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Author: Suurmond, Jolien **Title**: Immune regulation by mast cells **Issue Date**: 2016-03-22

SUMMARY & DISCUSSION

AIMS

This thesis focuses on the understanding of mast cell (and basophil) functions, with a special emphasis on the role of mast cells in autoimmune disease. Here, I will discuss the findings of this thesis, based on the specific aims:

- 1. To characterize the interaction between innate and Fc receptor triggers on mast cell and basophil function
- 2. To analyze the interaction between mast cells and CD4 $^+$ T cells
- 3. To understand the function of mast cells in chronic inflammation

INTERACTION BETWEEN INNATE & ADAPTIVE IMMUNITY

In the immune system, mast cells are considered to be involved in both innate and adaptive immunity. Their rapid degranulation makes them one of the first cells to respond in innate immune responses, whereas their expression of Fc receptors makes them important effector cells in antibody-mediated adaptive responses. However, these responses are often considered separately, and not much is known about the interaction between innate and adaptive immune triggers, which are often present, as discussed below.

In the first part of this thesis, I aimed at understanding the interaction between triggers of the innate immune system in combination with Fc receptor triggering. In chapters 2, 4, and 5, I showed that interactions of both FcεRI and FcγRIIA with TLR ligands induced synergy in mast cell and basophil activation. This interaction was characterized by a marked increase in cytokine secretion, without affecting Fc receptor-mediated degranulation.

Although the magnitude of the response was largely increased when TLR ligands and Fc receptor triggers were present at the same time, the specificity of the response remained tightly regulated. In particular, the cytokine profile was highly dependent on the TLR ligand that was present, in both basophils and mast cells.

Although we did not investigate the mechanism of synergy, some findings suggest that the enhanced cytokine production was mediated through interaction of the TLR and Fc receptor signaling pathways. The synergy remained present when mast cells or basophils were treated with brefeldin A to block the transport of proteins from the Golgi system to the cell surface or extracellular environment (Chapter 2 and unpublished observations).

This suggests that the synergy did not depend on upregulation of surface receptors or secreted mediators.

In human monocytes and dendritic cells, combined triggering of FcγRIIA and TLR was shown to induce synergy in transcription levels of several cytokines, but the exact signaling pathways leading to the synergy are not known (*[1,](#page-20-0) [2](#page-20-1)*). A study in murine mast cells investigated the phosphorylation events downstream of TLR and Fc receptors upon combined triggering of these receptors and showed that activation of the JNK kinase pathway was enhanced upon combined triggering of these receptors (*[3](#page-20-2)*). Many of the upstream molecules of the JNK kinase pathway induced by TLR activation (JNK1, MKK4, TAK1, IRAK1, IRAK2, MyD88, TRAM, and TRIF)(*[4](#page-20-3)*) or Fc receptor triggering (BTK, Syk) are expressed by human mast cells, at least at the mRNA level (Chapter 5 and unpublished observations). Therefore, a similar synergy in JNK kinase activation upon activation through TLR and Fc receptors may be underlying the synergy we observed.

As discussed below, the synergy in mast cell activation upon combined triggering of TLR and Fc receptors has important implications for both protective immunity as well as hypersensitivity reactions, such as during allergy or autoimmunity.

TLR AND FC RECEPTOR ACTIVATION IN RESPONSES TO PATHOGENS

The synergy observed upon TLR and Fc receptor triggering reflects a memory response, which is usually characterized by a shorter response time and an enhanced magnitude of the immune response after specific antibodies have been generated (*[5](#page-20-4)*). After primary responses to pathogens, antibodies and T cells can contribute to increased resistance to secondary or chronic infections, and it is this enhanced responsiveness of the immune system that forms the basis for vaccination (Figure 1).

As discussed in the introduction of this thesis, mast cells have been shown to play an important role in the immune response against a variety of pathogens, including bacteria, parasites and viruses. However, only few studies have evaluated the role of mast cells in recall responses against pathogens, when specific antibodies have been formed.

In the case of bacteria, IgG antibodies are most often generated after primary infection. IgG immune complex mediated reactions upon passive antibody transfer in the mouse are dependent on mast cells, in particular through FcγRIII (*[6](#page-20-5)*). The effect of mast cells in such reactions is mainly to recruit neutrophils, eosinophils and other immune cells to the site of antigen exposure. Eosinophil recruitment in a model of passive cutaneous anaphylaxis was augmented by pretreatment with LPS, suggesting that enhanced mast cell responses in presence of TLR ligands and immune complexes may lead to enhanced

protective responses against bacteria (*[7](#page-21-0)*). In addition, neutrophil recruitment upon Helicobacter infection in vaccinated mice was largely reduced in mast cell-deficient animals (*[8,](#page-21-1) [9](#page-21-2)*). The findings of this thesis are in line with these observations, showing that the production of several cytokines and chemokines, known to recruit and activate neutrophils and eosinophils, were markedly enhanced in the presence of combined TLR and Fc receptor activation.

In the case of parasites, most evidences suggest a dominant role for IgE in protection, although other IgA and IgG isotypes have been postulated to play a protective role as well (*[10-12](#page-21-3)*). In mice, protective immunity, such as induced via vaccination, is IgEdependent (*[13,](#page-21-4) [14](#page-21-5)*), and in humans, levels of parasite-specific IgE are correlated with resistance to parasitic infection (*[15-18](#page-21-6)*). Interestingly, a recent study showed that opsonized parasites could be directly killed in vitro by human mast cell-derived tryptase, through the formation of a so-called degranulation synapse (*[19](#page-21-7)*).

In addition to specific antibodies able to enhance mast cell responses during recall immunity, IgE bound to FcεRI can be crosslinked non-specifically by an S mansoni egg antigen and HIV antigen gp120 (*[20,](#page-21-8) [21](#page-21-9)*). Therefore, the synergy observed between TLR signaling and FcεRI crosslinking may also contribute to innate responses to these pathogens.

Figure 1. Synergy between TLR and Fc receptors can contribute to protective immunity against pathogens. Pathogens such as bacteria, parasites and viruses can be recognized by antibodies during secondary responses. These antibodies can trigger mast cell activation. When PAMPs from the pathogens trigger TLR activation, production of cytokines and chemokines by mast cells synergizes, thereby contributing to protective immunity.

Pathogen-specific responses

Importantly, we observed cytokine-specific responses, depending on the TLR that was triggered (Chapters 4 and 5), suggesting that the type of TLR-trigger (which typically depends on the type of pathogen encountered) can fine-tune the response mode of mast cells (Figure 2). There is a remarkable resemblance of the cytokines that were induced by specific TLRs in mast cells with their putative protective role for particular pathogens in vivo. For example, TLR-2 and -4 are mostly known for their involvement in bacterial and parasitic infection (*[22-25](#page-21-10)*). Ligands for these TLRs in mast cells induced a cytokine profile characterized by production of GM-CSF, IL-5, IL-13, and MIP-1ɑ. These cytokines may contribute to anti-bacterial immunity; IL-8 and MIP-1ɑ are particularly known to induce recruitment and activation of neutrophils, an important mechanism for the first line of defense against extracellular bacteria (*[26-28](#page-21-11)*). IL-5 and IL-13, in turn, are potent contributors to the clearance of parasites, through induction of eosinophil activation, mucus production and worm expulsion (*[29-31](#page-22-0)*).

Some of the cytokines involved in mast cell-mediated protection against bacteria in mice were not observed upon triggering of bacteria-associated TLRs in human mast cells. These include TNF-ɑ, CXCL1, CXCL2, IL-4 and IL-6 (*[8,](#page-21-1) [27,](#page-22-1) [32,](#page-22-2) [33](#page-22-3)*). However, for several of these cytokines produced by murine mast cells, their production by their human counterparts is not commonly observed (*[34](#page-22-4)*). Furthermore, the functions of these cytokines in bacterial infections largely overlap with the cytokines we observed, suggesting that human mast cells may serve a similar function be it through production of different cytokines.

In contrast, ligands which are associated with viruses, induced a different cytokine profile, which is more associated with recruitment and activation of T cells and NK cells, such as through production of MCP-1, MIP-1β (Chapter 4 and unpublished observations). Recruitment of T cells and NK cells by mast cells has been shown to contribute to antiviral immunity in mice (*[35-37](#page-22-5)*). Other cytokines produced by mast cells in response to the TLR-8 ligand ssRNA (Eotaxin, GRO-ɑ, TNF-ɑ) are associated to recruitment and activation of eosinophils and neutrophils. Although this process may not be generally appreciated in the context of viral infection, several studies now suggest that eosinophils can play an important role in viral infection, especially during pulmonary infections (*[38-40](#page-22-6)*). Eosinophils can secrete RNAses and cationic granule products, which are known to degrade single stranded RNA (TLR-8 ligand) (*[41-43](#page-22-7)*). Neutrophils can also contribute to anti-viral immunity, for example through secretion of antiviral peptides, formation of NETs, and phagocytosis of infected cells (*[44-46](#page-22-8)*). These results therefore suggest that mast cell-derived cytokines in response to viral TLR ligands may contribute to antiviral immunity through their effects of both innate and adaptive immune cells.

Figure 2. Pathogen-specific responses mediated through triggering of specific TLRs. The type of pathogen can fine-tune the response mode of mast cells. Bacterial and parasitic TLR ligands, through production of specific cytokines in response to triggering of TLR-2 and -4, can induce mucus production and recruitment of granulocytes to the site of infection. In contrast, mast cell-derived cytokines in response to viral TLR ligands are associated with recruitment of T cells and NK cells.

Differential responses determined by the type of TLR have been previously described for different types of dendritic cells and monocytes (*[47-49](#page-23-0)*), suggesting a model where triggering of different TLRs in multiple cell types shape the immune response against different pathogens. In Chapter 2, I described a similar phenomenon for basophils, where the TLRs associated with bacteria and parasites were able to induce type 2 cytokines, IL-4 and IL-13, whereas the TLRs associated with viruses induced secretion of RANTES, suggesting that both mast cells and basophils may contribute to skewing of the immune system to pathogen-specific responses. The specific role of mast cells and basophils seems to be the skewing of type 2 immunity and recruitment of neutrophils and eosinophils, thereby complementing protective responses induced by DCs and other cell types. The further enhancement of these TLR-specific responses by mast cells and basophils in the presence of Fc receptor triggering, suggests that mast cells and basophils may be potent enhancers of pathogen-directed immunity during memory responses.

TLR AND FC RECEPTOR ACTIVATION IN ALLERGY

Besides the role of TLR ligation in pathogen-specific immunity, a role for TLRs has been implicated in allergy as well. For example, viral and bacterial infections have been associated with asthma exacerbations (*[50,](#page-23-1) [51](#page-23-2)*). Furthermore, chronic allergy may lead to secretion of endogenous TLR ligands (*[52](#page-23-3)*), and some allergens have been shown to contain TLR ligands (*[53,](#page-23-4) [54](#page-23-5)*). The results described in this thesis suggest that both basophils and mast cell allergic responses are significantly enhanced in the presence of TLR ligands.

Allergy is usually hallmarked by two important processes. The sensitization phase takes place first, where Th2 and IgE responses against the allergen are initiated. During an allergic reaction, the immune system is triggered by re-exposure to the allergen. This effector phase consists of an FcεRI-dependent acute reaction, and a more prolonged reaction (late-phase reaction) caused by cytokines and inflammatory infiltrates.

TLR ligands during allergic sensitization

During the sensitization phase, allergen-specific Th2 responses are primed. Although dendritic cells are required as antigen presenting cells, they usually do not produce IL-4, a cytokine that is necessary for Th2 priming (*[55](#page-23-6)*). It is unclear which cell type provides these early innate type 2 cytokines, but basophils may contribute to this process, in addition to innate lymphoid cells or naïve T cells (*[56,](#page-23-7) [57](#page-23-8)*). Indeed, basophils were shown to enhance Th2 responses upon house dust mite inhalation in mice (*[58](#page-23-9)*).

The effect of TLR ligands, in particular endotoxin, has been studied in the context of allergic sensitization, although contradicting results have been obtained (*[59](#page-23-10)*). Several studies show that exposure to endotoxin or pathogens (such as in rural areas) is associated with a lower prevalence of allergy (*[60-62](#page-23-11)*). Furthermore, some studies in mice showed that exposure to LPS can reduce allergic sensitization ([63](#page-23-12)), presumably by inducing Th1 responses as a consequence of IL-12 production by dendritic cells in response to LPS (*[59,](#page-23-10) [64,](#page-23-13) [65](#page-23-14)*). In contrast, one study showed that a low level of endotoxin is required to mount a robust Th2 response against allergens, and several studies showed that TLR-4 is required for allergic sensitization (*[58,](#page-23-9) [66,](#page-23-15) [67](#page-24-0)*).

As I showed in this thesis, TLR ligands were able to induce IL-4 production by basophils, thereby providing a link between innate responses and Th2 immunity such as required for allergic sensitization. TLR ligands alone only led to a low level of IL-4, which in itself was not sufficient to induce Th2 skewing. However, TLR ligands together with protease allergens may be able to induce a more robust IL-4 production by basophils (*[68](#page-24-1)*), thereby potentially contributing to allergic sensitization.

TLR ligands during the effector phase of allergic reactions

In contrast to contradicting studies on the role of TLR ligation during allergic sensitization, much more is known about the enhancement of allergic responses by pathogens during the effector phase. Many studies suggest that pulmonary infections, with either virus or bacteria, are associated with asthma exacerbations (*[50,](#page-23-1) [51](#page-23-2)*), In allergic individuals, challenges with combined endotoxin and allergen induced a synergy in neutrophil and eosinophil recruitment to the nasal tissue, or increased wheal and flare reactions in the skin, depending on the tissue where the challenge took place (*[69,](#page-24-2) [70](#page-24-3)*), In experimental asthma in rodents, LPS enhanced eosinophilic airway inflammation (*[66,](#page-23-15) [71](#page-24-4)*). The latter was dependent on TLR-4 expression by mast cells.

Interestingly, we observed a significant enhancement of cytokine production by mast cells upon combined TLR-4 and FcεRI triggering, in particular those cytokines that are known for their role in recruitment and activation of neutrophils and eosinophils (*[71-](#page-24-4) [73](#page-24-4)*). Our results further suggest that basophils are an important source of IL -4 when triggered through TLRs and FcεRI, in line with findings in both human and mouse that basophils are the main source of IL-4 during viral infections and allergen challenge (*[74-](#page-24-5) [76](#page-24-5)*).

Therefore, synergy between TLRs and FcεRI triggering may significantly contribute to allergic reactions, mainly through their actions on basophils and mast cells during the effector phase of allergic responses (Figure 3).

TLR AND FC RECEPTOR ACTIVATION IN AUTOIMMUNITY

As described in chapter 12, many autoantibodies have specificity for TLR ligands, and we propose that this plays a role in the initiation of autoantibody responses, as triggering of TLRs in B cells is thought to mediate tolerance escape. Furthermore, the tissue damage that is associated with chronic inflammation in autoimmune disease often leads to release of endogenous TLR ligands, so-called DAMPs (*[77-81](#page-24-6)*).

Many studies have suggested a role for TLR signaling in the initiation of autoimmune responses, either through its effect on autoantibody production, or through enhancing autoreactive Th cell responses (*[82-89](#page-24-7)*). Knockout animals for TLRs or MyD88 have often reduced autoreactive T cell responses, and such responses are aggravated when immunization is done in the presence of TLR agonists or pathogens (*[90](#page-25-0)*). In vivo, a few studies suggest that TLR ligands can enhance inflammatory responses mediated by autoantibodies. TLR-4 signaling was shown to contribute significantly to thrombosis mediated by anti-phospholipid antibodies (*[91](#page-25-1)*). The use of a TLR-4 antagonist after induction of collagen-induced arthritis led to a reduction in arthritis, suggesting a contribution of TLR-4 to the inflammatory response mediated by Th cells and autoantibodies (*[92](#page-25-2)*). In addition, both TLR-2 and TLR-4 deficiency led to reduced arthritis symptoms in passive serum-transfer induced arthritis (K/BxN) (*[93,](#page-25-3) [94](#page-25-4)*).

Figure 3. Enhancement of allergic responses through triggering of TLR by pathogens or allergens. Allergic exacerbations are associated with the presence of viral or bacterial infections. In addition, some allergens have been suggested to directly trigger TLRs. When TLR ligands derived from these pathogens or allergens are present at the same time as the allergen to which mast cells and basophils are sensitized, synergy in the production of cytokines and chemokines occurs, thereby leading to enhancement of allergic symptoms.

We proposed in chapter 11-13, that TLR ligation may contribute to autoantibody-induced chronic inflammation, through their synergistic action on myeloid cells, including mast cells. Although the functional role of mast cells during autoantibody-mediated autoimmune disease is not yet clear, we were able to show for the first time that human mast cells can be activated by anti-citrullinated protein antibodies. The synergy observed when the activation by autoantibodies was combined with TLR ligands present in synovium of RA patients suggests that mast cells can significantly contribute to inflammatory responses in RA (Figure 4).

IL-33 AS IMMUNOMODULATORY CYTOKINE DURING ANTIBODY-MEDIATED RESPONSES

Whereas TLR-mediated activation of mast cells and basophils can generally lead to enhanced inflammatory responses, IL-33 is associated with modulatory effects, in particular through their interaction with monocytes. IL-33 specifically enhanced the production of type 2 cytokines (IL-5, IL-13, IL-10) induced by IgG immune complexes in mast cells. In basophils, IL-33 enhanced IL-4 and histamine release induced by FcεRI triggering. This led to dampened TNF-ɑ production by monocytes in response to LPS.

Figure 4. Enhancement of autoimmune responses through triggering of TLR and autoantibodies. Autoimmune disease is often associated with the presence of endogenous DAMPs, released as a consequence of tissue damage. Self-antigens against which autoantibodies are produced are also often recognized by TLRs. Combined activation of mast cells through TLR and Fc receptors during autoimmune responses can enhance their cytokine production, thereby leading to enhanced inflammation.

The reason why IL-33 may have dual roles in allergy and autoimmune disease may lie in its ability to specifically drive the release of type 2 cytokines while inhibiting TNF-ɑ by monocytes. As described above, allergy is driven largely by type 2 cytokines, IL-4, IL-5 and IL-13.

Therefore, enhancement of the production of these cytokines by basophils and mast cells may contribute to allergic symptoms. In particular mast cell-derived IL-13 has been shown to contribute to airway hyperresponsiveness in response to IL-33, although IL-13 derived from innate lymphoid cells is likely representing an additional important source of IL-13 as well (*[95-98](#page-25-5)*). Although studies in mice confirm the regulatory action of basophils on the monocyte/macrophage lineage (*[99,](#page-25-6) [100](#page-25-7)*), the strong enhancement of type 2 cytokines by IL-33 directly on basophils and mast cells may dominate these immunomodulatory effects, thereby enhancing allergic inflammation.

In autoimmunity, different effects have been observed with IL-33. Several studies showed a reduction in autoreactive responses, for example through reducing autoreactive Th17 responses or through induction of alternatively activated macrophages (*[101-104](#page-25-8)*). In contrast, IL-33 was shown to exacerbate disease in most, but not all, mouse models of arthritis (*[105-109](#page-25-9)*). Interestingly, IVIg was shown to upregulate IL-33, and, via the release of IL-4 by basophils, induced alternatively activated macrophages, thereby reducing inflammation in a mouse model of arthritis (*[110,](#page-26-0) [111](#page-26-1)*).

The results of this thesis suggest that IL-33 can have both pro-inflammatory and antiinflammatory actions through mast cells and basophils, by enhancing Th2 immune responses and at the same time reducing monocyte-mediated inflammation. This dual role may explain why contrasting results are obtained on the role of IL-33 in autoimmune disease.

T CELL INTERACTIONS

The second part of this thesis focused on the interaction between mast cells and $CD4^+$ T cells. I showed that human mast cells can contribute to T cell activation, through antigen presentation, co-stimulation, and expansion of Th17 cells (Chapters 7-9).

The antigen presenting capacity of mast cells is an area of debate (*[112](#page-26-2)*). Several recent studies, including the results of this thesis suggest that human mast cells can function as antigen presenting cells and can provide co-stimulation to T cells (Figure 5) (*[113,](#page-26-3) [114](#page-26-4)*). In mice, antigen presentation by mast cells can induce activation of memory Th cells, but not, or only poorly, of naïve CD4⁺ T cells, suggesting that mast cells are probably mostly involved in activation of memory $CD4^+$ T cell responses ([115](#page-26-5)). The results of this thesis support this hypothesis. First of all, although HLA class II expressing mast cells were present in tonsil, their frequency and expression levels of HLA class II are low, in particular compared to that of professional antigen presenting cells, such as dendritic cells. Furthermore, we found that human mast cells do not produce any of the cytokines required for skewing of naïve CD4⁺ T cells into classical Th cell subsets, such as IL-4, IL-12, or IL-23 (*[116](#page-26-6)*), in line with their inability to induce skewing of naïve Th cells (Chapter 9). In some studies, protein antigen processing and presentation by mast cells was more efficient when the protein was complexed with IgG or IgE antibodies, suggesting enhanced antigen presentation during memory responses (*[117-119](#page-26-7)*). Together, these findings suggests that mast cells probably do not represent a major cell population involved in the priming of naïve $CD4^+$ T cells in lymphoid organs.

Although the contribution of antigen presentation by mast cells has not been directly addressed in vivo, a number of studies have studied the role of mast cells in modulating T cell responses. First of all, many studies have shown that mast cells can direct the lymph node hypertrophy and migration and activation of both T cells and dendritic cells, for example through releasing exosomes containing TNF (in mice) (*[120-124](#page-26-8)*). This process occurs for example in response to infection, but may also play a role in allergic reactions (*[125-130](#page-26-9)*).

In contrast to these studies suggesting an activating role of mast cells in T cell responses, several studies have also suggested a role for mast cells in perpetuation of Treg responses (*[124,](#page-26-10) [131,](#page-27-0) [132](#page-27-1)*). This effect is most often mediated through mast cell-derived IL-10 (*[133-136](#page-27-2)*). Interestingly, a vast number of studies have shown that Tregs can also regulate mast cell-mediated responses, such as during anaphylaxis, suggesting a bidirectional crosstalk between T cells and mast cells in peripheral tissues (*[137-141](#page-27-3)*). Not much is known about the effect of conventional T cells on mast cell function, although our studies indicated that T cells can modulate mast cell phenotype inducing upregulation of HLA class II.

In addition to their role as accessory cell in priming of $CD4⁺$ T cell responses by dendritic cells, and their role in activating Treg cells, mast cells may play a role in the promotion of proinflammatory Th cell responses. This has been studied in particular in the context of autoimmunity. In EAE, a mouse model of multiple sclerosis, mast cells were required for activation of adoptively transferred T cells (*[142](#page-27-4)*). In line with this, both total and collagenspecific Th17 cells were reduced in mast cell deficient mice upon induction of collageninduced arthritis (*[143](#page-27-5)*). What determines this balance between promoting proinflammatory Th cells or Treg cells is not clear, but it seems to be influenced mostly by the inflammatory context; e.g. infection versus tolerance (*[144](#page-27-6)*). In agreement with this, one study showed that mast cells can convert Treg into Th17 cells during autoimmune disease (EAE) in mice, suggesting that mast cells can alter T cell phenotype when tolerance breaks (*[145](#page-27-7)*).

Figure 5. Mast cells interactions with CD4⁺ T cells lead to T cell activation and Th17 expansion. The results of this thesis show that human mast cells can directly activate T cells by providing all 3 signals required for T cell effector function: 1) Antigen presentation through HLA class II; 2) Co-stimulation, through a CD28 independent mechanism; 3) Expansion of Th17 cells through inflammasome-independent IL-1β.

Importantly, our study showed that mast cells could specifically expand Th17 cells when they were activated through TLR or Fc receptors. Both chronic allergy and autoimmunity are often associated with Th17 cell responses, which are thought to contribute to disease, through neutrophil recruitment as well as tissue-specific effects (*[146-153](#page-27-8)*). Our findings of Th17 cell expansion upon Fc receptor triggering of mast cells provides a potential link between antibody-mediated inflammation and pathogenic Th17 responses. These results together suggest that human mast cells have the complete molecular makeup to induce robust T cell activation, and when activated can drive the expansion of Th17 cells.

CHRONIC INFLAMMATION

Although the role of specific receptors such as TLRs and Fc receptors in chronic inflammation become increasingly known, there is a limited understanding of the behavior of immune cells during chronic inflammation. In chapter 10, we attempted to gain understanding of the function of mast cells in chronic Fc receptor mediated responses. We hypothesized that repeated activation through FcεRI could lead to mast cell-intrinsic changes reflecting what happens during chronic Fc receptor activation in tissue. We observed several changes in mast cell phenotype and function, which were correlated to gene expression in chronic allergy. These changes modified mast cell function, by increasing their expression of molecules involved in antigen processing and presentation, increasing their responsiveness to TLR stimulation, and increased expression and production of chemokines (Figure 6).

CHEMOTAXIS

Although some of the secreted molecules involved in acute IgE-mediated mast cell responses were downregulated, mast cells remained fully able to degranulate, and had increased expression of several chemokines, including CCL18 (Table 1). The chemokines that were upregulated are mostly involved in recruitment of T cells and granulocytes. As infiltration of these cell types can lead to local inflammation, epitope spreading and tissue remodeling, this suggests that mast cells, by retaining or even increasing their chemotactic capacity, may contribute to chronic inflammation by sustaining local leukocyte infiltration.

Interestingly, CCL18, the most highly upregulated gene, is specifically upregulated in chronic inflammatory conditions, such as different forms of chronic allergy and chronic autoimmune diseases (*[154-156](#page-28-0)*). There is no mouse homologue of this chemokine, therefore its role in vivo has been difficult to establish. CCL18 has been mainly shown to

play a role in chemotaxis, in particular that of naïve T cells and memory Th2 cells (*[157-](#page-28-1) [160](#page-28-1)*).

These results suggest that the chemotactic function of mast cells changes during chronic inflammatory stimulation, and can induce several pro-inflammatory effects including persistent recruitment of granulocytes and T cells.

Table 1. Chemokines specifically upregulated after repeated stimulation of mast cells through FcERI

Data obtained from gene expression analysis as shown in Chapter 10; information about chemokine receptors and recruited cell types were obtained from Zlotnik et al. (*[161](#page-28-2)*)

T CELL ACTIVATION

After repeated Fc receptor triggering of mast cells, we also observed enhanced expression of several genes involved in antigen processing and presentation. As described in the part about mast cell-T cell interactions, we showed that human mast cells can function as antigen presenting cells, and that these results therefore may have important implications for the capacity of mast cells to activate CD4⁺ T cells. CD4⁺ T cells play an important role in chronic inflammation, for example through driving the production of allergen-specific antibodies or autoantibodies (*[162-165](#page-28-3)*).

Although not much is known about the exact antigen processing pathways in mast cells, repeated triggering through Fc receptors induced upregulated expression of HLA class II molecules, costimulatory molecules (e.g. CD86), and cathepsins S. Therefore, our results suggest that mast cells may have increased capacity for antigen presentation during chronic Fc receptor-mediated inflammation, thereby potentially contributing to T cell activation in the local tissue environment.

TLR RESPONSES

Another important function of mast cells that was enhanced after repeated stimulation through Fc receptors was their response to bacteria. This enhancement was characterized by upregulated expression of several TLRs, and we were also able to show increased cytokine production in response to LPS, a prototype TLR-4 ligand.

Activation of TLRs is thought to initiate a positive feedback loop of inflammation through inducing tissue and cellular damage thereby leading to sustained release of endogenous TLR ligands (*[80,](#page-24-8) [81](#page-24-9)*). Interestingly, those TLRs most well-known for their involvement in responses to DAMPs, TLR-2 and -4, were upregulated in mast cells after repeated Fc receptor triggering (*[166](#page-28-4)*). Macrophages in synovium of RA patients were also found to exhibit increased expression of TLR-2 and -4, suggesting that upregulation of TLRs may be a common mechanism in different myeloid cell types during chronic inflammation (*[167](#page-28-5)*).

Given the importance of TLRs in the response to endogenous DAMPs, these findings suggests that TLR responses may be enhanced during chronic antibody-mediated responses and that this enhanced TLR responsiveness by mast cells may contribute to this positive feedback loop during chronic inflammation.

TISSUE REMODELING

Chronic inflammation is often characterized by tissue remodeling, leading to long-term, irreversible changes in tissue homeostasis. Possibly the most striking example is rheumatoid arthritis, where loss of bone and cartilage are characteristics of such tissue remodeling.

One of the biological pathways significantly upregulated in mast cells after repeated Fc receptor triggering was wound healing, a process closely related to tissue remodeling, and involving several different processes including coagulation, inflammation, angiogenesis, and tissue regeneration. We observed upregulated gene expression of several tissue-remodeling enzymes and extracellular structural proteins (MMP25, osteopontin, and possibly PADI4) allowing for a direct influence of mast cells on tissue homeostasis (*[168-171](#page-28-6)*).

In addition to these direct effects, mast cells have been shown to have indirect effects as well, such as through activation of stromal cells, and cleavage of pro-MMP enzymes secreted by other cell types (*[172-174](#page-28-7)*). For example, CCL18, although most well-known for its chemotactic functions, has also been shown to induce fibroblast activation and collagen production (*[175](#page-29-0)*). In addition, besides its role in bone metabolism, osteopontin

is associated with a variety of effects associated with tissue repair and fibrosis, presumably through its effect on fibroblasts, epithelial cells and macrophages (*[176-179](#page-29-1)*).

Importantly, although we used an in vitro model of mast cell activation, several changes observed were linked to gene expression in chronic inflammatory conditions. For example, both CCL18, osteopontin are highly upregulated in asthma and allergy, and were associated with markers of tissue remodeling in several chronic inflammatory diseases (*[180-189](#page-29-2)*). These molecules were also found to be expressed by mast cells in such chronic inflammatory diseases when analysed ex vivo (*[180,](#page-29-2) [190](#page-29-3)*). Therefore, our results show that mast cells upregulate several molecules involved in tissue remodeling after repeated Fc receptor triggering, suggesting that mast cells can significantly contribute to chronic inflammation through modulation of tissue homeostasis.

RESOLUTION OF INFLAMMATION

Inflammatory responses are tightly regulated to prevent tissue damage. When the inflammatory stimulus is eliminated, a resolution phase is initiated, characterized by phagocytosis of apoptotic neutrophils, the production of pro-resolving lipid mediators and anti-inflammatory molecules, as well as removal of tissue debris (*[191-193](#page-29-4)*).

Repeated stimulation of mast cells led to several changes that could potentially contribute to resolution of inflammation. First of all, we observed a dampening of the acute mast cell responses, such as several cytokines and chemokines. Furthermore, the gene expression of several regulatory molecules was upregulated. These include a number of inhibitory receptors (LAIR1, LILRB2, LILRB3, and VSTM1). These and other ITIM motif-bearing inhibitory receptors have been shown to potently inhibit mast cell responses through FcεRI (*[194-198](#page-30-0)*). Therefore, upregulation of these receptors may contribute to dampening of mast cell responses. In addition, heme oxygenase-1 (HO-1, HMOX1), a molecule with anti-inflammatory capacities, was upregulated in mast cells after repeated activation. HO-1 is most well-known as a stress-induced molecule that can suppress activation of myeloid cells (*[199-201](#page-30-1)*). Not much is known about the role of HO-1 in mast cells, but molecules that can upregulate expression of HO-1 diminished mast cell activation, suggesting an immunoregulatory role of HO-1 in mast cells as well (*[202,](#page-30-2) [203](#page-30-3)*).

Together, these results suggest that in addition to enhanced proinflammatory responses, mast cells also initiate an anti-inflammatory program, characterized by upregulation of immunoregulatory molecules, which could contribute to tissue protection. However, our results also show, that when antigen cannot be removed, therefore leading to repeated or continuous stimulation of mast cells through Fc receptors, proinflammatory effects may dominate over these anti-inflammatory mechanisms.

Figure 6. Potential roles of mast cells in chronic antibody-mediated inflammation. Upon repeated Fc receptor triggering, we observed changes in several processes related to chronic inflammation: 1) upregulation of inhibitory receptors, potentially contributing to resolution of inflammation; 2) enhanced antigen processing and presentation through HLA class II; 3) changes in production of chemokines; 4) enhanced TLR responses; 5) upregulation of tissue-remodeling enzymes.

IMPLICATIONS FOR AUTOIMMUNE DISEASE

Autoimmune diseases are characterized by immune responses during which the inflammatory stimulus (self-antigen) cannot be eliminated, leading to chronic inflammation. Our results suggest that mast cells can significantly contribute to chronic inflammation after repeated Fc receptor triggering, through changes in chemotaxis, enhancing T cell activation, upregulating their responsiveness to DAMPs, and upregulation of molecules involved in tissue remodeling. As we only studied FcεRI, it is unknown whether similar processes are upregulated upon repeated triggering of FcγRs in mast cells, which would be highly important in the context of autoimmunity. However, the mast cell response to triggering through FcεRI and FcγRIIA overlap considerably, suggesting similar responses may occur upon repeated stimulation of mast cells with IgG immune complexes (*[19](#page-21-7)*). Interestingly, the chemokine CCL18, which was highly

upregulated in mast cells after repeated FcεRI stimulation, has also been found in increased levels of expression in synovium of RA patients, suggesting that this molecule may be upregulated in mast cells during chronic autoimmune disease as well (*[204-206](#page-30-4)*). Even more interesting, synovial fluid levels of CCL18 were associated with levels of RF, one of the autoantibodies in RA, suggesting that repeated Fc receptor activation as observed in mast cells can present a novel pathway for secretion of CCL18 in response to autoantibodies (*[207](#page-30-5)*). Other molecules upregulated in mast cells after repeated Fc receptor triggering (such as osteopontin) are also associated with autoimmune disease and autoantibodies (*[208,](#page-30-6) [209](#page-30-7)*). Therefore, these studies may provide more insight into the role of Fc receptor activation during chronic autoimmune disease.

IMPLICATIONS FOR THERAPY OF CHRONIC INFLAMMATORY DISEASES

In chapter 13, we postulated several novel therapeutic approaches for the treatment of autoimmune diseases. As several pathogenic mechanisms overlap between autoimmune disease and chronic allergy, similar strategies may be employed for both types of diseases. Here, I will discuss the implications of these therapies for mast cell-mediated responses during chronic inflammation.

We described therapeutic targets that could influence the interaction between dendritic cells and T cells. Whereas dendritic cells are often viewed as the most potent antigen presenting cells, the second part of this thesis showed that mast cells can also function as antigen presenting cells and can specifically enhance Th17 responses, an effect of mast cells that was also shown in vivo in an mouse arthritis model (*[143](#page-27-5)*). Furthermore, several molecules involved in antigen processing and presentation had increased gene expression in our in vitro model of chronic inflammation, suggesting that chronic inflammatory responses could further enhance T cell activation by mast cells. Therefore, strategies to modulate APC-T cell interactions could inhibit T cell activation by mast cells as well.

First of all, we proposed to modulate activation of antigen presenting cells (dendritic cells and monocytes) by C1q or C1q-like agonists through its effect on LAIR1. In monocytes and dendritic cells, C1q has been shown to inhibit cytokine secretion and maturation, thereby potentially decreasing T cell activation or skewing by cytokines (*[210,](#page-30-8) [211](#page-30-9)*). Human mast cells also express LAIR1, but the effect of C1q and the functional consequences of ligation of LAIR1 in mast cells has not been studied (*[212](#page-30-10)*). However, it is conceivable that LAIR1 could modulate mast cell function in a similar manner as that of monocytes and dendritic cells, allowing for inhibition of mast cell activation through LAIR1 as therapeutic targets. As mast cell activation was shown to specifically enhance Th17 cell expansion, inhibition of mast cells using C1q has the potential to modulate chronic inflammatory diseases through its effect on Th17 responses.

Another therapeutic target that we identified to influence T cell activation in autoimmune disease was through inhibition of the presentation of self-antigens to autoreactive T cells by modulation of the levels of Blimp-1, IRF4 or cathepsin S in dendritic cells. Although the expression or function of Blimp-1 or IRF4 in mast cells is not established, we observed increased gene expression of cathepsin S in mast cells after repeated stimulation through Fc receptors. Cathepsin S in mast cells has been shown to be involved in processing of granule proteases, but its role in antigen processing has not been studied (*[213](#page-31-0)*). However, given its central role in antigen processing in other cell types, inhibitors of cathepsin S may potentially modulate antigen processing by mast cells during chronic inflammatory conditions.

This thesis further shows that co-stimulation of T cells by mast cells is B7/CD28 independent. Although blockade of CD28 co-stimulation by CTLA4-Ig (Abatacept) is an effective treatment for autoimmune diseases, inhibition of T cell activation with this therapy is not complete (*[214-217](#page-31-1)*). Furthermore, memory Th17 cells, an important T cell subset in various chronic inflammatory diseases, have been shown to be resistant to inhibition by CTLA-4-Ig, and a recent study indeed showed that treatment of RA patients with CTLA-4-Ig most potently inhibited Th1 responses and had no effect on Th17 cells (*[218,](#page-31-2) [219](#page-31-3)*). This is interesting as we showed that mast cells specifically enhance Th17 responses, and that T cell co-stimulation by mast cells was independent of B7/CD28. Therefore, blocking the interaction of antigen presenting cells and Th17 cells requires additional blockade besides B7/CD28, and inhibiting the interaction between mast cells and CD4⁺ T cells could potentially contribute to reducing Th17 cells. More research into the exact pathways that mediate T cell co-stimulation by mast cells may further contribute to inhibition of T cell activation in chronic inflammatory conditions.

Another therapeutic strategy that could affect mast cell-mediated responses is the use of decoy antigens. We proposed in chapter 13 that the use of decoy antigens can reduce the chronicity of inflammatory responses by neutralization of autoantibodies. As reviewed in chapter 12, autoantibodies often target ubiquitously expressed intracellular molecules, which need to be released to the extracellular environment, to allow for binding by autoantibodies. This process can initiate inflammatory responses leading to tissue damage. Inflammation is sustained as the tissue damage can lead to additional release of self-antigen and DAMPs, which further activate myeloid cells and B cells, thereby creating a vicious cycle of tissue damage and autoantibody production. Decoy antigens can bind autoreactive B cell receptors and secreted antibodies, and should be

designed in such as way that they do not trigger TLRs. They can thereby block activation of B cells as well as myeloid cells, and can inhibit tissue damage employed by immune complexes.

Although this strategy would not target mast cells specifically, they are an important target cell type in chronic inflammatory responses, as mast cells express only activating Fc receptors (in most tissues), and as they can release inflammatory cytokines as well as tissue modifying enzymes. We showed in this thesis that combined triggering through Fc receptors and TLR in mast cells can greatly enhance their cytokine release. Furthermore, repeated Fc receptor triggering augmented their responsiveness to TLR ligands. Therefore, blocking of Fc receptor activation in mast cells using decoy antigens has the potential to greatly reduce mast cell activation, also indirectly by reducing their response to DAMPs. Furthermore, repeated Fc receptor triggering led to a modulation of the mast cell response mode, characterized by upregulation of the expression of many genes involved in chronic inflammation, and it is likely that such effects are present in other cell types as well. Therefore, blocking Fc receptor activation might prevent the long-term effects of chronic activation on a cellular level, thereby breaking the vicious cycle of antibody-mediated inflammation.

In conclusion, in this thesis I showed that mast cells can significantly contribute to chronic inflammation through their activation by Fc receptors and TLRs, as well as their interaction with CD4⁺ T cells, thereby increasing our understanding of their role in allergy and autoimmunity and providing several therapeutic targets to prevent mast cellmediated immune responses.

REFERENCES

1. Vogelpoel LT, Hansen IS, Rispens T *et al.*, Fc gamma receptor-TLR cross-talk elicits proinflammatory cytokine production by human M2 macrophages. *Nat Commun* 5, 5444 (2014).

2. den Dunnen J, Vogelpoel LT, Wypych T *et al.*, IgG opsonization of bacteria promotes Th17 responses via synergy between TLRs and FcgammaRIIa in human dendritic cells. *Blood* 120, 112-21 (2012).

3. Qiao H, Andrade MV, Lisboa FA, Morgan K, Beaven MA, FcepsilonR1 and toll-like receptors mediate synergistic signals to markedly augment

production of inflammatory cytokines in murine mast cells. *Blood* 107, 610-8 (2006).

4. O'Neill LA, Golenbock D, Bowie AG, The history of Toll-like receptors - redefining innate immunity. *Nat Rev Immunol* 13, 453-60 (2013).

5. Abraham SN, St John AL, Mast cellorchestrated immunity to pathogens. *Nat Rev Immunol* 10, 440-52 (2010).

6. Sylvestre DL, Ravetch JV, A dominant role for mast cell Fc receptors in the Arthus reaction. *Immunity* 5, 387-90 (1996).

7. Macari DM, Teixeira MM, Hellewell PG, Priming of eosinophil recruitment in vivo by LPS pretreatment. *J Immunol* 157, 1684-92 (1996).

8. Ding H, Nedrud JG, Wershil B *et al.*, Partial protection against Helicobacter pylori in the absence of mast cells in mice. *Infect Immun* 77, 5543-50 (2009).

9. Velin D, Bachmann D, Bouzourene H, Michetti P, Mast cells are critical mediators of vaccineinduced Helicobacter clearance in the mouse model. *Gastroenterology* 129, 142-55 (2005).

10. Khalife J, Dunne DW, Richardson BA *et al.*, Functional role of human IgG subclasses in eosinophil-mediated killing of schistosomula of Schistosoma mansoni. *J Immunol* 142, 4422-7 (1989).

11. Gaze S, Driguez P, Pearson MS *et al.*, An immunomics approach to schistosome antigen discovery: antibody signatures of naturally resistant and chronically infected individuals from endemic areas. *PLoS Pathog* 10, e1004033 (2014).

12. Grzych JM, Grezel D, Xu CB *et al.*, IgA antibodies to a protective antigen in human Schistosomiasis mansoni. *J Immunol* 150, 527-35 (1993).

13. Abraham D, Leon O, Schnyder-Candrian S *et al.*, Immunoglobulin E and eosinophil-dependent protective immunity to larval Onchocerca volvulus in mice immunized with irradiated larvae. *Infect Immun* 72, 810-7 (2004).

14. Gurish MF, Bryce PJ, Tao H *et al.*, IgE enhances parasite clearance and regulates mast cell responses in mice infected with Trichinella spiralis. *J Immunol* 172, 1139-45 (2004).

15. Faulkner H, Turner J, Kamgno J *et al.*, Age- and infection intensity-dependent cytokine and antibody production in human trichuriasis: the importance of IgE. *J Infect Dis* 185, 665-72 (2002).

16. Rihet P, Demeure CE, Bourgois A, Prata A, Dessein AJ, Evidence for an association between human resistance to Schistosoma mansoni and high anti-larval IgE levels. *Eur J Immunol* 21, 2679-86 (1991).

17. Hagan P, Blumenthal UJ, Dunn D, Simpson AJ, Wilkins HA, Human IgE, IgG4 and resistance to reinfection with Schistosoma haematobium. *Nature* 349, 243-5 (1991).

18. Jiz M, Friedman JF, Leenstra T *et al.*, Immunoglobulin E (IgE) responses to paramyosin predict resistance to reinfection with Schistosoma japonicum and are attenuated by IgG4. *Infect Immun* 77, 2051-8 (2009).

19. Joulia R, Gaudenzio N, Rodrigues M *et al.*, Mast cells form antibody-dependent degranulatory synapse for dedicated secretion and defence. *Nat Commun* 6, 6174 (2015).

20. Schramm G, Mohrs K, Wodrich M *et al.*, Cutting edge: IPSE/alpha-1, a glycoprotein from Schistosoma mansoni eggs, induces IgE-dependent, antigen-independent IL-4 production by murine basophils in vivo. *J Immunol* 178, 6023-7 (2007).

21. Patella V, Florio G, Petraroli A, Marone G, HIV-1 gp120 induces IL-4 and IL-13 release from human Fc epsilon RI+ cells through interaction with the VH3 region of IgE. *J Immunol* 164, 589-95 (2000).

22. van der Kleij D, Latz E, Brouwers JF *et al.*, A novel host-parasite lipid cross-talk. Schistosomal lyso-phosphatidylserine activates toll-like receptor 2 and affects immune polarization. *J Biol Chem* 277, 48122-9 (2002).

23. Goodridge HS, Marshall FA, Else KJ *et al.*, Immunomodulation via novel use of TLR4 by the filarial nematode phosphorylcholine-containing secreted product, ES-62. *J Immunol* 174, 284-93 (2005).

24. Thomas PG, Carter MR, Atochina O *et al.*, Maturation of dendritic cell 2 phenotype by a helminth glycan uses a Toll-like receptor 4 dependent mechanism. *J Immunol* 171, 5837-41 (2003).

25. Akira S, Uematsu S, Takeuchi O, Pathogen recognition and innate immunity. *Cell* 124, 783-801 (2006).

26. Urban CF, Lourido S, Zychlinsky A, How do microbes evade neutrophil killing? *Cell Microbiol* 8, 1687-96 (2006).

14

27. Sutherland RE, Olsen JS, McKinstry A, Villalta SA, Wolters PJ, Mast cell IL-6 improves survival from Klebsiella pneumonia and sepsis by enhancing neutrophil killing. *J Immunol* 181, 5598-605 (2008).

28. Malaviya R, Ikeda T, Ross E, Abraham SN, Mast cell modulation of neutrophil influx and bacterial clearance at sites of infection through TNF-alpha. *Nature* 381, 77-80 (1996).

29. Sakamoto Y, Hiromatsu K, Ishiwata K *et al.*, Chronic intestinal nematode infection induces Stat6 independent interleukin-5 production and causes eosinophilic inflammatory responses in mice. *Immunology* 112, 615-23 (2004).

30. Le Goff L, Loke P, Ali HF, Taylor DW, Allen JE, Interleukin-5 is essential for vaccine-mediated immunity but not innate resistance to a filarial parasite. *Infect Immun* 68, 2513-7 (2000).

31. Oeser K, Schwartz C, Voehringer D, Conditional IL-4/IL-13-deficient mice reveal a critical role of innate immune cells for protective immunity against gastrointestinal helminths. *Mucosal Immunol*, (2014).

32. De Filippo K, Dudeck A, Hasenberg M *et al.*, Mast cell and macrophage chemokines CXCL1/CXCL2 control the early stage of neutrophil recruitment during tissue inflammation. *Blood* 121, 4930-7 (2013).

33. Rodriguez AR, Yu JJ, Guentzel MN *et al.*, Mast cell TLR2 signaling is crucial for effective killing of Francisella tularensis. *J Immunol* 188, 5604-11 (2012).

34. Bischoff SC, Role of mast cells in allergic and non-allergic immune responses: comparison of human and murine data. *Nat Rev Immunol* 7, 93-104 (2007).

35. Orinska Z, Bulanova E, Budagian V *et al.*, TLR3 induced activation of mast cells modulates CD8+ Tcell recruitment. *Blood* 106, 978-87 (2005).

36. St John AL, Rathore AP, Yap H *et al.*, Immune surveillance by mast cells during dengue infection promotes natural killer (NK) and NKT-cell recruitment and viral clearance. *Proc Natl Acad Sci U S A* 108, 9190-5 (2011).

37. Burke SM, Issekutz TB, Mohan K *et al.*, Human mast cell activation with virus-associated stimuli leads to the selective chemotaxis of natural killer cells by a CXCL8-dependent mechanism. *Blood* 111, 5467-76 (2008).

38. Su YC, Townsend D, Herrero LJ *et al.*, Dual proinflammatory and antiviral properties of pulmonary eosinophils in respiratory syncytial virus vaccine-enhanced disease. *J Virol* 89, 1564-78 (2015).

39. Percopo CM, Dyer KD, Ochkur SI *et al.*, Activated mouse eosinophils protect against lethal respiratory virus infection. *Blood* 123, 743-52 (2014).

40. Phipps S, Lam CE, Mahalingam S *et al.*, Eosinophils contribute to innate antiviral immunity and promote clearance of respiratory syncytial virus. *Blood* 110, 1578-86 (2007).

41. Domachowske JB, Dyer KD, Adams AG, Leto TL, Rosenberg HF, Eosinophil cationic protein/RNase 3 is another RNase A-family ribonuclease with direct antiviral activity. *Nucleic Acids Res* 26, 3358-63 (1998).

42. Domachowske JB, Dyer KD, Bonville CA, Rosenberg HF, Recombinant human eosinophilderived neurotoxin/RNase 2 functions as an effective antiviral agent against respiratory syncytial virus. *J Infect Dis* 177, 1458-64 (1998).

43. Rugeles MT, Trubey CM, Bedoya VI *et al.*, Ribonuclease is partly responsible for the HIV-1 inhibitory effect activated by HLA alloantigen recognition. *AIDS* 17, 481-6 (2003).

44. Tripathi S, White MR, Hartshorn KL, The amazing innate immune response to influenza A virus infection. *Innate Immun* 21, 73-98 (2015).

45. Saitoh T, Komano J, Saitoh Y *et al.*, Neutrophil extracellular traps mediate a host defense response to human immunodeficiency virus-1. *Cell Host Microbe* 12, 109-16 (2012).

46. Hashimoto Y, Moki T, Takizawa T, Shiratsuchi A, Nakanishi Y, Evidence for phagocytosis of influenza virus-infected, apoptotic cells by neutrophils and macrophages in mice. *J Immunol* 178, 2448-57 (2007).

47. Kadowaki N, Ho S, Antonenko S *et al.*, Subsets of human dendritic cell precursors express different toll-like receptors and respond to different microbial antigens. *J Exp Med* 194, 863-9 (2001).

48. Jarrossay D, Napolitani G, Colonna M, Sallusto F, Lanzavecchia A, Specialization and complementarity in microbial molecule recognition by human myeloid and plasmacytoid dendritic cells. *Eur J Immunol* 31, 3388-93 (2001).

49. Napolitani G, Rinaldi A, Bertoni F, Sallusto F, Lanzavecchia A, Selected Toll-like receptor agonist combinations synergistically trigger a T helper type 1-polarizing program in dendritic cells. *Nat Immunol* 6, 769-76 (2005).

50. Message SD, Laza-Stanca V, Mallia P *et al.*, Rhinovirus-induced lower respiratory illness is increased in asthma and related to virus load and Th1/2 cytokine and IL-10 production. *Proc Natl Acad Sci U S A* 105, 13562-7 (2008).

51. Sykes A, Johnston SL, Etiology of asthma exacerbations. *J Allergy Clin Immunol* 122, 685-8 (2008).

52. Willart MA, Lambrecht BN, The danger within: endogenous danger signals, atopy and asthma. *Clin Exp Allergy* 39, 12-9 (2009).

53. Michel O, Kips J, Duchateau J *et al.*, Severity of asthma is related to endotoxin in house dust. *Am J Respir Crit Care Med* 154, 1641-6 (1996).

54. Trompette A, Divanovic S, Visintin A *et al.*, Allergenicity resulting from functional mimicry of a Toll-like receptor complex protein. *Nature* 457, 585- 8 (2009).

55. Zhu J, Yamane H, Paul WE, Differentiation of effector CD4 T cell populations (*). *Annu Rev Immunol* 28, 445-89 (2010).

56. Liang G, Barker T, Xie Z *et al.*, Naive T cells sense the cysteine protease allergen papain through protease-activated receptor 2 and propel TH2 immunity. *J Allergy Clin Immunol* 129, 1377-86 e13 (2012).

57. Halim TY, Steer CA, Matha L *et al.*, Group 2 innate lymphoid cells are critical for the initiation of

adaptive T helper 2 cell-mediated allergic lung inflammation. *Immunity* 40, 425-35 (2014).

58. Hammad H, Plantinga M, Deswarte K *et al.*, Inflammatory dendritic cells--not basophils--are necessary and sufficient for induction of Th2 immunity to inhaled house dust mite allergen. *J Exp Med* 207, 2097-111 (2010).

59. Liu AH, Endotoxin exposure in allergy and asthma: reconciling a paradox. *J Allergy Clin Immunol* 109, 379-92 (2002).

60. Matricardi PM, Rosmini F, Riondino S *et al.*, Exposure to foodborne and orofecal microbes versus airborne viruses in relation to atopy and allergic asthma: epidemiological study. *BMJ* 320, 412-7 (2000).

61. Riedler J, Braun-Fahrlander C, Eder W *et al.*, Exposure to farming in early life and development of asthma and allergy: a cross-sectional survey. *Lancet* 358, 1129-33 (2001).

62. Gereda JE, Leung DY, Thatayatikom A *et al.*, Relation between house-dust endotoxin exposure, type 1 T-cell development, and allergen sensitisation in infants at high risk of asthma. *Lancet* 355, 1680-3 (2000).

63. Tulic MK, Wale JL, Holt PG, Sly PD, Modification of the inflammatory response to allergen challenge after exposure to bacterial lipopolysaccharide. *Am J Respir Cell Mol Biol* 22, 604- 12 (2000).

64. Langenkamp A, Messi M, Lanzavecchia A, Sallusto F, Kinetics of dendritic cell activation: impact on priming of TH1, TH2 and nonpolarized T cells. *Nat Immunol* 1, 311-6 (2000).

65. Manetti R, Parronchi P, Giudizi MG *et al.*, Natural killer cell stimulatory factor (interleukin 12 [IL-12]) induces T helper type 1 (Th1)-specific immune responses and inhibits the development of IL-4-producing Th cells. *J Exp Med* 177, 1199-204 (1993).

66. Eisenbarth SC, Piggott DA, Huleatt JW *et al.*, Lipopolysaccharide-enhanced, toll-like receptor 4 dependent T helper cell type 2 responses to inhaled antigen. *J Exp Med* 196, 1645-51 (2002).

14

67. Schmidt M, Raghavan B, Muller V *et al.*, Crucial role for human Toll-like receptor 4 in the development of contact allergy to nickel. *Nat Immunol* 11, 814-9 (2010).

68. Rosenstein RK, Bezbradica JS, Yu S, Medzhitov R, Signaling pathways activated by a protease allergen in basophils. *Proc Natl Acad Sci U S A* 111, E4963-71 (2014).

69. Eldridge MW, Peden DB, Allergen provocation augments endotoxin-induced nasal inflammation in subjects with atopic asthma. *J Allergy Clin Immunol* 105, 475-81 (2000).

70. Michel O, Ginanni R, Le Bon B, Duchateau J, Effect of endotoxin contamination on the antigenic skin test response. *Ann Allergy* 66, 39-42 (1991).

71. Nigo YI, Yamashita M, Hirahara K *et al.*, Regulation of allergic airway inflammation through Toll-like receptor 4-mediated modification of mast cell function. *Proc Natl Acad Sci U S A* 103, 2286-91 (2006).

72. Foster PS, Hogan SP, Ramsay AJ, Matthaei KI, Young IG, Interleukin 5 deficiency abolishes eosinophilia, airways hyperreactivity, and lung damage in a mouse asthma model. *J Exp Med* 183, 195-201 (1996).

73. Weber FC, Nemeth T, Csepregi JZ *et al.*, Neutrophils are required for both the sensitization and elicitation phase of contact hypersensitivity. *J Exp Med* 212, 15-22 (2015).

74. Moore ML, Newcomb DC, Parekh VV *et al.*, STAT1 negatively regulates lung basophil IL-4 expression induced by respiratory syncytial virus infection. *J Immunol* 183, 2016-26 (2009).

75. Schroeder JT, Lichtenstein LM, Roche EM, Xiao H, Liu MC, IL-4 production by human basophils found in the lung following segmental allergen challenge. *J Allergy Clin Immunol* 107, 265-71 (2001).

76. Mukai K, Matsuoka K, Taya C *et al.*, Basophils play a critical role in the development of IgEmediated chronic allergic inflammation independently of T cells and mast cells. *Immunity* 23, 191-202 (2005).

77. Seong SY, Matzinger P, Hydrophobicity: an ancient damage-associated molecular pattern that initiates innate immune responses. *Nat Rev Immunol* 4, 469-78 (2004).

78. Chen CJ, Kono H, Golenbock D *et al.*, Identification of a key pathway required for the sterile inflammatory response triggered by dying cells. *Nat Med* 13, 851-6 (2007).

79. Gondokaryono SP, Ushio H, Niyonsaba F *et al.*, The extra domain A of fibronectin stimulates murine mast cells via toll-like receptor 4. *J Leukoc Biol* 82, 657-65 (2007).

80. Midwood K, Sacre S, Piccinini AM *et al.*, Tenascin-C is an endogenous activator of Toll-like receptor 4 that is essential for maintaining inflammation in arthritic joint disease. *Nat Med* 15, 774-80 (2009).

81. Piccinini AM, Midwood KS, DAMPening inflammation by modulating TLR signalling. *Mediators Inflamm* 2010, (2010).

82. Reynolds JM, Martinez GJ, Chung Y, Dong C, Toll-like receptor 4 signaling in T cells promotes autoimmune inflammation. *Proc Natl Acad Sci U S A* 109, 13064-9 (2012).

83. Shi G, Vistica BP, Nugent LF *et al.*, Differential involvement of Th1 and Th17 in pathogenic autoimmune processes triggered by different TLR ligands. *J Immunol* 191, 415-23 (2013).

84. Viglianti GA, Lau CM, Hanley TM *et al.*, Activation of autoreactive B cells by CpG dsDNA. *Immunity* 19, 837-47 (2003).

85. Ehlers M, Fukuyama H, McGaha TL, Aderem A, Ravetch JV, TLR9/MyD88 signaling is required for class switching to pathogenic IgG2a and 2b autoantibodies in SLE. *J Exp Med* 203, 553-61 (2006).

86. Christensen SR, Shupe J, Nickerson K *et al.*, Toll-like receptor 7 and TLR9 dictate autoantibody specificity and have opposing inflammatory and regulatory roles in a murine model of lupus. *Immunity* 25, 417-28 (2006).

87. Lau CM, Broughton C, Tabor AS *et al.*, RNAassociated autoantigens activate B cells by combined

B cell antigen receptor/Toll-like receptor 7 engagement. *J Exp Med* 202, 1171-7 (2005).

88. Chaturvedi A, Dorward D, Pierce SK, The B cell receptor governs the subcellular location of Toll-like receptor 9 leading to hyperresponses to DNAcontaining antigens. *Immunity* 28, 799-809 (2008).

89. Reynolds JM, Pappu BP, Peng J *et al.*, Toll-like receptor 2 signaling in CD4(+) T lymphocytes promotes T helper 17 responses and regulates the pathogenesis of autoimmune disease. *Immunity* 32, 692-702 (2010).

90. Mills KH, TLR-dependent T cell activation in autoimmunity. *Nat Rev Immunol* 11, 807-22 (2011).

91. Pierangeli SS, Vega-Ostertag ME, Raschi E *et al.*, Toll-like receptor and antiphospholipid mediated thrombosis: in vivo studies. *Ann Rheum Dis* 66, 1327- 33 (2007).

92. Abdollahi-Roodsaz S, Joosten LA, Roelofs MF *et al.*, Inhibition of Toll-like receptor 4 breaks the inflammatory loop in autoimmune destructive arthritis. *Arthritis Rheum* 56, 2957-67 (2007).

93. Abdollahi-Roodsaz S, Koenders MI, Walgreen B *et al.*, Toll-like receptor 2 controls acute immune complex-driven arthritis in mice by regulating the inhibitory Fcgamma receptor IIB. *Arthritis Rheum* 65, 2583-93 (2013).

94. Kim HS, Chung DH, TLR4-mediated IL-12 production enhances IFN-gamma and IL-1beta production, which inhibits TGF-beta production and promotes antibody-induced joint inflammation. *Arthritis Res Ther* 14, R210 (2012).

95. Kaur D, Gomez E, Doe C *et al.*, IL-33 drives airway hyper-responsiveness through IL-13 mediated mast cell: airway smooth muscle crosstalk. *Allergy*, (2015).

96. Nagarkar DR, Ramirez-Carrozzi V, Choy DF *et al.*, IL-13 mediates IL-33-dependent mast cell and type 2 innate lymphoid cell effects on bronchial epithelial cells. *J Allergy Clin Immunol*, (2015).

97. Sjoberg LC, Gregory JA, Dahlen SE, Nilsson GP, Adner M, Interleukin-33 exacerbates allergic bronchoconstriction in the mice via activation of mast cells. *Allergy*, (2015).

98. Christianson CA, Goplen NP, Zafar I *et al.*, Persistence of asthma requires multiple feedback circuits involving type 2 innate lymphoid cells and IL-33. *J Allergy Clin Immunol*, (2015).

99. Van Dyken SJ, Locksley RM, Interleukin-4- and interleukin-13-mediated alternatively activated macrophages: roles in homeostasis and disease. *Annu Rev Immunol* 31, 317-43 (2013).

100. Egawa M, Mukai K, Yoshikawa S *et al.*, Inflammatory monocytes recruited to allergic skin acquire an anti-inflammatory M2 phenotype via basophil-derived interleukin-4. *Immunity* 38, 570-80 (2013).

101. Barbour M, Allan D, Xu H *et al.*, IL-33 attenuates the development of experimental autoimmune uveitis. *Eur J Immunol* 44, 3320-9 (2014).

102. Milovanovic M, Volarevic V, Ljujic B *et al.*, Deletion of IL-33R (ST2) abrogates resistance to EAE in BALB/C mice by enhancing polarization of APC to inflammatory phenotype. *PLoS One* 7, e45225 (2012).

103. Jiang HR, Milovanovic M, Allan D *et al.*, IL-33 attenuates EAE by suppressing IL-17 and IFN-gamma production and inducing alternatively activated macrophages. *Eur J Immunol* 42, 1804-14 (2012).

104. Zaiss MM, Kurowska-Stolarska M, Bohm C *et al.*, IL-33 shifts the balance from osteoclast to alternatively activated macrophage differentiation and protects from TNF-alpha-mediated bone loss. *J Immunol* 186, 6097-105 (2011).

105. Xu D, Jiang HR, Kewin P *et al.*, IL-33 exacerbates antigen-induced arthritis by activating mast cells. *Proc Natl Acad Sci U S A* 105, 10913-8 (2008).

106. Palmer G, Talabot-Ayer D, Lamacchia C *et al.*, Inhibition of interleukin-33 signaling attenuates the severity of experimental arthritis. *Arthritis Rheum* 60, 738-49 (2009).

107. Leung BP, Xu D, Culshaw S, McInnes IB, Liew FY, A novel therapy of murine collagen-induced arthritis with soluble T1/ST2. *J Immunol* 173, 145-50 (2004).

108. Xu D, Jiang HR, Li Y *et al.*, IL-33 exacerbates autoantibody-induced arthritis. *J Immunol* 184, 2620-6 (2010).

109. Martin P, Talabot-Ayer D, Seemayer CA *et al.*, Disease severity in K/BxN serum transfer-induced arthritis is not affected by IL-33 deficiency. *Arthritis Res Ther* 15, R13 (2013).

110. Anthony RM, Kobayashi T, Wermeling F, Ravetch JV, Intravenous gammaglobulin suppresses inflammation through a novel T(H)2 pathway. *Nature* 475, 110-3 (2011).

111. Tjon AS, van Gent R, Jaadar H *et al.*, Intravenous immunoglobulin treatment in humans suppresses dendritic cell function via stimulation of IL-4 and IL-13 production. *J Immunol* 192, 5625-34 (2014).

112. Kambayashi T, Laufer TM, Atypical MHC class II-expressing antigen-presenting cells: can anything replace a dendritic cell? *Nat Rev Immunol* 14, 719-30 (2014).

113. Kashiwakura J, Yokoi H, Saito H, Okayama Y, T cell proliferation by direct cross-talk between OX40 ligand on human mast cells and OX40 on human T cells: comparison of gene expression profiles between human tonsillar and lung-cultured mast cells. *J Immunol* 173, 5247-57 (2004).

114. Malaviya R, Twesten NJ, Ross EA, Abraham SN, Pfeifer JD, Mast cells process bacterial Ags through a phagocytic route for class I MHC presentation to T cells. *J Immunol* 156, 1490-6 (1996).

115. Kambayashi T, Allenspach EJ, Chang JT *et al.*, Inducible MHC class II expression by mast cells supports effector and regulatory T cell activation. *J Immunol* 182, 4686-95 (2009).

116. Banchereau J, Steinman RM, Dendritic cells and the control of immunity. *Nature* 392, 245-52 (1998).

117. Gaudenzio N, Espagnolle N, Mars LT *et al.*, Cell-cell cooperation at the T helper cell/mast cell immunological synapse. *Blood* 114, 4979-88 (2009).

118. Gong J, Yang NS, Croft M *et al.*, The antigen presentation function of bone marrow-derived mast cells is spatiotemporally restricted to a subset expressing high levels of cell surface FcepsilonRI and MHC II. *BMC Immunol* 11, 34 (2010).

119. Kambayashi T, Baranski JD, Baker RG *et al.*, Indirect involvement of allergen-captured mast cells in antigen presentation. *Blood* 111, 1489-96 (2008).

120. Suto H, Nakae S, Kakurai M *et al.*, Mast cellassociated TNF promotes dendritic cell migration. *J Immunol* 176, 4102-12 (2006).

121. Nakae S, Suto H, Kakurai M *et al.*, Mast cells enhance T cell activation: Importance of mast cellderived TNF. *Proc Natl Acad Sci U S A* 102, 6467-72 (2005).

122. Kunder CA, St John AL, Li G *et al.*, Mast cellderived particles deliver peripheral signals to remote lymph nodes. *J Exp Med* 206, 2455-67 (2009).

123. McLachlan JB, Hart JP, Pizzo SV *et al.*, Mast cell-derived tumor necrosis factor induces hypertrophy of draining lymph nodes during infection. *Nat Immunol* 4, 1199-205 (2003).

124. de Vries VC, Pino-Lagos K, Nowak EC *et al.*, Mast cells condition dendritic cells to mediate allograft tolerance. *Immunity* 35, 550-61 (2011).

125. Maurer M, Lopez Kostka S, Siebenhaar F *et al.*, Skin mast cells control T cell-dependent host defense in Leishmania major infections. *FASEB J* 20, 2460-7 (2006).

126. Reuter S, Dehzad N, Martin H *et al.*, Mast cells induce migration of dendritic cells in a murine model of acute allergic airway disease. *Int Arch Allergy Immunol* 151, 214-22 (2010).

127. Guermonprez P, Helft J, Claser C *et al.*, Inflammatory Flt3l is essential to mobilize dendritic cells and for T cell responses during Plasmodium infection. *Nat Med* 19, 730-8 (2013).

128. Shelburne CP, Nakano H, St John AL *et al.*, Mast cells augment adaptive immunity by orchestrating dendritic cell trafficking through infected tissues. *Cell Host Microbe* 6, 331-42 (2009).

129. Otsuka A, Kubo M, Honda T *et al.*, Requirement of interaction between mast cells and skin dendritic cells to establish contact hypersensitivity. *PLoS One* 6, e25538 (2011).

130. Jawdat DM, Rowden G, Marshall JS, Mast cells have a pivotal role in TNF-independent lymph node hypertrophy and the mobilization of Langerhans cells in response to bacterial peptidoglycan. *J Immunol* 177, 1755-62 (2006).

131. Nowak EC, de Vries VC, Wasiuk A *et al.*, Tryptophan hydroxylase-1 regulates immune tolerance and inflammation. *J Exp Med* 209, 2127-35 (2012).

132. Lu LF, Lind EF, Gondek DC *et al.*, Mast cells are essential intermediaries in regulatory T-cell tolerance. *Nature* 442, 997-1002 (2006).

133. Chacon-Salinas R, Limon-Flores AY, Chavez-Blanco AD, Gonzalez-Estrada A, Ullrich SE, Mast cellderived IL-10 suppresses germinal center formation by affecting T follicular helper cell function. *J Immunol* 186, 25-31 (2011).

134. Gan PY, Summers SA, Ooi JD *et al.*, Mast cells contribute to peripheral tolerance and attenuate autoimmune vasculitis. *J Am Soc Nephrol* 23, 1955- 66 (2012).

135. Chan CY, St John AL, Abraham SN, Mast cell interleukin-10 drives localized tolerance in chronic bladder infection. *Immunity* 38, 349-59 (2013).

136. Depinay N, Hacini F, Beghdadi W, Peronet R, Mecheri S, Mast cell-dependent down-regulation of antigen-specific immune responses by mosquito bites. *J Immunol* 176, 4141-6 (2006).

137. Gri G, Piconese S, Frossi B *et al.*, CD4+CD25+ regulatory T cells suppress mast cell degranulation and allergic responses through OX40-OX40L interaction. *Immunity* 29, 771-81 (2008).

138. Kashyap M, Thornton AM, Norton SK *et al.*, Cutting edge: CD4 T cell-mast cell interactions alter IgE receptor expression and signaling. *J Immunol* 180, 2039-43 (2008).

139. Frossi B, D'Inca F, Crivellato E *et al.*, Single-cell dynamics of mast cell-CD4+ CD25+ regulatory T cell interactions. *Eur J Immunol* 41, 1872-82 (2011).

140. Su W, Fan H, Chen M *et al.*, Induced CD4+ forkhead box protein-positive T cells inhibit mast cell function and established contact hypersensitivity through TGF-beta1. *J Allergy Clin Immunol* 130, 444- 52 e7 (2012).

141. Ganeshan K, Bryce PJ, Regulatory T cells enhance mast cell production of IL-6 via surfacebound TGF-beta. *J Immunol* 188, 594-603 (2012).

142. Gregory GD, Robbie-Ryan M, Secor VH, Sabatino JJ, Jr., Brown MA, Mast cells are required for optimal autoreactive T cell responses in a murine model of multiple sclerosis. *Eur J Immunol* 35, 3478- 86 (2005).

143. Schubert N, Dudeck J, Liu P *et al.*, Mast cells promote T cell driven antigen-induced arthritis despite being dispensable in T cell bypassing antibody-induced arthritis. *Arthritis Rheumatol*, (2014).

144. Forward NA, Furlong SJ, Yang Y, Lin TJ, Hoskin DW, Mast cells down-regulate CD4+CD25+ T regulatory cell suppressor function via histamine H1 receptor interaction. *J Immunol* 183, 3014-22 (2009).

145. Piconese S, Gri G, Tripodo C *et al.*, Mast cells counteract regulatory T-cell suppression through interleukin-6 and OX40/OX40L axis toward Th17-cell differentiation. *Blood* 114, 2639-48 (2009).

146. Nakae S, Saijo S, Horai R *et al.*, IL-17 production from activated T cells is required for the spontaneous development of destructive arthritis in mice deficient in IL-1 receptor antagonist. *Proc Natl Acad Sci U S A* 100, 5986-90 (2003).

147. Nakae S, Nambu A, Sudo K, Iwakura Y, Suppression of immune induction of collageninduced arthritis in IL-17-deficient mice. *J Immunol* 171, 6173-7 (2003).

148. Nakae S, Komiyama Y, Nambu A *et al.*, Antigen-specific T cell sensitization is impaired in IL-17-deficient mice, causing suppression of allergic cellular and humoral responses. *Immunity* 17, 375- 87 (2002).

149. Batten M, Li J, Yi S *et al.*, Interleukin 27 limits autoimmune encephalomyelitis by suppressing the development of interleukin 17-producing T cells. *Nat Immunol* 7, 929-36 (2006).

150. Molet S, Hamid Q, Davoine F *et al.*, IL-17 is increased in asthmatic airways and induces human

bronchial fibroblasts to produce cytokines. *J Allergy Clin Immunol* 108, 430-8 (2001).

151. Wilson RH, Whitehead GS, Nakano H *et al.*, Allergic sensitization through the airway primes Th17-dependent neutrophilia and airway hyperresponsiveness. *Am J Respir Crit Care Med* 180, 720-30 (2009).

152. Cosmi L, Liotta F, Maggi E, Romagnani S, Annunziato F, Th17 cells: new players in asthma pathogenesis. *Allergy* 66, 989-98 (2011).

153. Patel DD, Lee DM, Kolbinger F, Antoni C, Effect of IL-17A blockade with secukinumab in autoimmune diseases. *Ann Rheum Dis* 72 Suppl 2, ii116-23 (2013).

154. Kodera M, Hasegawa M, Komura K *et al.*, Serum pulmonary and activation-regulated chemokine/CCL18 levels in patients with systemic sclerosis: a sensitive indicator of active pulmonary fibrosis. *Arthritis Rheum* 52, 2889-96 (2005).

155. Xanthou G, Polihronis M, Tzioufas AG *et al.*, "Lymphoid" chemokine messenger RNA expression by epithelial cells in the chronic inflammatory lesion of the salivary glands of Sjogren's syndrome patients: possible participation in lymphoid structure formation. *Arthritis Rheum* 44, 408-18 (2001).

156. Schutyser E, Richmond A, Van Damme J, Involvement of CC chemokine ligand 18 (CCL18) in normal and pathological processes. *J Leukoc Biol* 78, 14-26 (2005).

157. Adema GJ, Hartgers F, Verstraten R *et al.*, A dendritic-cell-derived C-C chemokine that preferentially attracts naive T cells. *Nature* 387, 713- 7 (1997).

158. Gunther C, Bello-Fernandez C, Kopp T *et al.*, CCL18 is expressed in atopic dermatitis and mediates skin homing of human memory T cells. *J Immunol* 174, 1723-8 (2005).

159. Islam SA, Ling MF, Leung J, Shreffler WG, Luster AD, Identification of human CCR8 as a CCL18 receptor. *J Exp Med* 210, 1889-98 (2013).

160. de Nadai P, Charbonnier AS, Chenivesse C *et al.*, Involvement of CCL18 in allergic asthma. *J Immunol* 176, 6286-93 (2006).

161. Zlotnik A, Yoshie O, Chemokines: a new classification system and their role in immunity. *Immunity* 12, 121-7 (2000).

162. Tsai S, Santamaria P, MHC Class II Polymorphisms, Autoreactive T-Cells, and Autoimmunity. *Front Immunol* 4, 321 (2013).

163. Irigoyen P, Lee AT, Wener MH *et al.*, Regulation of anti-cyclic citrullinated peptide antibodies in rheumatoid arthritis: contrasting effects of HLA-DR3 and the shared epitope alleles. *Arthritis Rheum* 52, 3813-8 (2005).

164. van Heemst J, Jansen DT, Polydorides S *et al.*, Crossreactivity to vinculin and microbes provides a molecular basis for HLA-based protection against rheumatoid arthritis. *Nat Commun* 6, 6681 (2015).

165. Kontakioti E, Domvri K, Papakosta D, Daniilidis M, HLA and asthma phenotypes/endotypes: a review. *Hum Immunol* 75, 930-9 (2014).

166. McCormack WJ, Parker AE, O'Neill LA, Toll-like receptors and NOD-like receptors in rheumatic diseases. *Arthritis Res Ther* 11, 243 (2009).

167. Huang Q, Ma Y, Adebayo A, Pope RM, Increased macrophage activation mediated through toll-like receptors in rheumatoid arthritis. *Arthritis Rheum* 56, 2192-201 (2007).

168. English WR, Velasco G, Stracke JO, Knauper V, Murphy G, Catalytic activities of membrane-type 6 matrix metalloproteinase (MMP25). *FEBS Lett* 491, 137-42 (2001).

169. Chang X, Zhao Y, Wang Y, Chen Y, Yan X, Screening citrullinated proteins in synovial tissues of rheumatoid arthritis using 2-dimensional western blotting. *J Rheumatol* 40, 219-27 (2013).

170. Wegner N, Lundberg K, Kinloch A *et al.*, Autoimmunity to specific citrullinated proteins gives the first clues to the etiology of rheumatoid arthritis. *Immunol Rev* 233, 34-54 (2010).

171. Wang KX, Denhardt DT, Osteopontin: role in immune regulation and stress responses. *Cytokine Growth Factor Rev* 19, 333-45 (2008).

172. Magarinos NJ, Bryant KJ, Fosang AJ *et al.*, Mast cell-restricted, tetramer-forming tryptases induce

aggrecanolysis in articular cartilage by activating matrix metalloproteinase-3 and -13 zymogens. *J Immunol* 191, 1404-12 (2013).

173. Van Wart HE, Birkedal-Hansen H, The cysteine switch: a principle of regulation of metalloproteinase activity with potential applicability to the entire matrix metalloproteinase gene family. *Proc Natl Acad Sci U S A* 87, 5578-82 (1990).

174. Zenmyo M, Hiraoka K, Komiya S, Morimatsu M, Sasaguri Y, Histamine-stimulated production of matrix metalloproteinase 1 by human rheumatoid synovial fibroblasts is mediated by histamine H1 receptors. *Virchows Arch* 427, 437-44 (1995).

175. Atamas SP, Luzina IG, Choi J *et al.*, Pulmonary and activation-regulated chemokine stimulates collagen production in lung fibroblasts. *Am J Respir Cell Mol Biol* 29, 743-9 (2003).

176. Bulfone-Paus S, Paus R, Osteopontin as a new player in mast cell biology. *Eur J Immunol* 38, 338-41 (2008).

177. Denhardt DT, Noda M, O'Regan AW, Pavlin D, Berman JS, Osteopontin as a means to cope with environmental insults: regulation of inflammation, tissue remodeling, and cell survival. *J Clin Invest* 107, 1055-61 (2001).

178. Mori R, Shaw TJ, Martin P, Molecular mechanisms linking wound inflammation and fibrosis: knockdown of osteopontin leads to rapid repair and reduced scarring. *J Exp Med* 205, 43-51 (2008).

179. Berman JS, Serlin D, Li X *et al.*, Altered bleomycin-induced lung fibrosis in osteopontindeficient mice. *Am J Physiol Lung Cell Mol Physiol* 286, L1311-8 (2004).

180. Samitas K, Zervas E, Vittorakis S *et al.*, Osteopontin expression and relation to disease severity in human asthma. *Eur Respir J* 37, 331-41 (2011).

181. Xanthou G, Alissafi T, Semitekolou M *et al.*, Osteopontin has a crucial role in allergic airway disease through regulation of dendritic cell subsets. *Nat Med* 13, 570-8 (2007).

182. Simoes DC, Xanthou G, Petrochilou K *et al.*, Osteopontin deficiency protects against airway remodeling and hyperresponsiveness in chronic asthma. *Am J Respir Crit Care Med* 179, 894-902 (2009).

183. Hon KL, Ching GK, Ng PC, Leung TF, Exploring CCL18, eczema severity and atopy. *Pediatr Allergy Immunol* 22, 704-7 (2011).

184. Kollert F, Binder M, Probst C *et al.*, CCL18 - potential biomarker of fibroinflammatory activity in chronic periaortitis. *J Rheumatol* 39, 1407-12 (2012).

185. Kim HB, Kim CK, Iijima K, Kobayashi T, Kita H, Protein microarray analysis in patients with asthma: elevation of the chemokine PARC/CCL18 in sputum. *Chest* 135, 295-302 (2009).

186. Prasse A, Pechkovsky DV, Toews GB *et al.*, CCL18 as an indicator of pulmonary fibrotic activity in idiopathic interstitial pneumonias and systemic sclerosis. *Arthritis Rheum* 56, 1685-93 (2007).

187. Yang IV, Tomfohr J, Singh J et al., The clinical and environmental determinants of airway transcriptional profiles in allergic asthma. *Am J Respir Crit Care Med* 185, 620-7 (2012).

188. Gavala ML, Kelly EA, Esnault S *et al.*, Segmental allergen challenge enhances chitinase activity and levels of CCL18 in mild atopic asthma. *Clin Exp Allergy* 43, 187-97 (2013).

189. Blumenthal MN, Zhong W, Miller M *et al.*, Serum metalloproteinase leukolysin (MMP-25/MT-6): a potential metabolic marker for atopyassociated inflammation. *Clin Exp Allergy* 40, 859-66 (2010).

190. Peterson S, Poposki JA, Nagarkar DR *et al.*, Increased expression of CC chemokine ligand 18 in patients with chronic rhinosinusitis with nasal polyps. *J Allergy Clin Immunol* 129, 119-27 e1-9 (2012).

191. Perez DA, Vago JP, Athayde RM *et al.*, Switching off key signaling survival molecules to switch on the resolution of inflammation. *Mediators Inflamm* 2014, 829851 (2014).

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192. Buckley CD, Gilroy DW, Serhan CN, Stockinger B, Tak PP, The resolution of inflammation. *Nat Rev Immunol* 13, 59-66 (2013).

193. Serhan CN, Dalli J, Karamnov S *et al.*, Macrophage proresolving mediator maresin 1 stimulates tissue regeneration and controls pain. *FASEB J* 26, 1755-65 (2012).

194. Lebbink RJ, de Ruiter T, Verbrugge A, Bril WS, Meyaard L, The mouse homologue of the leukocyteassociated Ig-like receptor-1 is an inhibitory receptor that recruits Src homology region 2-containing protein tyrosine phosphatase (SHP)-2, but not SHP-1. *J Immunol* 172, 5535-43 (2004).

195. Steevels TA, Lebbink RJ, Westerlaken GH, Coffer PJ, Meyaard L, Signal inhibitory receptor on leukocytes-1 is a novel functional inhibitory immune receptor expressed on human phagocytes. *J Immunol* 184, 4741-8 (2010).

196. Masuda A, Nakamura A, Maeda T, Sakamoto Y, Takai T, Cis binding between inhibitory receptors and MHC class I can regulate mast cell activation. *J Exp Med* 204, 907-20 (2007).

197. Hitomi K, Tahara-Hanaoka S, Someya S *et al.*, An immunoglobulin-like receptor, Allergin-1, inhibits immunoglobulin E-mediated immediate hypersensitivity reactions. *Nat Immunol* 11, 601-7 (2010).

198. Nakahashi-Oda C, Tahara-Hanaoka S, Shoji M *et al.*, Apoptotic cells suppress mast cell inflammatory responses via the CD300a immunoreceptor. *J Exp Med* 209, 1493-503 (2012).

199. Yachie A, Niida Y, Wada T *et al.*, Oxidative stress causes enhanced endothelial cell injury in human heme oxygenase-1 deficiency. *J Clin Invest* 103, 129-35 (1999).

200. Tzima S, Victoratos P, Kranidioti K, Alexiou M, Kollias G, Myeloid heme oxygenase-1 regulates innate immunity and autoimmunity by modulating IFN-beta production. *J Exp Med* 206, 1167-79 (2009).

201. Lee TS, Chau LY, Heme oxygenase-1 mediates the anti-inflammatory effect of interleukin-10 in mice. *Nat Med* 8, 240-6 (2002).

202. Takamiya R, Murakami M, Kajimura M *et al.*, Stabilization of mast cells by heme oxygenase-1: an anti-inflammatory role. *Am J Physiol Heart Circ Physiol* 283, H861-70 (2002).

203. Ma YY, Yang MQ, Wang CF, Ding J, Li JY, Inhibiting mast cell degranulation by HO-1 affects dendritic cell maturation in vitro. *Inflamm Res* 63, 527-37 (2014).

204. Takayasu A, Miyabe Y, Yokoyama W *et al.*, CCL18 activates fibroblast-like synoviocytes in patients with rheumatoid arthritis. *J Rheumatol* 40, 1026-8 (2013).

205. Iwamoto T, Okamoto H, Toyama Y, Momohara S, Molecular aspects of rheumatoid arthritis: chemokines in the joints of patients. *FEBS J* 275, 4448-55 (2008).

206. van Lieshout AW, Fransen J, Flendrie M *et al.*, Circulating levels of the chemokine CCL18 but not CXCL16 are elevated and correlate with disease activity in rheumatoid arthritis. *Ann Rheum Dis* 66, 1334-8 (2007).

207. Momohara S, Okamoto H, Iwamoto T *et al.*, High CCL18/PARC expression in articular cartilage and synovial tissue of patients with rheumatoid arthritis. *J Rheumatol* 34, 266-71 (2007).

208. Zhang F, Luo W, Li Y, Gao S, Lei G, Role of osteopontin in rheumatoid arthritis. *Rheumatol Int* 35, 589-95 (2015).

209. Gazal S, Sacre K, Allanore Y *et al.*, Identification of secreted phosphoprotein 1 gene as a new rheumatoid arthritis susceptibility gene. *Ann Rheum Dis* 74, e19 (2015).

210. Son M, Santiago-Schwarz F, Al-Abed Y, Diamond B, C1q limits dendritic cell differentiation and activation by engaging LAIR-1. *Proc Natl Acad Sci U S A* 109, E3160-7 (2012).

211. Son M, Diamond B, C1q-Mediated Repression of Human Monocytes Is Regulated by Leukocyte-Associated Ig-Like Receptor 1 (LAIR-1). *Mol Med* 20, 559-68 (2015).

212. Florian S, Sonneck K, Czerny M *et al.*, Detection of novel leukocyte differentiation antigens on basophils and mast cells by HLDA8 antibodies. *Allergy* 61, 1054-62 (2006).

213. Henningsson F, Wolters P, Chapman HA, Caughey GH, Pejler G, Mast cell cathepsins C and S control levels of carboxypeptidase A and the chymase, mouse mast cell protease 5. *Biol Chem* 384, 1527-31 (2003).

214. Maxwell L, Singh JA, Abatacept for rheumatoid arthritis. *Cochrane Database Syst Rev*, CD007277 (2009).

215. Emery P, Burmester GR, Bykerk VP *et al.*, Evaluating drug-free remission with abatacept in early rheumatoid arthritis: results from the phase 3b, multicentre, randomised, active-controlled AVERT study of 24 months, with a 12-month, double-blind treatment period. *Ann Rheum Dis* 74, 19-26 (2015).

216. Gottenberg JE, Ravaud P, Cantagrel A *et al.*, Positivity for anti-cyclic citrullinated peptide is associated with a better response to abatacept: data from the 'Orencia and Rheumatoid Arthritis' registry. *Ann Rheum Dis* 71, 1815-9 (2012).

217. Pieper J, Herrath J, Raghavan S *et al.*, CTLA4-Ig (abatacept) therapy modulates T cell effector functions in autoantibody-positive rheumatoid arthritis patients. *BMC Immunol* 14, 34 (2013).

218. Riella LV, Sayegh MH, T-cell co-stimulatory blockade in transplantation: two steps forward one step back! *Expert Opin Biol Ther* 13, 1557-68 (2013).

219. Bouguermouh S, Fortin G, Baba N, Rubio M, Sarfati M, CD28 co-stimulation down regulates Th17 development. *PLoS One* 4, e5087 (2009).