

Immunogenetic and immunological aspects of rheumatoid arthritis: DERAA and anti-citrulline reactivity can make the difference Feitsma, A.L.

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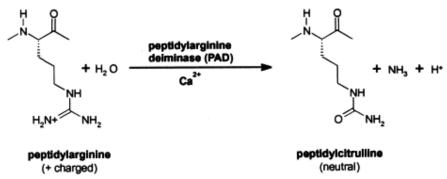
### Undifferentiated and Rheumatoid Arthritis

Arthritis is a group of conditions characterized by inflammation of the joints. This inflammation can lead to breakdown of the cartilage of the joints that can be caused by different mechanisms, e.g. autoimmunity, fractures, wearing, or infection. The different types of arthritis are diagnosed according to disease criteria, leaving cases that cannot be classified in one of the accepted categories of rheumatic diseases (usually referred to as 'undifferentiated arthritis' (UA)). The diagnosis of rheumatoid arthritis (RA), an inflammatory autoimmune disorder characterized by a chronic inflammation of the synovial tissue of several joints, is based on a list of seven criteria developed by the American College of Rheumatology (1). These criteria include clinical, radiological and laboratory findings; i.e. morning stiffness, arthritis of three or more joint areas, arthritis of hand joints, symmetric arthritis, serum rheumatoid factor, rheumatoid nodules, and radiographic changes. The RA patient population is clinically heterogeneous since only four of these seven ACR criteria have to be fulfilled for the diagnosis of RA. The occurrence of RA varies among countries and areas over the world, but has a prevalence of approximately 1% in Europe (2;3). In the Dutch population, women are affected by RA approximately two times more frequently than men (4).

In the Leiden Early Arthritis Clinic (EAC), which provides an inception cohort of patients with recent onset arthritis (5), 37% of the patients are diagnosed with UA and about 20% with RA at their first visit. After 1 year, 32% of the UA patients have qualified for the diagnosis of RA, indicating the complexity of a diagnosis at initial presentation.

RA patients can develop different kind of autoantibodies, amongst others against citrullinated proteins (anti-citrullinated protein antibodies (ACPA)). Citrullination is a post-translational conversion (deimination) of Arginine to Citrulline residues performed by the enzyme peptidylarginine deiminase (PAD) (*Figure 1*) that results in a small change in molecular mass (<1 Da) and the loss of one positive charge. Although citrullination is a common natural process, these ACPA are specific for RA, and can be measured already years before symptomatic disease (6;7). Recently, it has been shown that ACPA<sup>+</sup> and ACPA<sup>-</sup> RA patients show a different disease course, probably indicating that ACPA<sup>+</sup> and ACPA<sup>-</sup> RA reflect a totally different disease (8-10).





**Figure 1.** Citrullination. An Arginine residue is enzymatically converted by peptidylarginine deiminase (PAD) into a Citrulline residue in the presence of Ca<sup>2+</sup>.

# Progression and Severity scoring

RA is characterized by the proliferation of synovium and the destruction of cartilage and bone during the progression of the disease. Destruction of the cartilage is a consequence of pro-inflammatory cytokines and enzymes that are released during the chronic inflammation process inducing enhanced breakdown of cartilage matrix and reduced synthesis of matrix components by the articular chondrocytes (11;12), but joint erosion results more directly from osteoclasts (13). The formation of pannus, which results from the proliferating synovium (14-16), will eventually lead to joint space narrowing and joint erosions. This is, at least in part, mediated by fibroblast-like synoviocytes from the synovium (17).

Radiographic joint damage is an important outcome measure in RA, in addition to assessments of physical function and disease activity, which all associate with each other (18). It reflects cumulative disease activity and is related to overall disability (19;20). Therefore, progression rates are also influenced by the current therapy, i.e. disease modifying anti-rheumatic drugs (DMARDs) (21;22). Several scoring methods for the assessment of radiographic joint damage exist, from which the most well known are the Larsen (23) and Sharp-van der Heijde method (24;25). Both methods score the individual joints of the extremities but the Sharp-van der Heijde method scores hands and feet for the amount and severity of erosions and joint space narrowing separately (25). The Sharp-van der Heijde method is sensitive to detect changes over time and shows reliable results since it has a low measurement error (26). Analyses of radiographic progression can be analyzed on the individual patient or a patient group level. For both applies that the radiological progression is linear in the first

(approximately) five years but further in time, the curve levels off to a plateau (18;27-29).

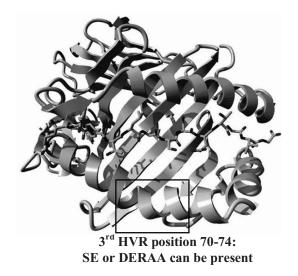
Because nowadays, RA patients are seen in an earlier phase of the disease, before the appearance of well established indicators of poor prognosis such as erosions and nodules, markers which have a good predictive value on radiographic damage in an early phase of the disease will become more important.



### Genetic Risk Factors for RA

The pathogenesis of RA is, as in many other autoimmune diseases, complex and largely unknown. It is generally accepted that both genetic and environmental factors contribute and probably also interact with each other. It has been described that genetic factors contribute for about 2/3 to the development of RA (30;31). The contributing risk factors can differ for the susceptibility to, and the progression of RA.

The strongest genetic risk factor, both for susceptibility and severity, has been mapped to the HLA-class II region, most probably DRB1. HLA-class II molecules consist of an  $\alpha$ - and  $\beta$ -chain which both have constant and variable regions. The variable regions constitute the binding groove for the peptide to be presented by HLA molecules to T cells of the immune system (*Figure 2*). A particular part of the binding groove, the third hypervariable region, is involved in the susceptibility to RA development. At position 70 to 74 in this third hypervariable region, different variants of amino acid sequences are present. Certain HLA-DRB1 alleles share common epitopes at this position. Regarding the risk for RA development, three variants can be discriminated; either amino acids of the so-called shared epitope (SE), the sequence "DERAA", or 'neutral' amino acids are present. Compared to the 'neutral' HLA-DRB1 alleles, carriership of HLA-DRB1 alleles with the SE increases the risk for RA development, and "DERAA"-containing HLA-DRB1 alleles decrease the risk. Both the SE and the "DERAA"-containing HLA-DRB1 allele effects will be discussed in more detail below.



**Figure 2**. HLA class II molecule (adapted from Boots et al. (32)). The variable regions of the  $\alpha$ - and  $\beta$ -chain build up the peptide binding groove. The rectangle indicated in the figure shows the position of the third hypervariable region (HVR) where the shared epitope (SE) or "DERAA"-sequence can be present.

#### Shared Epitope and ACPA

HLA-DRB1 molecules containing the amino acid sequence R(Q)K(R)RAA (i.e. the amino acids Arginine, (Glutamine), Lysine, (Arginine), Arginine, Alanine, Alanine) at position 70-74 in the third hypervariable region are belonging to the Shared Epitope (SE) alleles. This epitope is present in the HLA-DRB1\*0101, \*0102, \*0401, \*0404, \*0405, \*0408, \*0410, \*1001 and \*1402 alleles. The SE is associated with an increased risk (about 2.5 times (33)) to develop RA and a more severe disease course. It is postulated that this SE sequence, which is present in the binding cleft of the HLA-DR molecule, is directly related to the binding of RA inducing peptides to the SEcontaining HLA-DR molecules. These peptides are then presented to T cells thought to play an important role in the pathogenesis of RA. Since there is high linkage disequilibrium between HLA-DRB1 and HLA-DQB1 alleles, it is also hypothesized that the HLA-DQB1 alleles are associated with RA susceptibility, although these associations cannot be distinguished (34-36). RA-inducing peptides are not identified yet, but several findings indicate new directions for epitope discovery. Recently it has been shown that SE-containing HLA-DRB1 alleles do not confer risk to the development of RA itself, but predispose to the development of anti-citrullinated protein antibodies (ACPA). These antibodies are highly predictive for RA development as discussed in a previous section (6;7). ACPA+ UA Patients have approximately

fifteen times higher odds to develop RA within one year compared to ACPA UA patients. These ACPA are commonly measured in the IgG isotype, but are also present in the IgM and IgA isotype (37).

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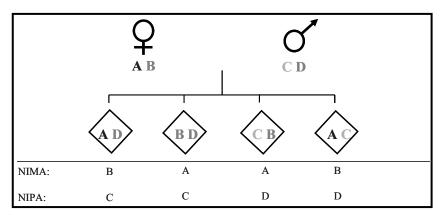
Chapter

The presence of IgG ACPA indicates the presence of T cell help. One of the proteins described to be citrullinated in vivo and present in the synovial fluid of RA patients is the cytoskeletal protein vimentin (38;39). We have recently shown that 90% of ACPA<sup>+</sup> RA patients recognizing a citrullinated peptide derived from vimentin carry one or two SE-containing HLA-DRB1 alleles (40), suggesting the involvement of helper T cells recognizing a citrullinated epitope from vimentin in the context of the SE-containing HLA-DRB1 alleles. The identification of citrullinated vimentin-derived T cell epitopes recognized by HLA-DR4 positive individuals is described in **Chapter 4** of this thesis.

#### "DERAA"

"DERAA" stands for the amino acid sequence of Aspartic acid-Glutamic acid-Arginine- Alanine-Alanine which is present in the HLA-DRB1 alleles of the subtype HLA-DRB1\*0103, \*0402, \*1102, \*1103, \*1301, \*1302 and \*1304. These alleles are present in 29% of the population (33;41-43). It has been shown by several groups that the frequency of "DERAA"-containing HLA-DRB1 alleles is reduced in RA patients compared to healthy controls and that the risk to develop RA is reduced by about 40% in DERAA positive individuals (33). Since the "DERAA" sequence is positioned at the same location as the SE in the HLA-DRB1 molecule, the effect of "DERAA" has to be evaluated after stratification for presence of the SE-containing HLA-DRB1 alleles. In this way, it has been shown that the effect of "DERAA"-containing HLA-DRB1 alleles is independent of the SE-containing HLA-DRB1 alleles (33).

"DERAA"-containing HLA-DRB1 alleles can be inherited, but can also be conferred as non-inherited maternal antigens (NIMA). NIMA can be conferred from the mother during and/or shortly after the pregnancy since the immune systems of mother and child are in close contact and cell trafficking will occur (44-47). The phenomenon of NIMA was described for the first time in 1954 for the RhD antigen (48) and is illustrated in *Figure 3*. The terminology is oriented from the point of view of the child in a family, since most studies are coming from the transplantation field. It has been described that haplo-identical NIMA-mismatched sibling transplants have a graft survival similar to that of HLA-identical siblings in contrast to NIPA-mismatched siblings, indicating tolerance for the HLA-mismatch from the mother (49-51).



**Figure 3.** Terminology of non-inherited maternal antigen (NIMA) and non-inherited paternal antigen (NIPA). The terminology is orientated from the point of view of the child.  $\diamond$  gender can be male or female.

The phenomenon of protection against RA by "DERAA"-containing HLA-DRB1 alleles as NIMA is studied in **Chapter 2** of this thesis and discussed in comparison with the inherited effect in **Chapter 3**.

Next to HLA-DRB1 there are several other genes involved in the risk for RA development. Below, only the additional risk alleles studied in the context of this thesis are discussed.

#### PTPN22

The protein Tyrosine phosphatase named Lyp is encoded by the protein Tyrosine phosphatase, non-receptor type 22 (PTPN22) gene and is expressed by many cell types present in haematopoietic tissues, like T cells, B cells, NK cells, monocytes, dendritic cells and neutrophils (52).

Genes can differ in their nucleotide sequence between different individuals in a population. When the frequency of a single nucleotide change is equal or higher than 1%, this is called a single nucleotide polymorphism (SNP). The most studied SNP of the PTPN22 gene, the C1858T missense single-nucleotide polymorphism, is associated with RA, UA (52-56) and several other autoimmune diseases (57-59). Upto now, it seems that the SNP does not have an effect on the severity of RA, only on the susceptibility (53;60).

There is a large variation in allele frequency among different ethnic populations among the world of the T-variant of the allele, with a variation only in Europe from 2-

3% in Italians to 15% in Finnish people (61). There are several articles describing the association of the PTPN22 T allele of the C1858T polymorphism with the development of ACPA<sup>+</sup> RA, therefore implicating a correlation of this SNP with the production of ACPA (62-64).

Chapter 1

We investigated in **Chapter 5** whether the C1858T polymorphism of the PTPN22 gene can give additive value to the prediction of progression from UA to RA when it is combined with presence of ACPA.

#### Genome wide association studies (GWAS)

In the past few years several genome wide association studies (GWAS) have been performed to scan the entire genome for common polymorphisms associating with different autoimmune diseases, including RA (65-68). Since thousands of patients and controls from different populations are studied in the GWAS, these studies may identify risk factors with modest effects on the risk for RA development, and also allow one to study subpopulations of patients for specific effects.

From all genes identified to be associated with RA, we studied based on a recent GWAS (67) six newly identified SNPs for their influence on the severity of RA (**Chapter 6**). These SNPs are located around genes that are either involved in activation of the immune response (CD40, TNFRSF14, CCL21 and PRKCQ) or play a role in intracellular processes involving cell cycle and homeostatis (MMEL1, KIF5A and CDK6).

## Statistical modelling

Several statistical models may be applied to the analysis of genetic associations. The kind of statistical model appropriate for the analysis is dependent on the correlation between the variables and measurement groups, the distribution of the data and the type of study performed. Below, two models are discussed that are used for the different genetic association chapters described in this thesis.

#### Theory of Bayes

To study the effect of numerous factors on the development of a disease, modelling of the risk factors studied is performed to fit the observed data with the expected frequencies. Modelling of all these risk factors preferably results in prediction of the outcome. A theory that calculates the probability that a hypothesis is true based on the available information is the theory of Bayes, which is used for the studies performed in

Chapter 2 and 5 of this thesis. With this theory, a prior probability (e.g. to develop RA), based on the risk of an individual in a certain population, is converted into a posterior probability, calculated on the basis of extra information derived from the risk factors studied. The prior probability is most often the prevalence of e.g. a certain disease (or symptom) in the population (69-71).

#### Linear Mixed Model

A mixed model is a multiple variance analysis often used to compare groups of individuals including multiple measurements from one individual e.g. in time. The data of every individual are plotted in a linear way, and combined for the whole group, thereby intrapolating missing values (72). In a mixed model both random and fixed effects are included. Fixed effects are categorical variables from which all levels are fixed and known, whereas random effects are variables where only a random sample of possible values is measured. A Gaussian distribution is assumed for the variables studied. Since all individual measurements are taken into account in one analysis, it is a powerful method with low intra-individual variability and therefore smaller group sizes are required (73). A linear mixed model was used in Chapter 6 of this thesis to study the effect of different SNPs on the severity of RA.

### Outline of the thesis

**Chapter 2** reports a family study in which we studied whether "DERAA"-containing HLA-DRB1 alleles protect against the development of rheumatoid arthritis not only when the alleles are inherited but also when they are present as a non-inherited maternal antigen (NIMA).

**Chapter 3** is a review that summarizes the epidemiological findings about "DERAA"-containing HLA-DRB1 alleles and RA and the observation we made in Chapter 2. Both associations are compared and possible explanations for the protective phenomenon are described.

On the same location in the HLA-DRB1 molecule as the "DERAA" epitope, the so-called shared epitope (SE) can be present. These SE-containing HLA-DRB1 alleles predispose to the formation of ACPA and therefore the development of RA. Since ACPA-producing B cells are helped by T cells, we studied in **Chapter 4** whether we

could identify possible T cell epitopes from an important ACPA target antigen, namely the citrullinated vimentin protein.

Both for ACPA and the C1858T polymorphism of the PTPN22 gene associations with RA development have been described. We studied the individual contribution of ACPA and a PTPN22 SNP on the prediction of RA development from UA in **Chapter** 5. We also studied the influence of the PTPN22 SNP on the level of ACPA.



Genes can be involved both in the susceptibility and severity of RA. This is the case e.g. for the SE-containing HLA-DRB1 alleles. Therefore, we studied in **Chapter 6** the contribution of several SNPs that were newly identified as risk factors for the development of RA and the severity of the disease.

The results obtained in the chapters 2-6 of this thesis are summarized and discussed in **Chapter 7**.

### References

- (1) Arnett FC, Edworthy SM, Bloch DA, McShane DJ, Fries JF, Cooper NS et al. The American Rheumatism Association 1987 revised criteria for the classification of rheumatoid arthritis. Arthritis Rheum 1988; 31(3):315-324.
- (2) Abdel-Nasser AM, Rasker JJ, Valkenburg HA. Epidemiological and clinical aspects relating to the variability of rheumatoid arthritis. Semin Arthritis Rheum 1997; 27(2):123-140.
- (3) Isenberg. Oxford Textbook of Rheumatology. 2004.
- (4) van der Linden. Nationaal Kompas Volksgezondheid. Bilthoven: RIVM; 2007.
- (5) van Aken J, van Bilsen JH, Allaart CF, Huizinga TW, Breedveld FC. The Leiden Early Arthritis Clinic. Clin Exp Rheumatol 2003; 21(5 Suppl 31):S100-S105.
- (6) van Gaalen FA, Linn-Rasker SP, van Venrooij WJ, de Jong BA, Breedveld FC, Verweij CL et al. Autoantibodies to cyclic citrullinated peptides predict progression to rheumatoid arthritis in patients with undifferentiated arthritis: a prospective cohort study. Arthritis Rheum 2004; 50(3):709-715.
- (7) van der Helm-van Mil AH, Verpoort KN, Breedveld FC, Huizinga TW, Toes RE, de Vries RR. The HLA-DRB1 shared epitope alleles are primarily a risk factor for anti-cyclic citrullinated peptide antibodies and are not an independent risk factor for development of rheumatoid arthritis. Arthritis Rheum 2006; 54(4):1117-1121.
- (8) van der Helm-van Mil AH, Huizinga TW. Advances in the genetics of rheumatoid arthritis point to subclassification into distinct disease subsets. Arthritis Res Ther 2008; 10(2):205.
- (9) Klareskog L, Ronnelid J, Lundberg K, Padyukov L, Alfredsson L. Immunity to citrullinated proteins in rheumatoid arthritis. Annu Rev Immunol 2008; 26:651-675.
- (10) Ding B, Padyukov L, Lundstrom E, Seielstad M, Plenge RM, Oksenberg JR et al. Different patterns of associations with anti-citrullinated protein antibody-positive and anti-citrullinated protein antibody-negative rheumatoid arthritis in the extended major histocompatibility complex region. Arthritis Rheum 2009; 60(1):30-38.
- (11) van den Berg WB. Joint inflammation and cartilage destruction may occur uncoupled. Springer Semin Immunopathol 1998; 20(1-2):149-164.
- (12) Tak PP, Bresnihan B. The pathogenesis and prevention of joint damage in rheumatoid arthritis: advances from synovial biopsy and tissue analysis. Arthritis Rheum 2000; 43(12):2619-2633.
- (13) Goldring SR, Gravallese EM. Pathogenesis of bone erosions in rheumatoid arthritis. Curr Opin Rheumatol 2000; 12(3):195-199.
- (14) Ainola MM, Mandelin JA, Liljestrom MP, Li TF, Hukkanen MV, Konttinen YT. Pannus invasion and cartilage degradation in rheumatoid arthritis: involvement of MMP-3 and interleukin-1beta. Clin Exp Rheumatol 2005; 23(5):644-650.
- (15) Okamoto H, Hoshi D, Kiire A, Yamanaka H, Kamatani N. Molecular targets of rheumatoid arthritis. Inflamm Allergy Drug Targets 2008; 7(1):53-66.
- (16) Fassbender HG, Gay S. Synovial processes in rheumatoid arthritis. Scand J Rheumatol Suppl 1988; 76:1-7.
- (17) Muller-Ladner U, Kriegsmann J, Franklin BN, Matsumoto S, Geiler T, Gay RE et al. Synovial fibroblasts of patients with rheumatoid arthritis attach to and invade normal human cartilage when engrafted into SCID mice. Am J Pathol 1996; 149(5):1607-1615.
- (18) Odegard S, Landewe R, van der Heijde DM, Kvien TK, Mowinckel P, Uhlig T. Association of early radiographic damage with impaired physical function in rheumatoid arthritis: a ten-year, longitudinal observational study in 238 patients. Arthritis Rheum 2006; 54(1):68-75.

- (19) Dawes PT. Radiological assessment of outcome in rheumatoid arthritis. Br J Rheumatol 1988; 27 Suppl 1:21-36.
- (20) van Leeuwen MA, van Rijswijk MH, Sluiter WJ, van Riel PL, Kuper IH, van de Putte LB et al. Individual relationship between progression of radiological damage and the acute phase response in early rheumatoid arthritis. Towards development of a decision support system. J Rheumatol 1997; 24(1):20-27.
- (21) Strand V, Cohen S, Schiff M, Weaver A, Fleischmann R, Cannon G et al. Treatment of active rheumatoid arthritis with leflunomide compared with placebo and methotrexate. Leflunomide Rheumatoid Arthritis Investigators Group. Arch Intern Med 1999; 159(21):2542-2550.
- (22) Finckh A, Liang MH, van Herckenrode CM, de PP. Long-term impact of early treatment on radiographic progression in rheumatoid arthritis: A meta-analysis. Arthritis Rheum 2006; 55(6):864-872.
- (23) Larsen A, Dale K, Eek M. Radiographic evaluation of rheumatoid arthritis and related conditions by standard reference films. Acta Radiol Diagn (Stockh) 1977; 18(4):481-491.
- (24) Sharp JT, Young DY, Bluhm GB, Brook A, Brower AC, Corbett M et al. How many joints in the hands and wrists should be included in a score of radiologic abnormalities used to assess rheumatoid arthritis? Arthritis Rheum 1985; 28(12):1326-1335.
- (25) van der Heijde DM. How to read radiographs according to the Sharp/van der Heijde method. J Rheumatol 1999; 26(3):743-745.
- (26) van der Heijde DM. Plain X-rays in rheumatoid arthritis: overview of scoring methods, their reliability and applicability. Baillieres Clin Rheumatol 1996; 10(3):435-453.
- (27) Hulsmans HM, Jacobs JW, van der Heijde DM, van Albada-Kuipers GA, Schenk Y, Bijlsma JW. The course of radiologic damage during the first six years of rheumatoid arthritis. Arthritis Rheum 2000; 43(9):1927-1940.
- (28) Plant MJ, Jones PW, Saklatvala J, Ollier WE, Dawes PT. Patterns of radiological progression in early rheumatoid arthritis: results of an 8 year prospective study. J Rheumatol 1998; 25(3):417-426.
- (29) Graudal NA, Jurik AG, de CA, Graudal HK. Radiographic progression in rheumatoid arthritis: a long-term prospective study of 109 patients. Arthritis Rheum 1998; 41(8):1470-1480.
- (30) MacGregor AJ, Snieder H, Rigby AS, Koskenvuo M, Kaprio J, Aho K et al. Characterizing the quantitative genetic contribution to rheumatoid arthritis using data from twins. Arthritis Rheum 2000; 43(1):30-37.
- (31) van der Woude D., Houwing-Duistermaat JJ, Toes RE, Huizinga TW, Thomson W, Worthington J et al. Quantitative heritability of anti-citrullinated protein antibody-positive and anti-citrullinated protein antibody-negative rheumatoid arthritis. Arthritis Rheum 2009; 60(4):916-923.
- (32) Boots AM, Hubers H, Kouwijzer M, den Hoed-van ZL, Westrek-Esselink BM, van DC et al. Identification of an altered peptide ligand based on the endogenously presented, rheumatoid arthritis-associated, human cartilage glycoprotein-39(263-275) epitope: an MHC anchor variant peptide for immune modulation. Arthritis Res Ther 2007; 9(4):R71.
- (33) van der Helm-van Mil AH, Huizinga TW, Schreuder GM, Breedveld FC, de Vries RR, Toes RE. An independent role of protective HLA class II alleles in rheumatoid arthritis severity and susceptibility. Arthritis Rheum 2005; 52(9):2637-2644.
- (34) Singal DP, D'Souza M, Reid B, Bensen WG, Kassam YB, Adachi JD. HLA-DQ beta-chain polymorphism in HLA-DR4 haplotypes associated with rheumatoid arthritis. Lancet 1987; 2(8568):1118-1120.
- (35) Zanelli E, Huizinga TW, Guerne PA, Vischer TL, Tiercy JM, Verduyn W et al. An extended HLA-DQ-DR haplotype rather than DRB1 alone contributes to RA predisposition. Immunogenetics 1998; 48(6):394-401.

- (36) Fugger L, Svejgaard A. The HLA-DQ7 and -DQ8 associations in DR4-positive rheumatoid arthritis patients. A combined analysis of data available in the literature. Tissue Antigens 1997; 50(5):494-500.
- (37) Verpoort KN, Jol-van der Zijde CM, Papendrecht-van der Voort EA, Ioan-Facsinay A, Drijfhout JW, van Tol MJ et al. Isotype distribution of anti-cyclic citrullinated peptide antibodies in undifferentiated arthritis and rheumatoid arthritis reflects an ongoing immune response. Arthritis Rheum 2006; 54(12):3799-3808.
- (38) Tabushi Y, Nakanishi T, Takeuchi T, Nakajima M, Ueda K, Kotani T et al. Detection of citrullinated proteins in synovial fluids derived from patients with rheumatoid arthritis by proteomics-based analysis. Ann Clin Biochem 2008; 45(Pt 4):413-417.
- (39) Bang H, Egerer K, Gauliard A, Luthke K, Rudolph PE, Fredenhagen G et al. Mutation and citrullination modifies vimentin to a novel autoantigen for rheumatoid arthritis. Arthritis Rheum 2007; 56(8):2503-2511.
- (40) Verpoort KN, Cheung K, Ioan-Facsinay A, van der Helm-van Mil AH, de Vries-Bouwstra JK, Allaart CF et al. Fine specificity of the anti-citrullinated protein antibody response is influenced by the shared epitope alleles. Arthritis Rheum 2007; 56(12):3949-3952.
- (41) Lundstrom E, Kallberg H, Smolnikova M, Ding B, Ronnelid J, Alfredsson L et al. Opposing effects of HLA-DRB1\*13 alleles on the risk of developing anti-citrullinated protein antibody-positive and anti-citrullinated protein antibody-negative rheumatoid arthritis. Arthritis Rheum 2009; 60(4):924-930.
- (42) Carrier N, Cossette P, Daniel C, de Brum-Fernandes A, Liang P, Menard HA et al. The DERAA HLA-DR alleles in patients with early polyarthritis: protection against severe disease and lack of association with rheumatoid arthritis autoantibodies. Arthritis Rheum 2009; 60(3):698-707.
- (43) Morgan AW, Haroon-Rashid L, Martin SG, Gooi HC, Worthington J, Thomson W et al. The shared epitope hypothesis in rheumatoid arthritis: evaluation of alternative classification criteria in a large UK Caucasian cohort. Arthritis Rheum 2008; 58(5):1275-1283.
- (44) Molitor ML, Haynes LD, Jankowska-Gan E, Mulder A, Burlingham WJ. HLA class I noninherited maternal antigens in cord blood and breast milk. Hum Immunol 2004; 65(3):231-239.
- (45) Petit T, Dommergues M, Socie G, Dumez Y, Gluckman E, Brison O. Detection of maternal cells in human fetal blood during the third trimester of pregnancy using allele-specific PCR amplification. Br J Haematol 1997; 98(3):767-771.
- (46) Lo YM, Lo ES, Watson N, Noakes L, Sargent IL, Thilaganathan B et al. Two-way cell traffic between mother and fetus: biologic and clinical implications. Blood 1996; 88(11):4390-4395.
- (47) Hall JM, Lingenfelter P, Adams SL, Lasser D, Hansen JA, Bean MA. Detection of maternal cells in human umbilical cord blood using fluorescence in situ hybridization. Blood 1995; 86(7):2829-2832.
- (48) Owen RD, Wood HR, Foord AG, Sturgeon P, Baldwin LG. EVIDENCE FOR ACTIVELY ACQUIRED TOLERANCE TO Rh ANTIGENS. Proc Natl Acad Sci U S A 1954; 40(6):420-424.
- (49) Burlingham WJ, Grailer AP, Heisey DM, Claas FH, Norman D, Mohanakumar T et al. The effect of tolerance to noninherited maternal HLA antigens on the survival of renal transplants from sibling donors. N Engl J Med 1998; 339(23):1657-1664.
- (50) Claas FH, Gijbels Y, van der Velden-de Munck, van Rood JJ. Induction of B cell unresponsiveness to noninherited maternal HLA antigens during fetal life. Science 1988; 241(4874):1815-1817.
- (51) van Rood JJ, Roelen DL, Claas FH. The effect of noninherited maternal antigens in allogeneic transplantation. Semin Hematol 2005; 42(2):104-111.

- (52) Begovich AB, Carlton VE, Honigberg LA, Schrodi SJ, Chokkalingam AP, Alexander HC et al. A missense single-nucleotide polymorphism in a gene encoding a protein tyrosine phosphatase (PTPN22) is associated with rheumatoid arthritis. Am J Hum Genet 2004; 75(2):330-337.
- (53) Wesoly J, van der Helm-van Mil AH, Toes RE, Chokkalingam AP, Carlton VE, Begovich AB et al. Association of the PTPN22 C1858T single-nucleotide polymorphism with rheumatoid arthritis phenotypes in an inception cohort. Arthritis Rheum 2005; 52(9):2948-2950.
- (54) Seldin MF, Shigeta R, Laiho K, Li H, Saila H, Savolainen A et al. Finnish case-control and family studies support PTPN22 R620W polymorphism as a risk factor in rheumatoid arthritis, but suggest only minimal or no effect in juvenile idiopathic arthritis. Genes Immun 2005; 6(8):720-722.
- (55) Hinks A, Barton A, John S, Bruce I, Hawkins C, Griffiths CE et al. Association between the PTPN22 gene and rheumatoid arthritis and juvenile idiopathic arthritis in a UK population: further support that PTPN22 is an autoimmunity gene. Arthritis Rheum 2005; 52(6):1694-1699.
- (56) Dieude P, Garnier S, Michou L, Petit-Teixeira E, Glikmans E, Pierlot C et al. Rheumatoid arthritis seropositive for the rheumatoid factor is linked to the protein tyrosine phosphatase nonreceptor 22-620W allele. Arthritis Res Ther 2005; 7(6):R1200-R1207.
- (57) Reddy MV, Johansson M, Sturfelt G, Jonsen A, Gunnarsson I, Svenungsson E et al. The R620W C/T polymorphism of the gene PTPN22 is associated with SLE independently of the association of PDCD1. Genes Immun 2005; 6(8):658-662.
- (58) Kahles H, Ramos-Lopez E, Lange B, Zwermann O, Reincke M, Badenhoop K. Sex-specific association of PTPN22 1858T with type 1 diabetes but not with Hashimoto's thyroiditis or Addison's disease in the German population. Eur J Endocrinol 2005; 153(6):895-899.
- (59) Skorka A, Bednarczuk T, Bar-Andziak E, Nauman J, Ploski R. Lymphoid tyrosine phosphatase (PTPN22/LYP) variant and Graves' disease in a Polish population: association and gene dosedependent correlation with age of onset. Clin Endocrinol (Oxf) 2005; 62(6):679-682.
- (60) Naseem H, Thomson W, Silman A, Worthington J, Symmons D, Barton A. The PTPN22\*C1858T functional polymorphism is associated with susceptibility to inflammatory polyarthritis but neither this nor other variants spanning the gene is associated with disease outcome. Ann Rheum Dis 2008; 67(2):251-255.
- (61) Gregersen PK, Lee HS, Batliwalla F, Begovich AB. PTPN22: setting thresholds for autoimmunity. Semin Immunol 2006; 18(4):214-223.
- (62) Orozco G, Pascual-Salcedo D, Lopez-Nevot MA, Cobo T, Cabezon A, Martin-Mola E et al. Autoantibodies, HLA and PTPN22: susceptibility markers for rheumatoid arthritis. Rheumatology (Oxford) 2008; 47(2):138-141.
- (63) Kokkonen H, Johansson M, Innala L, Jidell E, Rantapaa-Dahlqvist S. The PTPN22 1858C/T polymorphism is associated with anti-cyclic citrullinated peptide antibody-positive early rheumatoid arthritis in northern Sweden. Arthritis Res Ther 2007; 9(3):R56.
- (64) Kallberg H, Padyukov L, Plenge RM, Ronnelid J, Gregersen PK, van der Helm-van Mil AH et al. Gene-gene and gene-environment interactions involving HLA-DRB1, PTPN22, and smoking in two subsets of rheumatoid arthritis. Am J Hum Genet 2007; 80(5):867-875.
- (65) Wellcome Trust Case Control Consortium. Genome-wide association study of 14,000 cases of seven common diseases and 3,000 shared controls. Nature 2007; 447(7145):661-678.
- (66) Fung EY, Smyth DJ, Howson JM, Cooper JD, Walker NM, Stevens H et al. Analysis of 17 autoimmune disease-associated variants in type 1 diabetes identifies 6q23/TNFAIP3 as a susceptibility locus. Genes Immun 2009; 10(2):188-191.
- (67) Raychaudhuri S, Remmers EF, Lee AT, Hackett R, Guiducci C, Burtt NP et al. Common variants at CD40 and other loci confer risk of rheumatoid arthritis. Nat Genet 2008; 40(10):1216-1223.

- (68) Barton A, Thomson W, Ke X, Eyre S, Hinks A, Bowes J et al. Rheumatoid arthritis susceptibility loci at chromosomes 10p15, 12q13 and 22q13. Nat Genet 2008; 40(10):1156-1159.
- (69) Freedman L. Bayesian statistical methods. A natural way to assess clinical evidence. British medical journal 1996; 313:569-570.
- (70) Petrie A, Sabin C. Medical Statistics at a glance. 109-110. 2000. Ref Type: Serial (Book, Monograph)
- (71) Vandenbroucke JP, Hofman. Diagnostiek van ziekte. Grondslagen der Epidemiologie. 6th ed. 2004
- (72) van Houwelingen. Inleiding tot de medische statistiek. 1993.
- (73) Stevens. Intermediate statistics. A modern approach. 1990.