Genetic epidemiological approaches in complex neurological disorders
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CHAPTER 4

The 3p21.1-p21.3 hereditary vascular retinopathy locus increases the risk for Raynaud phenomenon and migraine

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Abstract

Previously, we described a large Dutch family with hereditary vascular retinopathy (HVR), Raynaud phenomenon, and migraine. A locus for HVR was mapped on chromosome 3p21.1-p21.3 but the gene has not yet been identified. The fact that all three disorders share a vascular aetiology prompted us to study whether the HVR haplotype also contributed to Raynaud phenomenon and migraine in this family. Despite of the low-powered parent-child transmission disequilibrium tests showing no significant association, the sibling transmission disequilibrium tests revealed that the HVR haplotype harbors a susceptibility factor for Raynaud phenomenon and migraine. Identification of the HVR gene may therefore improve the understanding of the pathophysiology of HVR, Raynaud phenomenon and migraine.

Keywords

Introduction

Migraine is a common neurovascular headache disorder affecting up to 18% of the general population[1,2]. The aetiology of migraine is complex, with probably various environmental and susceptibility genes involved[3,4]. Migraine can also be a part of autosomal dominant cerebrovascular syndromes, such as CADASIL and hereditary vascular retinopathy (HVR)[5-7]. Identification of genes involved in these cerebrovascular syndromes may therefore contribute to the understanding of migraine pathophysiology as well. Recently, HVR was mapped to chromosome 3p21.1-p21.3 in a large Dutch family[8]. Retinopathy in this family is characterized by microangiopathy of the retina, accompanied by micro-aneurysms and telangiectatic capillaries that appear preferentially around the macula and the posterior pole[9]. In later stages, capillary occlusions may develop leading to retinal ischemia and neovascularisation. Leukoencephalopathy was seen on MRI scans in some patients[7]. Genetic testing revealed that two additional families with autosomal dominant cerebroretinal vasculopathy (CRV) and hereditary endotheliopathy with retinopathy, nephropathy and stroke (HERNS), respectively, were also linked to this locus[10,11]. Despite the fact that all three families were linked to the locus, there was a considerable variation in clinical symptoms between the families[8,10,11]. The presence of different haplotypes suggests that the clinical variation might be related to different mutations in the same gene, although we cannot definitely exclude that different genes in the same chromosomal region may be involved in these three families.

Migraine was investigated and reported in the HERNS and HVR families[7,11]. Raynaud phenomenon, which is an episodic pathological vasomotor reaction of the digital vessels, was investigated and reported as a predominant feature in the HVR family[7]. Currently, no genes have been identified for Raynaud phenomenon, and only one linkage study for Raynaud phenomenon has been performed[12]. Several linkage and association studies have been done for migraine but no involvement of the 3p21.1-p21.3 region has been reported[13-20].

Here, we tested whether the haplotype co-segregating with HVR in the Dutch family also contributed to increased susceptibility for migraine and Raynaud phenomenon using parent-child and sibling-based transmission disequilibrium tests[21-26].

Materials and methods

Diagnosis of patients and family members

Detailed clinical information on the extended Dutch HVR family was published previously[7,8]. In total, 198 of the 289 family members were personally interviewed and information of the other individuals was obtained indirectly through relatives. Retinopathy was diagnosed by ophthalmologic examination, supplemented with fluorescence angiography of both eyes[9]. Migraine diagnosis was made based on a standard questionnaire, using the criteria of the International Headache Society (IHS)[27]. The diagnosis of Raynaud phenomenon was made according to standardized criteria of Miller et al.[28] All persons gave written informed consent and medical ethical approval was obtained from Leiden University Medical Center (LUMC).

Genotyping

For genotyping, genomic DNA was extracted from peripheral blood using a standard salting out extraction method[29]. To determine the presence of the HVR haplotype, DNA samples of 254 family members and spouses were genotyped for three genetic markers of the 3p21.1-p21.3 region; D3S3564, D3S1581 and D3S1289 [8]. All primer sequences are available through The Genome Database (http://www.gdb.org/). PCRs were performed in 15 μl reaction volume, containing 1.5 μl 10x PCR Buffer II (Applied Biosystems, Foster City, CA), 1.5 μl $MgCl_2$ (25 mM), 1.5 μl dNTPs (2.5 mM), 1.5 μl primer mix (5 pmol/μl), 5.85 μl H_2O, 0.15 μl Amplitaq Gold (5U/μl) (Applied Biosystems, Foster City, CA), and 3.0 μl of genomic DNA (15 ng/μl). The markers were amplified in two steps using 10 cycles of 30s at 94 °C, 30s at 55

°C, 30s at 72 °C followed by 25 cycles of 30s at 89 °C, 30s at 55 °C, 30s at 72 °C. PCRs were performed using a PTC-200 Thermocycler. Upon PCR, products were separated using an ABI 3700 DNA sequencer (Applied Biosystems, Foster City, CA). Genotypes were analyzed and independently scored by SKhK and KRJV using Genescan and Genotyper 2.1 software (Applied Biosystems, Foster City, CA). Haplotypes were constructed by inspection of allele segregation within the pedigree assuming a minimal number of recombinations.

Statistical analysis

A parent-child trio analysis was performed based on the principle of the transmission disequilibrium test (TDT)[21,23]. In TDT analysis the probability of transmission of the HVR haplotype is compared with the expected probability of 0.5. All trios in which a single parent is heterozygous for the HVR haplotype, and the child is affected with migraine and/or Raynaud phenomenon were selected from the HVR pedigree. Trios in which an affected child was also a heterozygous parent for a trio in a subsequent generation were excluded[30]. Accordingly, the TDT test is not biased by the fact that the trios are related. To evaluate significance, one-sided exact probabilities comparing the HVR haplotype transmission with the expected 0.50 *a priori* probability were calculated using the cumulative binomial distribution function of MS Excel 2000.

In addition to the parent-child TDT tests, sibling case-control associations were tested between the HVR haplotype and migraine or Raynaud phenomenon using the S-TDT design[22-26]. In this test the presence of the HVR haplotype is compared between sibling cases and controls from the nuclear families that make up the complete HVR pedigree. An overall statistic for the risk of carrying the HVR haplotype is subsequently calculated from the individual results of the nuclear families. From the HVR pedigree, a maximal number of nuclear families were selected in which one parent was a carrier of

the HVR haplotype. From these nuclear families all the siblings were selected for the association analysis. Risk estimates and significance were calculated with the Mantel-Haenszel extension (M-H) test, using nuclear family as stratification variable (SPSS for Windows 11.5)[25]. Furthermore, the Z' score approach implemented in the TDT/S-TDT 1.1 program was employed as a control statistic[24].

Results

Possible associations between Raynaud phenomenon, migraine and the HVR haplotype were tested in the large Dutch pedigree. Parent-affected child TDT analyses were used to test for deviations of the haplotype transmission probability, which *a priori* is 0.50 (table 1). For migraine, 23 trios and for Raynaud phenomenon, 26 trios were analyzed. Transmission probability of the HVR haplotype was only slightly increased for individuals with migraine and individuals with Raynaud phenomenon. However, the differences from the expected transmission did not reach significance.

Table 1

TDT test comparing the transmission of HVR haplotype from a heterozygous parent to offspring with Raynaud phenomenon or migraine.

Phenotype in child	HVR T+ N *(p)*	HVR T- N *(p)*	p value
Migraine (n = 23)	13 (0.57)	10 (0.44)	0.202
Raynaud phenomenon (n = 26)	16 (0.62)	10 (0.39)	0.084

HVR T+ = transmitted HVR haplotype, HVR T- = non-transmitted HVR haplotype,
N Number of children, *p* = transmission probability.

Next, sibling-control TDT tests were used to compare the risk of migraine and Raynaud phenomenon in carriers and non-carriers of the HVR haplotype. In total, 71 siblings were available from 19 nuclear families in which the haplotype was segregating (table 2). The risk of migraine was increased in HVR carriers. Of the 30 migraineurs, 13 were diagnosed with migraine without aura, 3 with migraine with aura, and 14 with mixed migraine with and without aura. Association analysis for the migraine types separately was not meaningful due to small numbers in each group. The risk of being affected with Raynaud phenomenon was also increased in carriers of the HVR haplotype (table 2).

Table 2

Associations of Raynaud phenomenon and migraine in relation to the presence of the HVR haplotype in siblings of nuclear families.

Siblings in the nuclear families	HVR+	HVR-	Odds ratio (95% CI)[a]	χ^2	p value	Z'	p value
Migraine	19	11	5.87 (1.06 - 32.60)	4.07	0.04	2.02	0.022
No migraine	15	26					
Raynaud phenomenon	25	10	11.36 (2.10 - 61.28)	11.87	0.001	2.70	0.003
No Raynaud phenomenon	9	27					

HVR+ = Hereditary Vascular Retinopathy haplotype carriers, HVR- = non Hereditary Vascular Retinopathy haplotype carriers. χ^2 = The χ^2 test of the common odds ratio equaling 1. Z' = The Z' score test of the TDT/S-TDT 1.1 program. Tests for homogeneity of the odds ratios between nuclear families were not significant for both disorders (p > 0.09).

[a] The 95% confidence interval of the Mantel-Haenszel common odds ratio.

Although there was a strong increase in risk in HVR haplotype carriers for having migraine and/or Raynaud phenomenon, the frequency of both disorders was also high in non-carriers of the HVR haplotype; 11 out of 37 had migraine and 10 out of 37 had Raynaud phenomenon. This clearly indicates that other migraine and Raynaud factors must be present in the HVR family.

Discussion

We tested whether the HVR haplotype, harboring the retinopathy gene, contributed to an increased susceptibility to migraine and Raynaud phenomenon in the Dutch family. Siblings with the HVR haplotype did show a significant increased risk of migraine and Raynaud phenomenon compared to non-carrier siblings. We found no significant increase in transmission of the haplotype with the less powerful parent-affected child TDT tests. We have provided genetic evidence that the HVR haplotype harbors a factor that increases the susceptibility for both vascular disorders in the Dutch family. However, the high incidence of these vascular diseases in non-HVR carriers suggests the presence of additional causative factors in this family.

Since retinal cerebrovascular disorders are rare, a sufficiently large sample of unrelated individuals for association analysis is difficult to obtain[8]. Testing for associations between the HVR haplotype and migraine and Raynaud phenomenon gave us a unique opportunity using a within-family approach. The fact that the rare autosomal dominant HVR haplotype is present in only one of the parents of the nuclear families allowed us to study the transmission unambiguously. Since only a single family was studied, a potential problem for testing significance is that the observations may be related[23-26]. Because transmission of the HVR haplotype from parents to offspring is random (i.e. Hardy-Weinberg equilibrium) the use of the applied parent-affected child TDT tests circumvents this potential bias. Furthermore, the results of the TDT test are independent of the disorder prevalence in controls[23]. Unfortunately, parent-child TDT tests lack sufficient detection power because of the low number of tested individuals. Moreover, small changes in the number of transmitted haplotypes have large effects on the significance of the outcome and we, therefore, tend to give less weight to these results.

For the sibling-based TDT test, taking case-control siblings from the same family provides a perfect match for confounding risk factors like population

stratification, but it may increase the risk of false-positive results[31,32]. Mantel-Haenszel statistics for related samples, and the Z' score approximation of Spielman and Ewens were used to adjust for the effects of related observations. Because the limited number of samples in each stratum may affect the p-values of the Mantel-Haenszel test, we used two tests[33].

In conclusion, we provided evidence that, within the HVR disease haplotype on chromosome 3p21.1-p21.3, a gene is present that enhances susceptibility for both Raynaud phenomenon and migraine. Future analysis will have to show whether it is the retinopathy gene itself that is associated with migraine and Raynaud phenomenon or whether it is a closely linked gene within the HVR haplotype.

Acknowledgements

References

1. Lipton RB, Stewart WF. Migraine headaches: epidemiology and comorbidity. Clin Neurosci 1998;5:2-9.
2. Launer LJ, Terwindt GM, Ferrari MD. The prevalence and characteristics of migraine in a population-based cohort: the GEM study. Neurology 1999;53:537-42.
3. Russell MB, Olesen J. Increased familial risk and evidence of genetic factor in migraine. BMJ 1995;311:541-4.
4. Russell MB, Iselius L, Olesen J. Inheritance of migraine investigated by complex segregation analysis. Hum Genet 1995;96:726-30.
5. Verin M, Rolland Y, Landgraf F et al. New phenotype of the cerebral autosomal dominant arteriopathy mapped to chromosome 19: migraine as the prominent clinical feature. J Neurol Neurosurg Psychiatry 1995;59:579-85.
6. Dichgans M, Mayer M, Uttner I et al. The phenotypic spectrum of CADASIL: clinical findings in 102 cases. Ann Neurol 1998;44:731-9.

7. Terwindt GM, Haan J, Ophoff RA, Groenen SM et al. Clinical and genetic analysis of a large Dutch family with autosomal dominant vascular retinopathy, migraine and Raynaud's phenomenon. Brain 1998;121:303-16.

8. Ophoff RA, DeYoung J, Service SK et al. Hereditary vascular retinopathy, cerebroretinal vasculopathy, and hereditary endotheliopathy with retinopathy, nephropathy, and stroke map to a single locus on chromosome 3p21.1-p21.3. Am J Hum Genet 2001;69:447-53.

9. Storimans CW, Van Schooneveld MJ, Oosterhuis JA, Bos PJ. A new autosomal dominant vascular retinopathy syndrome. Eur J Ophthalmol 1991;1:73-8.

10. Grand MG, Kaine J, Fulling K et al. Cerebroretinal vasculopathy. A new hereditary syndrome. Ophthalmology 1988;95:649-59.

11. Jen J, Cohen AH, Yue Q et al. Hereditary endotheliopathy with retinopathy, nephropathy, and stroke (HERNS). Neurology 1997;49:1322-30.

12. Susol E, MacGregor AJ, Barrett JH et al. A two-stage, genome-wide screen for susceptibility loci in primary Raynaud's phenomenon. Arthritis Rheum 2000;43:1641-6.

13. Lea RA, Shepherd AG, Curtain RP et al. A typical migraine susceptibility region localizes to chromosome 1q31. Neurogenetics 2002;4:1:17-22.

14. Nyholt DR, Dawkins JL, Brimage PJ et al. Evidence for an X-linked genetic component in familial typical migraine. Hum Mol Genet 1998;7:459-63.

15. Nyholt DR, Curtain RP, Griffiths LR. Familial typical migraine: significant linkage and localization of a gene to Xq24-28. Hum Genet 2000;107:18-23.

16. Cader ZM, Noble-Topham S, Dyment DA et al. Significant linkage to migraine with aura on chromosome 11q24. Hum Mol Genet 2003;12:2511-7.

17. Wessman M, Kallela M, Kaunisto MA et al. A susceptibility locus for migraine with aura, on chromosome 4q24. Am J Hum Genet 2002;70:652-62.

18. Bjornsson A, Gudmundsson G, Gudfinnsson E et al. Localization of a gene for migraine without aura to chromosome 4q21. Am J Hum Genet. 2003;73:986-93.

19. Carlsson A, Forsgren L, Nylander PO et al. Identification of a susceptibility locus for migraine with and without aura on 6p12.2-p21.1. Neurology 2002;59;11:1804-7.

20. Soranga D, Vettori A, Carraro G et al. A locus for migraine without aura maps on chromosome 14q21.2-q22.3. Am J Hum Genet. 2003;72:161-7.

21. Spielman RS, McGinnis RE, Ewens WJ. Transmission test for linkage disequilibrium: the insulin gene region and insulin-dependent diabetes mellitus (IDDM). Am J Hum Genet 1993;52:506-16.

22. Curtis D. Use of siblings as controls in case-control association studies. Ann Hum Genet 1997; 61:319-33.

23. Martin ER, Kaplan NL, Weir BS. Tests for linkage and association in nuclear families. Am J Hum Genet 1997;61:439-48.

24. Spielman RS, Ewens WJ. A sibship test for linkage in the presence of association: the sib transmission/disequilibrium test. Am J Hum Genet 1998;62:450-8.

25. Laird NM, Blacker D, Wilcox M. The sib transmission/disequilibrium test is a Mantel-Haenszel test. Am J Hum Genet 1998;63:1915-6

26. Horvath S, Laird NM. A discordant-sibship test for disequilibrium and linkage: no need for parental data. Am J Hum Genet 1998;63:1886-97.

27. Headache Classification Committee of the International Headache Society. Classification and diagnostic criteria for headache disorders, cranial neuralgia, and facial pain. Cephalalgia. 1988;8 Suppl 7:1-96.

28. Miller D, Waters DD, Warnica W et al. Is variant angina the coronary manifestation of a generalized vasospastic disorder? N Engl J Med 1981;304:763-6.

29. Miller SA, Dykes DD, Polesky HF. A simple salting out procedure for extracting DNA from human nucleated cells. Nucleic Acids Res 1988;16:1215.

30. Martin ER, Bass MP, Kaplan NL. Correcting for a potential bias in the pedigree disequilibrium test. Am J Hum Genet 2001;68:1065-7.

31. Witte JS, Gauderman WJ, Thomas DC. Asymptotic bias and efficiency in case-control studies of candidate genes and gene-environment interactions: basic family designs. Am J Epidemiol 1999;149:693-705.

32. Weinberg C. Asymptotic bias and efficiency in case-control studies of candidate genes and gene-environment interactions: basic family designs. Am J Epidemiol 2000;152:689-91.

33. Knapp M. Using exact P values to compare the power between the reconstruction-combined transmission/disequilibrium test and the sib transmission/disequilibrium test. Am J Hum Genet 1999;65:1208-10.