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Towards therapeutic disease control in inflammatory bowel diseases

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Chapter one

Introduction

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1 The immune system

The immune system protects the human body from disease. To accomplish this, many cell types are involved with different functions and tasks. When bacteria or viruses enter the body, the innate immune system first comes into play.¹ This system is non-specific, and provides an immediate inflammatory response which is characterized by redness, swelling, pain, heat and dysfunction. Once the innate immune system is activated, cells produce several cytokines and chemokines. In response to these secreted factors, other immune cells are recruited to the site of inflammation. The innate immune system then activates the adaptive immune system, consisting of highly specialized cells, B cells and T cells.² In the absence of antigen, T cells and B cells are naïve. They are activated by the innate immune system, when an antigen presenting cell (APC) presents an antigen to the B or T cell. Some of these activated B and T cells will become memory cells which provide long-term immune memory as these cells are able to respond to antigens without the help of the innate immune system. This will result in a more rapid and efficient response when the immune system encounters an antigen which has been recognized before.

1.1 The innate immune system

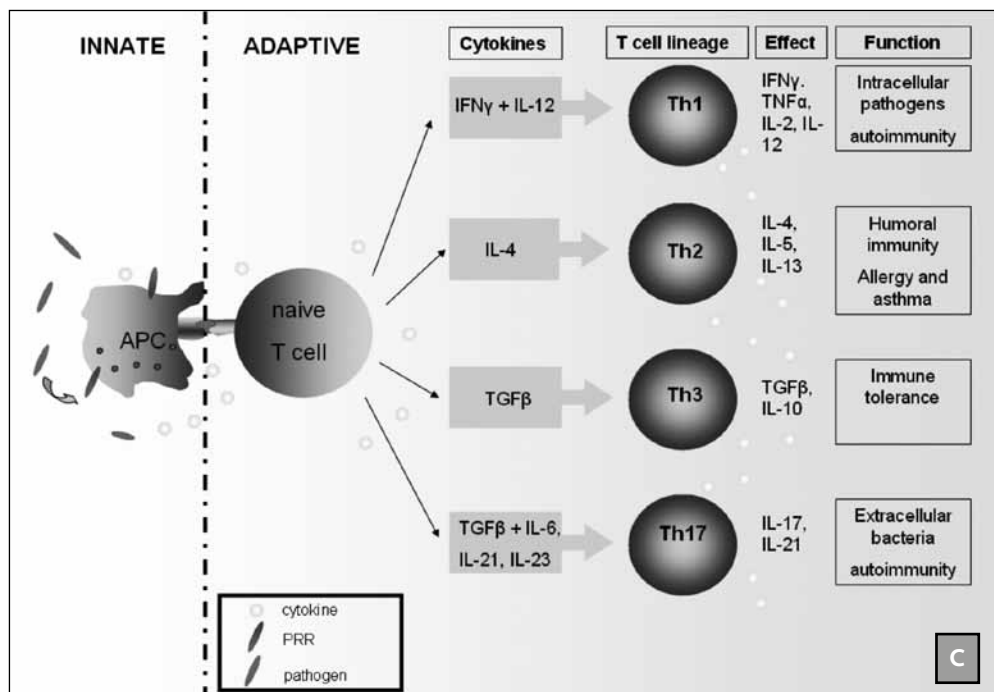
APCs are the key players of the innate immune system: they sense and process antigens, produce cytokines in response to pathogens, and activate the adaptive immune system. APCs express several pattern recognition receptor (PRR) molecules to sense pathogens in the environment. Two important types of PRRs are membrane-associated toll-like receptors (TLR) and cytosolic nucleotide-binding oligomerization domain (NOD proteins). The TLR family includes a family of 10 studied TLRs which all have different specificities for various pathogens. When lipopolysaccharide (LPS, a cell component of Gram-negative bacteria and responsible for septic shock) binds to TLR₄, the cell starts to produce several pro-inflammatory cytokines, among which tumor necrosis factor alpha (TNF α) and interleukin-1 β (IL-1 β).³ Next, other cells are recruited to the site of inflammation and an immune reaction is initiated. NOD₂ is located intracellular and recognizes molecules that contain muramyl dipeptide (MDP), which is present in several bacteria.⁴

Dendritic cells (DCs) are the most specialized APC and are usually one of the first cells that come into action once a bacterium or virus enters the body. When a DC senses a pathogen by ligation of a PRR, it phagocytoses (“eats”) the potential harmful foreigner, processes it inside the cell, and presents pieces of the protein on the cell membrane loaded on MHCII molecules.^{5,6} After ligation of PRRs, co-stimulatory molecules like CD80, CD83 and CD86 are upregulated. The antigen presented on the MHCII molecule is then recognized by the T cells receptor complex (TCR) on the T cell, and this provides the first signal to initiate an immune response.⁷ To achieve a full immune response, additional stimulation is often needed; the interaction of costimulatory molecules on DCs and T cells and the presence of various cytokines provide the second signal and finally determine the outcome of the immune response.^{8,9}

Like DCs, macrophages (M ϕ) are part of the innate immune system, derive from monocytes and function as APCs. They are present in many tissues and contribute to tissue homeostasis.^{10,11} M ϕ are a heterogeneous population of cells and have different functions depending on their differentiation status and the type of cytokines present in their environment.

¹²⁻¹⁴ Type 1 macrophages are typically induced in the presence of LPS or pro-inflammatory cytokines. In this setting, M ϕ are primed to become effector cells that are highly efficient in killing intracellular bacteria and in the production of pro-inflammatory cytokines. ¹⁵ In contrast to M ϕ 1, type 2 macrophages (M ϕ 2, regulatory macrophage or alternatively activated macrophage) have a more anti-inflammatory phenotype. They are induced by Th2 cytokines, glucocorticoids or immune complexes. M ϕ 2 have several characteristics that are functionally different from M ϕ 1: they are able to dampen immune responses by inhibiting T cell proliferation, production of anti-inflammatory cytokines and they contribute to wound healing. ¹⁶⁻¹⁸ In addition, M ϕ 2 inhibit Th1 responses by skewing the immune response towards a Th2 response.

Figure 1 Innate and adaptive immunity.



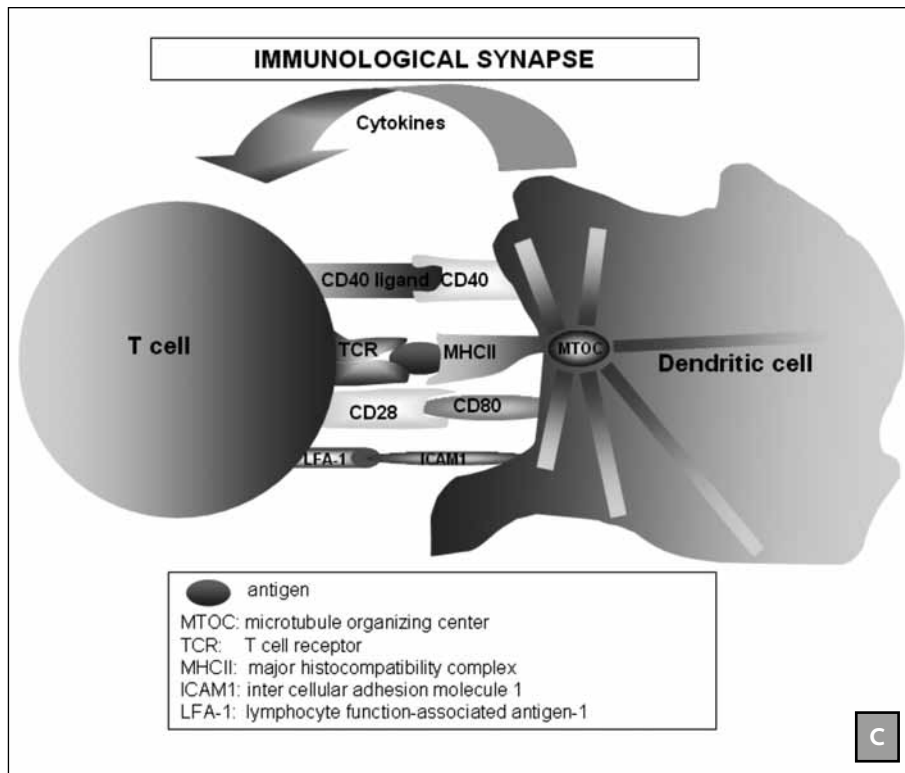
APC antigen presenting cell; IL interleukin; PRR pathogen recognition receptor

1.2 The adaptive immune system

The adaptive immune system consists of lymphocytes, i.e. B cells and T cells. B cells are involved in the production of Immunoglobulins (Ig, i.e. antibodies), whereas T cells differentiate into helper T cells that further support the ongoing immune response. After an APC has presented an antigen to the T cell, the T cell starts to proliferate, and differentiates into one of the known helper T cell lineages: Th1, Th2 or the more recently described Th3¹⁹ and Th17 (Figure 1).^{20, 21} The direction of differentiation is determined by the inflammatory environment and the presence of other factors, like cytokines. A Th1 response is characterized by the production of interferon- γ (IFN γ), TNF α and interleukin-2 (IL-2).

These cytokines stimulate T cell proliferation and activate macrophages. The activated macrophages produce cytokines that further promote Th₁ differentiation of T cells. During a Th₂ response, IL-4, IL-10 and IL-13 are typically produced, which activate B cells to produce immunoglobulins (Ig) and inhibit Th₁ responses. Cytokines produced by Th₂ effector cells further augment the differentiation signal towards a Th₂ response, thereby maintaining the Th₂ response. When the cells differentiate into the Th₃ lineage, the T cells start to produce large amounts of transforming growth factor beta (TGFβ), a factor which is known to be involved in tolerance and for the differentiation of regulatory T cells (Tregs).²² Tregs are cells with anti-inflammatory properties; they inhibit activation of the immune system and thereby maintain immune homeostasis.²³ A Th₁₇ response is characterized by the massive production of interleukin-17 by T cells, which is induced by IL-23, or IL-6 in combination with TGFβ.^{24, 25} IL-17 induces and promotes pro-inflammatory responses,²⁶ and triggers the production of pro-inflammatory cytokines like TNFα and IL-1β. Th₁, Th₂, Th₃ and Th₁₇ effector T cells all express CD4 on their membrane, making them CD4+ T cells. Another subset of T cells, CD8+ cells, plays an important role in the recognition and elimination of intracellular pathogens.²⁷ CD8+ T cells recognize antigens presented by MHC I molecules, and are able to kill infected cells by secreting perforin and enzymes.²⁸

Figure 2 The immunological synapse.



1.3 The immunological synapse

As mentioned before, cytokines present in the inflammatory environment and the interaction between the APC and the T cell together determine the outcome of the immune response. The immunological synapse (IS) is the initial site of interaction between the DC and the T cell.^{29, 30}

It is a highly organized structure, with the MHCII-TCR complex in the center, and the co-stimulatory molecules in the periphery (Figure 2). Importantly, a mature synapse is formed only upon recognition of a foreign antigen. When this is the case and an immunological synapse is formed, the actin skeleton polarizes towards the synapse.³¹ A proper formation of the IS is important for an efficient T cell response, as destabilization of the IS has been reported to result in decreased T cell signaling.³² As a consequence, formation of the IS plays a crucial role in the activation of T cells, and thereby the outcome of the immune response.

1.4 The immune system and tolerance in the gastrointestinal tract

The immune system in the gastrointestinal tract has several unique features. In the gut, an enormous amount of microorganisms (more than 500 different bacteria species)³³ is present that constitutes the gut flora (commensals), and intestinal APCs are constantly exposed to these microorganisms and food antigens. An immunologic response to the gut flora or other harmless antigens like food antigens would result in the recruitment of immune cells, production of cytokines, and consequently in inflammation, tissue damage and dysfunction. Therefore it is very important that intestinal APCs do not respond to these antigens, a process called tolerance.³⁴ The physical barrier separating the commensals from the underlying tissues consists of a single cell layer of epithelial cells and is the first line of defense. The presence of a mucus layer, antibacterial molecules (defensins) and IgA further helps to protect the invasion of antigens.³⁵ Since APC are then the first cells to respond to an antigen, M ϕ and DCs are the key players of innate immunity and tolerance.³⁶ To this end, intestinal M ϕ lack several innate response receptors, do not produce pro-inflammatory cytokines in response to various inflammatory signals,³⁷ and produce large amounts of IL-10 but no IL-12 and IL-23,³⁸ giving them an M ϕ 2-like appearance.³⁹ Loss of tolerance would lead to an immune reaction, and subsequently result in autoimmunity. In the absence of inflammation but in the presence of commensals, a close balance between effector T cells (i.e. Th1, Th2 or Th17 T cells) and regulatory T cells is maintained by a complex network of cytokines.

2 Inflammatory Bowel Diseases

2.1 Epidemiology, symptoms and diagnosis

Inflammatory bowel disease (IBD) refers to a chronic inflammation affecting the gastrointestinal tract. The highest incidence rates for both Crohn's disease (CD) and ulcerative colitis (UC) are reported in the western world.⁴⁰⁻⁴² However, the incidence in other parts of the world is increasing. In Europe and the US, incidence rates range from 1.5 to 20.3 per 100,000 person-years for UC, and from 0.7 to 14.6 for CD. The observation that IBD incidence is low in developing countries, suggests that environmental factors and diet play an important role in the pathogenesis.

Crohn's disease (CD) is characterized by deep ulcerations that can occur in the entire gastrointestinal tract, i.e. from the mouth to the anus.^{43,44} The disease may affect only one area in the gut, or several areas with healthy areas in between, so called "skip lesions". An early feature of CD is aphthoid ulceration, followed by deep ulcers and fissures in the mucosa at a later stage, which makes up the typical cobblestone pattern. Fistulae and abscesses are often present in a later stage. The inflammation is transmural (affecting all layers of the bowel), and lymphoid hyperplasia, an increase in inflammatory cells and granulomatous lesions are often observed.

Ulcerative colitis (UC) on the other hand, only affects the colon.^{43,44} It can affect the rectum alone (proctitis), it may involve the sigmoid and descending colon (left-sided colitis), or it may involve the whole colon. Typically, the mucosa has a red appearance, bleeds easily and is inflamed. The inflammation is restricted to the mucosa, and crypt abscesses and goblet cell depletion are common features.

Typically, a patient presents with abdominal pain, bloody stools, diarrhea and weight loss. Some patients may also have complaints of malaise, fever, nausea and vomiting. Also, CD can be complicated by anal or perianal disease.

The diagnosis IBD can usually be made based on clinical, radiographic and histologic data.⁴⁵ The distinct patterns of the two diseases often enable the final diagnosis and differentiation between UC and CD on histologic basis. However, this is not always possible, and sometimes the diagnosis interderminate inflammatory colitis is made. In both UC and CD, anemia is common, and erythrocyte sedimentation rate (ESR) is often raised. CRP has been described as a sensitive marker for CD; elevated CRP levels are detectable in 70 – 100% of the CD patients. On the other hand, only 50 – 60% of UC patients have an elevated CRP at diagnosis.^{46,47}

2.2 Pathogenesis of IBD

CD and UC are diseases of unknown etiology. The fact that higher incidence rates are reported in the Western world compared to developing countries, suggests that environmental factors and nutrition may play a role. However, different incidence rates might also result from differences in access to health care and thus lower incidence rates may be reported in developing countries. Next to environmental factors, genetic factors and defects in innate and adaptive immunity may contribute to inflammatory bowel diseases.

2.2.1 Genetics

IBD has a strong genetic component, since a positive family history for the disease is the largest independent risk factor. It has been reported that 2.2 – 16.2% of the CD patients have a first-degree relative with CD, and in 5.2 – 22% with IBD. For UC, this is 5.7 – 15.5% and 6.6 – 15.8% respectively.⁴⁸ Moreover, two studies performed in twins show a pooled estimated concordance in monozygotic twins of 37.3% for CD, and 10% for UC;^{49,50} pooled concordance in dizygotic twins shows 7% for CD and 3% for UC. This suggests that CD might have a stronger genetic component than UC.

However, the disease does not simply result from a single gene defect. Many studies investigating the contribution of genes have been performed, and several genes that are relevant in innate and adaptive immunity have been suggested to be involved in the pathogenesis of IBD. First, the involvement of the *CARD15/NOD2* gene in CD has been shown and con-

firmed in several studies.⁵⁴⁻⁵³ Since CARD15/NOD2 is a known PRR, mutations in this gene might result in altered sensing of bacterial products. Indeed, several mechanisms linking NOD2 dysfunction to CD pathogenesis have been reported, among which enhanced production of IL-1 β ,⁵⁴ abnormalities in TLR2 mediated inflammation in intestinal m ϕ ⁵⁵ and altered NOD2 dependent expression of microbicidal α -defensins.^{56, 57} Importantly, NOD2 knockout mice do not spontaneously develop colitis, indicating that defects in NOD2 only are not sufficient to induce inflammation.⁵⁸

SNPs (single nucleotide polymorphisms) in the tumor necrosis factor superfamily 15 (TNFSF15) gene have also been reported in the pathogenesis of IBD.^{59, 60} Briefly, TNFSF15 is a strong inducer of IFN γ production in T cells and is upregulated in CD4+/CD8+ T cells and macrophages in the lamina propria of CD patients.^{59, 61} Also, a role for SNPs in the IL-23 receptor has been verified in several studies.⁶²⁻⁶⁴ IL-23, together with IL-6 and TGF β , drives the differentiation of naïve T cells towards a Th17 response, thereby initiating an immune response. In addition, other genes in the IL-23 pathway have been implicated in the pathogenesis of IBD, including IL-12B (encodes the p40 subunit of IL23 and IL12) and signals transducer and activator of transcription 3 (STAT3), further suggesting a prominent role for this pathway.⁶⁵⁻⁶⁷

Many other genes have been identified that may play a role in IBD pathogenesis, but a full overview of these genetic defects is beyond the scope of this chapter.

2.2.2 Defects in the immune system in inflammatory bowel diseases

The underlying defect possibly lies in the loss of tolerance towards the mucosal flora, and several defects in innate and adaptive immunity have been reported that may play a role in the development of IBD.

It has been shown that lamina propria mononuclear cells (LPMNCs) from UC and CD patients spontaneously produce large amounts of pro-inflammatory cytokines, thereby triggering an immune response.⁶⁸⁻⁷⁰ In a mouse model of colitis, increased responsiveness to bacterial stimuli has been reported,⁷¹ resulting in aberrant immunity. This suggests that M ϕ from IBD patients display a more M ϕ 1 phenotype, while the ability of intestinal M ϕ to secrete pro-inflammatory cytokines is normally (and preferably) low compared to M ϕ 1. Indeed, lower amounts of M ϕ 2 were found in mucosal biopsies from active lesions in CD patients compared to non-affected colon of the same patient, and compared to healthy controls.⁷² In addition, DCs from both UC and CD patients show higher expression of TLR2 and TLR4 compared to healthy individuals,⁷³ making them hyperresponsive to bacterial antigens. Colonic macrophages from IBD patients have increased expression of the co-stimulatory molecules CD80 and CD86⁷⁴ resulting in an increased ability to activate T cells. As a consequence of these defects, a Th1 or possibly Th17 response is induced.

Leaks in the epithelial barrier, which is the first line of defense, also have been reported in IBD patients.⁷⁵ As a result, pathogens cross the epithelial layer more easily. Interestingly, this defect seems to precede the development of CD in individuals with familial risk,⁷⁶ suggesting a causal role for this defect. In addition, overgrowth of mucosa-associated *Escheria coli* has been observed in CD patients; adherent-invasive *E. coli* (AIEC) are found in 36.4% of the CD patients with ileal involvement. Although *E. coli* is considered a commensal, some strains acquire virulence factors. AIEC bind to the CEACAM6 receptor, which is over-expressed in ileal mucosa of CD patients, leading to abnormal colonization.⁷⁷ Next, they

invade the intestinal barrier, infect and replicate within mucosal macrophages⁷⁸ and induce production of TNF α .

Furthermore, defects in the T cell compartment might contribute to the induction and persistence of IBD. It has been shown that IBD patients have increased numbers of activated T cells in the circulation, and that activated T cells from CD patients are more resistant to apoptosis,⁷⁹ a mechanism involved in programmed cell death which takes place after T cell activation and thereby contributes to homeostasis. As a result, the balance between effector T cells and regulatory T cells is disturbed in IBD patients, followed by uncontrolled inflammation.

2.2.3 Autophagy

Another interesting but rather unexpected discovery is the contribution of the *autophagy-related 16-like 1* (ATG16L1) and *immunity-related GTPase family M* (IRGM) genes to CD pathogenesis,⁸⁰⁻⁸² two genes that are known to be involved in a process called autophagy (referring to the Greek work “autophagos”, i.e. “self-eating”). Autophagy was originally described as a cell survival mechanism. When a cell experiences nutrient depletion, autophagy is induced in order to remove damaged organelles.⁸³ Upon induction of autophagy, a membrane is formed, creating an autophagosome which surrounds the cellular contents and next fuses with lysosomes.⁸⁴ More recently, it became clear that autophagy also plays a crucial role in the clearance of intracellular bacteria,⁸⁵ and in the delivery of cytoplasmic antigens to MHCII molecules for antigen presentation to T cells.⁸⁶ In an experimental ATG16L1 knock-down system, cells showed defective autophagy in response to nutrient depletion and infection, demonstrating the importance of ATG16L1 in the autophagy process.⁸⁷ Since ATG16L1 and IRGM have been confirmed in several Genome Wide Association Studies (GWAS), and given the role of autophagy in general and in immunity, it is likely that autophagy plays an important role in the development of CD. Several mechanisms have been suggested that link defective autophagy to CD. In a DSS colitis model, mice lacking ATG16L1 in hematopoietic cells showed increased production of the pro-inflammatory cytokine IL-1 β .⁸⁸ Furthermore, abnormalities in paneth cells (cells specialized in the secretion of granule contents that contain antimicrobial contents) have been reported in ATG16L1 knockout mice and in CD patients carrying the risk allele.⁸⁹

In summary, environmental, genetic and immunologic defects all contribute to the development of IBD. Likely, the presence of a combination of these factors leads to a loss of response towards the mucosal flora, resulting in inflammation in the gut.

3 Treatment

The main treatment goals in IBD are improving quality of life, reducing hospitalization, surgery and steroid dependency, improving mucosal healing and maintaining clinical remission to control the disease while minimizing side effects. Mucosal healing can lead to significantly higher steroid-free remission rates and less relapses,⁹⁰ and is therefore an important goal to achieve in the treatment of CD patients. Induction therapy is concentrated on quickly reducing signs and symptoms of acute inflammation. However, the underlying disease cause remains unchanged and therefore maintenance therapy is often needed to

prevent relapses. Relapsing disease frequently leads to surgical interventions and hospitalization and for that reason maintaining remission is of great importance. Response to treatment is defined as a decrease in Crohn's Disease Activity Index (CDAI) of 70 points (70-points response) or 100 (100-points-response) after four weeks from baseline in non-fistulizing disease. Here, response is defined as a decrease of 70 points unless stated otherwise. In fistulizing disease, response is achieved when a decrease of at least 50% in the number of draining fistulas after ten weeks is observed. Remission is defined as a CDAI score below 150.^{91,92}

The final goal is to understand the course of disease and to finally alter the course towards a less aggressive phenotype. Several therapeutics are available with different effects, side effects and efficacy profiles to achieve the above described goals.

3.1 5-ASA

5-Aminosalicylic acid (5-ASA, mesalazine), a derivate of salicylic acid, is a non-steroidal anti-inflammatory drug (NSAID).

Whereas induction with 5-ASA therapy for patients with mild-to-moderate CD seems effective,⁹³ it is known that this agent is not effective in inducing remission.⁹⁴ In addition, patients with ileal disease do not benefit from 5-ASA and side effects occur in about one-third of the patients.⁹⁵ The clinical significance of the CDAI reduction obtained with 5-ASA is controversial, and therefore, 5-ASA has limited value in severe disease and in maintenance in CD patients.

On the other hand, 5-ASA is important in the treatment of UC; the efficacy of 5-ASA in severe UC has been shown in several systematic reviews and meta-analyses.^{96,97}

3.2 Steroids

Glucocorticoids are steroid hormones that bind to the glucocorticoid receptor and activate or suppress certain target genes. This then results in decreased production of pro-inflammatory cytokines and inhibition of T cell proliferation.^{98,99}

Although budesonide is more effective than placebo in inducing remission in acute active CD,¹⁰⁰ it is not effective in maintaining remission.¹⁰¹ Also, systemic corticosteroids are very effective in inducing remission in the first place,¹⁰² but do not induce long-term remission,^{103,104} mucosal healing¹⁰⁵ and do not reduce the risk for surgery.¹⁰⁶

Similar to CD, corticosteroids are effective in the induction of remission,¹⁰⁷ and are important in acute severe UC,¹⁰⁸ but are not useful in maintenance therapy.¹⁰⁹ Furthermore, corticosteroids are known to have serious side effects like diabetes mellitus,¹¹⁰ osteoporosis, depression, hypertension, and as a result these agents are associated with increased morbidity and mortality. Therefore, long-term corticosteroid use is discouraged.

3.3 Thiopurines

Azathioprine (AZA) and 6-mercaptopurine (6-MP) have been widely used in the treatment of IBD. AZA is a pro-drug which is converted to 6-MP, which is then metabolized to 6-thioguanine (6-TG). This acts as a DNA synthesis inhibitor, and thereby inhibits proliferation of cells, especially lymphocytes. In addition, it has been shown that azathioprine inhibits T cell proliferation by inhibiting APC-T cell conjugation¹¹¹ and it induces apoptosis in T cells by modulating Rac1 function.¹¹²

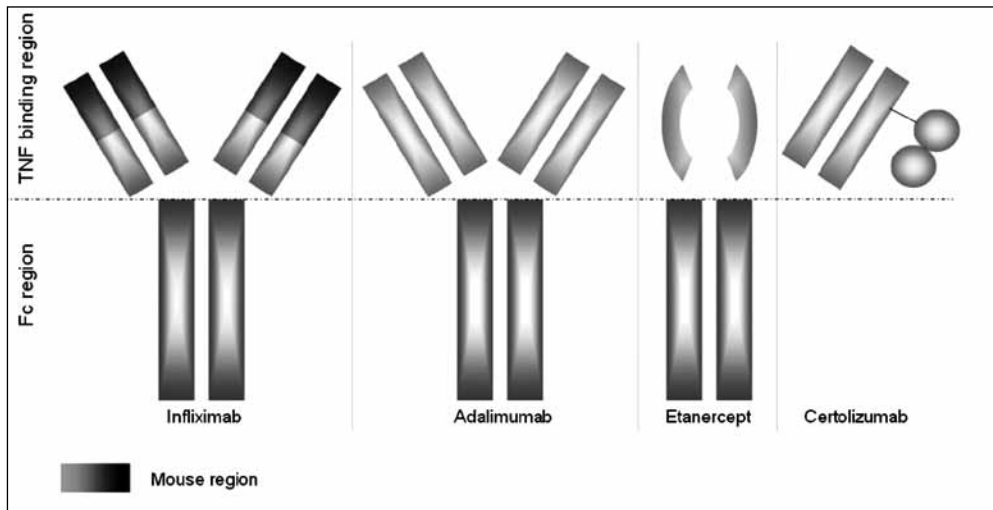
Thiopurines play an important role in controlling CD and have long term efficacy,^{113, 114} but more than half of the patients still depend on corticosteroids. In UC, azathioprine is effective in inducing clinical and endoscopic remission, and is a drug of first choice in patients who are steroid dependent.¹¹⁵ For maintenance therapy, the evidence for thiopurine in UC is weaker than in CD.

Unfortunately, thiopurine therapy has been associated with myelotoxicity, hepatotoxicity¹¹⁶ and lymphoma.¹¹⁷

3.4 Anti-TNF: structure and mechanism of action

Elevated levels of TNF α are detectable in serum and the intestine of IBD patients,^{118, 119} and therefore, blocking this cytokine would potentially alleviate the disease. The introduction of anti-TNF agents during the late '90s has proven to be an effective instrument to achieve the above described treatment goals. Anti-TNF agents induce mucosal healing, reduce steroid dependency, reduce the risk for surgery and hospitalization and improve the patient's quality of life.^{91, 120, 121} In addition, healing of endoscopic lesions^{121, 122} and reduction of chronic inflammatory infiltrates^{122, 123} was achieved.

Figure 3 Different anti-TNF agents.



Anti-TNF agents are designed to neutralize soluble TNF- α ,¹²⁴ an essential Th₁ cytokine produced by monocytes and T cells.¹²⁵ Several anti-TNF agents are available nowadays, and all of them have different structures and properties (Figure 3). Infliximab (Remicade®), is a chimeric monoclonal antibody, and the first anti-TNF antibody on the market for CD. Adalimumab (Humira®), is a completely humanized anti-TNF antibody, and was designed in the hope to reduce immunogenicity. Adalimumab and infliximab are quite similar in structure; both have an Fc region and a Fab region. Certolizumab (Cimzia®) is different from infliximab and adalimumab, since it does not contain an Fc region and thus can not interact with Fc receptors. Whereas certolizumab was approved by the FDA for the treatment of moderate-to-severe CD, it has not been approved by the EMEA. Etanercept (Enbrel®), a

soluble TNF receptor fusion protein, is an effective drug for the treatment of rheumatoid arthritis and also efficiently neutralizes TNF- α .^{124, 126, 127} Surprisingly, etanercept is not beneficial in CD,¹²⁸ implicating that neutralizing soluble TNF- α is not the only mechanism of action of anti-TNF agents responsible for their efficacy in CD. One of the differences between infliximab and etanercept is that infliximab induces apoptosis in lamina propria T cells,¹²⁹ but etanercept does not.¹³⁰ Infliximab, but not etanercept, binds to membrane bound TNF- α (mTNF), which can be cleaved by TNF- α -converting enzyme (TACE) to generate soluble TNF- α . Upon binding to mTNF, infliximab induces antibody dependent cytotoxicity (ADCC) and cell lysis.¹³¹ Mitoma *et al.* described reverse signaling through mTNF induced by infliximab and adalimumab (but not etanercept), leading to cell cycle arrest in Jurkat T cells.^{132, 133} Certolizumab does not induce apoptosis, but, like infliximab and adalimumab, inhibits LPS-induced IL-1 β production by monocytes.¹²⁴ In addition, infliximab induces apoptosis in monocytes from CD patients with active disease.¹³⁴ Furthermore, infliximab reduces VCAM-1 (vascular cell adhesion molecule-1) and CD40 expression on mucosal endothelium, thereby disrupting the CD40-CD40L dependent interaction between T cells and endothelium.¹³⁵ Infliximab also acts on wound healing: infliximab increases tissue inhibitor of metalloproteinases-1 (TIMP-1) production and reduces matrix metalloproteinase (MMP) activity, and enhances myofibroblast migration *in vitro*.¹³⁶

3.5 Anti-TNF therapy efficacy in clinical trials

The efficacy and safety of anti-TNF therapy has been widely evaluated in clinical studies Targan *et al.* reported response rates at week 4 of overall 65% in patients treated with infliximab vs. 17% in the placebo group.⁹¹ In the ACCENT I trial, 58% responded to infliximab at week 2.¹³⁷ In another study, much higher response rates were observed in patients naïve to immunomodulators and biologics with short duration of disease.¹³⁸ Response rates of anti-TNF-naïve patients treated with adalimumab were assessed in the CLASSIC-I trial. After 4 weeks, 54% (adalimumab 40/20 mg) to 59% (adalimumab 80/40 and 160/80) showed a clinical response.⁹² Clinical response rates of patients receiving certolizumab pegol were evaluated in the PRECISE-1 trial and were 44% at week 4¹³⁹. The 100-points response at week 6 (primary end point) after full induction therapy was 35%. Both response rates did not reach statistical significance. Patients who received anti-TNF therapy within the previous three months, or had a hypersensitivity or lack of response to a first anti-TNF dose were excluded. Infliximab, adalimumab, as well as certolizumab appeared safe.

Altogether, around one-third of patients treated with infliximab, 45% of patients treated with adalimumab and 56% of patients receiving certolizumab fail to show a clinical response at week 4 or week 6 after full induction therapy. Because there are no head-to-head trials, it is complex to directly compare the results of different studies. It is not known whether these primary non-responders represent a specific group of patients. Patients who do not show a response after a first infusion of infliximab, also fail to show response after subsequent infusions,^{91, 140} suggesting that lack of response is stable over time. In general, although many clinical trials have shown the efficacy and safety of anti-TNF therapy, there is a relatively large group of patients displaying lack of response after 4 weeks (primary non-responders). In addition to lack of response, a considerable group of patients lose response following an initial response after several months of treatment. Loss of response is generally defined as a history of initial response and lack of improvement or worsening of symptoms, including:

increased stool frequency, fever, rectal bleeding, daily abdominal pain and recurring drainage from a previously non-draining fistula.¹⁴¹ Furthermore, a significant number of patients become intolerant to anti-TNF, which is characterized by acute (during or within 24 h post treatment) or delayed (occurring 24 h – 15 days post treatment) infusion reactions.

Lack and loss of response to anti-TNF therapy is an obstacle in the treatment of CD. Predictive factors for lack and loss of response are needed to select patients for a certain approach. This may improve treatment, reduce side-effects and reduce morbidity. Many studies have been done to identify factors for lack and loss of response. Genetic, clinical and demographic factors have been described to play a role in lack of response. FcγRIIIa,¹⁴² TACE¹⁴³ and LTA¹⁴⁰ might be possible genetic factors, and young age,^{144, 145} luminal CD,¹⁴⁵ short duration of disease¹⁴⁶ and concurrent immunosuppression^{145, 147-150} possible clinical factors. Patients with lack of response can switch to another anti-TNF, or to a biologic with another mechanism of action. Though controversial, the formation of antibodies against antibodies has been associated with loss of response. The use of concurrent immunosuppressive drugs may reduce the formation of antibodies against antibodies. Dose intensification in patients with low drug through levels or switching to another anti-TNF might be a good option in patients with loss of response.

Also, side effects might complicate anti-TNF treatment. Anti-TNF use is associated with an approximately 21-fold increased risk of tuberculosis (TB) without appropriate safety measures.¹⁵¹ The TB incidence has been reported to decrease with 78% when suitable safety measures were undertaken. Most cases are presented during the first three months of treatment and have an atypical presentation, which makes the diagnosis more complicating.¹⁵² For that reason, international guidelines advise to assess the risk of TB before starting treatment with an anti-TNF agent, including an X-ray, tuberculin skin testing (depending on national guidelines) and careful evaluation of the TB history.¹⁵³ Latent TB may be suspected in case of a positive initial tuberculin skin test and when the patient has recently been exposed to the disease. Physicians should be aware of the possibility of false-negative skin tests, especially when patients are immunocompromized.

Next to reactivation of *Mycobacterium tuberculosis*, the use of immunosuppressive agents is associated with opportunistic infections. The risk of opportunistic infections in anti-TNF treated patients is estimated between 0.3 and 0.9%,¹⁵⁴ and an increased risk is observed in patients treated with concomittant immunosuppressives.¹⁵⁵ Indeed, in a large meta-analysis of 21 placebo-controlled trials including 5356 anti-TNF treated patients, the increased risk of opportunistic infections was likely due to disease severity and prednisone use, instead of merely anti-TNF.¹⁵⁶ In line with this observation, no increased risk was found in infections and mortality in 734 anti-TNF treated patients compared to controls, with a median follow-up of 58 months.¹⁴⁹

Finally, the development of malignancies, and especially lymphomas, is a major concern. Whereas some studies do not show an increased risk,¹⁵⁷⁻¹⁶⁰ other studies do find a moderately elevated risk, especially in patients on thiopurine therapy.^{117, 161, 162} Lethal hepatosplenic T cell lymphoma has been reported in young patients on azathioprine/infliximab combination therapy,¹⁶³⁻¹⁶⁶ and therefore long-term combination therapy in younger patients is not recommended. Still, the absolute risk appears to be low and should be weighed against the beneficial effects of immunomodulator therapy. In addition, differences in study design and patient recruitment complicate the interpretation of these data. Furthermore, in the

meta-analysis including 21 placebo-controlled trials,¹⁵⁶ no increased risk of malignancy was observed. These data were supported by another study¹⁴⁹ including 734 anti-TNF treated patients. However, long-term safety data are not available yet and therefore awareness of (serious) side effects is warranted.

3.6 Other therapies

There is not much data on the efficacy of methotrexate in UC, but the only randomized placebo-controlled trial did not show any benefit.¹⁶⁷ In contrast, methotrexate efficacy has been shown CD patients.¹⁶⁸ and at the present time, methotrexate therapy is used in patients with active or relapsing CD who are refractory or intolerant to thiopurine therapy or anti-TNF agents.¹⁶⁹ Other immunosuppressives, like ciclosporin or tacrolimus, may be of benefit in patients with severe UC who are intolerant to i.v. corticosteroids.¹⁰⁹ Ciclosporin is of limited value in CD,⁴⁵ but data are lacking on the efficacy of tacrolimus in CD.

3.7 Future treatment

Unfortunately, the medication used to control CD is not without risk. Although substantial progression has been made with regard to treatment, there is still no cure for IBD. In addition, side effects like lymphoma, *Mycobacterium tuberculosis* and opportunistic infections further complicate treatment. Moreover, lack and loss of response to anti-TNF therapy are problems in daily practice that are even now unsolved, and surgery is then often the only option left.

Our understanding of the mechanism of action and side effects of several therapies is still incomplete. In an ideal situation, it would be possible to select patients based on genotype or disease phenotype, age or other yet undefined factors for a certain therapeutic strategy. To accomplish this, a better understanding of the pathogenesis of CD and the complex mechanism of action of anti-TNF therapy is warranted in order to tailor therapy. In that way, it is possible to reduce side effects, surgery and chronic use of corticosteroids. In addition, therapy risks need to be re-assessed at any given time point, and if necessary, the therapeutic approach should be re-adjusted. Finally, we have to continue exploring new therapies with less side effects and high efficacy profiles. One interesting development in the treatment of inflammatory disorders, is the administration of mesenchymal stem cells (MSC). MSC are cells with immunosuppressive properties^{170, 171} and have been studied in various fields of medicine. Administration of MSCs to patients with severe steroid-refractory graft-versus-host disease (GvDH), including GvDH of the gut, has been shown to be effective.^{172, 173} This may be a promising strategy in the treatment of CD.

4 Scope of the thesis

In *chapter 2*, we give an overview of current treatment strategies for CD, especially the top-down approach. Since CD typically progresses from an inflammatory to a fibrotic phenotype, it may be beneficial to interfere in an early disease stage in patients with high risk at developing complicated disease.

In *chapter 3*, we demonstrate the role of autophagy in DCs in regulation of the immunological synapse and CD pathogenesis. We show that decreased levels of autophagy lead to

hyperstabilization of the immunological synapse. This results in increased interaction duration between DCs and T cells and increased T cell activation and IL-17 production. Also, we demonstrate that autophagosomes contain components of the synapse, suggesting that autophagy might be involved in the synaptic breakdown, and thereby plays a role in controlling T cell responses. In addition, we found the same results in patients carrying the ATG16L1 risk allele, indicating a novel role for autophagy in CD pathogenesis by modulating adaptive immune responses.

In *chapter 4*, we further dissect the mechanism of action of anti-TNF agents *in vitro*. We describe the Fc-receptor dependent induction of M ϕ 2 upon infliximab therapy and their immunosuppressive phenotype. Since loss of tolerance and hyperresponsiveness contribute to IBD, the induction of M ϕ 2 by infliximab might restore the dysbalance.

The induction of M ϕ 2 *in vivo* is shown in *chapter 5*, and a significant relation between response to infliximab and induction of M ϕ 2 is described. Also, we show the wound healing capacity of infliximab-induced macrophages, further supporting their role in mucosal healing. Furthermore, we show an enhanced induction of M ϕ 2 upon infliximab/azathioprine combination treatment, and that M ϕ 2 induced by combination treatment have a stronger immunosuppressive phenotype. This might explain the superiority of infliximab/azathioprine combination treatment observed in patients.

In *chapter 6* a Phase I study investigating the safety and feasibility of MSC therapy in steroid-refractory CD patients is described. We show the immunosuppressive properties of MSCs *in vitro* and that MSC therapy in CD patients is safe and feasible. Importantly, no serious side effects were reported during the study period. The efficacy of MSC therapy in CD patients should be further assessed in Phase II/III trials.

In *chapter 7* we further examine the safety profile of common IBD drugs in relation to lymphoma development. In a cohort of approximately 18000 patients, no increased risk was found compared to the general population, but a clear association was observed between thiopurine therapy and EBV positive lymphoma, especially in younger patients. These data give more insight in the risks in specific patient groups.

Reference List

1. Janeway CA, Jr., Medzhitov R. Innate immune recognition. *Annu Rev Immunol* 2002;20:197-216.
2. Medzhitov R, Janeway CA, Jr. Innate immune recognition and control of adaptive immune responses. *Semin Immunol* 1998;10:351-353.
3. Zhang G, Ghosh S. Molecular mechanisms of NF-kappaB activation induced by bacterial lipopolysaccharide through Toll-like receptors. *J Endotoxin Res* 2000;6:453-457.
4. Girardin SE, Boneca IG, Viala J et al. Nod2 is a general sensor of peptidoglycan through muramyl dipeptide (MDP) detection. *J Biol Chem* 2003;278:8869-8872.
5. Clark GJ, Angel N, Kato M et al. The role of dendritic cells in the innate immune system. *Microbes Infect* 2000;2:257-272.
6. Adema GJ. Dendritic cells from bench to bedside and back. *Immunol Lett* 2009;122:128-130.
7. Iwasaki A, Medzhitov R. Regulation of adaptive immunity by the innate immune system. *Science* 2010;327:291-295.

8. Kapsenberg ML. Dendritic-cell control of pathogen-driven T-cell polarization. *Nat Rev Immunol* 2003;3:984-993.
9. Reis e Sousa. Activation of dendritic cells: translating innate into adaptive immunity. *Curr Opin Immunol* 2004;16:21-25.
10. Naito M. Macrophage differentiation and function in health and disease. *Pathol Int* 2008;58:143-155.
11. Naito M, Umeda S, Yamamoto T et al. Development, differentiation, and phenotypic heterogeneity of murine tissue macrophages. *J Leukoc Biol* 1996;59:133-138.
12. Ma J, Chen T, Mandelin J et al. Regulation of macrophage activation. *Cell Mol Life Sci* 2003;60:2334-2346.
13. Gordon S, Martinez FO. Alternative activation of macrophages: mechanism and functions. *Immunity* 2010;32:593-604.
14. Martinez FO, Helming L, Gordon S. Alternative activation of macrophages: an immunologic functional perspective. *Annu Rev Immunol* 2009;27:451-483.
15. Verreck FA, de BT, Langenberg DM et al. Phenotypic and functional profiling of human proinflammatory type-1 and anti-inflammatory type-2 macrophages in response to microbial antigens and IFN-gamma- and CD40L-mediated costimulation. *J Leukoc Biol* 2006;79:285-293.
16. Schebesch C, Kodelja V, Muller C et al. Alternatively activated macrophages actively inhibit proliferation of peripheral blood lymphocytes and CD4+ T cells in vitro. *Immunology* 1997;92:478-486.
17. Kodelja V, Muller C, Tenorio S et al. Differences in angiogenic potential of classically vs alternatively activated macrophages. *Immunobiology* 1997;197:478-493.
18. Daley JM, Brancato SK, Thomay AA et al. The phenotype of murine wound macrophages. *J Leukoc Biol* 2010;87:59-67.
19. Carrier Y, Yuan J, Kuchroo VK et al. Th3 cells in peripheral tolerance. I. Induction of Foxp3-positive regulatory T cells by Th3 cells derived from TGF-beta T cell-transgenic mice. *J Immunol* 2007;178:179-185.
20. Murphy KM, Reiner SL. The lineage decisions of helper T cells. *Nat Rev Immunol* 2002;2:933-944.
21. Crome SQ, Wang AY, Levings MK. Translational mini-review series on Th17 cells: function and regulation of human T helper 17 cells in health and disease. *Clin Exp Immunol* 2010;159:109-119.
22. Weiner HL. Oral tolerance: immune mechanisms and the generation of Th3-type TGF-beta-secreting regulatory cells. *Microbes Infect* 2001;3:947-954.
23. Tang Q, Bluestone JA. The Foxp3+ regulatory T cell: a jack of all trades, master of regulation. *Nat Immunol* 2008;9:239-244.
24. Harrington LE, Hatton RD et al. Interleukin 17-producing CD4+ effector T cells develop via a lineage distinct from the T helper type 1 and 2 lineages. *Nat Immunol* 2005;6:1123-1132.
25. Mangan PR, Harrington LE, O'Quinn DB et al. Transforming growth factor-beta induces development of the T(H)17 lineage. *Nature* 2006;441:231-234.
26. Liu ZJ, Yadav PK, Su J et al. Potential role of Th17 cells in the pathogenesis of inflammatory bowel disease. *World J Gastroenterol* 2009;15:5784-5788.
27. Seder RA, Ahmed R. Similarities and differences in CD4+ and CD8+ effector and memory T cell generation. *Nat Immunol* 2003;4:835-842.

28. Yewdell JW, Norbury CC, Bennink JR. Mechanisms of exogenous antigen presentation by MHC class I molecules in vitro and in vivo: implications for generating CD8+ T cell responses to infectious agents, tumors, transplants, and vaccines. *Adv Immunol* 1999;73:1-77.
29. Grakoui A, Bromley SK, Sumen C et al. The immunological synapse: a molecular machine controlling T cell activation. *Science* 1999;285:221-227.
30. Rodriguez-Fernandez JL, Riol-Blanco L, Delgado-Martin C. What is the function of the dendritic cell side of the immunological synapse? *Sci Signal* 2010;3:re2.
31. Vicente-Manzanares M, Sanchez-Madrid F. Role of the cytoskeleton during leukocyte responses. *Nat Rev Immunol* 2004;4:110-122.
32. Al-Alwan MM, Liwski RS, Haeryfar SM et al. Cutting edge: dendritic cell actin cytoskeletal polarization during immunological synapse formation is highly antigen-dependent. *J Immunol* 2003;171:4479-4483.
33. Hughes JB, Hellmann JJ, Ricketts TH et al. Counting the uncountable: statistical approaches to estimating microbial diversity. *Appl Environ Microbiol* 2001;67:4399-4406.
34. Macpherson AJ, Harris NL. Interactions between commensal intestinal bacteria and the immune system. *Nat Rev Immunol* 2004;4:478-485.
35. Muller CA, Autenrieth IB, Peschel A. Innate defenses of the intestinal epithelial barrier. *Cell Mol Life Sci* 2005;62:1297-1307.
36. Johansson C, Kelsall BL. Phenotype and function of intestinal dendritic cells. *Seminars in Immunology* 2005;17:284-294.
37. Smythies LE, Sellers M, Clements RH et al. Human intestinal macrophages display profound inflammatory anergy despite avid phagocytic and bacteriocidal activity. *J Clin Invest* 2005;115:66-75.
38. Kamada N, Hisamatsu T, Okamoto S et al. Abnormally differentiated subsets of intestinal macrophage play a key role in Th1-dominant chronic colitis through excess production of IL-12 and IL-23 in response to bacteria. *J Immunol* 2005;175:6900-6908.
39. Smith PD, Ochsenbauer-Jambor C, Smythies LE. Intestinal macrophages: unique effector cells of the innate immune system. *Immunol Rev* 2005;206:149-159.
40. Russel MG, Dorant E, Volovics A et al. High incidence of inflammatory bowel disease in The Netherlands: results of a prospective study. The South Limburg IBD Study Group. *Dis Colon Rectum* 1998;41:33-40.
41. Bernstein CN, Blanchard JF, Rawsthorne P et al. Epidemiology of Crohn's disease and ulcerative colitis in a central Canadian province: a population-based study. *Am J Epidemiol* 1999;149:916-924.
42. Rubin GP, Hungin AP, Kelly PJ et al. Inflammatory bowel disease: epidemiology and management in an English general practice population. *Aliment Pharmacol Ther* 2000;14:1553-1559.
43. Podolsky DK. Inflammatory bowel disease. *N Engl J Med* 2002;347:417-429.
44. Baumgart DC, Sandborn WJ. Inflammatory bowel disease: clinical aspects and established and evolving therapies. *The Lancet* 2007;369:1641-1657.
45. Dignass A, Assche GV, Lindsay JO et al. The second European evidence-based Consensus on the diagnosis and management of Crohn's disease: Current management. 4 ed. 2010:28-62.
46. Beattie RM, Walker-Smith JA, Murch SH. Indications for investigation of chronic gastrointestinal symptoms. *Arch Dis Child* 1995;73:354-355.
47. Shine B, Berghouse L, Jones JE et al. C-reactive protein as an aid in the differentiation of functional and inflammatory bowel disorders. *Clin Chim Acta* 1985;148:105-109.

48. Russell RK, Satsangi J. IBD: a family affair. *Best Pract Res Clin Gastroenterol* 2004;18:525-539.
49. Thompson NP, Driscoll R, Pounder RE et al. Genetics versus environment in inflammatory bowel disease: results of a British twin study. *BMJ* 1996;312:95-96.
50. Tysk C, Lindberg E, Jarnerot G et al. Ulcerative colitis and Crohn's disease in an unselected population of monozygotic and dizygotic twins. A study of heritability and the influence of smoking. *Gut* 1988;29:990-996.
51. Hugot JP, Chamaillard M, Zouali H et al. Association of NOD2 leucine-rich repeat variants with susceptibility to Crohn's disease. *Nature* 2001;411:599-603.
52. Ogura Y, Bonen DK, Inohara N et al. A frameshift mutation in NOD2 associated with susceptibility to Crohn's disease. *Nature* 2001;411:603-606.
53. Hampe J, Cuthbert A, Croucher PJ et al. Association between insertion mutation in NOD2 gene and Crohn's disease in German and British populations. *Lancet* 2001;357:1925-1928.
54. Maeda S, Hsu LC, Liu H et al. Nod2 mutation in Crohn's disease potentiates NF-kappaB activity and IL-1beta processing. *Science* 2005;307:734-738.
55. Watanabe T, Kitani A, Murray PJ et al. Nucleotide binding oligomerization domain 2 deficiency leads to dysregulated TLR2 signaling and induction of antigen-specific colitis. *Immunity* 2006;25:473-485.
56. Kobayashi K, Inohara N, Hernandez LD et al. RICK/Rip2/CARDIAK mediates signalling for receptors of the innate and adaptive immune systems. *Nature* 2002;416:194-199.
57. Wehkamp J, Salzman NH, Porter E et al. Reduced Paneth cell alpha-defensins in ileal Crohn's disease. *Proc Natl Acad Sci U S A* 2005;102:18129-18134.
58. Pauleau AL, Murray PJ. Role of nod2 in the response of macrophages to toll-like receptor agonists. *Mol Cell Biol* 2003;23:7531-7539.
59. Thiebaut R, Kotti S, Jung C et al. TNFSF15 polymorphisms are associated with susceptibility to inflammatory bowel disease in a new European cohort. *Am J Gastroenterol* 2009;104:384-391.
60. Yamazaki K, McGovern D, Ragoussis J et al. Single nucleotide polymorphisms in TNFSF15 confer susceptibility to Crohn's disease. *Hum Mol Genet* 2005;14:3499-3506.
61. Picornell Y, Mei L, Taylor K et al. TNFSF15 is an ethnic-specific IBD gene. *Inflamm Bowel Dis* 2007;13:1333-1338.
62. Duerr RH, Taylor KD, Brant SR et al. A genome-wide association study identifies IL23R as an inflammatory bowel disease gene. *Science* 2006;314:1461-1463.
63. Lacher M, Schroepf S, Helmbrecht J et al. Association of the interleukin-23 receptor gene variant rs11209026 with Crohn's disease in German children. *Acta Paediatr* 2010;99:727-733.
64. Tremelling M, Cummings F, Fisher SA et al. IL23R variation determines susceptibility but not disease phenotype in inflammatory bowel disease. *Gastroenterology* 2007;132:1657-1664.
65. Lees CW, Barrett JC, Parkes M et al. New IBD genetics: common pathways with other diseases. *Gut* 2011. Feb 7. [Epub ahead of print]
66. Wang K, Zhang H, Kugathasan S et al. Diverse genome-wide association studies associate the IL12/IL23 pathway with Crohn Disease. *Am J Hum Genet* 2009;84:399-405.
67. Ferguson LR, Han DY, Fraser AG et al. Genetic factors in chronic inflammation: single nucleotide polymorphisms in the STAT-JAK pathway, susceptibility to DNA damage and Crohn's disease in a New Zealand population. *Mutat Res* 2010;690:108-115.
68. Reinecker HC, Steffen M, Witthoef T et al. Enhanced secretion of tumour necrosis factor-alpha, IL-6, and IL-1 beta by isolated lamina propria mononuclear cells from patients with ulcerative colitis and Crohn's disease. *Clin Exp Immunol* 1993;94:174-181.

69. Reimund JM, Wittersheim C, Dumont S et al. Mucosal inflammatory cytokine production by intestinal biopsies in patients with ulcerative colitis and Crohn's disease. *J Clin Immunol* 1996;16:144-150.
70. Zareie M, Singh PK, Irvine EJ et al. Monocyte/macrophage activation by normal bacteria and bacterial products: implications for altered epithelial function in Crohn's disease. *Am J Pathol* 2001;158:1101-1109.
71. Takakura R, Kiyohara T, Murayama Y et al. Enhanced macrophage responsiveness to lipopolysaccharide and CD40 stimulation in a murine model of inflammatory bowel disease: IL-10-deficient mice. *Inflamm Res* 2002;51:409-415.
72. Hunter MM, Wang A, Parhar KS et al. In Vitro-Derived Alternatively Activated Macrophages Reduce Colonic Inflammation in Mice. *Gastroenterology* 2010;138(4):1395-40.
73. Hart AL, Al-Hassi HO, Rigby RJ et al. Characteristics of intestinal dendritic cells in inflammatory bowel diseases. *Gastroenterology* 2005;129:50-65.
74. Rogler G, Hausmann M, Spottl T et al. T-cell co-stimulatory molecules are upregulated on intestinal macrophages from inflammatory bowel disease mucosa. *Eur J Gastroenterol Hepatol* 1999;11:1105-1111.
75. Soderholm JD, Olaison G, Peterson KH et al. Augmented increase in tight junction permeability by luminal stimuli in the non-inflamed ileum of Crohn's disease. *Gut* 2002;50:307-313.
76. Irvine EJ, Marshall JK. Increased intestinal permeability precedes the onset of Crohn's disease in a subject with familial risk. *Gastroenterology* 2000;119:1740-1744.
77. Barnich N, Carvalho FA, Glasser AL et al. CEACAM6 acts as a receptor for adherent-invasive *E. coli*, supporting ileal mucosa colonization in Crohn disease. *J Clin Invest* 2007;117:1566-1574.
78. Glasser AL, Boudeau J, Barnich N et al. Adherent invasive *Escherichia coli* strains from patients with Crohn's disease survive and replicate within macrophages without inducing host cell death. *Infect Immun* 2001;69:5529-5537.
79. Sturm A, de Souza HS, Fiocchi C. Mucosal T cell proliferation and apoptosis in inflammatory bowel disease. *Curr Drug Targets* 2008;9:381-387.
80. Hampe J, Franke A, Rosenstiel P et al. A genome-wide association scan of nonsynonymous SNPs identifies a susceptibility variant for Crohn disease in *ATG16L1*. *Nat Genet* 2007;39:207-211.
81. Kuballa P, Huett A, Rioux JD et al. Impaired autophagy of an intracellular pathogen induced by a Crohn's disease associated *ATG16L1* variant. *PLoS One* 2008;3:e3391.
82. Parkes M, Barrett JC, Prescott NJ et al. Sequence variants in the autophagy gene *IRGM* and multiple other replicating loci contribute to Crohn's disease susceptibility. *Nat Genet* 2007;39:830-832.
83. Mizushima N, Klionsky DJ. Protein turnover via autophagy: implications for metabolism. *Annu Rev Nutr* 2007;27:19-40.
84. Yorimitsu T, Klionsky DJ. Autophagy: molecular machinery for self-eating. *Cell Death Differ* 2005;12 Suppl 2:1542-1552.
85. Birmingham CL, Smith AC, Bakowski MA et al. Autophagy controls *Salmonella* infection in response to damage to the *Salmonella*-containing vacuole. *J Biol Chem* 2006;281:11374-11383.
86. Schmid D, Pypaert M, Munz C. Antigen-loading compartments for major histocompatibility complex class II molecules continuously receive input from autophagosomes. *Immunity* 2007;26:79-92.

87. Rioux JD, Xavier RJ, Taylor KD et al. Genome-wide association study identifies new susceptibility loci for Crohn disease and implicates autophagy in disease pathogenesis. *Nat Genet* 2007;39:596-604.
88. Saitoh T, Fujita N, Jang MH et al. Loss of the autophagy protein Atg16L1 enhances endotoxin-induced IL-1beta production. *Nature* 2008;456:264-268.
89. Cadwell K, Liu JY, Brown SL et al. A key role for autophagy and the autophagy gene Atg16L1 in mouse and human intestinal Paneth cells. *Nature* 2008;456:259-263.
90. Baert F, Moortgat L, van AG et al. Mucosal healing predicts sustained clinical remission in patients with early-stage Crohn's disease. *Gastroenterology* 2010;138:463-468.
91. Targan SR, Hanauer SB, van Deventer SJH et al. A Short-Term Study of Chimeric Monoclonal Antibody cA2 to Tumor Necrosis Factor {alpha} for Crohn's Disease. *N Engl J Med* 1997;337:1029-1036.
92. Hanauer SB, Sandborn WJ, Rutgeerts P et al. Human Anti-Tumor Necrosis Factor Monoclonal Antibody (Adalimumab) in Crohn's Disease: the CLASSIC-I Trial. *Gastroenterology* 2006;130:323-333.
93. Sandborn WJ, Feagan BG, Lichtenstein GR. Medical management of mild to moderate Crohn's disease: evidence-based treatment algorithms for induction and maintenance of remission. *Aliment Pharmacol Ther* 2007;26:987-1003.
94. Hanauer SB, Stromberg U. Oral Pentasa in the treatment of active Crohn's disease: A meta-analysis of double-blind, placebo-controlled trials. *Clin Gastroenterol Hepatol* 2004;2:379-388.
95. Taffet SL, Das KM. Sulfasalazine. Adverse effects and desensitization. *Dig Dis Sci* 1983;28:833-842.
96. Bebb JR, Scott BB. How effective are the usual treatments for ulcerative colitis? *Aliment Pharmacol Ther* 2004;20:143-149.
97. Sutherland L, Macdonald JK. Oral 5-aminosalicylic acid for induction of remission in ulcerative colitis. *Cochrane Database Syst Rev* 2006;CD000543.
98. Barnes PJ, Adcock I. Anti-inflammatory actions of steroids: molecular mechanisms. *Trends Pharmacol Sci* 1993;14:436-441.
99. Boumpas DT, Paliogianni F, Anastassiou ED et al. Glucocorticosteroid action on the immune system: molecular and cellular aspects. *Clin Exp Rheumatol* 1991;9:413-423.
100. Otley A, Steinhart AH. Budesonide for induction of remission in Crohn's disease. *Cochrane Database Syst Rev* 2005;CD000296.
101. Benchimol EI, Seow CH, Otley AR et al. Budesonide for maintenance of remission in Crohn's disease. *Cochrane Database Syst Rev* 2009;CD002913.
102. Benchimol EI, Seow CH, Steinhart AH et al. Traditional corticosteroids for induction of remission in Crohn's disease. *Cochrane Database Syst Rev* 2008;CD006792.
103. Faubion WA, Jr., Loftus EV, Jr., Harmsen WS et al. The natural history of corticosteroid therapy for inflammatory bowel disease: a population-based study. *Gastroenterology* 2001;121:255-260.
104. Steinhart AH, Ewe K, Griffiths AM et al. Corticosteroids for maintenance of remission in Crohn's disease. *Cochrane Database Syst Rev* 2003;CD000301.
105. Modigliani R, Mary JY, Simon JF et al. Clinical, biological, and endoscopic picture of attacks of Crohn's disease. Evolution on prednisolone. *Groupe d'Etude Therapeutique des Affections Inflammatoires Digestives. Gastroenterology* 1990;98:811-818.
106. Cosnes J, Nion-Larmurier I, Beaugerie L et al. Impact of the increasing use of immunosuppressants in Crohn's disease on the need for intestinal surgery. *Gut* 2005;54:237-241.

107. Lennard-Jones JE, Longmore AJ, Newell AC et al. An assessment of prednisone, salazopyrin, and topical hydrocortisone hemisuccinate used as out-patient treatment for ulcerative colitis. *Gut* 1960;1:217-222.
108. Campieri M, Adamo S, Valpiani D et al. Oral beclometasone dipropionate in the treatment of extensive and left-sided active ulcerative colitis: a multicentre randomised study. *Aliment Pharmacol Ther* 2003;17:1471-1480.
109. Travis SP, Stange EF, Lemann M et al. European evidence-based Consensus on the management of ulcerative colitis: Current management. *J Crohns Colitis* 2008;2:24-62.
110. Donihi AC, Raval D, Saul M et al. Prevalence and predictors of corticosteroid-related hyperglycemia in hospitalized patients. *Endocr Pract* 2006;12:358-362.
111. Poppe D, Tiede I, Fritz G et al. Azathioprine suppresses ezrin-radixin-moesin-dependent T cell-APC conjugation through inhibition of Vav guanosine exchange activity on Rac proteins. *J Immunol* 2006;176:640-651.
112. Tiede I, Fritz G, Strand S et al. CD28-dependent Rac1 activation is the molecular target of azathioprine in primary human CD4+ T lymphocytes. *J Clin Invest* 2003;111:1133-1145.
113. Bouhnik Y, Lemann M, Mary JY et al. Long-term follow-up of patients with Crohn's disease treated with azathioprine or 6-mercaptopurine. *Lancet* 1996;347:215-219.
114. Prefontaine E, Macdonald JK, Sutherland LR. Azathioprine or 6-mercaptopurine for induction of remission in Crohn's disease. *Cochrane Database Syst Rev* 2010;6:CD000545.
115. Ardizzone S, Maconi G, Russo A et al. Randomised controlled trial of azathioprine and 5-aminosalicylic acid for treatment of steroid dependent ulcerative colitis. *Gut* 2006;55:47-53.
116. de Boer NK, van Bodegraven AA, Jharap B et al. Drug Insight: pharmacology and toxicity of thiopurine therapy in patients with IBD. *Nat Clin Pract Gastroenterol Hepatol* 2007;4:686-694.
117. Beaugerie L, Brousse N, Bouvier AM et al. Lymphoproliferative disorders in patients receiving thiopurines for inflammatory bowel disease: a prospective observational cohort study. *The Lancet* 2009;374:1617-1625.
118. Plevy SE, Landers CJ, Prehn J et al. A role for TNF-alpha and mucosal T helper-1 cytokines in the pathogenesis of Crohn's disease. *J Immunol* 1997;159:6276-6282.
119. van Deventer SJH. Review article: targeting TNF as a key cytokine in the inflammatory processes of Crohns disease the mechanisms of action of infliximab. *Alimentary Pharmacology & Therapeutics* 1999;13:3-8.
120. Rutgeerts P, D'Haens G, Targan S et al. Efficacy and safety of retreatment with anti-tumor necrosis factor antibody (infliximab) to maintain remission in Crohn's disease. *Gastroenterology* 1999;117:761-769.
121. van Dullemen HM, van Deventer SJ, Hommes DW et al. Treatment of Crohn's disease with anti-tumor necrosis factor chimeric monoclonal antibody (cA2). *Gastroenterology* 1995;109:129-135.
122. D'Haens G, Van Deventer S, Van Hogezaand R et al. Endoscopic and histological healing with infliximab anti-tumor necrosis factor antibodies in Crohn's disease: A European multicenter trial. *Gastroenterology* 1999;116:1029-1034.
123. Baert FJ, D'Haens GR, Peeters M et al. Tumor necrosis factor alpha antibody (infliximab) therapy profoundly down-regulates the inflammation in Crohn's ileocolitis. *Gastroenterology* 1999;116:22-28.

124. Nesbitt A, Fossati G, Bergin M et al. Mechanism of action of certolizumab pegol (CDP870): in vitro comparison with other anti-tumor necrosis factor alpha agents. *Inflamm Bowel Dis* 2007;13:1323-1332.
125. Schreiber S, Nikolaus S, Hampe J et al. Tumour necrosis factor [alpha] and interleukin 1[beta] in relapse of Crohn's disease. *The Lancet* 1999;353:459-461.
126. Moreland LW, Schiff MH, Baumgartner SW et al. Etanercept Therapy in Rheumatoid Arthritis: A Randomized, Controlled Trial. *Ann Intern Med* 1999;130:478-486.
127. Moreland LW, Baumgartner SW, Schiff MH et al. Treatment of Rheumatoid Arthritis with a Recombinant Human Tumor Necrosis Factor Receptor (p75)-Fc Fusion Protein. *N Engl J Med* 1997;337:141-147.
128. Sandborn WJ, Hanauer SB, Katz S et al. Etanercept for Active Crohn's Disease: A Randomized, Double-Blind, Placebo-Controlled Trial. *Gastroenterology* 2001;121:1088-1094.
129. ten Hove T, van Montfrans C, Peppelenbosch MP et al. Infliximab treatment induces apoptosis of lamina propria T lymphocytes in Crohn's disease. *Gut* 2002;50:206-211.
130. Van den Brande JMH, Braat H, van den Brink G et al. Infliximab but not etanercept induces apoptosis in lamina propria T-lymphocytes from patients with Crohn's disease. *Gastroenterology* 2003;124:1774-1785.
131. Scallon BJ, Moore MA, Trinh H et al. Chimeric anti-TNF-[alpha] monoclonal antibody cA2 binds recombinant transmembrane TNF-[alpha] and activates immune effector functions. *Cytokine* 1995;7:251-259.
132. Mitoma H, Horiuchi T, Hata N et al. Infliximab induces potent anti-inflammatory responses by outside-to-inside signals through transmembrane TNF-[alpha]. *Gastroenterology* 2005;128:376-392.
133. Mitoma H, Horiuchi T, Tsukamoto H et al. Mechanisms for cytotoxic effects of anti-tumor necrosis factor agents on transmembrane tumor necrosis factor alpha-expressing cells: comparison among infliximab, etanercept, and adalimumab. *Arthritis Rheum* 2008;58:1248-1257.
134. Lügering A, Schmidt M, Lügering N et al. Infliximab Induces Apoptosis in Monocytes From Patients With Chronic Active Crohn's Disease by Using a Caspase-Dependent Pathway. *Gastroenterology* 2001;121:1145-1157.
135. Danese S, Sans M, Scaldaferrri F et al. TNF-[alpha] Blockade Down-Regulates the CD40/CD40L Pathway in the Mucosal Microcirculation: A Novel Anti-Inflammatory Mechanism of Infliximab in Crohn's Disease. *J Immunol* 2006;176:2617-2624.
136. Di Sabatino A, Pender SLF, Jackson C et al. Functional Modulation of Crohn's Disease Myofibroblasts by Anti-Tumor Necrosis Factor Antibodies. *Gastroenterology* 2007;133:137-149.
137. Hanauer SB, Feagan BG, Lichtenstein GR et al. Maintenance infliximab for Crohn's disease: the ACCENT I randomised trial. *The Lancet* 2002;359:1541-1549.
138. Colombel JF, Sandborn WJ, Reinisch W et al. Infliximab, azathioprine, or combination therapy for Crohn's disease. *N Engl J Med* 2010;362:1383-1395.
139. Sandborn WJ, Feagan BG, Stoinov S et al. Certolizumab Pegol for the Treatment of Crohn's Disease. *N Engl J Med* 2007;357:228-238.
140. Taylor KD, Plevy SE, Yang H et al. ANCA Pattern and LTA haplotype relationship to clinical responses to anti-TNF antibody treatment in crohn's disease. *Gastroenterology* 2001;120:1347-1355.

141. Sandborn WJ, Rutgeerts P, Enns R et al. Adalimumab Induction Therapy for Crohn Disease Previously Treated with Infliximab: A Randomized Trial. *Ann Intern Med* 2007;146:829-838.
142. Louis E, El GZ, Vermeire S et al. Association between polymorphism in IgG Fc receptor IIIa coding gene and biological response to infliximab in Crohn's disease. *Aliment Pharmacol Ther* 2004;19:511-519.
143. Dideberg V, Théâtre E, Farnir Fdr et al. The TNF/ADAM 17 system: implication of an ADAM 17 haplotype in the clinical response to infliximab in Crohn's disease. *Pharmacogenetics and Genomics* 2006 Oct;16(10):727-34
144. Kugathasan S, Werlin SL, Martinez A et al. Prolonged duration of response to infliximab in early but not late pediatric Crohn's disease. *Am J Gastroenterol* 2000;95:3189-3194.
145. Vermeire S, Louis E, Carbonez A et al. Demographic and clinical parameters influencing the short-term outcome of anti-tumor necrosis factor (infliximab) treatment in Crohn's disease. *Am J Gastroenterol* 2002;97:2357-2363.
146. Lionetti P, Bronzini F, Salvestrini C et al. Response to infliximab is related to disease duration in paediatric Crohn's disease. *Aliment Pharmacol Ther* 2003;18:425-431.
147. Arnott ID, McNeill G, Satsangi J. An analysis of factors influencing short-term and sustained response to infliximab treatment for Crohn's disease. *Aliment Pharmacol Ther* 2003;17:1451-1457.
148. Parsi MA, Achkar JP, Richardson S et al. Predictors of response to infliximab in patients with Crohn's disease. *Gastroenterology* 2002;123:707-713.
149. Schnitzler F, Fidler H, Ferrante M et al. Long-term outcome of treatment with infliximab in 614 patients with Crohn's disease: results from a single-centre cohort. *Gut* 2009;58:492-500.
150. Lemann M, Mary JY, Duclos B et al. Infliximab plus azathioprine for steroid-dependent Crohn's disease patients: a randomized placebo-controlled trial. *Gastroenterology* 2006;130:1054-1061.
151. Carmona L, Gomez-Reino JJ, Rodriguez-Valverde V et al. Effectiveness of recommendations to prevent reactivation of latent tuberculosis infection in patients treated with tumor necrosis factor antagonists. *Arthritis Rheum* 2005;52:1766-1772.
152. Gardam MA, Keystone EC, Menzies R et al. Anti-tumour necrosis factor agents and tuberculosis risk: mechanisms of action and clinical management. *Lancet Infect Dis* 2003;3:148-155.
153. Rahier JF, Ben-Horin S, Chowers Y. European evidence-based Consensus on the prevention, diagnosis and management of opportunistic infections in inflammatory bowel disease. *J Crohn's and colitis* (2009), doi:10.1016/j.crohns.2009.02.010
154. Sandborn WJ, Loftus EV. Balancing the risks and benefits of infliximab in the treatment of inflammatory bowel disease. *Gut* 2004;53:780-782.
155. Toruner M, Loftus EV, Jr., Harmsen WS et al. Risk factors for opportunistic infections in patients with inflammatory bowel disease. *Gastroenterology* 2008;134:929-936.
156. Lichtenstein GR, Feagan BG, Cohen RD et al. Serious Infections and Mortality in Association With Therapies for Crohn's Disease: TREAT Registry. *Clinical Gastroenterology and Hepatology* 2006;4:621-630.
157. Lewis JD, Bilker WB, Brensinger C et al. Inflammatory bowel disease is not associated with an increased risk of lymphoma. *Gastroenterology* 2001;121:1080-1087.
158. Askling J, Brandt L, Lapidus A et al. Risk of haematopoietic cancer in patients with inflammatory bowel disease. *Gut* 2005;54:617-622.
159. Persson PG, Karlen P, Bernell O et al. Crohn's disease and cancer: a population-based cohort study. *Gastroenterology* 1994;107:1675-1679.

160. Loftus EV, Jr, Tremaine WJ, Habermann TM et al. Risk of lymphoma in inflammatory bowel disease. *Am J Gastroenterol* 2000;95:2308-2312.
161. Kandiel A, Fraser AG, Korelitz BI et al. Increased risk of lymphoma among inflammatory bowel disease patients treated with azathioprine and 6-mercaptopurine. *Gut* 2005;54:1121-1125.
162. Farrell RJ, Ang Y, Kileen P et al. Increased incidence of non-Hodgkin's lymphoma in inflammatory bowel disease patients on immunosuppressive therapy but overall risk is low. *Gut* 2000;47:514-519.
163. Burger DC, Florin TH. Hepatosplenic T-cell lymphoma following infliximab therapy for Crohn's disease. *Med J Aust* 2009;190:341-342.
164. Mackey ACR, Green LP, Leptak CM et al. Hepatosplenic T Cell Lymphoma Associated with Infliximab Use in Young Patients Treated for Inflammatory Bowel Disease: Update. [Letter]. *Journal of Pediatric Gastroenterology & Nutrition* 2009;48:386-388.
165. Ochenrider MG, Patterson DJ, Aboulaifa DM. Hepatosplenic T-cell lymphoma in a young man with Crohn's disease: case report and literature review. *Clin Lymphoma Myeloma Leuk* 2010;10:144-148.
166. Shale M, Kanfer E, Panaccione R et al. Hepatosplenic T cell lymphoma in inflammatory bowel disease. *Gut* 2008;57:1639-1641.
167. Oren R, Arber N, Odes S et al. Methotrexate in chronic active ulcerative colitis: a double-blind, randomized, Israeli multicenter trial. *Gastroenterology* 1996;110:1416-1421.
168. Feagan BG, Rochon J, Fedorak RN et al. Methotrexate for the treatment of Crohn's disease. The North American Crohn's Study Group Investigators. *N Engl J Med* 1995;332:292-297.
169. Fraser AG. Methotrexate: first-line or second-line immunomodulator? *Eur J Gastroenterol Hepatol* 2003;15:225-231.
170. Krampera M, Glennie S, Dyson J et al. Bone marrow mesenchymal stem cells inhibit the response of naive and memory antigen-specific T cells to their cognate peptide. *Blood* 2003;101:3722-3729.
171. Aggarwal S, Pittenger MF. Human mesenchymal stem cells modulate allogeneic immune cell responses. *Blood* 2005;105:1815-1822.
172. Le Blanc K, Frassoni F, Ball L et al. Mesenchymal stem cells for treatment of steroid-resistant, severe, acute graft-versus-host disease: a phase II study. *Lancet* 2008;371:1579-1586.
173. Le Blanc K, Rasmusson I, Sundberg B et al. Treatment of severe acute graft-versus-host disease with third party haploidentical mesenchymal stem cells. *Lancet* 2004;363:1439-1441.

