-H2]: 1613.0584. Found: 1613.0599.

2-(2,4-Dichlorophenyl)-1-(4-hydroxyphenyl)-5-methyl-[13C]-imidazole-4-carboxamide (13g). A suspension of crude racemic 2-(2,4-dichlorophenyl)-1-(4-hydroxyphenyl)-5-methyl-[13C]-imidazole-4-carboxamide (1.17 g, 1.96 mmol) in CH2Cl2 (6 mL) and H2O (6 mL) was treated with Et3N (667 mg, 6.59 mmol) at rt. The resulting mixture was cooled to −78 °C, and 3,3,3-trifluoropropan-1-sulfonyl chloride (1.35 g, 2.20 mmol) was added dropwise. After stirring at −78 °C for 2 h 45 min, the reaction mixture was allowed to reach rt, upon which it was washed with water and evaporated. The stereoisomers were separated by HPLC (30–100% CH3CN in aqueous NH4OAc (0.1 M)) and by chiral chromatography (Chiralpak AD, heptane:iPrOH 85:15) to afford the title compound (30 mL) treated with Et3N (440 mg, 4.34 mmol) at rt. The resulting mixture was cooled to −78 °C, and 3,3,3-trifluoropropan-1-sulfonyl chloride (1.30 mg, 6.59 mmol) was added dropwise. After stirring at −78 °C for 2 h 30 min, the reaction mixture was allowed to reach rt, upon which it was washed with water and evaporated. The stereoisomers were separated by HPLC (30–100% CH3CN in aqueous NH4OAc (0.1 M)) and by chiral chromatography (Chiralpak AD, heptane:iPrOH 85:15) to afford the title compound (710 mg, 64%). 1H NMR (400 MHz, CDCl3) δ 7.06 (m, 8H), 3.82–3.70 (m, 2H), 2.49–2.47 (m, 2H), 2.17–2.15 (m, 2H), 1.75–1.61 (m, 2H), 1.34–1.12 (m, 4H). HMR Calc'd for [C31H31ClF3N4O5S+H]: 882.1826. Found: 882.1831. HMR Calc'd for [C31H31ClF3N4O5S-H]: 880.1650. Found: 880.1655. HMR Calc'd for [C31H31ClF3N4O5S-H2]: 878.1478. Found: 878.1483. Vibrational circular dichroism experiments were unable to unambiguously assign the absolute stereochemistry of the (−) enantiomer.


3.3), 29.3 (q, 1H) 3.89 (m, 1H), 3.49 (m, 2H), 2.86 (m, 2H), 2.39 (m, 2H). The resulting mixture was diluted with water and concentrated in vacuo to give a residue which was purified by HPLC (30–100% CH3CN in aqueous NH4OAc (0.1 M) over 35 min) to yield the title compound as a white solid (150 mg, 0.30 mmol) in dry CH2Cl2 (2 mL) was treated with Et3N (43 mg, 0.43 mmol) at rt. The resulting mixture was cooled to −78 °C, and 3,3,3-trifluoropropyl-1-sulfonol chloride (94 mg, 0.48 mmol) in dry CH2Cl2 (0.5 mL) was added dropwise. After stirring at −78 °C for 80 min, the reaction mixture was washed with water and evaporated. The product was purified by HPLC (30–100% CH3CN in aqueous NH4OAc (0.1 M) over 40 min) to yield the title compound as a white solid (131 mg, yield over 3 steps 52%). 1H NMR (400 MHz, CDCl3) δ 7.28 (d, J = 7.9 Hz, 1H), 7.15 (m, 4H), 3.54 (m, 2H), 3.45 (m, 2H), 3.38 (m, 2H), 2.80 (m, 2H), 2.72 (m, 2H), 2.53 (s, 3H). MS m/z 583 (M + H). HRMS Calcd for [C26H21Cl2F3N4O4S + H]: 617.0491. Found: 617.0473. HPLC: 100%.

2.2,2-Trichloroethyl-1-(4-(benzyloxyl)phenyl)-2-(4-dichlorophenyl)-5-methyl-1H-imidazole-4-carboxylate (15). A solution of compound 4 (10.0 g, 22.1 mmol) in CH2Cl2 (210 mL) was treated with oxalyl chloride (18.5 g, 145 mmol), followed by a few drops of DMF. The mixture was stirred at rt for 2 h, after which the solvents were evaporated. The residue was dissolved in CH2Cl2 (80 mL) and the mixture was cooled to 0 °C, upon which 2,2-trichloroethanol (3.63 g, 24.3 mmol) was added followed by DIPEA (3.42 g, 26.5 mmol). The ice bath was then removed, and the reaction mixture was stirred at rt for 3 h, adding DMAP (279 mg, 2.28 mmol) after 1 h 40 min. The reaction mixture was diluted with CH2Cl2, washed with water, dried (MgSO4), filtered, and concentrated in vacuo to yield the crude title compound (14.9 g). 1H NMR (400 MHz, CDCl3) δ 7.40–7.14 (m, 16, 7H), 6.85–6.56 (m, 2H), 6.42–6.08 (m, 7H), 6.16–5.82 (m, 4H), 5.09 (2H, s), 4.33 (s, 3H). MS m/z 583 (M + H).

2.2-Trichloroethyl-2-(4-dichlorophenyl)-1-(4-hydroxyphenyl)-5-methyl-1H-imidazole-4-carboxylate (16). Crude 15 (14.7 g) was dissolved in HBr (33% in AcOH, 200 mL) After having stirred at rt for an additional hour, the reaction mixture was cooled to 0 °C and EtOH was added. The mixture was stirred for 10 min before the solvents were evaporated. The residue was dissolved in MeOH and neutralized with aqueous NaHCO3 (1 M). The solvent was evaporated and the mixture dissolved in CH2Cl2. The organic phase was washed with water and dried (MgSO4), filtered, and concentrated in vacuo to yield the title compound (14.9 g). 1H NMR (400 MHz, CDCl3) δ 7.40–7.14 (m, 16, 7H), 6.85–6.56 (m, 2H), 6.42–6.08 (m, 7H), 6.16–5.82 (m, 4H), 5.09 (2H, s), 4.33 (s, 3H). MS m/z 583 (M + H).

2.2-Trichloroethyl-2-(4-dichlorophenyl)-5-methyl-1H-imidazole-4-carboxylate (17). A suspension of 16 (5.0 g, 10.13 mmol) in dry CH2Cl2 (100 mL) under nitrogen was treated with Et3N (1.23 g, 12.2 mmol) at rt. The resulting mixture was cooled to −78 °C, and 3,3,3-trifluoropropyl-1-sulfonol chloride (2.19 g, 11.1 mmol) was added dropwise. The reaction mixture was stirred at −78 °C for 3 h, adding more 3,3,3-trifluoropropyl-1-sulfonol chloride (0.28 g 1.43 mmol) after 2 h. Water was added and the phases were separated on a phase separator. The organic phase was concentrated in vacuo to yield the title compound (6.43 g, 97%). 1H NMR (400 MHz, CDCl3) δ 7.37–7.15 (m, 7H), 6.98–6.61 (m, 8H), 5.01 (2H, s), 4.23 (s, 3H). MS m/z 493 (M + H).

2-(4-Dichlorophenyl)-5-methyl-1-(4-(3,3,3-trifluoropropylsulfonyl)phenyl)-1H-imidazole-4-carboxylic acid (18). A solution of 17 (6.43 g, 9.82 mmol) in AcOH (100 mL) was treated with zinc dust (9.74 g, 148.91 mmol). The mixture was stirred at rt for 3 h, after which it was filtered through Celite and evaporated. The residue was dissolved in CH2Cl2 and washed with aqueous HCl (0.1 M), dried, filtered, and concentrated in vacuo to yield the crude title compound (5.28 g). MS m/z 523 (M + H).

2-(4-Dichlorophenyl)-4-(4-hydroxychlorocarbamoyl)-5-(methyl-1H-imidazol-1-yl)phenyl-3,3,3-trifluoropropane-1-sulfonate (19). A solution of 18 (crude 528 mg) in CH2Cl2 (25 mL) was treated with oxalyl chloride (641 mg, 5.00 mmol). A precipitate formed immediately after the addition so more CH2Cl2 (15 mL) was added, followed by a few drops of DMF. The reaction mixture was stirred at rt for 2 h, further extracted with water and evaporated. The product was dissolved by HPLC (30–100% CH3CN in aqueous NH4OAc (0.1 M) over 40 min) to yield the title compound as a white solid (131 mg, yield over 3 steps 52%). 1H NMR (400 MHz, CDCl3) δ 7.28 (d, J = 7.9 Hz, 1H), 7.15 (m, 4H), 3.54 (m, 2H), 3.45 (m, 2H), 3.38 (m, 2H), 2.80 (m, 2H), 2.72 (m, 2H), 2.53 (s, 3H).
(3 mL) was treated with TBAF (1.0 M THF, 237 mg, 0.91 mmol). The residue of the fully protected intermediate (610 mg, 0.91 mmol) in dry THF was evaporated to yield the crude desilylated intermediate (529 mg).

Racemic N-((cis)-2-Aminocyclohexyl)-2-(2,4-dichlorophenyl)-1-(4-(benzyloxy)phenyl)-3,3,3-trifluoropropane-1-sulfonate (24). A suspension of racemic 24 (104 mg, 0.21 mmol) in dry CH2Cl2 (1.5 mL) was treated with Et3N (23 mg, 0.22 mmol) and AcOH (0.2 mL) at rt. The resulting mixture was cooled to −78 °C and 3,3,3-trifluoropropane-1-sulfonil chloride (50 mg, 0.26 mmol) was added dropwise. After stirring at −78 °C for 3 h (including extra additions of 3,3,3-trifluoropropane-1-sulfonil chloride (2 × 43 mg, 0.22 mmol) after 1.5 and 2.5 h), the reaction mixture was washed with water and evaporated to yield the crude intermediate (655 mg). MS m/z 719 (M + H). To the solution of the Boc-protected intermediate (655 mg, 0.91 mmol) in MeOH (10 mL) at 0 °C was added dropwise, a solution of thionyl chloride in MeOH (prepared by dropwise addition of thionyl chloride (5.41 g, 45.5 mmol) to MeOH (10 mL) at −40 °C). After the addition, the ice bath was removed. The reaction mixture was stirred at rt for 1 h, after which the solvents were evaporated. The product was purified by HPLC (30–100% CH2CN (with 0.1% formic acid) in 0.1% formic acid (aq) during 40 min). The CH2CN was evaporated and the resulting mixture extracted with CH2Cl2. The organic phase was washed with aqueous NaHCO3 (1 M), dried (MgSO4), filtered, and concentrated in vacuo to yield the crude title compound (104 mg). MS m/z 487 (M + H).
(MgSO4), filtered, and concentrated in vacuo to yield the title compound as a slightly yellow oil (37 mg yield over 2 steps 20%). 1H NMR (400 MHz, CDCl3) δ 2.13 (d, J = 2.0 Hz, 1H), 7.29 (d, J = 8.9 Hz, 1H), 7.26 (d, J = 2.0 Hz, 1H), 1.71 (d, J = 8.9 Hz, 1H), 4.59—4.51 (m, 1H), 3.56—3.48 (m, 2H), 2.86—2.76 (m, 2H), 2.51 (s, 3H), 2.31 (s, 3H), 2.26—2.19 (m, 1H), 2.07—2.01 (s, 3H), 1.96 (d, J = 2.1 Hz, 2H), 1.85—1.77 (m, 1H), 1.54—1.25 (m, 5H). HRMS Calcld for [C26H18Cl2F6N4O4S + H]+: 667.0408. Found: 667.0540. HPLC: 100%.

Equilibrium Radioligand Displacement Assays. [3H]CP55940 displacement assays on 96-well plate were used for the determination of affinity (IC50 and Kd) values of antagonists for the cannabinoid CB1 receptors. The displacement experiments were performed using six concentrations of competing antagonists in 25 μL of assay buffer (50 mM Tris-HCl, 5 mM MgCl2, 0.1% BSA, pH 7.4) in the presence of another 25 μL of assay buffer with a final concentration of 3.5 nM [3H]CP55940. At this concentration, total radioligand binding did not exceed 10% of that added to prevent ligand depletion. Membrane aliquots containing 5 μg of CHOK1hCB1_bgal membrane in 100 μL of assay buffer were incubated at 30 °C for 60 min. Non-specific binding (NSB) was determined in the presence of 10 μM rimonabant. Incubation was terminated by rapid filtration performed on 96-well GF/C filter plates (PerkinElmer, Groningen, The Netherlands), presoaked for 30 min with 0.25% PEI (Polyethyleneimine), using a PerkinElmer Filtermate harvester (PerkinElmer, Groningen, The Netherlands). After 30 min of dehydration of the filter plate at 50 °C, the filter-bound radioactivity was determined by scintillation spectrometry using a 2450 MicroBeta 2 plate counter. The binding values were recorded in both counts per minute (CPM) and disintegrations per minute (DPM). Each antagonist was measured in duplicate, and at least three individual experiments were performed.

Classic Radioligand Kinetic Assays. Association experiments were performed by incubating membrane aliquots containing 5 μg of CHOK1hCB1_bgal membrane in a total volume of 100 μL of assay buffer at 30 °C with 3.5 nM [3H]CP55940. The amount of radioligand bound to the receptor was measured at different time intervals during a total incubation of 120 min. Dissociation experiments were performed by preincubating membrane aliquots containing 5 μg of protein in a total volume of 100 μL of assay buffer for 60 min. After the preincubation, radioligand dissociation was initiated by the addition of 10 μM unlabeled rimonabant. The amount of radioligand still bound to the receptor was measured at various time intervals for a total of 240 min to ensure that full dissociation from cannabinoid CB1 receptor was reached. Incubation was terminated by rapid filtration performed on GF/C filters (Whatman International, Maidstone, UK), presoaked for 30 min with 0.25% PEI, using a Brandel harvester (Brandel, Gaithersburg, MD). Filter-bound radioactivity was determined by scintillation spectrometry using a Tri-Carb 2000 TR liquid scintillation counter (PerkinElmer, Boston, MA).

Competition Association Assays. Kinetic rate index (KRI) values are an average of at least two independent experiments, each consisting of two replicates. Kinetic rate constant values are an average of at least three independent experiments, each consisting of two replicates. The binding kinetics of unlabeled ligands was quantified using the competition association assay based on the theoretical framework by Motulsky and Mahan. A concentration of 1—3-fold of the IC50 value was used to determine the binding kinetics of unlabeled cannabinoid CB1 receptor antagonists. The competition association assay was initiated by adding membrane aliquots (5 μg/well) at different time points for a total of 240 min to a total volume of 100 μL of assay buffer at 30 °C with 3.5 nM [3H]CP55940 in the absence or presence of competing CB1 receptor antagonists (1 to 3-fold IC50). Incubations were terminated, and samples obtained as described under Equilibrium Radioligand Displacement Assay. The “dual-point” competition association assays were run similarly, with only two time points, at 30 and 240 min, respectively. [35S]GTPγS Binding Assays. Antagonism assay: The antagonism of all tested compounds was evaluated at 30 °C in a [35S]GTPγS binding assay as reported earlier. 1 Insurmountability assay: Membrane homogenates containing the CB1 receptor (5 μg) were equilibrated in the assay buffer (50 mM Tris-HCl, 5 mM MgCl2, 1 mM EDTA, 100 mM NaCl, 0.05% BSA, pH 7.4) supplemented with 1 μM GDP, 1 mM DTT, and 2 μg of saponin. Membrane preparations were preincubated with or without antagonists (10-fold K values on the CB1 receptor) for 1 h prior to the
challenge of a CB1 receptor agonist, CP55940 at 25 °C with concentrations ranging from 1 μM to 0.1 nM. Subsequently, [35S]GTPγS (final concentration 0.3 nM) was added and incubation continued for another 30 min at 25 °C. Incubations were terminated, and samples were obtained as described under Equilibrium Radioligand Displacement Assays.

Data Analysis. All experimental data were analyzed using the nonlinear regression curve fitting program GraphPad Prism 6.0 (GraphPad Software, Inc., San Diego, CA). From displacement assays, IC50 values were obtained by nonlinear regression analysis of the displacement curves. The obtained IC50 values were converted into Kd values using the Cheng–Prusoff equation to determine the affinity of the ligands.32 The koff and kcat values for radiolabeled and unlabeled ligands were fitted and calculated, and the koff and kcat values were used to calculate residence times (in min) and kinetic dissociation binding constants (kinetic Kd). Association and dissociation rates for unlabeled compounds were calculated by fitting the data into the competition association model using “kinetics of competitive binding”:

\[
K_a = k_l [L] \cdot 10^{-9} + k_2 \\
K_b = k_l [I] \cdot 10^{-9} + k_3 \\
S = \sqrt{(K_a - K_b)^2 + 4 k_l k_3} \cdot L \cdot 10^{-18} \\
K_F = 0.5(K_a + K_b + S) \\
K_S = 0.5(K_a + K_b - S) \\
Q = \frac{B_{max} k_l L \cdot 10^{-9}}{K_F - K_S} \\
Y = Q \left( \frac{k_1 (K_F - K_S)}{K_F - K_S} + \frac{k_F - K_S}{K_F} e^{(-k_F x)} - \frac{k_F - K_S}{K_S} e^{(-k_F x)} \right)
\]

where \(k_l\) is the koff of the radioligand (M⁻¹ s⁻¹), \(k_2\) is the kcat of the radioligand (s⁻¹), \(L\) is the radioligand concentration (nM), \(I\) is the concentration of the unlabelled competitor (nM), and \(X\) is the time (min) and \(Y\) is the specific binding of the radioligand (DPM). During a competition association, these parameters are set, obtaining \(k_l\) from the control curve without competitor and \(k_j\) from previously performed dissociation assays described under Traditional Radioligand Kinetic Assays. With that, the \(k_j\), \(k_b\), and \(B_{max}\) can be calculated, where \(k_j\) represents the koff (M⁻¹ s⁻¹) of the unlabeled ligand, \(k_b\) stands for the kcat (s⁻¹) of the unlabeled ligand, and \(B_{max}\) equals the total binding (DPM). All competition association data were globally fitted. Residence times (RT, expressed in min) were calculated as RT = 1/(60 x koff).

Computational Studies. All computational studies were performed in the Schrödinger suite38 and based on the crystal structure of the CB1 receptor co-crystallized with 29 (PDB 5TGZ).39 The crystal structure was prepared with the Protein Preparation Wizard.53 Ligands were docked using induced fit docking,54 with core constraints on the 2,4-dichlorophenyl ring of 29 (all ligands share this moiety). To study whether the difference in RTs among 11d, 14f, and 28 could be explained by unfavorable hydration, we generated a WaterMap around 14f.55,66 Figures were rendered using PyMol.55

### ASSOCIATED CONTENT

#### Supporting Information

These materials are available free of charge via the Internet at The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.jmedchem.7b00861.

Target selectivity data for representative human CB1 receptor antagonists at human CB2 receptor physico-chemical properties of all antagonists, including their correlations with corresponding KRI values; proton NMR spectra for all final products and carbon NMRs spectra of 11b, 14f, and 28 (PDF)

Molecular formula strings (CSV)

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Author Contributions

Lizi Xia and Adriaan P. IJzerman conceived the study. Adriaan P. IJzerman, Robert J. Sheppard, Michael J. Waring, and Laura H. Heitman supervised the project. The chemical synthesis was designed and supervised by Leifeng Cheng and performed by Sara Pahlén, Maria J. Petersson, Peter Schell, and Roine I. Olsson. The bioassays were supervised by Adriaan P. IJzerman and Laura H. Heitman and performed by Lizi Xia and Henk de Vries. The computational work was performed by Eelke B. Lenselink. The manuscript was written by Lizi Xia, Julien Louvel, Robert J. Sheppard, and Adriaan P. IJzerman.

Notes

The authors declare no competing financial interest.

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### DEDICATION

We dedicate this study to the memory of Dr. Julien Louvel who passed away on November 5, 2017.

### ABBREVIATIONS USED

AEA, anandamide; 2-AG, 2-arachidonoylglycerol; CB, cannabinoid; CNS, central nervous system; ECS, endocannabinoid system; GPCRs, G-protein-coupled receptors; KRI, kinetic rate index; PNS, peripheral nervous system; PSA, Polar Surface Area; RT, residence time; SAR, structure–affinity relationship; SKR, structure-kinetic relationship; TMS, tetramethylsilane


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