

Molecular and cellular characterization of cardiac overload-induced hypertrophy and failure

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Citation

Umar, S. (2009, June 18). *Molecular and cellular characterization of cardiac overload-induced hypertrophy and failure*. Retrieved from https://hdl.handle.net/1887/13860

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CHAPTER 3

Myocardial collagen metabolism in failing hearts before and during cardiac resynchronisation therapy

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European Journal of Heart Failure 2008;10:878-883

Abstract

Background: In patients with heart failure cardiac resynchronization therapy (CRT) leads to reverse ventricular remodelling.

Aim: To evaluate whether myocardial collagen metabolism in patients with heart failure is implicated in adverse ventricular remodelling and response to CRT.

Methods: Collagen synthesis and degradation were assessed from the concentrations of aminoterminal propeptides of type I and type III collagen (PINP and PIIINP) and carboxyterminal telopeptide of type I collagen (ICTP), respectively, in serum of 64 patients with heart failure before and after 6 months of CRT. The pro-form of matrix metalloproteinase-1 (proMMP1) and tissue inhibitor of metalloproteinase-1 (TIMP1) were also assayed at these time points. Forty-six patients (72%) showed a >10% reduction in LV end-systolic volume at follow-up and were classified as responders to CRT, the other 18 patients (28%) were classified as non-responders.

Results: Responders demonstrated a mean (±SEM) increase of serum PINP and PIIINP during follow-up, from 32.9±2.2 to 46.7±4.0 µg/L (*p*<0.001) and from 4.59±0.24 to 5.13±0.36 µg/L (*p*<0.05), respectively. In non-responders, serum PINP and PIIINP remained unchanged during follow-up. At baseline, responders had significantly lower serum PINP than non-responders (32.9±2.2 vs. 41.8±4.3 µg/L; *p*<0.05). ICTP levels of responders at baseline tended to be higher than in non-responders (3.54±0.56 vs. 2.08±0.37 µg/L, *p*=ns), and in both groups ICTP levels did not change upon CRT.

Conclusion: Reverse LV remodelling following CRT is associated with increased collagen synthesis rate in the first 6 months of follow-up.

Key words: Heart failure, cardiac resynchronisation therapy, PINP, PIIINP, ICTP

Introduction

In addition to myocardial hypertrophy, overloaded hearts frequently demonstrate a reactive fibrosis due to alterations in collagen synthesis and degradation. Accumulation of interstitial collagen type I and type III increases myocardial stiffness resulting in changes in diastolic properties of the left ventricle (LV) [1].

Collagen type I and type III are secreted by interstitial fibroblasts as procollagens followed by splitting off of propeptides by endopeptidases and the release of their aminoterminal propeptides, PINP and PIIINP, and carboxyterminal propeptides, PICP and PIIICP, into the circulation. During degradation of type I collagen fibrils the carboxyterminal telopeptide of type I collagen (ICTP) is formed, which is a 12 kD peptide [2].

Extracellular matrix components, like collagen type I and type III, are continuously synthesized and degraded. With respect to degradation, matrix metalloproteinase-1 (MMP1), a collagenase, and MMP2 and MMP9, two gelatinases, can be upregulated at the mRNA level [3], can be activated at the protein level, for instance by MT₁-MMP (or MMP14)[4], and can be stimulated by lowered concentrations of MMP inhibitors like tissue inhibitor of metalloproteinases (TIMPs) [5].

The myocardial extracellular matrix is implicated in a number of conditions and diseases, such as hypertensive heart disease, dilated cardiomyopathy, and cardiac remodelling. In failing hearts myocardial MMP1 and MMP2 concentrations have been shown to be elevated, while TIMP1 levels were depressed or unchanged [6]. Rapid myocardial collagen synthesis rate and slow collagen degradation rate impair diastolic function and promote LV adverse remodelling, probably due to cardiomyocyte slippage secondary to increased LV diastolic pressures [7].

In the present study, the biochemical markers of collagen synthesis rate (PINP and PIIINP) and collagen degradation rate (ICTP), including proMMP1 and TIMP1, were assayed in patients with heart failure before and after 6 months of cardiac resynchronisation therapy (CRT), to evaluate whether responders and non-responders to CRT differ with regard to serum markers of collagen metabolism at those two time points and whether serum markers of collagen metabolism can predict response to CRT.

Methods

Patients

Sixty-four patients with heart failure, scheduled for implantation of a CRT device, were included in the study. Baseline characteristics of these patients have been reported in an earlier study evaluating the levels of circulating biomarkers of extracellular matrix metabolism [8]. The group had a mean age 64±1.2 years and consisted of 52 men (81%) and 12 women (19%). Ischaemic aetiology of heart failure was present in 45 patients (70%) and non-ischaemic aetiology in 19 patients (30%). Selection criteria for CRT were New York Heart Association (NYHA) functional class III-IV, LV ejection fraction ≤35%, and QRS duration >120ms. Patients with decompensated heart failure and patients with a recent myocardial infarction (<3 months) were excluded. LV dyssynchrony was quantified at baseline. Clinical status was assessed before CRT implantation (baseline) and at 6 months follow-up. At both time points, 2-dimensional echocardiography was performed to determine LV volumes and LV ejection fraction.

Clinical evaluation

At baseline and at 6 months follow-up clinical status was evaluated by (*i*) assessment of NYHA functional class, (*ii*) determination of quality-of-life score using the Minnesota Living with Heart Failure questionnaire, and (*iii*) assessment of exercise capacity using the distance of a 6-minute hall walk test.

Echocardiography

Echocardiography was performed as described previously [8,9]. A commercially available system (Vingmed system Seven, General Electric-Vingmed, Milwaukee, Wisconsin, USA) was used to acquire conventional apical 2- and 4-chamber images, which allow calculation of LV end-systolic and end-diastolic volumes (LVESV and LVEDV) and LV ejection fraction, using the biplane Simpson's technique [10] and commercial software (Echopac version 5.0.1, General Electric - Vingmed). Echocardiographic data were analyzed by two independent observers who were blinded to all other patient data. Inter- and intra-observer variabilities for assessment of LV ejection fraction and LV volumes were 90% and 96%, respectively [11]. Based on recent data [12], patients with a reduction of >10% in LVESV at 6 months follow-up were classified as responders to CRT, whereas patients with a reduction of ≤10% in LVESV or an increased LVESV at 6 months follow-up, were classified as non-responders. For the assessment of LV dyssynchrony, two regions of interest were selected in the basal portions of the septum and the LV lateral wall allowing calculation of the septal-to-lateral delay in time-to-peak systolic myocardial velocity [9]. A septal-to-lateral delay of \geq 65 ms was considered to represent significant LV dyssynchrony [13]. Additionally, wall motion score index (WMSI) was assessed in all subjects. The LV was divided into 16 segments. A semi quantitative scoring system (1, normal; 2, hypokinesia; 3, akinesia; 4, dyskinesia) was used to analyze each study. Global WMSI was calculated by the standard formula: sum of the segment scores divided by the number of segments scored [14].

Pacemaker implantation

The LV pacing lead was inserted via the subclavian vein. After a coronary sinus venogram was obtained, the LV pacing lead was introduced into the coronary sinus and placed as far as possible in the venous system, preferably in a (postero-) lateral vein. The right atrial and right ventricular leads were positioned according to standard procedures. Implantation of leads and CRT device was accomplished in all patients without major complications.

Biochemical analysis

Blood samples were obtained before CRT and after 6 months follow-up. Serum and EDTA-plasma samples were stored at -80°C prior to assay.

Serum levels of carboxyterminal cross-linked telopeptide of type I collagen (ICTP) were assayed by a competitive ELISA (Orion Diagnostica, Espoo, Finland) which has a measurement range of 1.0-50 μ g/L [15]. Inter-assay variations were 6% both at high (28 μ g/L) and low (3 μ g/L) ICTP concentrations. Detection limit of the assay was 0.3 μ g/L. Intra-assay variability was 11% at low ICTP concentrations, and 7% at high ICTP concentrations. Reference ranges of human serum ICTP concentrations were 1.6-4.2 μ g/L for women and 1.5-4.3 μ g/L for men.

To measure of rates of synthesis of collagen type I and type III, we chose to use serum levels of aminoterminal propeptides of type I collagen (PINP) and of type III collagen (PIINP). We acknowledge that there is ample experience with the use of the carboxyterminal propeptide of collagen I (PICP) (see for instance [16]); however, the commercial availability of assays for both PINP and PIIINP and the fact that there is a 1:1:1 stoichiometric relationship between the aminoterminal propeptide, the carboxyterminal propeptide and the collagen molecule formed for either collagen type, influenced our decision to use the assays to quantify PINP and PIIINP. Serum levels of PINP were assayed by radioimmunoassay (Orion Diagnostica) which has a measurement range of 5-250 μ g/L [17]. Inter-assay variations were 6% both at high (167 μ g/L) and 9.8% at low (12 μ g/L) PINP concentrations. Detection limit of the assay was 2 μ g/L. Intra-assay variability was 9.8% at low PINP concentrations (12 μ g/L) and 10.2% at high (173 μ g/L) PINP concentrations. Reference ranges of human serum PINP concentrations were 19-83 μ g/L for women and 22-87 μ g/L for men.

Serum levels of PIIINP were assayed by radioimmunoassay (Orion Diagnostica) which has a measurement range of 1.0-50 µg/L [18]. Inter-assay variations were 7.2 % at high (12.2 µg/L) and 6.5 % at low (2.7 µg/L) PIIINP concentrations. Detection limit of the assay was 0.3 µg/L. Intra-assay variability was 3.0 % at low PIIINP concentrations (2.8 µg/L) and 4.1% at high (11.9 µg/L) PIIINP concentrations. Reference ranges of adult human serum PIIINP concentrations were 2.3-6.4 µg/L.

The tissue inhibitor of metalloproteinases-1 (TIMP1) was assayed in EDTAplasma by ELISA (GE Healthcare, Little Chalfont, Buckinghamshire, UK) which has a measurement range of 3-50 μ g/L. Inter-assay variations were 15% at low (12.5 μ g/L) and 13% at high (47.3 μ g/L) TIMP1 concentrations. Intra-assay

variability was 11% at low (10.3 μ g/L) and 9% at high (39.4 μ g/L) TIMP1 concentrations. Reference range of adult human EDTA-plasma TIMP1 concentrations was 49 – 183 μ g/L [19].

Serum levels of pro-matrix metalloproteinase-1 (proMMP-1) were assayed by ELISA (Quantikine, R&D Systems, Abingdon Science Park, Abingdon, UK) which has a measurement range of $0.15 - 10 \ \mu g/L$. Inter-assay variations were 10.4% at low (0.74 $\mu g/L$) and 6.7% at high (4.66 $\mu g/L$) proMMP-1 concentrations. Intra-assay variability was 5.6% at low (0.70 $\mu g/L$) and 5.4% at high (4.43 $\mu g/L$) proMMP-1 concentrations. Detection limit of the assay was 0.02 $\mu g/L$. According to the manufacturer, average value of healthy adult human serum proMMP-1 concentration was 3.45 $\mu g/L$, with a range of 0.91 – 9.34 $\mu g/L$. Mean reference values (± SD) reported in literature are 8.5 ± 5.2 $\mu g/L$ [20] and 2.43 ± 0.37 $\mu g/L$ [21].

Statistical analysis

Data were expressed as mean ±SEM unless stated otherwise. Differences between the two groups (responders and non-responders) were tested with twotailed Student's t-test for paired or unpaired data when appropriate. Univariable and multivariable logistic regression analyses were performed to characterize predictors of good response to CRT. Continuous variables included baseline values of NYHA functional class, quality-of-life score, 6-minute hall walk distance, LVESV, LVEDV, LV ejection fraction, NT-proBNP, PINP, PIIINP, ICTP, TIMP1 and proMMP1. All variables with p<0.25 entered the multivariable regression analysis that was performed by stepwise backward deletion. All variables with p<0.25 remained in the final model. All statistical tests were performed with SPSS 12.1 software (SPSS Inc, Chicago, IL, USA). For all tests, a probability value <0.05 was considered statistically significant.

Results

None of the patients died during the 6 months follow-up period. CRT was successful in reducing LVESV by >10% in the first 6 months in 46 (72%) patients (responders), whereas in 18 (28%) patients LVESV was reduced by <10% or was even increased (non-responders). The clinical and echocardiographic variables at baseline and after 6 months of CRT in responders and non-responders have been presented previously [8]. At baseline, responders (1) were older (65.5±1.4 yrs vs. 59.5±2.6 yrs, p=0.03), (2) had more dyssynchrony (100±8 ms vs. 71±11 ms, p=0.05), and (3) had longer QRS duration (165±3 ms vs. 135±8 ms, p<0.001), compared to non-responders. There was no difference in baseline WMSI between responders and non-responders (2.30±0.27 vs. 2.18±0.20, n.s.). In responders, NYHA class improved from 3.1 ± 0.1 to 2.0 ± 0.1 (p<0.001), the 6-minute hall walk distance improved from 333 ± 18 m to 427 ± 17 m (p<0.001), and quality-of-life score improved from 35 ± 3 to 17 ± 3 (p<0.001).

Responders had a lower serum PINP level at baseline ($32.9\pm2.2 \ \mu g/L \ vs. 41.9\pm4.3 \ \mu g/L$, p=0.04) than non-responders. During follow-up, responders demonstrated an increase in serum PINP level from $32.9\pm2.2 \ \mu g/L$ to $46.7\pm4.0 \ \mu g/L$ (p<0.001), whereas non-responders demonstrated no significant change (see Figure). At baseline, plasma levels of PIIINP in responders did not differ from those of nonresponders. In responders, plasma PIIINP levels increased during follow-up (from 4.59 ± 0.24 to $5.13\pm0.36 \ \mu g/L$, p<0.05), whereas in non-responders plasma PIIINP levels remained unchanged (see Figure).



Figure 1. Levels of aminoterminal propeptide of type I procollagen (PINP), aminoterminal propeptide of type III procollagen (PIIINP), and carboxyterminal cross-linked telopeptide of type I collagen (ICTP) measured in serum of 64 patients with congestive heart failure at baseline (\Box) and at 6 months follow-up (**n**), divided in a group of patients who were successfully treated by CRT (responders) and a group of patients who responded poorly to CRT (non-responders).

Serum ICTP levels tended to be higher in responders at baseline and at 6 months follow-up than in non-responders at corresponding time points (see Figure). Plasma levels of proMMP1, TIMP1 and proMMP1/TIMP1 did not differ between responders and non-responders at baseline, nor during follow-up (see Table 1).

Table 1.	TIMP1	and prol	MMP1 cond	centra	atior	าร	in plasm	a of 64 pat	ients witl	h heart failı	Jre
determine	ed befo	re CRT	(baseline)	and	at	6	months	follow-up,	divided	according	to
response to CRT. (mean values ± SEM).											

	Responders n=46	Non-responders n=18	<i>p</i> -value	
TIMP1 (baseline) (μg/L)	124±5.2	111±7.1	0.16	
TIMP1 (6 months follow-up) (μg/L)	129±5.8	112±7.7	0.11	
proMMP1 (baseline) (µg/L)	7.55±0.72	8.04±1.12	0.71	
proMMP1 (6 mo follow-up) (µg/L)	7.89±0.85	8.08±1.11	0.90	
proMMP1/TIMP1 (baseline)	0.063±0.006	0.080±0.015	0.20	
proMMP1/TIMP1 (6 mo f-u)	0.064±0.007	0.083±0.017	0.21	

Abbreviations: CRT, cardiac resynchronisation therapy; TIMP1, tissue inhibitor of metalloproteinases-1; proMMP1, pro-matrix metalloproteinase-1.

Univariable correlation of the following baseline parameters, NYHA functional class, quality-of-life score, 6-minute hall walk distance, LVESV, LVEDV, LV ejection fraction, NT-proBNP, PINP, ICTP, PIIINP, TIMP1 and proMMP1, with good response to CRT demonstrated that PINP scored best (Table 2).

Multivariable correlation of all variables that had an univariable probability value <0.25 demonstrated that PINP, PIIINP and LVEDV at baseline were correlated with good response to CRT, but only PINP to a significant extent (Table 2).

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	Uni [,]	variable ana	lysis	 Multivariable analysis			
	OR	CI	<i>p</i> -value	 OR	CI	<i>p</i> -value	
NYHA functional class	0.74	0.28-1.96	0.55 0.55	 			
Quality-of-life score	0.99	0.96-1.01	0.45				
6-min hall walk distance	1.00	0.99-1.00	0.76				
LVESV	1.00	0.99-1.01	0.15				
LVEDV	1.00	0.99-1.01	0.20	1.00	0.99-1.01	0.14	
LV ejection fraction	0.94	0.88-1.02	2 0.15				
NT-proBNP	1.00	1.00-1.00	0.22				
PINP	0.99	0.93-1.00	0.05	0.96	0.93-0.99	0.03	
ICTP	1.24	0.93-1.66	6 0.13				
PIIINP	1.23	0.86-1.76	6 0.23	1.35	0.94-1.93	0.10	
TIMP1	1.01	0.99-1.03	0.16				
proMMP1	0.97	0.87-1.09	0.71				

Table 2. Univariable and multivariable correlation of baseline parameters with good response to CRT (reduction of LVESV by >10%).

Abbreviations: OR, odds ratio; CI, 95% confidence limits; CRT, cardiac resynchronisation therapy; NYHA, New York Heart Association; LVESV, left ventricular end-systolic volume; LVEDV, left ventricular end-diastolic volume; NT-proBNP, N-terminal pro B-type natriuretic peptide; PINP, aminoterminal propeptide of type I procollagen; ICTP, carboxyterminal cross-linked telopeptide of type I collagen; PIINP, aminoterminal propeptide of type III procollagen; TIMP1, tissue inhibitor of metalloproteinases-1; proMMP1, pro-matrix metalloproteinase-1.

Discussion

The present study demonstrated that patients with heart failure who were successfully treated with CRT had relatively low serum levels of PINP at baseline, a measure of collagen synthesis rate, and relatively high serum levels of ICTP at baseline, a measure of collagen degradation rate. After 6 months of CRT, serum PINP and PIIINP levels had increased significantly, whereas serum ICTP had hardly changed. Accordingly, a high serum level of PINP at baseline is associated with failure to respond to CRT, which is in line with reports showing that high levels of markers of collagen synthesis in serum of patients with heart failure are associated with poor outcome [16, 22-24].

In patients with hypertension and LV hypertrophy, elevated PINP levels in serum at baseline are associated with an increased myocardial collagen volume fraction (a measure of myocardial fibrosis) [25], low plasma levels of total and free MMP1, and high plasma levels of total and free TIMP1 [26]. Myocardial fibrosis is often associated with abnormal myocardial stiffness [27], diastolic abnormalities, and a decline in myocardial elastance during contraction, a measure of systolic dysfunction [28]. Perivascular accumulation of collagen may impair the vasodilator capacity of intramyocardial coronary arteries and contribute to the decrease in coronary reserve [29]. In patients with hypertrophic cardiomyopathy, high serum levels of PINP were associated with reduced diastolic function [30]. In patients with essential hypertension, serum levels of PINP and PIIINP were elevated compared to normotensives. In hypertensives, serum PIIINP levels were inversely correlated to diastolic (dys)function, whereas serum PINP levels were correlated positively to LV mass index [24]. In patients with dilated cardiomyopathy, the presence of a restrictive mitral pattern was associated with highest serum levels of PIIINP and high serum levels of PIIINP were related to poor outcome [31]. In patients with dilated cardiomyopathy, serum PIIINP levels were significantly elevated compared to corresponding levels in controls, these serum PIIINP levels were correlated to NYHA functional class, daily diuretics dosage, and mean right atrial pressure, and were inversely correlated to cardiac output [24].

In the present study, the responders to CRT tended to have higher serum ICTP levels than non-responders. Six months therapy with CRT had no significant effect on serum ICTP in responders, nor in non-responders. However, serum levels of proMMP1, a collagenase, did not differ between baseline and follow-up, nor did they differ between responders and non-responders at these two time points.

Previously it has been demonstrated that patients with dilated cardiomyopathy had higher serum levels of MMP1 than controls, which was associated with higher MMP1/TIMP1 ratio, and serum free MMP1 as well as MMP1/TIMP1 ratio which correlated positively with LV end-diastolic volume and negatively with cardiac index [21]. In failing hearts supported by a LV assist device myocardial MMP1 levels decreased, whereas TIMP1 and TIMP3 levels increased [32].

High serum levels of ICTP have also been observed in patients with dilated cardiomyopathy [21] and in patients with hypertrophic cardiomyopathy [30]. Klappacher et al. found that serum ICTP levels beyond a cut-off level of 7.6 µg/L

in patients with dilated cardiomyopathy were associated with increased risk of advanced clinical stage, increased risk of poor haemodynamic condition, increased risk of hyponatraemia (<138 mmol/L), and increased risk of heart transplantation [24].

The increase of serum PINP in responders to CRT (the patients demonstrating reverse LV remodelling) in the present study, is interpreted as an increased myocardial collagen synthesis in responders to CRT during 6 months follow-up. This increased myocardial collagen synthesis in responders may be a reaction of the myocardium induced by improved loading conditions (due to improved synchronicity of segmental wall motion); and thus LVESV and LVEDV were decreased by biochemical forces within the tissue that provide "passive tissue contraction".

The responders were the group of patients with the lowest collagen synthesis at baseline, which according to the above explanation, associates with adverse LV remodelling by weakening the myocardial structure, thereby allowing progressive LV dilatation. In a study using myocardial biopsies from patients with end-stage heart failure taken before and after 100-600 days of left ventricular assist device (LVAD) implantation, Bruggink and co-workers observed an increase in ECM volume in the first 200 days after LVAD implantation, followed by a gradual decrease of ECM volume in the following 200 days to a level that was still higher than that observed pre-LVAD [33]. Plasma PINP levels increased considerably (~3-fold) in the first month after LVAD implantation, and remained increased in the first 6 months after LVAD implantation. Thus, reverse LV remodelling induced by LV unloading was associated with increased collagen synthesis and expanded ECM space. Although in our study the tertiles of plasma PINP levels at baseline showed a tendency to associate with response to CRT (lowest tertile: 17/21 responders, 81%; middle tertile: 16/22 responders, 73%; highest tertile 13/21 responders, 62%), our study population was probably too small to find a significant association.

One or more patients with systemic diseases that would induce abnormalities in collagen metabolism may have been included in the study; however, signs and symptoms of such diseases were absent. Furthermore, echocardiography-derived backscatter data indicative of myocardial fibrosis were not available in this study.

In conclusion, elevated collagen synthesis rate at baseline is unfavourable in terms of therapeutic success of CRT. Reverse LV remodelling upon CRT is associated with increased collagen synthesis rate in the first 6 months of follow-up.

Acknowledgements

This study was supported by the Netherlands Heart Foundation (Grant nr 2001B124 and 2002B109).

References

1. Weber KT, Brilla CG. Pathological hypertrophy and cardiac interstitium. Fibrosis and the renin-angiotensin-aldosterone system. Circulation 1991;83:1849-65.

2. Laurent GJ. Dynamic state of collagen: pathways of collagen degradation in vivo and their possible role in regulation of collagen mass. Am J Physiol 1987;252:C1-C9.

3. Fini ME, Cook JR, Mohan R, Brinckerhoff CE. Regulation of matrix metalloproteinase gene expression. In: Parks WC, Mecham RP (editors). Matrix Metalloproteinases. San Diego, Academic Press, 1998; pp. 299-362.

4. Sato H, Takino T, Kinoshita T, *et al.* Cell surface binding and activation of gelatinase A induced by expression of membrane-type-1- matrix metalloproteinase (MT1-MMP). FEBS Lett 1996;385:238-40.

5. Gomez DE, Alonso DF, Yoshiji H, Thorgeirsson UP. Tissue inhibitors of metalloproteinases: structure, regulation and biological functions. Eur J Cell Biol 1997;74:111-22.

6. Sakata Y, Yamamoto K, Mano T, *et al.* Activation of matrix metalloproteinases precedes left ventricular remodeling in hypertensive heart failure rats. Circulation 2004;109:2143-9.

7. Burlew BS, Weber KT. Cardiac fibrosis as a cause of diastolic dysfunction. Herz 2002;27:92-8.

8. Hessel MHM, Bleeker GB, Bax JJ, *et al.* Reverse ventricular remodelling after cardiac resynchronisation therapy is associated with a reduction in serum tenascin-C and plasma matrix metalloproteinase-9 levels. Eur J Heart Fail 2007;9:1058-63.

9. Bax JJ, Bleeker GB, Marwick TH, *et al.* Left ventricular dyssynchrony predicts response and prognosis after cardiac resynchronization therapy. J Am Coll Cardiol 2004;44:1834-40. 10. Schiller NB, Shah PM, Crawford M, *et al.* Recommendations for quantitation of the left ventricle by two-dimensional echocardiography. American Society of Echocardiography Committee on Standards, Subcommittee on Quantitation of Two-Dimensional Echocardiograms. J Am Soc Echocardiogr 1989;2:358-67.

11. Ypenburg C, Roes SD, Bleeker GB, *et al.* Effect of total scar burden on contrastenhanced magnetic resonance imaging on response to cardiac resynchronisation therapy. Am J Cardiol 2007;99:657-60.

12. Yu CM, Bleeker GB, Fung JW, *et al.* Left ventricular reverse remodeling but not clinical improvement predicts long-term survival after cardiac resynchronization therapy. Circulation 2005;112:1580-6.

13. Bax JJ, Marwick TH, Molhoek SG, *et al.* Left ventricular dyssynchrony predicts benefit of cardiac resynchronization therapy in patients with end-stage heart failure before pacemaker implantation. Am J Cardiol 2003;92:1238-40.

14. Lang RM, Bierig M, Devereux RB, *et al.* Recommendations for chamber quantification. Eur J Echocardiography 2006;7:79-108.

15. Risteli J, Elomaa I, Niemi S, Novamo A, Risteli L. Radioimmunoassay for the pyridinoline cross-linked carboxy-terminal telopeptide of type I collagen: A new serum marker of bone collagen degradation. Clin Chem 1993;39:635-40.

16. Garcia-Bolao I, Macias A, Lopez B, *et al.* A biomarker of myocardial fibrosis predicts long-term response to cardiac resynchronization therapy. J Am Coll Cardiol 2006;47:23335-7.

17. Melkko J, Kauppila S, Niemi S, *et al.* Immunoassay for intact amino-terminal propeptide of human type I procollagen. Clin Chem 1996;42:947-54.

18. Risteli J, Niemi S, Trivedi P, Mäentausta O, Mowat AP, Risteli L. Rapid equilibrium radioimmunoassay for the amino-terminal propeptide of human type III procollagen. Clin Chem 1988;34:715-8.

19. Plumpton TA, Clark IM, Plumpton C, Calvin J, Cawston TE. Development of an enzyme-linked immunosorbent assay to measure total TIMP-1 (free TIMP-1 and TIMP-1 in

combination with matrix-metalloproteinases) and measurement of TIMP-1 and CRP in serum. Clin Chim Acta 1995;240:137-54.

20. Zhang J, Fujimoto N, Iwata K, Sakai T, Okada Y, Hayakawa T. A one-step sandwich enzyme immunoassay for human matrix metalloproteinase 1 (interstitial collagenase) using monoclonal antibodies. Clin Chim Acta 1993;219:1-14.

21. Schwartzkopff B, Fassbach M, Pelzer B, Brehm M, Strauer BE. Elevated serum markers of collagen degradation in patients with mild to moderate dilated cardiomyopathy. Eur J Heart Fail 2002;4:439-44.

22. Zannad F, Alla F, Dousset B, Perez A, Pitt B, on behalf of the RALES Investigators. Limitation of excessive extracellular matrix turnover may contribute to survival benefit of spironolactone therapy to patients with congestive heart failure. Circulation 2000;102:2700-6.

23. Cicoira M, Rossi A, Bonapace S, *et al.* Independent and additional prognostic value of aminoterminal propeptide of type III procollagen circulating levels in patients with chronic heart failure. J Card Failure 2004;10:403-11.

24. Klappacher G, Franzen P, Haab D, *et al.* Measuring extracellular matrix turnover in the serum of patients with idiopathic or ischemic dilated cardiomyopathy and impact on diagnosis and prognosis. Am J Cardiol 1995;75:913-8.

25. Querejeta R, Varo N, Lopez B, *et al.* Serum carboxy-terminal propeptide of procollagen type I is a marker of myocardial fibrosis in hypertensive heart disease. Circulation 2000;101:1729-35.

26. Laviades C, Varo N, Fernandez J, *et al.* Abnormalities of the extracellular degradation of collagen type I in essential hypertension. Circulation 1998;98:535-40.

27. Zannad F, Dousset B, Alla F. Treatment of congestive heart failure. Interfering the aldosterone-cardiac extracellular matrix relationship. Hypertension 2001;38:1227-32.

28. Maceira AM, Barba J, Varo N, Beloqui O, Diez J. Ultrasonic backscatter and serum marker of cardiac fibrosis in hypertensives. Hypertension 2002;39:923-8.

29. Lopez B, Gonzalez A, Varo N, Laviades C, Querejeta R, Diez J. Biochemical assessment of myocardial fibrosis in hypertensive heart disease. Hypertension 2001;38:1222-6.

30. Lombardi R, Betocchi S, Losi MA, *et al.* Myocardial collagen turnover in hypertrophic cardiomyopathy. Circulation 2003;108:1455-60.

31. Rossi A, Cicoira M, Golia G, *et al.* Amino-terminal propeptide of type III procollagen is associated with restrictive mitral filling pattern in patients with dilated cardiomyopathy : a possible link between diastolic dysfunction and prognosis. Heart 2004;90:650-4.

32. Li YY, Feng Y, McTiernan CF, *et al.* Downregulation of matrix metalloproteinases and reduction in collagen damage in the failing human heart after support with left ventricular assist devices. Circulation 2001;104:1147-52.

33. Bruggink AH, van Oosterhout MFM, de Jonge N, *et al.* Reverse remodeling of the myocardial extracellular matrix after prolonged left ventricular assist device support follows a biphasic pattern. J Heart Lung Transplant 2006;25:1091-8.