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Anti sense and sensibility : renal and skin effects of (antisense) oligonucleotides

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Citation

Meer, L. van. (2017, January 19). *Anti sense and sensibility : renal and skin effects of (antisense) oligonucleotides*. Retrieved from <https://hdl.handle.net/1887/45389>

Version: Not Applicable (or Unknown)

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Note: To cite this publication please use the final published version (if applicable).

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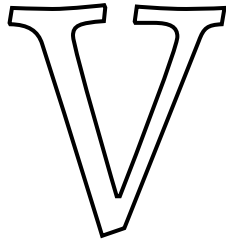


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Title: Anti sense and sensibility : renal and skin effects of (antisense) oligonucleotides

Issue Date: 2017-01-19



URINARY KIDNEY
BIOMARKERS FOR
EARLY DETECTION
OF NEPHROTOXICITY
IN CLINICAL DRUG
DEVELOPMENT

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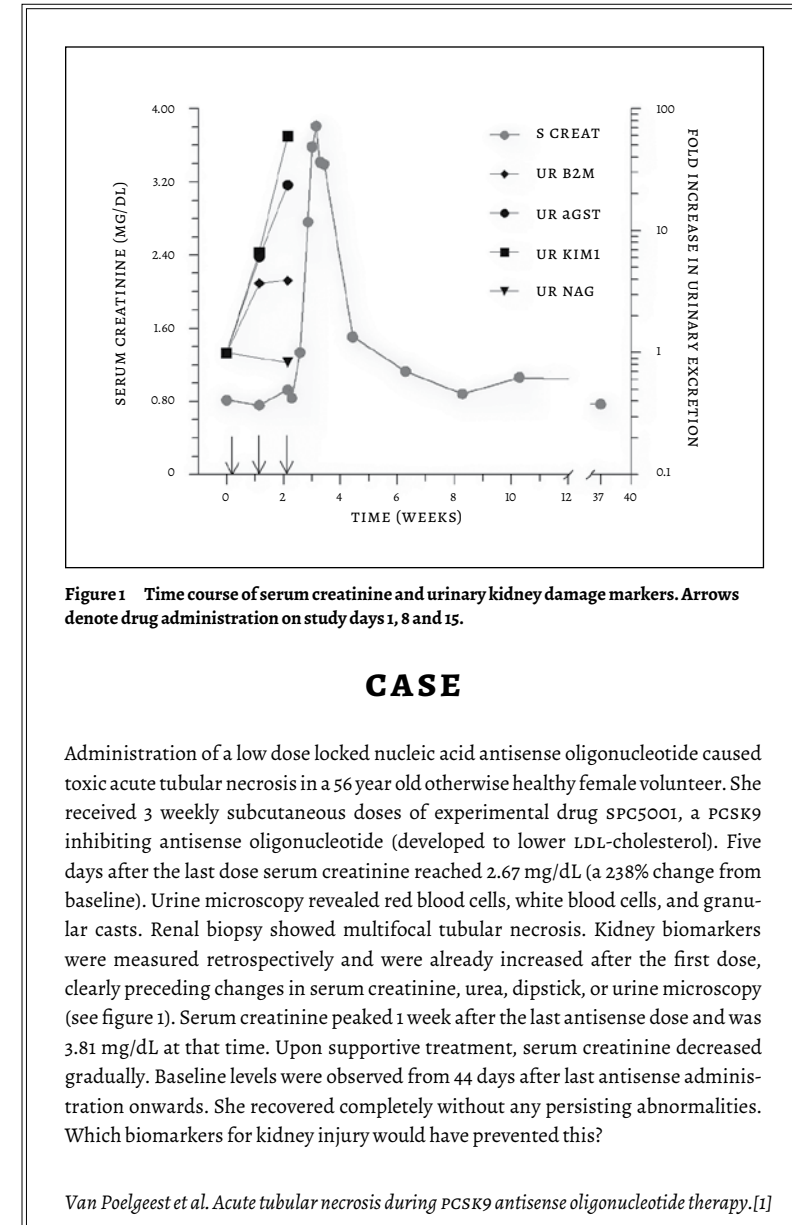


Abstract

Early detection of drug-induced kidney injury is vital in drug development. Generally accepted biomarkers such as creatinine and BUN lack sensitivity and early injury responses are missed. Many new biomarkers to detect nephrotoxicity for pre-clinical utilization have been described and their use is adopted in regulatory guidelines. However, guidance on appropriate biomarkers for clinical trials is minimal. We provide an overview of potentially useful kidney biomarkers that can be used in clinical trials. This includes guidance to select biomarkers suitable to capture specific characteristics of the (expected) kidney injury. We conclude that measurement of urinary Kidney Injury Marker-1 (KIM1) serves many purposes and is often an appropriate choice. Cystatin C captures effects on glomerular filtration rate, but this marker should preferably be combined with more specific markers to localize the origin of the observed effect. Untoward effects on tubules can be captured relatively well with several markers. Direct detection of glomerular injury is currently impossible since specific biomarkers are lacking. Indirect assessment of toxic effects on glomeruli is possible by using carefully selected panels of other injury markers. We conclude that it is possible to obtain appropriate information on nephrotoxicity in clinical drug development by using carefully selected panels of injury markers and suggest that identification and validation of specific glomerular biomarkers could be of great value.

Introduction

As illustrated by the case, described on the right hand side, early detection of drug-induced nephrotoxicity and prevention of clinical manifestations such as tubular necrosis are vital in early drug development in humans. The problem of drug-related renal toxicity is important as renal drug toxicities in animal studies account for more than 30 percent of the attrition of compounds from drug development [2]. Despite this pre-selection, prevalence of (acute) kidney injury due to drug toxicity in clinical practice is as high as 18 - 27% of all episodes of acute kidney injury [3]. In addition, efficacious interventions to reverse kidney damage are non-existing and clinicians can only apply supportive therapies while awaiting recovery of renal function. Animal models have shown that intervention



directly after early changes have been noted is preferable as the window of opportunity to apply treatment appears to be limited to a few hours [4]. Unfortunately, signals of early injury responses and the underlying mechanism of damage are often missed by traditional methods for monitoring renal function, such as serum creatinine, serum BUN, estimated GFR and actual GFR. Serum creatinine and BUN are easy to obtain and the assays are part of standard care measurements rendering the results readily available. However, as markers for kidney injury, both have several shortcomings. Creatinine concentration in serum depends on multiple other factors than decline in renal function that differ intra- and inter-individually, such as age, gender, muscle mass, protein intake, and certain drugs [4;5]. Also, BUN concentrations are influenced variable factors, such as diet, dehydration, liver function and tissue breakdown [6]. This lack of specificity complicates the interpretation of these markers. The problems can be partially overcome by calculating Glomerular Filtration Rate (GFR). Based upon data of large groups of patients different equations have been derived to estimate the actual (eGFR) using the serum creatinine value [7]. The eGFR is commonly used in clinical practice as an overall index for kidney function [8]. The Cockcroft-Gault formula, described in 1976 is based on a cohort of 249 patients and takes into account the factors age, weight and gender [9]. The eGFR Modification of Diet in Renal Disease (MDRD) was derived from a study with renal disease patients and includes age, race, gender, serum BUN and serum albumin as input parameters [10]. In addition, the Chronic Kidney Disease Epidemiology Collaboration (CKD-EPI) was derived from two study cohorts, one consisting of patients with known renal disease (mGFR < 90 mL/min), and a second cohort made up of kidney transplant donors all of whom had an mGFR > 90 mL/min [11]. The CKD-EPI formula includes the factors gender, age and race. Thus all formulas correct for certain variables and offer a tool to estimate GFR and monitor kidney function in chronic kidney disease patients. However, they still lack accuracy [12;13] and miss early and relatively small changes in GFR [14]. This is particularly true when monitoring healthy subjects with normal renal function and normal baseline serum creatinine values. In this situation very small changes in plasma creatinine, which are in the noise of the assay, may still reflect considerable loss of renal function, as illustrated in figure 2. Another disadvantage of serum creatinine (and eGFRs derived from this marker) and BUN is that a considerable time delay exist between the onset of kidney injury and the moment that the

signal is observed. For creatinine this is for example illustrated in figure 1. BUN levels correlate closely with histopathological changes in the kidney [15], however, the signal is delayed; by the time the damage is revealed by this marker 70-80% of renal epithelial mass is already lost [16].

It has been advocated to not estimate but measure GFR for a more reliable assessment of kidney function. This can be achieved by measuring clearance after intravenous administration of exogenous markers that are solely removed by glomerular filtration. This approach also avoids GFR estimates influenced by extra-glomerular clearance such as tubular secretion. Different markers are used among which inulin [17] and radio isotopic agents such as ^{99m}Tc-diethylenetriaminepentaacetic acid (DTPA), ¹⁶⁹Yb-DTPA, ¹²⁵I-iothalamate and ⁵¹Cr-ethylenediaminetetraacetic acid (⁵¹Cr-EDTA). These markers have a high accuracy and can detect subtle changes in GFR [17;18]. These methods are time consuming and therefore the results may not be readily available [19;20]. More importantly, however, GFR is only one measure of kidney function while many different mechanism of drug induced kidney injury exist, such as reduction in renal perfusion, direct tubular toxicity, intratubular obstruction, allergic interstitial nephritis and hemolytic-uremic syndrome [21]. All these mechanisms will lead directly or indirectly to reduction in the GFR, for instance by tubuloglomerular feedback after tubular toxicity [22]. However, these changes in GFR may be detected too late for adaptations in dose or schedule of dosing, and do not provide information on the mechanism of toxicity. In this review we focus on measures that may provide earlier and more specific warning signals of renal damage than a decreased GFR.

In preclinical research detection of early signs of kidney injury is done by histological examination, which is considered to be the gold standard. Histopathology provides accurate anatomical information on the kidney injury as well as its severity and leads to a diagnosis. However, performing a kidney biopsy in humans is rarely an option and is an inappropriate tool to be used in drug development trials.

These difficulties regarding early detection of kidney injury may substantially influence the path of drug development. For instance, development of compounds for which renal toxicity is observed during preclinical experiments may be ceased in order to avoid safety issues during the clinical development. However, it is known that only 40-60% of animal findings are predictive of toxicities in humans [23;24]. This results



in the undesirable situation in which potentially efficacious compounds are abandoned for the wrong reason. Moreover, as these compounds do not reach the clinical phase, no insights are gained regarding the (potentially avoidable) nephrotoxic mechanisms of injury. The development of immortalized human proximal tubule cell lines expressing functional influx and efflux transporters [25], which can be challenged with toxins [26] appears to be a promising approach that can be used preclinically. However, it is not yet confirmed that these cell lines and the subsequent incubations sufficiently mimic the *in vivo* situation. Therefore translating findings from these cell-lines directly to the clinical situation seems premature. Adequate use of renal biomarkers could thus be of great value during early drug development.

For preclinical assessment, many new potential biomarkers of toxicity have been identified and their possible benefit has been evaluated by comparing their performance to the traditional markers. This extensive research has resulted in an EMEA/FDA guideline (published in 2009), that identified acceptable biomarkers that can be used to detect drug-induced nephrotoxicity in preclinical research [27]. This panel includes the urinary excreted biomarkers Kidney Injury Molecule-1 (KIM1), albumin, total protein, Beta-2-microglobulin (B2M), clusterin, Trefoil Factor 3 (TFF3), and Cystatin C (CysC). These markers can be used to capture acute drug-induced nephrotoxicity of tubular or glomerular (with associated tubular involvement) origin [27]. These markers were shown to provide additional and complementary information to BUN and serum creatinine and correlate with histopathological alterations. However, it was recognized that for the clinical setting these markers were insufficiently qualified to justify their general use. It was suggested to further explore their potential as clinical biomarkers for acute drug-induced kidney injury and recovery/reversibility. The guideline also advised to consider the biomarkers in clinical trials on a case-by-case basis to gather further data on their usefulness to monitor drug-induced renal toxicity in man.

With this review, we aim to offer guidance to select the biomarkers that suit the specific characteristics of the (expected) kidney injury. We provide an overview of promising biomarkers for nephrotoxicity, focusing on their possible use and limitations in clinical trials during early drug development. Certain properties of biomarkers of kidney injury are considered to be critical. First, the origin of the biomarker and the mechanism and/or site of injury should be clarified as much as possible.

Furthermore the biomarker should be sensitive to early injury in order to outperform the traditional biomarkers and enable early intervention. High specificity and correlation with established outcome measures such as histopathology are also vital to avoid confusion on the value of the observed signal. As the majority of data on kidney injury biomarkers originates from preclinical research, the translational step to humans is of great importance. This could be achieved if supportive clinical evidence on biomarker profiles reflecting kidney injury in humans is available. Finally, reliable assays must be readily available.

The biomarkers were selected using the previously mentioned EMEA/FDA guidelines on preclinical markers, and on the condition that the pertaining assay is validated for human use and commercially available. The selection was expanded with the biomarkers for which promising data have been reported. These markers are Neutrophil Gelatinase-associated Lipocalin (NGAL), alpha Glutathione S-transferase (aGST), N-acetyl-beta-glucosaminidase (NAG) and Interleukin-18 (IL18). Established markers such as serum creatinine, serum BUN, urinary albumin and protein were not considered in detail, although these are used as reference. Based on the literature, several considerations and recommendations regarding selection of kidney biomarkers in clinical drug development are given.

Summary on selected markers

Cystatin C (CysC) is a small molecule cysteine proteinase inhibitor synthesized by all nucleated cells and filtered freely by the glomerulus. After filtration it is not secreted nor reabsorbed by the tubules, but catabolized completely and thus reflects true GFR when measured in blood. Preclinically, CysC appears to be the most sensitive marker for early proximal tubular damage in animals, although a consistent dose response relation is lacking [28]. CysC is suitable to assess kidney function in general, regardless of specific lesion site, as this marker is devoid of extra-glomerular clearance, variability in production and limited sensitivity that apply for BUN and serum creatinine [29]. It has been suggested that CysC measured in blood could be a suitable translational biomarker as it avoids laborious urine collection in animals [29]. Possible disadvantages of CysC are its dependency on factors other than decline of renal function alone, such as age, gender, weight and height, smoking, and

high serum C-reactive protein levels [5]. In the clinic, CysC has been shown to be a sensitive marker of early renal dysfunction following ischemic injury [30].

Neutrophil Gelatinase-associated Lipocalin (NGAL) is an acute-phase protein secreted as a response to acute injury of proximal and distal tubular epithelial cells. It is freely filtered by the glomerulus after which rapid clearance occurs via receptor binding and endocytosis [4]. NGAL has been reported to be the most sensitive marker for proximal tubular damage in the preclinical setting [28]. After gentamicin exposure in rats a clear signal is detected as early as 24 hours after exposure. However, specificity to the location of injury must be questioned, since in a rodent glomerular damage model increased NGAL levels were also observed [31]. Clinical research has demonstrated that urinary NGAL is increased in several forms of chronic kidney injury [32;33] and in patients with urinary tract infections [34].

Interleukin-18 (IL18) is a proinflammatory cytokine produced by leukocytes and renal parenchymal cells such as tubular epithelial cells, podocytes and mesangial cells. It plays an important role in the exacerbation of acute tubular necrosis [35] and the inflammatory pathways involved are partly clarified [36]. The IL18 receptor (IL-18R) is expressed on these cells in cisplatin-induced acute kidney injury and urinary IL18 excretion has proven to be an early diagnostic marker for acute kidney injury in humans, particularly in critically ill patients [37]. However, at present it is unclear if IL18 reflects location-specific injury and therefore its potential for preclinical and clinical use in drug development is unclear. An obvious disadvantage is that increased IL18 levels can also be observed upon many forms of inflammation not limited to the kidney.

N-acetyl-beta-glucosaminidase (NAG) is a lysosomal enzyme which is contained abundantly in the renal tubular epithelia and involved in the degradation of mucopolysaccharides and glycoproteins. Its size precludes glomerular filtration and elevated urinary concentrations are considered to reflect tubular dysfunction. In preclinical research the sensitivity of NAG is higher compared to serum creatinine and comparable to BUN [38]. The NAG response profile is dependent on the toxin causing proximal tubule injury [39]. For example, gentamicin triggers an early response that last for 8 hours after dosing, whereas chromium triggers a response after 8 hours and mercury does not trigger a significant NAG increase at all [39]. Nevertheless, clinical evidence supports the usefulness

of NAG as an early marker of mild tubular injury [40] and demonstrates that it has predictive properties regarding the development of acute tubular necrosis [41].

Alpha Glutathione S-transferase (aGST) is a detoxification enzyme that is produced in numerous tissues. Urinary aGST levels are very low under physiological conditions, but substantial amounts are excreted in case of various manifestations of tubular injury, including cisplatin- [42] and gentamicin-induced [43] toxicity and in acute tubular necrosis after mercuric chloride and potassium dichromate exposure [44]. aGST appears to be an adequate preclinical early toxicity biomarker to detect onset of epithelial necrosis, but is less suitable to monitor reversibility [29]. In drug-induced injury of proximal tubular cells with cisplatin and gentamicin, aGST correlated more closely to histopathological confirmed injury compared to NAG, BUN and serum creatinine [45]. In a preclinical model of tubular injury limited to the pars recta of proximal tubule cells, it was demonstrated that aGST-excretion reflects injury of low grade toxicity, outperforming numerous other markers, and with equal sensitivity as KIM1 [46]. However, injury to the collecting duct was associated with a decreased aGST, which is not well understood yet. As a consequence it has been suggested that qualification of this biomarker has to await further results [45]. Although limited information is available, clinical evidence suggests that aGST is informative for tubular dysfunction or injury [47;48].

Kidney Injury Molecule-1 (KIM1) is a transmembrane protein expressed by proximal tubular epithelial cells. KIM1 functions as a phosphatidylserine receptor and has phagocytic capacity [28]. Expression is markedly upregulated in response to injury [49]. Urinary KIM1 concentration provides a more sensitive predictor of histopathological confirmed injury in 11 well-established rat models of acute kidney injury when compared to BUN, serum creatinine or urinary NAG, even in cases of low grade toxicity [38]. One study reported that urinary KIM1 levels also correlate with different grades of kidney tubular histopathologies. This was supported by the finding of dose-dependent upregulation of the KIM1 gene in segment-specific toxicity models [50]. Whereas aGST appears to be a good early toxicity biomarker for epithelial necrosis, KIM1 and clusterin levels persist during regeneration and appear to reflect the triggering and continuation of the repair process [29;51]. KIM1 responses seem to depend on toxin, for example Sasaki et al. [52] report that after cisplatin exposure KIM1 increases (together with clusterin and aGST) after 3 days



(confirmed by Vinken et al.[15]), whereas in a model for papillary necrosis using 2-bromoethylamin hydrobromide KIM1 levels (together with clusterin, albumin and osteopontin) are convincingly increased as early as day 1 after exposure. Interestingly, measurement of urinary KIM1 also enables detection of subchronic and chronic injury and correlated closely with histopathology [28]. This study also showed that cysc and NGAL are the most sensitive markers for early kidney damage of proximal tubular damage, but subchronic or chronic injury was best reflected by KIM1 levels. In keeping with the notion that urinary KIM1 may be an early marker for chronic nephrotoxicity in animals, increased levels of urinary KIM1 levels were reported in an experiment using cadmium [53]. Clinical evidence that the results in animals translate to humans is currently limited. KIM1 did indeed show a significant signal following acute kidney injury, although sampling for KIM1 in this study was rather late, precluding assessment of its suitability as early marker [54].

Beta-2-microglobulin (B2M) is produced by all cells expressing major histocompatibility complex (MHC) class I antigen. Under normal conditions the main source are activated lymphocytes from which shedding from cell surface of the MHC occurs through proteolysis. Synthesis is stimulated in various conditions characterized by proliferation of lymphoid cells that occurs in various disease states, such as neoplasms, (auto-) immune disorders or infections [55-57]. B2M is filtered freely across the glomerulus and complete reabsorption occurs by proximal tubular cells [28]. Impaired tubular uptake results in increased B2M urinary excretion. Glomerular protein loss may also increase urinary B2M excretion as B2M shares a common rate-limited tubular reabsorption pathway with other proteins. In the preclinical setting B2M has a better diagnostic performance than BUN and serum creatinine to detect glomerular injury (together with urinary total protein and cysc) [58]. As tubular dysfunction without glomerular impairment also increases B2M [59;60] specificity regarding the location of damage using B2M alone is questionable. Interestingly, B2M might be excreted via other pathways as compared to other markers, as can be concluded from the findings on a model for papillary necrosis. This massive injury gave rise to impressive increases in KIM1, clusterin, albumin and osteopontin, but levels of B2M remained close to normal [52]. In the clinical setting, B2M has proven to be a marker for disease severity in autosomal polycystic kidney disease [61] and for renal damage by fumaric acid esters [62].

Clusterin is a glycosated protein associated with apoptosis and clearance of cellular debris and can be found in several tissues. Within kidney cells, clusterin has been suggested to possess anti-apoptotic properties and facilitates cell protection, lipid recycling and cell attachment, and aggregation [63]. Clusterin cannot be filtered by glomeruli due to its size and therefore urinary levels are specific for kidney injury. Clusterin performed better to detect proximal tubular injury than cysc, B2M and total protein [58] and evidence suggests it can be used as an early marker with a profile similar to KIM1 [15;52]. The clusterin response correlates well with tubular injury regardless of the location, particularly when regeneration is present [45]. Elevated clusterin levels persist during regeneration and appear to reflect the triggering and continuation of the repair process [29]. Clinical data on urinary clusterin in relation to kidney injury is not extensively available, however, it has been shown that clusterin expression is increased in renal injury and cystic diseases [64].

Trefoil factor 3 (TFF3) is a small peptide hormone secreted by mucus-producing and other epithelial cells [65]. In the kidney it is produced/secreted by cells of the collecting ducts [66]. TFF3 is involved in many functions including restoration of intestinal epithelium [67], but its physiological function within the kidney is still elusive. Because in ageing rats decreasing amount of kidney TFF3 are found, it has been suggested that TFF3 may have a general protective function [68]. Significantly decreased levels of TFF3 have been observed in different rat models of proximal tubular injury. Combining TFF3 with urinary albumin increases sensitivity to early injury compared to traditional markers [69]. However, studies comparing TFF3 to other novel biomarkers are lacking. In humans, it has been reported that certain populations (African descent, diabetes and antihypertensive medication use) have higher baseline urinary TFF3 levels, and that increase in urinary levels might indicate ongoing repair of chronic damage in the kidney [70].

Comparison & Considerations

Concerns regarding nephrotoxicity are often encountered during the development of novel drugs. Particularly when the compound is about to be tested in humans for the first time, all indications of possible kidney injury are weighed. This may regard compounds belonging to drug



classes that are notoriously associated with renal injury, drugs specifically targeted to the kidney, but also drugs that are considered 'suspicious' because of their mechanism of action or pharmacokinetic properties. Preclinical suggestions for nephrotoxicity can be present. In case of clearly dose-related adverse renal effects at the high end of the tested dose range, it is usually possible to estimate a safe range for dosing in humans provided that adequate monitoring is possible. However, it is more difficult if preclinical data point towards an incidentally occurring event. This might lead to a more variable risk for susceptible human subjects, which is difficult to catch. Whatever the findings in animals, close monitoring of the kidney in clinical trials to detect untoward effects as soon as possible is necessary in all cases.

Every drug has its specific features and other factors such as dosing schedule, cumulative dose, patient- and/or disease characteristics may play a role. Thus, each drug requires the selection of an appropriate biomarker or most likely a panel of biomarkers to provide comprehensive information. As subject safety is a primary goal in first into human trials, sensitivity for the event is crucial. A strategy for selecting biomarkers in humans could be to first focus on the localization of the site of injury and the mechanism by which the compound causes injury. This may be achieved by using histopathological information on the compound and comparing this to histopathological and biomarker profiles of known nephrotoxic agents that target similar sites (table 1).

The selection of a panel of biomarkers should obviously be based on the possibilities and limitations of individual markers and the aim should be to compose a panel of which the combination of biomarkers provides complimentary information (table 2).

We first point out that currently a specific marker for glomerular injury is unavailable and thus glomerular damage is frequently made by excluding other causes. Identification of a biomarker specific for glomerular injury would be of great value. cysc can be used to monitor general kidney function as it reflects GFR. Further, CYCC and NGAL are the most sensitive markers for early kidney damage of proximal tubular damage [28]. However, it should be realized that the organ-specificity of cysc is poor and other biomarkers are recommended to differentiate between tubular and glomerular damage. The dependency of cysc levels on other factors than renal function is important in diagnostic work-ups, but not so much in clinical drug research as these circumstances can be taken

into account by choice of population and by focusing on the time course and relative changes from baseline. If this is unfeasible or undesired, cysc appears to be a less suitable biomarker.

Although NGAL seems more tubular injury-specific compared to cysc, this has been challenged by the finding that NGAL also increases in a model of glomerular injury [31]. This could reflect generalized (glomerular and tubular) injury or that the biomarker is not injury-site specific. NGAL is not recommended as a monitoring tool, as it appears to possess equal sensitivity as serum BUN. agST and KIM1 exhibit similar sensitivity to detect tubular damage to the pars recta of the proximal tubule [46]. Nevertheless, KIM1 is preferred as there is more evidence regarding its performance compared to other biomarkers [38]. Moreover, the close correlation of KIM1 to histopathology [38;50] and the finding that it can be used to also monitor reversibility [29] suggest KIM1 to be the biomarker of choice. Furthermore, KIM1 can be used to detect subchronic or chronic injury [28;71]. The limited information available on TFF3 suggests that this biomarker does not have a clear advantage over other biomarkers for proximal tubular injury. We suggest to not use this marker unless more information is available on its profile and its performance compared to other biomarkers in animals and humans.

B2M is generally considered to be a glomerular marker, but increased urinary excretion could also reflect impaired reabsorption by proximal tubular cells. It appears that B2M as single measure does not have great value [72]. However, when included in a panel of biomarkers it allows distinguishing between glomerular and tubular dysfunction. The choice to incorporate clusterin depends largely on the expected toxicity. Clusterin is a sensitive marker and its urinary excretion increases as a response to tubular injury irrespectively of the site. This feature of non-specificity can be of value, when tubular injury to distal tubules is suspected or localization is unclear. It is advised to use clusterin only in a panel of biomarkers. The information on IL18 as suitable marker is too limited to justify its use in early clinical drug development. However, the finding that IL18 can be used to detect acute kidney injury very early in hospitalized patients [37] and its role in the pathophysiology of kidney injury [36] suggests that IL18 is potentially a useful marker but this should be explored further.

It is important to take into account that biomarkers might increase for other reasons than kidney injury (alone), which potentially results in



an inaccurate conclusion. For instance, increased levels of acute-phase proteins such as NGAL, but also B2M and IL18 may be observed upon any inflammatory condition regardless of the site of inflammation. Also the population that is studied is important as illustrated by the finding that patients with an impaired glucose tolerance show a higher urinary excretion of NAG [73]. These problems may be minimized by assessment of (changes in) renal function while taking all available information into account. In practice, it is advised to carefully consider the population that is studied, to have good information on the biomarkers at baseline, and to always measure chemistry and hematology at similar time points as the panel of renal biomarkers. This comprehensive approach prevents erroneous attribution of findings to the kidney.

Recommendation

We conclude that measurement of urinary KIM1 suits many purposes and is therefore often an appropriate choice. It allows for early detection of proximal tubule injury, differentiation between glomerular and tubular damage and assessment of reversibility and regeneration provided this occurs. KIM1 can also be used to detect sub-chronic and chronic kidney injury. NGAL is useful if tubular involvement is suspected, but no clear notion of tubular localization is present. Clusterin may add value in case of suspected injury in the distal tubule. If general kidney function and thus a measure of clearance is required, it is useful to include cysc, preferably combined with more specific markers to localize the origin of the observed effect. IL18 is considered less suitable for early phase trials, but might be helpful in phase II/III trials to be able to detect ATN in a very early stage. It may be useful to include B2M and α ST in the panel of biomarkers as this enables to relate findings on new compounds to existing knowledge of known nephrotoxic agents.

REFERENCES

- van Poelgeest EP, Swart RM, Betjes MG, Moerland M, Weening JJ, Tessier Y, Hodges MR, Levin AA, Burggraaf J: Acute Kidney Injury During Therapy With an Antisense Oligonucleotide Directed Against PCSK9. *Am J Kidney Dis* 2013;62:796-800.
- Kola I, Landis J: Can the pharmaceutical industry reduce attrition rates? *Nat Rev Drug Discov* 2004;3:711-715.
- Loghman-Adham M, Kiu Weber CI, Giocriaro C, Mann J, Meier M: Detection and management of nephrotoxicity during drug development. *Expert Opin Drug Saf* 2012;11:581-596.
- McIlroy DR, Wagener G, Lee HT: Biomarkers of acute kidney injury: an evolving domain. *Anesthesiology* 2010;112:998-1004.
- Knight EL, Verhave JC, Spiegelman D, Hillege HL, de ZD, Curhan GC, de Jong PE: Factors influencing serum cystatin C levels other than renal function and the impact on renal function measurement. *Kidney Int* 2004;65:1416-1421.
- Traynor J, Mactier R, Geddes CC, Fox JG: How to measure renal function in clinical practice. *BMJ* 2006;333:733-737.
- Herget-Rosenthal S, Bokenkamp A, Hofmann W: How to estimate GFR-serum creatinine, serum cystatin C or equations? *Clin Biochem* 2007;40:153-161.
- Stevens PE, Levin A: Evaluation and management of chronic kidney disease: synopsis of the kidney disease: improving global outcomes 2012 clinical practice guideline. *Ann Intern Med* 2013;158:825-830.
- Cockcroft DW, Gault MH: Prediction of creatinine clearance from serum creatinine. *Nephron* 1976;16:31-41.
- Levey AS, Bosch JP, Lewis JB, Greene T, Rogers N, Roth D: A more accurate method to estimate glomerular filtration rate from serum creatinine: a new prediction equation. Modification of Diet in Renal Disease Study Group. *Ann Intern Med* 1999;130:461-470.
- Levey AS, Stevens LA, Schmid CH, Zhang YL, Castro AF, III, Feldman HI, Kusek JW, Eggers P, Van LF, Greene T, Coresh J: A new equation to estimate glomerular filtration rate. *Ann Intern Med* 2009;150:604-612.
- Botev R, Mallie JP, Wetzels JF, Couchoud C, Schuck O: The clinician and estimation of glomerular filtration rate by creatinine-based formulas: current limitations and quo vadis. *Clin J Am Soc Nephrol* 2011;6:937-950.
- Stevens LA, Coresh J, Greene T, Levey AS: Assessing kidney function—measured and estimated glomerular filtration rate. *N Engl J Med* 2006;354:2473-2483.
- Murty MS, Sharma UK, Pandey VB, Kankare SB: Serum cystatin C as a marker of renal function in detection of early acute kidney injury. *Indian J Nephrol* 2013;23:180-183.
- Vinken P, Starckx S, Barale-Thomas E, Loozova A, Sonee M, Goeminne N, Vermissen L, Buyens K, Lampo A: Tissue Kim-1 and Urinary Clusterin as Early Indicators of Cisplatin-Induced Acute Kidney Injury in Rats. *Toxicol Pathol* 2012.
- Sieber M, Hoffmann D, Adler M, Vaidya VS, Clement M, Bonventre JV, Zidek N, Rached E, Amberg A, Callanan JJ, Dekant W, Mally A: Comparative analysis of novel noninvasive renal biomarkers and metabonomic changes in a rat model of gentamicin nephrotoxicity. *Toxicol Sci* 2009;109:336-349.
- Rahn KH, Heidenreich S, Bruckner D: How to assess glomerular function and damage in humans. *J Hypertens* 1999;17:309-317.
- Soares AA, Eyff TF, Campani RB, Ritter L, Camargo JL, Silveiro SP: Glomerular filtration rate measurement and prediction equations. *Clin Chem Lab Med* 2009;47:1023-1032.
- Prigent A: [Measurement of renal function in clinical practice: principles and limitations]. *J Radiol* 2011;92:274-279.
- Stevens LA, Levey AS: Measured GFR as a confirmatory test for estimated GFR. *J Am Soc Nephrol* 2009;20:2305-2313.
- Thadhani R, Pascual M, Bonventre JV: Acute renal failure. *N Engl J Med* 1996;334:1448-1460.
- Blantz RC, Thomson SC, Peterson OW, Gabbai FB: Physiologic adaptations of the tubuloglomerular feedback system. *Kidney Int* 1990;38:577-583.
- Olson H, Betton G, Stritar J, Robinson D: The predictivity of the toxicity of pharmaceuticals in humans from animal data—an interim assessment. *Toxicol Lett* 1998;102-103:535-538.
- Knight A: Systematic reviews of animal experiments demonstrate poor human clinical and toxicological utility. *Altern Lab Anim* 2007;35:641-659.
- Wilmer MJ, Saleem MA, Masereeuw R, Ni L, van der Velden TJ, Russel FG, Mathieson PW,



- Monnens LA, van den Heuvel LP, Levchenko EN: Novel conditionally immortalized human proximal tubule cell line expressing functional influx and efflux transporters. *Cell Tissue Res* 2010;339:449-457.
- 26 Moghadasali R, Mutsaers HA, Azarnia M, Aghdami N, Baharvand H, Torensma R, Wilmer MJ, Masereeuw R: Mesenchymal stem cell-conditioned medium accelerates regeneration of human renal proximal tubule epithelial cells after gentamicin toxicity. *Exp Toxicol Pathol* 2012.
- 27 EMEA/FDA guideline on detection of drug-induced nephrotoxicity: 2009.
- 28 Hoffmann D, Fuchs TC, Henzler T, Matheis KA, Herget T, Dekant W, Hewitt P, Mally A: Evaluation of a urinary kidney biomarker panel in rat models of acute and subchronic nephrotoxicity. *Toxicology* 2010;277:49-58.
- 29 Ozer JS, Dieterle F, Troth S, Perentes E, Cordier A, Verdes P, Staedtler F, Mahl A, Grenet O, Roth DR, Wahl D, Legay F, Holder D, Erdos Z, Vlasakova K, Jin H, Yu Y, Muniappa N, Forest T, Clouse HK, Reynolds S, Bailey WJ, Thudium DT, Topper MJ, Skopek TR, Sina JF, Glaab WE, Vonderscher J, Maurer G, Chibout SD, Sistare FD, Gerhold DL: A panel of urinary biomarkers to monitor reversibility of renal injury and a serum marker with improved potential to assess renal function. *Nat Biotechnol* 2010;28:486-494.
- 30 Vassalos A, Young D, MacArthur K, Pollock J, Lyall F, Danton MH: Cystatin C: influence of perfusion and myocardial injury on early (<24 h) renal function after pediatric cardiac surgery. *Paediatr Anaesth* 2011;21:1185-1191.
- 31 Tomomura Y, Tsuchiya N, Torii M, Uehara T: Evaluation of the usefulness of urinary biomarkers for nephrotoxicity in rats. *Toxicology* 2010;273:53-59.
- 32 Ding H, He Y, Li K, Yang J, Li X, Lu R, Gao W: Urinary neutrophil gelatinase-associated lipocalin (NGAL) is an early biomarker for renal tubulointerstitial injury in IgA nephropathy. *Clin Immunol* 2007;123:227-234.
- 33 Brunner HI, Mueller M, Rutherford C, Passo MH, Witte D, Grom A, Mishra J, Devarajan P: Urinary neutrophil gelatinase-associated lipocalin as a biomarker of nephritis in childhood-onset systemic lupus erythematosus. *Arthritis Rheum* 2006;54:2577-2584.
- 34 Yilmaz A, Sevketoglu E, Gedikbasi A, Karyagar S, Kiyak A, Mulazimoglu M, Aydogan G, Ozpacaci T, Hatipoglu S: Early prediction of urinary tract infection with urinary neutrophil gelatinase associated lipocalin. *Pediatr Nephrol* 2009;24:2387-2392.
- 35 Bani-Hani AH, Leslie JA, Asanuma H, Dinarello CA, Campbell MT, Meldrum DR, Zhang H, Hile K, Meldrum KK: IL-18 neutralization ameliorates obstruction-induced epithelial-mesenchymal transition and renal fibrosis. *Kidney Int* 2009;76:500-511.
- 36 Nozaki Y, Kinoshita K, Yano T, Asato K, Shiga T, Hino S, Niki K, Nagare Y, Kishimoto K, Shimazu H, Funauchi M, Matsumura I: Signaling through the interleukin-18 receptor alpha attenuates inflammation in cisplatin-induced acute kidney injury. *Kidney Int* 2012;82:892-902.
- 37 Parikh CR, Jani A, Melnikov VY, Faubel S, Edelstein CL: Urinary interleukin-18 is a marker of human acute tubular necrosis. *Am J Kidney Dis* 2004;43:405-414.
- 38 Vaidya VS, Ozer JS, Dieterle F, Collings FB, Ramirez V, Troth S, Muniappa N, Thudium D, Gerhold D, Holder DJ, Bobadilla NA, Marrer E, Perentes E, Cordier A, Vonderscher J, Maurer G, Goering PL, Sistare FD, Bonventre JV: Kidney injury molecule-1 outperforms traditional biomarkers of kidney injury in preclinical biomarker qualification studies. *Nat Biotechnol* 2010;28:478-485.
- 39 Zhou Y, Vaidya VS, Brown RP, Zhang J, Rosenzweig BA, Thompson KL, Miller TJ, Bonventre JV, Goering PL: Comparison of kidney injury molecule-1 and other nephrotoxicity biomarkers in urine and kidney following acute exposure to gentamicin, mercury, and chromium. *Toxicol Sci* 2008;101:159-170.
- 40 Ouchi M, Suzuki T, Hashimoto M, Motoyama M, Ohara M, Suzuki K, Igari Y, Watanabe K, Nakano H, Oba K: Urinary N-acetyl-beta-d-Glucosaminidase Levels are Positively Correlated With 2-Hr Plasma Glucose Levels During Oral Glucose Tolerance Testing in Prediabetes. *J Clin Lab Anal* 2012;26:473-480.
- 41 Herget-Rosenthal S, Poppen D, Husing J, Marggraf G, Pietruck F, Jakob HG, Philipp T, Kribben A: Prognostic value of tubular proteinuria and enzymuria in nonoliguric acute tubular necrosis. *Clin Chem* 2004;50:552-558.
- 42 Feinfeld DA, Fuh VL, Safirstein R: Urinary glutathione-S-transferase in cisplatin nephrotoxicity in the rat. *J Clin Chem Clin Biochem* 1986;24:529-532.
- 43 Feinfeld DA, Fleischner GM, Arias IM: Urinary ligandin and glutathione-S-transferase in gentamicin-induced nephrotoxicity in the rat. *Clin Sci (Lond)* 1981;61:123-125.
- 44 Feinfeld DA, Bourgoignie JJ, Fleischner G, Goldstein EJ, Biempica L, Arias IM: Ligandinuria in nephrotoxic acute tubular necrosis. *Kidney Int* 1977;12:387-392.
- 45 Harpur E, Ennulat D, Hoffman D, Betton G, Gautier JC, Riefke B, Bounous D, Schuster K, Beushausen S, Guffroy M, Shaw M, Lock E, Pettit S: Biological qualification of biomarkers of chemical-induced renal toxicity in two strains of male rat. *Toxicol Sci* 2011;122:235-252.
- 46 Swain A, Turton J, Scudamore C, Maguire D, Pereira I, Freitas S, Smyth R, Munday M, Stamp C, Gandhi M, Sondh S, Ashall H, Francis I, Woodfine J, Bowles J, York M: Nephrotoxicity of hexachloro-1,3-butadiene in the male Hanover Wistar rat; correlation of minimal histopathological changes with biomarkers of renal injury. *J Appl Toxicol* 2012;32:417-428.
- 47 Bruning T, Sundberg AG, Birner G, Lammert M, Bolt HM, Appelkvist EL, Nilsson R, Dallner G: Glutathione transferase alpha as a marker for tubular damage after trichloroethylene exposure. *Arch Toxicol* 1999;73:246-254.
- 48 Sundberg A, Appelkvist EL, Dallner G, Nilsson R: Glutathione transferases in the urine: sensitive methods for detection of kidney damage induced by nephrotoxic agents in humans. *Environ Health Perspect* 1994;102 Suppl 3:293-296.
- 49 Amin RP, Vickers AE, Sistare F, Thompson KL, Roman RJ, Lawton M, Kramer J, Hamadeh HK, Collins J, Grissom S, Bennett L, Tucker CJ, Wild S, Kind C, Oreffo V, Davis JW, Curtiss S, Naciff JM, Cunningham M, Tennant R, Stevens J, Car B, Bertram TA, Afshari CA: Identification of putative gene based markers of renal toxicity. *Environ Health Perspect* 2004;112:465-479.
- 50 Chiusolo A, Defazio R, Zanetti E, Mongillo M, Mori N, Cristofori P, Trevisan A: Kidney injury molecule-1 expression in rat proximal tubule after treatment with segment-specific nephrotoxicants: a tool for early screening of potential kidney toxicity. *Toxicol Pathol* 2010;38:338-345.
- 51 Rouse RL, Zhang J, Stewart SR, Rosenzweig BA, Espandiari P, Sadrieh NK: Comparative profile of commercially available urinary biomarkers in preclinical drug-induced kidney injury and recovery in rats. *Kidney Int* 2011;79:1186-1197.
- 52 Sasaki D, Yamada A, Umeno H, Kurihara H, Nakatsujii S, Fujihira S, Tsubota K, Ono M, Moriguchi A, Watanabe K, Seki J: Comparison of the course of biomarker changes and kidney injury in a rat model of drug-induced acute kidney injury. *Biomarkers* 2011;16:553-566.
- 53 Prozialeck WC, Vaidya VS, Liu J, Waalkes MP, Edwards JR, Lamar PC, Bernard AM, Dumont X, Bonventre JV: Kidney injury molecule-1 is an early biomarker of cadmium nephrotoxicity. *Kidney Int* 2007;72:985-993.
- 54 Vaidya VS, Waikar SS, Ferguson MA, Collings FB, Sunderland K, Gioulas C, Bradwin G, Matsouaka R, Betensky RA, Curhan GC, Bonventre JV: Urinary biomarkers for sensitive and specific detection of acute kidney injury in humans. *Clin Transl Sci* 2008;1:200-208.
- 55 Tsoukas CM, Bernard NF: Markers predicting progression of human immunodeficiency virus-related disease. *Clin Microbiol Rev* 1994;7:14-28.
- 56 Bethea M, Forman DT: Beta 2-microglobulin: its significance and clinical usefulness. *Ann Clin Lab Sci* 1990;20:163-168.
- 57 Grundy JE: Role of beta 2-microglobulin in cytomegalovirus infection. *Scand J Rheumatol Suppl* 1990;87:98-101.
- 58 Dieterle F, Perentes E, Cordier A, Roth DR, Verdes P, Grenet O, Pantano S, Moulin P, Wahl D, Mahl A, End P, Staedtler F, Legay F, Carl K, Laurie D, Chibout SD, Vonderscher J, Maurer G: Urinary clusterin, cystatin C, beta2-microglobulin and total protein as markers to detect drug-induced kidney injury. *Nat Biotechnol* 2010;28:463-469.
- 59 Gatanaga H, Tachikawa N, Kikuchi Y, Teruya K, Genka I, Honda M, Tanuma J, Yazaki H, Ueda A, Kimura S, Oka S: Urinary beta2-microglobulin as a possible sensitive marker for renal injury caused by tenofovir disoproxil fumarate. *AIDS Res Hum Retroviruses* 2006;22:744-748.
- 60 Del PM, Romero S, Casado JL: Proximal tubular renal dysfunction or damage in HIV-infected patients. *AIDS Rev* 2012;14:179-187.
- 61 Meijer E, Boertien WE, Nauta FL, Bakker SJ, van OW, Rook M, van der Jagt EJ, van GH, Peters DJ, Navis G, de Jong PE, Gansevoort RT: Association of urinary biomarkers with



- disease severity in patients with autosomal dominant polycystic kidney disease: a cross-sectional analysis. *Am J Kidney Dis* 2010;56:883-895.
- 62 Haring N, Mahr HS, Mundle M, Strohal R, Lhotta K: Early detection of renal damage caused by fumaric acid ester therapy by determination of urinary beta2-microglobulin. *Br J Dermatol* 2011;164:648-651.
- 63 Rosenberg ME, Silksen J: Clusterin: physiologic and pathophysiologic considerations. *Int J Biochem Cell Biol* 1995;27:633-645.
- 64 Dvergsten J, Manivel JC, Correa-Rotter R, Rosenberg ME: Expression of clusterin in human renal diseases. *Kidney Int* 1994;45:828-835.
- 65 Madsen J, Nielsen O, Tornoe I, Thim L, Holmskov U: Tissue localization of human trefoil factors 1, 2, and 3. *J Histochem Cytochem* 2007;55:505-513.
- 66 Chinery R, Poulsom R, Elia G, Hanby AM, Wright NA: Expression and purification of a trefoil peptide motif in a beta-galactosidase fusion protein and its use to search for trefoil-binding sites. *Eur J Biochem* 1993;212:557-563.
- 67 Kinoshita K, Taupin DR, Itoh H, Podolsky DK: Distinct pathways of cell migration and antiapoptotic response to epithelial injury: structure-function analysis of human intestinal trefoil factor. *Mol Cell Biol* 2000;20:4680-4690.
- 68 Debata PR, Panda H, Supakar PC: Altered expression of trefoil factor 3 and cathepsin L gene in rat kidney during aging. *Biogerontol* 2007;8:25-30.
- 69 Yu Y, Jin H, Holder D, Ozer JS, Villarreal S, Shughrue P, Shi S, Figueroa DJ, Clouse H, Su M, Muniappa N, Troth SP, Bailey W, Seng J, Aslamkhan AG, Thudium D, Sistare FD, Gerhold DL: Urinary biomarkers trefoil factor 3 and albumin enable early detection of kidney tubular injury. *Nat Biotechnol* 2010;28:470-477.
- 70 Astor BC, Kottgen A, Hwang SJ, Bhavsar N, Fox CS, Coresh J: Trefoil factor 3 predicts incident chronic kidney disease: a case-control study nested within the Atherosclerosis Risk in Communities (ARIC) study. *Am J Nephrol* 2011;34:291-297.
- 71 Prozialeck WC, Edwards JR: Early biomarkers of cadmium exposure and nephrotoxicity. *Biometals* 2010;23:793-809.
- 72 Dieterle F, Perentes E, Cordier A, Roth DR, Verdes P, Grenet O, Pantano S, Moulin P, Wahl D, Mahl A, End P, Staedtler F, Legay F, Carl K, Laurie D, Chibout SD, Vonderscher J, Maurer G: Urinary clusterin, cystatin C, beta2-microglobulin and total protein as markers to detect drug-induced kidney injury. *Nat Biotechnol* 2010;28:463-469.
- 73 Fujita H, Narita T, Morii T, Shimotomai T, Yoshioka N, Kakei M, Ito S: Increased urinary excretion of N-acetylglucosaminidase in subjects with impaired glucose tolerance. *Ren Fail* 2002;24:69-75.

Figure 2 Theoretical relationship between serum creatinine and glomerular filtration rate (GFR) calculated with creatinine clearance for a subject with normal muscle mass (12 mmol creat/24 hours). In the range of creatinine for healthy subjects large declines in GFR are associated with relatively small increases in serum creatinine. For example, if serum creatinine increases from 60 to 90 $\mu\text{mol/L}$ (black arrow), which is a 50% increase (a cut off value for safety often used in clinical trials), GFR decreases with 50 ml/min (grey arrow), which reflects a considerable functional loss of over 35%. When the GFR decreases below 60 ml/min, further decrements are associated with larger increases in serum creatinine, which makes the marker more sensitive for change.

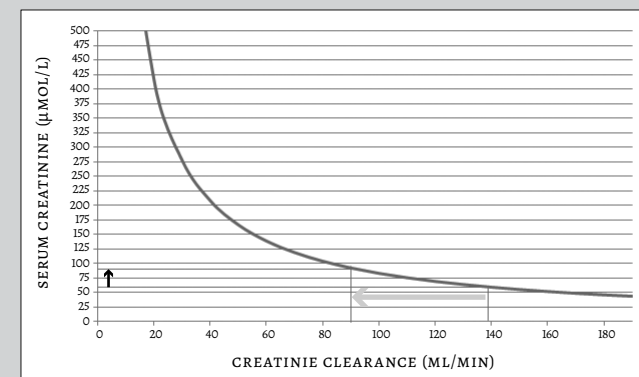


Table 1 Established nephrotoxic agents and accompanying biomarker signal

		Injury Models		Kidney Biomakers		
				Sensitive	Specific	
Glomerulus		Puromycin aminonucleoside (PAN) Doxorubicin		B2M, CysC		
Proximal Convoluted Tubule s1		Gentamicin	Cisplatin, Carbapenem A	TFF3, CysC, NGAL, (IL-18)	NAG	
Proximal Convoluted Tubule s2						
Proximal Straight Tubule s3		Vancomycin Hexachloro-1:3-butadiene (HCBD)		KIM-1, agST		
Distal Convoluted Tubule		Amphotericin B		clusterin, ngal	agst	
Collecting Duct						
Renal Papilla		2-bromoethylamin hydrobromide (BEA)		KIM-1, clusterin*		

* no biomarker identified to date covering this area

Table 2 Aims of research and appropriate biomarkers

Aim:	Suitable marker
Monitoring general kidney function (GFR)	CysC
Differentiate between glomerular and tubular damage	KIM1, clusterin, SGST
Monitor toxicity with suspected glomerular localization	B2M
Monitor toxicity with suspected non-glomerular localization	CysC, NGAL,
Detect early kidney injury with suspected proximal tubular localization	agST, KIM1
Monitor reversibility/regeneration with suspected proximal tubular localization	KIM1
Monitor toxicity with suspected distal tubular localization	Clusterin
Monitor reversibility/regeneration with suspected distal tubular localization	Clusterin
Elucidate pathophysiological mechanism with known proximal tubular injury site	NAG
High risk of ATN (Phase II/III trials)	IL-18