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Modulation of HDL metabolism : studies in APOE*3-Leiden.CETP mice

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Chapter 9

GENERAL DISCUSSION AND FUTURE PERSPECTIVES

Cardiovascular disease (CVD) is a major cause of morbidity and mortality in the western world, and is mainly caused by atherosclerosis. In the Netherlands, about one third of all deaths are due to CVD. Dyslipidemia (i.e. high plasma (V)LDL-cholesterol (C), high triglycerides and low HDL-C) is a major risk factor for atherosclerosis development and cardiovascular disease. Patients with dyslipidemia are usually treated with cholesterol lowering drugs including statins¹ and fibrates.² These drugs lower plasma cholesterol very efficiently (up to 40%),^{1,2} however, they prevent only a fraction (about 30%) of cardiovascular events. Therefore new therapeutic strategies to reduce CVD risk more efficiently are necessary. Since HDL is clearly inversely correlated with CVD risk, and has been attributed multiple protective effects in atherosclerosis by its role in reverse cholesterol transport (RCT) and its anti-inflammatory, anti-oxidative and anti-thrombotic properties, HDL-raising therapy is currently considered as a promising strategy to further reduce CVD risk.^{3,4}

The research described in this thesis was performed to elucidate the mechanisms underlying the HDL-C raising effects of classic lipid-lowering drugs (fenofibrate, atorvastatin, niacin) as well as an experimental HDL-raising compound (torcetrapib), and to evaluate the HDL-raising potential of novel strategies (PXR agonism, apoCI). The major conclusions and implications of our findings and future perspectives will be discussed here.

HDL modulation by classical lipid lowering drugs

The classical lipid-lowering drugs statins, fibrates and niacin have been shown to modestly raise HDL-C levels in humans up to 10, 15, and 30%, respectively.⁵ Interestingly, these drugs failed to show human-like HDL-increasing effects in either normolipidemic or classical hyperlipidemic mice (i.e. LDL receptor-knockout and apoE-knockout mice). Although fenofibrate has been shown to increase HDL in wild-type mice, the raised HDL had an increased particle size, as opposed to the raised HDL in humans. Likewise, although these drugs did evoke human-like lipid-lowering effects in ApoE*3Leiden transgenic (E3L) mice⁶ with respect to dose-dependent decreases of (V)LDL-C and triglycerides, they generally failed to raise HDL-C in E3L mice.⁷

As mice naturally lack the cholesteryl ester transfer protein (CETP), which is an important determinant of HDL metabolism in humans, we reasoned that the HDL-raising effect of the classical lipid-lowering drugs may relate to the presence of CETP. Therefore, we crossbred E3L mice with CETP mice to generate the novel E3L.CETP mouse model.⁸ In this thesis we show that E3L.CETP mice indeed respond with an increase in HDL besides a decrease in (V)LDL to both a fibrate (fenofibrate; chapter 2), a statin (atorvastatin; chapter 3) and niacin (chapter 4). This indicates not only that the E3L.CETP mouse is a valuable model to study the effect of HDL modulating drugs on lipid

metabolism, but also that CETP plays an essential role in the HDL increase observed with these drugs.

It is interesting to note that these various classes of drugs have a similar HDL-increasing effect, as dependent on CETP expression, whereas they evoke their lipid-lowering effects through different mechanisms. Statins, fibrates and niacin reduce plasma lipids by primarily by inhibition of the *de novo* hepatic cholesterol synthesis,^{9,10} stimulation of triglyceride hydrolysis in plasma,¹¹⁻¹³ and inhibition of lipolysis in adipocytes,¹⁴ respectively. Despite these different primary actions on lipid metabolism, our studies demonstrated that they all reduce the activity and mass of CETP in plasma. In fact, we showed that atorvastatin, fenofibrate and niacin all reduced the hepatic mRNA expression of CETP, which is likely the main causal factor for the reduction in plasma CETP. In line with this hypothesis, increasing the hepatic cholesterol content of CETP transgenic mice by cholesterol feeding increases hepatic CETP expression via LXR-dependent mechanisms as well as plasma CETP mass.^{15,16} The three classes of lipid-lowering drugs not only decrease CETP expression, but also decrease (V)LDL levels. Since (V)LDL is an acceptor of HDL-derived cholesteryl esters and, therefore, also a driving force for CETP activity, the decrease in (V)LDL adds to the drug-induced reduction in CETP activity.

Based on our observations, we speculate that lipid-lowering drugs in general will thus all increase HDL-C to a certain extent by reducing plasma CETP activity, as a result of a reduction in both hepatic CETP expression and plasma (V)LDL. In fact, observations in human subjects indeed showed that statins and fibrates both reduce CETP mass and activity.¹⁷⁻²¹ The effect of niacin on plasma CETP in humans has not been reported as yet, but niacin will thus probably also reduce CETP mass and activity. Interestingly, hyperlipidemic patients carrying the CETP TaqIB1 polymorphism, who have therefore higher plasma CETP levels than people with the TaqIB2 variant, benefit more from statin treatment with respect to the development of coronary atherosclerosis,²² suggesting that the reduction of CETP activity is indeed a relevant contributor to the protective effects of lipid-lowering drugs in atherosclerosis.

The apparent robust causal relation between hepatic lipid content and hepatic CETP expression raises the question whether the pathological condition of hepatic steatosis would be a causal factor for reducing HDL-levels by increasing CETP expression. Interestingly, a recent study in obese subjects showed that liver fat indeed inversely correlated with HDL levels.²³ However, since (V)LDL levels are also increased in patients with a fatty liver,²³ more research is needed to confirm the hypothesis that hepatic steatosis results in increased plasma CETP levels thereby decreasing HDL.

HDL modulation by CETP inhibition

CETP inhibition has been regarded as a novel promising HDL-C raising strategy to reduce atherosclerosis and cardiovascular disease. Large clinical studies with the CETP inhibitor torcetrapib have been performed to evaluate whether CETP inhibition is able to increase HDL levels and reduce atherosclerosis. Torcetrapib indeed increased HDL-C to a marked extent (+60% at 60 mg/day), but did not reduce atherosclerosis as measured by Intima Media Thickness (IMT) and Intravascular Ultrasound (IVUS)²⁴⁻²⁶ and had unwanted effects including increased overall mortality as well as fatal and non fatal cardiovascular events.²⁷

There are several possible explanations for the disappointing results of the torcetrapib studies, related to 1) inclusion criteria of the patients, 2) combination therapy with atorvastatin, 3) properties of the newly formed HDL, or 4) compound specific effects.

First, in the torcetrapib studies, patients were included who had undergone cardiac catheterization²⁶ or have (familial) dyslipidemia.^{24,25} These broad inclusion criteria led to the inclusion of a very heterogeneous patient group that may not all benefit from CETP inhibition, as the metabolic context, including baseline CETP activity, HDL-C levels and TG levels, is likely to be an important determinant of the effect of CETP inhibition on atherosclerosis development and CVD risk. The view that the metabolic context is important in the effect of CETP inhibition on atherosclerosis is supported by mouse studies, which in general show that CETP expression is atheroprotective in mice with increased HDL levels while CETP expression is atherogenic in mice with elevated (V)LDL levels. CETP expression in mice with high HDL because of LCAT overexpression or SR-BI deficiency reduces atherosclerosis.^{28,29} In line with these mouse data, it has been shown that subjects with highly elevated HDL-C levels which were mainly associated with CETP mutations, have higher prevalence of ischemic ECG changes.³⁰ CETP inhibition in subjects who have already high HDL may, therefore, not lead to protection against atherosclerosis and CVD. These findings may also suggest that HDL elevation by CETP inhibition is only protective when this normalizes or elevates HDL mildly compared to normolipidemic subjects and CETP inhibitors should not be used to further increase HDL in subjects who have cardiovascular risk factors but normal or elevated HDL levels. In contrast, CETP is a clear atherogenic factor in hyperlipidemic mouse models with impaired (V)LDL clearance, including apoE knockout,³¹ LDL receptor knockout³¹ and E3L mice.⁸ When translated to humans, dyslipidemic subjects with high plasma TG may therefore benefit from CETP inhibition since CETP transfers HDL-CE to (V)LDL that is inefficiently cleared from plasma as indicated by high TG. This suggests that CETP inhibition could be particularly promising in those dyslipidemic patients, who besides elevated (V)LDL also have low HDL and/or high CETP activity.

Further subgroup analyses should be performed in the recent torcetrapib studies to study whether patients with low HDL, high CETP activity and/or high TG at baseline benefit from torcetrapib with respect to atherosclerosis development.

Second, the clinical studies with torcetrapib were all performed in combination with atorvastatin,²⁴⁻²⁶ which by itself reduces CETP activity in plasma.^{17-20,32,33}

By investigating the effect of torcetrapib on atherosclerosis development in E3L.CETP mice, either in combination with atorvastatin or alone (chapter 5), we showed that torcetrapib *per se* did in fact reduce atherosclerosis while torcetrapib did not reduce atherosclerosis in mice that were also treated with atorvastatin, indicating that combination treatment with atorvastatin and other lipid lowering drugs may attenuate or mask the effect of torcetrapib on atherosclerosis.

Third, it is not clear whether the increased HDL as induced by CETP inhibition by torcetrapib is atheroprotective or not. HDL is thought to be protective in atherosclerosis via mediating RCT and by its proposed anti-inflammatory, anti-oxidative and anti-thrombotic properties. Torcetrapib alters HDL by increasing its size, by the formation of a torcetrapib/CETP complex that associates with HDL, and torcetrapib may alter the protein composition of HDL which may alter HDL functionality. In addition, torcetrapib may reduce HDL-CE clearance as it has been described that in humans, the majority of HDL-CE reaches the liver via (V)LDL after CETP-mediated transfer of HDL-CE from HDL to (V)LDL, and this pathway is blocked by torcetrapib.^{34,35}

Fourth, torcetrapib may have had compound-specific adverse side effects with respect to increased mortality and increased cardiovascular events, and compound-specific side effects may explain why torcetrapib did not add to the atherosclerosis-reducing effect of atorvastatin. Torcetrapib has been found to increase blood pressure by approx. 5 mm Hg,^{24-26,36} which is not observed with novel CETP inhibitors including anacetrapib and JTT-705. This mild increase in blood pressure is unlikely to completely have counteracted potential protective effects of the HDL increase and is unlikely to have caused increased mortality, but it may be indicative for other underlying adverse effects. Analysis of plasma samples from patients treated with torcetrapib showed an increase in sodium, bicarbonate and aldosterone levels.²⁷ Torcetrapib also increased plasma aldosterone in E3L.CETP mice (chapter 5). The raise in aldosterone may not only explain the increase in blood pressure, but animal studies have also shown that aldosterone causally increases atherosclerosis, inflammation and oxidative stress,³⁷⁻⁴⁰ indicating that the increase in aldosterone may have counteracted potentially atheroprotective effects of the torcetrapib-induced increase in HDL. We have demonstrated that torcetrapib increases the macrophage to collagen ratio within atherosclerotic plaques in E3L.CETP mice (chapter 5). Whereas plaques of mice do not easily rupture spontaneously, such a phenotype is considered more prone to spontaneous rupture in humans. Extrapolation of our data to humans may thus suggest that the increase in cardiovascular events and

death may have been caused, at least partly, by increased incidence of plaque rupture. As aldosterone is associated with more inflammation and increased activity of matrix metalloproteinases (MMP),^{39,41} which cause breakdown of collagen, it can be reasoned that the increased aldosterone levels may have contributed to the plaque phenotype. However, this hypothesis should be underscored by experimental studies. The effects of torcetrapib on aldosterone and blood pressure are compound-specific and independent of CETP, as torcetrapib induced these effects in both humans, CETP transgenic mice and non-transgenic mice.⁴² In contrast, anacetrapib does not increase blood pressure and aldosterone levels.^{42,43} The effect of novel CETP inhibitors on plaque composition and cardiovascular events is thus eagerly awaited.

In addition to chemical CETP inhibitors, endogenous CETP inhibitors have also been described. Whereas the lipid transfer inhibitor protein (LTI) as present on LDL inhibits CETP activity,⁴⁴ apoCI is the major endogenous CETP inhibitor present on HDL.⁴⁵ being an endogenous protein, apoCI may be a lead for novel safe CETP inhibitors. However, apoCI is also an inhibitor of LPL.⁴⁶ Because of apoCI-induced LPL inhibition, human apoCI overexpression in CETP transgenic mice not only reduces specific CETP activity, but also largely increases VLDL levels. The increase in VLDL levels consequently increases hepatic CETP gene expression, which precludes an increase in HDL-C resulting from CETP inhibition only.⁴⁷

To find an apoCI derived CETP inhibitor without LPL inhibitory properties, we performed structure-function analysis using an array of apoCI derived peptides. ApoCI₃₂₋₅₇ was the most efficient CETP inhibitory peptide tested, and this peptide had only minimal effects on LPL. Therefore we expect that this peptide should be able to increase HDL without inducing hyperlipidemia. Pilot experiments showed that intravenous injections with apoCI and apoCI₃₂₋₅₇ were unable to modulate lipid levels *in vivo* in CETP transgenic mice (unpublished data), which may relate to dosing and/or adverse pharmacokinetics of the peptides. Short-term elevation of plasma levels of apoCI and apoCI₃₂₋₅₇, e.g. by using recombinant adenoviruses that induce a relatively high hepatic protein expression of apoCI and apoCI₃₂₋₅₇ in CETP expressing mice could therefore be useful as a tool to show the potential of apoCI₃₂₋₅₇ to increase HDL levels without affecting plasma TG. If so, apoCI₃₂₋₅₇ mimicking agents could be developed that can be used orally.

In addition to CETP, apoCI also affects other HDL modifying enzymes, including LCAT^{48,49} HL^{50,51} and SR-BI (chapter 7). These combined actions increase HDL in naturally CETP-deficient wild-type mice (chapter 7), but functionality of the increased HDL is unknown. It is also unknown how apoCI₃₂₋₅₇ would act on these various proteins involved in HDL metabolism. Therefore, additional *in vitro* and *in vivo* experiments in wild-type mice should be performed to show whether potential HDL-increasing effects of apoCI₃₂₋₅₇

are CETP dependent. In addition, studies in E3L.CETP mice will be useful to reveal whether apoCI₃₂₋₅₇ will reduce atherosclerosis. It would be interesting to study the effect of apoCI₃₂₋₅₇ on the plaque phenotype as well as to evaluate whether the effect of torcetrapib on plaque composition was indeed compound specific, especially because apoCI has a different mechanism to inhibit CETP as compared to torcetrapib, i.e. reduction of affinity of CETP with HDL⁵² versus formation of an inactive HDL/CETP complex.⁵³

Novel strategies to reduce cardiovascular disease

Cholesterol lowering is a proven effective strategy to reduce cardiovascular disease. Therefore a large number of new drugs to reduce plasma lipid levels is under development, including microsomal triglyceride transfer protein (MTP) inhibitors, squalene synthase inhibitors, and apoB expression inhibitors that are all aimed at reducing lipid production by the liver.⁵⁴⁻⁵⁸ However, as statins already efficiently reduce plasma (V)LDL-C levels without severe side effects, and additional safe (V)LDL-C lowering agents to treat patients that do not respond well to statins are available (e.g. cholesterol binding resins) it will be difficult to develop novel lipid lowering drugs that lead to more clinical benefit with respect to protection against cardiovascular disease.

After the disappointing results from the torcetrapib studies, it is difficult to predict whether novel HDL raising agents will prevent CVD in the future. Despite that there is no direct evidence for the protective effect of HDL in atherosclerosis, HDL is thought to play a role in RCT, and is thought to be anti-inflammatory, anti-oxidative and anti-thrombotic. HDL raising may be achieved by 1) reducing HDL clearance, 2) increasing HDL maturation and modification of HDL metabolism or 3) enhancing HDL production.

In mice, clearance of HDL-C is almost exclusively mediated via SR-BI. In humans, Cla-1 (i.e. the human orthologue of SR-BI) is thought to play a less important role, since HDL-CE is mainly cleared from plasma after CETP-mediated transfer to (V)LDL.³⁴ Albeit that raising HDL by reducing its clearance may increase the anti-inflammatory and anti-oxidative properties of HDL, the role of HDL in RCT (i.e. transfer of cholesterol to the liver, followed by excretion via the feces) is possibly the most important protective function of HDL. Together with the fact that SR-BI-deficiency in mice aggravates atherosclerosis, reducing HDL clearance may probably not be the most valid HDL-raising strategy.

LCAT plays an important role in the maturation of HDL as this enzyme esterifies HDL-associated cholesterol into CE. Mutations in LCAT which lower LCAT activity reduce HDL-C levels and mildly enhance atherosclerosis development (as measured by IMT).⁵⁹ However, since overexpression of LCAT in mice increases atherosclerosis, LCAT-targeted interventions to raise HDL should be pursued with care. As niacin increases HDL via CETP reduction

(chapter 4) niacin can also be considered as a compound that increases HDL via HDL modulation. Niacin is however not well tolerated because it induces severe flushing. A recent study shows that addition of the prostaglandin D₂ receptor 1 blocker laropiprant reduces niacin mediated flushing which may increase niacin use.⁶⁰ In addition novel compounds targeting the niacin receptor GPR109A are under development.⁶¹ However, it is uncertain whether the protective effects of niacin on IMT progression and mortality are mediated via its HDL raising or via its (V)LDL reducing properties^{62,63}

HDL production is presumably mainly initiated by the synthesis and secretion of apoAI by the liver and intestine, and lipidation of apoAI in plasma via ABCA1. Indeed, apoAI-deficiency and ABCA1-deficiency in mice largely decrease HDL-C. Interestingly, humans carrying mutations in ABCA1 and apoAI⁵⁹ have a more severe increase in atherosclerosis (as measured by IMT) as compared to carriers of CETP or LCAT mutations, suggesting that enhancing HDL production would be the most promising strategy in the prevention of atherosclerosis and CVD. Upregulation of ABCA1 in macrophages can be achieved with compounds like LXR agonists.⁶⁴ However, these compounds will also induce the expression of lipogenic genes in the liver which counteract potential protective effects of ABCA1 upregulation in atherosclerosis. In addition, LXR activation will result in upregulation of multiple proteins involved in cholesterol efflux, including apoE, which hampers evaluation of the effect of upregulation of ABCA1 only. Therefore, compounds that specifically upregulate ABCA1 expression in macrophages, preferably in the atherosclerotic vessel wall, should be developed. Increase of apoAI can be achieved via administration of apoAI or apoAI mimicking compounds, or via upregulation of apoAI expression. ApoAI, infused either as a lipid-free protein or contained in recombinant HDL, increases HDL levels. Clinical studies show that short-term apoAI infusion lead to a quick reduction of atheroma volumes in patients with acute coronary syndrome.⁶⁵ However, a drawback of apoAI infusion is that treatment is invasive and very expensive, and that long term effects are still unknown.⁶⁶ Alternatively, apoAI mimicking agents have been developed which can be given orally.⁶⁷ Agents that increase apoAI production would also be promising to increase endogenous HDL production. It has been suggested from studies in wild-type mice that PXR agonism would increase apoAI production and raise HDL levels.⁶⁸ However, we observed in our more human-like E3L.CETP mice that PXR agonists reduced HDL of all sizes concomitant with a reduction (rather than an increase) in apoAI expression (chapter 6). This study thus indicates that PXR agonism is likely also unable to raise apoAI and HDL in humans. Recently, rvx-208 has been developed to induce apoAI expression and raise HDL levels, as shown in mice and non-human primates. Provided that this compound is specific for apoAI, it would be promising new lead in the ongoing search for HDL-raising strategies to prevent CVD.⁶⁹

Concluding remarks

Besides a large number of new lipid-lowering agents, drugs that are aimed to specifically raise HDL are currently under development. The therapeutic value of such HDL-raising therapies, however, is still unclear, especially since the first HDL-raising strategy by torcetrapib failed in large human trials. The question whether raising HDL will add to the atheroprotective effect of lipid-lowering therapy is thus still unanswered. Albeit that high HDL-C is clearly associated with reduced CVD risk, high HDL-C is also associated with low (V)LDL-C and TG, a balance that is probably dictated by bidirectional transfer of neutral lipids by CETP. Therefore, it is difficult to predict the importance of HDL independent of other risk factors, and virtually no direct evidence is currently available to show that HDL *per se* is protective in atherosclerosis development. Therefore, other therapeutic strategies such as inhibition of inflammation, which besides dyslipidemia is also a driving force for atherosclerosis, should not be overlooked.

The torcetrapib trials taught us that care should be taken in selecting appropriate subjects in human studies that are expected to benefit most from a novel experimental approach to reduce CVD. Future studies will show which of these new strategies will eventually be used in the future therapy of those patients that are prone to develop CVD. Moreover, our studies indicate the importance of testing effects of experimental drugs on lipid metabolism, atherosclerosis and plaque composition in appropriate animal models. We expect that our newly developed E3L.CETP mouse model with a human-like lipoprotein metabolism will largely contribute to the development of such compounds by reliably predicting human responses to experimental drugs and revealing underlying mechanisms.

References

1. Baigent C, Keech A, Kearney PM, Blackwell L, Buck G, Pollicino C, Kirby A, Sourjina T, Peto R, Collins R, Simes R. Efficacy and safety of cholesterol-lowering treatment: prospective meta-analysis of data from 90,056 participants in 14 randomised trials of statins. *Lancet*. 2005; 366:1267-1278.
2. Saha SA, Kizhakepunnur LG, Bahekar A, Arora RR. The role of fibrates in the prevention of cardiovascular disease--a pooled meta-analysis of long-term randomized placebo-controlled clinical trials. *Am.Heart J.* 2007; 154:943-953.
3. Gordon DJ, Probstfield JL, Garrison RJ, Neaton JD, Castelli WP, Knoke JD, Jacobs DR, Jr., Bangdiwala S, Tyroler HA. High-density lipoprotein cholesterol and cardiovascular disease. Four prospective American studies. *Circulation*. 1989; 79:8-15.
4. Chapman MJ. Therapeutic elevation of HDL-cholesterol to prevent atherosclerosis and coronary heart disease. *Pharmacol.Ther.* 2006; 111:893-908.
5. Singh IM, Shishehbor MH, Ansell BJ. High-density lipoprotein as a therapeutic target: a systematic review. *JAMA*. 2007; 298:786-798.

6. van den Maagdenberg AM, Hofker MH, Krimpenfort PJ, de B, I, van Vlijmen B, van der BH, Havekes LM, Frants RR. Transgenic mice carrying the apolipoprotein E3-Leiden gene exhibit hyperlipoproteinemia. *J.Biol.Chem.* 1993; 268:10540-10545.
7. Zadelaar S, Kleemann R, Verschuren L, de Vries-Van der Weij, van der HJ, Princen HM, Kooistra T. Mouse models for atherosclerosis and pharmaceutical modifiers. *Arterioscler.Thromb.Vasc.Biol.* 2007; 27:1706-1721.
8. Westerterp M, van der Hoogt CC, de Haan W, Offerman EH, Dallinga-Thie GM, Jukema JW, Havekes LM, Rensen PC. Cholesteryl ester transfer protein decreases high-density lipoprotein and severely aggravates atherosclerosis in APOE*3-Leiden mice. *Arterioscler.Thromb.Vasc.Biol.* 2006; 26:2552-2559.
9. Endo A, Kuroda M, Tanzawa K. Competitive inhibition of 3-hydroxy-3-methylglutaryl coenzyme A reductase by ML-236A and ML-236B fungal metabolites, having hypocholesterolemic activity. *FEBS Lett.* 1976; 72:323-326.
10. Istvan ES and Deisenhofer J. Structural mechanism for statin inhibition of HMG-CoA reductase. *Science.* 2001; 292:1160-1164.
11. Staels B, Vu-Dac N, Kosykh VA, Saladin R, Fruchart JC, Dallongeville J, Auwerx J. Fibrates downregulate apolipoprotein C-III expression independent of induction of peroxisomal acyl coenzyme A oxidase. A potential mechanism for the hypolipidemic action of fibrates. *J.Clin.Invest.* 1995; 95:705-712.
12. Malmendier CL, Lontie JF, Delcroix C, Dubois DY, Magot T, De Roy L. Apolipoproteins C-II and C-III metabolism in hypertriglyceridemic patients. Effect of a drastic triglyceride reduction by combined diet restriction and fenofibrate administration. *Atherosclerosis.* 1989; 77:139-149.
13. Schoonjans K, Staels B, Auwerx J. The peroxisome proliferator activated receptors (PPARS) and their effects on lipid metabolism and adipocyte differentiation. *Biochim.Biophys.Acta.* 1996; 1302:93-109.
14. Bodor ET and Offermanns S. Nicotinic acid: an old drug with a promising future. *Br.J.Pharmacol.* 2008; 153 Suppl 1:S68-75. Epub@2007 Nov 26.:S68-S75.
15. Luo Y and Tall AR. Sterol upregulation of human CETP expression in vitro and in transgenic mice by an LXR element. *J.Clin.Invest* 2000; 105:513-520.
16. Jiang XC, Agellon LB, Walsh A, Breslow JL, Tall A. Dietary cholesterol increases transcription of the human cholesteryl ester transfer protein gene in transgenic mice. Dependence on natural flanking sequences. *J.Clin.Invest.* 1992; 90:1290-1295.
17. Ahnadi CE, Berthezene F, Ponsin G. Simvastatin-induced decrease in the transfer of cholesterol esters from high density lipoproteins to very low and low density lipoproteins in normolipidemic subjects. *Atherosclerosis* 1993; 99:219-228.
18. Guerin M, Lassel TS, Le Goff W, Farnier M, Chapman MJ. Action of atorvastatin in combined hyperlipidemia : preferential reduction of cholesteryl ester transfer from HDL to VLDL1 particles. *Arterioscler.Thromb.Vasc.Biol.* 2000; 20:189-197.
19. Guerin M, Dolphin PJ, Talussot C, Gardette J, Berthezene F, Chapman MJ. Pravastatin modulates cholesteryl ester transfer from HDL to apoB-containing lipoproteins and lipoprotein subspecies profile in familial hypercholesterolemia. *Arterioscler.Thromb.Vasc.Biol.* 1995; 15:1359-1368.
20. Guerin M, Egger P, Soudant C, Le Goff W, Van Tol A, Dupuis R, Chapman MJ. Dose-dependent action of atorvastatin in type IIB hyperlipidemia: preferential and progressive reduction of atherogenic apoB-containing lipoprotein subclasses (VLDL-2, IDL, small dense LDL) and stimulation of cellular cholesterol efflux. *Atherosclerosis.* 2002; 163:287-296.
21. Guerin M, Bruckert E, Dolphin PJ, Turpin G, Chapman MJ. Fenofibrate reduces plasma cholesteryl ester transfer from HDL to VLDL and normalizes the atherogenic, dense

- LDL profile in combined hyperlipidemia. *Arterioscler.Thromb.Vasc.Biol.* 1996; 16:763-772.
22. Kuivenhoven JA, Jukema JW, Zwinderman AH, de Knijff P, McPherson R, Bruschke AV, Lie KI, Kastelein JJ. The role of a common variant of the cholesteryl ester transfer protein gene in the progression of coronary atherosclerosis. The Regression Growth Evaluation Statin Study Group. *N.Engl.J.Med.* 1998; 338:86-93.
 23. Cali AM, Zern TL, Taksali SE, de Oliveira AM, Dufour S, Otvos JD, Caprio S. Intrahepatic fat accumulation and alterations in lipoprotein composition in obese adolescents: a perfect proatherogenic state. *Diabetes Care.* 2007; 30:3093-3098.
 24. Kastelein JJ, van Leuven SI, Burgess L, Evans GW, Kuivenhoven JA, Barter PJ, Revkin JH, Grobbee DE, Riley WA, Shear CL, Duggan WT, Bots ML. Effect of torcetrapib on carotid atherosclerosis in familial hypercholesterolemia. *N.Engl.J.Med.* 2007; 356:1620-1630.
 25. Bots ML, Visseren FL, Evans GW, Riley WA, Revkin JH, Tegeler CH, Shear CL, Duggan WT, Vicari RM, Grobbee DE, Kastelein JJ. Torcetrapib and carotid intima-media thickness in mixed dyslipidaemia (RADIANCE 2 study): a randomised, double-blind trial. *Lancet.* 2007; 370:153-160.
 26. Nissen SE, Tardif JC, Nicholls SJ, Revkin JH, Shear CL, Duggan WT, Ruzyllo W, Bachinsky WB, Lasala GP, Tuzcu EM. Effect of torcetrapib on the progression of coronary atherosclerosis. *N.Engl.J.Med.* 2007; 356:1304-1316.
 27. Barter PJ, Caulfield M, Eriksson M, Grundy SM, Kastelein JJ, Komajda M, Lopez-Sendon J, Mosca L, Tardif JC, Waters DD, Shear CL, Revkin JH, Buhr KA, Fisher MR, Tall AR, Brewer B. Effects of torcetrapib in patients at high risk for coronary events. *N.Engl.J.Med.* 2007; 357:2109-2122.
 28. Harder C, Lau P, Meng A, Whitman SC, McPherson R. Cholesteryl ester transfer protein (CETP) expression protects against diet induced atherosclerosis in SR-BI deficient mice. *Arterioscler.Thromb.Vasc.Biol.* 2007; 27:858-864.
 29. Foger B, Chase M, Amar MJ, Vaisman BL, Shamburek RD, Paigen B, Fruchart-Najib J, Paiz JA, Koch CA, Hoyt RF, Brewer HB, Jr., Santamarina-Fojo S. Cholesteryl ester transfer protein corrects dysfunctional high density lipoproteins and reduces aortic atherosclerosis in lecithin cholesterol acyltransferase transgenic mice. *J.Biol.Chem.* 1999; 274:36912-36920.
 30. Hirano K, Yamashita S, Nakajima N, Arai T, Maruyama T, Yoshida Y, Ishigami M, Sakai N, Kameda-Takemura K, Matsuzawa Y. Genetic cholesteryl ester transfer protein deficiency is extremely frequent in the Omagari area of Japan. Marked hyperalphalipoproteinemia caused by CETP gene mutation is not associated with longevity. *Arterioscler.Thromb.Vasc.Biol.* 1997; 17:1053-1059.
 31. Plump AS, Masucci-Magoulas L, Bruce C, Bisgaier CL, Breslow JL, Tall AR. Increased atherosclerosis in ApoE and LDL receptor gene knock-out mice as a result of human cholesteryl ester transfer protein transgene expression. *Arterioscler.Thromb.Vasc.Biol.* 1999; 19:1105-1110.
 32. de Haan W, van der Hoogt CC, Westerterp M, Hoekstra M, Dallinga-Thie GM, Princen HM, Romijn JA, Jukema JW, Havekes LM, Rensen PC. Atorvastatin increases HDL cholesterol by reducing CETP expression in cholesterol-fed APOE*3-Leiden.CETP mice. *Atherosclerosis.* 2008; 197:57-63.
 33. Lagrost L, Athias A, Lemort N, Richard JL, Desrumaux C, Chatenet-Duchene L, Courtois M, Farnier M, Jacotot B, Braschi S, Gambert P. Plasma lipoprotein distribution and lipid transfer activities in patients with type IIb hyperlipidemia treated with simvastatin. *Atherosclerosis.* 1999; 143:415-425.

34. Schwartz CC, VandenBroek JM, Cooper PS. Lipoprotein cholesteryl ester production, transfer, and output in vivo in humans. *J.Lipid Res.* 2004; 45:1594-1607.
35. Goldberg DI, Beltz WF, Pittman RC. Evaluation of pathways for the cellular uptake of high density lipoprotein cholesterol esters in rabbits. *J.Clin.Invest.* 1991; 87:331-346.
36. Davidson MH, McKenney JM, Shear CL, Revkin JH. Efficacy and safety of torcetrapib, a novel cholesteryl ester transfer protein inhibitor, in individuals with below-average high-density lipoprotein cholesterol levels. *J.Am.Coll.Cardiol.* 2006; 48:1774-1781.
37. Hirono Y, Yoshimoto T, Suzuki N, Sugiyama T, Sakurada M, Takai S, Kobayashi N, Shichiri M, Hirata Y. Angiotensin II receptor type 1-mediated vascular oxidative stress and proinflammatory gene expression in aldosterone-induced hypertension: the possible role of local renin-angiotensin system. *Endocrinology.* 2007; 148:1688-1696.
38. Hashikabe Y, Suzuki K, Jojima T, Uchida K, Hattori Y. Aldosterone impairs vascular endothelial cell function. *J.Cardiovasc.Pharmacol.* 2006; 47:609-613.
39. Suzuki J, Iwai M, Mogi M, Oshita A, Yoshii T, Higaki J, Horiuchi M. Eplerenone with valsartan effectively reduces atherosclerotic lesion by attenuation of oxidative stress and inflammation. *Arterioscler.Thromb.Vasc.Biol.* 2006; 26:917-921.
40. Keidar S, Hayek T, Kaplan M, Pavlotzky E, Hamoud S, Coleman R, Aviram M. Effect of eplerenone, a selective aldosterone blocker, on blood pressure, serum and macrophage oxidative stress, and atherosclerosis in apolipoprotein E-deficient mice. *J.Cardiovasc.Pharmacol.* 2003; 41:955-963.
41. Rude MK, Duhaney TA, Kuster GM, Judge S, Heo J, Colucci WS, Siwik DA, Sam F. Aldosterone stimulates matrix metalloproteinases and reactive oxygen species in adult rat ventricular cardiomyocytes. *Hypertension.* 2005; 46:555-561.
42. Forrest MJ, Bloomfield D, Briscoe RJ, Brown PN, Cumiskey AM, Ehrhart J, Hershey JC, Keller WJ, Ma X, McPherson HE, Messina E, Peterson LB, Sharif-Rodriguez W, Siegl PK, Sinclair PJ, Sparrow CP, Stevenson AS, Sun SY, Tsai C, Vargas H, Walker M, III, West SH, White V, Woltmann RF. Torcetrapib-induced blood pressure elevation is independent of CETP inhibition and is accompanied by increased circulating levels of aldosterone. *Br.J.Pharmacol.* 2008; 154:1465-1473.
43. Krishna R, Anderson MS, Bergman AJ, Jin B, Fallon M, Cote J, Rosko K, Chavez-Eng C, Lutz R, Bloomfield DM, Gutierrez M, Doherty J, Bieberdorf F, Chodakewitz J, Gottesdiener KM, Wagner JA. Effect of the cholesteryl ester transfer protein inhibitor, anacetrapib, on lipoproteins in patients with dyslipidaemia and on 24-h ambulatory blood pressure in healthy individuals: two double-blind, randomised placebo-controlled phase I studies. *Lancet.* 2007; 370:1907-1914.
44. Morton RE, Gnizak HM, Greene DJ, Cho KH, Paromov VM. Lipid transfer inhibitor protein (apolipoprotein F) concentration in normolipidemic and hyperlipidemic subjects. *J.Lipid Res.* 2008; 49:127-135.
45. Gautier T, Masson D, de Barros JP, Athias A, Gambert P, Aunis D, Metz-Boutigue MH, Lagrost L. Human apolipoprotein C-I accounts for the ability of plasma high density lipoproteins to inhibit the cholesteryl ester transfer protein activity. *J.Biol.Chem.* 2000; 275:37504-37509.
46. Berbee JF, van der Hoogt CC, Sundararaman D, Havekes LM, Rensen PC. Severe hypertriglyceridemia in human APOC1 transgenic mice is caused by apoC-I-induced inhibition of LPL. *J.Lipid Res.* 2005; 46:297-306.
47. Gautier T, Masson D, Jong MC, Pais de Barros JP, Duverneuil L, Le Guern N, Deckert V, Dumont L, Bataille A, Zak Z, Jiang XC, Havekes LM, Lagrost L. Apolipoprotein CI overexpression is not a relevant strategy to block cholesteryl ester transfer protein (CETP) activity in CETP transgenic mice. *Biochem.J.* 2005; 385:189-195.

48. Soutar AK, Garner CW, Baker HN, Sparrow JT, Jackson RL, Gotto AM, Smith LC. Effect of the human plasma apolipoproteins and phosphatidylcholine acyl donor on the activity of lecithin: cholesterol acyltransferase. *Biochemistry*. 1975; 14:3057-3064.
49. Jonas A, Sweeny SA, Herbert PN. Discoidal complexes of A and C apolipoproteins with lipids and their reactions with lecithin: cholesterol acyltransferase. *J.Biol.Chem.* 1984; 259:6369-6375.
50. Kinnunen PK and Ehnolm C. Effect of serum and C-apoproteins from very low density lipoproteins on human postheparin plasma hepatic lipase. *FEBS Lett.* 1976; 65:354-357.
51. Conde-Knape K, Bensadoun A, Sobel JH, Cohn JS, Shachter NS. Overexpression of apoC-I in apoE-null mice: severe hypertriglyceridemia due to inhibition of hepatic lipase. *J.Lipid Res.* 2002; 43:2136-2145.
52. Dumont L, Gautier T, de Barros JP, Laplanche H, Blache D, Ducoroy P, Fruchart J, Fruchart JC, Gambert P, Masson D, Lagrost L. Molecular mechanism of the blockade of plasma cholesteryl ester transfer protein by its physiological inhibitor apolipoprotein CI. *J.Biol.Chem.* 2005; 280:38108-38116.
53. Clark RW, Ruggeri RB, Cunningham D, Bamberger MJ. Description of the torcetrapib series of cholesteryl ester transfer protein inhibitors, including mechanism of action. *J.Lipid Res.* 2006; 47:537-552.
54. Samaha FF, McKenney J, Bloedon LT, Sasiela WJ, Rader DJ. Inhibition of microsomal triglyceride transfer protein alone or with ezetimibe in patients with moderate hypercholesterolemia. *Nat.Clin.Pract.Cardiovasc.Med.* 2008; 5:497-505.
55. Cuchel M, Bloedon LT, Szapary PO, Kolansky DM, Wolfe ML, Sarkis A, Millar JS, Ikewaki K, Siegelman ES, Gregg RE, Rader DJ. Inhibition of microsomal triglyceride transfer protein in familial hypercholesterolemia. *N.Engl.J.Med.* 2007; 356:148-156.
56. Charlton-Menys V and Durrington PN. Human cholesterol metabolism and therapeutic molecules. *Exp.Physiol.* 2008; 93:27-42.
57. Merki E, Graham MJ, Mullick AE, Miller ER, Crooke RM, Pitas RE, Witztum JL, Tsimikas S. Antisense oligonucleotide directed to human apolipoprotein B-100 reduces lipoprotein(a) levels and oxidized phospholipids on human apolipoprotein B-100 particles in lipoprotein(a) transgenic mice. *Circulation.* 2008; 118:743-753.
58. Crooke RM, Graham MJ, Lemonidis KM, Whipple CP, Koo S, Perera RJ. An apolipoprotein B antisense oligonucleotide lowers LDL cholesterol in hyperlipidemic mice without causing hepatic steatosis. *J.Lipid Res.* 2005; 46:872-884.
59. Hovingh GK, de Groot E, van der SW, Boekholdt SM, Hutten BA, Kuivenhoven JA, Kastelein JJ. Inherited disorders of HDL metabolism and atherosclerosis. *Curr.Opin.Lipidol.* 2005; 16:139-145.
60. Paolini JF, Mitchel YB, Reyes R, Kher U, Lai E, Watson DJ, Norquist JM, Meehan AG, Bays HE, Davidson M, Ballantyne CM. Effects of laropiprant on nicotinic acid-induced flushing in patients with dyslipidemia. *Am.J.Cardiol.* 2008; 101:625-630.
61. Deng Q, Frie JL, Marley DM, Beresis RT, Ren N, Cai TQ, Taggart AK, Cheng K, Carballo-Jane E, Wang J, Tong X, Waters MG, Tata JR, Colletti SL. Molecular modeling aided design of nicotinic acid receptor GPR109A agonists. *Bioorg.Med.Chem.Lett.* 2008; 18:4963-4967.
62. Taylor AJ, Sullenberger LE, Lee HJ, Lee JK, Grace KA. Arterial Biology for the Investigation of the Treatment Effects of Reducing Cholesterol (ARBITER) 2: a double-blind, placebo-controlled study of extended-release niacin on atherosclerosis progression in secondary prevention patients treated with statins. *Circulation.* 2004; 110:3512-3517.

63. Canner PL, Berge KG, Wenger NK, Stamler J, Friedman L, Prineas RJ, Friedewald W. Fifteen year mortality in Coronary Drug Project patients: long-term benefit with niacin. *J.Am.Coll.Cardiol.* 1986; 8:1245-1255.
64. Joseph SB and Tontonoz P. LXRs: new therapeutic targets in atherosclerosis? *Curr.Opin.Pharmacol.* 2003; 3:192-197.
65. Conca P and Franceschini G. Synthetic HDL as a new treatment for atherosclerosis regression: has the time come? *Nutr.Metab Cardiovasc.Dis.* 2008; 18:329-335.
66. Shah PK. High-density lipoprotein mimetics: focus on synthetic high-density lipoprotein. *Am.J.Cardiol.* 2007; 100:S62-S67.
67. Nicholls SJ and Nissen SE. New targets of high-density lipoprotein therapy. *Curr.Opin.Lipidol.* 2007; 18:421-426.
68. Bachmann K, Patel H, Batayneh Z, Slama J, White D, Posey J, Ekins S, Gold D, Sambucetti L. PXR and the regulation of apoA1 and HDL-cholesterol in rodents. *Pharmacol.Res.* 2004; 50:237-246.
69. Johansson J, Jahagirdar R, Genest J, Krimbou L, Chiacchia F, Wagner G, Hansen H, McLure K, Wong NC. Use of RVX-208 to increase apolipoprotein A-I and HDL in animals and phase I clinical trials. *Arterioscler.Thromb.Vasc.Biol.* 2008; 28:E32.