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Daysleeper : from genomic parasite to indispensable gene

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Citation

Knip, M. (2012, November 22). *Daysleeper : from genomic parasite to indispensable gene*. Retrieved from <https://hdl.handle.net/1887/20170>

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Title: Daysleeper : from genomic parasite to indispensable gene

Date: 2012-11-22

Chapter 6

DAYSLEEPER plays an important role in several plant processes, through its involvement in the ubiquitylation machinery

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Summarizing discussion

D*AYSLEEPER* is essential in Arabidopsis [1] and we have shown that it is conserved in higher plants. We identified and cloned homologous genes from the grapevine (*Vitis vinifera*) and rice (*Oryza sativa*) genomes and obtained evidence of expression of homologous genes in many other angiosperms, including lower angiosperms (Chapter 2, this thesis). *DAYSLEEPER* is widely expressed throughout the plant, although this is most pronounced in meristems and developing flowers. *DAYSLEEPER* is located both within the nucleus and in vesicular structures of plant cells (Chapter 3, this thesis).

We have found that overexpression of *DAYSLEEPER* disturbs the regulation of many genes (Chapter 4, this thesis) and also that the *DAYSLEEPER* protein interacts with several other proteins and seems to influence the abundance of CULLIN1 (Chapter 5, this thesis). In this discussion we try to place our experimental data in perspective and put forward a model of the functional context we think *DAYSLEEPER* is involved in (Figure 1).

DAYSLEEPER and the Endosomal Sorting Complex Required for Transport-machinery III (ESCRT-III)

6 *DAYSLEEPER* is able to interact with the proteins VPS2.1, VPS2.2 and VPS2.3 (chapter 5, this thesis). These proteins are homologs of the canonical VPS2 subunit of the Endosomal Sorting Complex Required for Transport-machinery III (ESCRT-III) [2, 3]. The ESCRT-III machinery is involved in sorting ubiquitylated cargo proteins into vesicles before degradation [4]. *Daysleeper* mutants display a disrupted cellular organization in the root, a problem which also occurs in *vps2.2* and *vps2.3* plantlets, albeit less severe (Chapter 5, this thesis) [5, 6]. This might suggest that *DAYSLEEPER* and *VPS2.2* and *VPS2.3* are involved in cytokinesis in the root, although *VPS2.1 Vps2.1* plants are embryo lethal [6, 7]. *Vps2.1* has the most overlapping expression pattern with *DAYSLEEPER* in the root.

It was shown that that ESCRT-III machinery is necessary for the establishment of proper auxin maxima in Arabidopsis [8]. It is thought that this is caused by the fact that auxin transporters, such as PIN1, need vesicular transport to be targeted to specific sites of the cellular membrane [8, 9]. *Daysleeper* mutants also display disturbed auxin maxima (Chapter 5, this thesis). We found that RFP:*DAYSLEEPER* co-localizes in vesicular

structures with PIN1:GFP in protoplasts (Chapter 5, this thesis). However, although maxima are irregularly shaped in *daysleeper* plants, they are formed, which indicates that PIN1 transport does not require DAYSLEEPER. It seems therefore more likely that the aberrant auxin maxima observed in *daysleeper* mutants might be the result of problems with cellular organization, possibly caused by disturbance of a functional connection of DAYSLEEPER to (one of the) VPS2 proteins.

In our transcriptomics data, we found the downregulation of AT1G13340 (Chapter 4, this thesis) [1]. This gene is a putative negative regulator of *VPS4*, which is a subunit of the ESCRT-III machinery [10]. AT1G13340 contains an InterPro domain IPR005061 [11], which is also found in the negative regulator of *VPS4* in yeast, *Ist1* [11, 12]. Although this data does not provide a direct functional link, it is a further indication of the involvement of DAYSLEEPER with ESCRT-III related proteins.

The ESCRT-III machinery has also been shown to be involved in membrane budding and cytokinesis and there are also indications that proteins in this complex might fulfill a nuclear function [2, 3]. ESCRT-related protein CHMP1/*VPS46* has been reported to interact with Polycomb-like factor Pcl on condensed chromatin in human cells [13]. Furthermore, CHMP2 overexpression induces condensed chromatin regions, to which Polycomb group protein *Rac1* and phosphorylated histone H3 are recruited [14]. Recently it was also described that ESCRTIII protein CHMP4C in human cells (homologous to *SNF7* in Arabidopsis) has a role in maintaining genomic stability by regulating the phosphorylation of chromatin associated protein *Aurora B* [15]. An interesting observation is therefore the nuclear localization of the interaction of the VPS2-proteins and DAYSLEEPER (Chapter 5, this thesis). From a proteomic analysis it has become apparent that *VPS2.2* in Arabidopsis may play a role in the nucleus [16], which is in line with our finding that DAYSLEEPER has been shown to interact with VPS2-proteins in the nucleus (Chapter 3 and 5, this thesis).

DAYSLEEPER, the COP9-signalosome (CSN) and CULLIN-ring ligases (CRL's)

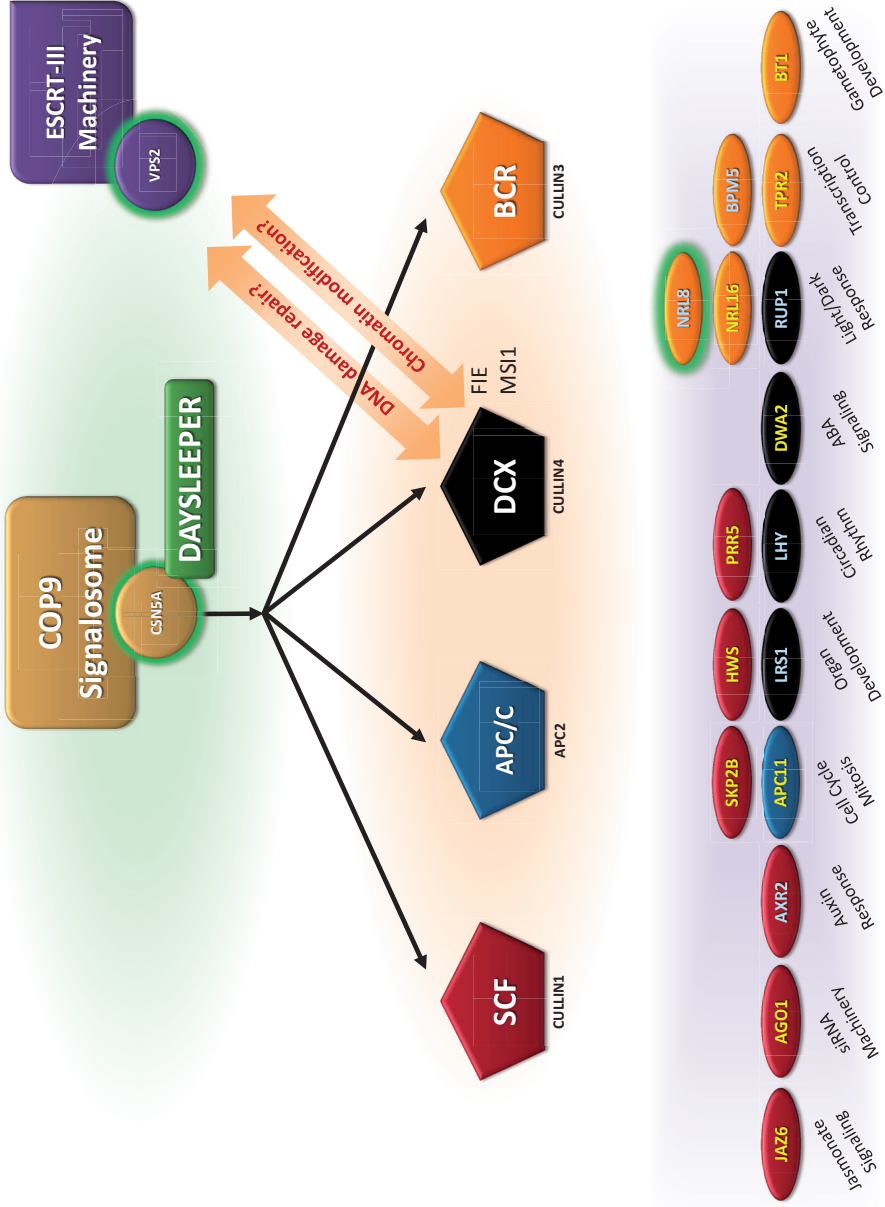
We found that DAYSLEEPER interacts with the COP9-signalosome (CSN) subunit *CSN5A* (Chapter 3, this thesis). *Daysleeper* plantlets display similarities to *cop* mutants [1, 17, 18]. *Cop* mutants are similar to *daysleeper* mutants in that they do not progress past the seedling stage and accumulate DNA-damage (Chapter 5, this thesis) [19]. The light-grown phenotype of *cop* mutants is not shared by *daysleeper* plantlets and also the accumulation of anthocyanin is not observed [1].

We found that *daysleeper* seedlings have an accumulation of CULLIN1 (Chapter 5, this thesis). Furthermore, we found that induced *DAYSLEEPER* overexpression prompts the differential regulation of several genes, among which many CRL-related genes and COP9-signalosome subunits, as well as genes linked to the Anaphase Promoting Complex/Cyclosome (APC/C) (Chapter 4, this thesis).

We have found that *DAYSLEEPER* has a very similar expression pattern to chromatin related genes *MSI1* and *FIE* (Chapter 4, this thesis). *FIE* is a subunit in a CUL4-containing DCX-type CRL [20]. Furthermore, there is evidence that *DAYSLEEPER* can also interact with a CUL4-related protein *in vivo* (personal communication Alexander Kenzior, Columbia, MI, USA). DCX complexes have been known to play a role in DNA damage repair, chromatin remodeling and several other processes [21–23].

The abovementioned findings lead us to conclude that *DAYSLEEPER* functions in concert with both CSN and CRL complexes. *DAYSLEEPER*'s precise mode of activity remains unknown, but the increased amount of CUL1 in *daysleeper* mutants could point to a role in regulating CULLIN expression or stability. However, we did not find an effect of *DAYSLEEPER* overexpression on CULLIN transcript abundance (Chapter 4, this thesis). Therefore, it is likely that *DAYSLEEPER* has an effect in CULLIN translation or stability. Also *DAYSLEEPER* has a highly correlated expression pattern to *ECR-1* (Chapter 4, this thesis). *ECR-1* is responsible for the rubylation of CULLINs, which is thought to regulate their activity [24]. It is interesting that *DAYSLEEPER* was first identified as a protein binding to the promoter of the DNA damage repair gene *KU70* and that both *cop* and *daysleeper* mutants accumulate DNA damage [18, 19] (Chapter 5, this thesis). *DAYSLEEPER* may therefore be associated with DCX-type CRL's, possibly in a complex associated with DNA damage repair or maintenance of genomic stability [22, 23]. Alternatively, *DAYSLEEPER* might be part of a chromatin remodeling related complex with *FIE*, which is highly coexpressed with *DAYSLEEPER* (Chapter 5), and CUL4, similar to a complex recently described by Pazhouhandeh *et al.* (2011) [20]. Furthermore, we found that several chromatin-related genes are misregulated when *DAYSLEEPER* is overexpressed and that overlap exists between our expression data set and a data set obtained by studying gene expression in the histone methyltransferase *sdg4* [25] mutant (Chapter 4, this thesis).

Figure 1. Model of *DAYSLEEPER*'s influence on the regulation of CRL's and the CSN. The CSN derubylates CRL's, thereby regulating their activity (reviewed in Hotton *et al.* (2008) [26]). Genes are depicted in ovals with colors referring to the type of CRL that regulates them. Burgundy = SCF, Black = DCX, APC/C = Blue and BCR = orange. The color of the text of the gene refers to the up- or downregulation in the SAGE-seq dataset (Chapter 4, this thesis; 24 hours of *DAYSLEEPER* induction). Genes written with blue letters are downregulated and yellow-written genes are upregulated. Ovals that have a green glow (i.e. *NRL8* and *CSN5A*) are genes that have been shown to interact with *DAYSLEEPER* (Chapter 5, this thesis). Genes that have a role in chromatin modifying complexes and are found to have a highly correlated expression pattern to *DAYSLEEPER* are placed to the right from DCX (i.e. *MSI1* and *FIE*) (Chapter 5, this thesis). The ESCRT-III machinery (purple box) is involved in sorting ubiquitylated cargo, before degradation [3]. *DAYSLEEPER* also interacts with one of the components (*VPS2*) of this complex (Chapter 5, this thesis).



Conclusion: A graphical representation of DAYSLEEPER's field of function

We created a schematic representation of the interactions of DAYSLEEPER based on the interactions and influence of induced DAYSLEEPER overexpression discussed in the paragraphs above. *DAYSLEEPER* is likely to have a role in the interplay between the CSN and CRL complexes and the related APC/C complex, in connection to protein ubiquitylation (Figure 1). We have found many genes that are part of CRL's or genes that are regulated by CRL's (Chapter 4, this thesis) that are misregulated when *DAYSLEEPER* is overexpressed. This indicates that DAYSLEEPER plays a role in the fundamental processes of the interplay between the CSN and CRL complexes, which is substantiated by the severe phenotype of both the *daysleeper* and the overexpression mutants.

We speculate that the main functionality of the interaction between DAYSLEEPER and the ESCRTIII-complex through its interaction partner VPS2 could be the involvement of DAYSLEEPER in DCX-type CRL's, involved in DNA damage repair or chromatin modifying (PRC2-related) complexes (Figure 1).

In this work we have made steps towards understanding *DAYSLEEPER* evolution and function. Although the exact molecular role of DAYSLEEPER remains elusive, our findings suggest a role in elemental cellular processes and offers an enticing starting point for further understanding the role of *DAYSLEEPER* in Arabidopsis, and *SLEEPER* genes in other plant species.

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References

1. Bundoek P, Hooykaas P: **An Arabidopsis hAT-like transposase is essential for plant development.** *Nature* 2005, **436**:282–4.
2. Wollert T, Wunder C, Lippincott-schwartz J, Hurley JHJH, H J: **Membrane scission by the ESCRT-III complex.** *Nature* 2009, **458**:172–7.
3. Lata S, Schoehn G, Solomons J, Pires R, Göttlinger HG, Weissenhorn W: **Structure and function of ESCRT-III.** *Biochem. Soc. Trans.* 2009, **37**:156–60.
4. Babst M, Katzmann DJ, Estepa-Sabal EJ, Meerloo T, Emr SD: **Escrt-III: an endosome-associated heterooligomeric protein complex required for mvb sorting.** *Dev. Cell* 2002, **3**:271–82.
5. Müller S, Fuchs E, Ovecka M, Wysocka-Diller J, Benfey PN, Hauser MT: **Two new loci, PLEIADE and HY-ADE, implicate organ-specific regulation of cytokinesis in Arabidopsis.** *Plant Physiol.* 2002, **130**:312–324.
6. Katsiarimpa A, Anzenberger F, Schlager N, Neubert S, Hauser M-T, Schwechheimer C, Isono E: **The Arabidopsis Deubiquitinating Enzyme AMSH3 Interacts with ESCRT-III Subunits and Regulates Their Localization.** *Plant Cell* 2011, **23**:3026–3040.
7. Sessions A, Burke E, Presting G, Aux G, McElver J, Patton D, Dietrich B, Ho P, Bacwaden J, Ko C, Clarke JD, Cotton D, Bullis D, Snell J, Miguel T, Hutchison D, Kimmerly B, Mitzel T, Katagiri F, Glazebrook J, Law M,

- Goff SA: **A high-throughput Arabidopsis reverse genetics system.** *Plant Cell* 2002, **14**:2985–94.
8. Spitzer C, Reyes FC, Buono R, Sliwinski MK, Haas TJ, Otegui MS: **The ESCRT-related CHMP1A and B proteins mediate multivesicular body sorting of auxin carriers in Arabidopsis and are required for plant development.** *Plant Cell* 2009, **21**:749–66.
9. Grunewald W, Friml J: **The march of the PINs: developmental plasticity by dynamic polar targeting in plant cells.** *EMBO J.* 2010, **29**:2700–14.
10. Obita T, Saksena S, Ghazi-Tabatabai S, Gill DJ, Perisic O, Emr SD, Williams RL: **Structural basis for selective recognition of ESCRT-III by the AAA ATPase Vps4.** *Nature* 2007, **449**:735–9.
11. Hunter S, Apweiler R, Attwood TK, Bairoch A, Bateman A, Binns D, Bork P, Das U, Daugherty L, Duquenne L, Finn RD, Gough J, Haft D, Hulo N, Kahn D, Kelly E, Laugraud A, Letunic I, Lonsdale D, Lopez R, Madera M, Maslen J, McAnulla C, McDowall J, Mistry J, Mitchell A, Mulder N, Natale D, Orengo C, Quinn AF, Selengut JD, Sigrist CJA, Thimma M, Thomas PD, Valentin F, Wilson D, Wu CH, Yeats C: **InterPro: the integrative protein signature database.** *Nucleic Acids Res.* 2009, **37**:D211–5.
12. Rue SM, Mattei S, Saksena S, Emr SD: **Novel Ist1-Did2 complex functions at a late step in multivesicular body sorting.** *Molecular Biology of the Cell* 2008, **19**:475–484.
13. Stauffer DR, Howard TL, Nyun T, Hollenberg SM: **CHMP1 is a novel nuclear matrix protein affecting chromatin structure and cell-cycle progression.** *J. Cell. Sci.* 2001, **114**:2383–93.
14. Hodges E, Redelius JS, Wu W, Höög C: **Accelerated discovery of novel protein function in cultured human cells.** *Mol. Cell Proteomics* 2005, **4**:1319–27.
15. Carlton JG, Caballe a., Agromayor M, Kloc M, Martin-Serrano J: **ESCRT-III Governs the Aurora B-Mediated Abscission Checkpoint Through CHMP4C.** *Science (80-)* 2012, **220**.
16. Ibl V, Csaszar E, Schlager N, Neubert S, Spitzer C, Hauser M-T: **Interactome of the plant-specific ESCRT-III component AtVPS2.2 in Arabidopsis thaliana.** *J. Proteome Res.* 2012, **11**:397–411.
17. Wei N, Deng XW: **The COP9 signalosome.** *Annu. Rev. Cell Dev. Biol.* 2003, **19**:261–86.
18. Gusmaroli G, Feng S, Deng XW: **The Arabidopsis CSN5A and CSN5B subunits are present in distinct COP9 signalosome complexes, and mutations in their JAMM domains exhibit differential dominant negative effects on development.** *Plant Cell* 2004, **16**:2984–3001.
19. Chamovitz D a: **Revisiting the COP9 signalosome as a transcriptional regulator.** *EMBO Rep.* 2009, **10**:352–8.
20. Pazhouhandeh M, Molinier J, Berr A, Genschik P: **MSI4/FVE interacts with CUL4-DDB1 and a PRC2-like complex to control epigenetic regulation of flowering time in Arabidopsis.** *Proc. Natl. Acad. Sci. U.S.A.* 2011, **108**:3430–5.
21. Hua Z, Vierstra RD: **The cullin-RING ubiquitin-protein ligases.** *Annu Rev Plant Biol* 2011, **62**:299–334.
22. Biedermann S, Hellmann H: **The DDB1a interacting proteins ATCSA-1 and DDB2 are critical factors for UV-B tolerance and genomic integrity in Arabidopsis thaliana.** *Plant J.* 2010, **62**:404–15.
23. Castells E, Molinier J, Benvenuto G, Bourbousse C, Zabulon G, Zalc A, Cazzaniga S, Genschik P, Barneche F, Bowler C: **The conserved factor DE-ETIOLATED 1 cooperates with CUL4-DDB1/DDB2 to maintain genome integrity upon UV stress.** *EMBO J.* 2011, **30**:1162–72.
24. Pozo JC, Timpte C, Tan S, Callis J, Estelle M: **The ubiquitin-related protein RUB1 and auxin response in Arabidopsis.** *Science (80-)* 1998, **280**:1760–3.
25. Cartagena J a, Matsunaga S, Seki M, Kurihara D, Yokoyama M, Shinozaki K, Fujimoto S, Azumi Y, Uchiyama S, Fukui K: **The Arabidopsis SDG4 contributes to the regulation of pollen tube growth by methylation of histone H3 lysines 4 and 36 in mature pollen.** *Dev. Biol.* 2008, **315**:355–68.
26. Hotton SK, Callis J: **Regulation of cullin RING ligases.** *Annu Rev Plant Biol* 2008, **59**:467–89.

