

Chemokines in atherosclerotic lesion development and stability : from mice to man

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1. Atherosclerosis

Cardiovascular diseases are the major cause of morbidity and mortality in western societies¹. The most common clinical manifestations are stroke and acute myocardial infarction and in both ailments atherosclerosis is the underlying culprit. In general, atherosclerosis is regarded a progressive, multi-factorial disease, already initiated during early adolescence^{2, 3}. Initially lesion progression remains at a subclinical stage due to arterial plasticity or in other words the capacity of vessels remodelling to compensate for luminal loss⁴. Depending on the site, plaque composition and affected vessel, advanced atherosclerotic lesions are prone to rupture⁵⁻⁷. Upon rupture, the highly thrombogenic content of the plaque will be exposed to the circulation, triggering blood coagulation and thrombus formation^{8, 9}. The ensuing (total) arterial occlusion can induce acute complications like cerebral ischemia (stroke), angina pectoris, peripheral arterial occlusive disease and myocardial infarction and might eventually lead to death.

 Several locations in the vasculature appear to be pre-disposed for atherosclerosis, especially vascular segments with curves and branches, like the left anterior descending artery (LAD) of the coronary arteries, the common carotid arteries at the bifurcation and all main branching points of the aorta^{10, 11}. These site-specific effects are attributable to hemodynamic factors, such as low shear stress, oscillatory flow and turbulent flow¹². Apart from genetic and spatial predisposition to atherogenesis, several behavioural features can affect disease progression, such as smoking¹³, high fat diet¹⁴, stress^{15, 16} and physical inactivity^{17, 18}. Also hypertension^{13, 19}, hyperhomocysteinemia^{20, and physical inactivity^{17, 18}. Also hypertension^{13, 19}, hyperhomocysteinemia^{20,}} ²¹, diabetes²²⁻²⁵ and obesity^{26, 27} generate an increased risk for cardiovascluar disease. Surgical intervention by e.g. bypass surgery, percutaneous transluminal coronary angioplasty (PTCA), stenting or atherectomy is frequently required to restore obstructed blood flow, however the success rate of these interventions is often impaired by re-stenosis28.

2. Atherosclerotic Plaque Development

2.1. Plaque Initiation

Atherosclerosis mostly occurs in the medium and large sized arteries^{29, 30}. Under normal conditions the artery consists of an endothelial layer covering the media of smooth muscle cells that is framed by the internal and external elastic lamina. On the perivascular site, the artery is surrounded by adventitial tissue. As already discussed above, atherosclerosis is initiated, at predisposed sites (e.g. arterial branches or bi-furcations), by endothelial dysfunction caused by low turbulent or oscillatory shear stress in combination with the presence of atherogenic factors like high lipoproteins (VLDL, LDL) levels and hypertension: the so-called 'response to injury' theory^{31, 32}. The first crucial step in atherogenesis is the interaction of atherogenic lipoproteins with freely exposed proteoglycans just underneath the endothelial layer³³, consequently resulting in lipid accumulation. Simultaneously, endothelial cells activated by flow disturban-ces increase the expression of cellular adhesion molecules such as E- and P-selectin34, 35 on their cell surface, which mediates the rolling of monocytes to the endothelial layer (*Figure 1*). Subsequently circulating leukocytes are arrested to the vascular endothelium during rolling by cellular activation via several members of the chemokine family^{36, 37} or by interaction with cellular integrins³⁴. Firm adhesion of leukocytes is induced by integrin clustering or structural rearrangement^{38, 39,} which in turn will initiate intracellular signalling pathways thereby further strengthening cell adhesion. The final step in the process of leukocyte emigration into the subendothelial intimal area in atherosclerosis is transmigration or diapedesis. Transmigration is usually preceeded by a process currently known as crawling^{40, 41} as leukocytes scan for the most convenient site for transmigration. During extravasation, leukocytes will have to penetrate the endothelial cell barrier, the endothelial cell basement membrane and finally the pericytes. Probably the unique combination of adhesion molecules, chemokines and integrins presented in the

context of atherosclerosis results in recruitment of specific leukocyte subsets to 'coded areas' of the atherosclerotic lesion.

Figure 1: Leukocyte rolling, arrest, adherence and diapedesis through the vascular wall are mediated by adhesion molecules and chemokines. Adapted from Ley et al.214

As a result of locally produced pro-inflammatory signals, such as Macrophage Colony Stimulating Factor (M-CSF), Tumor Necrosis Factor α (TNF α), Interferon γ (IFN γ), Interleukin-1 and growth factors (like Placental Derived Growth Factor (PDGF)) the migrated leukocytes will differentiate into tissue macrophages⁴²⁻⁴⁴. The maintenance crew of the immune system: the intimal macrophages, will ingest accumulated cholesterol and modified lipoprotein particles, thereby converting into foam cells⁴⁵ (*Figure 2*). This initial, quiescent plaque is classified as a type I lesion according to the classification criteria of the American Heart Association (AHA) system⁴⁶⁻⁴⁹. Type I lesions will further accumulate lipid-laden macrophages, attract T-lymphocytes and develop into type II fatty streaks⁵⁰. Under the influence of several growth factors, as Fibroblast Growth Factor (FGF) and Tumor Growth Factor β (TGFβ), medial vascular smooth muscle cells (vSMC) will start to migrate toward the endothelial lining and consequently the lesion progresses into a type III lesion.

2.2. Plaque Progression

Type III intermediate lesions contain small extracellular lipid deposits under a layer of migrated vSMCs. This class of plaques can be regarded as the transition stage between the fatty streak and an advanced atherosclerotic lesion and are also referred to as preatheroma plaques $48, 49$. In type IV lesions the intimal lipid deposits have expanded into large a-cellular lipid pools containing a substantial amount of cholesterol crystals, due to either apoptosis or necrosis of intimal foam cells or to accumulation of infiltrated lipoprotein particles. The type IV atheroma is the first stage of an advanced atherosclerotic lesion and is distinguished by a large lesion core and by intimal capillaries that most likely originate from the vasa vasorum. Type IV atheromas can induce clinical symptoms known as angina pectoris. During further progression, even more fibroblasts and vSMCs accumulate subendothelially to produce, via interaction with extracellular matrix material like collagen and proteoglycans, a fibrous cap covering the lipid core. However due to local death signals apoptosis can occur in fibrous cap cells resulting in rupture-prone vulnerable areas in the atherosclerotic lining. The type V lesion is known as the fibro-atheroma⁵¹ and rupture preferentially occurs in this lesion type, as these lesions are biomechanically vulnerable and are constantly exposed to high blood flow forces⁵. In fact, type V lesions are subdivided into 3 subcategories, of which the first (type Va) is described above, type Vb that is highly calcified and type Vc lesion, is relatively lipid poor. In practice, type IV and V lesion are difficult to discriminate, and nowadays frequently termed as 'thick' and 'thin' fibrous cap atheroma, respectively⁵.

Figure 2: Leukocyte adherence to the endothelial layer during atherosclerotic lesion initiation (panel I). During lesion progression transmigrated monocytes differentiate into macrophages, which release pro-inflammatory cytokines resulting in accumulation of T cells in the plaque (panel II). Thinning of the fibrous cap can eventually *lead to plaque rupture (panel III). Adapted from Packard et al.215*

Ruptured lesions with an intramural or luminal thrombus or lesions containing intra-plaque haemorrhage are classified type VI atherosclerotic lesions (*Figure 2*). Type VI lesions without noticeable cap breaks are referred to as eroded^{52,53}. The various subclasses can be distinguished on the basis of three different criteria: The 'fibrous cap atheroma with erosion', which has a thick fibrous cap and a luminal thrombus but without physical lumen-plaque core interaction. Next is the 'thin fibrous cap with plaque rupture', where a luminal thrombus is in direct contact with the lipid core of the lesion. The third subtype describes the 'calcified nodule with erosion', with an eruptive nodular calcification with overlying luminal thrombus.

3. The Unstable Plaque

All plaques that have progressed beyond type IV are considered 'unstable' and are accountable for the majority of clinical manifestations as stroke and myocardial infarction. In the pathobiology of atherosclerosis there is a delicate balance between necro-tic core size and fibrous cap rigidity. Disturbance of this delicate balance may lead to fibrous cap rupture. Due to rupture the highly thrombogenic lipid core will come in immediate contact with the circulation leading to coagulation, thrombus formation and finally results in acute cardiovascular syndromes or stroke. Several mechanisms have been implicated in the induction or acceleration of plaque destabilization. In particular, extra cellular matrix degradation⁵⁴⁻⁵⁶, vascular wall cell apoptosis, intimal macrophage apoptosis^{54,} $57-59$ and platelets adherence^{60, 61} are regarded to be key regulators of sta-bility. In turn many of these processes are influenced by the local inflammatory status. Atherosclerotic plaques and in particular the unstable plaque, contains many leukocyte subsets that can induce various pro-inflammatory interleukins, cytokines, chemokines⁶²⁻⁶⁴, which in turn regulate leukocyte homeostasis.

4. Leukocyte Homeostasis in Atherosclerosis

4.1. Monocytes/Macrophages

At present time atherosclerosis is broadly accepted as a lipid driven process with features of a chronic inflammatory disease. The first notion for this was provided by pathology studies revealing the presence of inflammatory infiltrates in atherosclerotic l esions 65.67 . One of the major effector cells in the initiation and progression of atherosclerosis is the macrophage. Macrophages are part of the innate immune system and normally function in immediate host defence against pathogens. Plaque macrophages engulf lipid particles, such as oxidized LDL (oxLDL), via several scavenger receptors (e.g. SR-A, SR-B1, CD36, CD68 and CXCL16)68-73. Uptake of oxLDL results in cellular activation and differentiation and as lipid particles further accumulate macrophages gradu-

ally transform into foam cells (Figure 3A).

Once activated intimal macrophages produce and release a broad panel of proinflammatory cytokines and growth factors. These soluble mediators can either influence the endothelial lining of the vascular wall or they can further stimulate foam cell formation, macrophage activation or T cell stimulation (Figure 3B). For instance IL1α, TNF α and IFN v^{74-76} have been implicated in the induction of adhesion molecules and chemokines, particularly MCP-1 (CCL2), IL-8 (CXCL8) and Fractalkine (CX3CL1)⁷⁷⁻⁸⁰, on the vascular wall thereby promoting further cellular infiltration in the plaque. Furthermore some cytokines can influence foam cell formation either by attenuation or augmentation of lipid uptake. For instance IFN γ can inhibit scavenger receptor expression^{81, 82} thereby influencing cholesterol uptake, while on the other hand it also attenuates cholesterol efflux $^{\rm 83}$. Conversely TGFβ was seen to inhibit scavenger receptor activity in human macrophages84. Although macrophages are not specialized in antigen presentation they are capable of presenting antigen on their cell surface. Oxidized LDL particles are processed by macrophages leading to oxLDL peptide presentation on MHC-II molecules and subsequent T cel activation⁸⁵. Indeed blockade of MHC-II molecules resulted in decreased activation and proliferation of oxLDL specific T cells⁸⁶.

4.2. Lymphocytes

Already a few decades ago Jonasson *et al.* demonstrated the precensce of T cells in human atherosclerotic plaques 87 . The majority of these T cells are of the CD4⁺ subtype 88 . Ablation of CD4⁺ T cells⁸⁹⁻⁹¹ attenuated lesion formation in LDLr^{-/-} mice, while adoptive transfer of CD4* T cells to immune deprived B and T cell deficient RAG^{-/-} mice accelerated atherogenesis⁸⁹. Furthermore it has become clear that T cell activation predominantly occurs during progression of atherosclerosis, but it is virtually absent during initiation 92 . Cytotoxic CD8* T cells have also been identified in atherosclerosis, but their role in the disease progress is not unambiguous⁹³⁻⁹⁵. Classically CD4⁺ helper T cells are categorized into two subclasses based on their cytokine profile. The T helper 1 (Th1) subset is classified as an inducer of cellular immunity, while the T helper 2 (Th2) subset induce a humoral response^{96, 97}, with regards to atherosclerosis these subsets are considered pro- and anti-inflammatory respectively. Under normal conditions the balance between these two T cells subsets is static, while during episodes of inflammation this balance is polarized towards Th1 or Th2. Atherosclerosis has been identified as a pro-inflammatory Th1 driven disease 98 (Figure 3C). Conceivably modulation of the immune response towards Th2 might favourably influence atherogenesis or plaque stability. In keeping with this conception, immunization of mice against MDA-LDL or ox-LDL attenuated atherosclerotic lesion formation or neo-intima formation respectively^{99,} 100. Interestingly recent studies revealed that protein vaccination might prove a useful strategy to prevent atherosclerosis. For instance protein vaccination against interleukin-12 and VEGF-RII both inhibit atherogenesis¹⁰¹,¹⁰². Moreover tolerance induction to oxidized LDL was shown to emeliorate atherogenesis, mainly due to increased levels of regulatory T cells¹⁰³.

Already in the late seventies an immune suppressive T cell was identified 104 , however scientific attention was quenched during the 90s due to lack of suitable markers. Recently this cell type recurred in science as the regulatory T cell. Regulatory T cells (Treg) represent a novel subset of CD4+ T cells with the capacity to regulate local immune responses105, 106. Currently 3 different subsets of Treg cells have been identi- -ied. First naturally occurring Tregs, which derive from thymic selection and are distinguished as CD4+, CD25 $^{\rm high}$, FoxP3 expressing and TGFβ responsive cells $^{\rm 107\text{-}109}$. Second antigen specific inducible Tregs have been described, the so-called Tr-1 cells, whose major effector protein is interleukin $10^{110,111}$. The third identified subset of Tregs, Th3 cells are antigen non-specific and exert their regulatory potential as bystander inhibitors, mainly via TGF $\beta^{110,\,111}$. Also within the field of vascular biology regulatory T cells play an important role. Induction of ovalbumin specific Tr1 cells was shown to attenu-

ate atherosclerosis development in apolipoprotein E (ApoE) deficient mice. Adoptive transfer of these Tr1 cells resulted in decreased T cell and macrophage numbers and increased interleukin-10 expression within the atherosclerotic lesion¹¹². Moreover, combined hematopoietic deficiency of co-stimulatory molecules CD80 and CD86 resulted in decreased CD4+ CD25high naturally occurring Treg and subsequent acceleration of atherogenesis. Additionally, transfer of Treg depleted splenocytes induced atherosclerotic lesion in T cell deficient mice, whereas adoptive transfer of naturally occurring Tregs attenuated atherogenesis 113 .

Figure 3: Modification of intimal LDL cholesterol and subsequent uptake by macrophages (A). Monocyte *differentiation and release of pro-in-lammaroty cytokines from macrophages (B) results in in-lammtory cell accumulation (C). Adapted from Hansson63*

4.3. Mast Cells

Mast cells (MC) are part of the innate immune system and are notorious for their role in allergy and asthma. MCs are large granular cells with the unique ability to actively release their granules into the surrounding tissue. Already in 1878, MCs were identi- -ied by Paul Ehrlich who believed that this curious cell type contained nutrients for its neighbouring cells^{114, 115}. He therefore termed these 'fertilizing' cells Mastzellen, which resulted in the english term Mast cells. MCs originate from CD34+ progenitors in the bone marrow under control of interleukin-3 and Stem Cell Factor and are released into the circulation in an immature form $116-118$. These immature MCs migrate into different tissues were they fully maturate into either mucosal or connective tissue mast cells depending on their surroundings^{119, 120}. MCs express the high affinity IgE receptor, FcɛRI, via which they usually are activated during episodes of allergies or asthma¹²¹⁻¹²⁴. Binding of multiple IgE molecules leads to cross linking of the FcεRI and subsequent cellular activation resulting in the release of MC granules $^{125\text{-}128}.$ Next to this classical activation of MCs it was recently shown that activation can also occur by binding of immunoglobulin Light Chain (IgLC) to a currently unkown receptor on the MCs¹²⁹. Neurogenic stimulation by for instance substance \tilde{P} can also lead to activation and degranulation of MCs, while several inflammatory stimuli (e.g. TNF and IL-1) and complement factors (e.g. C3a and C5a) act similarly. MC granules contain a plethora of proteases (histamine, chymase, tryptase), cytokines (TNFa, IFNg, IL-2, IL13, IL15), chemokines (CCL2, CCL3, CCL4, CCL5, CKCL10) but also growth factors (SCF, VEGF, TGF B)¹³⁰⁻¹³⁵.

Relevant to atherosclerosis MCs have been identified in the shoulder region of human plaques and they were associated with plaque rupture^{136, 137}. Interestingly, acti-

vated mast cells were found to be abundantly present also in the adventitia of atherosclerotic lesions and their number were seen to correlate with the stage of atherosclerotic plaque development and the incidence of plaque rupture138, 139. *In vitro* studies have revealed that the release of heparin proteoglycans from MCs can induce the uptake of lipids by macrophages^{140, 141}, suggesting that MCs have the potential to modulate atherogenesis. More so the MC protease chymase can effectively proteolyse Apo-A1 containing lipoproteins, thereby reducing cholesterol efflux 142 . Next to the effects on foam cell formation, heparin proteoglycans have the potential to inhibit SMC proliferation, while MC derived chymase can provoke SMC apoptosis and induce matrix degradation¹⁴³⁻¹⁴⁶. Moreover MC can release angiogenic factors (e.g. VEGF and bFGF) and therefore have been implied in plaque neovascularisation147, 148. Only very recent direct experimental evidence for MC involvement in atherosclerosis was provided by use of MC deficient mice. The absence of MC diminished aortic plaque progression by 50%. The percentage of macrophages and T cells was significantly reduced, while collagen deposition was enhanced. Reconstitution of these mice with *ex vivo* cultured mast cells normalized plaque growth to that of of control MC⁺ mice. Moreover adoptive transfer of TNF α , IL6 and IFNγ deficient MC revealed a significant role for both MC derived IL6 and IFNγ in lesion progression, while MC derived TNF α does not influence lesion development 149 . Clearly these findings provide evidence for a key role of the MCs in atherogenesis (*Figure 4*).

Adventitia

Figure 4: Activation of adventitial and intimal MCs results in plaque destabilization as a result of increased intimal apoptosis, matrix degradation and erythrocyte extravasation. Adapted from Libby et al.216

4.4. Chemokines

Chemokines are members of the cytokine family, with strong chemotactic capacity^{150,} ¹⁵¹. Chemokines and their receptors have conventionally been divided into four families on the basis of the structural arrangement of the N-terminal conserved cystein residues (CXC, CC, C and CX3C). Next to their structural classification, chemokines can also be

functionally classified as being either homeostatic or inflammatory chemokines. Homeostatic chemokines are constitutively expressed and regulate leukocyte navigation during immune surveillance. However, the vast majority of chemokines are inducible and regulate cellular recruitment especially to sites of inflammation¹⁵². Chemokines are soluble proteins that can be released from many inflammatory cell types including, endothelial cells, platelets, MCs, macrophages and lymphocytes^{134, 153-155}. Chemokines contain one to three disulfide bonds, with the exception of CX3CL1 and CXCL16, which contain a membrane-anchored mucin stalk156, 157. Chemokines characteristically fold to a structure that consists of an N-terminal domain, a three-stranded β-sheet and a C-terminal helix. Chemokines bind to dedicated receptors of the G Protein Coupled Receptor (GPCR) family of 7 transmembrane receptors coupled to heterotrimeric G-proteins and ligand binding generally induces Gi mediated calcium release and subsequent activation of downstream signalling cascades 158, 159. A detailed overview of chemokine participation in the pathology of cardiovascular disease and possible treatment strategies is provided in chapter 2.

4.5. G Protein Coupled Receptor Kinases

The activity of most GPCRs is regulated not only at the level of receptor expression but also at a functional level. An important mechanism for controlling receptor activity involves receptor desensitization, which dampens the response to prolonged or repeated stimuli^{160, 161}. Desensitization occurs within seconds after receptor stimulation and is primarily mediated by uncoupling of the GPCR from associated \tilde{G} -proteins^{162, 163}. Dedicated GPCR kinases (GRKs) can induce receptor desensitization by phosphorylation of the ligand occupied receptor, which subsequently enhances its affinity for cytosolic inhibitor proteins, so-called arrestin family members. Binding of arrestins to the phosphorylated receptor results in uncoupling and internalization of the receptor 164, ¹⁶⁵. Currently the GRK family consist of 7 ubiquitously expressed serine/threonine kinase members¹⁶³. The GRK family has been categorized into three subfamilies based on functional and structural similarities: (1) rhodopsin kinases (GRK1 and GRK-7) (2) βadrenergic receptor kinases (GRK2 and GRK3) and (3) GRK4 like kinases (GRK4, GRK5 and $GRK6$ ¹⁶².

While several of the GRK family members have been implicated in human pathology, GRK2 has been most frequently related to cardiovascular diseases. GRK2 contributes to chronic heart failure by desensitization of the β1-adrenergic receptor, resulting in loss of cardiac contractility^{166, 167}. Furthermore GRK2 was shown to influence vascular resistance and induce hypertension by inhibiting β-adrenergic agonist stimulation¹⁶⁸⁻¹⁷⁰. Transgenic mice with vSMC specific overexpression of GRK2 have an attenuated vasodilatory response to beta adrenergic stimuli¹⁷¹. Moreover GRK2 can affect hypertension by regulation of epithelial Na * channels activity 172 and by impairment of endothelial cell nitric oxide synthase (eNOS) activity¹⁷³. GRKs also regulate inflammatory responses that may be relevant to atherosclerosis. Indeed patients suffering from rheumatoid arthritis, an inflammatory disease which shares many features with atherosclerosis, were shown to have decreased GRK2 levels. Rheumatoid arthritis specific cytokines (IFNγ, interleukin-6) are able to decrease GRK2 synthesis^{174, 175}. In a rat model of experimental arthritis GRK activity was significantly down regulated during disease manifestation, where it was most evident in B- and CD4⁺ T cells¹⁷⁶. Moreover GRK2 protein levels were demonstrated to be reduced in patients with both active and secondary progressive multiple sclerosis (MS) and interestingly the decrease in protein level was similar during remission. Initiation of experimental MS was similar in both wild-type and GRK2^{+/-} mice. However GRK2^{+/-} mice only developed an acute phase of the disease, which was accompanied by massive influx of both T cells and macrophages, without any episodes of relapse¹⁷⁷. Collectively these findings indicate that *in* vivo inflammation induces tissue- and immune cell-specific downregulation of various GRKs. Conceivably, downregulation of GRKs might result in excessive cellular migration towards inflammatory sites like the atherosclerotic plaque, possibly influencing plaque

progression or stability, hence rendering GRK modulation an intriguing target for the treatment of atherosclerosis.

5. Adventitial Inflammation

The adventitia, or perivasuclar tissue is increasingly recognized as an important substrate in atherosclerosis research. The adventitia consists of extracellular matrix material, a network of capillary blood vessels (vasa vasorum), sensory nerves, tertiary lymphoid structures, fibroblasts, progenitor cells and also inflammatory cells^{139, 178-180}. During progression of atherosclerotic lesion development the adventitia expands and adventitial inflammation is gradually enhanced¹⁸¹⁻¹⁸³. Moreover expression of cytokines in aortic adventitia was shown to be associated with advanced atherosclerosis¹⁸⁴. The adventitia of ruptured lesions was shown to contain significantly more inflammatory cells, such as monocytes, T-lymphocytes and MCs138, 182, 185, 186, than that of non-ruptured lesions. Illustratively, expression of the chemokine MCP-1 and its receptor CCR2 are reported to be abundantly present on adventitial macrophages during early atherogenesis. Interestingly while MCP-1 was also expressed in intimal macrophages, the expression of CCR2 appeared specific for adventitial macrophages¹⁸⁷. In culprit lesions, significantly more CD4⁺ and CD8⁺ lymphocytes were observed at the adventitial rim, accompanied by an increased amount of capillaries, compared to stable athero-sclerotic lesions¹⁸⁸. Furthermore the adventitia, rather than the media, was suggested to be the major source of myofibroblast proliferation after balloon angioplasty and thereby implicated in re-stenosis as well 189 . Adventitial fibroblast were also shown to be activated during atherogenesis in ApoE^{-/-} mice before the formation of intimal lesions^{190, 191}. Post mortem examination of human atherosclerotic lesions provided evidence that ruptured lesions displayed enhanced adventitial inflammation, accompanied by increased elastic lamina breaks¹⁸². Finally adventitial innervation has been proposed as the link between diabetes, smoking, exercise or aging and atherosclerosis, all as a result of dysfunctional autonomic adventitial innervation 1^{192}

The vasa vasorum, a network of adventitial capillaries, is increasingly recognized as an important factor in atherosclerotic lesion development, as it is a major source of intimal neovessels¹⁹³⁻¹⁹⁶. Although luminal infiltration of neovessels may occur as well, this particularly occurs at earlier stages of lesion formation. The exact mechanism of neovessel formation from the vasa vasorum into the plaque is only poorly understood¹⁹⁷. Possibly, intimal hypoxia and ischemia may induce the expression of Hypoxia-Inducible Factor $(HIF-1)^{198}$, which in turn upregulates the expression of Vascular Endothelial Growth Factor (VEGF) and other angiogenic factors by inflammatory cells of the vasa vasorum. Additionally, activated macrophages, particularly in the inner core of the atheroma, stimulate the angiogenic system by inducing endothelial cell secretion of basic Fibroblast Growth Factor (bFGF) and VEGF197, which further induce endothelial cell proliferation. Moreover adventitial MCs can release a whole set of angiogenic factors such as Histamine, IL-8 and VEGF upon receptor mediated activati on, thereby possibly regulating angiogenesis from the adventitia toward the plaque intima199-201. *In vivo* models have revealed that vasa vasorum neovascularisation is correlated to aortic plaque progression in both ApoE and LDL receptor knock out mice^{202, 203.} The vasa vasorum also represents an alternative entry point for inflammatory cells and plasma constituents into the plaque, which is critical in plaque progression²⁰⁴. In post mortem studies, hyperplasia of the vasa vasorum and the consequential macrophage infiltration were found to be associated with plaque rupture $^{205}\!$. Currently, a high density of vasa vasorum is considered as one of the determinants of a "vulnerable plaque"194.

Angiogenesis and ensuing adventitial vasa vasorum neovascularization of the intima may predispose to intraplaque hemorrhage (IPH), which has been associated with plaque instability¹⁹⁶. Kolodgie and colleagues²⁰⁶ have recently provided compelling evidence that intraplaque hemorrhage often colocalizes with leaky microvessels and may significantly contribute to the expansion of the lipid core. Extravasated erythrocytes form a rich source of free cholesterol, which will be deposited in the core, thereby unbalancing the equilibrium between lipid core size and cap thickness. Moreover excess cholesterol that is taken up by macrophages may induce apoptosis of these cells²⁰⁷. Finally, IPH will increase macrophage infiltration, platelet deposition and foam cell formation, all factors that destabilize plaques. Macrophage apoptosis will be accompanied by enhanced TF activity in the plaque which in turn increases VEGF expression and angiogenesis, thus creating a self-perpetuating circuit. In patients with peripheral artery disease, both VEGF and TF levels were significantly increased and the expression of both factors appeared to be interrelated, suggesting a direct link between thrombosis and angiogenesis¹⁹⁵. Focal inhibition of angiogenesis could results in reduced vasa vasorum development and decreased plaque formation²⁰⁸.

6. Study Aims

The most common clinical manifestations of cardiovascular disorders, stroke and acute myocardial infarction, are a result of atherosclerotic plaque rupture and subsequent thrombosis. As chemokines are generally considered key regulators of leukocyte transmigration into the vessel wall, we anticipated that specific chemokines might have a distinctive role in leukocyte homeostasis at specific stages of atherosclerotic disease progression and during ischemia-reperfusion injury. We also suggest that patient specific local regulation of leukocyte homeostasis by means of modulated chemokine-directed leukocyte migration might therapeutically modulate atherosclerotic plaque progression and stability and additionally could improve tissue recovery after ischemic injury. In this thesis, the first aim was to identify cardiovascular disease specific chemokine or chemokine pattern regulation in humans by use of multiplex chemokine analysis. Secondly we aimed to mechanistically validate the chemokine markers obtained from the human profiling studies and attenuate lesion progression by modulation of several other chemokine targets *in vivo*. Finally, we aimed to integrate human and mouse studies in order to identify new chemokine targets or chemokine patterns for future therapeutic intervention.

7. Thesis Outline

Prevention of clinical complications as myocardial infarction or stroke due to plaque rupture in unstable angina pectoris patients is likely to result in a decreased cardiovascular death rate in the Western Society. Currently angina pectoris treatment is mainly focused on anti-coagulants and cholesterol lowering, often followed by invasive treatments such as primary percutaneous intervention (PCI). To prevent patient hospitalization and invasive surgery, strategies to attenuate plaque progression and even so improve plaque stability of an atherosclerotic lesion could offer a suitable therapeutic alternative.

In this thesis, it was aimed to improve plaque stability by modulation of the local and/or systemic leukocyte homeostasis. **Chapter 2** provides an overview on the current status of chemokine research in several cardiovascular disorders. Furthermore possible treatment strategies are suggested. In **Chapters 3 and 4**, patient material from two different cohorts comprised of acute myocardial infarction patients (MISSION!) and angina pectoris patients (APRAIS) was analyzed for chemokine distribution patterns by use of a multiplex immuno assay. The chemokines CCL3 (MIP1 α), CCL5 (RANTES) and CCL18 (PARC) emerged as the most promising therapeutic targets. In **Chapter 5** one of the targets obtained from the human profiling, CCL3 was studied for its contribution to atherogenesis. CCL3 is an inducible inflammatory chemokine also known as Macrophage Inflammatory Protein-1 α (MIP-1 α), which is higly expressed by macrophages, lymphocytes and MCs. CCL3 binds to three different chemokine receptors, CCR1, CCR3 and CCR5 and it can form dimers with CCL4. In this study we pursued a bone marrow transplant (BMT) approach where wild type bone marrow was replaced by CCL3 defi-

cient bone marrow in $LDLr^{-/-}$ recipients, thereby inducing leukocyte specific CCL3 knock out. After a full engraftment of the bone marrow the animals received a high fat, high cholesterol diet after which lesion formation and composition in the aortic leaflet area was determined. In **Chapter 6** a CXCR3 antagonist (NBI-74330) was used to establish the role of CXCR3 expressing leukocytes in atherosclerosis. LDL $r^{/-}$ mice were treated daily with NBI-74330 during the entire experiment and the effect of CXCR3 blockage on both collar induced and 'natural' atherosclerosis was evaluated. In **Chapter 7** the TGFβ family member Macrophage Inhibitory Cytokine-1 (MIC-1) was studied for its role in early atherogenesis and plaque stability. MIC-1 was first implicated in atherosclerosis in patient studies, where it was shown to be an independent risk factor for acute coronary syndromes^{209, 210}. Furthermore MIC-1 was shown to co-localize with intimal macrophages and is a potent inducer of p53 mediated apoptosis^{211, 212}, thereby rendering this growth factor a detrimental player in atherosclerosis. For this study we used the BMT strategy to study the effects of leukocyte specific MIC-1 deficiency on atherogenesis. In **Chapter 8** we studied the effect of chemokine receptor desensitization on atherogenesis and plaque stability. Receptor desensitization, by for instance GRKs, is an important mechanism for controlling receptor activity, which dampens the response to prolonged or repeated stimuli^{12, 13}. GRK2 has been frequently related to cardiovascular diseases. For instance, GRK2 contributes to chronic heart failure by desensitization of the β1 adrenergic receptor, resulting in loss of cardiac contractility^{18, 19}. Furthermore GRK2 was shown to influence vascular resistance and induce hypertension by inhibiting βadrenergic agonist stimulation²⁰⁻²². For this study we used the BMT strategy to study the effects of leukocyte specific partial GRK2 deficiency on atherogenesis and plaque morphology. In **Chapter 9** we have studied an inflammatory cell type, the mast cell, which, in human plaque rupture, has been shown to be abundantly present in the adventitia of affected arteries. Although the MC content of the adventitia was linked to the severity of disease, it remained unclear whether adventitial mast cells causally contribute to or are recruited in response to plaque rupture. In this study, we have persued a novel adapted delayed type hypersensitivity approach to attract MCs to the adventitia of carotid artery lesions in ApoE-/- mice and evaluated its impact on plaque morphology. **Chapter 10** represents a detour to the field of immuno haematology as we have by serendipity found that CCR7 may be critical to the development of Graft versus Host Disease (GvHD). Lethally irradiated animals were transplanted with either WT or $CCR7$ ^{-/-} bone marrow. After approximately 5 weeks post transplantation the CCR7-/- recipients started to show signs of chronic GvHD. In an additional experiment we have attempted to rescue $CCR7^{-/-}$ recipients from chronic GvHD by dilution of CCR7 deficiency. Finally, **Chapter 11** provides a discussion of the most relevant findings of this thesis and offers an overview of future perspectives the therapeutic implications.

References

- Lopez AD, Mathers CD, Ezzati M, et al. Global and regional burden of disease and risk factors, 2001: systematic analysis of population health data. *Lancet*. 2006;367(9524):1747-1757. 1.
- Stary HC. Macrophage foam cells in the coronary artery intima of human infants. *Ann N Y Acad Sci*. 1985;454:5-8. 2.
- Stary HC. Evolution and progression of atherosclerotic lesions in coronary arteries of children and young adults. *Arteriosclerosis*. 1989;9(1 Suppl):I19-32. 3.
- Pasterkamp G, Wensing PJ, Post MJ, et al. Paradoxical arterial wall shrinkage may contribute to luminal narrowing of human atherosclerotic femoral arteries. *Circulation.* 1995;91(5):1444-1449. 4.
- Virmani R, Kolodgie FD, Burke AP, et al. Lessons from sudden coronary death: a comprehensive morphological classification scheme for atherosclerotic lesions. *Arterioscler Thromb Vasc Biol*. 2000;20(5):1262-1275. 5.
- Libby P. Molecular bases of the acute coronary syndromes. *Circulation*. 1995;91(11):2844-2850. 6.
- Lee RT, Libby P. The unstable atheroma. *Arterioscler Thromb Vasc Biol.* 1997;17(10):1859-1867. 7.
- Davies MJ, Richardson PD, Woolf N, et al. Risk of thrombosis in human atherosclerotic plaques: role of extracellular lipid, macrophage, and smooth muscle cell content. *Br Heart J.* 1993;69(5):377-381. 8.
- Shah PK. Plaque disruption and coronary thrombosis: new insight into pathogenesis and prevention. *Clin Cardiol.* 1997;20(11 Suppl 2):II-38-44. 9.
- 10. Caro CG, Fitz-Gerald JM, Schroter RC. Arterial wall shear and distribution of early atheroma in man. *Nature*. 1969;223(5211):1159-1160.
- Zarins CK, Giddens DP, Bharadvaj BK, et al. Carotid bifurcation atherosclerosis. Quantitative correlation of plaque localization with flow velocity profiles and wall shear stress. *Circ Res.* 1983;53(4):502-514. 11.
- VanderLaan PA, Reardon CA, Getz GS. Site specificity of atherosclerosis: site-selective responses to atherosclerotic modulators. *Arterioscler Thromb Vasc Biol*. 2004;24(1):12-22. 12.
- Glasser SP, Selwyn AP, Ganz P. Atherosclerosis: risk factors and the vascular endothelium. *Am Heart J.* 1996;131(2):379-384. 13.
- Kritchevsky D. Diet and atherosclerosis. *Am Heart J.* 1999;138(5 Pt 2):S426-430. 14.
- Everson-Rose SA, Lewis TT. Psychosocial factors and cardiovascular diseases. *Annu Rev Public Health.* 15. 2005;26:469-500.
- 16. Hauss WH, Bauch HJ, Schulte H. Adrenaline and noradrenaline as possible chemical mediators in the pathogenesis of arteriosclerosis. *Ann N Y Acad Sci.* 1990;598:91-101.
- Kamphuis MH, Geerlings MI, Tijhuis MA, et al. Physical inactivity, depression, and risk of cardiovascular mortality. *Med Sci Sports Exerc*. 2007;39(10):1693-1699. 17.
- Kadoglou NP, Iliadis F, Liapis CD. Exercise and carotid atherosclerosis. *Eur J Vasc Endovasc Surg.* 2008;35(3):264-272. 18.
- Chobanian AV. Hypertension, antihypertensive drugs, and atherogenesis. Mechanisms and clinical impli-19. Chobanian AV. Hypertension, antihypertensive drugs,
cations. *J Clin Hypertens*. 1986;2(3 Suppl):148S-157S.
- 20. Clarke R, Fitzgerald D, O'Brien C, et al. Hyperhomocysteinaemia: a risk factor for extracranial carotid artery atherosclerosis. *Ir J Med Sci.* 1992;161(3):61-65.
- 21. Nygard O, Nordrehaug JE, Refsum H, et al. Plasma homocysteine levels and mortality in patients with coronary artery disease. *N Engl J Med.* 1997;337(4):230-236.
- Steiner G. Diabetes and atherosclerosis: an overview. *Diabetes.* 1981;30(Suppl 2):1-7. 22.
- Criqui MH. Epidemiology of atherosclerosis: an updated overview. *Am J Cardiol.* 1986;57(5):18C-23C. 23.
- 24. Ordovas JM. Genetic links between diabetes mellitus and coronary atherosclerosis. *Curr Atheroscler Rep.* 2007;9(3):204-210.
- Dominiczak MH. Obesity, glucose intolerance and diabetes and their links to cardiovascular disease. Implications for laboratory medicine. *Clin Chem Lab Med.* 2003;41(9):1266-1278. 25.
- Shively CA, Clarkson TB. Regional obesity and coronary artery atherosclerosis in females: a non-human primate model. *Acta Med Scand Suppl.* 1988;723:71-78. 26.
- Egan BM, Bassett DR, Block WD. Comparative effects of overweight on cardiovascular risk in younger versus older men. *Am J Cardiol.* 1991;67(4):248-252. 27.
- Dangas G, Fuster V. Management of restenosis after coronary intervention. *Am Heart J.* 1996;132 (2 Pt 1):428-436. 28.
- Lusis AJ. Atherosclerosis. *Nature.* 2000;407(6801):233-241. 29.
- Ross R. Atherosclerosis--an inflammatory disease. *N Engl J Med.* 1999;340(2):115-126.
- 30. Ross R. Atherosclerosis--an inflammatory disease. *N Engl J Med.* 1999;340(2):115-126.
31. Ross R, Glomset J, Harker L. Response to injury and atherogenesis. *Am J Pathol.* 1977;86(3):675-684. 31.
- 32. Ross R, Glomset JA. Atherosclerosis and the arterial smooth muscle cell: Proliferation of smooth
- muscle is a key event in the genesis of the lesions of atherosclerosis. *Science.* 1973;180(93):1332-1339. Camejo G, Hurt-Camejo E, Wiklund O, et al. Association of apo B lipoproteins with arterial proteoglycans: pathological significance and molecular basis. *Atherosclerosis*. 1998;139(2):205-222. 33.
- Hakkert BC, Kuijpers TW, Leeuwenberg JF, et al. Neutrophil and monocyte adherence to and migration across monolayers of cytokine-activated endothelial cells: the contribution of CD18, ELAM-1, and VLA-4. *Blood*. 1991;78(10):2721-2726. 34.
- Murphy JF, Bordet JC, Wyler B, et al. The vitronectin receptor (alpha v beta 3) is implicated, in cooperation with P-selectin and platelet-activating factor, in the adhesion of monocytes to activated endothelial cells. *Biochem J*. 1994;304 (Pt 2):537-542. 35.
- Weber KS, Nelson PJ, Grone HJ, et al. Expression of CCR2 by endothelial cells : implications for MCP-1 mediated wound injury repair and In vivo inflammatory activation of endothelium. *Arterioscler Thromb Vasc Biol.* 1999;19(9):2085-2093. 36.
- Umehara H, Imai T. Role of fractalkine in leukocyte adhesion and migration and in vascular injury. *Drug* 37.

News Perspect. 2001;14(8):460-464.

- Sigal A, Bleijs DA, Grabovsky V, et al. The LFA-1 integrin supports rolling adhesions on ICAM-1 under physiological shear flow in a permissive cellular environment. *J Immunol*. 2000;165(1):442-452. 38.
- Grabovsky V, Feigelson S, Chen C, et al. Subsecond induction of alpha4 integrin clustering by immobilized chemokines stimulates leukocyte tethering and rolling on endothelial vascular cell adhesion molecule 1 under flow conditions. *J Exp Med.* 2000;192(4):495-506. 39.
- Schenkel AR, Mamdouh Z, Muller WA. Locomotion of monocytes on endothelium is a critical step during extravasation. *Nat Immunol.* 2004;5(4):393-400. 40.
- Phillipson M, Heit B, Colarusso P, et al. Intraluminal crawling of neutrophils to emigration sites: a molec-41. ularly distinct process from adhesion in the recruitment cascade. *J Exp Med.* 2006;203(12):2569-2575.
- Watanabe Y, Inaba T, Gotoda T, et al. Role of macrophage colony-stimulating factor in the initial process of atherosclerosis. *Ann N Y Acad Sci.* 1995;748:357-364; discussion 364-356. 42.
- Libby P, Galis ZS. Cytokines regulate genes involved in atherogenesis. *Ann N Y Acad Sci*. 1995;748:158- 43. 168; discussion 168-170.
- Ross R, Raines E, Bowen-Pope D. Growth factors from platelets, monocytes, and endothelium: their role in cell proliferation. *Ann N Y Acad Sci.* 1982;397:18-24. 44.
- Aqel NM, Ball RY, Waldmann H, et al. Monocytic origin of foam cells in human atherosclerotic plaques. 45. *Atherosclerosis.* 1984;53(3):265-271.
- 46. Stary HC. Natural history and histological classification of atherosclerotic lesions: an update. *Arterioscler Thromb Vasc Biol.* 2000;20(5):1177-1178.
- 47. Stary HC, Blankenhorn DH, Chandler AB, et al. A definition of the intima of human arteries and of its atherosclerosis-prone regions. A report from the Committee on Vascular Lesions of the Council on Arteriosclerosis, American Heart Association. *Arterioscler Thromb.* 1992;12(1):120-134.
- Stary HC, Chandler AB, Dinsmore RE, et al. A definition of advanced types of atherosclerotic lesions and a histological classification of atherosclerosis. A report from the Committee on Vascular Lesions of the Council on Arteriosclerosis, American Heart Association. *Arterioscler Thromb Vasc Biol.* 1995;15(9):1512-1531. 48.
- 49. Stary HC, Chandler AB, Glagov S, et al. A definition of initial, fatty streak, and intermediate lesions of atherosclerosis. A report from the Committee on Vascular Lesions of the Council on Arteriosclerosis, American Heart Association. *Circulation.* 1994;89(5):2462-2478.
- Masuda J, Ross R. Atherogenesis during low level hypercholesterolemia in the nonhuman primate. I. Fatty streak formation. *Arteriosclerosis.* 1990;10(2):164-177. 50.
- Masuda J, Ross R. Atherogenesis during low level hypercholesterolemia in the nonhuman primate. II. Fatty streak conversion to fibrous plaque. *Arteriosclerosis.* 1990;10(2):178-187. 51.
- 52. Arbustini E, Dal Bello B, Morbini P, et al. Plaque erosion is a major substrate for coronary thrombosis in acute myocardial infarction. *Heart*. 1999;82(3):269-272.
- Virmani R, Burke AP, Farb A. Plaque rupture and plaque erosion. *Thromb Haemost.* 1999;82 Suppl 1:1- 3. 53.
- Bennett MR. Apoptosis in the cardiovascular system. *Heart.* 2002;87(5):480-487. 54.
- Newby AC. Dual role of matrix metalloproteinases (matrixins) in intimal thickening and atherosclerotic plaque rupture. *Physiol Rev.* 2005;85(1):1-31. 55.
- Newby AC, Johnson JL. Genetic strategies to elucidate the roles of matrix metalloproteinases in atherosclerotic plaque growth and stability. *Circ Res.* 2005;97(10):958-960. 56.
- 57. Kolodgie FD, Burke AP, Farb A, et al. The thin-cap fibroatheroma: a type of vulnerable plaque: the major
- precursor lesion to acute coronary syndromes. *Curr Opin Cardiol.* 2001;16(5):285-292.
Bjorkerud S, Bjorkerud B. Apoptosis is abundant in human atherosclerotic lesions, especially in inflammatory cells (macrophages and T cells), and may contribute to the accumulation of gruel and plaque instability. *Am J Pathol.* 1996;149(2):367-380. 58.
- Kockx MM, Herman AG. Apoptosis in atherogenesis: implications for plaque destabilization. *Eur Heart J.* 1998;19 Suppl G:G23-28. 59.
- 60. Marutsuka K, Hatakeyama K, Yamashita A, et al. Role of thrombogenic factors in the development of atherosclerosis. *J Atheroscler Thromb.* 2005;12(1):1-8.
- Sato Y, Hatakeyama K, Yamashita A, et al. Proportion of fibrin and platelets differs in thrombi on ruptured and eroded coronary atherosclerotic plaques in humans. *Heart.* 2005;91(4):526-530. 61.
- Ikeda U. Inflammation and coronary artery disease. *Curr Vasc Pharmacol.* 2003;1(1):65-70. 62.
- 63. Hansson GK. Inflammation, atherosclerosis, and coronary artery disease. NEngl J Med. 2005;352(16):1685-1695.
- Corti R, Hutter R, Badimon JJ, et al. Evolving concepts in the triad of atherosclerosis, inflammation and thrombosis. *J Thromb Thrombolysis.* 2004;17(1):35-44. 64.
- Pomerance A. Pathological and clinical study of calcification of the mitral valve ring. *J Clin Pathol.* 1970;23(4):354-361. 65.
- 66. Poston RN, Davies DF. Immunity and inflammation in the pathogenesis of atherosclerosis. A review. Ath*erosclerosis*. 1974;19(3):353-367.
- Parums D, Mitchinson MJ. Demonstration of immunoglobulin in the neighbourhood of advanced atherosclerotic plaques. *Atherosclerosis*. 1981;38(1-2):211-216. 67.
- Van Berkel TJ, De Rijke YB, Kruijt JK. Different fate in vivo of oxidatively modified low density lipoprotein and acetylated low density lipoprotein in rats. Recognition by various scavenger receptors on Kupffer and endothelial liver cells. *J Biol Chem.* 1991;266(4):2282-2289. 68.
- Endemann G, Stanton LW, Madden KS, et al. CD36 is a receptor for oxidized low density lipoprotein. *J Biol* 69. *Chem.* 1993;268(16):11811-11816.
- 70. De Rijke YB, Biessen EA, Vogelezang CJ, et al. Binding characteristics of scavenger receptors on liver en-
- *22*

dothelial and Kupffer cells for modified low-density lipoproteins. *Biochem J.* 1994;304 (Pt 1):69-73.

- Sakai M, Miyazaki A, Hakamata H, et al. The scavenger receptor serves as a route for internalization of lysophosphatidylcholine in oxidized low density lipoprotein-induced macrophage proliferation. *J Biol Chem.* 1 1996;271(44):27346-27352. 71.
- Ling W, Lougheed M, Suzuki H, et al. Oxidized or acetylated low density lipoproteins are rapidly cleared by the liver in mice with disruption of the scavenger receptor class A type I/II gene. *J Clin Invest.* 1997;100(2):244-252. 72.
- Minami M, Kume N, Shimaoka T, et al. Expression of SR-PSOX, a novel cell-surface scavenger receptor for phosphatidylserine and oxidized LDL in human atherosclerotic lesions. *Arterioscler Thromb Vasc Biol.* 2001;21(11):1796-1800. 73.
- Stolpen AH, Guinan EC, Fiers W, et al. Recombinant tumor necrosis factor and immune interferon act singly and in combination to reorganize human vascular endothelial cell monolayers. *Am J Pathol.* 1986;123(1):16-24. 74.
- Pober JS. Effects of tumour necrosis factor and related cytokines on vascular endothelial cells. *Ciba Found Symp.* 1987;131:170-184. 75.
- Pober JS, Gimbrone MA, Jr., Lapierre LA, et al. Overlapping patterns of activation of human endothelial cells by interleukin 1, tumor necrosis factor, and immune interferon. *J Immunol.* 1986;137(6):1893- 1896. 76.
- 77. Rollins BJ, Yoshimura T, Leonard EJ, et al. Cytokine-activated human endothelial cells synthesize and secrete a monocyte chemoattractant, MCP-1/JE. *Am J Pathol.* 1990;136(6):1229-1233.
- Brown Z, Gerritsen ME, Carley WW, et al. Chemokine gene expression and secretion by cytokine-activated human microvascular endothelial cells. Differential regulation of monocyte chemoattractant protein-1 and interleukin-8 in response to interferon-gamma. *Am J Pathol.* 1994;145(4):913-921. 78.
- Lukacs NW, Strieter RM, Elner V, et al. Production of chemokines, interleukin-8 and monocyte chemoattractant protein-1, during monocyte: endothelial cell interactions. *Blood.* 1995;86(7):2767-2773. 79.
- 80. Imaizumi T, Matsumiya T, Fujimoto K, et al. Interferon-gamma stimulates the expression of CX3CL1/fractalkine in cultured human endothelial cells. *Tohoku J Exp Med.* 2000;192(2):127-139.
- Geng YJ, Hansson GK. Interferon-gamma inhibits scavenger receptor expression and foam cell formation 81. in human monocyte-derived macrophages. *J Clin Invest.* 1992;89(4):1322-1330.
- Garner B, Baoutina A, Dean RT, et al. Regulation of serum-induced lipid accumulation in human monocyte-derived macrophages by interferon-gamma. Correlations with apolipoprotein E production, lipoprotein lipase activity and LDL receptor-related protein expression. *Atherosclerosis.* 1997;128(1):47- 58. 82.
- Panousis CG, Zuckerman SH. Regulation of cholesterol distribution in macrophage-derived foam cells by 83. interferon-gamma. *J Lipid Res.* 2000;41(1):75-83.
- Bottalico LA, Wager RE, Agellon LB, et al. Transforming growth factor-beta 1 inhibits scavenger receptor activity in THP-1 human macrophages. *J Biol Chem.* 1991;266(34):22866-22871. 84.
- Nicoletti A, Caligiuri G, Tornberg I, et al. The macrophage scavenger receptor type A directs modified proteins to antigen presentation. *Eur J Immunol.* 1999;29(2):512-521. 85.
- Huang YH, Ronnelid J, Frostegard J. Oxidized LDL induces enhanced antibody formation and MHC class II-dependent IFN-gamma production in lymphocytes from healthy individuals. *Arterioscler Thromb Vasc Biol.* 1995;15(10):1577-1583. 86.
- 87. Jonasson L, Holm J, Skalli O, et al. Regional accumulations of T cells, macrophages, and smooth muscle cells in the human atherosclerotic plaque. *Arteriosclerosis.* 1986;6(2):131-138.
- Frostegard J, Ulfgren AK, Nyberg P, et al. Cytokine expression in advanced human atherosclerotic plaques: dominance of pro-inflammatory (Th1) and macrophage-stimulating cytokines. *Atherosclerosis.* 1999;145(1):33-43. 88.
- Zhou X, Nicoletti A, Elhage R, et al. Transfer of CD4(+) T cells aggravates atherosclerosis in immunodeficient apolipoprotein E knockout mice. *Circulation.* 2000;102(24):2919-2922. 89.
- Reardon CA, Blachowicz L, White T, et al. Effect of immune deficiency on lipoproteins and atherosclerosis in male apolipoprotein E-deficient mice. *Arterioscler Thromb Vasc Biol.* 2001;21(6):1011-1016. 90.
- Song L, Leung C, Schindler C. Lymphocytes are important in early atherosclerosis. *J Clin Invest.* 2001;108(2):251-259. 91.
- 92. Khallou-Laschet J, Caligiuri G, Groyer E, et al. The proatherogenic role of T cells requires cell division and is dependent on the stage of the disease. *Arterioscler Thromb Vasc Biol.* 2006;26(2):353-358.
- Elhage R, Gourdy P, Brouchet L, et al. Deleting TCR alpha beta+ or CD4+ T lymphocytes leads to op-93. posite effects on site-specific atherosclerosis in female apolipoprotein E-deficient mice. Am J Pathol. 2004;165(6):2013-2018.
- Ludewig B, Freigang S, Jaggi M, et al. Linking immune-mediated arterial inflammation and cholesterolinduced atherosclerosis in a transgenic mouse model. *Proc Natl Acad Sci U S A.* 2000;97(23):12752- 12757. 94.
- van Wanrooij EJ, Happe H, Hauer A, et al. HIV Entry Inhibitor TAK-779 Attenuates Atherogenesis in Low-Density Lipoprotein Receptor-Deficient Mice. *Arterioscler Thromb Vasc Biol.* 2005(12):2642-2647. 95.
- Garside P, Mowat AM. Polarization of Th-cell responses: a phylogenetic consequence of nonspecific immune defence? *Immunol Today.* 1995;16(5):220-223. 96.
- Del Prete G. The concept of type-1 and type-2 helper T cells and their cytokines in humans. *Int Rev Immunol.* 1998;16(3-4):427-455. 97.
- Hansson GK. In-lammation and immune response in atherosclerosis. *Curr Atheroscler Rep.* 1999;1(2):150- 155. 98.
- George J, Afek A, Gilburd B, et al. Hyperimmunization of apo-E-deficient mice with homologous malondialdehyde low-density lipoprotein suppresses early atherogenesis. *Atherosclerosis.* 1998;138(1):147- 99.

152.

- 100. Nilsson J, Calara F, Regnstrom J, et al. Immunization with homologous oxidized low density lipoprotein reduces neointimal formation after balloon injury in hypercholesterolemic rabbits. *J Am Coll Cardiol.* 1997;30(7):1886-1891.
- 101. Hauer AD, Uyttenhove C, de Vos P, et al. Blockade of interleukin-12 function by protein vaccination attenuates atherosclerosis. *Circulation.* 2005;112(7):1054-1062.
- 102. Hauer AD, van Puijvelde GH, Peterse N, et al. Vaccination against VEGFR2 attenuates initiation and progression of atherosclerosis. *Arterioscler Thromb Vasc Biol.* 2007;27(9):2050-2057.
- 103. van Puijvelde GH, Hauer AD, de Vos P, et al. Induction of oral tolerance to oxidized low-density lipoprotein ameliorates atherosclerosis. *Circulation.* 2006;114(18):1968-1976.
- 104. Dutton RW. Suppressor T cells. *Transplant Rev.* 1975;26:39-55.
- Papiernik M, Banz A. Natural regulatory CD4 T cells expressing CD25. *Microbes Infect.* 2001;3(11):937- 105. 945.
- 106. Sakaguchi S, Sakaguchi N, Shimizu J, et al. Immunologic tolerance maintained by CD25+ CD4+ regulatory T cells: their common role in controlling autoimmunity, tumor immunity, and transplantation tolerance. *Immunol Rev.* 2001;182:18-32.
- 107. Itoh M, Takahashi T, Sakaguchi N, et al. Thymus and autoimmunity: production of CD25+CD4+ naturally anergic and suppressive \overline{T} cells as a key function of the thymus in maintaining immunologic self-tolerance. *J Immunol.* 1999;162(9):5317-5326.
- 108. Feunou P, Poulin L, Habran C, et al. CD4+CD25+ and CD4+CD25- T cells act respectively as inducer and effector T suppressor cells in superantigen-induced tolerance. *J Immunol.* 2003;171(7):3475-3484.
- 109. Zheng SG, Wang JH, Gray JD, et al. Natural and induced CD4+CD25+ cells educate CD4+CD25- cells to develop suppressive activity: the role of IL-2, TGF-beta, and IL-10. *J Immunol.* 2004;172(9):5213-5221.
- Beissert S, Schwarz A, Schwarz T. Regulatory T cells. *J Invest Dermatol.* 2006;126(1):15-24. 110.
- 111. Damoiseaux J. Regulatory T cells: back to the future. *Neth J Med.* 2006;64(1):4-9.
- 112. Mallat Z, Gojova A, Brun V, et al. Induction of a regulatory T cell type 1 response reduces the development of atherosclerosis in apolipoprotein E-knockout mice. *Circulation.* 2003;108(10):1232-1237.
- 113. Ait-Oufella H, Salomon BL, Potteaux S, et al. Natural regulatory T cells control the development of atherosclerosis in mice. *Nat Med.* 2006;12(2):178-180.
- Ehrlich P. Beitrage zur Theorie und Praxis der Histologischen Farbung. *Thesis.* 1878. 114.
- 115. Crivellato E, Beltrami C, Mallardi F, et al. Paul Ehrlich's doctoral thesis: a milestone in the study of mast cells. *Br J Haematol.* 2003;123(1):19-21.
- 116. Kirshenbaum AS, Kessler SW, Goff JP, et al. Demonstration of the origin of human mast cells from CD34+ bone marrow progenitor cells. *J Immunol.* 1991;146(5):1410-1415.
- 117. Rottem M, Kirshenbaum AS, Metcalfe DD. Early development of mast cells. *Int Arch Allergy Appl Immunol.* 1991;94(1-4):104-109.
- 118. Kirshenbaum AS, Goff JP, Kessler SW, et al. Effect of IL-3 and stem cell factor on the appearance of human basophils and mast cells from CD34+ pluripotent progenitor cells. *J Immunol.* 1992;148(3):772-777.
- 119. Kitamura Y, Kanakura Y, Fujita J, et al. Differentiation and transdifferentiation of mast cells; a unique member of the hematopoietic cell family. *Int J Cell Cloning.* 1987;5(2):108-121.
- 120. Gurish MF, Boyce JA. Mast cell growth, differentiation, and death. *Clin Rev Allergy Immunol.* 2002;22(2):107-118.
- 121. Moller G, Konig W. Binding characteristics of aggregated IgGa to rat basophilic leukaemia (RBL) cells and rat mast cells. *Immunology.* 1980;41(3):605-615.
- 122. Hogg JC, Pare PD, Boucher RC, et al. The pathophysiology of asthma. *Can Med Assoc J.* 1979;121(4):409-414.
- 123. Marquardt DL, Wasserman SI. Mast cells in allergic diseases and mastocytosis. West J Med. 1982;137(3):195-212.
- 124. Holgate ST, Hardy C, Robinson C, et al. The mast cell as a primary effector cell in the pathogenesis of asthma. *J Allergy Clin Immunol.* 1986;77(2):274-282.
- 125. Healicon RM, Foreman JC. Rat mast cell activation and inactivation: differences when various ligands are used to induce secretion. *Agents Actions.* 1985;16(3-4):155-159.
- 126. Kane P, Erickson J, Fewtrell C, et al. Cross-linking of IgE-receptor complexes at the cell surface: synthesis and characterization of a long bivalent hapten that is capable of triggering mast cells and rat basophilic leukemia cells. *Mol Immunol*. 1986;23(7):783-790.
- 127. Erickson J, Kane P, Goldstein B, et al. Cross-linking of IgE-receptor complexes at the cell surface: a fluorescence method for studying the binding of monovalent and bivalent haptens to IgE. *Mol Immunol.* 1986;23(7):769-781.
- 128. Reischl IG, Coward WR, Church MK. Molecular consequences of human mast cell activation following immunoglobulin E-high-affinity immunoglobulin E receptor (IgE-FcepsilonRI) interaction. *Biochem Pharmacol.* 1999;58(12):1841-1850.
- 129. Redegeld FA, van der Heijden MW, Kool M, et al. Immunoglobulin-free light chains elicit immediate hypersensitivity-like responses. *Nat Med.* 2002;8(7):694-701.
- 130. Schwartz LB. Mast cells: function and contents. Curr Opin Immunol. 1994;6(1):91-97.
- 131. Pejler G, Abrink M, Ringvall M, et al. Mast cell proteases. Adv Immunol. 2007;95:167-255.
- 132. Gordon JR, Burd PR, Galli SJ. Mast cells as a source of multifunctional cytokines. *Immunol Today.* 1990;11(12):458-464.
- 133. Galli SJ, Gordon JR, Wershil BK. Mast cell cytokines in allergy and inflammation. Agents Actions Suppl. 1993;43:209-220.
- 134. Lukacs NW, Strieter RM, Chensue SW, et al. Activation and regulation of chemokines in allergic airway in-lammation. *J Leukoc Biol.* 1996;59(1):13-17.
- 135. Krishnaswamy G, Ajitawi O, Chi DS. The human mast cell: an overview. *Methods Mol Biol.* 2006;315:13-34.
- 136. Kaartinen M, Penttila A, Kovanen PT. Accumulation of activated mast cells in the shoulder region of human coronary atheroma, the predilection site of atheromatous rupture. *Circulation.* 1994;90(4):1669- 1678.
- 137. Kovanen PT, Kaartinen M, Paavonen T. Infiltrates of activated mast cells at the site of coronary atheromatous erosion or rupture in myocardial infarction. *Circulation.* 1995;92(5):1084-1088.
- Laine P, Kaartinen M, Penttila A, et al. Association between myocardial infarction and the mast cells in 138. the adventitia of the infarct-related coronary artery. *Circulation.* 1999;99(3):361-369.
- 139. Laine P, Naukkarinen A, Heikkila L, et al. Adventitial mast cells connect with sensory nerve fibers in
- atherosclerotic coronary arteries. *Circulation.* 2000;101(14):1665-1669. Kokkonen JO, Kovanen PT. Proteolytic enzymes of mast cell granules degrade low density lipoproteins 140. and promote their granule-mediated uptake by macrophages in vitro. *J Biol Chem*. 1989;264(18):10749- 10755.
- Lindstedt KA, Kokkonen JO, Kovanen PT. Soluble heparin proteoglycans released from stimulated mast 141. cells induce uptake of low density lipoproteins by macrophages via scavenger receptor-mediated phagocytosis. *J Lipid Res.* 1992;33(1):65-75.
- Lindstedt L, Lee M, Castro GR, et al. Chymase in exocytosed rat mast cell granules effectively proteolyzes 142. apolipoprotein AI-containing lipoproteins, so reducing the cholesterol efflux-inducing ability of serum and aortic intimal fluid. *J Clin Invest.* 15 1996;97(10):2174-2182.
- 143. Wang Y, Shiota N, Leskinen MJ, et al. Mast cell chymase inhibits smooth muscle cell growth and collagen expression in vitro: transforming growth factor-beta1-dependent and -independent effects. *Arterioscler Thromb Vasc Biol.* 2001;21(12):1928-1933.
- Leskinen M, Wang Y, Leszczynski D, et al. Mast cell chymase induces apoptosis of vascular smooth mus-144. cle cells. *Arterioscler Thromb Vasc Biol.* 2001;21(4):516-522.
- Leskinen MJ, Kovanen PT, Lindstedt KA. Regulation of smooth muscle cell growth, function and death in 145. vitro by activated mast cells--a potential mechanism for the weakening and rupture of atherosclerotic plaques. *Biochem Pharmacol.* 2003;66(8):1493-1498.
- Leskinen MJ, Heikkila HM, Speer MY, et al. Mast cell chymase induces smooth muscle cell apoptosis by 146. disrupting NF-kappaB-mediated survival signaling. *Exp Cell Res.* 2006;312(8):1289-1298.
- 147. Kaartinen M, Penttila A, Kovanen PT. Mast cells accompany microvessels in human coronary atheromas: implications for intimal neovascularization and hemorrhage. *Atherosclerosis.* 1996;123(1-2):123-131.
- Lappalainen H, Laine P, Pentikainen MO, et al. Mast cells in neovascularized human coronary plaques 148. store and secrete basic fibroblast growth factor, a potent angiogenic mediator. *Arterioscler Thromb Vasc Biol.* 2004;24(10):1880-1885.
- Sun J, Sukhova GK, Wolters PJ, et al. Mast cells promote atherosclerosis by releasing proinflammatory 149. Sun J, Sukhova GK, Wolters PJ, et al. Mas
.cytokines. *Nat Med.* 2007;13(6):719-724
- 150. Rossi D, Zlotnik A. The biology of chemokines and their receptors. Annu Rev Immunol. 2000;18:217-242.
- 151. Bazan JF, Bacon KB, Hardiman G, et al. A new class of membrane-bound chemokine with a CX3C motif. *Nature.* 1997;385(6617):640-644.
- 152. Moser B, Willimann K. Chemokines: role in inflammation and immune surveillance. *Ann Rheum Dis.* 2004;63 Suppl 2:ii84-ii89.
- 153. Burke-Gaffney A, Brooks AV, Bogle RG. Regulation of chemokine expression in atherosclerosis. *Vascul Pharmacol.* 2002;38(5):283-292.
- Weber C. Platelets and chemokines in atherosclerosis: partners in crime. *Circ Res.* 2005;96(6):612-616. 154.
- 155. Reape TJ, Groot PH. Chemokines and atherosclerosis. Atherosclerosis. 1999;147(2):213-225.
- 156. Lau EK, Allen S, Hsu AR, et al. Chemokine-receptor interactions: GPCRs, glycosaminoglycans and viral chemokine binding proteins. *Adv Protein Chem*. 2004;68:351-391.
- 157. Allen SJ, Crown SE, Handel TM. Chemokine: receptor structure, interactions, and antagonism. Annu Rev *Immunol.* 2007;25:787-820.
- 158. Thelen M. Dancing to the tune of chemokines. *Nat Immunol.* Feb 2001;2(2):129-134.
- 159. Arai H, Charo IF. Differential regulation of G-protein-mediated signaling by chemokine receptors. *J Biol Chem.* 1996;271(36):21814-21819.
- 160. Muller S, Lohse MJ. The role of G-protein beta gamma subunits in signal transduction. *Biochem Soc Trans.* 1995;23(1):141-148.
- 161. Hausdorff WP, Caron MG, Lefkowitz RJ. Turning off the signal: desensitization of beta-adrenergic receptor function. *Faseb J.* 1990;4(11):2881-2889.
- 162. Premont RT, Inglese J, Lefkowitz RJ. Protein kinases that phosphorylate activated G protein-coupled receptors. *Faseb J.* 1995;9(2):175-182.
- 163. Freedman NJ, Lefkowitz RJ. Desensitization of G protein-coupled receptors. *Recent Prog Horm Res.* 1996;51:319-351; discussion 352-313.
- 164. Arriza JL, Dawson TM, Simerly RB, et al. The G-protein-coupled receptor kinases beta ARK1 and beta ARK2 are widely distributed at synapses in rat brain. *J Neurosci.* 1992;12(10):4045-4055.
- 165. Attramadal H, Arriza JL, Aoki C, et al. Beta-arrestin2, a novel member of the arrestin/beta-arrestin gene family. *J Biol Chem.* 1992;267(25):17882-17890.
- 166. Ungerer M, Bohm M, Elce JS, et al. Altered expression of beta-adrenergic receptor kinase and beta 1-adrenergic receptors in the failing human heart. *Circulation.* 1993;87(2):454-463.
- 167. Ungerer M, Parruti G, Bohm M, et al. Expression of beta-arrestins and beta-adrenergic receptor kinases in the failing human heart. *Circ Res.* 1994;74(2):206-213.
- 168. Gros R, Chorazyczewski J, Meek MD, et al. G-Protein-coupled receptor kinase activity in hypertension

: increased vascular and lymphocyte G-protein receptor kinase-2 protein expression. *Hypertension.* 2000;35(1 Pt 1):38-42.

- Feldman RD. Beta-adrenergic inhibition of Na-K-Cl cotransport in lymphocytes. *Am J Physiol.* 1992;263(5 169. Pt 1):C1015-1020.
- Gros R, Benovic JL, Tan CM, et al. G-protein-coupled receptor kinase activity is increased in hypertension. *J Clin Invest.* 1997;99(9):2087-2093. 170.
- 171. Eckhart AD, Ozaki T, Tevaearai H, et al. Vascular-targeted overexpression of G protein-coupled receptor kinase-2 in transgenic mice attenuates beta-adrenergic receptor signaling and increases resting blood pressure. *Mol Pharmacol.* 2002;61(4):749-758.
- 172. Dinudom A, Fotia AB, Lefkowitz RJ, et al. The kinase Grk2 regulates Nedd4/Nedd4-2-dependent control of epithelial Na+ channels. *Proc Natl Acad Sci U S A.* 2004;101(32):11886-11890.
- 173. Liu S, Premont RT, Kontos CD, et al. A crucial role for GRK2 in regulation of endothelial cell nitric oxide synthase function in portal hypertension. *Nat Med.* 2005;11(9):952-958.
- 174. Lombardi MS, Kavelaars A, Schedlowski M, et al. Decreased expression and activity of G-protein-coupled receptor kinases in peripheral blood mononuclear cells of patients with rheumatoid arthritis. *Faseb J.* 1999;13(6):715-725.
- 175. Levine JD, Coderre TJ, Helms C, et al. Beta 2-adrenergic mechanisms in experimental arthritis. *Proc Natl* **A** *Acad Sci U S A.* 1988;85(12):4553-4556.
- 176. Lombardi MS, Kavelaars A, Cobelens PM, et al. Adjuvant arthritis induces down-regulation of G proteincoupled receptor kinases in the immune system. *J Immunol.* 2001;166(3):1635-1640.
- 177. Vroon A, Kavelaars A, Limmroth V, et al. G protein-coupled receptor kinase 2 in multiple sclerosis and experimental autoimmune encephalomyelitis. *J Immunol.* 2005;174(7):4400-4406.
- 178. Houtkamp MA, de Boer OJ, van der Loos CM, et al. Adventitial infiltrates associated with advanced atherosclerotic plaques: structural organization suggests generation of local humoral immune responses. *J Pathol.* 2001;193(2):263-269.
- 179. McGeachie J, Campbell P, Simpson S, et al. Arterial vasa vasorum: a quantitative study in the rat. *J Anat.* 1982;134(Pt 2):193-197.
- 180. Gulbenkian S, Saetrum Opgaard O, Ekman R, et al. Peptidergic innervation of human epicardial coronary arteries. *Circ Res.* 1993;73(3):579-588.
- 181. Maiellaro K, Taylor WR. The role of the adventitia in vascular inflammation. *Cardiovasc Res.* 2007;75(4):640-648.
- 182. Moreno PR, Purushothaman KR, Fuster V, et al. Intimomedial interface damage and adventitial inflammation is increased beneath disrupted atherosclerosis in the aorta: implications for plaque vulnerability. *Circulation.* 2002;105(21):2504-2511.
- 183. Higuchi ML, Gutierrez PS, Bezerra HG, et al. Comparison between adventitial and intimal inflammation of ruptured and nonruptured atherosclerotic plaques in human coronary arteries. *Arq Bras Cardiol.* 2002;79(1):20-24.
- 184. Ramshaw AL, Roskell DE, Parums DV. Cytokine gene expression in aortic adventitial inflammation associated with advanced atherosclerosis (chronic periaortitis). *J Clin Pathol.* 1994;47(8):721-727.
- Atkinson JB, Harlan CW, Harlan GC, et al. The association of mast cells and atherosclerosis: a morphologic study of early atherosclerotic lesions in young people. *Hum Pathol.* 1994;25(2):154-159. 185.
- 186. Schumacher H, Kaiser E, Schnabel PA, et al. Immunophenotypic characterisation of carotid plaque: increased amount of inflammatory cells as an independent predictor for ischaemic symptoms. *Eur J Vasc Endovasc Surg.* 2001;21(6):494-501.
- 187. Rayner K, Van Eersel S, Groot PH, et al. Localisation of mRNA for JE/MCP-1 and its receptor CCR2 in atherosclerotic lesions of the ApoE knockout mouse. *J Vasc Res.* 2000;37(2):93-102.
- 188. Milei J, Parodi JC, Fernandez Alonso G, et al. Carotid atherosclerosis. Immunocytochemical analysis of the vascular and cellular composition in endarterectomies. *Cardiologia.* 1996;41(6):535-542.
- Wilcox JN, Scott NA. Potential role of the adventitia in arteritis and atherosclerosis. *Int J Cardiol.* 1996;54 189. Suppl:S21-35.
- 190. Xu F, Ji J, Li L, et al. Adventitial fibroblasts are activated in the early stages of atherosclerosis in the apolipoprotein E knockout mouse. *Biochem Biophys Res Commun.* 2007;352(3):681-688.
- 191. Xu F, Ji J, Li L, et al. Activation of adventitial fibroblasts contributes to the early development of atherosclerosis: a novel hypothesis that complements the "Response-to-Injury Hypothesis" and the "Inflammation Hypothesis". *Med Hypotheses.* 2007;69(4):908-912.
- 192. Yun AJ, Doux JD, Bazar KA, et al. Adventitial dysfunction: an evolutionary model for understanding atherosclerosis. *Med Hypotheses.* 2005;65(5):962-965.
- 193. Moreno PR, Purushothaman KR, Fuster V, et al. Plaque neovascularization is increased in ruptured atherosclerotic lesions of human aorta: implications for plaque vulnerability. *Circulation.* 2004;110(14):2032- 2038.
- 194. Carlier S, Kakadiaris IA, Dib N, et al. Vasa vasorum imaging: a new window to the clinical detection of vulnerable atherosclerotic plaques. *Curr Atheroscler Rep.* 2005;7(2):164-169.
- 195. Fuchs S, Kornowski R, Leon MB, et al. Anti-angiogenesis: A new potential strategy to inhibit restenosis. *Int J Cardiovasc Intervent.* 2001;4(1):3-6.
- 196. Hayden MR, Tyagi SC. Vasa vasorum in plaque angiogenesis, metabolic syndrome, type 2 diabetes mellitus, and atheroscleropathy: a malignant transformation. *Cardiovasc Diabetol.* 2004;3:1.
- 197. Conway EM. Angiogenesis: a link to thrombosis in athero-thrombotic disease. Pathophysiol Haemost *Thromb.* 2003;33(5-6):241-248.
- 198. Stenmark KR, Davie NJ, Reeves JT, et al. Hypoxia, leukocytes, and the pulmonary circulation. *J Appl Physiol.* 2005;98(2):715-721.
- 199. Norrby K. Mast cells and de novo angiogenesis: angiogenic capability of individual mast-cell mediators

such as histamine, TNF, IL-8 and bFGF. *In-lamm Res.* 1997;46 Suppl 1:S7-8.

- Grutzkau A, Kruger-Krasagakes S, Baumeister H, et al. Synthesis, storage, and release of vascular endothelial growth factor/vascular permeability factor (VEGF/VPF) by human mast cells: implications for the biological significance of VEGF206. *Mol Biol Cell.* 1998;9(4):875-884. 200.
- Levi-Schaffer F, Pe'er J. Mast cells and angiogenesis. *Clin Exp Allergy.* 2001;31(4):521-524. 201.
- Langheinrich AC, Michniewicz A, Sedding DG, et al. Correlation of vasa vasorum neovascularization and 202. plaque progression in aortas of apolipoprotein E(-/-)/low-density lipoprotein(-/-) double knockout mice. *Arterioscler Thromb Vasc Biol.* 2006;26(2):347-352.
- Langheinrich AC, Michniewicz A, Bohle RM, et al. Vasa vasorum neovascularization and lesion dis-203. tribution among different vascular beds in ApoE-/-/LDL-/- double knockout mice. *Atherosclerosis.* 2007;191(1):73-81.
- 204. Moos MP, John N, Grabner R, et al. The lamina adventitia is the major site of immune cell accumulation in standard chow-fed apolipoprotein E-deficient mice. *Arterioscler Thromb Vasc Biol.* 2005;25(11):2386-2391.
- 205. Fleiner M, Kummer M, Mirlacher M, et al. Arterial neovascularization and inflammation in vulnerable patients: early and late signs of symptomatic atherosclerosis. *Circulation.* 2004;110(18):2843-2850.
- 206. Kolodgie FD, Gold HK, Burke AP, et al. Intraplaque hemorrhage and progression of coronary atheroma. *N Engl J Med.* 2003;349(24):2316-2325.
- 207. Makin AJ, Chung NA, Silverman SH, et al. Vascular endothelial growth factor and tissue factor in patients with established peripheral artery disease: a link between angiogenesis and thrombogenesis? *Clin Sci (Lond).* 2003;104(4):397-404.
- 208. Moulton KS, Vakili K, Zurakowski D, et al. Inhibition of plaque neovascularization reduces macrophage accumulation and progression of advanced atherosclerosis. *Proc Natl Acad Sci U S A.* 2003;100(8):4736- 4741.
- 209. Brown DA, Breit SN, Buring J, et al. Concentration in plasma of macrophage inhibitory cytokine-1 and risk of cardiovascular events in women: a nested case-control study. *Lancet.* 2002;359(9324):2159-2163.
- 210. Wollert KC, Kempf T, Peter T, et al. Prognostic value of growth-differentiation factor-15 in patients with non-ST-elevation acute coronary syndrome. *Circulation.* 2007;115(8):962-971.
- 211. Schlittenhardt D, Schober A, Strelau J, et al. Involvement of growth differentiation factor-15/macrophage inhibitory cytokine-1 (GDF-15/MIC-1) in oxLDL-induced apoptosis of human macrophages in vitro and in arteriosclerotic lesions. *Cell Tissue Res.* 2004;318(2):325-333.
- Zimmers TA, Jin X, Hsiao EC, et al. Growth differentiation factor-15: induction in liver injury through p53 212. and tumor necrosis factor-independent mechanisms. *J Surg Res.* 2006;130(1):45-51.
- 213. Trogan E, Feig JE, Dogan S, et al. Gene expression changes in foam cells and the role of chemokine receptor CCR7 during atherosclerosis regression in ApoE-deficient mice. *Proc Natl Acad Sci U S A.* 2006;103(10):3781-3786.
- 214. Ley K, Laudanna C, Cybulsky MI, et al. Getting to the site of inflammation: the leukocyte adhesion cascade updated. *Nat Rev Immunol.* Sep 2007;7(9):678-689.
- 215. Packard RR, Libby P. Inflammation in Atherosclerosis: From Vascular Biology to Biomarker Discovery and Risk Prediction. *Clin Chem.* 2008;54(1):24-38.
- Libby P, Shi GP. Mast cells as mediators and modulators of atherogenesis. *Circulation*. 2007;115(19):2471- 216. 2473.

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