

Cover Page



Universiteit Leiden



The handle <http://hdl.handle.net/1887/20942> holds various files of this Leiden University dissertation.

Author: Wammes, Linda Judith

Title: Immune regulation during parasitic infections : from bench to field

Issue Date: 2013-06-11



CHAPTER 8

Global worming or deworming? Epidemiological, immunological and clinical perspectives

Linda J. Wammes, Harriet Mpairwe, Alison M. Elliott, Maria Yazdanbakhsh

– Manuscript in preparation –

Summary

Helminth infections still affect billions of people worldwide. Deworming is advocated to prevent the morbidities induced by helminths. At the same time, in resource-rich settings, the possibility of infecting patients with different life cycle stages of worms is being explored, specifically for those suffering from a range of inflammatory diseases. It has been recognized that helminths have immunomodulatory properties, which can impair not only parasite-specific immune responses but also responses to third party antigens. By using these properties, parasitic helminths allow their long-term survival within their human host but at the same time curtail overt immunological responses that lead to allergic, inflammatory bowel or autoimmune diseases. However, this would also suggest that deworming could lead to the development of inflammatory conditions, due to the removal of regulatory mechanisms associated with helminths. This can result in unfavorable situations in countries that are not prepared for the emergence of such pathological conditions. This review summarizes the consequences of deworming versus helminthic therapy.

Background

Parasitic worms have accompanied us throughout human history¹. The fact that mortality due to helminth infections is rare, and that worm infections are often asymptomatic, with only a minority of infected individuals having intense infections², suggests that this is as a result of evolutionary co-adaptation between parasitic worms and man. A key component of this partnership is the immunological interaction between helminths and their mammalian hosts. Helminths modulate immunological processes: multiple mechanisms of helminth-induced immunological tolerance and regulatory pathways have been revealed over the past 50 years, that may explain chronicity of these infections³. Helminths also appear to fundamentally affect the host's genetic composition; studies into the recent evolutionary history of human interleukin genes reveal that pathogens have driven selection of certain genetic adaptations and that helminths have had a stronger influence than other classes of pathogens in this process⁴. Pressure from helminth infections seems to have promoted the selection of alleles likely to protect against infection, while predisposing humans to immune-mediated diseases such as allergies.

In the twentieth century, great effort was put into the worldwide control of infectious diseases by improved hygiene, vaccination programs and drug treatment. However, the decline in parasitic and other infectious diseases was associated with a marked increase in prevalence of chronic inflammatory disorders such as asthma, auto-immune disease (type 1 diabetes, multiple sclerosis) and inflammatory bowel disease⁵. Although the prevalence of asthma and allergic disorders now seems to have stabilized in developed countries^{6,7}, the prevalence is on the increase in developing countries^{7,8}. These epidemiological observations accord with the Hygiene Hypothesis⁹, which suggests that removal of the regulatory effects of pathogens such as helminths (from populations genetically adapted to live with them) tends to lead to an imbalance in the immune system followed by a number of pathological conditions.

Consequently, the question arises whether helminths should be considered as harmful pathogens or as beneficial commensals. While in low-resource settings deworming is advocated to prevent worm-associated morbidity, several research groups in the Western world are currently investigating the therapeutic potential of worms and their secreted products in inflammatory diseases. Whereas deworming may result in an increase of inflammatory disorders, introduction of helminths experimentally can potentially be harmful. Experimental therapeutic infections using worms and their products may lead to unforeseen immunological and pathological consequences. This review aims to summarize current knowledge on the immunological effects of worms, the beneficial aspects of chronic helminth infections, the possible consequences of global deworming, and the current

evidence as to whether the controlled use of worms or their products for treating patients is beneficial.

T cell responses in infectious and inflammatory diseases

The immune system is equipped with different cell types involved in recognition of pathogens and their elimination. So far, a number of T-cell subsets have been identified that appear to be key to the control of distinct classes of incoming pathogens. T-helper (Th1) cells are mainly involved in defence against intracellular pathogens, Th2 are there to combat helminths and ectoparasites and Th17 cells appear to be important for defence against extracellular bacteria and fungi¹⁰. However, it is also clear that these cells can inflict damage to tissues and organs if uncontrolled. Thus whereas Th1 and Th17 cells that release pro-inflammatory cytokines are involved in recruitment and activation of macrophages and neutrophils that can attack bacteria, viruses, protozoa and fungi, their overt activation is associated with autoimmune and inflammatory diseases. Th2 cells trigger responses that disable, degrade and dislodge parasites, as recently reviewed¹¹. Interestingly, Th2 cytokines seem to be involved in tissue repair as well^{12,13}, which may encapsulate the parasite and prevent excessive inflammation¹⁴. However, an overactivated Th2 immune response can lead to allergies and allergic diseases. Therefore it is not surprising that an important component of the immune system is the regulatory network, spearheaded by the regulatory T cells, that are capable of controlling activated effector T cells through expression of inhibitory molecules¹⁵. It should be noted that additional components of the regulatory network are the so-called modified Th1 and Th2 cells which express the signature cytokines, IFN- γ and IL-4/IL-5/IL-13, respectively, along with IL-10¹⁶. It is thought that over time, effector Th1 and Th2 cells start to express IL-10 to control the damage that might otherwise be inflicted by these cells.

Characteristics of immune responses in helminth infections

Helminth-induced immune regulatory network

In the late half of the 20th century, Ottesen and others started to elucidate the immunological basis of helminth-host interaction¹⁷⁻¹⁹. Interestingly, they observed that in helminth-infected individuals, the proliferative response of lymphocytes to specific antigens was lower than in uninfected subjects¹⁷⁻²⁰. These and other studies²¹ have led to the concept that cellular immune "hyporesponsiveness" induced by helminths is one of the key mechanisms used by these parasites to evade the host immune system. Numerous studies have since contributed to our understanding of a sophisticated immune regulatory network operative during helminth infections¹⁶. First, the presence of non-lymphocytic adherent cells was

demonstrated in the blood of patients with *Brugia malayi* microfilariae²². The adherent cells could suppress anti-filarial immune responses when cultured with patient-derived lymphocytes²². These cells are probably the precursors for regulatory antigen-presenting cells that are now studied, such as alternatively activated macrophages and monocytes or regulatory dendritic cells²³⁻²⁶. Furthermore, the existence of suppressor CD8⁺ T cells was shown in individuals with patent *B. malayi* or *B. timori* microfilaremia in Indonesia; antibody-mediated depletion of these cells enhanced proliferative responses to parasite antigens²⁷. Thereafter, the focus was shifted to a population of CD4⁺ T cells, termed regulatory T cells (Tregs)²⁸. Tregs are an essential component of the immune system, helping the immune response to reach an optimal balance between sufficiently strong anti-pathogen responses and carefully gauged control of overt and pathogenic inflammation¹⁵. Helminths are thought to exploit this immune suppressory mechanism to ensure their long-term survival. Natural thymus-derived Tregs are currently characterized as CD4⁺CD25^{hi}FOXP3⁺ cells, but several peripherally inducible subsets of regulatory T cells with variable phenotypes have also been described^{29,30}. Several cross-sectional studies in human helminthiases have indeed provided evidence for the expansion of Tregs during infection³¹⁻³⁶. Not only numbers, but also functional properties of Tregs have been studied. In individuals with patent *Wuchereria bancrofti* microfilaremia, suppressed filarial-specific proliferative T and B cell responses as well as IL-13 production *in vitro* were restored after removal of CD4⁺CD25^{hi} Tregs³⁷.

The mechanisms for Treg expansion by these parasites are not fully understood, but lately a number of helminth-derived molecules have been identified that can directly drive Treg induction. Molecules derived from *Schistosoma mansoni* can condition dendritic cells to induce IL-10 producing Tregs³⁸. The excretory/secretory products of *Heligmosomoides polygyrus* as well as omega-1, a glycoprotein of *S. mansoni* eggs, can directly induce Foxp3 expression in murine CD4⁺ T cells^{39,40}. These studies indicate that there might be great potential in utilising helminth-derived molecules in the manipulation of the human immune system such that regulatory T cells are induced or enhanced.

Besides T cells, several other cells appear to be involved in immune regulation, as illustrated by emergence of regulatory B cells, a subset producing IL-10 during human helminthiases^{41,42}, and the importance of regulatory subsets of antigen-presenting cells as mentioned above.

Induction of type 2 responses by helminths

One of the hallmarks of helminth infections is the expansion of the Th2 responses; elevated cytokines such as IL-4, IL-5 and IL-13, along with high levels of IgE characterize helminth-infected subjects⁴³. As there is evidence for a degree of counter-regulation between Th1 and Th2 responses, the strong type 2 inducing

capacity of helminths might also play a role in keeping Th1- (and Th17-) mediated diseases such as inflammatory bowel diseases (IBD) and arthritis at bay. Therefore, understanding the mechanisms whereby helminths lead to Th2 responses might, in analogy with understanding how Tregs are induced, lead to potential therapeutic interventions. Recent studies have identified helminth antigens with the ability to drive Th2 responses in murine models. Th2-inducing properties of molecules derived from *Fasciola hepatica*⁴⁴, *S. mansoni*⁴⁴ and *H. polygyrus*⁴⁵ was demonstrated by injection into mice. ES-62, a glycoprotein secreted by the filarial nematode *Acanthocheilonema viteae*, and omega-1, a molecule found in excretory secretory (ES) products of *S. mansoni* eggs, have been shown to endow DC with the ability to skew immune responses toward type 2 in a mouse model^{46,47}. The Th2-inducing capacities of omega-1 may seem in contrast to the previously mentioned Foxp3-inducing activity, however in the latter NOD mice were used⁴⁰. Some of these Th2-inducing capacities have been confirmed in experiments using human cells⁴⁶.

More recently, research into tissues at the sites of worm infection has revealed that epithelial cells, the first line of defence in the gut and lung, produce a set of cytokines, IL-25, IL-33 and thymic stromal lymphopoietin (TSLP), so-called 'alarmins'⁴⁸⁻⁵⁰, which in turn induce innate lymphocytes to produce type 2 cytokines^{51,52}. These cells, termed natural helper cells (NHC), are found in peripheral lymphoid clusters and produce IL-5 and IL-13 enhancing B and T cell function⁵³; as yet very little is known about the equivalent of these cells in humans.

The association between helminth infections and inflammatory diseases

To date, human studies of the possible influence of worm infections on inflammatory diseases have included cross-sectional studies, anthelmintic trials and trials of helminth therapy. We shall now look at each in turn, discussing the potential pitfalls and advantages of each approach. We will start with summarizing the associations between helminth infections and different inflammatory diseases.

Allergy & asthma

Based on the ability of helminth infections to induce a regulatory network and to modify the Th2 response, we have postulated that helminths have a special capacity to protect against allergic conditions. There is evidence from several murine models that helminth infections might be protective against allergic airway inflammation⁵⁴⁻⁵⁷. However, the findings from cross-sectional studies in humans regarding the association between worm infections and allergy-related conditions have so far been inconsistent (reviewed by Leonardi-Bee et al⁵⁸ and Flohr et al⁵⁹).

For atopy, usually assessed as a positive skin prick test (SPT) response to a panel of allergens, an inverse association with worms has commonly been observed⁶⁰⁻⁶⁵, with occasional exceptions⁶⁶. For allergy-related clinical syndromes, results have been more mixed. Both negative^{67, 68} and positive⁶⁹ associations have been reported for eczema. Negative associations have been observed for wheezing in some studies^{70,71}; in others, an absence of association between worms and wheeze⁷¹ or asthma⁷², or a positive association between worms and asthma⁶⁶ have been reported. Factors that may influence the relationships between worms and allergy observed in these studies are likely to be complex but may include the timing, or chronicity of helminth infection – with recent infections having a different effect from chronic, established infections⁶⁴ – as well as the intensity of the infections, the specific parasite species and the host genetics^{73, 74}.

Inflammatory Bowel Diseases

Studies in murine models have shown that infection with worms protects against several forms of experimentally induced colitis⁷⁵⁻⁷⁸ but studies in humans are hindered by the fact that inflammatory bowel disease (IBD) and parasitic infections do usually not coincide. A small study in India showed that patients with Crohn's disease had lower immune responses to different hookworm antigens compared to healthy controls, suggesting that patients had less often harbored hookworm infections, however the infection was not assessed directly⁷⁹.

Multiple sclerosis

A case-control study in 1966 already pointed to the contribution of environmental factors in MS; the presence of piped water, a flush toilet and sharing a room with 1 person or less were more often recorded as environmental factors associated with MS patients compared to healthy controls⁸⁰. Reasoning from the Hygiene Hypothesis, the country prevalences of MS and *T. trichiura* infections were compared and shown to be almost mutually exclusive⁸¹. Correale and colleagues took this further by assessing a prospective cohort of MS patients. When comparing helminth-infected and -uninfected patients, the appearance of new MRI lesions was much less frequent in the infected individuals over 5 years of follow-up. This difference seemed to be associated with higher production of IL-10 and TGF- β by peripheral blood mononuclear cells (PBMC) and with suppressive activity of Tregs⁸². Furthermore, when a few of the helminth-infected patients were treated with anthelmintics due to intestinal symptoms, the number of clinical relapses and MRI lesions increased, parallel to a decrease in regulatory immune responses⁸³. Using the murine model for MS, experimental autoimmune encephalomyelitis (EAE), evidence was further obtained for a protective effect of helminths or their products⁸⁴⁻⁸⁸ on the clinical course and CNS inflammation.

Diabetes mellitus

Another autoimmune condition that has been linked to immune modulatory properties of helminths is diabetes mellitus. In the non-obese diabetes (NOD) mouse model, it was shown that several types of helminthic infections could prevent type 1 diabetes (T1D)⁸⁹⁻⁹². In humans, an inverse association has been observed between prevalence of T1D and neglected tropical diseases, which helminth infections form a substantial part of. They showed an inverse relation between the prevalence of T1D and the access to sanitation and clean water, although this was not statistically tested⁹³. Within the Chennai Urban Rural Epidemiology Study (CURES) in India, the prevalence of lymphatic filariasis was lower in T1D subjects⁹⁴. They furthermore measured total anti-filarial IgG and IgG4 as proxies for past and current infections, respectively. Whereas total IgG levels were not different between the groups, IgG4 was higher in the non-diabetic group, indicating that current infections might lead to this inverse relationship.

For type 2 diabetes (T2D), it is more difficult to establish experimental models. This multifactorial disease is currently regarded as an inflammatory disease⁹⁵, but is also associated with genetic as well as nutritional and other life style factors. Another report from the CURES study has shown lower prevalence of LF in T2D patients⁹⁶. Moreover, the study showed lower serum levels of pro-inflammatory cytokines in LF-positive versus LF-negative subjects with T2D, suggesting a role for anti-inflammatory properties of LF infection in protection against T2D.

Rheumatoid arthritis

Although some helminth infections can induce reactive arthritis which resembles rheumatoid arthritis⁹⁷ several studies in different rodent models have shown that helminth infections or extracts can suppress or prevent arthritis⁹⁸⁻¹⁰⁰. The filarial-derived glycoprotein ES-62 was shown to be effective as a therapy for murine collagen-induced arthritis and furthermore synovial cells from rheumatoid arthritis (RA) patients treated with ES-62 produced less pro-inflammatory cytokines after LPS stimulation¹⁰¹.

Other autoimmune or inflammatory diseases

Aoyama et al. showed that *Strongyloides stercoralis* infections were less prevalent in patients with autoimmune liver diseases than in other individuals visiting the hospital¹⁰², however there has been no further follow-up study to this. In a mouse model of Graves' disease, an autoimmune disease of the thyroid, *S. mansoni* appeared to have a protective effect¹⁰³, but this has not been examined in humans. Furthermore, it has been suggested that helminths might have therapeutic potential in atherosclerosis and other cardiovascular diseases¹⁰⁴, based on a study that showed *S. mansoni* to have anti-atherogenic effects in a mouse model. The

exact mechanism has not been resolved but it might be caused by the consumption of lipids by *S. mansoni*, rather than by immune-mediated mechanisms¹⁰⁵.

Studies of anthelmintic treatment

Cross-sectional and observational studies are inherently flawed due to difficulty in ascertaining cause and effect, issues of confounding and reverse causation. Therefore, evidence from such studies need to be complemented by other study designs. Since remote areas still suffer from a high burden of helminthic infections¹⁰⁶, mass deworming programs are currently advocated¹⁰⁷. This creates opportunities to investigate prospectively whether deworming paves the way for the increased prevalence of allergic and other inflammatory diseases.

Effect of deworming on atopic diseases

Anthelmintic trials are designed to study the influence of worms on allergic conditions indirectly, by studying the effects of treating the worms. Such trials have been conducted among school-going children from worm-endemic areas using different anthelmintic drugs for various periods of follow-up. An initial non-randomized study in Venezuela compared treated children with children who declined treatment, and suggested that anthelmintic treatment was associated with increased prevalence of atopy¹⁰⁸. The first randomized trial was carried out among 317 Gabonese school children using open-label three-monthly praziquantel and mebendazole, versus no treatment, over 30 months; it found that anthelmintic treatment was associated with an increased rate of developing positive skin responses to house dust mites¹⁰⁹. A cluster-randomized trial among 1632 children from Ecuador using two-monthly albendazole versus placebo over 12 months found no effect on SPT responses¹¹⁰. An individually randomized trial, among 1566 rural children from Vietnam using initially three-monthly mebendazole versus placebo but later three-monthly albendazole versus placebo over 12 months, found an increased risk of positive SPT responses to allergens¹¹¹. The latest trial in Indonesia, assessed the effect of three-monthly albendazole treatment compared to placebo on SPT reactivity to different allergens. A significant increase in cockroach reactivity was observed after 21 months, but overall SPT responses were not altered¹¹². None of these four trials showed any effects of anthelmintic treatment on clinical allergy outcomes. Given the fact that clinical allergy is relatively rare in these areas, it should be considered that the power of these studies might have been not sufficient to detect significant effects. Moreover, it has to be noted that differences in species of prevalent helminths, co-prevalence of other immunomodulating infections (such as oro-faecal infections^{113,114} or malaria¹¹⁵), exposure to environmental pollutants and duration and timing of treatment, can have major impact on trial outcomes.

Pertinent to the duration of treatment, a study in Ecuador compared SPT reactivity and allergy-related symptoms among school-age children from communities that had received 15-17 years of periodic ivermectin treatment to school-age children from adjacent communities that had not received treatment¹¹⁶. This study observed that the prevalence of skin reactivity to allergens among children from the treated communities was double that for the children from untreated communities. The children from the treated communities also had a higher prevalence of recent eczema symptoms, but not any other allergy-related symptoms¹¹⁶. While the design of this study is again limited by the fact that the communities were not randomized relative to the intervention, the results do suggest that long-term intervention against helminths may be required to alter responsiveness to allergens and to influence allergy-related clinical outcomes.

Regarding the timing of treatment, a trial among 2507 pregnant women in Uganda using single doses of albendazole and praziquantel versus matching placebos (in a 2x2 factorial design) found that albendazole during pregnancy was associated with an increased risk of eczema in infancy¹¹⁷ and in the first five years of life, whether the mother had hookworm infection or not¹¹⁸. Praziquantel during pregnancy was associated with an increased risk of eczema in infancy among children whose mothers were infected with *S. mansoni* infection¹¹⁷. In this same trial, children themselves were randomized to three-monthly albendazole versus placebo from fifteen months to five years of age; this treatment was not associated with an increased risk of eczema in early childhood¹¹⁸. These results suggest that in-utero events may be more important in priming or programming the child's immune system (thereby influencing the risk of eczema, and perhaps other allergy-related conditions) than events in early childhood. Surprisingly, anthelmintic treatment with albendazole and praziquantel during pregnancy had no beneficial effects on the immune responses to BCG, tetanus and measles vaccines in early childhood and none of the anticipated benefits for birth weight, resistance to infectious diseases, or improved child development^{119,120}, that would have compensated for the observed adverse effect on eczema.

All the above studies examined effects of anthelmintics in general populations. Another approach has been to examine effects of anthelmintic treatment in people already suffering from allergy-related disease. For example, a recent small trial conducted among individuals aged five to fifty years with a history of asthma in the last twelve months from a schistosomiasis-endemic area in Brazil has been published¹²¹. Study participants received single dose albendazole and praziquantel or placebos. No differences in asthma severity between the two treatment arms were observed over three months. When all participants were treated with both drugs after about three months of follow-up, the study observed the worsening of clinical asthma symptoms at fifteen months¹²¹. However, this worsening cannot be confidently attributed to anthelmintic treatment since there was no comparison

arm in the second part of the study. Larger studies investigating the effect of treating worms in people with established allergy-related diseases would be warranted.

Effect of deworming on other inflammatory diseases

The effect of deworming on other chronic inflammatory diseases has not extensively been studied, partly because naturally (and inherent to our hypothesis) the presence of helminth infections and these conditions may not overlap. An interesting study by Bager and colleagues did assess the effect of mebendazole treatment retrospectively in a population cohort in Denmark¹²². 14% of the more than 900.000 children had been prescribed mebendazole, for probable infection with *Enterobius vermicularis*, the pinworm that is still endemic in the US and in Europe¹²³. The incidence rate ratios for the chronic inflammatory diseases asthma, T1D, juvenile arthritis and IBD were not significantly higher in treated children. However, mebendazole was prescribed based on presumption and not diagnosis of pinworm infection. Moreover, enterobiasis in these children might not have been sufficiently chronic to induce immune regulation¹²² and thus the treatment might have masked the possible benefits.

The main drawback of anthelmintic trials in studying the influence of worms on inflammatory diseases is that they are based on a number of assumptions whose validity is currently unknown. For instance, the trials assume that the effect of worms is immediately removed following treatment and that development of allergy symptoms follows soon after anthelmintic treatment. What if the protective effects of worms persist long after anthelmintic treatment? What if the development of clinical allergy and other conditions is not immediate or could only be observed in the next generation? This concern might be answered by longer follow-up and different study designs, such as the Ugandan trial¹¹⁷, which showed that the effect of treating worms in the mother could be observed in the next generation as increased risk of childhood eczema. Follow-up for this Ugandan study is currently ongoing to determine whether the risk of asthma is also increased as the children grow older. The second drawback in using anthelmintic drugs to study the possible influence of worms on allergy is that any observed effect might be due to the anthelmintic drug itself, or to broader spectrum effects, and not due to the elimination of worms. This seems to be the most likely explanation for the observed effect of albendazole treatment in pregnancy on the increased risk of eczema in the Ugandan trial described above. Albendazole acts by binding to tubulin thereby interfering with the formation of microtubules in the cytoskeleton¹²⁴ and hence can affect protozoa^{125, 126}, fungi¹²⁷ and mammalian cells. Whether maternal albendazole increased the risk of childhood eczema through a direct effect or through acting on other organisms, was not established in the

Ugandan study. Therefore, results from anthelmintic trials should be interpreted with caution and it might be helpful to examine effects of a variety of drugs in different trials.

Immunological consequences of deworming

A number of studies have been able to assess immune responses after anthelmintic treatment. Helminth-specific responses have been examined in Vietnamese school children in the context of the larger study on allergy¹¹¹. Cytokine responses were only evaluated in hookworm-positive children, in which albendazole treatment led to lower IL-10 responses to hookworm antigens. A similar effect was seen in *Ascaris* responses in children treated with albendazole in Ecuador, although this study was not placebo-controlled¹²⁸. A decrease in IL-10 might be expected after clearance of immunoregulatory helminth infections, however in earlier studies, treatment of schistosomiasis and STH infections with praziquantel and mebendazole, respectively, had been associated with enhanced IL-10 responses^{129,130}. Also a placebo-controlled trial of praziquantel treatment of pregnant women resulted in higher *S. mansoni*-specific IL-10 production¹³¹. This latter study also found a treatment-induced increase of Th1 and Th2 responses to schistosomal antigens, similar to the enhanced Th1 and Th2 cytokines in response to *Ascaris* after albendazole treatment of school children in Ecuador¹²⁸.

Overall, immune responses tend to increase after anthelmintic treatment, which is in line with the results of our randomized controlled trial (RCT) in Indonesia, showing enhanced pro-inflammatory cytokine responses to *Ascaris* and malaria antigens as well as mitogen after albendazole treatment (Wammes et al. manuscript in preparation). However, it is not known how long this immune stimulatory effect may persist and whether this predisposes to hyperinflammatory conditions.

Very few studies have looked at allergen-specific immune responses after anthelmintic treatment. The trial in Ecuador found no differences in cytokine production in response to cockroach and house mite *Dermatophagoides pteronyssinus* antigens after repeated treatment with albendazole¹²⁸. More trials might be needed to confirm this observation.

Helminth therapy in humans

A more direct approach, which avoids the pitfalls described above, is to study the direct effects of helminths through clinical trials using whole helminths or helminth-derived molecules.

The first scientists to undertake clinical evaluation of infecting patients with helminths were Joel Weinstock and David Elliott in the 1990s, who have recently

summarized the progress in translational studies on helminthic therapy in a review¹³². Up to now, *Trichuris suis* and *Necator americanus* worms have been used in a clinical trial setting, as discussed below.

Trichuris eggs

Most clinical trials so far were performed with *Trichuris suis* ova (TSO) in patients with IBD. *T. suis* is a pig whipworm, which is able to colonize the human gut for a short period of time, but without overt pathology. After two open-label trials assessing the safety of *T. suis* infection in IBD patients and showing promising results (about 70% remission in Crohn's disease^{133,134}), Summers and colleagues set out to study the effect of TSO in a first placebo-controlled double blind randomized trial including 54 ulcerative colitis (UC) patients¹³⁵. The UC Disease Activity Index (UCDAI) improved more in the TSO group compared to the placebo group, however numbers of remissions were not significantly different. The group of P'ng Loke, working on characterization of the local immune responses surrounding the location of *Trichuris* worms, studied a patient who infected himself with *T. trichiura*¹³⁶. In this patient, during colitis, T cells only producing IL-17 were abundant, while after *Trichuris* infection more multifunctional T cells were induced, mainly producing IL-22. At the same time goblet cell hyperplasia and enhanced mucus production was seen, which suggests that IL-22 is involved in the tissue repair response, possibly together with the canonical Th2 cytokines¹³⁶. In addition to the previous observations of helminth-associated *de novo* induction of regulatory cells, it appears that *Trichuris* worms are able to modify the cytokine signature of local inflammatory cells. In rhesus macaques with idiopathic chronic diarrhoea, which resembles UC in intestinal inflammation, the Loke group further examined the effects of *T. trichiura* treatment¹³⁷. Interestingly, four of the five monkeys showed clinical improvement. Colonic T cells produced more IL-4 but had less Tregs after treatment, however, no data could be generated on IL-17 and IL-22 responses, which makes it difficult to correlate these clinical effects to the previously reported immunological alterations. However, IL-22 might be a promising candidate as correlate of protection and this may be further explored in helminthic therapy trials, with *T. trichiura* or other helminths.

Recently, the results of a safety trial of TSO in MS patients were reported, as a starting point for a planned phase 2 trial (trial identifier NCT00645749). The trial followed 5 patients with relapsing-remitting MS after inoculation with TSO¹³⁸. Although the majority experienced mild gastrointestinal symptoms, the number of new lesions revealed by MRI was lower during TSO treatment than the number before treatment, or after treatment had been discontinued. This was not accompanied by a change in circulating Tregs or monocytes expressing typical molecules of alternatively activated macrophages, raising the question whether it

is difficult to detect these cells in peripheral blood of patients rather than in the intestine.

A relatively large RCT in 100 allergic rhinitis patients showed the induction of gastrointestinal symptoms and immunological response to TSO, but no effect in reducing symptom scores, medication use or skin prick test reactivity¹³⁹. However, this trial has been criticized due to the fact that infection with TSO was too short before the hay fever season started and therefore might not have allowed sufficient time for the treatment to work¹⁴⁰.

As of December 2012, there are a total of 11 clinical trials registered assessing safety and/or efficacy of TSO in IBD, MS, allergies and even autism (Table 1).

Necator americanus larvae

Relatively new is the therapeutic potential of worms known to establish chronic infections in humans. An advantage of this could be that administration is only needed once, whereas TSO should be administered at two- or three-weekly intervals. The safety and efficacy of *Necator americanus* larvae has been assessed in three trials. Inoculation with 50 larvae or more was shown to cause considerable gastrointestinal symptoms in healthy volunteers¹⁴¹⁻¹⁴³. A dose-ranging trial to establish the dose which would achieve an infection intensity of 50 eggs per gram of faeces in healthy individuals showed that inoculation with 10 larvae was sufficient. This dose also induced a modest immunological response, as measured by eosinophil counts, IgE and hookworm-specific IgG levels¹⁴³.

Before starting a RCT of *N. americanus* infection in asthma patients, two other issues were addressed, which could potentially affect the safety of these studies. It was shown in allergic rhinitis patients that the lung passage of hookworm larvae did not cause deterioration in airway reactivity¹⁴⁴ and that hookworm-induced type 2 responses did not potentiate an allergen-specific IgE response¹⁴⁵. In an asthma trial that followed, although infection using 10 larvae was well tolerated, it did not show any beneficial effect against asthma symptoms¹⁴⁶. One RCT has been reported on *N. americanus* infection in celiac disease patients under restricted diet¹⁴⁷. Wheat challenge was performed to assess the effect that helminthic immune modulation would have, however no differences in mucosal T cell counts, gluten-specific IFN- γ production by PBMC or clinical responses to challenge were observed comparing groups of 10 patients. Currently five clinical trials are registered to use *N. americanus* larvae for celiac disease, allergic rhinitis, asthma and MS (Table 1).

Table 1. Overview of registered clinical trials on helminthic therapy

Trial identifier	Sponsor	Phase	Status	Condition	Intervention
ACTRN12608000241336	Asphelia Pharmaceuticals	1	not yet recruiting	Crohn's disease	TSO
EUCTR2007-006099-12-DK	Statens Serum Institut, Denmark	2	not recruiting	Allergic rhinitis	TSO
NCT00645749	University of Wisconsin, Madison, US	2	recruiting	MS	TSO
NCT01005941	Rigshospitalet, Copenhagen, Denmark	2	completed	MS	TSO
NCT01040221	Montefiore, New York, US	1	not yet recruiting	Autism	TSO
NCT01070498	Beth Israel, Boston, US	1	completed	Food allergy	TSO
NCT01279577 / EUCTR2006-000720-13-DE	Dr. Falk Pharma, Frankfurt, Germany	2	recruiting	Crohn's disease	TSO
NCT01413243 / EUCTR2009-015319-41-DE	Charite, Berlin, Germany	2	recruiting	MS	TSO
NCT01433471	NYU, New York, US	?	recruiting	Ulcerative colitis	TSO
NCT01434693	Coronado Biosciences, US	1	ongoing	Crohn's disease	TSO
NCT01576471	Coronado Biosciences, US	2	recruiting	Crohn's disease	TSO
NCT01734941	Hadassah Medical Organization, Jerusalem, Israel	2	not yet recruiting	Autism	TSO
NCT00232518	University of Nottingham, UK	1	completed	Allergic rhinoconjunctivitis	N.a. larvae
NCT00465989	University of Nottingham, UK	1	completed	Asthma	N.a. larvae
NCT00671138	Brisbane, Australia	2	unknown	Celiac Disease	N.a. larvae
EUCTR2008-005008-24-GB	University of Nottingham, UK	2	recruiting	MS	N.a. larvae
NCT01661933	Brisbane, Australia	1&2	recruiting*	Celiac Disease	N.a. larvae

TSO *Trichuris suis* ova; N.a. *Necator americanus*; *enrolling by invitation only

Helminth-derived molecules

Regarding the fact that helminth species currently used or planned to be used in trials are able to colonize the human intestine or could have clinical and pathological consequences, there has been a shift on focusing on helminth-derived molecules to substitute whole parasite treatment approach (reviewed by William and Margaret Harnett¹⁴⁸).

As already alluded to earlier, several helminthic products with immune modulating properties have been defined. For several of these, cellular immunological responses induced have been investigated and some have been tested in disease models, however none has been administered to humans. Currently, ES-62 is the most well-characterized candidate molecule for therapeutic trials. This phosphorylcholine-coupled glycoprotein was first identified in 1994¹⁴⁹ and, as discussed, has been used to treat arthritis in a murine model¹⁰¹. Furthermore, through inhibition of mast cell histamine release, ES-62 may protect against allergic diseases¹⁵⁰.

The *Heligmosomoides* excreted-secreted (ES) products⁷⁶, characterized by the group of Rick Maizels, have been shown to suppress allergic airway inflammation⁵⁵. AvCystatin, another molecule secreted by *A.viteae*, inhibits the development of allergic airway inflammation and acute colitis¹⁵¹. Moreover, it has been shown that *in vitro* Th2 responses of PBMC from grass pollen allergic patients are markedly reduced by adding AvCystatin to cultures¹⁵². Furthermore, there is data indicating that soluble products from *S. mansoni*, *T. suis* and *Trichinella spiralis* can suppress clinical signs of EAE by modulation of DC⁸⁸.

In murine colitis, extracts from *S. mansoni* adult worms and ES products of the canine hookworm *Ankylostoma caninum* have displayed beneficial effects¹⁵³. Although treatment with *S. mansoni* extracts did not improve the clinical score, it diminished local inflammation and myeloid cell infiltration in colonic tissue. In parallel, lower mucosal Th1 and Th17 responses and enhanced expression of IL-10 and TGF- β in T cells was found¹⁵³. These results illustrate that altered immune responses do not always lead to clinical improvement, and a longer time period may be needed for a detectable clinically beneficial effect. Lastly, Lewis χ -containing glycan from *S. mansoni* eggs, lacto-N-fucopentaose III (LNFP III), was tested in mouse models of inflammatory conditions. LNFP III suppressed EAE by enhancement of IL-10 and Th2 cytokines¹⁵⁴ and was also shown to be beneficial in psoriasis¹⁵⁵. Taken together these results in animal models encourage further studies, and possibly clinical trials, to evaluate their beneficial effects.

Challenges in helminth immunotherapy

There are some drawbacks in the use of helminthic therapy as currently proposed. As mentioned before, infection with *N. americanus* but possibly also *T. suis* could have pathogenic effects in humans. Patients undergoing helminth infection should

be monitored closely for infection intensity and possible extra-intestinal manifestations of the infection¹⁵⁶. The long-term consequences of this approach have not been assessed; the trials with the longest follow-up time were for 24 weeks. The advantage for introducing hookworm infections would be that only single inoculation is needed, whereas TSO should be provided every two to three weeks. However, this also implies that hookworm infections are less controllable, as they lead to chronic infestations. The timing of infection is another issue, since under natural conditions, the protective effects of helminthic infections might have been acquired in early life. Moreover, possible relevant immunomodulatory effects could need substantial length of time to fully develop.

Another issue is the helminth-induced immune modulation itself. Immune regulatory responses are desirable to counteract inflammatory disorders, but could be detrimental for other immune-associated conditions. Not only defence against incoming pathogens may be impaired, but also anti-tumor immune responses may be compromised. Immunosuppression is a strategy for tumor cells to evade host immune responses and efforts are being made to inhibit Tregs in cancer by immunotherapy¹⁵⁷. Therefore, non-specific blanket immune suppression, although mild, should be considered critically.

Emerging area – Immunometabolism

Immunometabolism is an emerging concept, which explores the interaction between nutrients, metabolism and the immune system. Since metabolic disorders such as T2D and obesity are associated with immune alterations, they can be regarded as inflammatory diseases⁹⁵. It has become clear that some of the pathways involved in pathogen sensing are also involved in the induction of inflammation that leads to metabolic disorders. Pathogen-associated molecules are known to induce pro-inflammatory cytokine production, such as TNF, through toll-like receptor (TLR) signaling¹⁵⁸. Large amounts of TNF are found in adipose tissue of obese mice¹⁵⁹. The intracellular signaling pathways of TNF involve serine kinases, which have been shown to phosphorylate serine residues in insulin receptor substrates^{160,161}. As serine phosphorylation is the inhibitory counterpart of tyrosine phosphorylation usually induced by insulin binding to the receptor, the signaling events of TNF lead to inhibition of insulin signaling, resulting in insulin resistance¹⁶¹.

Combatting microorganisms has high energy demand and moreover, worms and other parasites use host nutrients for their long-term survival. As a consequence, parasitic worms might need to tightly co-regulate immune responses and nutrient metabolism, to avoid depriving their host from resources necessary for survival. Therefore, these metabolic properties of parasitic infections could be of interest. Recently the effects of LNFPIII, which is not only present on *S. mansoni* eggs but

also found in human breast milk¹⁶², was assessed on immune metabolism. LNFP III was already shown to induce an immunoregulatory phenotype in macrophages¹⁶³ but in addition, Bhargava and colleagues reported that this glycoconjugate improves insulin signaling and thereby sensitivity in white adipose tissue in high fat diet-fed mice, in part through IL-10¹⁶⁴. Furthermore, some detailed studies of the interaction of Th2 cytokines and metabolic homeostasis have shown that IL-4 shifts cellular energy resources from fatty acids to glucose oxidation, by enhancing the activity of insulin¹⁶⁵. These studies highlight the possible metabolic advantages of harboring helminth infections.

Final remarks

In summary, there is an apparent contrast between efforts to deworm populations in remote areas suffering from helminth-associated morbidities and initiatives to test effects of helminthic therapy on patients with hyperinflammatory diseases (Figure 1).

Murine models have supported the hypothesis that helminths or their products may be beneficial for inflammatory conditions. Human studies in poor-resource settings have been less consistent, which may be explained by the presence of other modulating infections and by the fact that the clinical effects of deworming may take longer to establish. Further deworming trials should take these issues into account and plan for longer follow-up periods. Human studies of the therapeutic use of helminths in resource-rich settings may shed more light on the role of helminths in human physiology and immunology. Although some positive results have been obtained in IBD and MS, not much benefit was seen in treatment of asthma and allergy. Further trials should be less modest in numbers of patients included. Furthermore, assessment of locally – and importantly – systemically induced immune (regulatory) responses deserves further attention, since the underlying mechanisms and consequences of voluntary helminth infection could influence other health outcomes in patients.

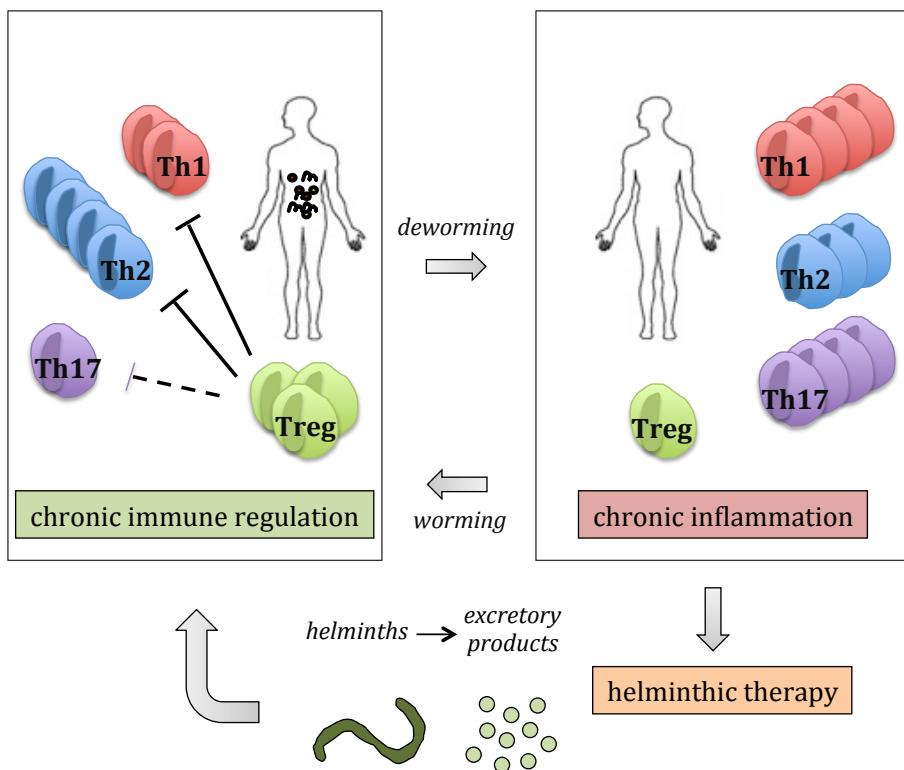


Figure 1. The immunological effects of deworming and worming. Helminth-infected individuals (left panel) harbor enhanced Th2 responses. The number and immunosuppressive capacity of Tregs may also be increased during infection. After deworming, removal of immune suppression could lead to over-inflammation, characteristic of several inflammatory diseases (right panel). Th1, Th2 or Th17 responses are more abundant, whereas Tregs are less in number and function. Treatment with experimental helminth infection or with helminth-derived immunomodulatory molecules could restore the immune regulation as observed during natural chronic helminth infections. The resulting suppression of inflammatory T-cell responses may curtail symptoms of several immune-mediated diseases.

References

1. Hoepli, R. The knowledge of parasites and parasitic infections from ancient times to the 17th century. *Experimental parasitology* **5**, 398-419 (1956).
2. Bundy, D.A. & Medley, G.F. Immuno-epidemiology of human geohelminthiasis: ecological and immunological determinants of worm burden. *Parasitology* **104 Suppl**, S105-119 (1992).
3. Allen, J.E. & Maizels, R.M. Diversity and dialogue in immunity to helminths. *Nature reviews. Immunology* **11**, 375-388 (2011).
4. Fumagalli, M., et al. Parasites represent a major selective force for interleukin genes and shape the genetic predisposition to autoimmune conditions. *The Journal of experimental medicine* **206**, 1395-1408 (2009).
5. Bach, J.F. The effect of infections on susceptibility to autoimmune and allergic diseases. *The New England journal of medicine* **347**, 911-920 (2002).
6. Anderson, H.R., Gupta, R., Strachan, D.P. & Limb, E.S. 50 years of asthma: UK trends from 1955 to 2004. *Thorax* **62**, 85-90 (2007).
7. Pearce, N., et al. Worldwide trends in the prevalence of asthma symptoms: phase III of the International Study of Asthma and Allergies in Childhood (ISAAC). *Thorax* **62**, 758-766 (2007).
8. Odhiambo, J.A., Williams, H.C., Clayton, T.O., Robertson, C.F. & Asher, M.I. Global variations in prevalence of eczema symptoms in children from ISAAC Phase Three. *The Journal of allergy and clinical immunology* **124**, 1251-1258 e1223 (2009).
9. Strachan, D.P. Hay fever, hygiene, and household size. *BMJ* **299**, 1259-1260 (1989).
10. Zhu, J. & Paul, W.E. CD4 T cells: fates, functions, and faults. *Blood* **112**, 1557-1569 (2008).
11. Maizels, R.M., Hewitson, J.P. & Smith, K.A. Susceptibility and immunity to helminth parasites. *Current opinion in immunology* **24**, 459-466 (2012).
12. Chen, F., et al. An essential role for TH2-type responses in limiting acute tissue damage during experimental helminth infection. *Nature medicine* **18**, 260-266 (2012).
13. Mentink-Kane, M.M., et al. Accelerated and progressive and lethal liver fibrosis in mice that lack interleukin (IL)-10, IL-12p40, and IL-13Ralpha2. *Gastroenterology* **141**, 2200-2209 (2011).
14. Allen, J.E. & Wynn, T.A. Evolution of Th2 immunity: a rapid repair response to tissue destructive pathogens. *PLoS pathogens* **7**, e1002003 (2011).
15. Belkaid, Y. Regulatory T cells and infection: a dangerous necessity. *Nature reviews. Immunology* **7**, 875-888 (2007).
16. Maizels, R.M. & Yazdanbakhsh, M. Immune regulation by helminth parasites: cellular and molecular mechanisms. *Nature reviews. Immunology* **3**, 733-744 (2003).
17. Colley, D.G., Todd, C.W., Lewis, F.A. & Goodgame, R.W. Immune responses during human schistosomiasis mansoni. VI. In vitro nonspecific suppression of phytohemagglutinin responsiveness induced by exposure to certain schistosomal preparations. *J Immunol* **122**, 1447-1453 (1979).
18. Lewert, R.M., Yogore, M.G., Jr. & Blas, B.L. Lymphocyte responsiveness to phytohemagglutinin and to worm and egg antigens in human schistosomiasis japonica. *The American journal of tropical medicine and hygiene* **28**, 92-98 (1979).
19. Ottesen, E.A., Weller, P.F. & Heck, L. Specific cellular immune unresponsiveness in human filariasis. *Immunology* **33**, 413-421 (1977).
20. Ottesen, E.A., Hiatt, R.A., Cheever, A.W., Sotomayor, Z.R. & Neva, F.A. The acquisition and loss of antigen-specific cellular immune responsiveness in acute

and chronic schistosomiasis in man. *Clinical and experimental immunology* **33**, 37-47 (1978).

21. Maizels, R.M., Bundy, D.A., Selkirk, M.E., Smith, D.F. & Anderson, R.M. Immunological modulation and evasion by helminth parasites in human populations. *Nature* **365**, 797-805 (1993).
22. Piessens, W.F., et al. Antigen-specific suppressor cells and suppressor factors in human filariasis with *Brugia malayi*. *The New England journal of medicine* **302**, 833-837 (1980).
23. Babu, S., Kumaraswami, V. & Nutman, T.B. Alternatively activated and immunoregulatory monocytes in human filarial infections. *The Journal of infectious diseases* **199**, 1827-1837 (2009).
24. Semnani, R.T., Mahapatra, L., Moore, V., Sanprasert, V. & Nutman, T.B. Functional and phenotypic characteristics of alternative activation induced in human monocytes by interleukin-4 or the parasitic nematode *Brugia malayi*. *Infection and immunity* **79**, 3957-3965 (2011).
25. Van den Bossche, J., et al. Alternatively activated macrophages engage in homotypic and heterotypic interactions through IL-4 and polyamine-induced E-cadherin/catenin complexes. *Blood* **114**, 4664-4674 (2009).
26. Li, Z., et al. The phenotype and function of naturally existing regulatory dendritic cells in nematode-infected mice. *International journal for parasitology* **41**, 1129-1137 (2011).
27. Piessens, W.F., et al. Antigen-specific suppressor T lymphocytes in human lymphatic filariasis. *The New England journal of medicine* **307**, 144-148 (1982).
28. MacDonald, T.T. Suppressor T cells, rebranded as regulatory T cells, emerge from the wilderness bearing surface markers. *Gut* **51**, 311-312 (2002).
29. Levings, M.K. & Roncarolo, M.G. T-regulatory 1 cells: a novel subset of CD4 T cells with immunoregulatory properties. *The Journal of allergy and clinical immunology* **106**, S109-112 (2000).
30. Mills, K.H. & McGuirk, P. Antigen-specific regulatory T cells--their induction and role in infection. *Seminars in immunology* **16**, 107-117 (2004).
31. Babu, S., Blauvelt, C.P., Kumaraswami, V. & Nutman, T.B. Regulatory networks induced by live parasites impair both Th1 and Th2 pathways in patent lymphatic filariasis: implications for parasite persistence. *J Immunol* **176**, 3248-3256 (2006).
32. Metenou, S., et al. At homeostasis filarial infections have expanded adaptive T regulatory but not classical Th2 cells. *J Immunol* **184**, 5375-5382 (2010).
33. Nausch, N., Midzi, N., Mduluza, T., Maizels, R.M. & Mutapi, F. Regulatory and activated T cells in human *Schistosoma haematobium* infections. *PloS one* **6**, e16860 (2011).
34. Ricci, N.D., et al. Induction of CD4(+)CD25(+)FOXP3(+) regulatory T cells during human hookworm infection modulates antigen-mediated lymphocyte proliferation. *PLoS neglected tropical diseases* **5**, e1383 (2011).
35. Teixeira-Carvalho, A., et al. Cytokines, chemokine receptors, CD4+CD25HIGH+ T-cells and clinical forms of human schistosomiasis. *Acta tropica* **108**, 139-149 (2008).
36. Watanabe, K., et al. T regulatory cell levels decrease in people infected with *Schistosoma mansoni* on effective treatment. *The American journal of tropical medicine and hygiene* **77**, 676-682 (2007).
37. Wammes, L.J., et al. Regulatory T cells in human lymphatic filariasis: stronger functional activity in microfilaremics. *PLoS neglected tropical diseases* **6**, e1655 (2012).

38. van der Kleij, D., *et al.* A novel host-parasite lipid cross-talk. Schistosomal lyso-phosphatidylserine activates toll-like receptor 2 and affects immune polarization. *The Journal of biological chemistry* **277**, 48122-48129 (2002).
39. Grainger, J.R., *et al.* Helminth secretions induce de novo T cell Foxp3 expression and regulatory function through the TGF-beta pathway. *The Journal of experimental medicine* **207**, 2331-2341 (2010).
40. Zaccone, P., *et al.* The *S. mansoni* glycoprotein omega-1 induces Foxp3 expression in NOD mouse CD4(+) T cells. *European journal of immunology* **41**, 2709-2718 (2011).
41. Correale, J., Farez, M. & Razzitte, G. Helminth infections associated with multiple sclerosis induce regulatory B cells. *Annals of neurology* **64**, 187-199 (2008).
42. van der Vlugt, L.E., *et al.* Schistosomes induce regulatory features in human and mouse CD1d(hi) B cells: inhibition of allergic inflammation by IL-10 and regulatory T cells. *PloS one* **7**, e30883 (2012).
43. Anthony, R.M., Rutitzky, L.I., Urban, J.F., Jr., Stadecker, M.J. & Gause, W.C. Protective immune mechanisms in helminth infection. *Nature reviews. Immunology* **7**, 975-987 (2007).
44. Donnelly, S., *et al.* Helminth 2-Cys peroxiredoxin drives Th2 responses through a mechanism involving alternatively activated macrophages. *FASEB journal : official publication of the Federation of American Societies for Experimental Biology* **22**, 4022-4032 (2008).
45. Rzepecka, J., *et al.* Calreticulin from the intestinal nematode *Heligmosomoides polygyrus* is a Th2-skewing protein and interacts with murine scavenger receptor-A. *Molecular immunology* **46**, 1109-1119 (2009).
46. Everts, B., *et al.* Omega-1, a glycoprotein secreted by *Schistosoma mansoni* eggs, drives Th2 responses. *The Journal of experimental medicine* **206**, 1673-1680 (2009).
47. Whelan, M., *et al.* A filarial nematode-secreted product signals dendritic cells to acquire a phenotype that drives development of Th2 cells. *J Immunol* **164**, 6453-6460 (2000).
48. Fallon, P.G., *et al.* Identification of an interleukin (IL)-25-dependent cell population that provides IL-4, IL-5, and IL-13 at the onset of helminth expulsion. *The Journal of experimental medicine* **203**, 1105-1116 (2006).
49. Humphreys, N.E., Xu, D., Hepworth, M.R., Liew, F.Y. & Grencis, R.K. IL-33, a potent inducer of adaptive immunity to intestinal nematodes. *J Immunol* **180**, 2443-2449 (2008).
50. Hurst, S.D., *et al.* New IL-17 family members promote Th1 or Th2 responses in the lung: in vivo function of the novel cytokine IL-25. *J Immunol* **169**, 443-453 (2002).
51. Koyasu, S., Moro, K., Tanabe, M. & Takeuchi, T. Natural helper cells: a new player in the innate immune response against helminth infection. *Advances in immunology* **108**, 21-44 (2010).
52. Saenz, S.A., *et al.* IL25 elicits a multipotent progenitor cell population that promotes T(H)2 cytokine responses. *Nature* **464**, 1362-1366 (2010).
53. Moro, K., *et al.* Innate production of T(H)2 cytokines by adipose tissue-associated c-Kit(+)Sca-1(+) lymphoid cells. *Nature* **463**, 540-544 (2010).
54. Mangan, N.E., *et al.* Helminth infection protects mice from anaphylaxis via IL-10-producing B cells. *J Immunol* **173**, 6346-6356 (2004).
55. McSorley, H.J., *et al.* Suppression of type 2 immunity and allergic airway inflammation by secreted products of the helminth *Heligmosomoides polygyrus*. *European journal of immunology* **42**, 2667-2682 (2012).

56. Smits, H.H., *et al.* Protective effect of *Schistosoma mansoni* infection on allergic airway inflammation depends on the intensity and chronicity of infection. *The Journal of allergy and clinical immunology* **120**, 932-940 (2007).

57. Wilson, M.S., *et al.* Suppression of allergic airway inflammation by helminth-induced regulatory T cells. *The Journal of experimental medicine* **202**, 1199-1212 (2005).

58. Leonardi-Bee, J., Pritchard, D. & Britton, J. Asthma and current intestinal parasite infection: systematic review and meta-analysis. *American journal of respiratory and critical care medicine* **174**, 514-523 (2006).

59. Flohr, C., Quinnell, R.J. & Britton, J. Do helminth parasites protect against atopy and allergic disease? *Clinical and experimental allergy : journal of the British Society for Allergy and Clinical Immunology* **39**, 20-32 (2009).

60. Araujo, M.I. & de Carvalho, E.M. Human schistosomiasis decreases immune responses to allergens and clinical manifestations of asthma. *Chemical immunology and allergy* **90**, 29-44 (2006).

61. Cooper, P.J., *et al.* Reduced risk of atopy among school-age children infected with geohelminth parasites in a rural area of the tropics. *The Journal of allergy and clinical immunology* **111**, 995-1000 (2003).

62. Flohr, C., *et al.* Poor sanitation and helminth infection protect against skin sensitization in Vietnamese children: A cross-sectional study. *The Journal of allergy and clinical immunology* **118**, 1305-1311 (2006).

63. Mpairwe, H., *et al.* Skin prick test reactivity to common allergens among women in Entebbe, Uganda. *Transactions of the Royal Society of Tropical Medicine and Hygiene* **102**, 367-373 (2008).

64. Rodrigues, L.C., *et al.* Early infection with *Trichuris trichiura* and allergen skin test reactivity in later childhood. *Clinical and experimental allergy : journal of the British Society for Allergy and Clinical Immunology* **38**, 1769-1777 (2008).

65. van den Biggelaar, A.H., *et al.* Decreased atopy in children infected with *Schistosoma haematobium*: a role for parasite-induced interleukin-10. *Lancet* **356**, 1723-1727 (2000).

66. Palmer, L.J., *et al.* *Ascaris lumbricoides* infection is associated with increased risk of childhood asthma and atopy in rural China. *American journal of respiratory and critical care medicine* **165**, 1489-1493 (2002).

67. Schafer, T., Meyer, T., Ring, J., Wichmann, H.E. & Heinrich, J. Worm infestation and the negative association with eczema (atopic/nonatopic) and allergic sensitization. *Allergy* **60**, 1014-1020 (2005).

68. Wordemann, M., *et al.* Association of atopy, asthma, allergic rhinoconjunctivitis, atopic dermatitis and intestinal helminth infections in Cuban children. *Tropical medicine & international health : TM & IH* **13**, 180-186 (2008).

69. Haileamlak, A., *et al.* Early life risk factors for atopic dermatitis in Ethiopian children. *The Journal of allergy and clinical immunology* **115**, 370-376 (2005).

70. Dagoye, D., *et al.* Wheezing, allergy, and parasite infection in children in urban and rural Ethiopia. *American journal of respiratory and critical care medicine* **167**, 1369-1373 (2003).

71. Scrivener, S., *et al.* Independent effects of intestinal parasite infection and domestic allergen exposure on risk of wheeze in Ethiopia: a nested case-control study. *Lancet* **358**, 1493-1499 (2001).

72. Cooper, P.J., Chico, M.E., Bland, M., Griffin, G.E. & Nutman, T.B. Allergic symptoms, atopy, and geohelminth infections in a rural area of Ecuador. *American journal of respiratory and critical care medicine* **168**, 313-317 (2003).

73. Peisong, G., *et al.* An asthma-associated genetic variant of STAT6 predicts low burden of *Ascaris* worm infestation. *Genes and immunity* **5**, 58-62 (2004).

74. Moller, M., *et al.* Genetic haplotypes of Th-2 immune signalling link allergy to enhanced protection to parasitic worms. *Human molecular genetics* **16**, 1828-1836 (2007).
75. Elliott, D.E., *et al.* Exposure to schistosome eggs protects mice from TNBS-induced colitis. *American journal of physiology. Gastrointestinal and liver physiology* **284**, G385-391 (2003).
76. Khan, W.I., *et al.* Intestinal nematode infection ameliorates experimental colitis in mice. *Infection and immunity* **70**, 5931-5937 (2002).
77. Reardon, C., Sanchez, A., Hogaboam, C.M. & McKay, D.M. Tapeworm infection reduces epithelial ion transport abnormalities in murine dextran sulfate sodium-induced colitis. *Infection and immunity* **69**, 4417-4423 (2001).
78. Melon, A., Wang, A., Phan, V. & McKay, D.M. Infection with *Hymenolepis diminuta* is more effective than daily corticosteroids in blocking chemically induced colitis in mice. *Journal of biomedicine & biotechnology* **2010**, 384523 (2010).
79. Kabeerdoss, J., Pugazhendhi, S., Subramanian, V., Binder, H.J. & Ramakrishna, B.S. Exposure to hookworms in patients with Crohn's disease: a case-control study. *Alimentary pharmacology & therapeutics* **34**, 923-930 (2011).
80. Leibowitz, U., *et al.* Epidemiological study of multiple sclerosis in Israel. II. Multiple sclerosis and level of sanitation. *Journal of neurology, neurosurgery, and psychiatry* **29**, 60-68 (1966).
81. Fleming, J.O. & Cook, T.D. Multiple sclerosis and the hygiene hypothesis. *Neurology* **67**, 2085-2086 (2006).
82. Correale, J. & Farez, M. Association between parasite infection and immune responses in multiple sclerosis. *Annals of neurology* **61**, 97-108 (2007).
83. Correale, J. & Farez, M.F. The impact of parasite infections on the course of multiple sclerosis. *Journal of neuroimmunology* **233**, 6-11 (2011).
84. Donskow-Lysoniewska, K., Krawczak, K. & Doligalska, M. *Heligmosomoides polygyrus*: EAE remission is correlated with different systemic cytokine profiles provoked by L4 and adult nematodes. *Experimental parasitology* **132**, 243-248 (2012).
85. Sewell, D., *et al.* Immunomodulation of experimental autoimmune encephalomyelitis by helminth ova immunization. *International immunology* **15**, 59-69 (2003).
86. Walsh, K.P., Brady, M.T., Finlay, C.M., Boon, L. & Mills, K.H. Infection with a helminth parasite attenuates autoimmunity through TGF-beta-mediated suppression of Th17 and Th1 responses. *J Immunol* **183**, 1577-1586 (2009).
87. La Flamme, A.C., Ruddenklau, K. & Backstrom, B.T. Schistosomiasis decreases central nervous system inflammation and alters the progression of experimental autoimmune encephalomyelitis. *Infection and immunity* **71**, 4996-5004 (2003).
88. Kuijk, L.M., *et al.* Soluble helminth products suppress clinical signs in murine experimental autoimmune encephalomyelitis and differentially modulate human dendritic cell activation. *Molecular immunology* **51**, 210-218 (2012).
89. Cooke, A., *et al.* Infection with *Schistosoma mansoni* prevents insulin dependent diabetes mellitus in non-obese diabetic mice. *Parasite immunology* **21**, 169-176 (1999).
90. Hubner, M.P., Stocker, J.T. & Mitre, E. Inhibition of type 1 diabetes in filaria-infected non-obese diabetic mice is associated with a T helper type 2 shift and induction of FoxP3+ regulatory T cells. *Immunology* **127**, 512-522 (2009).
91. Saunders, K.A., Raine, T., Cooke, A. & Lawrence, C.E. Inhibition of autoimmune type 1 diabetes by gastrointestinal helminth infection. *Infection and immunity* **75**, 397-407 (2007).

92. Robinson, M.W., Dalton, J.P., O'Brien, B.A. & Donnelly, S. *Fasciola hepatica*: The therapeutic potential of a worm secretome. *International journal for parasitology* (2012).

93. Zaccone, P., Fehervari, Z., Phillips, J.M., Dunne, D.W. & Cooke, A. Parasitic worms and inflammatory diseases. *Parasite immunology* **28**, 515-523 (2006).

94. Aravindhan, V., et al. Decreased prevalence of lymphatic filariasis among subjects with type-1 diabetes. *The American journal of tropical medicine and hygiene* **83**, 1336-1339 (2010).

95. Donath, M.Y. & Shoelson, S.E. Type 2 diabetes as an inflammatory disease. *Nature reviews. Immunology* **11**, 98-107 (2011).

96. Aravindhan, V., et al. Decreased prevalence of lymphatic filariasis among diabetic subjects associated with a diminished pro-inflammatory cytokine response (CURES 83). *PLoS neglected tropical diseases* **4**, e707 (2010).

97. van Kuijk, A.W., Kerstens, P.J., Perenboom, R.M., Dijkmans, B.A. & Voskuyl, A.E. Early-onset polyarthritis as presenting feature of intestinal infection with *Strongyloides stercoralis*. *Rheumatology (Oxford)* **42**, 1419-1420 (2003).

98. Osada, Y., Shimizu, S., Kumagai, T., Yamada, S. & Kanazawa, T. *Schistosoma mansoni* infection reduces severity of collagen-induced arthritis via down-regulation of pro-inflammatory mediators. *International journal for parasitology* **39**, 457-464 (2009).

99. Rocha, F.A., et al. Protective effect of an extract from *Ascaris suum* in experimental arthritis models. *Infection and immunity* **76**, 2736-2745 (2008).

100. Salinas-Carmona, M.C., et al. Spontaneous arthritis in MRL/lpr mice is aggravated by *Staphylococcus aureus* and ameliorated by *Nippostrongylus brasiliensis* infections. *Autoimmunity* **42**, 25-32 (2009).

101. McInnes, I.B., et al. A novel therapeutic approach targeting articular inflammation using the filarial nematode-derived phosphorylcholine-containing glycoprotein ES-62. *J Immunol* **171**, 2127-2133 (2003).

102. Aoyama, H., et al. An inverse relationship between autoimmune liver diseases and *Strongyloides stercoralis* infection. *The American journal of tropical medicine and hygiene* **76**, 972-976 (2007).

103. Nagayama, Y., Watanabe, K., Niwa, M., McLachlan, S.M. & Rapoport, B. *Schistosoma mansoni* and alpha-galactosylceramide: prophylactic effect of Th1 Immune suppression in a mouse model of Graves' hyperthyroidism. *J Immunol* **173**, 2167-2173 (2004).

104. Magen, E., Borkow, G., Bentwich, Z., Mishal, J. & Scharf, S. Can worms defend our hearts? Chronic helminthic infections may attenuate the development of cardiovascular diseases. *Medical hypotheses* **64**, 904-909 (2005).

105. Doenhoff, M.J., Stanley, R.G., Griffiths, K. & Jackson, C.L. An anti-atherogenic effect of *Schistosoma mansoni* infections in mice associated with a parasite-induced lowering of blood total cholesterol. *Parasitology* **125**, 415-421 (2002).

106. Hotez, P.J., et al. Helminth infections: the great neglected tropical diseases. *The Journal of clinical investigation* **118**, 1311-1321 (2008).

107. Utzinger, J. A research and development agenda for the control and elimination of human helminthiases. *PLoS neglected tropical diseases* **6**, e1646 (2012).

108. Lynch, N.R., et al. Effect of anthelmintic treatment on the allergic reactivity of children in a tropical slum. *The Journal of allergy and clinical immunology* **92**, 404-411 (1993).

109. van den Biggelaar, A.H., et al. Long-term treatment of intestinal helminths increases mite skin-test reactivity in Gabonese schoolchildren. *The Journal of infectious diseases* **189**, 892-900 (2004).

110. Cooper, P.J., *et al.* Effect of albendazole treatments on the prevalence of atopy in children living in communities endemic for geohelminth parasites: a cluster-randomised trial. *Lancet* **367**, 1598-1603 (2006).
111. Flohr, C., *et al.* Reduced helminth burden increases allergen skin sensitization but not clinical allergy: a randomized, double-blind, placebo-controlled trial in Vietnam. *Clin Exp Allergy* (2009).
112. Wiria, A.E., *et al.* The effect of three-monthly albendazole treatment on malarial parasitemia and allergy: a household-based cluster-randomized, double-blind, placebo-controlled trial. *PloS one* **8**, e57899 (2013).
113. Matricardi, P.M., *et al.* Exposure to foodborne and orofecal microbes versus airborne viruses in relation to atopy and allergic asthma: epidemiological study. *BMJ* **320**, 412-417 (2000).
114. Pelosi, U., *et al.* The inverse association of salmonellosis in infancy with allergic rhinoconjunctivitis and asthma at school-age: a longitudinal study. *Allergy* **60**, 626-630 (2005).
115. Lell, B., Borrman, S., Yazdanbakhsh, M. & Kremsner, P.G. Atopy and malaria. *Wiener klinische Wochenschrift* **113**, 927-929 (2001).
116. Endara, P., *et al.* Long-term periodic anthelmintic treatments are associated with increased allergen skin reactivity. *Clinical and experimental allergy : journal of the British Society for Allergy and Clinical Immunology* **40**, 1669-1677 (2010).
117. Mpairowe, H., *et al.* Anthelmintic treatment during pregnancy is associated with increased risk of infantile eczema: randomised-controlled trial results. *Pediatric allergy and immunology : official publication of the European Society of Pediatric Allergy and Immunology* **22**, 305-312 (2011).
118. Ndirbazza, J., *et al.* Impact of anthelmintic treatment in pregnancy and childhood on immunisations, infections and eczema in childhood: a randomised controlled trial. *PloS one* **7**, e50325 (2012).
119. Ndirbazza, J., *et al.* Effects of deworming during pregnancy on maternal and perinatal outcomes in Entebbe, Uganda: a randomized controlled trial. *Clinical infectious diseases : an official publication of the Infectious Diseases Society of America* **50**, 531-540 (2010).
120. Webb, E.L., *et al.* Effect of single-dose anthelmintic treatment during pregnancy on an infant's response to immunisation and on susceptibility to infectious diseases in infancy: a randomised, double-blind, placebo-controlled trial. *Lancet* **377**, 52-62 (2011).
121. Almeida, M.C., *et al.* The effect of antihelminthic treatment on subjects with asthma from an endemic area of schistosomiasis: a randomized, double-blinded, and placebo-controlled trial. *Journal of parasitology research* **2012**, 296856 (2012).
122. Bager, P., Vinkel Hansen, A., Wohlfahrt, J. & Melbye, M. Helminth infection does not reduce risk for chronic inflammatory disease in a population-based cohort study. *Gastroenterology* **142**, 55-62 (2012).
123. Cook, G.C. Enterobius vermicularis infection. *Gut* **35**, 1159-1162 (1994).
124. Lacey, E. Mode of action of benzimidazoles. *Parasitol Today* **6**, 112-115 (1990).
125. Skinner-Adams, T.S., Davis, T.M., Manning, L.S. & Johnston, W.A. The efficacy of benzimidazole drugs against *Plasmodium falciparum* in vitro. *Transactions of the Royal Society of Tropical Medicine and Hygiene* **91**, 580-584 (1997).
126. MacDonald, L.M., Armon, A., Thompson, A.R. & Reynoldson, J.A. Characterisation of benzimidazole binding with recombinant tubulin from *Giardia duodenalis*, *Encephalitozoon intestinalis*, and *Cryptosporidium parvum*. *Molecular and biochemical parasitology* **138**, 89-96 (2004).

127. Cruz, M.C. & Edlind, T. beta-Tubulin genes and the basis for benzimidazole sensitivity of the opportunistic fungus *Cryptococcus neoformans*. *Microbiology* **143** (Pt 6), 2003-2008 (1997).

128. Cooper, P.J., et al. Repeated treatments with albendazole enhance Th2 responses to *Ascaris Lumbricoides*, but not to aeroallergens, in children from rural communities in the Tropics. *The Journal of infectious diseases* **198**, 1237-1242 (2008).

129. van den Biggelaar, A.H., Borrmann, S., Kremsner, P. & Yazdanbakhsh, M. Immune responses induced by repeated treatment do not result in protective immunity to *Schistosoma haematobium*: interleukin (IL)-5 and IL-10 responses. *The Journal of infectious diseases* **186**, 1474-1482 (2002).

130. Wright, V.J., et al. Early exposure of infants to GI nematodes induces Th2 dominant immune responses which are unaffected by periodic anthelmintic treatment. *PLoS neglected tropical diseases* **3**, e433 (2009).

131. Tweyongyere, R., et al. Effect of praziquantel treatment during pregnancy on cytokine responses to schistosome antigens: results of a randomized, placebo-controlled trial. *The Journal of infectious diseases* **198**, 1870-1879 (2008).

132. Weinstock, J.V. & Elliott, D.E. Translatability of helminth therapy in inflammatory bowel diseases. *International journal for parasitology* (2012).

133. Summers, R.W., et al. *Trichuris suis* seems to be safe and possibly effective in the treatment of inflammatory bowel disease. *The American journal of gastroenterology* **98**, 2034-2041 (2003).

134. Summers, R.W., Elliott, D.E., Urban, J.F., Jr., Thompson, R. & Weinstock, J.V. *Trichuris suis* therapy in Crohn's disease. *Gut* **54**, 87-90 (2005).

135. Summers, R.W., Elliott, D.E., Urban, J.F., Jr., Thompson, R.A. & Weinstock, J.V. *Trichuris suis* therapy for active ulcerative colitis: a randomized controlled trial. *Gastroenterology* **128**, 825-832 (2005).

136. Broadhurst, M.J., et al. IL-22+ CD4+ T cells are associated with therapeutic *trichuris trichiura* infection in an ulcerative colitis patient. *Science translational medicine* **2**, 60ra88 (2010).

137. Broadhurst, M.J., et al. Therapeutic helminth infection of macaques with idiopathic chronic diarrhea alters the inflammatory signature and mucosal microbiota of the colon. *PLoS pathogens* **8**, e1003000 (2012).

138. Fleming, J.O., et al. Probiotic helminth administration in relapsing-remitting multiple sclerosis: a phase 1 study. *Mult Scler* **17**, 743-754 (2011).

139. Bager, P., et al. *Trichuris suis* ova therapy for allergic rhinitis: a randomized, double-blind, placebo-controlled clinical trial. *The Journal of allergy and clinical immunology* **125**, 123-130 e121-123 (2010).

140. Summers, R.W., Elliott, D.E. & Weinstock, J.V. *Trichuris suis* might be effective in treating allergic rhinitis. *The Journal of allergy and clinical immunology* **125**, 766-767 (2010).

141. Croese, J., et al. A proof of concept study establishing *Necator americanus* in Crohn's patients and reservoir donors. *Gut* **55**, 136-137 (2006).

142. Croese, J., Wood, M.J., Melrose, W. & Speare, R. Allergy controls the population density of *Necator americanus* in the small intestine. *Gastroenterology* **131**, 402-409 (2006).

143. Mortimer, K., et al. Dose-ranging study for trials of therapeutic infection with *Necator americanus* in humans. *The American journal of tropical medicine and hygiene* **75**, 914-920 (2006).

144. Feary, J., et al. Safety of hookworm infection in individuals with measurable airway responsiveness: a randomized placebo-controlled feasibility study. *Clinical and experimental allergy : journal of the British Society for Allergy and Clinical Immunology* **39**, 1060-1068 (2009).

145. Blount, D., *et al.* Immunologic profiles of persons recruited for a randomized, placebo-controlled clinical trial of hookworm infection. *The American journal of tropical medicine and hygiene* **81**, 911-916 (2009).
146. Feary, J.R., *et al.* Experimental hookworm infection: a randomized placebo-controlled trial in asthma. *Clinical and experimental allergy : journal of the British Society for Allergy and Clinical Immunology* **40**, 299-306 (2010).
147. Daveson, A.J., *et al.* Effect of hookworm infection on wheat challenge in celiac disease--a randomised double-blinded placebo controlled trial. *PloS one* **6**, e17366 (2011).
148. Harnett, W. & Harnett, M.M. Helminth-derived immunomodulators: can understanding the worm produce the pill? *Nature reviews. Immunology* **10**, 278-284 (2010).
149. Harnett, W., Frame, M.J., Nor, Z.M., MacDonald, M. & Houston, K.M. Some preliminary data on the nature/structure of the PC-glycan of the major excretory-secretory product of *Acanthocheilonema viteae* (ES-62). *Parasite* **1**, 179-181 (1994).
150. Melendez, A.J., *et al.* Inhibition of Fc epsilon RI-mediated mast cell responses by ES-62, a product of parasitic filarial nematodes. *Nature medicine* **13**, 1375-1381 (2007).
151. Schnoeller, C., *et al.* A helminth immunomodulator reduces allergic and inflammatory responses by induction of IL-10-producing macrophages. *J Immunol* **180**, 4265-4272 (2008).
152. Danilowicz-Luebert, E., *et al.* A nematode immunomodulator suppresses grass pollen-specific allergic responses by controlling excessive Th2 inflammation. *International journal for parasitology* (2012).
153. Ruyssers, N.E., *et al.* Therapeutic potential of helminth soluble proteins in TNBS-induced colitis in mice. *Inflammatory bowel diseases* **15**, 491-500 (2009).
154. Zhu, B., *et al.* Immune modulation by Lacto-N-fucopentaose III in experimental autoimmune encephalomyelitis. *Clin Immunol* **142**, 351-361 (2012).
155. Atochina, O. & Harn, D. Prevention of psoriasis-like lesions development in fsn/fsn mice by helminth glycans. *Experimental dermatology* **15**, 461-468 (2006).
156. Van Kruiningen, H.J. & West, A.B. Potential danger in the medical use of *Trichuris suis* for the treatment of inflammatory bowel disease. *Inflammatory bowel diseases* **11**, 515 (2005).
157. Zou, W. Regulatory T cells, tumour immunity and immunotherapy. *Nature reviews. Immunology* **6**, 295-307 (2006).
158. Janssens, S. & Beyaert, R. Role of Toll-like receptors in pathogen recognition. *Clinical microbiology reviews* **16**, 637-646 (2003).
159. Hotamisligil, G.S., Shargill, N.S. & Spiegelman, B.M. Adipose expression of tumor necrosis factor-alpha: direct role in obesity-linked insulin resistance. *Science* **259**, 87-91 (1993).
160. Law, J.P., *et al.* The importance of Foxp3 antibody and fixation/permeabilization buffer combinations in identifying CD4+CD25+Foxp3+ regulatory T cells. *Cytometry. Part A : the journal of the International Society for Analytical Cytology* **75**, 1040-1050 (2009).
161. Hotamisligil, G.S. Inflammation and metabolic disorders. *Nature* **444**, 860-867 (2006).
162. Stahl, B., *et al.* Oligosaccharides from human milk as revealed by matrix-assisted laser desorption/ionization mass spectrometry. *Analytical biochemistry* **223**, 218-226 (1994).

163. Atochina, O., Da'dara, A.A., Walker, M. & Harn, D.A. The immunomodulatory glycan LNFPIII initiates alternative activation of murine macrophages in vivo. *Immunology* **125**, 111-121 (2008).
164. Bhargava, P., et al. Immunomodulatory glycan LNFPIII alleviates hepatosteatosis and insulin resistance through direct and indirect control of metabolic pathways. *Nature medicine* **18**, 1665-1672 (2012).
165. Ricardo-Gonzalez, R.R., et al. IL-4/STAT6 immune axis regulates peripheral nutrient metabolism and insulin sensitivity. *Proceedings of the National Academy of Sciences of the United States of America* **107**, 22617-22622 (2010).

