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## The role of p53.S389 phosphorylation in DNA damage response pathways and tumorigenesis

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# Chapter 5

## **Delayed expression of apoptotic and cell cycle control genes in carcinogen-exposed bladders of mice lacking p53.S389 phosphorylation**

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## Abstract

Mice with non-phosphorylated Ser389 in p53 are susceptible for bladder tumors induced by 2-AAF. Since p53 is a transcription factor, this might well be preceded by differences in the regulation of gene expression. Microarray analysis was used to determine early transcriptional changes that might underlie this cancer-prone phenotype. Interestingly, lack of Ser389 phosphorylation led to endogenously different gene expression levels. The number of genes affected was, however, rather small. Conversely, after short-term exposure to 2-AAF, wild-type and p53.S389A bladders demonstrated a significant number of differentially-expressed genes. Differences between wild-type and p53.S389A could mainly be attributed to a delayed, rather than complete absence of, transcriptional response of a group of genes, including well-known p53 target genes involved in apoptosis and cell cycle control like *Bax*, *Perp* and *P21*. An analysis of differentially-expressed genes in non-tumorigenic tissue and bladder tumors of p53.S389A after long-term exposure to 2-AAF revealed 319 genes. Comparison of these with those found after short-term exposure resulted in 23 transcripts. These possible marker genes might be useful for the early prediction of bladder tumor development. In conclusion, our data indicate that, lack of Ser389 phosphorylation results in aberrant expression of genes needed to execute vital responses to DNA damage. Post-translational modifications, like Ser389 phosphorylation, seem crucial for fine-tuning the transcription of a specific set of genes and do not appear to give rise to major changes in transcription patterns. As such, Ser389 phosphorylation is needed for some, but certainly not all, p53 functions.

## Introduction

When chemical or physical compounds damage DNA, several defense mechanisms are activated to prevent the DNA from harmful structural alterations that may ultimately lead to cancer. All of these defense mechanisms are aimed at eliminating DNA damage. Examples are cell cycle arrest, DNA repair and apoptotic processes. The latently present p53 protein becomes stabilized and activated in response to DNA damage. Once activated, p53 acts as a tumor suppressor through transcriptional activation of a large variety of target genes [1-3]. Activation of p53 occurs exclusively at the post-translational level through a broad range of modifications; namely (de)phosphorylation, acetylation, ubiquitination, sumoylation, glycosylation, methylation and neddylation. Of these processes, phosphorylation appears to be the most frequent modification of the protein [4-6].

Phosphorylation of the p53 N-terminal region, in particular Ser15 and Ser20, disrupts the interaction with the MDM2 protein, a ubiquitin ligase that inactivates p53 by targeting the protein for proteasomal degradation [7;8]. Ser315 and Ser392 were the first phosphorylation sites identified at the C-terminal domain [9]. Ser315 is phosphorylated in response to both ionizing and UV radiation, whereas Ser392 is only phosphorylated specifically after UV radiation [4;10-12]. Kinases targeting this Ser392 site *in vitro* are Casein Kinase II (CKII) [13;14], the double-stranded, RNA-activated protein kinase (PKR) [15], p38 Map kinase [16;17] and the recently identified cyclin-dependent kinase 9 (cdk9) through direct physical interaction [18;19]. However, it is still not clear how certain types of DNA damage result in phosphorylation of Ser392, and what the ultimate effect on specific cellular defense systems is *in vivo*.

To investigate the function of the Ser392 phosphorylation site (equivalent of mouse Ser389), we generated mice with a single point mutation in the *p53* gene that resulted in a substitution of a serine to an alanine; the p53.S389A mouse model [20]. Phosphorylation of Ser389 is not required

for p53 functioning in either spontaneous or ionizing radiation-induced lymphoma tumor development, since tumor responses were highly comparable in p53.S389A and wild-type mice [20;21]. However, p53.S389A mice appeared to be more susceptible for 2-acetylaminofluorene (2-AAF)-induced urinary bladder tumors and UV-B induced skin tumors [20;21]. This is in line with our *in vitro* findings where cells of these mice demonstrated an impaired DNA-binding capacity and a reduced apoptotic response after UV radiation [20]. We therefore hypothesized that phosphorylation of p53 at codon 389 is substrate dependent, and that lack of Ser389 phosphorylation only has an adverse effect on the functioning of p53 as a tumor suppressor in the case of specific substrates.

In the current study, we further explored which *in vivo* molecular responses are affected when phosphorylation of p53 at codon 389 is impossible. For this, mice were exposed to 2-AAF and gene-expression profiles were analyzed in urinary bladder tissue (further referred to as bladder) of male mice. This was done because the increased incidence of tumors in p53.S389A mice was observed in this specific tissue, and was furthermore restricted to male mice only [21]. Given the well-known function of p53 as a transcription factor, we hypothesized that this increased incidence of bladder tumors is likely to be preceded early in the exposure period by a different gene-expression profile in bladders of p53.S389A mice compared to wild-type mice not susceptible to exposure to 2-AAF. To investigate this, we performed a microarray analysis on wild-type and p53.S389A male bladders exposed for 1 or 2 weeks (short-term) to 2-AAF. Interestingly, a significant number of genes are differentially-expressed in bladders of p53.S389A compared to wild-type mice, including known p53 target genes involved in apoptotic and cell cycle processes. Finally, we also analyzed gene expression patterns in tumorigenic bladders of p53.S389A mice after long-term exposure to 2-AAF, and compared these to the differentially-expressed genes found after short-term exposure to 2-AAF. Several overlapping genes could be identified. Possibly these genes are useful as early markers for bladder cancer development.

## Materials and Methods

### *Wild-type and p53.S389A mice exposed to 2-AAF*

Mice were exposed to 300 ppm 2-AAF mixed in feed as used in previous studies [21] for 1 or 2 weeks (short-term) or 39 weeks (long-term) (n=4). The 2-AAF containing feed (Altromin, Lage, Germany), normal feed and water were available ad libitum throughout the course of the studies. All mice used were males of 6-9 weeks of age and at least ten times backcrossed in C57BL/6. Furthermore control mice received normal diet.

### *RNA isolation*

Total RNA was isolated from mouse bladder using the RNeasy Mini kit (Qiagen, Valencia, USA), followed by a DNase treatment with RNase-Free DNase Set (Qiagen Valencia, CA, USA). RNA quality was assessed with the Bioanalyzer 2100 (Agilent Technologies, Palo Alto, USA) All assays were performed according to the manufacturer's protocols.

### *Microarray analysis*

The mouse oligonucleotide libraries (Cat # MOULIBST & Cat # MOULIB384B) were obtained from Sigma-Compugen Incorporated. Technical support was supplied by LabOnWeb ([http://www.labonweb.com/cgi-bin/chips/full\\_loader.cgi](http://www.labonweb.com/cgi-bin/chips/full_loader.cgi)). The libraries represent in total 21,766 LEADS™ clusters plus 231 controls. The oligonucleotide library was printed with a Lucidea

Spotter (Amersham Pharmacia Biosciences, Piscataway, USA) on commercial UltraGAPS slides (amino-silane-coated slides, Corning 40017) and processed according to the manufacturer's instructions. The slides contained 65-mer oligonucleotides and the batch was checked for the quality of spotting by hybridizing with SpotCheck Cy3 labeled nonamers (Genetix, New Milton Hampshire, UK).

1.5 µg total RNA samples were used in randomized batches, according to a common reference design, with a mix of all samples as common reference. The RNA was amplified using the Amino Allyl MessageAmp aRNA kit (Ambion, Austin, USA), and labeled with Cy3 (experimental samples) and Cy5 (common reference) reactive dye according to the manufacturer's instructions. The microarrays were hybridized overnight with 200 µl hybridization mixture, consisting of 50 µl Cy3- and Cy5-labeled aRNA (in a 1 to 3 molar ratio), 100 µl Formamide and 50 µl 4 x RPK0325 MicroArray Hybridization Buffer (Amersham Pharmacia Biosciences, Piscataway, USA) at 37°C, washed in an Automated Slide Processor (Amersham Pharmacia Biosciences, Piscataway, USA), and subsequently scanned (Agilent DNA MicroArray Scanner, Agilent Technologies, Palo Alto, USA).

#### *Data analysis and statistics*

Microarray spot intensities were quantified as artifact removed densities, using Array Vision software (version 6.0). Further processing of the data was performed using R (version 2.2.1) and the Bioconductor MAANOVA package (version 0.98.8). All slides were subjected to a set of quality control checks, i.e., visual inspection of the scans, examining the consistency among the replicated samples by principal components analysis (PCA), testing against criteria for signal to noise ratios, testing for consistent performance of the labeling dyes, pen grid plots to check consistent pen performance, and visual inspection of pre- and post-normalized data with box plots and RI plots.

After log<sub>2</sub> transformation, the data were normalized by a spatial lowess smoothing procedure, and analyzed using a two-stage mixed-model ANOVA model [22;23]. First, array, dye and array-by-dye effects were modeled globally. Subsequently, the residuals from this first model are fed into the gene-specific model to fit treatment, and spot effects on a gene-by-gene basis using a mixed-model ANOVA. These residuals are normalized expression values and throughout used in the graphs to depict (differential) gene expression. For hypothesis testing, a permutation based F1 test was used which allows relaxing the assumption that the data is normally distributed [24]. To account for multiple testing, p-values from the permutation procedure were adjusted to represent a False Discovery Rate (FDR) of 10% [25]. Four tests were performed to identify differential gene expression. 1) A test to find differences between control, 1 or 2 weeks 2-AAF exposed wild-type bladder. 2) A test to find differences between control, 1 or 2 weeks 2-AAF exposed p53.S389A bladder. 3) A test to find differences between genotypes across control, 1 or 2 weeks 2-AAF exposed bladder. 4) A test to find differences between tumorigenic and non-tumorigenic p53.S389A bladder.

The differences in gene expression between the wild-type and p53.S389A mice in their response to 2-AAF was qualitatively determined as follows. First, the normalized expression values for all genes found to be involved in 2-AAF exposure were calculated (the union of list (1) and (2)). Second, for each time point the difference in gene expression between 2-AAF exposed and control bladder was calculated; this difference represents a rate of change in expression. Third, for each time point this difference in gene expression between 2-AAF exposed and control bladder

of wild-type and p53.S389A were compared in a scatter plot. A deviation from a one-to-one relationship in these plots indicated a difference in gene expression rate between wild-type and p53.S389A in their response to 2-AAF exposure. This enabled the identification of delayed up-regulated and delayed down-regulated genes in p53.S389A bladder.

To relate the differences in gene expression to changes in functional biological processes, lists of differentially-expressed genes were analyzed for overrepresentation of specific Gene Ontology (GO) terms using Onto Express (<http://vortex.cs.wayne.edu/projects.htm>). The list of all gene products on the microarray was then used as the reference set. Relevant GO-terms were selected according to FDR-corrected p-values and the number of genes involved. The F1-statistics from list (1), (2) and (3) were used for Gene Set Enrichment Analysis (GSEA) [26]. All pathways and gene expression signatures contained in the c2 database of the by Molecular Signature Database (MsigDb 1.0; [http://www.broad.mit.edu/gsea/msigdb/msigdb\\_index.html](http://www.broad.mit.edu/gsea/msigdb/msigdb_index.html)), and an additional p53 pathway reported by Harris and Levine [27] were tested for significance using the Gene-Set-Test facility provided by the Limma package (version 2.7.3) in Bioconductor. Pathways and gene expression signatures with p-values <0.05 and with at least one gene from either list (1), (2) or (3) were reported.

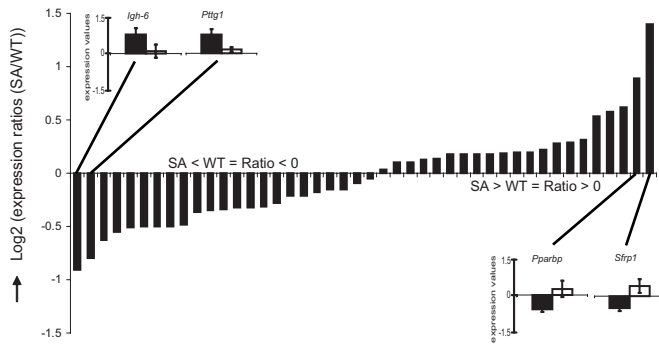
#### *Real-time PCR*

To verify the microarray results, cDNA was generated from RNA by using the high-Capacity cDNA archive kit containing random hexamer primers (Applied Biosystems). mRNA presence was measured with Taqman gene expression assays (Applied Biosystems) on a 7500 Fast Real-Time PCR System, with a two-step PCR procedure according to the manufacturer's protocol. The assays we used were *Ccnb2*; Mm00432351\_m1, *Ccng1*; Mm00438084\_m1, *Pttg1* Mm00479224\_m1 and *Pmaip1*; Mm00451763\_m1.

## **Results**

### **Differentially-expressed genes in unexposed wild-type and p53.S389A bladders**

Absence of biologically active p53 protein has a dramatic influence on expression profiles, even in unexposed cells [28;29]. To test comparable effects in cells carrying modified p53 protein, we compared gene-expression profiles between unexposed wild-type and p53.S389A bladders. A total of 44 genes with a clear statistically-significant difference between the genotypes were identified and these are presented in a bar plot in Figure 1. Twenty-three genes had a lower ( $\log_2$  (expression ratio (p53.S389A/wild-type)) <0) and 21 genes had a higher ( $\log_2$  (expression ratio (p53.S389A/wild-type)) >0) expression level in p53.S389A bladders compared to wild-type bladders. Interestingly, the two genes with the highest induced ratios were the bladder tumor-related gene *Sfrp1* and the cofactor of transcription *Pparbp* [30]. The two genes with the most reduced ratio were *Igh-6* and the presumed proto-oncogene *Pttg1* [31]. The biological categories, based on GO terms (analyzed using Onto-Express [32]), containing the most affected genes were development, regulation of transcription and transport. The complete lists of differentially-regulated genes found on the basis of genotype differences between wild-type and p53.S389A bladders together with their  $\log_2$  (expression ratios) is provided in Supplementary Table I. Lack of Ser389 phosphorylation therefore results in clear differences in gene expression levels in unexposed bladders.



**Figure 1 – Bar plot of differentially-expressed genes in wild-type and p53.S389A bladders**

Differences in expression of genes across genotypes were tested as described in Materials and Methods. A bar plot representing ratios of the log<sub>2</sub> (expression values) of p53.S389A (SA) divided by wild-type (WT) is shown. Each bar represents an individual gene; corresponding values are depicted in Supplementary Table I. The genes with the highest ratios are magnified and represented as a bar plot with the average of log<sub>2</sub> (expression values) of both WT (wild-type; black) and SA (p53.S389A; white) control samples.

### Differentially-expressed genes in wild-type and p53.S389A bladders after exposure to 2-AAF

To analyze whether differences in gene expression shortly after exposure to 2-AAF can explain differences in tumor outcome later in life, we exposed wild-type and p53.S389A mice for one and two weeks to 2-AAF and analyzed gene expression by microarrays in RNA isolated from bladder tissues.

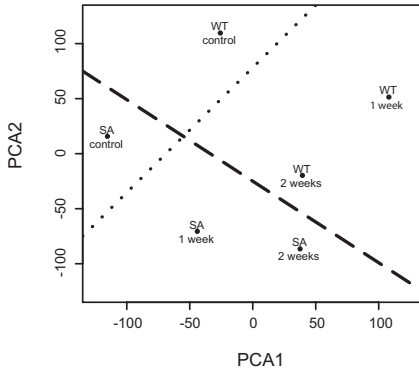
A PCA with a two-dimensional representation of the relationship between the wild-type and p53.S389A samples demonstrated the segregation between the control and samples exposed to 2-AAF for 1 or 2 weeks (Figure 2A). Importantly, a clear segregation between the genotypes was observed as well. Apparently, lack of Ser389 phosphorylation of p53 resulted in different gene expression responses *in vivo* after exposure to 2-AAF.

Differential expression of the genes found to have significantly changed over time was analyzed with the help of a one-way ANOVA analysis on the different groups of mice (control, 1 or 2 weeks 2-AAF), and the construction of Self-Organizing Maps (SOMs). Figure 2B shows the expression profiles of both genotypes as well as the number of genes per SOM. The complete lists of differentially-expressed genes found in wild-type and p53.S389A bladders, together with fold-change levels based on their corresponding controls is provided in Supplementary Tables II and III, respectively. Expression levels measured by microarray analysis were highly similar to results obtained with real-time PCR (Supplementary Figure 1).

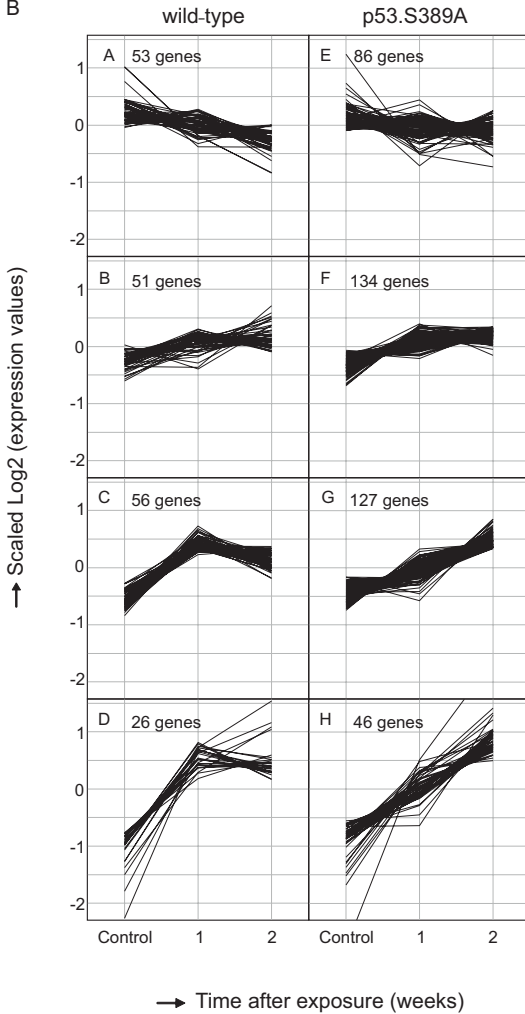
As a consequence of 2-AAF exposure, there were groups of genes in both genotypes that exhibited continuous changes in gene expression. However, induction and repression levels were rather small (panels A, B, E, and F). Furthermore, there were groups with more pronounced expression profiles (panels C, D, G, and H). With respect to differences between wild-type and p53.S389A, a group of 82 genes (panels C and D) in the wild-type demonstrated a clear increase in gene expression after 1 week and a marked decrease after 2 weeks, which was not observed in p53.S389A (panels G and H). A careful evaluation revealed that those genes peak at 1 week in wild-type, whereas in p53.S389A they only reached this peak after 2 weeks. Interestingly, this group included apoptosis-related genes such as *Pmaip1* (*Noxa*) [33], *Bax* [34] and *Birc5* (*Survivin*) [35], genes involved in damage recovery and growth promotion like *Ccng1* (*Cyclin G1*) and *Ccnb2* (*Cyclin B2*) [36], and the DNA repair gene *Mgmt*.



A



B



**Figure 2 - Presentation of differentially-expressed genes in wild-type and p53.S389A bladders after short-term exposure to 2-AAF**

A) PCA of differentially-expressed genes in wild-type (WT) and p53.S389A (SA) bladders, providing a two-dimensional representation of the relationships between wild-type and p53.S389A gene expression of control bladders and bladders after short-term exposure to 2-AAF. Each group represents 4 individual replicates. The dashed line shows segregation between wild-type and p53.S389A, and the dotted line between the control group and groups exposed short-term (1 week and 2 weeks) to 2-AAF.

B) SOM of differentially-expressed genes in bladders wild-type and p53.S389A mice after short-term exposure to 2-AAF. All differentially-expressed genes were grouped into four clusters per genotype (panel A-D for wild-type and panel E-H for p53.S389A). Each SOM cluster represents the gene expression pattern for genes within the specific cluster, with the number of genes in each cluster indicated. Log<sub>2</sub> (expression values) scaled relative to the mean are shown on the y-axis, with the corresponding time points on the x-axis.

1 = one week exposure to 2-AAF, 2 = two weeks exposure to 2-AAF

## Exploring the differentially-expressed genes in wild-type and p53.S389A bladders after exposure to 2-AAF

To gain a better understanding of the processes involved in mouse bladders after short-term exposure to 2-AAF, we analyzed all genes that were differentially-expressed in wild-type and/or p53.S389A (i.e., union, see Materials and Methods) using Onto-Express [32]. Supplementary Table IV shows the GO categories with a corrected p-value  $\leq 0.05$  and at least 3 genes present. Within the GO categories found to be significant, the most interesting were: cell cycle (arrest), DNA repair, and induction of apoptosis. These are processes known to be affected after exposure to a genotoxic agent [37]. As expected after exposure to 2-AAF, the category ‘response to DNA damage stimulus’ was also found. Furthermore, mitosis and mitotic-related processes such as DNA replication, cytokinesis and DNA metabolism are significantly present. In addition to the GO analysis, we used GSEA (recently described as a powerful analytical method for interpreting microarray data) to obtain more insight into the biological pathways involved [26]. Table I provides a complete list of pathways that were significantly affected ( $p \leq 0.005$ ). Four processes with known p53 involvement were ranked in the top ten when selecting for highest p-values in wild-type bladders. For example, the p53 signaling pathway was highly significantly altered, as well as a general p53 pathway and the p53 hypoxia pathway. Furthermore, a variety of cell cycle and apoptotic-related processes were found. In conclusion, short-term *in vivo* exposure to 2-AAF of wild-type mice resulted in altered expression of genes in bladders mostly involved in p53-related pathways, such as cell cycle regulation and apoptosis. Given this observation, it might be expected that these p53-related pathways in particular, are aberrantly regulated by exposure to 2-AAF when p53 functioning is inhibited through mutational inactivation of the Ser389 phosphorylation site. Indeed, GSEA of the differentially-regulated genes in bladders from p53.S389A mice exposed to 2-AAF resulted in an almost entirely different order of this list (Table I). Only two of the p53-related processes, p53-UP and p53 signaling, were also found to

**Table I – Biological pathways in mouse bladders after exposure to 2-AAF that were found to be differential using GSEA analysis**

Wild type			p53.S389A		
Pathway	Description	p-value*	Pathway	Description	p-value*
p53_signalling	BioCarta	<0.0001	PGC	Manually Curated	0.0004
PGC	Manually Curated	<0.0001	MAP00600_Sphingoglycolipid_metabolism	GenMAPP	0.0009
<b>p53_UP</b>	Kannan_et_al_2001	<0.0001	MYC_MUT	BLACK	0.001
GLUT_UP	Peng_at_al_2002	<0.0001	PROLIF_GENES	Manually Curated	0.0013
CBF_LEUKEMIA_DOWNING_AML	Manually Curated	<0.0001	MAP00531_Glycosaminoglycan_degradation	GenMAPP	0.0018
<b>p53Pathway</b>	BioCarta	6.00E-04	MAP00480_Glutathione_metabolism	GenMAPP	0.0019
<b>p53hypoxiaPathway</b>	BioCarta	7.00E-04	GLUT_UP	Peng_at_al_2002	0.0021
PROLIF_GENES	Manually Curated	8.00E-04	CR_CELL_CYCLE	PNAS_2004	0.0022
MAP00512_O_Glycans_biosynthesis	GenMAPP	0.0015	ST_Dictyostelium_discoideum_cAMP_Chemotaxis_Pathway	Signalling Transduction KE	0.0024
radiation_sensitivity	BioCarta	0.0016	<b>p53_UP</b>	Kannan_et_al_2001	0.0025
cell_cycle_arrest		0.0018	<b>p53_signalling</b>	BioCarta	0.0026
BRCA_UP	Welcsh_et_al_2002	0.002	MAP00052_Galactose_metabolism	GenMAPP	0.0034
cellcyclePathway		0.0021	cell_cycle_arrest		0.0036
drug_resistance_and_metabolism	BioCarta	0.0021	GO_ROS	FO	0.0036
GO_ROS	FO	0.0021	breast_cancer_estrogen_signalling	GEArray	0.0037
MAP00480_Glutathione_metabolism	GenMAPP	0.0024	AR_MOUSE_PLUS_TESTO_FROM_NETAFFX	na	0.0038
SA_PROGRAMMED_CELL_DEATH	SigmaAldrich	0.0024	CBF_LEUKEMIA_DOWNING_AML	Manually Curated	0.0038
CR_CELL_CYCLE	PNAS_2004	0.0027	freePathway		0.0043
breast_cancer_estrogen_signalling	GEArray	0.0032	g1Pathway	BioCarta	0.0049
DNA_DAMAGE_SIGNALLING	GO	0.0034	cell_proliferation		0.005
badPathway		0.0035			
FRASOR_ER_DOWN	Frasor_et_al_2004	0.0036			
g1Pathway	BioCarta	0.0038			
ceramidePathway		0.0042			
MAP00230_Purine_metabolism	GenMAPP	0.0049			

\*Pathways with a p-value < 0.005 are shown

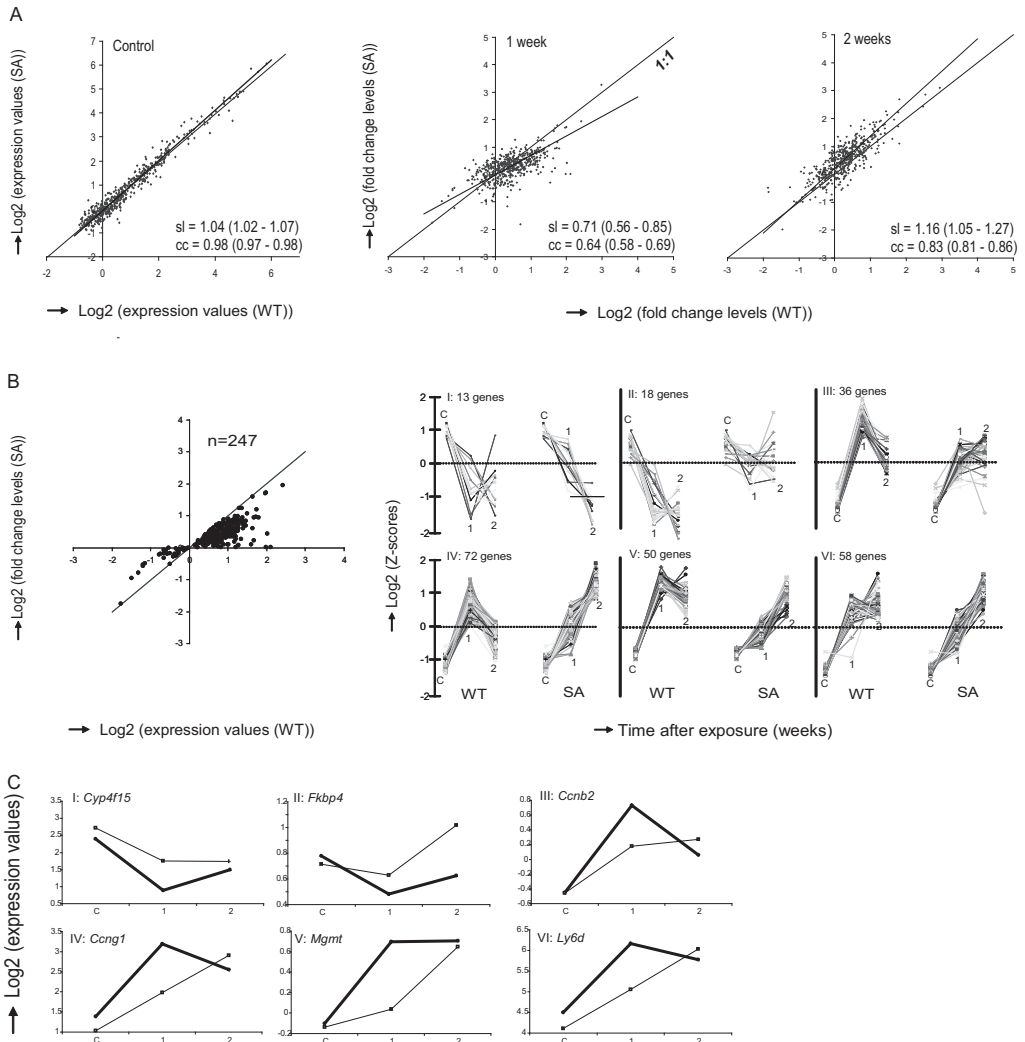
be significantly present. However for the corresponding processes, the associated p-values were considerably higher in the p53.S389A bladders than those found in wild-type mice, indicating that these p53-related pathways are activated/de-activated to a lesser extent in p53.S389A bladders exposed to 2-AAF.

### **Delayed gene expression in p53.S389A bladders exposed to 2-AAF**

To unravel the exact nature of the observed differences in gene expression between wild-type and p53.S389A mice, the union of the 2-AAF induced differential genes was tested for its distribution of genes along the log<sub>2</sub> (expression values) axis. As depicted in Figure 3A, left panel, the line chart with the log<sub>2</sub> (expression values) of genes in wild-type and p53.S389A control bladders had a slope of 1.04 and a correlation coefficient of 0.98, indicating equal log<sub>2</sub> (expression values) of the genes in unexposed bladders of mice of both genotypes (i.e., lack of significant differences). However after 1 week of exposure to 2-AAF, the distribution along the 1:1 line of these genes was divergent since the slope was only 0.71 and the correlation coefficient 0.64. This means the log<sub>2</sub> values (fold change levels) compared to their corresponding control values of genes in p53.S389A bladders were either higher or lower than those of the same genes in wild-type bladders. Intriguingly, after 2 weeks of exposure to 2-AAF, the distribution of log<sub>2</sub> (fold change levels) of the selected genes was becoming more comparable again, with a slope of 1.16 and a correlation coefficient of 0.83. These results indicated that although expression levels in unexposed samples were highly comparable, a mutation at Ser389 resulted in different kinetics of gene expression after short-term exposure to 2-AAF compared to wild-type mice. Since this difference in transcriptional response was most evident after 1 week, and seemed to decrease again after 2 weeks of exposure towards endogenous levels, responses in p53.S389A bladders appeared to be delayed compared to their wild-type counterparts.

K-means clustering was subsequently performed on a selection of these genes (n=247), in which induction or repression were delayed in p53.S389A compared to wild-type bladders after 1 week of exposure to 2-AAF (Figure 3B). Although the resulting groups of genes did not have similar response patterns, responses in p53.S389A bladders were consistently delayed compared to wild-type responses, both in down-regulation as well as in up-regulation of genes. In some groups (I, IV and V) the expression value in p53.S389A bladders after 2 weeks of exposure to 2-AAF reached more or less the same value as observed in wild-type bladders after 1 week of exposure to 2-AAF. In the remaining groups the minimum expression values in p53.S389A bladders were either higher (II), or reached a lower (III) or higher (VI) maximum than those in the wild-type. This indicated that although all 247 genes demonstrated delayed responses, the exact nature of this delay was highly dependent on the individual gene concerned. A representative gene was selected for each group (I to VI) and the expression levels of these are depicted in Figure 3C. Use of the GO categories revealed that these delayed genes were involved in transport and mitotic-related processes (cytokinesis, DNA replication, electron transport, metabolism, mitosis). In addition, categories involved in the p53-dependent processes like apoptosis and cell cycle regulation were also present (see Supplementary Table V).

Based on the above-mentioned results, we investigated the behavior of the known p53-related pathways of cell cycle arrest and apoptosis, and the p53 target genes involved in these processes. Harris and Levine recently reviewed the downstream events of the p53 pathway and this led to a model of important p53 target genes and their function [27]. Figure 4 shows an adapted version of this model, provides an overview of the genes present on our microarray and lists

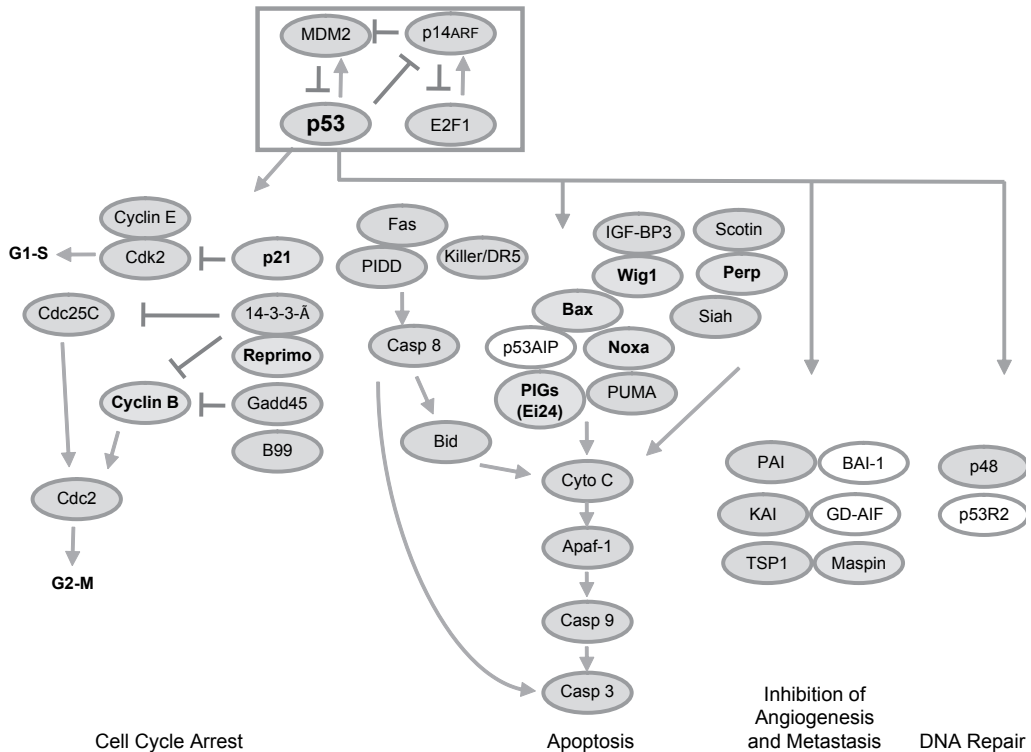


**Figure 3 - Graphical configuration of differentially-expressed genes in wild-type and p53.S389A bladders after short-term exposure to 2-AAF**

A) All differentially-expressed genes are displayed in a scatter plot to compare the expression values of the differentially-expressed genes found in bladders of wild-type (WT) and p53.S389A (SA) mice after short-term exposure to 2-AAF. Each spot represents an individual gene, where the axis shows the log<sub>2</sub> (expression values) for the 1<sup>st</sup> plot and log<sub>2</sub> (fold change levels) based on their corresponding controls for the 2<sup>nd</sup> and 3<sup>rd</sup> plot of wild-type (x-axis) and p53.S389A (y-axis). An orthogonal regression line is added to show the slope of the data points. Equal expression values of a gene in both wild-type and p53.S389A give a slope of 1. The slope (sl) of this line is displayed in the chart, as well as the corresponding correlation coefficient (cc).

B) A selection of genes (n=247) with a delayed response in p53.S389A (SA) compared to wild-type (WT) mice after 1 week of exposure (represented in A) is shown. Six pairs of line graphs have been made for these genes. In each pair the left plot are wild-type (WT) and the right plot are p53.S389A (SA) values of the selected genes. The number of genes in each plot is indicated above the corresponding pair of graphs.

C) Log<sub>2</sub> (expression values) of 6 individual genes corresponding with the line graphs depicted under B. Bold line: wild-type, Thin line: p53.S389A, C = control, 1 = one week exposure to 2-AAF exposure, 2 = two weeks exposure to 2-AAF.



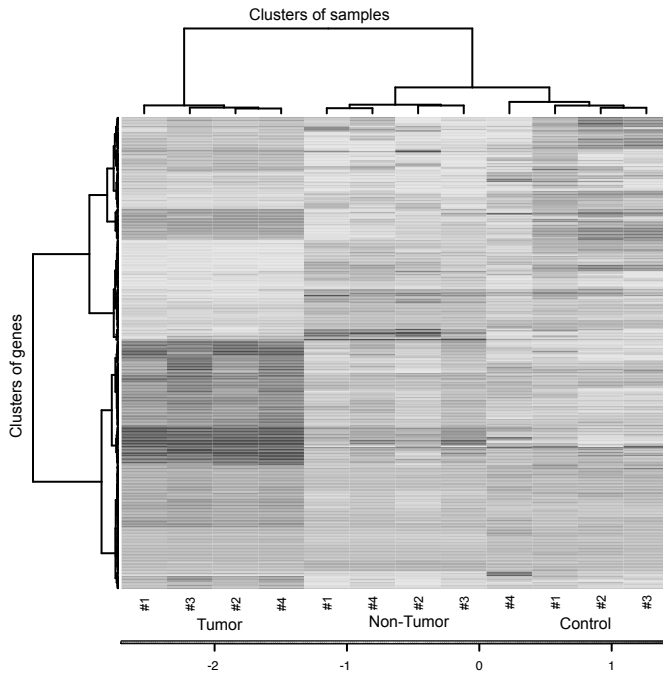
**Figure 4 - Adversely affected p53 target genes in bladders of p53.S389A mice after short-term exposure to 2-AAF**

Known p53 target genes involved in the apoptotic, cell cycle arrest, inhibition of angiogenesis and metastasis and DNA repair processes are presented (model adapted from [27]). Genes represented in yellow show a delayed response in bladders of p53.S389A mice compared to wild-type after exposure to 2-AAF. Genes represented in blue are present on our microarray but not differential between p53.S389A and wild-type; genes in white are absent on our microarray. For color figure, see page 183.

the genes showing a delayed response in our mutant mouse model. Several genes involved in these p53-dependent pathways were severely altered by the mutation at Ser389. In the cell cycle arrest pathway, three of the ten p53 target genes, *Cdkn1a* (*P21*), *Reprimo* and *Ccnb2* (*Cyclin B*), demonstrated a delayed response. Also, five of the nine analyzed genes involved in the intrinsic apoptosis pathway were affected by the p53.S389A mutation; *Wig1* (*PAG608*), *Perp*, *Bax*, *Pmaip1* (*Noxa*) and *Ei24* (*Pig8*). Interestingly, the extrinsic apoptotic pathway seemed not to be affected, at least not those genes described in the Harris and Levine model. Even in the pathway involving angiogenesis and metastasis expression, one of the four genes analyzed was affected; *Maspin*. In summary, differences in gene expression between p53.S389A and wild-type bladders after exposure to 2-AAF, as detected by microarray analysis, could be mapped to specific p53-dependent pathways.

### Analysis of gene expression in p53.S389A bladder tumors

The detected differences in gene-expression profiles of bladders of wild-type and p53.S389A mice after short-term exposure to 2-AAF, might explain the increased susceptibility of our mutant model for development of bladder tumors. To analyze whether these differentially-expressed



**Figure 5 - Hierarchical clustering of genes found to be differentially-expressed in tumorigenic versus non-tumorigenic p53.S389A bladders after long-term exposure to 2-AAF**

A total of 319 differentially-expressed transcripts were found by analysis of tumorigenic versus non-tumorigenic bladders of p53.S389A mice after long-term exposure to 2-AAF. Clusters were created using hierarchical clustering of the gene-expression profiles, where each row represents an individual gene. The degree of redness and greenness represent induction and repression respectively.

non-tumor = bladders after 39 weeks exposure to 2-AAF without tumorigenic bladder, tumor = bladders after 39 weeks of exposure to 2-AAF with tumorigenic bladder. For color figure, see page 183.

genes could be directly linked to bladder tumor development, we compared gene-expression profiles in both non-tumorigenic bladders and bladder tumor tissues. Since wild-type mice did not develop bladder tumors using the current exposure protocol, these analyses were restricted to p53.S389A mice only. A total of 319 significant genes were found between non-tumorigenic and bladder tumors of p53.S389A mice (Supplementary Table VI). Hierarchical clustering of the individual samples of the control, tumorigenic and non-tumorigenic bladders separated the groups, as a strong correlation across samples was found between the 319 differentially-expressed genes (Figure 5). Non-tumorigenic tissue exposed long-term to 2-AAF, clustered more closely to the controls than tumor-bearing bladders. In addition, absolute expression levels of the majority of genes appeared to be higher in the tumorigenic bladders compared to the non-tumorigenic bladders or controls. This seemed plausible given the high levels of cell proliferation or suppression of survival genes known to be present in tumors in general. Examples of significant biological processes found after GO analysis in this group of differential genes were metabolism, cell adhesion, muscle development, apoptosis and cytoskeleton organization. All of these are processes known to be involved in or affected by tumor formation (Supplementary Table VII). Some processes demonstrated a reduced expression level of genes in tumorigenic compared to non-tumorigenic bladder (e.g., cytoskeleton organization and muscle development), whereas others show an increased expression of most genes (e.g., cell adhesion).

To find out whether altered gene expression after short-term exposure coincided with genes affected in tumors, a comparison was made of genes found to be differentially-expressed in tumorigenic bladders (long-term exposure to 2-AAF) and genes found in p53.S389A bladders exposed short-term to 2-AAF. Interestingly, this comparison revealed 23 genes found both in short-term exposed bladders as well as in bladder tumors. Ratios of short-term exposed (i.e., 1 week) p53.S389A/wild-type and long-term exposed tumorigenic/non-tumorigenic

**Table II - Differentially-expressed genes detected in p53.S389A bladders after both long-term and short-term exposure to 2-AAF**

Accno	GO category	symbol	Ratios	
			tumor/non-tumor*	SA/WT 1 week**
NM_013834	Signal Transduction	<b>Sfrp1</b>	2,0	2,2
NM_019645	Signal Transduction	<b>Pkp1</b>	1,9	-0,1
NM_009704	Cell cycle	<b>Areg</b>	1,6	-0,2
NM_019736	Lipid Metabolism	<b>Acate2</b>	1,3	0,0
NM_009448	Nuclear Congression	<b>Tuba6</b>	1,2	0,3
NM_008385	Cell Growth and/or Maintenance	<b>Inpp5b</b>	1,0	-0,4
AF036898	DNA Replication	<b>Pole2</b>	0,8	-0,1
NM_010282	Lipid Metabolism	<b>Ggps1</b>	0,6	-0,1
NM_026662	Neurogenesis	<b>Prps2</b>	0,5	0,0
AK016670	Germ-Cell Development	<b>Bcl2l14</b>	-0,4	-0,1
AF109905	Chromatin Silencing at Ribosomal DNA	<b>Ly6g6c</b>	-0,5	-0,2
AK017677		<b>Cds2</b>	-0,7	-0,2
AK008108		<b>Sulf2</b>	-0,7	-0,7
NM_011663	Developmental Processes	<b>U2af1-rs1</b>	-0,7	-0,5
NM_007554	Cell Growth and/or Maintenance	<b>Bmp4</b>	-0,7	-0,5
NM_011390	Transport	<b>Slc12a7</b>	-0,7	0,0
NM_011812	Cell Matrix Adhesion	<b>Fbln5</b>	-1,1	-0,8
AK004418		<b>Synpo2</b>	-1,2	0,3
NM_009994	Electron Transport	<b>Cyp1b1</b>	-1,5	-0,7
NM_010145	Nitrogen Metabolism	<b>Ephx1</b>	-1,5	-1,4
U12961	Heat Shock Response	<b>Nqo1</b>	-1,6	-0,2
NM_007436	Aldehyde Metabolism	<b>Aldh3a1</b>	-1,9	-0,6
NM_008182	Stress Response	<b>Gsta2</b>	-1,9	-1,3

\* Ratio > 0 = tumor > non-tumor, \*\* Ratio > 0 = SA > WT  
SA = p53.S389A, WT = wild-type

were subsequently calculated, and are presented in Table II. Of the genes overexpressed in tumorigenic bladders after long-term exposure to 2-AAF, only two genes (*Sfrp1* and *Tuba6*) were also overexpressed in p53.S389A bladders exposed short-term, compared to wild-type. However, most of the genes repressed in the tumors were also decreased in p53.S389A bladders exposed short-term, compared to wild-type.

## Discussion

Previously we demonstrated that 39 weeks of *in vivo* 2-AAF exposure results in an increased incidence of bladder tumors in p53.S389A compared to wild-type mice [21]. Although the underlying molecular and cellular mechanisms remained unclear, it was thought that changes in gene expression might, at least in part, provide some insight in these processes.

Microarray experiments were performed with bladders from wild-type and p53.S389A mice that were exposed to the genotoxic carcinogen 2-AAF for either 1 or 2 weeks. These time points were chosen because a clear and significant induction of cell proliferation and apoptotic cells were previously observed after exposure to 2-AAF in wild-type and p53 knock-out mice [38]. Short-term 2-AAF exposure resulted in a large group of differentially-expressed genes, confirming the involvement of gene expression in response to a genotoxic carcinogen at early time points.

## Differentially-expressed genes between unexposed wild-type and p53.S389A bladders

Merely the lack of Ser389 phosphorylation resulted in a different expression pattern without the introduction of damage by 2-AAF, even though only 44 genes were found. An explanation



for the difference in genotypes under unexposed conditions could lie in different responses to spontaneously formed DNA damages, like reactive oxygen species (ROS) or depurination [reviewed in [39]]. Apparently, the basal p53 tumor suppressor system is readjusted to cope with spontaneous DNA damages, since p53.S389A mice are not tumor prone under unexposed conditions [20]. However, these readjusted responses together with an altered response to genotoxic compounds, might contribute to an increased susceptibility for bladder tumor development. A good example is the gene with the highest fold endogenous change level; *Sfrp1*. This gene is frequently affected in bladder tumors, and is used as a biomarker for bladder cancer detection in humans [40]. Another interesting example with one of the lowest fold endogenous change levels is *Pttg1* (*Securin*), which plays a role in p53-mediated cellular response to DNA damage. Rustgi *et al.* have already demonstrated that the C-terminus of p53 can interact with the amino-terminus of *Pttg1*, regulating apoptosis and transcription activity [41]. The S389A alteration at the C-terminus of p53 might therefore prohibit binding of *Pttg1*. In our experiment *Pttg1* was not specifically induced after exposure to 2-AAF but the expression level was already found to be reduced in p53.S389A unexposed bladders compared to wild-type. A constitutive expression level of *Pttg1* probably gives an overall steady level of some target genes involved in DNA damage response, so that wild-type cells can immediately react to 2-AAF exposure. Alternatively, *Pttg1* might be necessary to respond to endogenous DNA damage, and this response is constitutively activated through Ser389 phosphorylation. *Pttg1* is also known to be involved in the induction of apoptosis through the p53-dependent, pro-apoptotic gene *Bax* [42], and mice lacking *Pttg1* have aberrant cell cycle progression, premature centromere division and problems with chromosomal stability [43]. In our experiment, levels of *Bax* induction are indeed reduced in p53.S389A compared to wild-type bladders after exposure to 2-AAF, and this might finally lead to a less effective defense against carcinogens. Altogether, it seems likely that readjustments of basal gene expression level in response to the p53.S389A mutation plays a role in the bladder tumorigenesis upon exposure to 2-AAF.

### Processes involved in the responses to 2-AAF

Short-term 2-AAF exposure induced rather large effects on gene expression levels in bladders of mice, with most of the differentially-expressed genes functioning in apoptotic and cell cycle control processes. These findings are in line with a previous study using 2-AAF as the damaging agent, where it was found that apoptotic and cell proliferation related genes were up-regulated, whereas immune system related genes were down-regulated (note that these assays were examined in rat livers and B-cells respectively [44]). In our present study, however, we did not find differences in expression of immune response related genes in the bladders of mice after short-term exposure to 2-AAF.

### Delayed transcriptional response to 2-AAF in p53.S389A bladders

For the first time we demonstrated *in vivo* that the mutation at Ser389, a site involved in the post-translational modification of p53, caused altered gene expression patterns in bladders after exposure to a genotoxic carcinogen (Figure 4). In particular, we identified a set of genes whose change of expression after 1 week of exposure was clearly delayed in the p53.S389A mutant bladders. This group of 'delayed genes' included several p53 target genes, such as *Mgmt* (O-6-methylguanine-DNA methyltransferase), which is involved in DNA repair [45]. Recently, a microarray study demonstrated the largest induction for *Mgmt* after 28 days of 2-AAF administration [46]. Even



though this was specifically described for rat livers, our experiment also clearly revealed an induction of *Mgmt* in wild-type bladders after short-term exposure to 2-AAF, with a reduced activation of *Mgmt* after 1 week in p53.S389A mutant bladders. Apparently, phosphorylation of Ser389 is, at least in part, necessary for a rapid induction of *Mgmt* by p53.

The group of delayed genes also included the well-known p53 target genes involved in the p53-dependent apoptotic or cell cycle arrest responses to DNA damage (see also model by Harris and Levine, Figure 4 and [27]) like *Pmaip1* (*Noxa*), [47], *Bax* [48], *Wig1* [49], *Perp* [50], *Ei24* (*Pig8*) [51], *P21* [52], *Reprimo* [53], *Ccnb2* (*Cyclin B2*) [54] and *Ccng1* (*Cyclin G1*) [55]. Inactivation of these targets commonly results in an oncogenic phenotype. Therefore, the reduced expression levels in p53.S389A mutant bladders after exposure to 2-AAF, as observed in this study, can be clearly linked to increased tumor susceptibility. Preliminary results using immunohistochemical analysis of apoptosis in epithelial tissue of bladders after short-term exposure to 2-AAF, indicate a slight reduction of apoptotic levels *in vivo* in the p53.S389A mice compared to wild-type.

### **Genes found differentially-expressed in tumors and after short-term exposure to 2-AAF**

To identify possible tumor markers, i.e., genes that are associated with tumor formation, differentially-expressed genes found with the analysis of tumorigenic versus non-tumorigenic bladders after long-term exposure to 2-AAF were compared with genes found differentially-expressed in bladders after short-term exposure. A fraction of these genes, 23, indeed overlapped, indicating that these tumor-related genes are already affected by 2-AAF exposure at early time points. The list of genes as presented in Table II, contains some genes related to drug metabolism and tumor development, like *Fbln5* and *Sfrp1*. Expression of *Fbln5* has previously been associated with the suppression of tumor formation through its control of cell proliferation [56], and reduced expression of this gene has also been associated with the progression of malignant lymphoma [57]. In line with this, expression levels in tumorigenic tissue were decreased compared to non-tumorigenic p53.S389A bladders in our study, indicating phosphorylation of Ser389 is needed for optimal functioning of *Fbln5* in tumor suppression in the case of bladders after long-term exposure to 2-AAF. Other examples of overlapping genes are the cytochrome P450 family member *Cyp1b1* and the key enzyme involved in defense against reactive forms of oxygen *Nqo1*, which was also found to be required for the stabilization of p53 protein in response to DNA-damaging stimuli. Both are repressed after short-term 2-AAF exposure in p53.S389A bladders and in the bladder tumor. A final example is the Wnt-antagonist *Sfrp1* with induced expression levels both in unexposed p53.S389A bladders and in the bladder tumor. However, for the above-mentioned genes in our experiment, the ratio of expression levels of genes found in tumorigenic compared to non-tumorigenic tissue was the opposite of that in studies performed by others [40;58-62]. This might be because our experiment was performed in bladders of mutant mice, while these other studies mainly used bladder cancer samples or related cell lines from human patients that probably contain other genetic alterations that affect gene expression. In summary, our work has identified some genes differentially-expressed after short-term exposure to 2-AAF, which are associated with tumor formation in bladders of mice lacking the Ser389 phosphorylation event.

### **Concluding remarks**

As tumor formation is a process of several sequential stages like initiation, promotion and progression, the differences in response to bladder tumors between wild-type and p53.S389A mice

are expected to be present at all of these stages. This was particularly the case in our experimental set up, since it was previously reported that arylamines (such as 2-AAF) are involved in both initiating as well as promoting activities in carcinogenesis [63]. In this study we were interested in the gene-expression profiles after 1 and 2 weeks of exposure to 2-AAF in relation to a p53.S389A mutation, as this can give an impression of the initiation step of tumor development. In general we measured many subtle differences in gene expression. Mapping these findings to a recently published model of the p53 activation pathway demonstrated that a considerable number of these model genes were differentially-expressed in bladders of p53.S389A compared to wild-type mice after exposure to 2-AAF. These subtle changes in gene expression can be important for tumor development, as bladder cancer develops due to the accumulation of various molecular changes: i) chromosomal alterations, ii) loss of cell cycle regulation, iii) growth control events such as angiogenesis, resulting in metastasis and iiiii) decrease in cellular apoptosis [64]. Our results demonstrated a possible effect on chromosomal stability, as *Pttg1* expression is differential between the genotypes. Further, a clear effect on cell cycle regulation and apoptotic responses could be detected. Although angiogenesis-related genes and pathways were not found, the regulation of these processes might arise at a later stage in tumor development, i.e., far beyond the 2 weeks time point. In conclusion, a mutation at the phosphorylation site Ser389 results, *in vivo*, in specific altered gene-expression profiles in bladders exposed short term to carcinogens, which finally leads to increased susceptibility for tumor development in the bladder. Since responses of several (p53) target genes are not completely absent but only partially disturbed, and since only 14% of the p53.S389A mice developed tumors after 39 weeks, p53 still seems to be partially active in response to DNA damage induced by 2-AAF. Activation of p53 in the p53.S389A mice might presumably go through other post-translational modifications [21]. Mouse models with mutations at other phosphorylation sites also displayed intermediate cellular responses, underlining the idea of p53 functioning being fine tuned through post-translational modifications [65-68]. These results do, however, indicate the importance of post-translational modifications for proper p53 functioning. Phosphorylation of Ser389 appears to play a key role in fine-tuning the expression of a variety of important genes, as opposed to being the result of major changes in a few genes, because a reduction or delay in expression is measurable as opposed to a complete elimination. Finally, we demonstrated that phosphorylation of Ser389 is involved in some, but not all, functions of p53.

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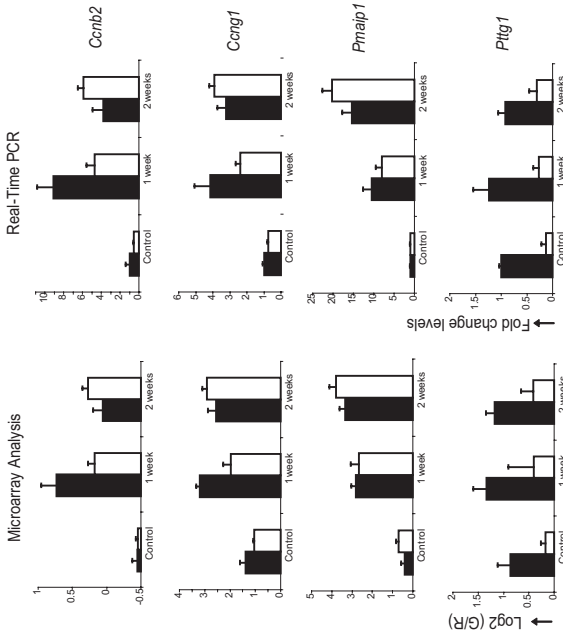
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## **Supplementary tables and figures**

**Sup. Table 1 – Log2 (expression ratios (p53.S389A/wild-type) of differentially expressed genes in unexposed bladders, based on genotype differences**

Symbol	Accession Number	Log2 (expression ratios)
Igh-6	AF052834	-0.90
Ptfg1	NM_013917	-0.80
Cd209a	AF373408	-0.63
Trf	J03299	-0.55
Dap	BC010828	-0.51
Siat7b	NM_009180	-0.50
4432412D15Rik	AK014490	-0.50
Fbln5	NM_011812	-0.48
Mmp2	NM_008610	-0.48
Ly6c	NM_010741	-0.36
Alp6ap1	NM_018794	-0.34
Trim16	AF220134	-0.34
Pja1	NM_008853	-0.33
Bmp4	NM_007554	-0.32
U5701	U5701	-0.32
Ccl11	NM_011330	-0.28
D11E1td99e	AK019025	-0.21
Pecan1	NM_008816	-0.21
Trim11	AF220124	-0.18
U93277	U93277	-0.15
Alox15b	NM_015754	-0.15
Rbbp9	NM_019647	-0.09
Rpl21	NM_011920	-0.05
Abcg2	AK012224	0.03
2700008N14Rik	AK009349	0.10
2310015A16Rik	AK017548	0.10
Tie4	AK016041	0.12
4930544O15Rik	AK016041	0.12
5730409G07Rik	AK016076	0.14
1300015D01Rik	AK005014	0.18
Ap3d1	NM_007460	0.18
Efcbp1	AK005641	0.18
4930484H19Rik	AK015621	0.18
4933434L15Rik	AK013967	0.19
1700047E16Rik	AK006705	0.20
3630403N18Rik	AK014417	0.22
2900053O20Rik	AK013680	0.22
Pcpb	NM_008796	0.28
Meis1	NM_010789	0.29
M30432	M30432	0.31
Prkwnk4	AK008047	0.53
5730405I09Rik	AK017493	0.57
1700052K11Rik	AK006771	0.62
Pparbp	NM_013634	0.89
Sfp1	NM_013834	1.40

**Sup. Figure 1 – Validation of gene-expression profiles obtained from microarray analysis by real time PCR**



A real-time quantitative gene-expression analysis using a Taqman-based detection method was used to verify differentially-expressed genes found by means of microarray analysis. Four genes, *Ccng1* (*Cyclin G1*), *Pmaip1* (*Noxa*), *Ccnb2* (*Cyclin B2*) and *Ptfg1* were randomly selected for this verification. Left plots show average expression level ( $\log_2$  (G/R)) values calculated with microarray analysis; right plots represent the fold changes found with real-time PCR analysis (unexposed wild-type samples were taken as basal level with value 1).



Sup. Table II – Differentially-expressed genes in wild-type bladders after short-term exposure to 2-AAF

Symbol	Acc. Nr.	FC 1 wk	FC 2 wks	Symbol	Acc. Nr.	FC 1 wk	FC 2 wks	Symbol	Acc. Nr.	FC 1 wk	FC 2 wks	Symbol	Acc. Nr.	FC 1 wk	FC 2 wks
Krt20	AK018567	-2.3	-3.6	301000IK23Rik	X04211	1.0	1.1	R605	AK004796	1.6	2.0	Sprf2c	NM_011479	2.5	2.0
9230106L14Rik	AF276974	-2.3	-3.6	BCW05542	AK011489	1.6	1.3	Lgn	NM_016753	1.7	1.3	Cdkn2b	NM_007670	2.5	2.6
D12Bwg1266e	V13832	-2.2	-2.2	5033428C03Rik	AKU17200	1.1	1.1	Lxn	NM_018636	1.7	1.1	Krt24a	NM_008475	2.6	2.2
231002012Rik	AK009423	-1.6	-1.2	Lgnb3	X16834	1.1	1.1	91300191SRik	AK018636	1.7	1.4	Pp4d3	NM_008975	2.6	2.1
4930426D05Rik	AK015205	-1.6	-1.5	2900073G15Rik	NM_026064	1.1	1.2	D11Erd603e	NM_026023	1.7	1.3	Aker1b3	NM_009658	2.6	1.8
Chrm4	NM_007699	-1.5	-1.4	ltpkb	AK004067	1.1	-1.2	D11Erd603e	NM_008360	1.7	1.5	Mkr67	X82786	2.6	1.5
Rnat1	AK002717	-1.5	-1.4	Aox1	NM_009676	1.1	-1.4	I118	NM_009523	1.7	1.3	Dmp1	U65020	2.6	2.3
Fatp9	NM_011598	-1.5	-1.6	Grb14	NM_016719	1.1	-1.2	Wnt4	AF091101	1.7	1.8	Gsta2	NM_008182	2.6	1.7
Beas1	AK008957	-1.4	-1.4	D10Erd610e	AK010452	1.1	-1.3	Mgmt	NM_009523	1.7	1.3	S100a11	U41341	2.8	2.1
4930524007Rik	AK001966	-1.4	-1.6	Trim35	AKM03000	1.1	-1.3	Mgmt	NM_007629	1.7	1.4	Spr1a	NM_009264	2.8	2.7
D6Bwg1452e	AK004185	-1.4	-1.8	Scpn	NM_010656	1.1	-1.3	Mmp53	NM_008598	1.7	1.7	Epbl1	NM_010145	3.0	2.3
1300013J1SRik	M94308	-1.4	-1.5	3110050N22Rik	AK012273	1.1	1.2	Hsp22	AK005439	1.8	1.8	Mel	NM_010762	3.1	2.3
1300013J1SRik	NM_026183	-1.4	-1.9	Sgol2	AK003555	1.2	1.1	Hsp22	NM_019487	1.8	1.8	Nos3	AK010032	3.2	5.5
L1b4dh	NM_025968	-1.3	1.3	Cap1	NM_007598	1.2	1.2	Birc5	NM_009689	1.8	1.4	Lys6d	NM_010742	3.2	2.4
Adss1	NM_007421	-1.3	-1.4	Helz	AK020667	1.2	1.1	Birc5	NM_009689	1.8	1.4	Anxa3	NM_013470	3.3	2.7
Lmed1	AF070470	-1.3	-1.5	Nud5	NM_016918	1.2	1.3	Birc5	NM_009689	1.8	1.4	Anxa3	NM_013470	3.3	2.7
Col19a1	NM_007753	-1.3	-1.8	Dp111	AB039933	1.2	-1.1	493342G05Rik	AK016892	1.8	1.5	Cyp1b1	NM_009994	3.3	2.2
Knmbp4	NM_021452	-1.2	-1.1	Acp2	AK009134	1.2	-1.1	Msh6	NM_010830	1.9	1.1	Krt1-15	NM_008469	3.3	2.8
Fkbp4	NM_010219	-1.2	-1.1	493342G06Rik	AK005558	1.2	1.8	Pgaml1	AF283667	1.9	1.4	Ceng1	NM_009831	3.5	2.2
Sympo2	AK004418	-1.2	-2.1	Utr315	AK014780	1.3	1.4	Nqo1	U12961	1.9	1.2	Areg	NM_009704	3.9	5.8
Nymb	NM_026523	-1.2	-1.3	Hn1	NM_008258	1.3	1.0	Gfer	AF148688	1.9	1.5	Krt1-23	NM_033373	4.0	3.5
Nmb	NM_009722	-1.2	-1.9	Abhd9	AK010021	1.3	1.3	Gpr115	AK019508	1.9	1.5	Cttnl1	NM_009897	4.0	3.4
Apr2a2	NM_008168	-1.2	-1.2	Daf6	AKM10356	1.3	1.4	Trim16	AF220134	1.9	2.6	Pnaip1	NM_021451	5.3	7.7
Grk5	NM_008168	-1.2	-1.2	Pole2	AF056898	1.3	1.1	AK1	NM_021515	2.0	1.8	Spr2f	NM_011472	7.9	13.9
Chn1	AK019340	-1.2	-1.2	130006M19Rik	BC015459	1.3	1.1	AK1	NM_021515	2.0	1.8				
Ehlt	AF121981	-1.2	-1.3	Ard4d	AK010352	1.3	1.3	Igfb4	NM_025520	2.0	1.6				
Sing2	NM_010210	-1.2	1.4	Rrm2	NM_009104	1.3	1.3	Igfb4	L04678	2.0	1.6				
Alox12	AF367760	-1.2	-1.6	Fabp2	NM_007980	1.3	1.3	Serpinc5	NM_009257	2.0	1.4				
C78915	AJ409496	-1.1	-1.5	Dlx28	AKM10396	1.3	1.5	Epha1	AF131197	2.0	1.8				
1110057K04Rik	NM_007440	-1.1	-1.2	Gnal3	NM_010303	1.3	1.1	Bov1	NM_025824	2.0	1.4				
Ly6g6c	NM_011390	-1.1	-1.6	Pp1r14b	NM_008889	1.3	1.3	Css	NM_008180	2.0	2.1				
Fanel	NM_019704	-1.1	-1.1	U2ad1-ns1	NM_011663	1.4	1.1	Spr2g	NM_025889	2.1	1.7				
Sod1	AK004285	-1.1	-1.4	Mrel1a	NM_029043	1.4	1.2	923017N10Rik	NM_011473	2.1	2.4				
GIG18	AF109905	-1.1	-1.2	Lzic	NM_018736	1.4	1.2	Rrm1	NM_009103	2.1	1.6				
GIG18	BC012698	-1.1	-1.1	Spr2j	AK007657	1.4	1.5	Rpk3	NM_019955	2.2	1.9				
Trm25	NM_025923	-1.1	-1.3	Tuft1	NM_011476	1.4	2.2	Anxa8	NM_013473	2.2	2.2				
Pkl1	AF374476	-1.0	1.1	Bbs2	NM_011656	1.4	1.5	Rbx1	NM_019712	2.2	1.7				
3110001N18Rik	D63902	-1.0	1.3	BC004728	AF342737	1.4	1.4	Searb2	NM_007644	2.2	1.7				
4632432E15Rik	NM_013650	-1.0	-1.3	Dgka	NM_016811	1.4	1.4	Caa	NM_008064	2.2	1.5				
4930542G03Rik	NM_026517	-1.0	-1.5	Elovl4	AF277093	1.4	1.4	Tusc4	NM_018879	2.2	1.7				
Ptprz1	AK014604	-1.0	2.1	Rargamo	AK008785	1.4	1.2	Bax	NM_019645	2.2	2.3				
Stac	BC002298	1.0	-1.2	Gbl1	AK043586	1.4	1.8	Txn2f	NM_007527	2.2	2.0				
Cd82	NM_016853	1.0	-1.4	Gcfr1	BC002135	1.4	2.1	Gsta1	AF118650	2.2	1.7				
Cdkn1a	AK017677	1.0	-1.6	Col7a1	NM_009732	1.5	1.6	Rps27l	NM_008181	2.2	1.8				
Ptprn1	NM_007669	1.0	1.1	C1gnt1	AK003420	1.5	1.6	Wig1	NM_026467	2.3	1.9				
1110021P09Rik	BC003980	1.0	-1.2	1210005A06Rik	AF157962	1.5	1.2	Cemb2	NM_009517	2.3	1.4				
4921504P20Rik	AK003907	1.0	-1.1	Pold3	AF294329	1.5	1.8	181002720Rik	AK007617	2.3	2.1				
	AK014816	1.0	1.2	Sral1	AF092039	1.5	1.5	-	NM_019669	2.3	2.2				
				Galm3	NM_015736	1.5	1.2	S100a14	NM_018806	2.3	2.4				
				Dpp8	AK016546	1.6	1.4	2210023G05Rik	AK008775	2.4	1.3				
				Rkn	NM_009106	1.6	2.2	Ube2c	AK003722	2.4	1.5				
								1190003J15Rik	AK004470	2.4	2.6				

Acc.Nr. = Accession Number  
FC = Fold Change  
wk = week





Symbol	Acc.Nr.	FC 1 wk	FC 2 wks	Symbol	Acc.Nr.	FC 1 wk	FC 2 wks	Symbol	Acc.Nr.	FC 1 wk	FC 2 wks
Msi2h	AB156103	1.3	1.8	A830059120Rik	NM_021427	1.5	1.5	Mcm5	NM_008566	1.6	1.5
Tm7s1f	NM_054337	1.3	1.3	Cst6	AK003744	1.5	1.7	3830408G10Rik	NM_008566	1.6	2.1
Cde42ap5	NM_021454	1.3	1.9	Hmxo1	NM_010442	1.5	2.0	Inhba	NM_008380	1.6	2.1
D11Etrid603e	NM_026023	1.3	1.6	1700020114Rik	AK006166	1.5	1.7	BC006059	NM_009312	2.4	3.5
Hn1	NM_008258	1.3	1.7	Vamp5	NM_016872	1.5	2.6	6820402018Rik	AK018359	1.7	2.0
Nxrl1	NM_019761	1.3	1.5	130006M19Rik	BC015459	1.5	1.7	Spr24g	NM_011473	1.7	3.7
Rxrl1	NM_019712	1.3	2.2	Cast	AB026997	1.5	2.2	Dnmp1	L65020	1.7	2.4
2310058J06Rik	AJ299430	1.3	1.5	AK1	NM_021515	1.5	1.7	Gm33	AK013750	1.7	2.1
493054ZG03Rik	BC005347	1.3	2.4	2010005A06Rik	AK008111	1.5	1.7	Dpys4	NM_019389	1.7	1.7
Dpkx28	AK010396	1.3	1.7	Mue2	AF121215	1.5	1.5	-	NM_007827	1.7	2.3
Ppss2	NM_026662	1.3	1.5	Tae3	NM_011524	1.5	1.5	Dat2	NM_008181	1.7	2.1
Dpp8	AK016546	1.3	1.7	Hgfac	NM_019447	1.5	1.4	Gstal	NM_008181	1.7	2.1
Aug-np1	AK004318	1.3	1.7	E030027H19Rik	BC011340	1.5	2.5	Nasp	NM_009265	1.7	2.1
Sprf10	U22517	1.3	1.4	Dnaj9	BC014686	1.5	2.2	Spr1b	NM_025415	1.7	1.9
Skmt1	NM_009997	1.3	3.3	Pknox1a	BC003461	1.5	2.4	Cks2	NM_013754	1.7	2.2
Trmpss2	NM_015775	1.3	1.7	Brcal	U36475	1.5	1.6	Ins16	NM_013601	1.7	1.7
Gho	NM_008094	1.3	2.0	1700007K13Rik	AK005731	1.5	2.4	Msc2	NM_019955	1.7	2.2
1110059M19Rik	AK004321	1.3	1.4	Scamp5	NM_020270	1.5	2.3	Rpk3	NM_009658	1.7	3.1
Notch3	NM_008716	1.4	1.7	-	NM_007629	1.5	1.5	Akrb3	NM_016917	1.7	3.4
-	NM_018752	1.4	1.7	4632411J06Rik	AK019489	1.5	1.4	Sle4b1	NM_008879	1.7	3.4
Indo	NM_008324	1.4	2.6	Pole3	NM_021498	1.5	1.6	Tusc4	NM_018879	1.7	2.4
Stat4	NM_011487	1.4	1.8	Hexa	NM_010421	1.5	1.7	Gsta4	NM_010357	1.7	3.3
Cd44	M30655	1.4	1.5	Tmml6	AF220134	1.5	2.2	Phlda3	NM_013750	1.7	2.1
Panx1	NM_019482	1.4	1.7	2610034P21Rik	AK011679	1.5	1.4	Gaa	NM_008064	1.8	2.7
Hdh	U24233	1.4	1.4	Mal	NM_010762	1.5	1.9	Slc1	NM_011519	1.8	2.1
2610034E01Rik	AK011660	1.4	1.4	Robin	NM_009708	1.5	2.0	2210023G05Rik	AK008775	1.8	2.9
Ela9l4	AF277093	1.4	1.5	1190003J15Rik	AK004470	1.5	2.8	Sik	NM_009289	1.8	1.9
Gih1	NM_009752	1.4	1.4	Gzmc	NM_010373	1.5	1.6	Clu	NM_013492	1.8	3.8
Auss2	NM_007422	1.4	1.5	5730507H05Rik	AJ237585	1.5	1.4	Tmem27	NM_020626	1.8	1.6
Tmfrs18	NM_009400	1.4	1.5	Cebpg	BC011319	1.5	1.8	Rtkn	NM_009106	1.8	2.7
Pknox3	NM_019587	1.4	1.4	Bax	NM_007527	1.5	1.9	S100a11	U41341	1.8	2.6
0710001D07Rik	AK002941	1.4	1.4	Gsta2	NM_008182	1.5	3.3	Wfrc2	AK005519	1.8	3.2
Nudf5	NM_016918	1.4	2.2	4632432E15Rik	AK014604	1.5	1.6	Bzwl	NM_025824	1.8	2.2
8030444G23Rik	AK020201	1.4	1.6	Acate2	NM_019736	1.5	1.7	1810037117Rik	BC002135	1.8	3.1
Axa8	NM_013473	1.4	2.5	1600029D21Rik	AK005558	1.5	1.5	Inscp	NM_016692	1.8	1.6
6330414G21Rik	AK018173	1.4	2.0	Ccnb2	NM_007650	1.5	1.7	Cdca2b	NM_007670	1.8	3.4
Pgaml	AF283667	1.4	1.6	Psat1	BC004827	1.6	1.1	Pgpl	NM_019645	1.8	2.2
Ephal1	AF131197	1.4	2.1	Gss	NM_008180	1.6	2.4	Ppfd43	NM_008975	1.9	2.9
Dxrl9	NM_007916	1.4	1.6	Rrm1	NM_009103	1.6	1.9	S100a14	NM_025393	1.9	2.7
Yotbab	NM_018753	1.4	1.8	Dpagl1	NM_007875	1.6	1.4	Tuesq2	NM_020047	1.9	2.3
1700026D08Rik	AK006375	1.4	1.2	H18	NM_008360	1.6	1.8	9130019115Rik	AK018636	1.9	2.6
Wigl1	NM_009517	1.4	1.9	1700013H19Rik	AK005954	1.6	1.7	Ly6a	NM_010738	1.9	3.0
Gfer	AF148688	1.4	2.3	Homer3	AF093261	1.6	1.5	Birc5	NM_009689	1.9	1.7
Sat1	NM_009121	1.4	2.5	Sprria	NM_009264	1.6	3.5	Ceng1	NM_009831	1.9	3.7
5730596K12Rik	AF309554	1.4	2.7	Dusp	AF091101	1.6	1.6	Mki67	X82786	1.9	2.3
2700038L12Rik	BC002149	1.4	1.4	1190002A23Rik	AK004440	1.6	1.8	Fsn1	NM_007984	1.9	1.8
Fmo5	NM_010232	1.4	1.7	Fgf10p2	AB041650	1.6	2.0	Ly6d	NM_010742	1.9	3.8
Hmfs	NM_012057	1.4	1.9	Srebp5	NM_009257	1.6	2.1	Ancx3	NM_013470	2.0	3.1
Clcl	BC004658	1.4	1.4	Cd14	NM_009841	1.6	2.0	Slc2a8	NM_019488	2.0	2.6
1200003E16Rik	AK004560	1.4	1.9	Hsd17b12	NM_019657	1.6	2.2	Hfbp2	NM_019487	2.0	1.9
Dazp1	AF225910	1.4	1.8	Gap43	NM_008083	1.6	1.9	Ube2c	AK003722	2.0	2.0
Tmm8b	NM_013899	1.4	1.9	Cug	NM_007599	1.6	2.2	Coll17a1	NM_007732	2.0	2.2
Fbp2	NM_007994	1.4	1.8	4833427G06Rik	AK014780	1.6	2.0	Alb33a1	NM_007436	2.1	2.0
Lzic	AK007657	1.4	1.2	Dnajc2	NM_019794	1.6	1.7	2310061G07Rik	BC003749	2.1	2.2
2310020F24Rik	AK009417	1.4	1.5	Ccpna	NM_007681	1.6	1.5	Cyp11b1	NM_009594	2.1	2.7

Acc.Nr. = Accession Number  
 FC = Fold Change  
 wk = week

**Sup. Table IV – Biological processes (i.e., GO categories) containing genes involved in 2-AAF responses in mouse bladders**

GO category	Total*	Corrected p-value*
Cell cycle	14	0.0084
Electron transport	13	0.0224
Cytokinesis	10	0.0035
DNA replication	9	<0.001
Response to DNA damage stimulus	9	0.0051
Carbohydrate metabolism	8	0.0298
Protein ubiquitination	9	0.0309
DNA repair	7	0.0406
Lipid metabolism	7	0.0291
Mitosis	7	0.0027
Induction of apoptosis	6	0.0033
Angiogenesis	4	0.0466
Cell cycle arrest	4	0.0043
Negative regulation of cell proliferation	4	0.0249
Neuropeptide signaling pathway	4	0.0464
Nucleobase, nucleoside, nucleotide and nucleic acid metabolism	4	0.0028
Rho protein signal transduction	4	0.0033
DNA metabolism	3	0.0291
DNA replication and chromosome cycle	3	<0.001
Induction of apoptosis by intracellular signals	3	<0.001
One-carbon compound metabolism	3	0.0323
Protein catabolism	3	0.0115
Protein localization	3	0.0079
Response to UV	3	<0.001
Thyroid hormone generation	3	<0.001

\*Processes with at least 3 genes present and a corrected p-value <0.05 are shown

**Sup. Table V – Biological processes (i.e., GO categories) containing genes with a delayed response in p53.S389A bladders compared to wild-type bladders**

GO category	Total
Transport	15
Cell cycle	9
Proteolysis and peptidolysis	9
Cytokinesis	7
Development	7
DNA replication	7
Electron transport	6
Metabolism	6
Regulation of transcription, DNA dependent	6
Signal transduction	6
Induction of apoptosis	5
Mitosis	5
Protein amino acid phosphorylation	5
Response to DNA damage stimulus	5
Apoptosis	4
Carbohydrate metabolism	4
Cell adhesion	4
Cell differentiation	4
DNA repair	4
G-protein coupled receptor protein signaling pathway	4
Lipid metabolism	4
Protein biosynthesis	4
Protein folding	4
Protein transport	4
Immune response	4
Induction of apoptosis by intracellular signals	3
Integrin-mediated signaling pathway	3
Intracellular protein transport	3
Protein targeting	3
Protein ubiquitination	3
Regulation of apoptosis	3
Rho protein signal transduction	3
Transcription	3
Ubiquitin cycle	3

**Sup. Table VI – Differentially-expressed genes in tumorigenic compared to non-tumorigenic p53.5389A bladders after long-term exposure to 2-AAF**

Symbol	Acc.Nr.	Fold Change Tumorigenic	Symbol	Acc.Nr.	Fold Change Tumorigenic	Symbol	Acc.Nr.	Fold Change Tumorigenic	Symbol	Acc.Nr.	Fold Change Tumorigenic
Myh11	NM_013607	-8.2	Scn11g	NM_011326	-2.2	Map3k7ip1	AK009321	-1.6	4932412D3Rik	AK016521	-1.0
Gsta3	NM_010356	-7.7	Snr2c	AF367760	-2.2	Sash1	AK0003151	-1.6	BC005561	BC005561	-1.0
Adh1	NM_007409	-6.8	Akap12	AB020886	-2.2	Cks2	AK017677	-1.6	Chk2	AK005681	-1.0
Acta2	NM_007392	-4.9	Ehlns	NM_011812	-2.1	3110048E14Rik	AK014192	-1.6	Chk2	NM_016681	-1.0
Actg2	NM_009610	-4.6	Sh3bp1	NM_015625	-2.1	-	NM_028785	-1.6	1700026G02Rik	AK005362	-1.0
4631428E05Rik	AK014536	-4.6	Abcc3	AK006128	-2.1	Cirbp	NM_007705	-1.6	2310005A03Rik	AK009160	-1.0
Ugr1a6	U16818	-4.6	Col14a1	AJ131395	-2.1	Gpc6	AK020118	-1.6	Nfia	U57633	1.0
Upr1a	AF073956	-4.6	S3-12	NM_020568	-2.1	Gpr124	AF378759	-1.5	Leimd1	AK003833	1.1
Upr3a	AF222750	-4.6	1110003E01Rik	NM_009097	-2.1	Acdy9	NM_009624	-1.5	Slo6d1	AK014872	1.1
Upr3a	AF222750	-4.4	Upk2	NM_009476	-2.1	AKp81	NM_017476	-1.5	4930459C07Rik	AK015487	1.1
Myi9	AK007972	-4.3	Tm4s16	NM_019656	-2.1	2900060P06Rik	AK013739	-1.5	4930554H23Rik	AK019764	1.1
Gsta2	NM_008182	-4.3	Gst4	BC003903	-2.1	Sic14a1	AK012066	-1.5	Fgf14	NM_010201	1.1
Alch3a1	NM_007436	-3.9	Tnxb	BC003288	-2.1	2900441A09Rik	AK013631	-1.5	Dhnt	NM_008345	1.1
1110032A04Rik	AK036876	-3.7	Igfbp6	NM_008344	-2.0	Aig2	NM_009705	-1.4	AF358859	AF358859	1.1
Lhd2	NM_008492	-3.6	Ptx2	NM_011098	-2.0	Ly6g6c	AF109905	-1.4	BC006036	BC006036	1.2
Cyb5	NM_025797	-3.5	061001015Rik	AK002481	-2.0	Traf6	NM_017633	-1.4	NM_025469	NM_025469	1.2
Dcs	L22560	-3.4	Sic16a7	NM_011391	-2.0	Hoxd10	NM_013554	-1.4	Clps	NM_025469	1.2
Norg2	D00926	-3.4	Gata3	NM_008091	-2.0	Asb2	AF155353	-1.4	BC013520	BC013520	1.2
Tcea3	NM_013864	-3.4	Sgpl1	NM_009163	-2.0	2010316F05Rik	AK008574	-1.4	Vrk2	AJ278264	1.2
Smtn	NM_013870	-3.2	Jph2	AK003288	-2.0	Map4k2	NM_009006	-1.4	Tnfrsf23	AJ278264	1.2
Tnnt2	NM_011619	-3.2	9530014B07Rik	NM_024255	-1.9	8430421H08Rik	AK018430	-1.4	1700120G07Rik	AK007216	1.2
Deptd6	BC004774	-3.1	IV ADH	2610207116Rik	-1.9	8430436L14Rik	AK018464	-1.3	En2	NM_010134	1.2
Upl1a	AK004014	-3.0	1810041L15Rik	AK007745	-1.9	Tbtd	NM_009378	-1.3	Rbm22	AK018452	1.2
Prom2	AF128113	-3.0	Rip	NM_011125	-1.9	Bcl2l14	NM_016670	-1.3	Gfz2a1l	NM_023630	1.2
4631408O11Rik	AK014514	-2.9	Cbr1	NM_007620	-1.9	Tk2	NM_011902	-1.3	1810053B23Rik	AK007854	1.2
Ngn1	U12961	-2.9	Pcp41	AK002772	-1.9	9530405J04Rik	AK013190	-1.3	2600001B17Rik	AK011126	1.2
Gstm2	NM_008183	-2.9	Fmo5	AK017179	-1.9	1700009P17Rik	AK005804	-1.3	AF149205	AF149205	1.2
4632417N05Rik	AK014566	-2.9	Sorsb1	AK010043	-1.8	2310005P05Rik	NM_026189	-1.3	Suv39h2	AF219945	1.2
Ephx1	NM_010145	-2.9	Adh3b1	AK005615	-1.8	Dslp1	NM_010286	-1.3	Tulp4	AF219945	1.2
Cyp11b1	NM_009994	-2.9	Rassf3	BCO11511	-1.8	Phn2	NM_019410	-1.3	6030444E23Rik	AK020061	1.2
Pdlim3	NM_016798	-2.8	Gjb6	NM_008128	-1.8	Coh1	AK007749	-1.3	J05609	J05609	1.2
mt-Nd5	AK018737	-2.8	Sic2a4	NM_009204	-1.7	Tp2	AK019456	-1.3	2700003A03Rik	AK012205	1.2
C030033M19Rik	AK021118	-2.8	Myk	AF314149	-1.7	D11Vslu68e	NM_028776	-1.2	1500005K14Rik	AK005164	1.2
-	U50959	-2.8	Pparg	NM_011146	-1.7	Abcc2	NM_011984	-1.2	3930401E15Rik	AK013936	1.2
Coch	NM_007728	-2.8	1110032A03Rik	AJ250230	-1.7	Scn3	BC015296	-1.2	X56613	X56613	1.2
Alp2a2	NM_009722	-2.7	Pdk2	AF267660	-1.7	Sern3	BC015296	-1.2	Psg28	AF113598	1.3
Taf1h	NM_011526	-2.7	A930010G16Rik	AK020843	-1.7	2900003A17Rik	AK013479	-1.2	Smurf2	AK012600	1.3
Alf3	AK016257	-2.7	4930469P12Rik	AF412298	-1.7	Sox17	NM_011441	-1.2	Seh1	AK011330	1.3
Fmo2	AK009753	-2.6	Tripv4	AF208206	-1.7	1810013C16Rik	AK007472	-1.2	Zfp51	AB010357	1.3
Rhnt18	AB041548	-2.6	Fbnp1	AK004278	-1.7	Sic29a2	NM_007854	-1.2	Rb1	U27177	1.3
Home2	AK001736	-2.6	Dscr5	NM_019543	-1.7	Pex11b	NM_011069	-1.2	Fbxo28	AK018321	1.3
2200001115Rik	AK008614	-2.6	Bmp4	NM_011390	-1.7	Ceb	AK009844	-1.2	Dusp10	AB037908	1.3
Ogn	AK014259	-2.6	1810019D21Rik	NM_011863	-1.7	Aes	NM_010347	-1.2	C030039E19Rik	AK021134	1.3
Sytl2	M35725	-2.5	U2af1rs1	AK007545	-1.7	6330407A06Rik	AK020091	-1.2	4830431P22Rik	AK015281	1.3
Sod1	NM_010354	-2.4	Sorr1	AK013673	-1.6	Ry11	X39332	-1.1	Hoxd13	NM_008267	1.4
Tcf21	NM_011545	-2.4	Sulf2	AK008108	-1.6	5430400D12Rik	AK017243	-1.1	Cenph	NM_021886	1.4
Lmo61	AF237627	-2.4	Capz3	NM_007605	-1.6	Ahr	NM_009644	-1.1	Tbc1d1	NM_019636	1.4
Prplp	AK020180	-2.3	Prdk3	NM_007452	-1.6	Tscot	NM_021053	-1.1	Drd1a	AK021124	1.4
Synpo2	AK004418	-2.3	Pdh2	NM_008811	-1.6	Tpr40	NM_011906	-1.1	Sag	NM_009118	1.4
P2rx1	NM_008771	-2.3	Lmo1	AJ296304	-1.6	Th2	NM_021028	-1.1	Prps2	NM_026662	1.4
-	-	-	1700021N21Rik	AK006224	-1.6	1700003F17Rik	AK005634	-1.1	Pigt	AK019717	1.4
-	-	-	Sh3p2	NM_019535	-1.6	1500004F05Rik	AK005145	-1.1	Phca	AK004287	1.4
-	-	-	Ctnnb2	NM_025771	-1.6	4930444E06Rik	AK015377	-1.0	-	NM_029207	1.4
-	-	-	-	-	-	D730003K21Rik	AK021326	-1.0	Skil	NM_011386	1.4

**Sup. Table VIII – Biological processes (i.e., GO categories) containing genes involved in 2-AAF induced tumor response in mouse bladders**

GO category	Total	Corrected p-value
Metabolism	11	0.0044
Cell adhesion	10	0.0287
Muscle development	9	<0.001
Apoptosis	7	0.0456
Cytoskeleton organization and biogenesis	7	0.0005
Small GTPase mediated signal transduction	5	0.0366
Pattern specification	4	0.0064
Skeletal development	4	0.0023
Cell fate commitment	3	0.0064
Chromatin assembly or disassembly	3	0.0046
Chromatin modification	3	0.0324
DNA replication	3	0.0453
Glycolysis	3	0.0085
Inflammatory response	3	0.0465
Neuropeptide signaling pathway	3	0.0309
Perception of sound	3	0.0103
Regulation of growth	3	<0.001
Regulation of muscle contraction	3	0.0021

Symbol	Acc.Nr.	Fold Change Tumorigenic	Symbol	Acc.Nr.	Fold Change Tumorigenic
2010002N04Rik	AF313412	1.4	Rhoj	AJ276568	1.9
Gtf2f2	AK003999	1.4	Mmp9	NM_013599	1.9
-	U23095	1.5	Dlr	NM_010415	1.9
2310008M10Rik	NM_025509	1.5	Neb	U58108	1.9
-	U50960	1.5	Inpp5b	NM_008385	1.9
-	M12194	1.5	Celsr1	NM_009886	2.0
Ggpps1	AF120320	1.5	Lamb3	NM_008484	2.0
2410004D21Rik	NM_010282	1.5	Irfid1	NM_013662	2.0
Ddit4	BC003216	1.5	Mek1	NM_010790	2.0
Ddit4	AK017926	1.5	1500012M23Rik	AK005238	2.0
10E10E16e	AK009683	1.5	S100a3	NM_011310	2.1
Ttk	NM_009445	1.5	1700083E07Rik	BC013063	2.1
2300006M17Rik	BC006061	1.5	Piscr2	NM_008880	2.1
2610206B13Rik	AK011896	1.5	Shcnp1	NM_011369	2.1
Has3	NM_008217	1.5	Hk2	NM_013820	2.1
Psmc14	AF290967	1.5	Igf4a	NM_018770	2.1
Ptdc6ip	NM_021526	1.5	Tnfrsf11b	NM_008764	2.1
Tlf2	NM_011052	1.5	Rab38	NM_028238	2.1
Bmp1er	AK014221	1.6	Gdf1	NM_008107	2.1
Hoxb6	X56461	1.6	Serp1nbt1	NM_025429	2.1
Rala	NM_019491	1.6	Rras2	NM_025846	2.2
Tex292	X80433	1.6	Piscr1	NM_011636	2.2
Gpr87	NM_032399	1.6	2310076G05Rik	AK010197	2.3
-	AK006925	1.6	Fnbp3	NM_018785	2.3
Jub	NM_010590	1.7	Tuba6	NM_009448	2.3
1700030C10Rik	AK006529	1.7	Cd44	M30655	2.3
Sdcd1	BC004579	1.7	Tmd4sf1	NM_008536	2.4
Olf64	NM_013616	1.7	Birc5	NM_009689	2.5
BC003236	NM_030249	1.7	Sfn	NM_018754	2.5
PspH	BC002251	1.7	Acate2	NM_019736	2.5
-	NM_019509	1.7	1200002N14Rik	AK004552	2.5
Ccng2	NM_007635	1.7	Trim15	AK007445	2.5
Gjb4	NM_008127	1.7	Sic14a2	AF258602	2.7
Syncrip	AB035725	1.7	Timp1	NM_011593	2.7
Nodal	NM_013611	1.7	Adam8	NM_007403	2.7
Nolc1	BC003244	1.7	Vsnl1	NM_012038	2.8
Sgpl2	AK003555	1.7	Tgfb1	NM_009369	2.8
Tll4	AK014557	1.8	Sox2	NM_011443	2.9
Nola2	NM_026651	1.8	Gm566	AF014453	2.9
6720460F02Rik	AK011345	1.8	E030027H19Rik	BC011340	2.9
Fg66	AK016940	1.8	Bnc1	U88064	3.1
Ubigln1	NM_026842	1.8	Areg	NM_009704	3.1
Pole2	AF036898	1.8	C12a2a	NM_007796	3.1
Fosl1	NM_010235	1.8	Serpine1	NM_008871	3.4
Map4k4	NM_008696	1.8	Pkp1	NM_019645	3.7
Trip13	AK010336	1.8	Sfrp1	NM_013834	4.1
Smarca5	AF325921	1.9	Col17a1	NM_007732	4.3
Agpl2	NM_007426	1.9	Mmp13	NM_008607	4.6
Pkp3	NM_019762	1.9			
Bhlh2	NM_011498	1.9			
Irf6	NM_016851	1.9			
Ch25h	NM_009890	1.9			
1110015M06Rik	AK003729	1.9			
AA545217	NM_013726	1.9			

Acc.Nr. = Accession Number



