

From NSD1 to Sotos syndrome : a genetic and functional analysis

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Citation

Visser, R. (2011, May 26). *From NSD1 to Sotos syndrome : a genetic and functional analysis*. Retrieved from https://hdl.handle.net/1887/17673

Note: To cite this publication please use the final published version (if applicable).

Chapter 2

Overgrowth syndromes: from classical to new

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Pediatr Endocrinol Rev 2009; 6: 375-394

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Abstract

Overgrowth syndromes are a group of growth disorders which have gained joint attention from the fields of pediatrics, endocrinology and genetics. Major progress such as the identification of genetic causes has recently enhanced the delineation of the characteristic and non-characteristic manifestations, phenotype-genotype correlations and knowledge of the underlying pathophysiological mechanisms. As a consequence, the possibilities for distinction between the different overgrowth disorders have increased. Patients with either typical or non-typical features in whom no molecular abnormalities are found, form a basis for further research. Identification of new pathogenic alterations in these patients, best exemplified by the Marfan-related syndromes, has provided further understanding of the regulatory gene network involved.

In light of the recent developments and as an aid to the diagnostic process, the aim of this review is to give a comprehensive overview of the clinical, molecular genetic and pathophysiological aspects of each of the classic and new overgrowth syndromes.

Introduction

Longitudinal growth is a complex process which is influenced pre- and postnatally by an interaction of genetic, endocrine, nutritional, environmental and socioeconomic factors (1). Due to the complexity to attain normal height, usually disturbance of even a single factor results in a delayed growth velocity and short stature. Less common are conditions causing increased height. However, from a patient's and physician's perspective concerning diagnosis, prognosis, counselling and therapy, tall stature, either as the only symptom or as part of an overgrowth syndrome, is not less important. In the last two decades, the causative genetic defects in the classic overgrowth syndromes such as Beckwith-Wiedemann, Simpson-Golabi-Behmel and Sotos syndrome have been identified (2-4). These discoveries have resulted in the delineation of the phenotypic spectrum and genotype-phenotype correlations. They also stimulated and still stimulate further research in patients with either characteristic or non-characteristic manifestations but without a confirmed genetic defect. A successful example of these investigations is the identification of transforming growth factor beta receptor (*TGFBR1* and *TGFBR2)* mutations; originally detected in patients with Marfan-like syndromes and subsequently also in classic Marfan syndrome patients without a fibrillin 1 (*FBN1)* mutation (5,6). Furthermore, genes implicated in overgrowth syndromes resulted in the identification of reciprocal short stature syndromes; hypomethylation of an imprinting center region at 11p15 was identified as a cause of Silver-Russell syndrome, while hypermethylation of the same region is responsible for 2-7% of patients with Beckwith-Wiedemann syndrome (7,8). In accordance with this, other reports of patients with gene alterations leading to an opposite growth phenotype have emerged recently, for example a missense mutation of the fibroblast growth factor receptor 3 (*FGFR3)* gene is thought to cause a tall stature syndrome (9) while activating mutations of *FGFR3* are well-known to be responsible for several growth deficient disorders such as hypochondroplasia and achondroplasia (10). Furthermore, a duplication of the insulin-like growth factor 1 receptor (*IGF1R)* is associated with tall stature while haploinsufficiency results in short stature (11).

With regard to the recent progress, this review gives a comprehensive overview of the clinical, molecular genetic and pathophysiological aspects of the classic overgrowth syndromes and new overgrowth disorders. The syndromes are selected based on the fact that during the process of diagnosis they should be considered and hence distinguished from each other. Although in the past classificatory schemes have been postulated (reviewed in (12)), these

seem nowadays artificial by absence of good grouping definitions other than the common classification "overgrowth syndrome". Endocrine disorders and skeletal dysplasias with an overgrowth component will not be discussed here.

Diagnosis of overgrowth syndromes

The starting point for diagnostic evaluation of an overgrowth problem is usually tall stature, which is defined as a height of $> +2$ standard deviation score (SDS; i.e. ~98th percentile) above the mean height for that age, corrected for sex and ethnicity. It is important also to suspect children growing outside their target height range (> +2 SDS of the mid-parental height corrected for gender- and secular trend) of having a growth disorder. Furthermore, recent growth acceleration warrants further diagnostic analysis as well, with specific attention for endocrine disorders with tall stature (reviewed in (13)). However, it should be noted that although tall stature is used as a common denominator, many patients of the so called overgrowth syndromes might not fulfil the $> + 2$ SDS criterion. Therefore we propose that typical dysmorphic features either with or without tall stature should be a second starting point for diagnostic consideration of the overgrowth syndromes discussed in this review. A diagnostic flow chart is shown in Figure 1 and the respective diagnostic tests for the overgrowth disorders are shown in Table 1. Detailed recording and description of the dysmorphic features will be indispensable to discern the correct clinical diagnosis. Afterwards, molecular confirmation of a defect of the involved gene should be undertaken. In groups belonging to the same molecular pathways, for example the Marfan-related syndromes, a stepwise screening of the candidate genes known in this pathway is recommended. In other overgrowth syndromes, routine molecular screening of multiple genes implicated in overgrowth syndromes cannot be advised based on the very low and often zero detection rate. Therefore, molecular testing for different genes should be reserved for patients with overlapping features of different overgrowth syndromes.

Table 1. Overgrowth causing disorders and their respective diagnostic tests

 1 Order of disorders is according the diagnostic flow chart presented in Figure 1.

Abbreviations: 17OHP: 17 hydroxyprogesterone; ACTH: adrenocorticotropic hormone; BA: bone age; BMI: body mass index; DHEAS: dehydroepiandrosterone sulphate; FSH: follicle-stimulating hormone; GH: Growth Hormone; GnRH: gonadotropin releasing hormone; IGF-I: Insulin-like Growth Factor I; IGFBP-3: Insulin-like Growth Factor Binding Protein 3; LH: luteinizing hormone; TSH: thyroid-stimulating hormone

Classic syndromes

Marfan syndrome

Marfan syndrome (MFS; OMIM 154700) is an autosomal dominant connective tissue disorder with an estimated incidence of 2-3 per 10.000 individuals (16). It is caused by pathogenic mutations of the fibrillin 1 (*FBN1*) gene at 15q21.1 (17) and approximately 25% of the mutations occur *de novo* (16). In MFS multi-organ systems are affected with manifestations primarily in the skeletal (Marfan habitus), cardiovascular (aortic root dilatation and mitral valve prolaps and regurgitation) and ocular systems (ectopia lentis). Also the skin/integument, lung or dura can be affected (18). The characteristic Marfan habitus includes a tall disproportionate stature with long slender limbs (dolichostenomelia), arachnodactyly with a positive thumb and wrist sign and pectus excavatum or carinatum (19). Typical craniofacial features are a long and narrow face, downslant of the palpebral fissures, enophtalmos, malar hypoplasia, micrognathia or retrognathia and a high-arched palate (16). Other prominent manifestations are scoliosis, pes planus, joint laxity and lumbosacral dural ectasia (20). Acute dissection of the ascending aorta due to progressive dilatation is the major cause of mortality in MFS. The clinical diagnosis is made on the criteria defined by the Ghent nosology (Table 2) (18). For the diagnosis of MFS, a major criterion in two different organ systems and involvement of a third is required. Since the occurrence of certain Marfan features such as aortic dilation and ectopia lentis are age-related, caution is warranted to use this nosology for the exclusion of MFS in patients younger than 18 years (19). Furthermore, the phenotypic expression can vary greatly between affected family members or between affected members of different families.

Although height itself is not included in the Ghent nosology, the mean birth length for boys and girls with clinically MFS is near the $90th$ percentile (21). Boys then continue to grow approximately between the 50th en 95th percentiles for the first three years of life. Girls show similar increased body length in the first year of life with a further increase along the $95th$ percentile between 1 and 3 years of age. Height of boys and girls after three years of age till adulthood is consistently above the $95th$ percentile of the normal population. Furthermore, the peak of the pubertal growth spurt is advanced on average with 2.4 and 2.2 years for boys and girls, respectively (21).

Table 2. Diagnostic criteria of Marfan syndrome and frequencies of common features with confirmed *FBN1* **alterations**

Clinical diagnosis of MFS requires a major criterion in two systems and involvement of a third

Table 2. *(continued)*

Table 2 was adapted from Faivre et al. (20) with permission from BMJ Publishing Group Ltd.Diagnostic criteria

are according the Ghent nosology (18)

1 n.r. : not reported

The detection rate of *FBN1* mutations varies widely from 9-90%, depending on the type of screening method employed as well as on the criteria used (reviewed in (22)). A strong association between the Ghent nosology and the rate of *FBN1* mutations is shown by two large studies which detected *FBN1* mutations in 51-66% in patients fulfilling the Gent criteria versus 12% not meeting the diagnostic criteria (23,24). *FBN1* mutations consist mainly of missense mutations (~56%) and the majority of the missense mutations are located in the

calcium-binding epidermal growth factor domains (cb-EGF; in 74%) or in the transforming growth factor β binding protein-like domain (TB; in 15%) (25). Approximately 60% of the missense mutations substitute or create a cysteine residue and consequentially affect disulfide bonds and the tertiary protein structure (25). Mutations in *FBN1* have been also found in a broad group of disorders including for example isolated ectopia lentis, Shprintzen-Goldberg craniosynostosis, familial thoracic aortic aneurysms and dissections (TAAD) and autosomal dominant Weill-Marchesani syndrome (reviewed in (22)).

Until recently, no strong genotype-phenotype correlation had been established in MFS, except for the association of mutations in exons 24-32 with a neonatal form of MFS (26). This neonatal presentation is more severe with additional manifestations such as flexion contractures, pulmonary emphysema and loose skin and most patients do no survive 2 years of life. A recent large study of 1013 *FBN1* mutation carriers showed that mutations in exons 24-32 were also correlated with a more severe phenotype in MFS patients with a younger age at diagnosis and shorter survival (25). Furthermore, a higher probability to develop ectopia lentis, ascending aortic root dilatation, mitral valve defects and scoliosis was found for this region. In the same study, missense mutations affecting a cysteine residue were correlated with a higher probability of ectopia lentis compared to other missense mutations.

FBN1 encodes for a large extracellular matrix (ECM) protein and this monomer fibrillin-1 polymerizes in bundles of microfibrils. In interaction with other ECM proteins these bundles form an assembly and give elasticity to the connective tissues (27). Although the exact mechanisms are not clear, the loss of connective tissue integrity in MFS could be explained by either a reduced incorporation of fibrillin-1 in the microfibrils or that after incorporation of mutated fibrillins an increased loss of tissue, possibly due to proteolysis, would occur. The molecular biology of fibrillin and fibrillinopathies has been extensively reviewed elsewhere (22). However, the loss of connective tissue integrity would not provide enough support for manifestations such as increased statural growth and other skeletal features. Interestingly, an interaction between fibrillin and the latent TGFβ binding protein 1 (LTBP1) has been reported and a model was proposed in which LTBP1 by sequestering latent TGFβ complexes regulates TGFβ bioactivity in the ECM (28). This involvement of TGFβ signaling is further supported by the detection of TGFβ receptor 2 *(TGFBR2)* mutations as the cause of MFS type II (29) (see further). In addition, *TGFBR1* and *TGFBR2* mutations have been identified in individuals fullfiling the Ghent criteria (5,6), emphasizing the shared underlying pathogenic

mechanism. However conflicting data exists, showing a decreased signaling activity for mutations found in MFS type 2 (29) while increased signaling was found in the related Loeys-Dietz syndrome (LDS) (30) and in the lungs of *Fbn1* deficient mice (31). Nevertheless, from *in vitro* studies the role of TGFβ in chondrogenesis has been well established (32) and perturbation of this signaling pathway is likely to contribute to the skeletal manifestations in MFS.

Marfan syndrome type II / Loeys-Dietz syndrome

Marfan syndrome type II (MFS2 ; OMIM 154705) is a dominant autosomal disorder and was first described in a large French family (33). Similar involvement as in Marfan syndrome was found for the skeletal system (tall stature, increased arm span, arachnodactyly and chest deformities, scoliosis) and the cardiovascular system (aortic root dilatation, aortic dissection or rupture, mitral valve prolapse), but without involvement of the ocular system.

A cosegregating loss-of-function missense mutation of *TGFBR2* at 3p24.1 was identified as the cause in this family (29) and *TGBR2* mutations were detected in several other patients with features of Marfan-syndrome but without major ocular involvement (5,6,34,35). However, recently one member of the original French family was reported to have ectopia lentis (36). Furthermore, *TGFBR2* and also *TGFBR1* mutations were found in LDS (type I and II) which is characterised by arterial tortuosity and aneurysms, hypertelorism, bifid uvula and cleft palate (30,37) and in a spectrum of other disorders (36). Whether MFS2 should be regarded as the phenotypic spectrum of classic MFS or of LDS remains a topic for discussion (29,37). With regard to molecular diagnosis, it is important to consider *TGFBR2* and *TGFBR1* analysis in patients with features of MFS without *FBN1* abnormalities.

TGFBR2 encodes a transmembrane receptor with an intracellular kinase domain. Primarily missense mutations are detected which cluster in or close to the serine-threonine kinase domain and alter TGFβ signaling (29,30,37). For discussion on TGFβ signaling we refer to MFS.

Congenital contractural arachnodactyly / Beals syndrome

Congenital contractural arachnodactyly or Beals syndrome (CCA; OMIM 121050) is an autosomal dominantly inherited disorder with overlapping manifestations with MFS. CCA is caused by abnormalities of the fibrillin 2 (*FBN2*) gene at 5q23.3 (38). Characteristic features include multiple congenital contractures (knees, elbows, fingers), arachnodactyly, tall stature with dolichostenomelia, severe kyphoscoliosis, muscular hypoplasia and crumpled ears (22). Although initially aortic involvement was thought to be absent in CCA, aortic root dilatation has been reported in four patients (39).

Mutations in *FBN2* cluster in exons 24 through 34 which is associated in *FBN1* with neonatal MFS and a more severe phenotype (see MFS). Mutations are primarily located in the cbEGF domains or are affecting splicing (39). Although the pathogenesis is thought to be in general similar to *FBN1* defects, differences in spatial and temporal gene expression for *FBN1* and *FBN2*, which might underlie the different phenotypic manifestations, have been shown (22).

Homocystinuria

Homocystinuria (OMIM 236200) is an autosomal recessive disorder of sulfur amino acid metabolism and is caused by a cystathionine β-synthase (CBS) deficiency due to mutations of the CBS gene at 21q22.3 (40). The clinical features include skeletal (Marfanoid habitus, osteoporosis and scoliosis), ocular (ectopia lentis and myopia), vascular manifestations (thromboembolism, malar flush and livido reticularis) and manifestations affecting the central nervous system (mental retardation and psychiatric disorders) (41). Height is at or above the 95th percentile in 50% of the patients (42). Extremely high plasma levels of total homocysteine and high levels of methionine are measured with decreased levels of cystathionine and cysteine (40). Treatment options include pyridoxine (vitamin B6), betaine and restriction of dietary methionine. In a small study group, a correlation between growth and free homcysteine levels was found with a mean height SDS of -0.01 SDS \pm 0.81 in the optimally treated group and $+1.73$ SDS \pm 0.88 in the suboptimally treated group (43).

Interestingly, an interaction between FBN1 and homocysteine was detected, in which homocysteine reduces the disulfide bonds of the cbEGF domains of FBN1 and consequentially modifies protein folding and increases proteolytic degradation of the protein (44,45). This interaction would explain why in homocystinuria patients, manifestations similar to Marfan syndrome are part of the phenotype.

Lujan-Fryns syndrome

Lujan-Fryns syndrome (LFS ; OMIM 309520) is a rare X-linked mental retardation syndrome which is characterised by a marfanoid habitus becoming evident after puberty (tall stature, long thin hyper-extensible fingers and toes), typical craniofacial dysmorphism (a long, narrow face, prominent forehead, maxillary hypoplasia, a small mandible) and behavioural problems (46). Recently mutations in the *MED12* gene and in the *UPF3B* gene were found as the cause of Lujan-Fryns syndrome (47,48). Furthermore, mutations in the *ZDHHC9* gene cause another X-linked mental retardation syndrome with a marfanoid habitus and should be considered in the differential diagnosis of LFS (49).

Sotos syndrome

Sotos syndrome (SoS; OMIM 117550) is an autosomal dominant disorder and has an estimated incidence of 1 in 15.000 newborns (50). It is caused by haploinsufficiency of the nuclear receptor binding SET domain protein 1 (*NSD1*) at 5q35.2-35.3 (4). The cardinal features (i.e. ≥ 90% of the patients) for the diagnosis of SoS are characteristic facial features, overgrowth (height and/or head circumference ≥ 98 th percentile) and a certain degree of learning disability (51). The characteristic Sotos craniofacial features include a triangular shaped ("inverted pear-like") face with a prominent chin, macrodolichocephaly, frontal bossing with a high hairline, (apparent) hypertelorism and downslanting of the palpebral fissures. These facial features are probably the most consistent diagnostic criterion (52), but become less apparent in adolescence and adulthood. SoS is furthermore associated with a large variety of additional features such as advanced bone age, scoliosis, seizures and neonatal problems (Table 3) (51-53). Cardiac and genitourinary anomalies are also frequently reported. The overall tumor incidence in patients with an *NSD1* abnormality is ~2% with an increased incidence of neural crest tumors and sacrococcygeal teratomas (54). Although the risk for malignancy in SoS is very low, some patients carrying an *NSD1* alteration have been described with acute lymphoblastic leukaemia, T-cell lymphoma, neuroblastoma, hepatoblastoma and small-cell lung cancer (52,54).

The growth pattern shows an accelerated growth, which starts pre- or postnatally and is especially increased in the early years of childhood. The final adult height however is found to be within the (high) normal range (53). Although overgrowth is a cardinal feature, children carrying a pathogenic *NSD1* mutation with normal heights and normal head circumference

Table 3. Clinical diagnostic criteria and common features in Sotos syndrome with confirmed *NSD1* **abnormalities**

¹ Major features were adapted from (51-53)

2 n.r.: not reported

have been described (51,52). The degree of learning disability varies widely, from mild to severe.

Pathogenic alterations of *NSD1* are found in approximately 60-90% of the patients depending on the stringency of the inclusion criteria used (reviewed in (55)). Most of the alterations occur *de novo*, but familial cases with autosomal dominant inheritance have also been described. The main causes are intragenic point mutations (~80-85%) , whole-gene microdeletions (~10%) and exon-deletions (~5%) (55). However, based on ethnicity this spectrum can be different. In the Japanese population a common 1.9 Mb-microdeletion encompassing *NSD1* and neighbouring genes is the main cause and is detected in ~50% of the patients while intragenic point mutations only account for ~10% (56,57). As an explanation,

we have suggested that a genomic inversion polymorphism increases the susceptibility to microdeletions in the Japanese population (57). Mutations resulting in protein truncation are found throughout the *NSD1* gene without specific hotspot locations. In contrast, missense mutations are preferentially located in the functional domains of *NSD1* (reviewed in *(55)*). Statistically convincing data are lacking but a correlation between a milder phenotype and missense mutations compared with truncating mutations has been suggested (52). This would possibly explain the preferential detection of missense mutations in familial cases (51). Although SoS is primarily caused by a reduced level of proper functioning NSD1, a correlation for a more severe level of mental impairment and smaller height has been found for patients harboring a microdeletion in comparison with patients carrying a pointmutation (51,58). Furthermore, anomalies of the cardiovascular and genitourinary systems seem correlated with microdeletions (52,58).

NSD1 alterations have also been detected in 6 Weaver syndrome patients (59,60), in two patients with Beckwith-Wiedemann (61) and in one patient with Nevo syndrome (62). Without additional patients reported so far, it seems likely that these patients should be considered as having overlapping phenotypic features, rather than *NSD1* alterations being responsible for a subset of patients with these respective syndromes.

NSD1 was identified as interacting with nuclear hormone receptors (retinoic receptor, thyroid receptor, retinoid X and estrogen receptors) through its nuclear receptor interaction domains (63). It was postulated that NSD1 could interact both as a co-repressor and coactivator of nuclear hormone receptors and would therefore be a bifunctional transcriptional regulator (63). Additionally, NSD1 is able to control chromatin transcription through the histone methyltransferase activity of its SET domain (Su(var) 3-9, Enhancer of zeste, Trithorax domain) (64) and through its interaction with the NIZP1 protein (65). In this function as a transcriptional regulator it can be hypothesized that NSD1 reduces the transcription of growth promoting genes and loss of this activity would consequentially result in overgrowth (4). Unfortunately no further evidence for the involvement of NSD1 in growth regulation could be derived from mouse models since heterozygous *Nsd1* mutant mice did not express an overgrowth phenotype and homozygous *Nsd1* deficient mice died during embryogenesis (64). We detected endocrine and paracrine changes in the IGF-system between SoS patients with a confirmed *NSD1* defect and controls, although the actual contribution of these differences to overgrowth is unclear (66).

Weaver syndrome

Weaver syndrome (OMIM 277590) is a rare disorder with less than hundred patients described in the literature. It is characterized by pre- or postnatal overgrowth, typical craniofacial features, developmental delay, a hoarse low-pitched cry, advanced bone maturation and finger- and nail abnormalities such as camptodactyly and deep set nails (67-70). The craniofacial features include macrocephaly, flat occiput, hypertelorism, micrognathia, long and prominent philtrum and large ears. There is a phenotypical overlap with Sotos syndrome and six Weaver syndrome patients have indeed been reported to carry an *NSD1* point mutation (59,60),although two of them were later reclassified as typical Sotos and one as Sotos-like (51). In 16 additional patients no alterations were detected (59,60,71- 73) and it can be questioned whether *NSD1* abnormalities are the cause of classical Weaver syndrome (51). Furthermore, no *RNF135* (see further) abnormalities were found in classical Weaver syndrome patients.

Nevo syndrome / Ehlers-Danlos type VIA

Nevo syndrome (OMIM 601451) is a rare autosomal recessive disorder and is characterized by an increased perinatal length, kyphoscoliosis, talipes calcaneovalgus, generalized hypotonia, volar edema and spindle shaped fingers (74-76). In 2005, a homozygous *PLOD1* mutation (p.R319X) was found in 6 patients from Arab ancestry and one Dutch patient carried a homozygous deletion of exon 17 (77). Deficiency of the procollagen-lysine 1, 2-

1 Revised nosology, Villefranche 1997 (78)

oxoglutarate 5-dioxygenase 1 (*PLOD1)* gene was previously described in the kyphoscoliotic type of Ehlers-Danlos syndrome (EDS VIA; OMIM 225400) which is characterized by severe muscular hypotonia present at birth, progressive kyphoscoliosis, joint hypermobility, scleral fragility and an elevated ratio of total lysyl pyridinoline to hydroxylysyl pyridinoline in the urine (Table 4) (78). It was therefore concluded that Nevo syndrome and EDS VIA were a single entity (77).

Nevo syndrome was originally regarded as an overgrowth syndrome, but this remains equivocal since height information of patients with confirmed *PLOD1* abnormalities is scarce. Four patients were reported to have birth lengths $> 90th$ percentile, three between the 50th-90th percentile and one at the 10th percentile (77,79,80). Follow-up height in infancy or childhood in 7 of these patients varied between the $25th$ -90th percentile, while two patients had a height >90th percentile. However, progressive kyphoscoliosis might have influenced follow-up height measurements.

PLOD1 is located on chromosome 1p36.22 and encodes for a protein which hydroxylates specific lysyl residues in collagen proteins (81). These residues are attachment sites for carbohydrates and are essential for the formation of collagen cross-links. Deficiency of PLOD1 results in a weakened cross-linking formation and consequentially an impaired biomechanical instability of the connective tissues (79). However, also since *Plod1 * mice did not show overgrowth (82), it is not known how PLOD1 would be involved in longitudinal growth.

Beckwith-Wiedemann syndrome

Beckwith-Wiedemann syndrome (BWS; OMIM 130650) is characterised by three cardinal features: neonatal macrosomia or postnatal overgrowth, abdominal wall defects and macroglossia. Additional features include ear abnormalities (earlobe creases or posterior helical pits), neonatal hypoglycaemia, nevus flammeus, hemihypertrophy, organomegaly, polyhydramnios, midfacial hypoplasia, cardiomyopathy and embryonal tumors (83,84). Although there are no consensus diagnostic criteria, in general the diagnosis can be made if three of the cardinal features are present or two cardinal and one of the additional features (Table 5) (85). BWS has an estimated incidence of approximately 1 in 13700 newborns (86). Approximately 85% of the patients with BWS are sporadic while 10-15% show an autosomal inherited pattern with a preferential maternal transmission (8).

Table 5. Clinical diagnostic criteria in BWS and common features

The diagnosis is made if three of the sentinel features are present¹ . If less than three are present the additional features may support the diagnosis.

1 Clinical diagnostic criteria are according to (85). No consensus clinical criteria exist. Cohen (84) suggested at least the combination of macrosomia, macroglossia and abdominal wall defects or two of those in combination with hypoglycaemia, hemihyperplasia, ear anomalies, midface hypoplasia, nevus flammeus, cardiomegaly and enlarged placenta/long umbilical cord and/or polyhydramnios, advanced bone age and tumors.

2 Common features are adapted from (83,84,86,87) and references therein. Percentages include data from clinical studies (83,84,86,87) without molecular confirmation.

3 Only 9 patients reported in (86)

4 n.r. not reported

Increased infant mortality is caused by hypoglycaemia, respiratory or feeding problems due to macroglossia, prematurity or cardiovascular problems. Usually there is a normal mental development in BWS, although moderate to severe developmental delay has been reported in 4% (83). There is an estimated tumor risk of 5-10% with a predisposition for embryonal tumors, specifically Wilms tumor (54).

In a review of 134 clinically diagnosed patients, the average birth length and weight for boys was above the 95th percentile and height was at or above the 95th percentile throughout adolescence, parallel to the growth curve of the normal population (87). Weight followed the 95th percentile, but weight follow-up data was only available till three years of age. For girls, average birth length and weight was at the $75th$ percentile and increased towards the 95th percentile at 18 months of age. The statural growth throughout adolescence was similar to the males, while weight data were only available till nine years of age. There is an advanced bone maturation especially during the first 4 years of life and adult height is supposed to be within the normal range (84).

BWS is caused by abnormalities, either cytogenetic, molecular genetic or epigenetic, of a cluster of imprinted genes on 11p15 (reviewed in (8,88)). This cluster can be divided into two domains. The telomeric domain 1 contains the genes *H19* and insulin-like growth factor 2 (*IGF2*) and is regulated by the imprinting center region 1 (ICR1 or DMR1; differentially methylated region 1). The centromeric domain 2 contains *CDKN1C*, *KCNQ1* and *KCNQ1OT1* as the most important genes and is regulated by imprinting center region 2 (ICR2 or DMR2). In a normal situation, these imprinting centres are differentially methylated which means that instead of bi-allelic expression either the genes from the paternal allele (*IGF2*, *KCNQ1OT1*) or from the maternal allele (*H19*, *CDKN1C* and *KCNQ1*) are expressed and that the genes on the opposite allele are silenced. In general, disturbance of this differentially regulated gene-expression pattern is the cause of BWS. In sporadic BWS patients, loss of methylation at the DMR2 accounts for 50-60%, intragenic-point mutations of *CDKN1C* for 5-10% and hypermethylation of DMR1 and *H19* for 2-7% of the abnormalities detected (reviewed in (8,88)). In approximately 20% of the sporadic patients, paternal uniparental disomy (UPD) with loss of the maternal allele is found. In autosomal dominant pedigrees, intragenic-point mutations of *CDKN1C* are found in ~30-50% of the patients. Less frequent causes, either sporadic and/or inherited, include balanced chromosomal translocations, duplications or inversions involving 11p15 and microdeletions of DMR1 and DMR2. In 10-15% of the

patients the etiology is unknown. Recently, constitutional DMR2 defects were shown to be also responsible for 3% of patients with sporadic Wilms tumor but without features of BWS (89). This suggests that defects in this region might result in a broad phenotypic spectrum.

Interestingly, hypomethylation of DMR1 has been found to be the cause of Silver-Russell syndrome (7). This syndrome is characterised by intrauterine and postnatal growth retardation, dysmorphic facial features and frequent body asymmetry, an opposite phenotype of the BWS (90). No abnormalities, except for a maternal duplication in a single patient (91), have been detected so far in the centromeric 11p15 domain.

In BWS, genotype/epigenotype-phenotype correlations show a strong association of UPD with hemihypertrophy (92), which is probably due to that fact that many cases of UPD display somatic mosaicism. Different associations can be found for the two imprinting centers with exomphalos being associated with an DMR2 alteration or a *CDKN1C* defect (92,93), a higher birth weight with DMR1 defects (92) and a higher risk of tumors for patients with DMR1 alterations and UPD patients (92,94,95). Although there is a lower risk, in patients with DMR2 defects tumors such as hepatoblastoma and rhabdomyosarcoma have still been reported (54). On the other hand, the risk for Wilms tumor for patients with an DMR2 defect seems very low and in fact no such patients have yet been reported (54,92).

Originally increased expression of *IGF2* was thought to be the cause of the BWS phenotype (2,96). This was supported by transgenic mice overexpressing *Igf2* which showed prenatal and postnatal overgrowth, macroglossia, polyhydramnios and organomegaly (97). In addition, loss of imprinting and increased *IGF2* expression was found in Wilms tumor (98). However, significant biallelic expression of *IGF2* was also observed in the leukocytes of 10% of normal individuals without a BWS phenotype (99) and detection of abnormalities in BWS patients at DMR2 and in *CDKN1C* called for a more complex explanatory mechanism. Evidence came from a mouse model with a null-mutation in *CDKN1C* and loss of imprinting of *Igf2*, which showed features reminiscent of BWS (100), while mice carrying *CDKN1C* mutations alone exhibited omphalocele and kidney dysplasia but no macrosomia (101). Additional support for an interaction model can be derived from the data that increased *IGF2* expression causes decreased *CDKN1C* expression both *in vitro* and *in vivo* (102). Therefore, both genes seem to act in a concerted fashion influencing the same biochemical pathway and perturbation of this signaling cascade is likely to be the cause of BWS.

Simpson-Golabi-Behmel syndrome

Simpson-Golabi-Behmel syndrome (SGBS; OMIM 312870) is an X-linked condition and is caused by defects of the glypican 3 (*GPC3*) gene at Xq26.2 (3). Characteristic features are pre- and postnatal overgrowth, a "coarse" facial appearance (short nose with anteverted nares, low set posteriorly rotated ears, hypertelorism, downslanting of the palpebral fissures and epicanthic folds), supernumerary nipples and congenital heart defects (Table 6). Other common features include skeletal, hand and renal tract anomalies, macroglossia, midline groove of the lower lip and or tongue, macrognathia, cleft palate and inguinal and umbilical hernias (103-105). There is a predisposition for embryonal tumors especially Wilms tumor and furthermore hepatoblastoma and nephroblastomatosis (104). Although speech and fine and/or gross motor delay are commonly found, mental development can be normal in SGBS-patients (103). The phenotypic spectrum varies from very mild in female carriers to early lethal forms in affected boys (106). Due to the neonatal macrosomia, macroglossia, abdominal wall defects and a predisposition for Wilms tumor, BWS is usually considered in the differential diagnosis although the coarse facial features, supernumerary nipples, cardiac defects, polydactyly and an X-linked inheritance pattern are more typical of SGBS (106).

The mutational spectrum of GPC3 abnormalities includes microdeletions affecting a variable number of exons (most frequently), deletions of the whole gene and intragenic point mutations (3,104,107). In roughly 40-70% of the typical SGBS patients alterations of *GPC3* were identified (104,108) and no genotype-phenotype correlation was observed, which leaves nonfunctional GPC3 as the most likely cause of SGBS (107). In contrast, the causative role of GPC3 in Wilms tumor is less clear because no abrogating alterations of *GPC3* were detected (109) and even an increased expression of *GPC3* was found (110). A second SGBS locus was mapped to ~6Mb region on chromosome Xp22 in a family expressing a more severe from of SGBS, so called SGBS type 2 (111). Recently a novel X-linked mental retardation syndrome including macrocephaly and ciliary dysfunction was described by Budney et al. which was caused by mutation in the *OFD1* gene (112). This gene is also responsible for the oral-facial-digital syndrome type 1 (113). Because of the overlapping phenotype of their families and the fact that *OFD1* is located in the 6Mb region on Xp22, Budney et al. speculated that *OFD1* might be the responsible gene in SGBS type 2 (112).

GPC3 is one of the six members of the glypican family, which is a group of heparan sulphate proteoglycans (HSPG). These proteins are involved in the regulation of heparin-

Table 6. Common features (>10%) in Simpson-Golabi-Behmel syndrome with confirmed *GPC3* **alterations**

¹ Major features are adapted from (103), (122) and references therein.

binding growth factors, such as Wnts and fibroblast growth factors (reviewed in (114)). It was demonstrated *in vitro* that GPC3 binds directly to IGF2 and sequestering of this ligand might inhibit its activity (3). Recently this direct interaction between GPC3 and IGF2 and additionally also with the IGF1R was shown again (115)*.* A model was proposed in which absent GPC3 would result in high level of IGF2 signaling, resulting in overgrowth in a fashion similar to BWS (3,116). This model was supported by double mutant mice overexpressing

Igf2 due to absence of the Igf receptor type 2 (*Igf2r*; a downregulator of Igf2) and the *H19* locus (117). These mice demonstrated BWS features but additionally also skeletal defects and cleft palate reminiscent of SGBS. In contrast, in a rat model the direct interaction between GPC3 and IGF2 could not be reproduced (118). In addition, Gpc3 mutant mice which did exhibit an increased body size, did not show elevated levels of circulating Igf2 or an increased expression in tissues (119) and overgrowth due to *Gpc3* deficiency was postulated to occur without directly influencing Igf2 signaling (120). It remains therefore unclear if and to what extent increased IGF2 signaling is responsible for SGBS. Alternative explanations such as the regulation by GPC3 of other heparin-binding growth factors (for example Wnts, fibroblast growth factors, bone morphogenetic proteins and hepatocyte growth factors) should be taken into consideration (105,114). Recently, deficient GPC3 was shown to cause upregulation of Hedgehog signaling and it was suggested that hyperactivation of this pathway plays a role in overgrowth in SGBS (121).

Bannayan-Riley-Ruvalcaba syndrome

Bannayan-Riley-Ruvalcaba syndrome (BRRS; OMIM 153480) is an autosomal dominant disorder of unknown incidence which is caused by haploinsufficiency of the phosphatase and tensin homolog (*PTEN*) gene at 10q23.31 (123,124). Diagnostic criteria were defined as at least two of the following features: macrocephaly, hamartomas (including at least one lipoma, haemangioma or intestinal polyp) and penile macules in males (125) or as at least three of the four following characteristics: macrocephaly, lipomatosis, haemangiomas and pigmented macules of the glans penis in males (126) (Table 7). The latter criteria were found to result in a more sex-biased diagnosis towards males (127). Other common detected features in *PTEN*-mutation positive patients include developmental delay/mental retardation, hypotonia, downslanting of the palpebral fissures, postnatal childhood overgrowth, a high arched palate and joint hypermobility (125,128) (Table 7). Mental retardation is reported in *PTEN*-mutation positive patients in 93-100% (125,128). However, this might be an overestimation due to ascertainment bias as two recent studies showed learning difficulties in only 2 of 17 non-proband patients (12%) and mental retardation or global development delay in 3 of 26 patients (12%), respectively (127,129).

Abnormalities of *PTEN* are detected in approximately 57-60% of the BRRS patients (126,130,131). In the majority, intragenic point-mutations are found, while only a few patients have been described carrying exonic or whole *PTEN*-gene deletions (132). *PTEN*

Table 7. Clinical diagnostic criteria and common features in BRRS

Clinical diagnostic criteria used by Parisi et al. (125)

At least two of the following features:

Macrocephaly

Hamartomas (including at least one lipoma, haemangioma or intestinal polyp) Penile macules in males

Clinical diagnostic criteria used by Marsh et al. (126)

At least three of the four following characteristics:

Macrocephaly

Lipomatosis

Haemangiomas

Pigmented maculae of the glans penis in males

Common features of BRRS with a confirmed *PTEN* **mutation, adapted from (128) and references therein**

abnormalities are also responsible for ~80% of Cowden syndrome syndrome (CS) patients (131). This is an adult-onset hamartomatous disorder with pathognomic mucocutaneous lesions including trichilemmomas (133). There is an increased incidence of malignancy with lifetime risks of 3-10% for thyroid cancer, 25-50% for breast cancer and an unknown risk for endometrial cancer (reviewed in (134)). Because of the overlapping features between BRRS and CS, the fact that they are caused by alterations of the same gene and the lack of consistent genotype-phenotype correlations, it was suggested that BRRS and CS are the same entity with a variable expression and age-related penetrance (126,127). It was proposed that they should be classified as the "PTEN-hamartoma-tumour syndrome" (PHTS) (126). In relation to this and since a correlation was found with cancer or breast fibroadenoma in BRRS and BRRS/CS families (126), BRRS patient should undergo similar cancer surveillance as proposed for CS (133,135). *PTEN* aberrations are further reported in Proteus syndrome (~20%) and Proteus-like syndrome (~60%) (136) although in other Proteus(like) populations no *PTEN* mutations could be detected (137-139). Furthermore, somatic *PTEN*-alterations were found in a variety of sporadic neoplasias (reviewed in (135)).

The *PTEN*-gene encodes for a tumor suppressor protein which exerts phosphatase activity (i.e. removal of phosphate groups from macromolecules) targeting both proteins as well as lipids (reviewed in (140)). PTEN has an inhibitory function on the PI3K/Akt pathway and decreased signaling activity of this pathway usually limits proliferation and cell survival. Furthermore, PTEN dephosphorylates the focal adhesion factor (FAK) which is involved in inhibiting cell migration and spreading (141). Hence, loss of function of PTEN would consequentially promote cell growth, survival and migration. Most of the functional studies were performed to elucidate the role of PTEN in neoplasias and much less is known about its role in skeletogenesis. However, recently mice lacking *Pten* in their osteochondroprogenitor cells were reported (142). These mice displayed an increased skeletal size, especially enlargement of the vertebrae, and disorganised epiphyseal growth plates. Furthermore, there was a significant increase in the amount of trabecular and cortical bone. In a second report, mice deficient for *Pten* in their osteoblasts were of normal size but showed a highly increased bone mass (143). In both studies an increased PI3K/Akt signaling was demonstrated. These reports are very interesting because they provide the first direct evidence of the role of *PTEN* in skeletogenesis and create an explanatory basis for the macrocephaly and longitudinal overgrowth found in BRRS.

Fragile X syndrome

Fragile X syndrome (FXS; OMIM 300624) is an X-linked disorder and is caused by a silenced expression of the fragile X mental retardation 1 (*FMR1*) gene at Xq27.3. It has a prevalence of approximately 1 in 4000 - 9000 Caucasian males and 1 in 8000-9000 females (144), although a higher frequency of the full mutation allele (see further) of approximately 1 in 2500 individuals (males and females) is proposed (145). FXS is therefore the most common inherited cause of cognitive impairment. The degree of mental retardation is however variable; from mild learning difficulties with normal IQ to very severe mental impairment (146). The dysmorphic facial features can be subtle and include a narrow and elongated face, high forehead, prominent lower jaw and large protruding ears (Table 8) (147). Other physical features are macroorchidism and hyperextensibility of the joints (147). In general, the craniofacial features and macroorchidism become more outspoken in adolescence and adulthood. In males, the head circumference is consistently increased compared to the normal population and although height is close to the normal growth charts (148,149), childhood and preadolescent height and/or weight overgrowth has been associated with FXS (148,150,151). Adult height for males and females is lower than normal (148,151). Behavioural problems are frequent in FXS and include hyperactivity, impulsivity, sensory oversensitivity, tactile defensiveness, shyness, hand flapping and biting and autism spectrum disorders (147,152). In general, females are more mildly affected, probably due to X-inactivation. Diagnostic testing for FXS is advised in all individuals with mental retardation, developmental delay or autism, especially when other features of FXS are present or if there is a positive family history (153).

FXS is caused by hypermethylated CGG repeat expansions in the 5'UTR of the *FMR1* gene at Xq27.3 (154). These repeats co-localise with a constriction of the long arm seen in metaphases which is known as the fragile site at chromosome X (FRAXA) (155). In addition to transcriptional silencing due to repeat expansions, less frequently microdeletions of (parts of) *FMR1* and intragenic mutations have been described as well (reviewed in (156)). Nowadays, four allelic classes of repeat expansions can be distinguished: normal or common (6-44 repeats), intermediate (45-54 repeats), premutation (55-200 repeats) and full mutation (> 200 repeats) (146). The full mutation is hypermethylated and consequentially results in a silenced transcription. The premutation is non-methylated and is meiotically but also mitotically instable and can expand to larger repeats or to full mutations in the next generation (157). Full-mutation expansions were detected in the female germline and not in sperm cells,

Table 8. Common features of FXS with confirmed full mutation¹

¹ Common features are adapted from (149)

which would explain why expansion from premutation to full-mutation only occurs through female transmission (158). Although premutation carriers do not exhibit a characteristic FXS phenotype, mild physical manifestations have been reported and neurocognitive and behavioural functioning might be affected (reviewed in (159)). Furthermore, premutations are associated with premature ovarian failure and with Fragile-X Tremor Ataxia Syndrome (FXTAS) (159). This is a late-onset neurological disorder characterized by cerebellar ataxia, intention tremor and a progressive cognitive decline.

FMR1 is widely expressed, especially in the brain and the testis (160). The protein (FMRP) exhibits selective RNA binding capacity and controls local protein production by inhibiting translation of mRNA (reviewed in (161)). In addition, the translational control by FMRP might be mediated by influencing a microRNA pathway, although this needs further investigation (161). The suppression of local protein synthesis takes place at the neuron dendrites, which have been shown to display structural and numerical anomalies in FXS (162). One identified mechanism is that deficiency of FMRP results in an increased signaling of group I metabotropic receptors (mGluR) with an increased synaptic protein production and consequentially affecting synaptic plasticity and neuronal signal transmission (163). Since most of the functional research is focussing on the role of FMRP in mental retardation and neuronal development, the role in growth regulation is yet an unexplored field. Although Fmr1 deficient knockout mice showed macroorchidism and behavioural abnormalities similar to FXS, no other features were reported (164). Recently, evidence of a direct involvement of FMRP in skeletogenesis was shown by a zebrafish *fmr1* knockdown model in which mild craniofacial abnormalities were demonstrated to be caused by abnormal cartilage formation (165).

Klinefelter syndrome

Klinefelter syndrome (KS) is the most common disorder of sex chromosome aneuploidy with a prevalence of 1.09 - 1.72 per 1000 newborns (166). It is caused by an extra X chromosome which usually results in a 47,XXY karyotype. Other karyotypes observed include 48,XXXY; 48,XXYY; 49,XXXXY and 46,XY/47,XXY mosaicism. The classical pubertal KS phenotype is characterised by tall stature and features of androgen deficiency such as eunuchoidal body proportions with an increased arm span and long legs, sparse or absent pubic and axillary hair, decreased muscle mass, small testis and reduced penile length, infertility, a feminine distribution of adipose tissue and gynecomastia (Table 9) (167). A recent study identified

 1 Common features are adapted from (167), (168), (169), (183) and references therein.

clinodactyly, hypertelorism, elbow dysplasia, a high arched palate and hypotonia as additional frequent findings (168). Although birth length was reported to be less compared to controls (169), from around 2 years of age there is a tall stature with a mean height just below or at +1 SDS (168,170). Associations of KS with breast cancer, autoimmune and endocrine disorders, venous disease, osteoporosis and taurodontism have been observed (reviewed in (167)). The overall cognitive development is apparently normal (171), but delayed speech and language development are common findings in KS patients (169,171). Although there is no characteristic personality or behavioural phenotype, most KS boys were observed to be quiet, unassertive, passive, with an increased level of anxiety and a tendency of withdrawing from group activities (reviewed in (171)).

In early puberty, serum testosterone levels are normal, while at a later age in mid-pubertal patients levels are decreased with increased levels of LH, FSH and estradiol (172). These hormonal imbalances are addressed by testosterone replacement at the beginning of puberty, which improves the secondary sex characteristics, body proportions, bone mineral density and strength but does not improve fertility, testicular size or gynecomastia (173).

Nondisjunction during meiotic divisions in parental gametogenesis or during early mitotic cell divisons cause the extra X chromosome in KS. Paternal nondisjunction at meiosis I accounts for 53% of the cases, while maternal nondisjunction occurs either during meiosis I (34%) or meiosis II (9%) (174). Postzygotic disjunction errors during mitosis accounts for 3% of the cases (174). In some studies, a correlation was observed between the maternal age and nondisjunction at meiosis I (174-176) and between the paternal age and meiosis I (176), but no association with parental age was found in another study (177).

The exact molecular mechanisms underlying KS remain to be elucidated. On testicular biopsy of adult KS patients, fibrosis and hyalinization of the seminiferous tubules is found (reviewed in (178)). It is however not known whether testicular disfunction is due to intrinsic germ cell defects or that Sertoli cells are not able to support normal spermatogenesis (177,178). Furthermore, expressed phenotypic features such as height, gynaecomastia and smaller testes were reported to be regulated by the activity of the X-located androgen receptor (179). A positive correlation was found with a long CAGn repeat in this receptor, which is thought to result in decreased functionality of the receptor and consequentially in stronger effects of the androgen deficiency. However, only a correlation with penile length was found in another study (180). Because haploinsufficiency of the short stature homeobox (*SHOX*) gene is shown to be the cause of Leri-Weill syndrome, short stature in Turner syndrome and in a subset of patients with idiopathic short stature (181), a plausible explanation for the increased height in KS is the overdosage of *SHOX* in combination with hypogonadism (182).

New syndromes

Overexpression of the natriuretic peptide precursor C (*NPPC***) gene**

Recently, three patients were described each carrying a unique balanced translocation of the same 2q37.1 locus and chromosomes 7, 8 and 13, respectively (184,185). These patients presented with a similar phenotype of postnatal statural overgrowth $(>97th$ percentile), a marfanoid habitus, scoliosis, very long halluces and metaphyseal-epiphyseal dysplasia. All three translocation breakpoints localized in the vicinity of the *NPPC* gene, which encodes for the C-type natriuretic peptide (CNP). An increased expression of *NPPC* was found in fibroblasts, chondrocytes and lymphoblasts of these patients (184,185). It was proposed that the translocations caused a separation of *NPPC* from a negative regulatory element, hence resulting in overexpression (185). This finding is in line with transgenic mice overexpressing *NPPC* in growth-plate cartilage*,* which showed general skeletal overgrowth (186), while *Npcc* / mice were dwarfed (187). Chondrocytic overexpression of CNP was also shown to be able to rescue achondroplasia in mice with an activated fibroblast growth factor receptor 3 (*Fgfr3*) in their cartilage (188). Furthermore, homozygous loss-of-function mutations of the *NPR2*, the membrane receptor with the highest affinity for CNP, cause acromesomelic dysplasia type Maroteaux with a disproportionate short stature and radiographic skeletal changes (189). Heterozyogous *NPR2* mutations have been associated with proportionate, short stature (190). In general it can be concluded that CNP plays an important role in the regulation of endochondral bone growth and cartilage homeostasis (reviewed in (191)).

CATSHL syndrome

In a large pedigree of 27 living affected family members spanning four generations an autosomal dominant syndrome was identified which was characterised by *C*amptodactyly (18/20; 90%), *T*all stature (13/14 93%) and Sensoneurinal *H*earing Loss (17/20; 85%). (CATSHL; OMIM 610474) (9). Height was >97th percentile for 5 out of 5 affected males and 8 out of 9 affected females. Additional features were development delay (12/20; 60%), microcephaly, scoliosis and/or pectus excavatum. A heterozygous missense mutation (p.R621H) in the tyrosine kinase domain of the fibroblast growth factor receptor 3 (*FGFR3*) was found, which is thought to impair the signaling function of FGFR3 through a negative dominant mechanism (9). Furthermore, *Fgfr3⁻*/ mice showed a reminiscent skeletal phenotype in combination with deafness (192). In contrast, gain-of-function alterations of the *FGFR3* gene are found in several well-known growth failure disorders due to impaired endochondral bone formation

such as achondroplasia and hypochondroplasia (reviewed in (10)).

RNF135 **alterations**

Recently a new overgrowth disorder due to haploinsufficiency of the ring finger protein 135 (*RNF135*) (OMIM 611358) was reported in six patients (193). The alterations detected included four heterozygous truncating mutations, one missense mutation and a microdeletion including four neighbouring genes of *RNF135*. The patients showed a postnatal overgrowth phenotype with tall stature (height > +2.0 SDS) and macrocephaly (head circumference ≥ +2.0 SDS). The height of the single patient carrying a missense mutation was within normal range (+1.1 SDS). Dysmorphic characteristics included a broad forehead, anti-mongoloid slant of the eyes, a broad nasal tip, a long philtrum, a thin upper lip and a full lower lip. Furthermore, a spectrum of additional features was reported: for example advanced bone age (3/6), hearing problems (2/6) and eye abnormalities (2/6). A varying degree of developmental delay was present. The fathers or the mothers and also two siblings were carriers of the mutations as well. However, the dysmorphic features of these carriers were, except for macrocephaly, less distinctive and three of them showed a normal intellectual development.

RNF135 is located on 17q11.2 and and expression is found in several tissues (193). It is one of the 14 genes deleted in the common 1.4 Mb microdeletion, which causes neurofibromatosis type 1 (NF1) in approximately 5% of the cases (194,195). Due to the tall stature phenotype of these patients in comparison to patients with intragenic *NF1* mutations, *RNF135* was considered as a candidate gene responsible for this overgrowth (193,196). Furthermore, a Weaver syndrome-like phenotype was described in two familial patients carrying a deletion of the *NF1* region (197). However, investigations of *RNF135* mutations in classical Weaver syndrome patients did not yield positive results (193). Due to the phenotypic overlap with Sotos syndrome, a cohort of 160 patients referred for *NSD1* analysis on suspicion of Sotos syndrome features was investigated, but no *RNF135* abnormalities were detected (198). Additional patients harbouring pathogenic *RNF135* alterations are necessary for a further phenotype-delineation and also for elucidation of the very mild phenotype found in carrierparents.

Conclusion and future perspective

In this review we have given a comprehensive overview of the classical and new overgrowth disorders with a delineation of the clinical phenotype and the molecular genotype. Whether the new disorders will actually justify their position within the group of classic overgrowth syndromes, will largely depend on the discovery of additional patients. However, based on for example the unequivocal importance of proper CNP signaling in both under-and overgrowth, it is likely that additional patients with defects affecting the same pathway indeed will be identified. Furthermore, we expect that technological developments such as high resolution genome-wide array approaches targeting copy number variations and increased capacity of long range sequencing will result in the identification of additional defects of known genes as well as in the discovery of new genes. With regard to this, a combined approach with studies in the fields of genetics, genomics and proteomics are necessary to elucidate the underlying pathophysiological mechanisms. However, despite technological advances, thorough clinical assessment of a patient suspected of an overgrowth syndrome remains the first important step in the diagnostic process. This will not only increase the likelihood of attaining a molecular diagnosis for the patient, but will also create the basis for further research targeting new molecular defects in patients with certain features but without a confirmed molecular abnormality.

Acknowledgements

R.Visser was supported by grant number 920-03-325 from The Netherlands Organisation for Health Research and Development and our Sotos syndrome research was supported by Stinafo ("Dutch fund for disabled children").

References

- 1. Reiter EO, Rosenfeld RG. Normal and aberrant growth. In: Wilson JD, Foster DW, Kronenberg HM, Larsen P, eds. Williams Textbook of Endocrinology. Philadelphia: W.B. Saunders Company, 1998; 1427-1507
- 2. Ogawa O, Becroft DM, Morison IM, Eccles MR, Skeen JE, Mauger DC, et al. Constitutional relaxation of insulin-like growth factor II gene imprinting associated with Wilms' tumour and gigantism. Nat Genet 1993; 5: 408-412
- 3. Pilia G, Hughes-Benzie RM, MacKenzie A, Baybayan P, Chen EY, Huber R, et al. Mutations in GPC3, a glypican gene, cause the Simpson-Golabi-Behmel overgrowth syndrome. Nat Genet 1996; 12: 241-247
- 4. Kurotaki N, Imaizumi K, Harada N, Masuno M, Kondoh T, Nagai T, et al. Haploinsufficiency of NSD1 causes Sotos syndrome. Nat Genet 2002; 30: 365-366
- 5. Singh KK, Rommel K, Mishra A, Karck M, Haverich A, Schmidtke J, et al. TGFBR1 and TGFBR2 mutations in patients with features of Marfan syndrome and Loeys-Dietz syndrome. Hum Mutat 2006; 27: 770-777
- 6. Disabella E, Grasso M, Marziliano N, Ansaldi S, Lucchelli C, Porcu E, et al. Two novel and one known mutation of the TGFBR2 gene in Marfan syndrome not associated with FBN1 gene defects. Eur J Hum Genet 2006; 14: 34-38
- 7. Gicquel C, Rossignol S, Cabrol S, Houang M, Steunou V, Barbu V, et al. Epimutation of the telomeric imprinting center region on chromosome 11p15 in Silver-Russell syndrome. Nat Genet 2005; 37: 1003-1007
- 8. Weksberg R, Shuman C, Smith AC. Beckwith-Wiedemann syndrome. Am J Med Genet C Semin Med Genet 2005; 137C: 12-23
- 9. Toydemir RM, Brassington AE, Bayrak-Toydemir P, Krakowiak PA, Jorde LB, Whitby FG, et al. A novel mutation in FGFR3 causes camptodactyly, tall stature, and hearing loss (CATSHL) syndrome. Am J Hum Genet 2006; 79: 935-941
- 10. Ornitz DM, Marie PJ. FGF signaling pathways in endochondral and intramembranous bone development and human genetic disease. Genes Dev 2002; 16: 1446-1465
- 11. Kant SG, Kriek M, Walenkamp MJ, Hansson KB, van Rhijn A, Clayton-Smith J, et al. Tall stature and duplication of the insulin-like growth factor I receptor gene. Eur J Med Genet 2007; 50: 1-10
- 12. Cohen MM, Jr. Perspectives on overgrowth syndromes. Am J Med Genet 1998; 79: 234-237
- 13. Kant SG, Wit JM, Breuning MH. Genetic analysis of tall stature. Horm Res 2005; 64: 149-156
- 14. Drop SL, Greggio N, Cappa M, Bernasconi S. Current concepts in tall stature and overgrowth syndromes. J Pediatr Endocrinol Metab 2001; 14 Suppl 2: 975-984
- 15. Iughetti L, Bergomi A, Bernasconi S. Diagnostic approach and therapy of overgrowth and tall stature in childhood. Minerva Pediatr 2003; 55: 563-582
- 16. Ramirez F, Dietz HC. Marfan syndrome: from molecular pathogenesis to clinical treatment. Curr Opin Genet Dev 2007; 17: 252-258
- 17. Dietz HC, Cutting GR, Pyeritz RE, Maslen CL, Sakai LY, Corson GM, et al. Marfan syndrome caused by a recurrent de novo missense mutation in the fibrillin gene. Nature 1991; 352: 337-339
- 18. De Paepe A, Devereux RB, Dietz HC, Hennekam RC, Pyeritz RE. Revised diagnostic criteria for the Marfan syndrome. Am J Med Genet 1996; 62: 417-426
- 19. Dean JC. Marfan syndrome: clinical diagnosis and management. Eur J Hum Genet 2007; 15: 724-733
- 20. Faivre L, Collod-Beroud G, Child A, Callewaert B, Loeys BL, Binquet C, et al. Contribution of molecular analyses in diagnosing Marfan syndrome and type I fibrillinopathies: an international study of 1009 probands. J Med Genet 2008; 45: 384-390
- 21. Erkula G, Jones KB, Sponseller PD, Dietz HC, Pyeritz RE. Growth and maturation in Marfan syndrome. Am J Med Genet 2002; 109: 100-115
- 22. Robinson PN, Arteaga-Solis E, Baldock C, Collod-Beroud G, Booms P, De Paepe A, et al. The molecular genetics of Marfan syndrome and related disorders. J Med Genet 2006; 43: 769- 787
- 23. Loeys B, Nuytinck L, Delvaux I, De Bie S, De Paepe A. Genotype and phenotype analysis of 171 patients referred for molecular study of the fibrillin-1 gene FBN1 because of suspected Marfan syndrome. Arch Intern Med 2001; 161: 2447-2454
- 24. Katzke S, Booms P, Tiecke F, Palz M, Pletschacher A, Turkmen S, et al. TGGE screening of the entire FBN1 coding sequence in 126 individuals with marfan syndrome and related fibrillinopathies. Hum Mutat 2002; 20: 197-208
- 25. Faivre L, Collod-Beroud G, Loeys BL, Child A, Binquet C, Gautier E, et al. Effect of mutation type and location on clinical outcome in 1,013 probands with Marfan syndrome or related phenotypes and FBN1 mutations: an international study. Am J Hum Genet 2007; 81: 454-466
- 26. Robinson PN, Booms P, Katzke S, Ladewig M, Neumann L, Palz M, et al. Mutations of FBN1 and genotype-phenotype correlations in Marfan syndrome and related fibrillinopathies. Hum Mutat 2002; 20: 153-161
- 27. Kielty CM, Sherratt MJ, Marson A, Baldock C. Fibrillin microfibrils. Adv Protein Chem 2005; 70: 405-436
- 28. Isogai Z, Ono RN, Ushiro S, Keene DR, Chen Y, Mazzieri R, et al. Latent transforming growth factor beta-binding protein 1 interacts with fibrillin and is a microfibril-associated protein. J Biol Chem 2003; 278: 2750-2757
- 29. Mizuguchi T, Collod-Beroud G, Akiyama T, Abifadel M, Harada N, Morisaki T, et al. Heterozygous TGFBR2 mutations in Marfan syndrome. Nat Genet 2004; 36: 855-860
- 30. Loeys BL, Chen J, Neptune ER, Judge DP, Podowski M, Holm T, et al. A syndrome of altered cardiovascular, craniofacial, neurocognitive and skeletal development caused by mutations in TGFBR1 or TGFBR2. Nat Genet 2005; 37: 275-281
- 31. Neptune ER, Frischmeyer PA, Arking DE, Myers L, Bunton TE, Gayraud B, et al. Dysregulation of TGF-beta activation contributes to pathogenesis in Marfan syndrome. Nat Genet 2003; 33: 407-411
- 32. Pogue R, Lyons K. BMP signaling in the cartilage growth plate. Curr Top Dev Biol 2006; 76: 1- 48
- 33. Boileau C, Jondeau G, Babron MC, Coulon M, Alexandre JA, Sakai L, et al. Autosomal dominant Marfan-like connective-tissue disorder with aortic dilation and skeletal anomalies not linked to the fibrillin genes. Am J Hum Genet 1993; 53: 46-54
- 34. Matyas G, Arnold E, Carrel T, Baumgartner D, Boileau C, Berger W, et al. Identification and in silico analyses of novel TGFBR1 and TGFBR2 mutations in Marfan syndrome-related disorders. Hum Mutat 2006; 27: 760-769
- 35. Sakai H, Visser R, Ikegawa S, Ito E, Numabe H, Watanabe Y, et al. Comprehensive genetic analysis of relevant four genes in 49 patients with Marfan syndrome or Marfan-related phenotypes. Am J Med Genet A 2006; 140: 1719-1725
- 36. Mizuguchi T, Matsumoto N. Recent progress in genetics of Marfan syndrome and Marfanassociated disorders. J Hum Genet 2007; 52: 1-12
- 37. Loeys BL, Schwarze U, Holm T, Callewaert BL, Thomas GH, Pannu H, et al. Aneurysm syndromes caused by mutations in the TGF-beta receptor. N Engl J Med 2006; 355: 788-798
- 38. Putnam EA, Zhang H, Ramirez F, Milewicz DM. Fibrillin-2 (FBN2) mutations result in the Marfanlike disorder, congenital contractural arachnodactyly. Nat Genet 1995; 11: 456-458
- 39. Frederic MY, Monino C, Marschall C, Hamroun D, Faivre L, Jondeau G, et al. The FBN2 gene: new mutations, locus-specific database (Universal Mutation Database FBN2), and genotypephenotype correlations. Hum Mutat 2008; [Epub ahead of print]
- 40. Gaustadnes M, Wilcken B, Oliveriusova J, McGill J, Fletcher J, Kraus JP, et al. The molecular basis of cystathionine beta-synthase deficiency in Australian patients: genotype-phenotype correlations and response to treatment. Hum Mutat 2002; 20: 117-126
- 41. Mudd SH, Levy HL & Kraus JP: Disorders of transsulfuration; in Scriver CR, Beaudet AL, SLy WS & Valle D (eds): The Metabolic and Molecular Bases of Inherited Disease (8th edition). 2001. McGraw-Hill, New York, pp 2001-2056. 2001;
- 42. Brenton DP. Skeletal abnormalities in homocystinuria. Postgrad Med J 1977; 53: 488-496
- 43. Topaloglu AK, Sansaricq C, Snyderman SE. Influence of metabolic control on growth in homocystinuria due to cystathionine B-synthase deficiency. Pediatr Res 2001; 49: 796-798
- 44. Hutchinson S, Aplin RT, Webb H, Kettle S, Timmermans J, Boers GH, et al. Molecular effects of homocysteine on cbEGF domain structure: insights into the pathogenesis of homocystinuria. J Mol Biol 2005; 346: 833-844
- 45. Hubmacher D, Tiedemann K, Bartels R, Brinckmann J, Vollbrandt T, Batge B, et al. Modification of the structure and function of fibrillin-1 by homocysteine suggests a potential pathogenetic mechanism in homocystinuria. J Biol Chem 2005; 280: 34946-34955
- 46. Van Buggenhout G, Fryns JP. Lujan-Fryns syndrome (mental retardation, X-linked, marfanoid habitus). Orphanet J Rare Dis 2006; 1: 26
- 47. Schwartz CE, Tarpey PS, Lubs HA, Verloes A, May MM, Risheg H, et al. The original Lujan syndrome family has a novel missense mutation (p.N1007S) in the MED12 gene. J Med Genet 2007; 44: 472-477
- 48. Tarpey PS, Raymond FL, Nguyen LS, Rodriguez J, Hackett A, Vandeleur L, et al. Mutations in UPF3B, a member of the nonsense-mediated mRNA decay complex, cause syndromic and nonsyndromic mental retardation. Nat Genet 2007; 39: 1127-1133
- 49. Raymond FL, Tarpey PS, Edkins S, Tofts C, O'Meara S, Teague J, et al. Mutations in ZDHHC9, which encodes a palmitoyltransferase of NRAS and HRAS, cause X-linked mental retardation associated with a Marfanoid habitus. Am J Hum Genet 2007; 80: 982-987
- 50. Tatton-Brown K, Rahman N. Sotos syndrome. Eur J Hum Genet 2007; 15: 264-271
- 51. Tatton-Brown K, Douglas J, Coleman K, Baujat G, Cole TR, Das S, et al. Genotype-phenotype associations in Sotos syndrome: an analysis of 266 individuals with NSD1 aberrations. Am J Hum Genet 2005; 77: 193-204
- 52. Saugier-Veber P, Bonnet C, Afenjar A, Drouin-Garraud V, Coubes C, Fehrenbach S, et al. Heterogeneity of NSD1 alterations in 116 patients with Sotos syndrome. Hum Mutat 2007; 28: 1098-1107
- 53. Tatton-Brown K, Rahman N. Clinical features of NSD1-positive Sotos syndrome. Clin Dysmorphol 2004; 13: 199-204
- 54. Rahman N. Mechanisms predisposing to childhood overgrowth and cancer. Curr Opin Genet Dev 2005; 15: 227-233
- 55. Visser R, Matsumoto N. NSD1 and Sotos Syndrome. In: Epstein CJ, Erickson RP, Wynshaw-Boris A, eds. Inborn errors of development (2nd edition). New York, Oxford University Press, 2008; 1032-1037
- 56. Kurotaki N, Harada N, Shimokawa O, Miyake N, Kawame H, Uetake K, et al. Fifty microdeletions among 112 cases of Sotos syndrome: low copy repeats possibly mediate the common deletion. Hum Mutat 2003; 22: 378-387
- 57. Visser R, Shimokawa O, Harada N, Kinoshita A, Ohta T, Niikawa N, et al. Identification of a 3.0-kb major recombination hotspot in patients with sotos syndrome who carry a common 1.9-Mb microdeletion. Am J Hum Genet 2005; 76: 52-67
- 58. Nagai T, Matsumoto N, Kurotaki N, Harada N, Niikawa N, Ogata T, et al. Sotos syndrome and haploinsufficiency of NSD1: clinical features of intragenic mutations and submicroscopic deletions. J Med Genet 2003; 40: 285-289
- 59. Rio M, Clech L, Amiel J, Faivre L, Lyonnet S, Le Merrer M, et al. Spectrum of NSD1 mutations in Sotos and Weaver syndromes. J Med Genet 2003; 40: 436-440
- 60. Douglas J, Hanks S, Temple IK, Davies S, Murray A, Upadhyaya M, et al. NSD1 mutations are the major cause of Sotos syndrome and occur in some cases of Weaver syndrome but are rare in other overgrowth phenotypes. Am J Hum Genet 2003; 72: 132-143
- 61. Baujat G, Rio M, Rossignol S, Sanlaville D, Lyonnet S, Le Merrer M, et al. Paradoxical NSD1 mutations in Beckwith-Wiedemann syndrome and 11p15 anomalies in Sotos syndrome. Am J Hum Genet 2004; 74: 715-720
- 62. Kanemoto N, Kanemoto K, Nishimura G, Kamoda T, Visser R, Shimokawa O, et al. Nevo syndrome with an NSD1 deletion: A variant of Sotos syndrome? Am J Med Genet A 2006; 140: 70-73
- 63. Huang N, vom BE, Garnier JM, Lerouge T, Vonesch JL, Lutz Y, et al. Two distinct nuclear receptor interaction domains in NSD1, a novel SET protein that exhibits characteristics of both corepressors and coactivators. EMBO J 1998; 17: 3398-3412
- 64. Rayasam GV, Wendling O, Angrand PO, Mark M, Niederreither K, Song L, et al. NSD1 is essential for early post-implantation development and has a catalytically active SET domain. EMBO J 2003; 22: 3153-3163
- 65. Nielsen AL, Jorgensen P, Lerouge T, Cervino M, Chambon P, Losson R. Nizp1, a novel multitype zinc finger protein that interacts with the NSD1 histone lysine methyltransferase through a unique C2HR motif. Mol Cell Biol 2004; 24: 5184-5196
- 66. De Boer L, Van Duyvenvoorde HA, Willemstein-Van Hove EC, Hoogerbrugge CM, Van Doorn J, Maassen JA, et al. Mutations in the NSD1 gene in patients with Sotos syndrome associate with endocrine and paracrine alterations in the IGF system. Eur J Endocrinol 2004; 151: 333-341
- 67. Cole TR, Dennis NR, Hughes HE. Weaver syndrome. J Med Genet 1992; 29: 332-337
- 68. Weaver DD, Graham CB, Thomas IT, Smith DW. A new overgrowth syndrome with accelerated skeletal maturation, unusual facies, and camptodactyly. J Pediatr 1974; 84: 547-552
- 69. Opitz JM, Weaver DW, Reynolds JF, Jr. The syndromes of Sotos and Weaver: reports and review. Am J Med Genet 1998; 79: 294-304
- 70. Proud VK, Braddock SR, Cook L, Weaver DD. Weaver syndrome: autosomal dominant inheritance of the disorder. Am J Med Genet 1998; 79: 305-310
- 71. Turkmen S, Gillessen-Kaesbach G, Meinecke P, Albrecht B, Neumann LM, Hesse V, et al. Mutations in NSD1 are responsible for Sotos syndrome, but are not a frequent finding in other overgrowth phenotypes. Eur J Hum Genet 2003; 11: 858-865
- 72. Cecconi M, Forzano F, Milani D, Cavani S, Baldo C, Selicorni A, et al. Mutation analysis of the NSD1 gene in a group of 59 patients with congenital overgrowth. Am J Med Genet A 2005; 134: 247-253
- 73. Tong TM, Hau EW, Lo IF, Chan DH, Lam ST. Spectrum of NSD1 gene mutations in southern Chinese patients with Sotos syndrome. Chin Med J (Engl) 2005; 118: 1499-1506
- 74. Nevo S, Zeltzer M, Benderly A, Levy J. Evidence for autosomal recessive inheritance in cerebral gigantism. J Med Genet 1974; 11: 158-165
- 75. Hilderink BG, Brunner HG. Nevo syndrome. Clin Dysmorphol 1995; 4: 319-323
- 76. Al Gazali LI, Bakalinova D, Varady E, Scorer J, Nork M. Further delineation of Nevo syndrome. J Med Genet 1997; 34: 366-370
- 77. Giunta C, Randolph A, Al Gazali LI, Brunner HG, Kraenzlin ME, Steinmann B. Nevo syndrome is allelic to the kyphoscoliotic type of the Ehlers-Danlos syndrome (EDS VIA). Am J Med Genet A 2005; 133: 158-164
- 78. Beighton P, De Paepe A, Steinmann B, Tsipouras P, Wenstrup RJ. Ehlers-Danlos syndromes: revised nosology, Villefranche, 1997. Ehlers-Danlos National Foundation (USA) and Ehlers-Danlos Support Group (UK). Am J Med Genet 1998; 77: 31-37
- 79. Giunta C, Randolph A, Steinmann B. Mutation analysis of the PLOD1 gene: an efficient multistep approach to the molecular diagnosis of the kyphoscoliotic type of Ehlers-Danlos syndrome (EDS VIA). Mol Genet Metab 2005; 86: 269-276
- 80. Yis U, Dirik E, Chambaz C, Steinmann B, Giunta C. Differential diagnosis of muscular hypotonia in infants: the kyphoscoliotic type of Ehlers-Danlos syndrome (EDS VI). Neuromuscul Disord 2008; 18: 210-214
- 81. Steinmann B, Eyre DR, Shao P. Urinary pyridinoline cross-links in Ehlers-Danlos syndrome type VI. Am J Hum Genet 1995; 57: 1505-1508
- 82. Takaluoma K, Hyry M, Lantto J, Sormunen R, Bank RA, Kivirikko KI, et al. Tissue-specific changes in the hydroxylysine content and cross-links of collagens and alterations in fibril morphology in lysyl hydroxylase 1 knock-out mice. J Biol Chem 2007; 282: 6588-6596
- 83. Elliott M, Bayly R, Cole T, Temple IK, Maher ER. Clinical features and natural history of Beckwith-Wiedemann syndrome: presentation of 74 new cases. Clin Genet 1994; 46: 168-174
- 84. Cohen MM, Jr. Beckwith-Wiedemann syndrome: historical, clinicopathological, and etiopathogenetic perspectives. Pediatr Dev Pathol 2005; 8: 287-304
- 85. Cytrynbaum CS, Smith AC, Rubin T, Weksberg R. Advances in overgrowth syndromes: clinical classification to molecular delineation in Sotos syndrome and Beckwith-Wiedemann syndrome. Curr Opin Pediatr 2005; 17: 740-746
- 86. Thorburn MJ, Wright ES, Miller CG, Smith-Read EH. Exomphalos-macroglossia-gigantism syndrome in Jamaican infants. Am J Dis Child 1970; 119: 316-321
- 87. Pettenati MJ, Haines JL, Higgins RR, Wappner RS, Palmer CG, Weaver DD. Wiedemann-Beckwith syndrome: presentation of clinical and cytogenetic data on 22 new cases and review of the literature. Hum Genet 1986; 74: 143-154
- 88. Enklaar T, Zabel BU, Prawitt D. Beckwith-Wiedemann syndrome: multiple molecular mechanisms. Expert Rev Mol Med 2006; 8: 1-19
- 89. Scott RH, Douglas J, Baskcomb L, Huxter N, Barker K, Hanks S, et al. Constitutional 11p15 abnormalities, including heritable imprinting center mutations, cause nonsyndromic Wilms tumor. Nat Genet 2008; Epub ahead of print
- 90. Rossignol S, Netchine I, Le Bouc Y, Gicquel C. Epigenetics in Silver-Russell syndrome. Best Pract Res Clin Endocrinol Metab 2008; 22: 403-414
- 91. Schonherr N, Meyer E, Roos A, Schmidt A, Wollmann HA, Eggermann T. The centromeric 11p15 imprinting centre is also involved in Silver-Russell syndrome. J Med Genet 2007; 44: 59-63
- 92. Cooper WN, Luharia A, Evans GA, Raza H, Haire AC, Grundy R, et al. Molecular subtypes and phenotypic expression of Beckwith-Wiedemann syndrome. Eur J Hum Genet 2005; 13: 1025- 1032
- 93. Lam WW, Hatada I, Ohishi S, Mukai T, Joyce JA, Cole TR, et al. Analysis of germline CDKN1C (p57KIP2) mutations in familial and sporadic Beckwith-Wiedemann syndrome (BWS) provides a novel genotype-phenotype correlation. J Med Genet 1999; 36: 518-523
- 94. Bliek J, Maas SM, Ruijter JM, Hennekam RC, Alders M, Westerveld A, et al. Increased tumour risk for BWS patients correlates with aberrant H19 and not KCNQ1OT1 methylation: occurrence of KCNQ1OT1 hypomethylation in familial cases of BWS. Hum Mol Genet 2001; 10: 467-476
- 95. Weksberg R, Nishikawa J, Caluseriu O, Fei YL, Shuman C, Wei C, et al. Tumor development in the Beckwith-Wiedemann syndrome is associated with a variety of constitutional molecular 11p15 alterations including imprinting defects of KCNQ1OT1. Hum Mol Genet 2001; 10: 2989-3000
- 96. Weksberg R, Shen DR, Fei YL, Song QL, Squire J. Disruption of insulin-like growth factor 2 imprinting in Beckwith-Wiedemann syndrome. Nat Genet 1993; 5: 143-150
- 97. Sun FL, Dean WL, Kelsey G, Allen ND, Reik W. Transactivation of Igf2 in a mouse model of Beckwith-Wiedemann syndrome. Nature 1997; 389: 809-815
- 98. Ogawa O, Eccles MR, Szeto J, McNoe LA, Yun K, Maw MA, et al. Relaxation of insulin-like growth factor II gene imprinting implicated in Wilms' tumour. Nature 1993; 362: 749-751
- 99. Sakatani T, Wei M, Katoh M, Okita C, Wada D, Mitsuya K, et al. Epigenetic heterogeneity at imprinted loci in normal populations. Biochem Biophys Res Commun 2001; 283: 1124-1130
- 100. Caspary T, Cleary MA, Perlman EJ, Zhang P, Elledge SJ, Tilghman SM. Oppositely imprinted genes p57(Kip2) and igf2 interact in a mouse model for Beckwith-Wiedemann syndrome. Genes Dev 1999; 13: 3115-3124
- 101. Zhang P, Liegeois NJ, Wong C, Finegold M, Hou H, Thompson JC, et al. Altered cell differentiation and proliferation in mice lacking p57KIP2 indicates a role in Beckwith-Wiedemann syndrome. Nature 1997; 387: 151-158
- 102. Grandjean V, Smith J, Schofield PN, Ferguson-Smith AC. Increased IGF-II protein affects p57kip2 expression in vivo and in vitro: implications for Beckwith-Wiedemann syndrome. Proc Natl Acad Sci U S A 2000; 97: 5279-5284
- 103. Mariani S, Iughetti L, Bertorelli R, Coviello D, Pellegrini M, Forabosco A, et al. Genotype/ phenotype correlations of males affected by Simpson-Golabi-Behmel syndrome with GPC3 gene mutations: patient report and review of the literature. J Pediatr Endocrinol Metab 2003; 16: 225-232
- 104. Li M, Shuman C, Fei YL, Cutiongco E, Bender HA, Stevens C, et al. GPC3 mutation analysis in a spectrum of patients with overgrowth expands the phenotype of Simpson-Golabi-Behmel syndrome. Am J Med Genet 2001; 102: 161-168
- 105. DeBaun MR, Ess J, Saunders S. Simpson Golabi Behmel syndrome: progress toward understanding the molecular basis for overgrowth, malformation, and cancer predisposition. Mol Genet Metab 2001; 72: 279-286
- 106. Neri G, Gurrieri F, Zanni G, Lin A. Clinical and molecular aspects of the Simpson-Golabi-Behmel syndrome. Am J Med Genet 1998; 79: 279-283
- 107. Hughes-Benzie RM, Pilia G, Xuan JY, Hunter AG, Chen E, Golabi M, et al. Simpson-Golabi-Behmel syndrome: genotype/phenotype analysis of 18 affected males from 7 unrelated families. Am J Med Genet 1996; 66: 227-234
- 108. Veugelers M, Cat BD, Muyldermans SY, Reekmans G, Delande N, Frints S, et al. Mutational analysis of the GPC3/GPC4 glypican gene cluster on Xq26 in patients with Simpson-Golabi-Behmel syndrome: identification of loss-of-function mutations in the GPC3 gene. Hum Mol Genet 2000; 9: 1321-1328
- 109. Jakubovic BD, Jothy S. Glypican-3: from the mutations of Simpson-Golabi-Behmel genetic syndrome to a tumor marker for hepatocellular carcinoma. Exp Mol Pathol 2007; 82: 184-189
- 110. Toretsky JA, Zitomersky NL, Eskenazi AE, Voigt RW, Strauch ED, Sun CC, et al. Glypican-3 expression in Wilms tumor and hepatoblastoma. J Pediatr Hematol Oncol 2001; 23: 496-499
- 111. Brzustowicz LM, Farrell S, Khan MB, Weksberg R. Mapping of a new SGBS locus to chromosome Xp22 in a family with a severe form of Simpson-Golabi-Behmel syndrome. Am J Hum Genet 1999; 65: 779-783
- 112. Budny B, Chen W, Omran H, Fliegauf M, Tzschach A, Wisniewska M, et al. A novel X-linked recessive mental retardation syndrome comprising macrocephaly and ciliary dysfunction is allelic to oral-facial-digital type I syndrome. Hum Genet 2006; 120: 171-178
- 113. Ferrante MI, Giorgio G, Feather SA, Bulfone A, Wright V, Ghiani M, et al. Identification of the gene for oral-facial-digital type I syndrome. Am J Hum Genet 2001; 68: 569-576
- 114. Filmus J. Glypicans in growth control and cancer. Glycobiology 2001; 11: 19R-23R
- 115. Cheng W, Tseng CJ, Lin TT, Cheng I, Pan HW, Hsu HC, et al. Glypican-3-mediated oncogenesis involves the IGF signaling pathway. Carcinogenesis 2008; 29: 1319-1326
- 116. Pellegrini M, Pilia G, Pantano S, Lucchini F, Uda M, Fumi M, et al. Gpc3 expression correlates with the phenotype of the Simpson-Golabi-Behmel syndrome. Dev Dyn 1998; 213: 431-439
- 117. Eggenschwiler J, Ludwig T, Fisher P, Leighton PA, Tilghman SM, Efstratiadis A. Mouse mutant embryos overexpressing IGF-II exhibit phenotypic features of the Beckwith-Wiedemann and Simpson-Golabi-Behmel syndromes. Genes Dev 1997; 11: 3128-3142
- 118. Song HH, Shi W, Filmus J. OCI-5/rat glypican-3 binds to fibroblast growth factor-2 but not to insulin-like growth factor-2. J Biol Chem 1997; 272: 7574-7577
- 119. Cano-Gauci DF, Song HH, Yang H, McKerlie C, Choo B, Shi W, et al. Glypican-3-deficient mice exhibit developmental overgrowth and some of the abnormalities typical of Simpson-Golabi-Behmel syndrome. J Cell Biol 1999; 146: 255-264
- 120. Chiao E, Fisher P, Crisponi L, Deiana M, Dragatsis I, Schlessinger D, et al. Overgrowth of a mouse model of the Simpson-Golabi-Behmel syndrome is independent of IGF signaling. Dev Biol 2002; 243: 185-206
- 121. Capurro MI, Xu P, Shi W, Li F, Jia A, Filmus J. Glypican-3 inhibits Hedgehog signaling during development by competing with patched for Hedgehog binding. Dev Cell 2008; 14: 700-711
- 122. Sakazume S, Okamoto N, Yamamoto T, Kurosawa K, Numabe H, Ohashi Y, et al. GPC3 mutations in seven patients with Simpson-Golabi-Behmel syndrome. Am J Med Genet A 2007; 143A: 1703-1707
- 123. Marsh DJ, Dahia PL, Zheng Z, Liaw D, Parsons R, Gorlin RJ, et al. Germline mutations in PTEN are present in Bannayan-Zonana syndrome. Nat Genet 1997; 16: 333-334
- 124. Zigman AF, Lavine JE, Jones MC, Boland CR, Carethers JM. Localization of the Bannayan-Riley-Ruvalcaba syndrome gene to chromosome 10q23. Gastroenterology 1997; 113: 1433-1437
- 125. Parisi MA, Dinulos MB, Leppig KA, Sybert VP, Eng C, Hudgins L. The spectrum and evolution of phenotypic findings in PTEN mutation positive cases of Bannayan-Riley-Ruvalcaba syndrome. J Med Genet 2001; 38: 52-58
- 126. Marsh DJ, Kum JB, Lunetta KL, Bennett MJ, Gorlin RJ, Ahmed SF, et al. PTEN mutation spectrum and genotype-phenotype correlations in Bannayan-Riley-Ruvalcaba syndrome suggest a single entity with Cowden syndrome. Hum Mol Genet 1999; 8: 1461-1472
- 127. Lachlan KL, Lucassen AM, Bunyan D, Temple IK. Cowden syndrome and Bannayan Riley Ruvalcaba syndrome represent one condition with variable expression and age-related penetrance: results of a clinical study of PTEN mutation carriers. J Med Genet 2007; 44: 579- 585
- 128. Hendriks YM, Verhallen JT, van der Smagt JJ, Kant SG, Hilhorst Y, Hoefsloot L, et al. Bannayan-Riley-Ruvalcaba syndrome: further delineation of the phenotype and management of PTEN mutation-positive cases. Fam Cancer 2003; 2: 79-85
- 129. Tan WH, Baris HN, Burrows PE, Robson CD, Alomari AI, Mulliken JB, et al. The spectrum of vascular anomalies in patients with PTEN mutations: implications for diagnosis and management. J Med Genet 2007; 44: 594-602
- 130. Longy M, Coulon V, Duboue B, David A, Larregue M, Eng C, et al. Mutations of PTEN in patients with Bannayan-Riley-Ruvalcaba phenotype. J Med Genet 1998; 35: 886-889
- 131. Marsh DJ, Coulon V, Lunetta KL, Rocca-Serra P, Dahia PL, Zheng Z, et al. Mutation spectrum and genotype-phenotype analyses in Cowden disease and Bannayan-Zonana syndrome, two hamartoma syndromes with germline PTEN mutation. Hum Mol Genet 1998; 7: 507-515
- 132. Zhou XP, Waite KA, Pilarski R, Hampel H, Fernandez MJ, Bos C, et al. Germline PTEN promoter mutations and deletions in Cowden/Bannayan-Riley-Ruvalcaba syndrome result in aberrant PTEN protein and dysregulation of the phosphoinositol-3-kinase/Akt pathway. Am J Hum Genet 2003; 73: 404-411
- 133. Gustafson S, Zbuk KM, Scacheri C, Eng C. Cowden syndrome. Semin Oncol 2007; 34: 428-434
- 134. Eng C. Will the real Cowden syndrome please stand up: revised diagnostic criteria. J Med Genet 2000; 37: 828-830
- 135. Eng C. PTEN: one gene, many syndromes. Hum Mutat 2003; 22: 183-198
- 136. Zhou X, Hampel H, Thiele H, Gorlin RJ, Hennekam RC, Parisi M, et al. Association of germline mutation in the PTEN tumour suppressor gene and Proteus and Proteus-like syndromes. Lancet 2001; 358: 210-211
- 137. Barker K, Martinez A, Wang R, Bevan S, Murday V, Shipley J, et al. PTEN mutations are uncommon in Proteus syndrome. J Med Genet 2001; 38: 480-481
- 138. Biesecker LG, Rosenberg MJ, Vacha S, Turner JT, Cohen MM. PTEN mutations and proteus syndrome. Lancet 2001; 358: 2079-2080
- 139. Thiffault I, Schwartz CE, Der K, V, Foulkes WD. Mutation analysis of the tumor suppressor PTEN and the glypican 3 (GPC3) gene in patients diagnosed with Proteus syndrome. Am J Med Genet A 2004; 130: 123-127
- 140. Sansal I, Sellers WR. The biology and clinical relevance of the PTEN tumor suppressor pathway. J Clin Oncol 2004; 22: 2954-2963
- 141. Tamura M, Gu J, Matsumoto K, Aota S, Parsons R, Yamada KM. Inhibition of cell migration, spreading, and focal adhesions by tumor suppressor PTEN. Science 1998; 280: 1614-1617
- 142. Ford-Hutchinson AF, Ali Z, Lines SE, Hallgrimsson B, Boyd SK, Jirik FR. Inactivation of Pten in osteo-chondroprogenitor cells leads to epiphyseal growth plate abnormalities and skeletal overgrowth. J Bone Miner Res 2007; 22: 1245-1259
- 143. Liu X, Bruxvoort KJ, Zylstra CR, Liu J, Cichowski R, Faugere MC, et al. Lifelong accumulation of bone in mice lacking Pten in osteoblasts. Proc Natl Acad Sci U S A 2007; 104: 2259-2264
- 144. Crawford DC, Acuna JM, Sherman SL. FMR1 and the fragile X syndrome: human genome epidemiology review. Genet Med 2001; 3: 359-371
- 145. Hagerman PJ. The fragile X prevalence paradox. J Med Genet 2008; 45: 498-499
- 146. Garber KB, Visootsak J, Warren ST. Fragile X syndrome. Eur J Hum Genet 2008; 16: 666-672
- 147. Maes B, Fryns JP, Ghesquiere P, Borghgraef M. Phenotypic checklist to screen for fragile X syndrome in people with mental retardation. Ment Retard 2000; 38: 207-215
- 148. Butler MG, Brunschwig A, Miller LK, Hagerman RJ. Standards for selected anthropometric measurements in males with the fragile X syndrome. Pediatrics 1992; 89: 1059-1062
- 149. Merenstein SA, Sobesky WE, Taylor AK, Riddle JE, Tran HX, Hagerman RJ. Molecular-clinical correlations in males with an expanded FMR1 mutation. Am J Med Genet 1996; 64: 388-394
- 150. de Vries BB, Robinson H, Stolte-Dijkstra I, Tjon Pian Gi CV, Dijkstra PF, van Doorn J, et al. General overgrowth in the fragile X syndrome: variability in the phenotypic expression of the FMR1 gene mutation. J Med Genet 1995; 32: 764-769
- 151. Loesch DZ, Huggins RM, Hoang NH. Growth in stature in fragile X families: a mixed longitudinal study. Am J Med Genet 1995; 58: 249-256
- 152. Hagerman RJ. Lessons from fragile X regarding neurobiology, autism, and neurodegeneration. J Dev Behav Pediatr 2006; 27: 63-74
- 153. Sherman S, Pletcher BA, Driscoll DA. Fragile X syndrome: diagnostic and carrier testing. Genet Med 2005; 7: 584-587
- 154. Verkerk AJ, Pieretti M, Sutcliffe JS, Fu YH, Kuhl DP, Pizzuti A, et al. Identification of a gene (FMR-1) containing a CGG repeat coincident with a breakpoint cluster region exhibiting length variation in fragile X syndrome. Cell 1991; 65: 905-914
- 155. Lubs HA. A marker X chromosome. Am J Hum Genet 1969; 21: 231-244
- 156. Penagarikano O, Mulle JG, Warren ST. The pathophysiology of fragile x syndrome. Annu Rev Genomics Hum Genet 2007; 8: 109-129
- 157. Fu YH, Kuhl DP, Pizzuti A, Pieretti M, Sutcliffe JS, Richards S, et al. Variation of the CGG repeat at the fragile X site results in genetic instability: resolution of the Sherman paradox. Cell 1991; 67: 1047-1058
- 158. Malter HE, Iber JC, Willemsen R, de Graaff E, Tarleton JC, Leisti J, et al. Characterization of the full fragile X syndrome mutation in fetal gametes. Nat Genet 1997; 15: 165-169
- 159. Van Esch H. The Fragile X premutation: new insights and clinical consequences. Eur J Med Genet 2006; 49: 1-8
- 160. Devys D, Lutz Y, Rouyer N, Bellocq JP, Mandel JL. The FMR-1 protein is cytoplasmic, most abundant in neurons and appears normal in carriers of a fragile X premutation. Nat Genet 1993; 4: 335-340
- 161. Garber K, Smith KT, Reines D, Warren ST. Transcription, translation and fragile X syndrome. Curr Opin Genet Dev 2006; 16: 270-275
- 162. Irwin SA, Galvez R, Greenough WT. Dendritic spine structural anomalies in fragile-X mental retardation syndrome. Cereb Cortex 2000; 10: 1038-1044
- 163. Bear MF, Huber KM, Warren ST. The mGluR theory of fragile X mental retardation. Trends Neurosci 2004; 27: 370-377
- 164. Fmr1 knockout mice: a model to study fragile X mental retardation. The Dutch-Belgian Fragile X Consortium. Cell 1994; 78: 23-33
- 165. Tucker B, Richards RI, Lardelli M. Contribution of mGluR and Fmr1 functional pathways to neurite morphogenesis, craniofacial development and fragile X syndrome. Hum Mol Genet 2006; 15: 3446-3458
- 166. Morris JK, Alberman E, Scott C, Jacobs P. Is the prevalence of Klinefelter syndrome increasing? Eur J Hum Genet 2008; 16: 163-170
- 167. Smyth CM, Bremner WJ. Klinefelter syndrome. Arch Intern Med 1998; 158: 1309-1314
- 168. Zeger MP, Zinn AR, Lahlou N, Ramos P, Kowal K, Samango-Sprouse C, et al. Effect of ascertainment and genetic features on the phenotype of Klinefelter syndrome. J Pediatr 2008; 152: 716-722
- 169. Ratcliffe S. Long-term outcome in children of sex chromosome abnormalities. Arch Dis Child 1999; 80: 192-195
- 170. Schibler D, Brook CG, Kind HP, Zachmann M, Prader A. Growth and body proportions in 54 boys and men with Klinefelter's syndrome. Helv Paediatr Acta 1974; 29: 325-333
- 171. Mandoki MW, Sumner GS, Hoffman RP, Riconda DL. A review of Klinefelter's syndrome in children and adolescents. J Am Acad Child Adolesc Psychiatry 1991; 30: 167-172
- 172. Salbenblatt JA, Bender BG, Puck MH, Robinson A, Faiman C, Winter JS. Pituitary-gonadal function in Klinefelter syndrome before and during puberty. Pediatr Res 1985; 19: 82-86
- 173. Visootsak J, Graham JM, Jr. Klinefelter syndrome and other sex chromosomal aneuploidies. Orphanet J Rare Dis 2006; 1: 42
- 174. Jacobs PA, Hassold TJ, Whittington E, Butler G, Collyer S, Keston M, et al. Klinefelter's syndrome: an analysis of the origin of the additional sex chromosome using molecular probes. Ann Hum Genet 1988; 52: 93-109
- 175. Carothers AD, Filippi G. Klinefelter's syndrome in Sardinia and Scotland. Comparative studies of parental age and other aetiological factors in 47,XXY. Hum Genet 1988; 81: 71-75
- 176. Lorda-Sanchez I, Binkert F, Maechler M, Robinson WP, Schinzel AA. Reduced recombination and paternal age effect in Klinefelter syndrome. Hum Genet 1992; 89: 524-530
- 177. Lanfranco F, Kamischke A, Zitzmann M, Nieschlag E. Klinefelter's syndrome. Lancet 2004; 364: 273-283
- 178. Wikstrom AM, Dunkel L. Testicular function in Klinefelter syndrome. Horm Res 2008; 69: 317- 326
- 179. Zitzmann M, Depenbusch M, Gromoll J, Nieschlag E. X-chromosome inactivation patterns and androgen receptor functionality influence phenotype and social characteristics as well as pharmacogenetics of testosterone therapy in Klinefelter patients. J Clin Endocrinol Metab 2004; 89: 6208-6217
- 180. Zinn AR, Ramos P, Elder FF, Kowal K, Samango-Sprouse C, Ross JL. Androgen receptor CAGn repeat length influences phenotype of 47,XXY (Klinefelter) syndrome. J Clin Endocrinol Metab 2005; 90: 5041-5046
- 181. Ogata T, Matsuo N, Nishimura G. SHOX haploinsufficiency and overdosage: impact of gonadal function status. J Med Genet 2001; 38: 1-6
- 182. Rao E, Weiss B, Fukami M, Rump A, Niesler B, Mertz A, et al. Pseudoautosomal deletions encompassing a novel homeobox gene cause growth failure in idiopathic short stature and Turner syndrome. Nat Genet 1997; 16: 54-63
- 183. Kamischke A, Baumgardt A, Horst J, Nieschlag E. Clinical and diagnostic features of patients with suspected Klinefelter syndrome. J Androl 2003; 24: 41-48
- 184. Bocciardi R, Giorda R, Buttgereit J, Gimelli S, Divizia MT, Beri S, et al. Overexpression of the C-type natriuretic peptide (CNP) is associated with overgrowth and bone anomalies in an individual with balanced t(2;7) translocation. Hum Mutat 2007; 28: 724-731
- 185. Moncla A, Missirian C, Cacciagli P, Balzamo E, Legeai-Mallet L, Jouve JL, et al. A cluster of translocation breakpoints in 2q37 is associated with overexpression of NPPC in patients with a similar overgrowth phenotype. Hum Mutat 2007; 28: 1183-1188
- 186. Yasoda A, Komatsu Y, Chusho H, Miyazawa T, Ozasa A, Miura M, et al. Overexpression of CNP in chondrocytes rescues achondroplasia through a MAPK-dependent pathway. Nat Med 2004; 10: 80-86
- 187. Chusho H, Tamura N, Ogawa Y, Yasoda A, Suda M, Miyazawa T, et al. Dwarfism and early death in mice lacking C-type natriuretic peptide. Proc Natl Acad Sci U S A 2001; 98: 4016-4021
- 188. Yasoda A, Komatsu Y, Chusho H, Miyazawa T, Ozasa A, Miura M, et al. Overexpression of CNP in chondrocytes rescues achondroplasia through a MAPK-dependent pathway. Nat Med 2004; 10: 80-86
- 189. Bartels CF, Bukulmez H, Padayatti P, Rhee DK, Ravenswaaij-Arts C, Pauli RM, et al. Mutations in the transmembrane natriuretic peptide receptor NPR-B impair skeletal growth and cause acromesomelic dysplasia, type Maroteaux. Am J Hum Genet 2004; 75: 27-34
- 190. Olney RC, Bukulmez H, Bartels CF, Prickett TC, Espiner EA, Potter LR, et al. Heterozygous mutations in natriuretic peptide receptor-B (NPR2) are associated with short stature. J Clin Endocrinol Metab 2006; 91: 1229-1232
- 191. Pejchalova K, Krejci P, Wilcox WR. C-natriuretic peptide: an important regulator of cartilage. Mol Genet Metab 2007; 92: 210-215
- 192. Colvin JS, Bohne BA, Harding GW, McEwen DG, Ornitz DM. Skeletal overgrowth and deafness in mice lacking fibroblast growth factor receptor 3. Nat Genet 1996; 12: 390-397
- 193. Douglas J, Cilliers D, Coleman K, Tatton-Brown K, Barker K, Bernhard B, et al. Mutations in RNF135, a gene within the NF1 microdeletion region, cause phenotypic abnormalities including overgrowth. Nat Genet 2007; 39: 963-965
- 194. Kluwe L, Siebert R, Gesk S, Friedrich RE, Tinschert S, Kehrer-Sawatzki H, et al. Screening 500 unselected neurofibromatosis 1 patients for deletions of the NF1 gene. Hum Mutat 2004; 23: 111-116
- 195. Cnossen MH, van der Est MN, Breuning MH, van Asperen CJ, Breslau-Siderius EJ, van der Ploeg AT, et al. Deletions spanning the neurofibromatosis type 1 gene: implications for genotypephenotype correlations in neurofibromatosis type 1? Hum Mutat 1997; 9: 458-464
- 196. Spiegel M, Oexle K, Horn D, Windt E, Buske A, Albrecht B, et al. Childhood overgrowth in patients with common NF1 microdeletions. Eur J Hum Genet 2005; 13: 883-888
- 197. van Asperen CJ, Overweg-Plandsoen WC, Cnossen MH, van Tijn DA, Hennekam RC. Familial neurofibromatosis type 1 associated with an overgrowth syndrome resembling Weaver syndrome. J Med Genet 1998; 35: 323-327
- 198. Visser R, Koelma N, Vijfhuizen L, van der Wielen MJ, Kant SG, Breuning MH, et al. RNF135 mutations are not present in patients with Sotos syndrome-like features. Am J Med Genet A 2009; 149A: 806-808