

Cover Page



Universiteit Leiden



The handle <http://hdl.handle.net/1887/32582> holds various files of this Leiden University dissertation

Author: Wijngaarden, Marjolein A.

Title: Metabolic and endocrine adaptations to fasting in lean and obese individuals

Issue Date: 2015-03-26

Chapter 1

1

Introduction

1

Fasting and energy-sensing

A prolonged fast has tremendous effects on metabolism. Mainly insulin, but also glucose levels quickly decline over the course of a couple of days. During the fed state energy is mainly derived from carbohydrate oxidation. This switches during fasting towards lipid oxidation. The amount of carbohydrate and lipid oxidation can be estimated by indirect calorimetry. This method uses the CO₂ and O₂ in the exhaled air to calculate substrate oxidation ¹. During a fast, the increase in lipid oxidation leads to the production of ketone bodies (beta-hydroxybutyrate and acetoacetate) in the liver, which can be used as fuel by the brain when glucose levels drop ². After 2-3 days of fasting, liver glucose stores (glycogen) are depleted. At this point, glucose is synthesized in the kidney and also in the liver from lactate, pyruvate, glycerol and amino acids such as alanine (gluconeogenesis) ³.

While fasting, the activity of hormonal axes such as the thyroid-axis – important for metabolism – and the reproductive axis are downregulated. With respect to the thyroid-hormone axis this is characterized by decreased thyroid-stimulating hormone (TSH) and triiodothyronine (T₃) levels whereas T₄ levels remain stable which is due to an altered deiodinase activity during fasting and leptin might be involved as well ^{4,5}. Likely, the reduction in T₃ levels saves energy during fasting. The pituitary-gonadal axis is downregulated during fasting as well; plasma levels of follicle-stimulating hormone (FSH), luteinizing hormone (LH) and testosterone decrease during fasting whereas sex-hormone binding globulin (SHBG) levels increase ⁶. Most likely, this is an evolutionary mechanism to prevent reproduction when food is sparse.

On a molecular level, so-called energy-sensing pathways are affected during fasting. The adenosine monophosphate-activated kinase (AMP-activated kinase, AMPK) is of special interest. This enzyme is phosphorylated by liver kinase B1 (LKB1), in response to energy depletion and the coinciding increase in AMP/ATP ratio. Generally speaking, the actions of AMPK aim to restore energy balance; AMPK inhibits anabolic processes and stimulates catabolic processes. AMPK is also involved in the metabolic shift towards lipid oxidation during fasting; AMPK phosphorylates and thereby deactivates acetyl-CoA carboxylase 1 (ACC). This results in an increased fatty acid oxidation and reduced lipid storage ⁷. The most important effects of AMPK on glucose metabolism are that it increases glucose uptake ^{8,9} and inhibits the hepatic glucose production ¹⁰⁻¹². Besides AMPK, there are many upstream and downstream (in)direct targets that play a role during fasting with respect to either energy-sensing or the

metabolic shift such as the sirtuins (SIRT, from silent information regulators), histone deacetylase 4 (HDAC4), mammalian target of rapamycin (mTOR), Forkhead box O (FOXO), protein kinase B (PKB/Akt), pyruvate dehydrogenase kinase isozyme 4 (PDK4), glucose transporter type 4 (GLUT4), cluster of differentiation 36 (CD36) and ACC.

Obesity

The body mass index (BMI) is used to define obesity; the BMI is calculated by dividing body weight (in kilograms) by the square of height (in meters). The World Health Organization defines obesity as a BMI ≥ 30 kg/m² and overweight as a BMI between 25 and 30 kg/m². The prevalence of obesity has doubled since 1980. In 2008 there were 1.5 billion overweight and 500 million obese adults (>20 years) ¹³. Obesity is a risk factor for cardiovascular diseases, diabetes, musculoskeletal disorders and many forms of cancer (e.g. endometrial, breast and colon cancer). Since it became clear that AMPK is important for sensing and repairing energy balance disturbances, and obesity is clearly a result from disrupted energy balance, the role of AMPK in type 2 diabetes mellitus and obesity has been investigated. Skeletal muscle is very flexible with respect to the metabolic shift from glucose towards fatty acid oxidation. Since the *musculus vastus lateralis* is easily accessible, energy-sensing pathways are often studied in human biopsy materials of this muscle. With respect to AMPK, thus far only few alterations in obesity and type 2 diabetes mellitus (T2DM) have been found. One study has shown that AMPK activity is reduced in muscle from obese subjects ¹⁴. In contrast, other studies showed that skeletal muscle AMPK activity is similar between lean and obese individuals or obese subjects and T2DM patients ^{15;16}.

Brain

The hypothalamus is probably the most important area of the brain involved in the homeostasis of metabolism, food intake and energy expenditure ¹⁷. The hypothalamus is located in the ventral part of the diencephalon, below the thalamus. The hypothalamus has important hormonal effects, since it secretes factors and hormones that stimulate the pituitary to secrete hormones (with neuro-endocrine feedback). The hypothalamic arcuate nucleus receives information about the peripheral blood through the median eminence, where the blood-brain barrier is absent ¹⁸. Claude

Bernard was the first to suggest a role for the hypothalamus in glucose homeostasis in 1855¹⁹. It is now known that different hypothalamic areas have different roles in weight maintenance. Animal studies show that damaging the lateral hypothalamus induces weight loss whereas damaging the ventromedial part (VMH) of the hypothalamus induces obesity (3-6). On a neuronal level, food intake is regulated by hypothalamic neurons that either stimulate (orexigenic) or inhibit (anorexigenic) food intake. The orexigenic neurons contain NPY (neuropeptide Y) and AGRP (agouti-related protein), neurotransmitters that stimulate food intake²³⁻²⁵. Anorexigenic neurons contain CART (cocaine- and amphetamine-regulated transcript) and POMC (proopiomelanocortin), neurotransmitters that inhibit food intake²⁶⁻²⁸.

Functional Magnetic Resonance Imaging

When a human body is placed in a magnetic resonance imaging (MRI) scanner its hydrogen nuclei get excited by radiofrequency pulses in two directions; longitudinal and transversal. Contrasts on MR images are based on the relaxation of these excited nuclei; longitudinal relaxation (T1) and transverse relaxation (T2). Functional MRI (fMRI) uses blood-oxygen-level dependent (BOLD) signals as a measure of brain activity. The basics behind this mechanism were described at first by Linus Pauling in 1936 which was later implemented by Seiji Ogawa^{29;30}. Oxygenated hemoglobin (Hb) is diamagnetic, whereas deoxygenated Hb is paramagnetic. Brain activity in a certain brain region increases the blood flow to this region. This alters the ratio of oxygenated Hb versus deoxygenated Hb in favor of the oxygenated hemoglobin in the blood stream itself and in the surrounding tissues. Paramagnetic molecules reduce signal intensity (T2 value) and result in a dark image. When the amount of paramagnetic molecules in the blood decreases (during increased brain activity) the opposite occurs; the signal intensity increases. Since this was discovered, a lot of research has been performed in the neuropsychological field to correlate brain function with specific brain regions.

Within the brain, the hypothalamus is involved in the regulation of food intake and energy expenditure¹⁷ whereas the rewarding effects of food are mainly regulated in the amygdala³¹. As reviewed, the brain – mainly the hypothalamus again - also plays a role in glucose homeostasis³². To date, however, there are not many functional MRI studies that looked at the effect of fasting on neuronal activity. A positron emission tomography (PET) study showed that a 36 hour fast (“hunger”), compared to the

satiated state, increased regional cerebral bloodflow (rCBF) in the hypothalamus, insula, (para)limbic areas (such as the ACC), thalamus, caudate, precuneus, putamen and cerebellum³³. Two other PET studies looked at the response to respectively the taste or ingestion of a meal after a 36 hour fast in lean and obese individuals. rCBF increased in the midbrain and insula and decreased in the PCC, temporal cortex and OFC in obese compared to lean participants in response to tasting a meal after the fast³⁴. Meal ingestion, after a 36 hour fast, led to a higher increase in rCBF the prefrontal cortex in obese compared to lean individuals³⁵. Besides, a larger rCBF decrease was seen in the (para)limbic areas in obese compared to lean individuals. Decreases in the hypothalamus, ACC and thalamus upon satiation were smaller in obese than lean participants³⁵.

Heart Rate Variability

During a short-term fast in humans, the activity of the sympathetic nervous system (SNS) increases^{36,37}. A derivative of SNS activity, heart rate variability (HRV), can easily be measured in humans by electrocardiography. Indeed, heart rate is under control of the autonomic nervous system (as reviewed³⁸). The autonomic nervous system consists of the sympathetic nervous system that increases heart rate and the parasympathetic nervous system that decreases heart rate. The balance between these autonomic branches is extremely dynamic and quickly adapts to environmental cues, such as fasting. The sympathetic nervous system mobilizes energy whereas the parasympathetic nervous system is important for the digestion of food and energy storage. Disbalances of the autonomic nervous system mostly result in hyperactivity of the sympathetic nervous system and a decrease in parasympathetic nervous system activity. In obesity, an increased SNS activity has been previously described³⁹.

Outline Thesis

The general aim of this thesis was to study the physiological adaptations to a prolonged-fast in different populations on several parameters. In these studies, we were mainly interested in the response of energy-sensing pathways in skeletal muscle. First, we hoped to contribute to the understanding of normal fasting physiology. Besides, a better understanding of the response of these energy-sensing pathways to a prolonged-fast - and the possible differences in this response between lean and

obese subjects - may lead to the identification of factors that are involved in the pathophysiology of obesity. A bit far stretched, baseline differences between lean and obese subjects in the functioning of their energy-sensing systems might give cues for therapeutic targets.

In the current thesis, we hypothesized that the response to fasting would be different between lean and obese subjects. Next to the energy-sensing pathways, we also studied the neuronal response with functional MRI scanning and we assessed the response of the autonomic nervous system by heart rate variability measurements. These studies are described in the “middle” chapters of this thesis. The thesis starts with a chapter that is dedicated to the time-course of energy-sensing adaptations during a 24 hour fast in healthy young men. At the very end of this thesis, we describe the result of a study in which we hypothesized that the adaptations to a 60 hour fast would be altered by the presence of food related odours during fasting.

In **chapter two**, we hypothesized that the response to a prolonged fast of 48 hours would be different between lean and obese individuals. We compared this response on several levels in 14 obese and 12 lean individuals: we made fMRI scans, took blood samples, performed an indirect calorimetry, an electrocardiogram to measure HRV and took muscle biopsies. All measurements were performed both before and after the 48 hour fast (the indirect calorimetry and blood sampling were also performed after 24 hours of fasting). The results of the HRV measurements, used to study the effect of fasting on SNS activity in lean and obese individuals, are described in **chapter three**. In **chapter four** we describe the differential effect of fasting on the hypothalamus, amygdala and posterior cingulate cortex functional connectivity networks (fMRI) in lean and obese individuals. In **chapter five**, we investigated the time course of metabolic adaptations in response to a 24 hour fast. We investigated this time course by taking blood samples, by performing an indirect calorimetry (to measure lipid and carbohydrate oxidation) and by taking muscle biopsies in a group of 12 healthy young male volunteers at 3 time points during a 24 hour fast (different in that respect compared to chapter two). In the muscle biopsies we were mainly interested in the so-called “energy-sensing” pathways. In **chapter six** we evaluated the hypothesis that the response to fasting would be influenced by the presence or absence of visual and odorous food cues, based on a study in *Drosophila Melanogaster*⁴⁰. Twelve lean men fasted twice during 60 hours; once in the presence and once in the absence of food cues. We studied the effects of the presence and absence of the

1

food cues on blood parameters, lipid and glucose oxidation (measured by indirect calorimetry) and the hypothalamic BOLD signal (measured by fMRI). In **chapter seven** we discuss all chapters described above.

References

1. Simonson DC, DeFronzo RA. Indirect calorimetry: methodological and interpretative problems. *Am J Physiol* 1990; 258(3 Pt 1): E399-E412.
2. Owen OE, Morgan AP, Kemp HG, Sullivan JM, Herrera MG, Cahill GF, Jr. Brain metabolism during fasting. *J Clin Invest* 1967; 46(10): 1589-1595.
3. Cahill GF, Jr. Fuel metabolism in starvation. *Annu Rev Nutr* 2006; 26:1-22.: 1-22.
4. Heemstra KA, Soeters MR, Fliers E, Serlie MJ, Burggraaf J, van Doorn MB, van der Klaauw AA, Romijn JA, Smit JW, Corssmit EP, Visser TJ. Type 2 iodothyronine deiodinase in skeletal muscle: effects of hypothyroidism and fasting. *J Clin Endocrinol Metab* 2009; 94(6): 2144-2150.
5. Boelen A, Wiersinga WM, Fliers E. Fasting-induced changes in the hypothalamus-pituitary-thyroid axis. *Thyroid* 2008; 18(2): 123-129.
6. Bergendahl M, Aloï JA, Iranmanesh A, Mulligan TM, Veldhuis JD. Fasting suppresses pulsatile luteinizing hormone (LH) secretion and enhances orderliness of LH release in young but not older men. *J Clin Endocrinol Metab* 1998; 83(6): 1967-1975.
7. Davies SP, Sim AT, Hardie DG. Location and function of three sites phosphorylated on rat acetyl-CoA carboxylase by the AMP-activated protein kinase. *Eur J Biochem* 1990; 187(1): 183-190.
8. Nakano M, Hamada T, Hayashi T, Yonemitsu S, Miyamoto L, Toyoda T, Tanaka S, Masuzaki H, Ebihara K, Ogawa Y, Hosoda K, Inoue G, Yoshimasa Y, Otaka A, Fushiki T et al. alpha2 Isoform-specific activation of 5'adenosine monophosphate-activated protein kinase by 5-aminoimidazole-4-carboxamide-1-beta-d-ribo nucleoside at a physiological level activates glucose transport and increases glucose transporter 4 in mouse skeletal muscle. *Metabolism* 2006; 55(3): 300-308.
9. Kurth-Kraczek EJ, Hirshman MF, Goodyear LJ, Winder WW. 5' AMP-activated protein kinase activation causes GLUT4 translocation in skeletal muscle. *Diabetes* 1999; 48(8): 1667-1671.
10. Andreelli F, Foretz M, Knauf C, Cani PD, Perrin C, Iglesias MA, Pillot B, Bado A, Tronche F, Mithieux G, Vaulont S, Burcelin R, Viollet B. Liver adenosine monophosphate-activated kinase-alpha2 catalytic subunit is a key target for the control of hepatic glucose production by adiponectin and leptin but not insulin. *Endocrinology* 2006; 147(5): 2432-2441.

-
- 1
11. Shaw RJ, Lamia KA, Vasquez D, Koo SH, Bardeesy N, Depinho RA, Montminy M, Cantley LC. The kinase LKB1 mediates glucose homeostasis in liver and therapeutic effects of metformin. *Science* 2005; 310(5754): 1642-1646.
 12. Koo SH, Flechner L, Qi L, Zhang X, Sreaton RA, Jeffries S, Hedrick S, Xu W, Boussouar F, Brindle P, Takemori H, Montminy M. The CREB coactivator TORC2 is a key regulator of fasting glucose metabolism. *Nature* 2005; %20;437(7062): 1109-1111.
 13. World Health Organization. Obesity and Overweight, Fact Sheet No 311. 2011.
 14. Bandyopadhyay GK, Yu JG, Ofrecio J, Olefsky JM. Increased malonyl-CoA levels in muscle from obese and type 2 diabetic subjects lead to decreased fatty acid oxidation and increased lipogenesis; thiazolidinedione treatment reverses these defects. *Diabetes* 2006; 55(8): 2277-2285.
 15. Steinberg GR, Smith AC, van Denderen BJ, Chen Z, Murthy S, Campbell DJ, Heigenhauser GJ, Dyck DJ, Kemp BE. AMP-activated protein kinase is not down-regulated in human skeletal muscle of obese females. *J Clin Endocrinol Metab* 2004; 89(9): 4575-4580.
 16. Hojlund K, Mustard KJ, Staehr P, Hardie DG, Beck-Nielsen H, Richter EA, Wojtaszewski JF. AMPK activity and isoform protein expression are similar in muscle of obese subjects with and without type 2 diabetes. *Am J Physiol Endocrinol Metab* 2004; 286(2): E239-E244.
 17. Schwartz MW, Woods SC, Porte D, Jr., Seeley RJ, Baskin DG. Central nervous system control of food intake. *Nature* 2000; 404(6778): 661-671.
 18. Broadwell RD, Brightman MW. Entry of peroxidase into neurons of the central and peripheral nervous systems from extracerebral and cerebral blood. *J Comp Neurol* 1976; 166(3): 257-283.
 19. Bernard C. *Lecons de Physiologie Experimentale Appliqué a la Medicine Faites au College de France*. Bailere et Fils: Paris, France, 1855.
 20. Marshall NB, Barnett RJ, Mayer J. Hypothalamic lesions in goldthioglucose injected mice. *Proc Soc Exp Biol Med* 1955; 90(1): 240-244.
 21. Brecher G, Waxler SH. Obesity in albino mice due to single injections of goldthioglucose. *Proc Soc Exp Biol Med* 1949; 70(3): 498-501.
 22. Nagamachi Y. Effect of satiety center damage on food intake, blood glucose and gastric secretion in dogs. *Am J Dig Dis* 1972; 17(2): 139-148.
 23. Kalra SP, Dube MG, Sahu A, Phelps CP, Kalra PS. Neuropeptide Y secretion increases in the paraventricular nucleus in association with increased appetite for
-

-
- food. *Proc Natl Acad Sci U S A* 1991; 88(23): 10931-10935.
24. Stanley BG, Kyrkouli SE, Lampert S, Leibowitz SF. Neuropeptide Y chronically injected into the hypothalamus: a powerful neurochemical inducer of hyperphagia and obesity. *Peptides* 1986; 7(6): 1189-1192.
25. Zarjevski N, Cusin I, Vettor R, Rohner-Jeanrenaud F, Jeanrenaud B. Chronic intracerebroventricular neuropeptide-Y administration to normal rats mimics hormonal and metabolic changes of obesity. *Endocrinology* 1993; 133(4): 1753-1758.
26. Fan W, Boston BA, Kesterson RA, Hruby VJ, Cone RD. Role of melanocortineric neurons in feeding and the agouti obesity syndrome. *Nature* 1997; 385(6612): 165-168.
27. Hagan MM, Rushing PA, Pritchard LM, Schwartz MW, Strack AM, Van Der Ploeg LH, Woods SC, Seeley RJ. Long-term orexigenic effects of AgRP-(83—132) involve mechanisms other than melanocortin receptor blockade. *Am J Physiol Regul Integr Comp Physiol* 2000; 279(1): R47-R52.
28. Rossi M, Kim MS, Morgan DG, Small CJ, Edwards CM, Sunter D, Abusnana S, Goldstone AP, Russell SH, Stanley SA, Smith DM, Yagaloff K, Ghatei MA, Bloom SR. A C-terminal fragment of Agouti-related protein increases feeding and antagonizes the effect of alpha-melanocyte stimulating hormone in vivo. *Endocrinology* 1998; 139(10): 4428-4431.
29. Pauling L, Coryell CD. The Magnetic Properties and Structure of Hemoglobin, Oxyhemoglobin and Carbonmonoxyhemoglobin. *Proc Natl Acad Sci U S A* 1936; 22(4): 210-216.
30. Ogawa S, Lee TM, Kay AR, Tank DW. Brain magnetic resonance imaging with contrast dependent on blood oxygenation. *Proc Natl Acad Sci U S A* 1990; 87(24): 9868-9872.
31. Baxter MG, Murray EA. The amygdala and reward. *Nat Rev Neurosci* 2002; 3(7): 563-573.
32. Lam CK, Chari M, Lam TK. CNS regulation of glucose homeostasis. *Physiology (Bethesda)* 2009; 24:159-70.: 159-170.
33. Tataranni PA, Gautier JF, Chen K, Uecker A, Bandy D, Salbe AD, Pratley RE, Lawson M, Reiman EM, Ravussin E. Neuroanatomical correlates of hunger and satiation in humans using positron emission tomography. *Proc Natl Acad Sci U S A* 1999; 96(8): 4569-4574.
-

- 1
34. DelParigi A, Chen K, Salbe AD, Reiman EM, Tataranni PA. Sensory experience of food and obesity: a positron emission tomography study of the brain regions affected by tasting a liquid meal after a prolonged fast. *Neuroimage* 2005; 24(2): 436-443.
 35. Gautier JF, Chen K, Salbe AD, Bandy D, Pratley RE, Heiman M, Ravussin E, Reiman EM, Tataranni PA. Differential brain responses to satiation in obese and lean men. *Diabetes* 2000; 49(5): 838-846.
 36. Chan JL, Mietus JE, Raciti PM, Goldberger AL, Mantzoros CS. Short-term fasting-induced autonomic activation and changes in catecholamine levels are not mediated by changes in leptin levels in healthy humans. *Clin Endocrinol (Oxf)* 2007; 66(1): 49-57.
 37. Webber J, Macdonald IA. The cardiovascular, metabolic and hormonal changes accompanying acute starvation in men and women. *Br J Nutr* 1994; 71(3): 437-447.
 38. Dampney RA. Functional organization of central pathways regulating the cardiovascular system. *Physiol Rev* 1994; 74(2): 323-364.
 39. Muscelli E, Emdin M, Natali A, Pratali L, Camastra S, Gastaldelli A, Baldi S, Carpeggiani C, Ferrannini E. Autonomic and hemodynamic responses to insulin in lean and obese humans. *J Clin Endocrinol Metab* 1998; 83(6): 2084-2090.
 40. Libert S, Zwiener J, Chu X, Vanvoorhies W, Roman G, Pletcher SD. Regulation of *Drosophila* life span by olfaction and food-derived odors. *Science* 2007; 315(5815): 1133-1137.