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Leiden
The Netherlands

Celiac disease : how complicated can it get?

Tjon, J.M.L.

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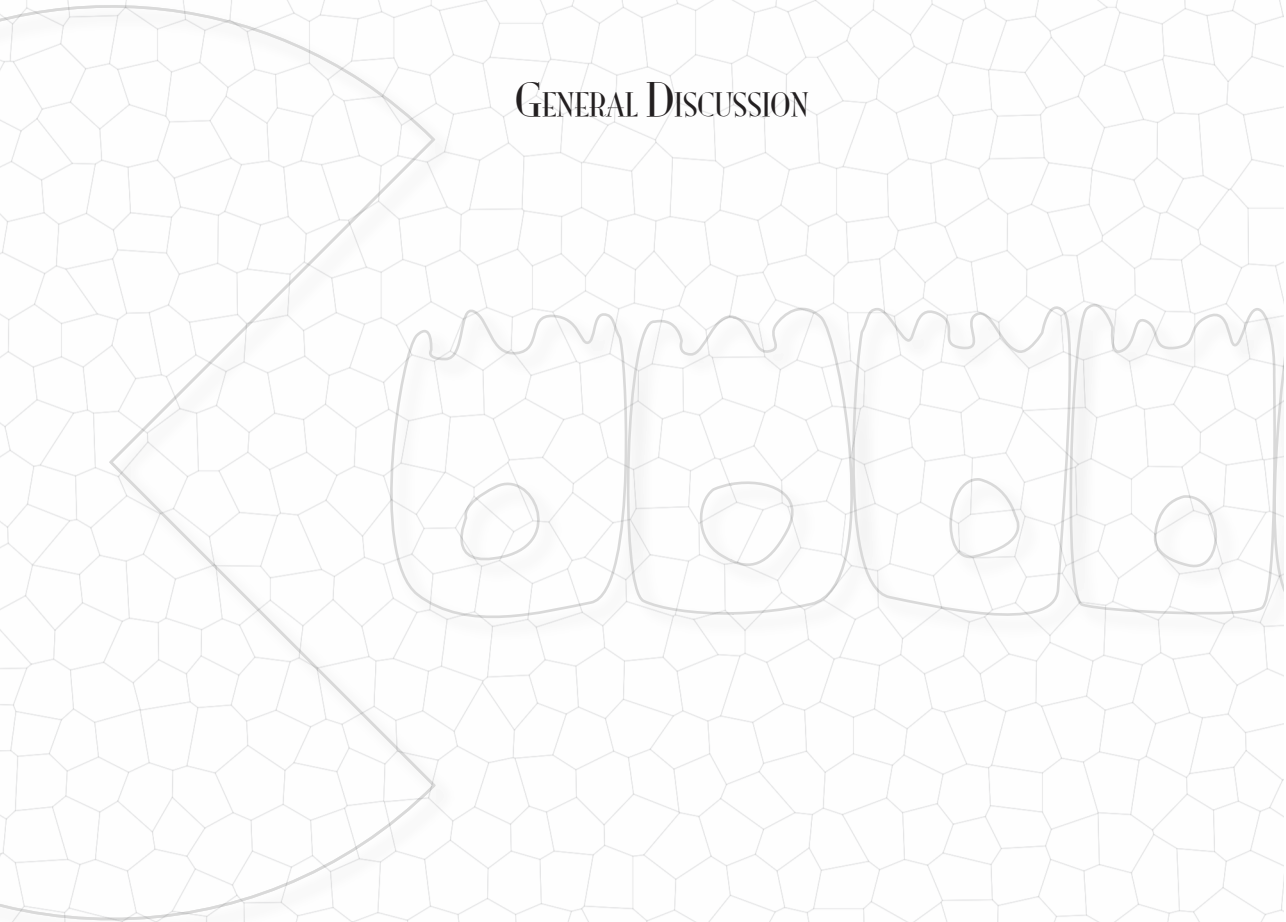
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CHAPTER 6

GENERAL DISCUSSION



GENERAL DISCUSSION

Although celiac disease (CD) is a relatively benign condition where elimination of gluten from the diet can reverse disease, a small percentage of adult-onset CD patients fail to recover on a gluten-free diet. These refractory celiac disease (RCD) patients have persisting villous atrophy and elevated levels of intraepithelial lymphocytes (IELs) in the small intestine. In a subset of these patients an aberrant monoclonal IEL population is present that lacks expression of the T cell receptor (TCR)-CD3 complex on the cell surface, but expresses CD3 intracellularly. This aberrant IEL population is now regarded as an intraepithelial lymphoma that can develop into overt lymphoma¹. Therefore, this disease state -RCD type II (RCD II)- is considered an intermediate stage between CD and lymphoma. This thesis reports on novel insights on aberrant IELs that increased our knowledge on these (pre)malignant cells and that might aid in the development of future therapies for RCD II and RCD-associated lymphoma.

RCD CELL LINES: A MODEL FOR ABERRANT IELS IN RCD II AND RCD-ASSOCIATED LYMPHOMA

About a decade ago, aberrant IELs in RCD II were identified as the missing link between TCR⁺ IELs in CD and the TCR⁻ lymphoma cells of enteropathy associated T cell lymphoma (EATL)^{2,3} Although the aberrant IELs did not express the TCR-CD3 complex on the cell surface, CD3 was expressed intracellularly and clonal TCR- γ gene rearrangements were commonly found. One view is, therefore, that aberrant IELs are mature TCR⁺ IELs that lost cell surface expression of the TCR-CD3 complex due to overstimulation⁴. Studies on aberrant IELs were mainly performed on IELs *in situ*, which limited the type of experiments that could be performed to investigate molecular events that are linked to malignant transformation. The isolation and propagation of aberrant IEL lines isolated from small intestinal biopsies of RCD II patients (Chapter 2) allowed us to perform more in-depth analysis of aberrant IELs. These RCD cell lines had the characteristic aberrant IEL phenotype: surface TCR-CD3⁻CD4⁻CD8⁻CD7⁺CD103⁺, intracellular CD3⁺. In addition, the RCD cell lines displayed monoclonal TCR- γ gene rearrangements identical to the predominant rearrangements found in the aberrant IELs of the patients from which the cell lines were derived. The proliferative response of the RCD cell lines to IL-15, which is highly upregulated in RCD II and RCD-associated lymphoma⁵, further supported the notion that RCD cell lines could serve as a model for aberrant IELs (Chapter 2).

ABERRANT IELS CONTRIBUTE TO THE PROPAGATION OF TISSUE DAMAGE AND INFLAMMATION IN RCD II.

In active CD, the number of TCR⁺ IELs is markedly increased. It has become clear that TCR⁺ IELs can acquire an activating NK cell receptor repertoire, presumably under the influence of IL-15⁶. These NK cell receptors co-stimulate TCR-mediated lysis of epithelial cells⁷. Upon stimulation with IL-15, which can alter the NK cell receptor function, IELs can even display TCR-independent cytotoxicity⁸. It seems, therefore, likely that these TCR⁺ IELs that have acquired an NK cell phenotype play a role in perpetuating tissue damage in RCD.

In addition to a role in tissue damage, TCR⁺ IELs in active CD contribute to the pro-inflammatory milieu by the secretion of high levels of IFN- γ ^{9;10}. Analogous to IFN- γ secretion by gluten-specific CD4⁺ T cells¹¹, IFN- γ secretion by TCR⁺ IELs seems to be linked to gluten-intake as TCR⁺ IELs in active CD patients and CD patients after gluten-challenge secreted higher levels of IFN- γ when compared to healthy controls or CD patients on a gluten-free diet¹⁰. Furthermore, TCR⁺ IELs also secreted high amounts of IFN- γ after stimulation with IL-15^{12;13}. Strikingly, none of the other T helper 1 (Th1) or T helper 2 (Th2) cytokines seemed to have a role as important as IFN- γ (and to a lesser extent IL-10) in the inflammatory response of active CD.

Much less is known about cytotoxicity and cytokine secretion of the aberrant TCR⁺ IEL population in RCD II and the subsequent state of lymphoma. The availability of the RCD cell lines as a model for aberrant IELs allowed us to address these issues *in vitro*. In analogy with TCR⁺ IELs in active CD, the TCR⁺ RCD cell lines expressed multiple activating NK cell receptors (Chapter 3). In contrast to the TCR⁺ IELs, however, epithelial-cell specific cytotoxicity was mainly mediated by DNAM-1¹⁴ with only a minor role for other activating NK cell receptors (Chapter 3). We postulated that in a subset of RCD II patients the aberrant IELs acquire the ability to lyse epithelial cells via DNAM-1.

Analogous to TCR⁺ IELs, the RCD cell lines secreted IFN- γ but none of the other Th1 cytokines upon stimulation with IL-15. This indicated that aberrant IELs can play a role in the propagation of the inflammatory response in RCD II (Chapter 4). Interestingly, simultaneous stimulation with an anti-CD30 antibody and IL-15 had a synergistic effect on IFN- γ secretion (Chapter 4). As CD30 is expressed on aberrant IELs in a subset of RCD II patients¹⁵ and on virtually all RCD-associated lymphoma cells, combined stimulation of the CD30 receptor with IL-15 stimulation could occur *in vivo* and might contribute to the ongoing inflammation in the transition from RCD II into lymphoma.

Together, these results indicate that aberrant IELs could contribute to the ongoing tissue damage and cytokine secretion in RCD II and RCD-associated lymphoma.

ABERRANT IELs POSSIBLY ORIGINATE FROM A UNIQUE CD3⁺CD7⁺ LYMPHOCYTE POPULATION

Knowledge on the functional characteristics of aberrant IELs has thus greatly increased. The exact cellular origin of aberrant IELs, however, remains unclear. Following the hypothesis that aberrant IELs used to be TCR⁺ IELs, we assessed the presence of the TCR-CD3 complex in our RCD cell lines. Intracellularly, all CD3 α -chains and ζ -chains were present and could form the CD3 $\alpha\zeta$ -, CD3 $\delta\zeta$ - and $\zeta\zeta$ -dimers (Chapter 2). The TCR-chains, however, were not always present (Chapter 2). Introduction of exogenous TCR-chains, however, restored surface expression and functionality (proliferation and IFN- γ production) in the RCD cell lines (Chapter 2 and unpublished). This indicates that the CD3-complex was functional in the RCD cell lines. Analysis of TCR-gene rearrangements with PCR and Southern blot revealed that clonal TCR-gene rearrangements could be found in the RCD cell lines. These TCR-gene rearrangements were, however, either incomplete, non-functional or out-of frame and can therefore not assemble into a functional TCR. Furthermore, one of the RCD cell lines only displayed germline configuration of the TCR genes (Chapter 5).

The RCD cell lines thus have characteristics of T cells, yet their phenotype does not match that of mature T cells. Phenotypic analysis of the RCD cell lines indicated that aberrant IELs have a unique phenotype: CD45⁺CD19⁻CD14⁻CD3⁻CD7⁺CD56⁻CD34⁻CD103⁺CD127⁻ (Chapter 5). This phenotype is distinct from any known lymphocyte class. Furthermore, IELs with this phenotype were present in the small intestine of healthy adults, children and thymic cell suspensions (chapter 5). We hypothesized, therefore, that aberrant IELs do not derive from mature T cells but instead derive from cells in an early stage of extrathymic lymphocyte development.

DIRECTIONS FOR FURTHER RESEARCH

The different aspects of aberrant IELs described in this thesis help understand the events leading from uncomplicated CD to RCD II and gastrointestinal lymphoma. Many issues, however, remain to be answered.

First, most patients do well on a gluten-free diet but some individuals develop RCD II and lymphoma. It has been suggested that HLA-DQ2 gene dose is involved in the risk to develop complicated CD as RCD II patients and gastrointestinal lymphoma patients were more often HLA-DQ2.5 homozygous¹⁶. Genome-wide association studies (GWAS) on uncomplicated CD patients have identified several risk variants on non-HLA-loci for development of CD¹⁷. The presence of a higher number of additional non-HLA risk alleles is directly correlated with an increase in the risk to develop CD¹⁸. An interesting issue is whether these genes also influence the risk to develop complicated forms of CD. A GWAS on RCD I, RCDII and RCD-associated lymphoma patients with uncomplicated CD patients and healthy individuals as controls would be a way to uncover the role of non-HLA-genes on the development of complicated CD. Screening of CD patients on potential risk alleles might then lead to earlier intervention. Recent GWAS have indicated that to identify risk variants with small effect sizes the amount of samples should be increased to obtain significant results. As only 2-5% of adult-onset CD patients develop complicated CD, even pooling of sample collections from multiple European countries would not obtain a sufficient sample-size for such a study.

Second, cells with a TCR-CD3-negative phenotype are present in the intestine of healthy adults and children (Chapter 5). These cells could be an immature lymphocyte type that represents the physiological counterpart of the aberrant IELs in RCD II. To test this hypothesis, future studies are needed to determine whether these cells can differentiate into mature T cells and/or NK cells. Hematopoietic precursor cells from fetal liver and postnatal thymus were shown to differentiate into T cells on OP9 cells expressing Notch ligand Delta-like-1(DL1)^{19,20}. In analogy, cells with the CD45⁺CD19⁻CD14⁻CD3⁻CD7⁺CD56⁻CD34⁻CD103⁺CD127⁻ phenotype could be isolated from the small intestine of a healthy individual and cultured in the presence of stromal cell lines OP9 or OP9-delta-like-1¹⁹. Such co-culturing could indicate whether the cell population identified in Chapter 5 can indeed differentiate into a mature lymphocyte type. Alternatively, the presence of essential transcription factors Notch1 and GATA3 could indicate in which stage of development these lymphocytes are²¹. Microarray analysis might inform whether these genes are differentially expressed when compared to mature T and NK cells. In addition, in mice the transcription factor E4BP4²² has been appointed as a critical factor in NK cell development. It remains to be determined if this factor is also important in human NK cell development.

Third, a subset of RCD II patients acquires the ability to lyse epithelial cells via DNAM-1 (Chapter 3). It remains unclear, why DNAM-1 that normally acts as a co-receptor, can now act autonomously. Recently, the inhibitory receptor TIGIT was identified that can be co-expressed with DNAM-1 on T cells and NK cells and competes for binding to the ligands CD155 and CD112^{23;24}. TIGIT was shown to have a 100-fold higher affinity for CD155 than DNAM-1 suggesting that, when co-expressed, TIGIT has the dominant function and can regulate the function of DNAM-1²³. Dysregulation of the balance of expression of TIGIT and/or DNAM-1 might enable DNAM-1 to induce cytotoxicity autonomously. Determining expression of TIGIT and DNAM-1 in different stages of CD and in healthy controls could be a way to address this issue.

FUTURE THERAPEUTIC POSSIBILITIES FOR RCD II AND RCD-ASSOCIATED LYMPHOMA

RCD II and RCD-associated lymphoma are severe complications of CD with 5-year survival rates of 44-58% and <20% respectively²⁵. RCD II is very resistant to therapy and transition into lymphoma cannot be prevented. At present, only Cladribine therapy²⁶ or autologous stem cell therapy²⁷ might have an effect on reducing the amount of (pre) malignant aberrant IELs.

Blocking of IL-15 has been suggested as a therapy as this cytokine influences many aspects of aberrant IELs including apoptosis¹, proliferation (Chapter 2), cytotoxicity (Chapter 3) and cytokine secretion (Chapter 4). Alternatively, we suggest that blocking of DNAM-1 and CD30 may be interesting as these molecules seem to be linked to malignant transformation of the aberrant IELs into lymphoma cells. Monoclonal antibody therapy against these two receptors might have immediate effects on cytotoxicity and cytokine production, thus limiting tissue destruction and inflammation.

In conclusion, the research described in this thesis improved our understanding of the pathogenesis of complicated CD and gave us suggestions on possibilities for future therapy.

REFERENCES

1. Malamut G, El MR, Montcuquet N et al. IL-15 triggers an antiapoptotic pathway in human intraepithelial lymphocytes that is a potential new target in celiac disease-associated inflammation and lymphomagenesis. *J.Clin.Invest* 2010;120:2131-2143.
2. Cellier C, Delabesse E, Helmer C et al. Refractory sprue, coeliac disease, and enteropathy-associated T-cell lymphoma. French Coeliac Disease Study Group. *Lancet* 2000;356:203-208.
3. Bagdi E, Diss TC, Munson P, Isaacson PG. Mucosal intra-epithelial lymphocytes in enteropathy-associated T-cell lymphoma, ulcerative jejunitis, and refractory celiac disease constitute a neoplastic population. *Blood* 1999;94:260-264.
4. Cellier C, Patey N, Mauvieux L et al. Abnormal intestinal intraepithelial lymphocytes in refractory sprue. *Gastroenterology* 1998;114:471-481.
5. Mention JJ, Ben AM, Begue B et al. Interleukin 15: a key to disrupted intraepithelial lymphocyte homeostasis and lymphomagenesis in celiac disease. *Gastroenterology* 2003;125:730-745.
6. Meresse B, Curran SA, Ciszewski C et al. Reprogramming of CTLs into natural killer-like cells in celiac disease. *J.Exp.Med.* 2006;203:1343-1355.
7. Hue S, Mention JJ, Monteiro RC et al. A direct role for NKG2D/MICA interaction in villous atrophy during celiac disease. *Immunity*. 2004;21:367-377.
8. Meresse B, Chen Z, Ciszewski C et al. Coordinated induction by IL15 of a TCR-independent NKG2D signaling pathway converts CTL into lymphokine-activated killer cells in celiac disease. *Immunity*. 2004;21:357-366.
9. Olausson RW, Johansen FE, Lundin KE et al. Interferon-gamma-secreting T cells localize to the epithelium in coeliac disease. *Scand.J.Immunol.* 2002;56:652-664.
10. Forsberg G, Hernell O, Melgar S et al. Paradoxical coexpression of proinflammatory and down-regulatory cytokines in intestinal T cells in childhood celiac disease. *Gastroenterology* 2002;123:667-678.
11. Nilsen EM, Lundin KE, Krajci P et al. Gluten specific, HLA-DQ restricted T cells from coeliac mucosa produce cytokines with Th1 or Th0 profile dominated by interferon gamma. *Gut* 1995;37:766-776.
12. Di Sabatino A, Ciccocioppo R, Cupelli F et al. Epithelium derived interleukin 15 regulates intraepithelial lymphocyte Th1 cytokine production, cytotoxicity, and survival in coeliac disease. *Gut* 2006;55:469-477.
13. Ebert EC. IL-15 converts human intestinal intraepithelial lymphocytes to CD94 producers of IFN-gamma and IL-10, the latter promoting Fas ligand-mediated cytotoxicity. *Immunology* 2005;115:118-126.
14. Shibuya A, Campbell D, Hannum C et al. DNAM-1, a novel adhesion molecule involved in the cytolytic function of T lymphocytes. *Immunity*. 1996;4:573-581.
15. Farstad IN, Johansen FE, Vlatkovic L et al. Heterogeneity of intraepithelial lymphocytes in refractory sprue: potential implications of CD30 expression. *Gut* 2002;51:372-378.
16. Al-Toma A, Goerres MS, Meijer JW et al. Human leukocyte antigen-DQ2 homozygosity and the development of refractory celiac disease and enteropathy-associated T-cell lymphoma. *Clin.Gastroenterol.Hepatol.* 2006;4:315-319.
17. Dubois PC, Trynka G, Franke L et al. Multiple common variants for celiac disease

- influencing immune gene expression. *Nat.Genet.* 2010
18. Romanos J, van Diemen CC, Nolte IM et al. Analysis of HLA and non-HLA alleles can identify individuals at high risk for celiac disease. *Gastroenterology* 2009;137:834-40, 840.
 19. Schmitt TM, Zuniga-Pflucker JC. Induction of T cell development from hematopoietic progenitor cells by delta-like-1 in vitro. *Immunity.* 2002;17:749-756.
 20. van Coppennolle S, Verstichel G, Timmermans F et al. Functionally mature CD4 and CD8 TCRalpha cells are generated in OP9-DL1 cultures from human CD34+ hematopoietic cells. *J.Immunol.* 2009;183:4859-4870.
 21. Blom B, Spits H. Development of human lymphoid cells. *Annu.Rev.Immunol.* 2006;24:287-320.
 22. Gascoyne DM, Long E, Veiga-Fernandes H et al. The basic leucine zipper transcription factor E4BP4 is essential for natural killer cell development. *Nat.Immunol.* 2009;10:1118-1124.
 23. Yu X, Harden K, Gonzalez LC et al. The surface protein TIGIT suppresses T cell activation by promoting the generation of mature immunoregulatory dendritic cells. *Nat.Immunol.* 2009;10:48-57.
 24. Stanietsky N, Simic H, Arapovic J et al. The interaction of TIGIT with PVR and PVRL2 inhibits human NK cell cytotoxicity. *Proc.Natl.Acad.Sci.U.S.A* 2009;106:17858-17863.
 25. Malamut G, Afchain P, Verkarre V et al. Presentation and long-term follow-up of refractory celiac disease: comparison of type I with type II. *Gastroenterology* 2009;136:81-90.
 26. Al-Toma A, Goerres MS, Meijer JW et al. Cladribine therapy in refractory celiac disease with aberrant T cells. *Clin.Gastroenterol.Hepatol.* 2006;4:1322-1327.
 27. Al-Toma A, Visser OJ, van Roessel HM et al. Autologous hematopoietic stem cell transplantation in refractory celiac disease with aberrant T cells. *Blood* 2007;109:2243-2249.