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**Title:** Novel modulators of lipoprotein metabolism : implications for steatohepatitis and atherosclerosis

**Issue Date:** 2013-11-06

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## PROLONGED CALORIC RESTRICTION IN OBESE PATIENTS WITH TYPE 2 DIABETES MELLITUS DECREASES PLASMA CETP AND INCREASES APOLIPOPROTEIN AI LEVELS WITHOUT IMPROVING THE CHOLESTEROL EFFLUX PROPERTIES OF HDL

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*Diabetes Care* 2010; 33: 1625-8

## ABSTRACT

4

Using a mouse model for human-like lipoprotein metabolism we previously observed that reduction of the hepatic triglyceride (TG) content resulted in a decrease in plasma cholesteryl ester transfer protein (CETP) and an increase in HDL levels. The aim of the present study was to investigate the effects of prolonged caloric restriction in obese patients with type 2 diabetes mellitus, resulting in a major reduction in hepatic TG content, on plasma CETP and HDL levels. We studied 27 obese (BMI:  $37.2 \pm 0.9$  kg/m<sup>2</sup>) insulin-dependent patients with type 2 diabetes mellitus (14 men, 13 women, age  $55 \pm 2$  years) who received a 16-week very low calorie diet (VLCD). At baseline and after a 16-week VLCD, plasma lipids, lipoproteins and CETP were measured. Furthermore, functionality of HDL with respect to inducing cholesterol efflux from human monocyte cells (THP-1) was determined. A 16-week VLCD markedly decreased plasma CETP concentration (-18%,  $P < 0.01$ ) and increased plasma apoAI levels (+16%,  $P < 0.05$ ), without significantly affecting plasma HDL-cholesterol and HDL-phospholipids. Although a VLCD results in HDL that is less lipidated, the functionality of HDL with respect to inducing cholesterol efflux in vitro was unchanged. In conclusion, the marked decrease in hepatic TG content induced by a 16-week VLCD is accompanied by a decrease in plasma CETP concentration and an increase in ApoAI levels, without improving the cholesterol efflux properties of HDL in vitro.

## INTRODUCTION

Patients with type 2 diabetes mellitus display a typical atherogenic dyslipidemia marked by increased plasma triglycerides (TG) and very low density lipoprotein (VLDL)-cholesterol concentrations, and decreased high density lipoprotein (HDL)-cholesterol levels. Furthermore, hepatic steatosis, which is also strongly associated with cardiovascular disease (CVD) risk <sup>1,2</sup> is frequently observed in patients with type 2 diabetes mellitus <sup>3-5</sup>.

Previously, we demonstrated that the HDL-raising effect of various classical lipid-lowering drugs was caused by a reduction in plasma cholesteryl ester transfer protein (CETP) that mediates the net transfer of cholesteryl esters from HDL to (V)LDL. In *APOE\*3-Leiden.CETP* mice, a well-established animal model for human-like lipoprotein metabolism, statins <sup>6</sup>, fibrates <sup>7</sup>, and niacin <sup>8</sup> decrease the hepatic lipid content (i.e. both TG and cholesterol) resulting in a decreased hepatic CETP expression accompanied by decreased plasma CETP levels, and a consequently increased plasma HDL. Recently, we showed that a similar mechanism may account for the HDL-raising effect of pioglitazone in humans. In patients with type 2 diabetes mellitus, pioglitazone decreased hepatic TG content <sup>9</sup>, accompanied by a decrease in plasma CETP concentration and increase in HDL level <sup>10</sup>. In contrast, metformin did not affect either hepatic TG, plasma CETP or HDL levels <sup>10</sup>.

Lifestyle interventions such as diet-induced weight reduction and exercise are very important in the treatment of obese patients with type 2 diabetes mellitus. Recently, we reported that a 16-week very low calorie diet (VLCD) in obese patients with type 2 diabetes mellitus significantly decreased plasma total cholesterol and TG levels and markedly reduced hepatic TG content <sup>11</sup>, but the potential beneficial effect of a VLCD on plasma CETP and HDL levels has not been studied. Therefore, using plasma samples from that study <sup>11</sup>, we now investigated whether prolonged caloric restriction reduces CETP concentration and thereby increases HDL levels in obese patients with type 2 diabetes mellitus.

## METHODS

### Patients

The study protocol has previously been described in detail <sup>11</sup>. Twenty-seven obese patients with insulin-dependent type 2 diabetes mellitus (14 men and 13 women) were included (mean  $\pm$  standard error of the mean: age: 55 $\pm$ 2 years, BMI: 37.2 $\pm$ 0.9 kg/m<sup>2</sup>).

HbA1c:  $7.8 \pm 0.2\%$ ). At baseline patients used  $82 \pm 11$  units of insulin per day with or without concomitant use of metformin and/or sulfonyl ureum derivatives. Exclusion criteria were: smoking, unstable weight during 3 months before inclusion or any other chronic disease. In the previously published paper <sup>11</sup>, we described only 12 out of the 27 patients, from whom <sup>1</sup>H-MRS scans could be obtained. The local ethics committee approved this protocol. All patients gave written informed consent and the study was performed in accordance with the Declaration of Helsinki.

## Study design

Patients were studied before start and after completion of the 16-week VLCD. Three weeks before start of the VLCD all oral blood glucose lowering medication was stopped and insulin therapy was intensified. The day before the start of the VLCD intervention, only short-acting insulin was prescribed. Patients did not use any blood glucose-lowering medication, including insulin, during the 16-week VLCD. The VLCD consisted of 3 liquid food shakes (Modifast Intensive; kindly provided by Nutrition & Santé, Antwerp, Belgium) containing a total of 450 kcal/day and all essential micro- and macronutrients. Thirteen of the 27 subjects simultaneously followed an exercise program in addition to the VLCD. Since exercise had no effect on outcome parameters (Supplemental Table 1), data of all subjects were pooled for the present analyses.

**Supplemental Table 1. Changes in plasma CETP and (apo)lipoprotein values induced by 16 weeks of VLCD in obese patients with type 2 diabetes mellitus without or with an exercise program.**

Plasma parameters	Non-exercise (n=14)	Exercise (n=13)	P value
Δ CETP (μg/mL)	-0.41 ± 0.14	-0.49 ± 0.23	0.7651
Δ Total cholesterol (mM)	-0.67 ± 0.34	-0.86 ± 0.20	0.6356
Δ Triglycerides (mM)	-0.87 ± 0.19	-1.32 ± 0.50	0.4128
Δ Phospholipids (mM)	-0.37 ± 0.08	-0.45 ± 0.12	0.5822
Δ LDL-C (mM)	-0.64 ± 0.30	-0.63 ± 0.24	0.9866
Δ ApoB100 (mg/dL)	-11.3 ± 7.4	-26.8 ± 7.3	0.1477
Δ HDL-C (mM)	0.00 ± 0.09	0.15 ± 0.06	0.1896
Δ HDL-PL (mM)	-0.03 ± 0.08	0.05 ± 0.05	0.4477
Δ ApoAI (mg/dL)	34.5 ± 12.2	8.2 ± 11.3	0.1270

Changes (Δ) are calculated by subtracting values obtained after VLCD from those obtained at baseline. Data are presented as means ± SEM. P-values are calculated using Unpaired Student's t test. CETP, cholesteryl ester transfer protein; VLCD, very low calorie diet; PL, phospholipids; LDL-C, low density lipoprotein-cholesterol; HDL-C, high density lipoprotein-cholesterol; HDL-PL, high density lipoprotein-phospholipids.

## Hepatic triglyceride content

Hepatic TG content was measured in supine position using <sup>1</sup>H-MRS on a 1.5 Tesla whole-body MR scanner (Gyrosan ACS/NT15; Philips, Best, The Netherlands), exactly as previously described <sup>11</sup>.

## Plasma (apo)lipoprotein and CETP analyses

All plasma samples were obtained after an overnight fast before the start (i.e. after stopping all glucose-lowering medication including insulin) and after the 16-week VLCD protocol, stored in aliquots at -80°C, and analyzed after thawing once in a single laboratory (Leiden, The Netherlands). To ascertain that we could make adequate correlations, all analyses were performed within the same blood samples in the same assay runs. Plasma cholesterol and TG concentrations were determined using enzymatic kits (no. 236691 and 11488872, respectively, Roche Molecular Biochemicals, Indianapolis, IN, USA). Plasma phospholipids were determined using the phospholipids B kit (Wako Chemicals, Neuss, Germany). Plasma CETP concentration was quantified using kit 'CETP ELISA Daiichi' (Daiichi Pure Chemicals Co, Ltd, Tokyo, Japan). HDL fractions were obtained after precipitation of ApoB-lipoproteins from 50 µL plasma by adding 25 µL 36% polyethylene glycol 6000 (PEG6000, no.81260, Sigma Aldrich, Inc, USA.) The HDL-cholesterol and phospholipids were determined as described above. Plasma ApoAI and ApoB100 levels were determined with the 'Human ApoAI ELISA kit' (no. 3710-1H, Mabtech AB, Sweden) and 'Human ApoB ELISA kit' (no. 3715-1H; Mabtech AB, Sweden), respectively.

## Cholesterol efflux study

Cholesterol efflux to total plasma and ApoB-depleted human plasma were determined using the human monocyte cell line THP-1 as cholesterol donor. THP-1 cells were obtained from European Collection of Cell Cultures (ECACC), and maintained in medium A (RPMI 1640 with 25 mM HEPES Buffer, supplemented with 10% fetal bovine serum, 1% L-glutamine, 100 U/mL penicillin and 100 µg/mL streptomycin) at 37°C in 5% CO<sub>2</sub>. Before the experiment, THP-1 cells were seeded into 24 well plates at density of 5x10<sup>5</sup> cells per well and differentiated into macrophages with 0.1 µM phorbol 12-myristate-13-acetate (PMA; no. P1585, Sigma Aldrich, Inc, USA) within 3 days. Macrophages were washed three times with PBS and incubated in medium B (RPMI 1640 with 25 mM HEPES buffer, supplemented with 2% fetal bovine serum, 1% L-glutamine, 100 U/mL penicillin, 100 µg/mL streptomycin, 50 µg/mL acetyl-LDL and 10 µCi/ml [1α,2α(n)-<sup>3</sup>H]-cholesterol (no.NET139001MC, Perkin Elmer, Netherlands) for 1 day at 37°C in 5% CO<sub>2</sub>. After incubation, cells were washed 3 times with PBS and the efflux assay was started

by adding total human plasma or ApoB-depleted human plasma diluted to 1% in medium C (RPMI 1640 with 25 mM HEPES buffer, supplemented with 1% L-glutamine, 100 U/ml penicillin, 100 µg/ml streptomycin, and 0.5 mg/ml BSA). The whole assay was carried out in triplicate. To be able to normalize results between series of experiments and to correct for plate-to-plate variation, efflux to a standard preparation of HDL (50 µg protein/mL) was determined in triplicate. After 4 hours incubation, medium was collected and centrifuged. Subsequently, [<sup>3</sup>H]cholesterol was quantified by liquid scintillation counting. Total cellular <sup>3</sup>H-cholesterol was determined after extraction of the cells with 0.1 M NaOH. Cholesterol efflux rate was calculated by dividing the <sup>3</sup>H-activity in the medium by the sum of the <sup>3</sup>H-activity in the medium and the cell extract. Background values (the efflux in the absence of plasma) were subtracted.

### Statistical analysis

Data are expressed as means ± SEM. Paired *t*-tests were used for the statistical comparisons between measurements at baseline and after 16 weeks of caloric restriction. For correlation analysis, Pearson's correlation analysis was used. A *P*-value < 0.05 was considered statistically significant.

## RESULTS

In line with our previous observations in 12 patients <sup>11</sup>, a subset of the 27 patients who were included in the present study, the VLCD profoundly reduced bodyweight from 113.1±3.7 to 87.7±2.9 kg (*P*<0.05) and decreased BMI from 37.2±0.9 to 28.9±0.8 kg/m<sup>2</sup> (*P*<0.05). In addition, in the 12 patients from whom <sup>1</sup>H-MRS scans could be obtained, hepatic TG content considerably reduced from 21.2±4.2 to 3.0±0.9% (n =12, *P*<0.001) as reported previously <sup>11</sup>.

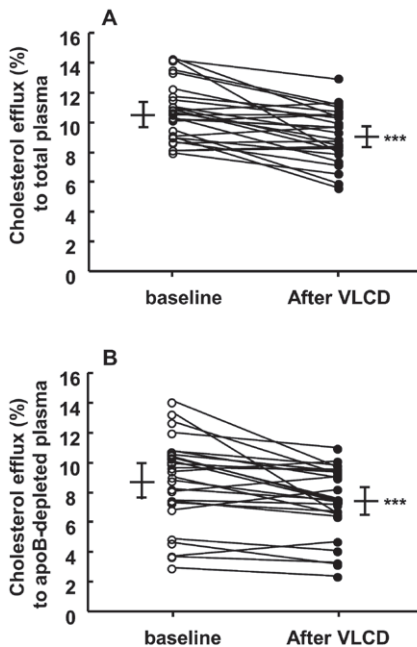
### Plasma CETP and (apo)lipoproteins

Compared to baseline, VLCD decreased plasma CETP concentration (-18.2%, *P*<0.01). In addition, VLCD reduced plasma levels of total cholesterol (-13.1%, *P*<0.001), TG (-45.1%, *P*<0.001), phospholipid (-15.2%, *P*<0.0001), LDL-cholesterol (-15.8%, *P*<0.01) and ApoB100 (-13.9%, *P*<0.01). VLCD did not alter plasma HDL-cholesterol and HDL-phospholipids, but increased ApoA1 (+16.2%; *P*<0.05) (Table 1). The change in bodyweight after 16 weeks of VLCD did not correlate with the change in either plasma triglycerides (*R*<sup>2</sup>=0.0000; *P*=0.9952), total cholesterol (*R*<sup>2</sup>=0.0471; *P*=0.2769), phospholipid (*R*<sup>2</sup>=0.0305; *P*=0.3837), LDL-cholesterol (*R*<sup>2</sup>=0.0320; *P*=0.3717), HDL-cholesterol (*R*<sup>2</sup>=0.0086; *P*=0.6460) or HDL-phospholipid (*R*<sup>2</sup>=0.0013; *P*=0.8570).

**Table 1. Plasma CETP and (apo)lipoprotein levels in obese patients with type 2 diabetes mellitus and hepatic steatosis in response to 16 weeks of VLCD.**

Plasma parameters	Baseline	After VLCD	Delta (%)	P value
CETP ( $\mu\text{g/mL}$ )	$2.48 \pm 0.15$	$2.03 \pm 0.14$	-18.2	0.0021
Total cholesterol (mM)	$5.76 \pm 0.30$	$5.00 \pm 0.22$	-13.1	0.0007
Triglycerides (mM)	$2.41 \pm 0.28$	$1.32 \pm 0.10$	-45.1	0.0003
Phospholipids (mM)	$2.69 \pm 0.11$	$2.28 \pm 0.07$	-15.2	0.0000
LDL-C (mM)	$3.99 \pm 0.27$	$3.36 \pm 0.20$	-15.8	0.0028
ApoB100 (mg/dL)	$130 \pm 6$	$111 \pm 5$	-13.9	0.0016
HDL-C (mM)	$0.84 \pm 0.04$	$0.91 \pm 0.06$	-	NS
HDL-PL (mM)	$0.98 \pm 0.03$	$0.98 \pm 0.05$	-	NS
ApoAI (mg/dL)	$135 \pm 10$	$156 \pm 11$	+16.2	0.0174

Delta-values are calculated by comparing values obtained after VLCD to those obtained at baseline from obese patients with T2DM. Data are presented as means  $\pm$  SEM (n=27). *P*-values are calculated using Paired Student's *t* test. CETP, cholesteryl ester transfer protein; VLCD, very low calorie diet; PL, phospholipids; LDL-C, low density lipoprotein-cholesterol; HDL-C, high density lipoprotein-cholesterol; HDL-PL, high density lipoprotein-phospholipids; NS, not significant.

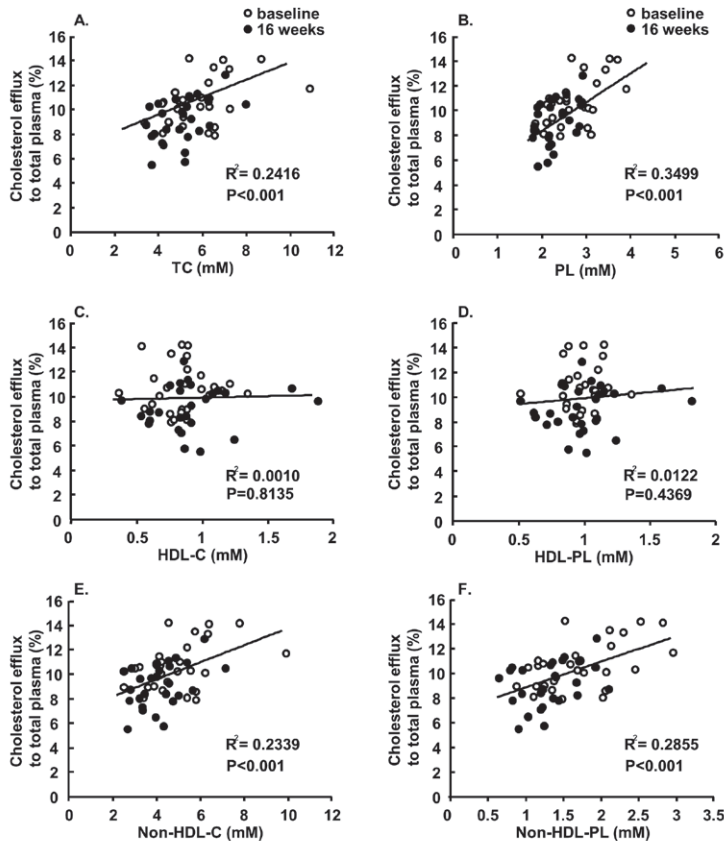


**Figure 1. Cholesterol efflux to total plasma and apoB-depleted plasma.** THP-1 cells were loaded with  $[^3\text{H}]$ cholesterol and incubated for 4 h at  $37^\circ\text{C}$  with total plasma (1% v/v) (A) or ApoB-depleted plasma (1% v/v) (B), obtained before and after VLCD from 27 obese patients with T2DM. Cholesterol efflux rate is calculated by dividing  $^3\text{H}$ -activity in the medium by the sum of the  $^3\text{H}$ -radioactivity in the medium and cell extract. Data are means  $\pm$  SEM. *P*-values are calculated using Paired Student's *t* test. \*\*\**P*<0.001 as compared to baseline. ApoB, Apolipoprotein B; HDL, high density lipoprotein; VLCD, very low calorie diet.



## Cholesterol efflux

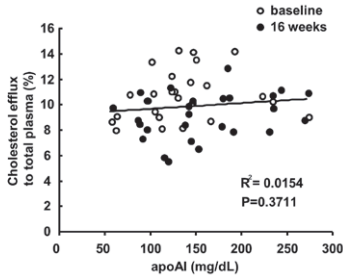
VLCD decreased cholesterol efflux from THP-1 cells to total plasma obtained from patients compared to plasma from baseline (-14.5%,  $P < 0.001$ ) (Fig. 1A). Similarly, the capacity of apoB-depleted plasma obtained after VLCD to promote cholesterol efflux was lower than that of apoB-depleted plasma obtained at baseline (-14.9%;  $P < 0.001$ ) (Fig. 1B).



**Supplemental Figure 1. Correlation between cholesterol efflux and plasma lipids.** THP-1 cells were loaded with [ $^3\text{H}$ ]cholesterol and incubated for 4 h at 37°C with total plasma (1% v/v) obtained at baseline and after VLCD from 27 obese patients with T2DM. Cholesterol efflux rate was calculated by dividing  $^3\text{H}$ -activity in the medium by the sum of the  $^3\text{H}$ -radioactivity in the medium and cell extract. Cholesterol efflux was plotted against total cholesterol (TC, A), phospholipid (PL, B), HDL-C (C), HDL-PL (D), Non-HDL-C (E), and Non-HDL-PL (F), and correlations were calculated.

Correlation analysis showed that cholesterol efflux to total plasma positively correlated with plasma total cholesterol ( $R^2 = 0.2416$ ;  $P < 0.001$ ) and plasma total

phospholipid ( $R^2=0.3499$ ;  $P<0.001$ ). Cholesterol efflux to total plasma positively correlated with non-HDL-cholesterol ( $R^2=0.2339$ ;  $P<0.001$ ) and non-HDL-phospholipid ( $R^2=0.2855$ ;  $P<0.001$ ) rather than HDL-cholesterol or HDL-phospholipid (both  $P>0.05$ ) (Supplemental Fig. 1). Moreover, no significant correlation was observed between cholesterol efflux to plasma and apoAI (Supplemental Fig. 2).



**Supplemental Figure 2. Correlation between cholesterol efflux and plasma apoAI.** THP-1 cells were loaded with [ $^3\text{H}$ ]cholesterol and incubated for 4 h at 37°C with total plasma (1% v/v) obtained at baseline and after VLCD from 27 obese patients with T2DM. Cholesterol efflux rate was calculated by dividing  $^3\text{H}$ -activity in the medium by the sum of the  $^3\text{H}$ -radioactivity in the medium and cell extract. Cholesterol efflux was plotted against apoAI, and correlations were calculated.

## DISCUSSION

A main finding from the present study is that prolonged caloric restriction by a 16-week VLCD in obese patients with type 2 diabetes mellitus and hepatic steatosis, which considerably reduces hepatic TG content (-85%)<sup>11</sup>, also markedly decreases plasma CETP concentration (-18.2%). This observation corroborates our recent finding that a reduction of the hepatic lipid content (-30.5%), as induced by pioglitazone, also associates with a reduction in plasma CETP concentration (-11.6%) in patients with type 2 diabetes mellitus<sup>10</sup>. However, the potency of prolonged caloric restriction to reduce hepatic lipid content and plasma CETP concentration exceeds that of pioglitazone treatment considerably.

These data are in full accordance with our previous observations that lowering hepatic lipids (i.e. TG as well as cholesterol) in *APOE\*3-Leiden.CETP* mice by classical lipid-lowering drugs decreased hepatic CETP mRNA expression, resulting in decreased plasma CETP concentration<sup>6-8</sup>. Since CETP expression is regulated by liver X receptor alpha (LXR $\alpha$ ) for which oxysterols are natural ligands<sup>12</sup> and the liver cholesterol level determines LXR $\alpha$  activation<sup>13</sup>, we concluded from those studies that a decrease in hepatic cholesterol content, associated with a decrease in cholesterol derivatives,

reduces hepatic LXR $\alpha$  activation, thereby downregulating CETP mRNA transcription. Although it is unknown whether hepatic TG levels reflect levels of hepatic cholesterol and oxysterols in the present study, as we cannot assess hepatic (oxy)sterols noninvasively in humans, hepatic TG and cholesterol levels were highly correlated ( $r=0.867$ ) in 33 Chinese subjects (Dr. P. Parini, personal communication). Therefore, it is likely that the reduction in plasma CETP concentration induced by VLCD also reduces hepatic LXR $\alpha$ -activated CETP mRNA transcription thereby reducing plasma CETP. Our data corroborate those of Laimer *et al.*<sup>14</sup> who showed that substantial weight loss in morbidly obese women induced by laparoscopic gastric banding surgery also decreased plasma CETP mass (-8.3%) at 1 year after surgery.

The effect of caloric restriction on HDL levels is still under debate. Although a single study reported that an average of 6 years of caloric restriction in 18 subjects increased HDL-cholesterol<sup>15</sup>, other studies demonstrated that both 6 months and 2 years of caloric restriction in 8 subjects in fact decreased plasma HDL-cholesterol<sup>16,17</sup>. Moreover, caloric restriction had conflicting effects on HDL-cholesterol among different diet groups even in one study: HDL-cholesterol was either increased or unaffected<sup>18</sup>. The current study is the first to show that caloric restriction by a VLCD increased the plasma level of the main HDL protein constituent ApoAI (+16.2%), which is supposed to be a good (negative) predictor of CVD risk<sup>19</sup>. However, the VLCD did not increase HDL-cholesterol levels. Our previous studies in mice<sup>6-8</sup> and in patients with type 2 diabetes mellitus<sup>10</sup> showed that reduction of hepatic lipids and plasma CETP, as induced by drugs, were in fact related to increased plasma HDL-cholesterol levels. In fact, treatment of only 20 patients with type 2 diabetes mellitus with pioglitazone resulted in a significant increase in HDL-cholesterol despite less pronounced decreases in hepatic lipid and plasma CETP<sup>10</sup>, suggesting that our present study including 27 patients would not be underpowered to detect an potential effect on HDL-C. It is known that the LXR $\alpha$  target ATP binding cassette A1 (ABCA1) plays a crucial role in HDL maturation by mediating the lipidation of plasma ApoAI<sup>20</sup>. Furthermore, hepatic ABCA1 is the main contributor to the loading of HDL with cholesterol as evidenced by studies in mice that selectively lack ABCA1 from the liver<sup>21</sup>. Collectively, it is thus conceivable that the dramatic reduction in hepatic lipids largely downregulates the hepatic expression of ABCA1, resulting in reduced lipidation of plasma ApoAI with liver-derived cholesterol, thereby counteracting the expected rise in HDL-cholesterol due to the reduction in CETP.

The reduced ratio of HDL-cholesterol over apoAI as induced by VLCD may be expected to result in an increased ability of total plasma and ApoB-depleted plasma to induce cholesterol efflux from cholesterol-laden macrophages. However, we observed that the VLCD actually reduced the capacity of total plasma and ApoB-depleted plasma

to induce cholesterol efflux from THP-1 cells, even after discarding the three subjects showing the largest decrease in cholesterol efflux from the analysis. Correlation analysis revealed that HDL constituents (cholesterol, phospholipid and apoAI) did not correlate with cholesterol efflux to plasma. Instead, the decrease in cholesterol efflux to plasma was mainly related to the decrease in total phospholipid levels in plasma. Collectively, these data indicate that the total lipoprotein surface area in plasma, rather than the HDL level, determines the capacity of plasma to induce cholesterol efflux.

Our findings are in line with those of a recent study showing that the cholesterol efflux capacity of plasma was independent of total HDL-C or ApoAI levels<sup>22</sup>. In fact, independently of HDL-cholesterol, sera with high efflux capacity had a significant increase in ABCA1-mediated efflux due to the presence of pre $\beta$ -1 HDL. Likewise, another study showed that, although both pioglitazone and statins increase HDL-cholesterol, pioglitazone but not statins increases the cholesterol efflux capacity of plasma, and no correlation was noted between the change in HDL-cholesterol and the change in cholesterol efflux capacity<sup>23</sup>. Overall, the cholesterol efflux capacity of plasma is thus not simply related to HDL-cholesterol or apoAI, although the capacity of HDL to mediate cholesterol efflux from macrophages has recently been established to strongly inversely associate with carotid intima-media thickness and likelihood of angiographic coronary artery disease<sup>23</sup>. The fact that we were unable to detect a correlation between cholesterol efflux and HDL-cholesterol or apoAI thus probably indicates that VLCD does not improve the functionality of HDL with respect to mediating cholesterol efflux.

A limitation of the current study may be the small study group. Although we initially included 27 patients for the VLCD intervention, hepatic TG quantification by MRS was possible in only 12 patients, mainly related to limitations for maximum weight and circumference of the MRI scanner. Secondly, the design of the current human study does not permit to assess causal relationships between hepatic lipid content and plasma CETP or ApoAI level. Nonetheless, the results are in accordance with data obtained from mechanistic studies in a mouse model relevant for human lipoprotein metabolism<sup>6-8</sup>.

In conclusion, this study indicates that prolonged caloric restriction, which considerably reduces hepatic TG content, also markedly decreases plasma CETP concentration. This is in full accordance with our previous findings in *APOE\*3-Leiden.CETP* mice, in which we showed that classical lipid-lowering drugs concurrently lowered hepatic lipids resulting in decreased plasma CETP concentration. Furthermore, the VLCD increased plasma apoAI levels without improving functionality of HDL with respect to cholesterol efflux from macrophages.

## ACKNOWLEDGMENTS

This research was supported by the Center for Translational Molecular Medicine (CTMM), project PREDICt (grant 01C-104), by the Netherlands Heart Foundation (grant 2007B081) and by Roba Metals B.V., IJsselstein, The Netherlands. P.C.N.R. is an Established Investigator of the Netherlands Heart Foundation (grant 2009T038).

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