



Universiteit
Leiden
The Netherlands

Tissue factor isoforms and cancer

Berg, Y.W. van den

Citation

Berg, Y. W. van den. (2013, October 8). *Tissue factor isoforms and cancer*. Retrieved from <https://hdl.handle.net/1887/21936>

Version: Corrected Publisher's Version

License: [Licence agreement concerning inclusion of doctoral thesis in the Institutional Repository of the University of Leiden](#)

Downloaded from: <https://hdl.handle.net/1887/21936>

Note: To cite this publication please use the final published version (if applicable).

Cover Page



Universiteit Leiden



The handle <http://hdl.handle.net/1887/21936> holds various files of this Leiden University dissertation.

Author: Berg, Yascha Wilfred van den
Title: Tissue factor isoforms and cancer
Issue Date: 2013-10-08

Chapter 1 - General introduction and outline of the thesis

Full-length tissue factor (fTF) is a 47 kDa transmembrane glycoprotein that is primarily responsible for activation of the coagulation cascade along the extrinsic pathway. Upon vessel damage, subendothelial expressed fTF forms a complex with circulating factor VII/VIIa (FVII/FVIIa) that can subsequently activate factor X (FX) and thrombin, resulting in localized formation of fibrin required for hemostasis. fTF expression is classically thought to be limited to subendothelial tissues in order to serve as a hemostatic envelope surrounding the vasculature^{1;2}. However, in specific conditions, circulating monocytes, endothelium and circulating microparticles can bear membrane-bound fTF as well^{3;4}.

Besides its important role in initiating the coagulation cascade, fTF is involved in numerous physiological and pathological processes like angiogenesis and inflammation. fTF expression on endothelium and circulating microparticles underlies thrombotic complications in cancer and sepsis due to fTF's coagulant activity, but non-hemostatic properties of fTF also influence tumor biology and inflammatory processes.

The study of fTF^{-/-} mice showed that fTF is indispensable for embryonic pericyte function and therefore to be critical for embryonic angiogenesis⁵. In adults, fTF's role in angiogenesis comprises wound healing, neovascularization and tumor angiogenesis. The last years, extensive research provided insight in how fTF triggers several cellular signaling pathways involved in cell growth, cell migration and cell survival⁶⁻¹². Examination of mice that express TF lacking the cytoplasmic domain indicated that this domain is not required for and may even inhibit angiogenesis. These results support the notion that fTF may only affect angiogenesis indirectly via downstream coagulation factors^{13;14}. In subsequent studies, however, direct activation of the membrane bound G-protein coupled protease activated receptor 2 (PAR2) by either the fTF/FVIIa/fXa-complex or the fTF/FVIIa-complex was reported^{15;16}. These direct activation routes obviate the need for downstream coagulation factors and the formation or breakage of the disulfide bond between Cys186 and Cys209 has been claimed to switch between either signaling or pro-coagulant activity of fTF. It should be noted though that there remains controversy regarding the role of disulfide switching and this remains an area of ongoing research^{12;17-19}.

Binding and activation of integrins is a second FVII/FVIIa-independent and non-hemostatic property of fTF. Integrins are heterodimeric complexes consisting of an α and β subunit that play a critical role in cellular mechanisms involved in angiogenesis, wound healing and cancer biology²⁰. fTF can directly bind to β 1 integrins and this binding results in activation of integrins, but also in an active state of the fTF:FVIIa:PAR2 complex, thus allowing for fTF signaling²¹.

In 2003 Bogdanov and colleagues reported a soluble, circulating splicing variant of fITF²². Through alternative splicing, exon 5 is excluded, leading to a frameshift that alters the amino acid sequence. This alternatively spliced tissue factor (asTF) lacks a transmembrane domain and has a unique C-terminus of which the consequences remain unknown. Since its discovery, the role of asTF in coagulation has been a matter of debate, although a role in tumor growth, potentially through angiogenesis by integrin ligation, has been suggested²²⁻²⁴.

Outline

In this thesis, the properties of asTF regarding angiogenesis, inflammatory processes and cancer are investigated. In addition, the relative contributions of each TF isoform to tumor angiogenesis are assessed in a large cohort of breast cancer patients.

Chapter 2 examines how recombinant human asTF promotes angiogenesis in a non-hemostatic, but integrin-dependant fashion. **Chapter 3** is a review that describes the current debate on the role of asTF in coagulation and its suggested non-hemostatic properties, with a focus on cancer, but also a potential role in atherosclerosis will be discussed. **Chapter 4** outlines how asTF promotes the interaction between inflammatory cells and the endothelium. In addition, data are provided that support the notion that asTF mediates adherence of inflammatory cells to the endothelium, which is relevant to both atherosclerosis and cancer. Whether murine asTF has similar properties as human asTF is investigated in **chapter 5**. Furthermore, this chapter describes the distribution of asTF in experimental murine models for atherosclerosis and cancer. **Chapter 6** proceeds with an assessment of the role of asTF in human breast cancer. A large cohort of human breast cancer is studied in order to provide insight in the clinical and pathological relevance of both asTF and fITF. **Chapter 7** continues with a study on ectopically produced FVII in breast cancer. The results suggest that auto-activation of the fITF:FVIIa:PAR2 axis influences the prognosis of young women with breast cancer. In **chapter 8** the evolutionary conservation of the cysteines involved in fITF disulfide bonding is assessed and a comparison is made with other proteins of which the activity is also regulated by isomerization of an allosteric disulfide. **Chapter 9** reviews the current knowledge on the non-hemostatic effects of TF isoforms in cancer. The chapter describes the regulation of TF expression in cancer, its effects *in vitro* and in mice, and how studies on human tumor material support the findings from experimental studies. **Chapter 10** provides a general summary and discussion.

References

1. Drake TA, Morrissey JH, Edgington TS. Selective cellular expression of tissue factor in human tissues. Implications for disorders of hemostasis and thrombosis. *Am.J.Pathol.* 1989;134:1087-1097.
2. Monroe DM, Hoffman M. What does it take to make the perfect clot? *Arterioscler.Thromb.Vasc.Biol.* 2006;26:41-48.
3. Drake TA, Cheng J, Chang A, Taylor FB, Jr. Expression of tissue factor, thrombomodulin, and E-selectin in baboons with lethal *Escherichia coli* sepsis. *Am.J.Pathol.* 1993;142:1458-1470.
4. Osterud B, Flaegstad T. Increased tissue thromboplastin activity in monocytes of patients with meningococcal infection: related to an unfavourable prognosis. *Thromb.Haemost.* 1983;49:5-7.
5. Carmeliet P, Mackman N, Moons L et al. Role of tissue factor in embryonic blood vessel development. *Nature* 1996;383:73-75.
6. Versteeg HH, Ruf W. Emerging insights in tissue factor-dependent signaling events. *Semin.Thromb.Hemost.* 2006;32:24-32.
7. Versteeg HH, Spek CA, Peppelenbosch MP, Richel DJ. Tissue factor and cancer metastasis: the role of intracellular and extracellular signaling pathways. *Mol.Med.* 2004;10:6-11.
8. Versteeg HH, Spek CA, Slofstra SH et al. FVIIa:TF induces cell survival via G12/G13-dependent Jak/STAT activation and BclXL production. *Circ.Res.* 2004;94:1032-1040.
9. Versteeg HH, Peppelenbosch MP, Spek CA. Tissue factor signal transduction in angiogenesis. *Carcinogenesis* 2003;24:1009-1013.
10. Versteeg HH, Peppelenbosch MP, Spek CA. The pleiotropic effects of tissue factor: a possible role for factor VIIa-induced intracellular signalling? *Thromb.Haemost.* 2001;86:1353-1359.
11. Versteeg HH, Hoedemaeker I, Diks SH et al. Factor VIIa/tissue factor-induced signaling via activation of Src-like kinases, phosphatidylinositol 3-kinase, and Rac. *J.Biol.Chem.* 2000;275:28750-28756.
12. Versteeg HH, Schaffner F, Kerver M et al. Inhibition of tissue factor signaling suppresses tumor growth. *Blood* 2007
13. Melis E, Moons L, Arnout J et al. Thrombophilia in mice expressing a tissue factor variant lacking its transmembrane and cytosolic domain. *Biochem.Biophys.Res.Commun.* 2005;333:488-495.
14. Riewald M, Ruf W. Mechanistic coupling of protease signaling and initiation of coagulation by tissue factor. *Proc.Natl.Acad.Sci.U.S.A* 2001;98:7742-7747.
15. Belting M, Ahamed J, Ruf W. Signaling of the tissue factor coagulation pathway in angiogenesis and cancer. *Arterioscler.Thromb.Vasc.Biol.* 2005;25:1545-1550.
16. Belting M, Dorrell MI, Sandgren S et al. Regulation of angiogenesis by tissue factor cytoplasmic domain signaling. *Nat.Med.* 2004;10:502-509.
17. Ahamed J, Versteeg HH, Kerver M et al. Disulfide isomerization switches tissue factor from coagulation to cell signaling. *Proc.Natl.Acad.Sci.U.S.A* 2006;103:13932-13937.
18. Versteeg HH, Ruf W. Tissue factor coagulant function is enhanced by protein disulfide isomerase independent of oxido-reductase activity. *J.Biol.Chem.* 2007
19. Pendurthi UR, Ghosh S, Mandal SK, Rao LV. Tissue factor activation: is disulfide bond switching a regulatory mechanism? *Blood* 2007
20. Hynes RO. Integrins: bidirectional, allosteric signaling machines. *Cell* 2002;110:673-687.
21. Dorfleutner A, Hintermann E, Tarui T, Takada Y, Ruf W. Cross-talk of integrin alpha3beta1 and tissue factor in cell migration. *Mol.Biol.Cell* 2004;15:4416-4425.
22. Bogdanov VY, Balasubramanian V, Hathcock J et al. Alternatively spliced human tissue factor: a circulating, soluble, thrombogenic protein. *Nat.Med.* 2003;9:458-462.
23. Censarek P, Bobbe A, Grandoch M, Schror K, Weber AA. Alternatively spliced human tissue factor (asHTF) is not pro-coagulant. *Thromb.Haemost.* 2007;97:11-14.

Chapter 1

24. Hobbs JE, Zakarija A, Cundiff DL et al. Alternatively spliced human tissue factor promotes tumor growth and angiogenesis in a pancreatic cancer tumor model. *Thromb.Res.* 2007;120 Suppl 2:S13-S21.